Soil-surface $CO₂$ fluxes in a *Deyeuxia angustifolia* wetland in Sanjiang Plain, China

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Received 13 February 2003; accepted in revised form 7 August 2003

Key words: CO₂ flux, Deyeuxia angustifolia wetland, Sanjiang Plain soil-surface, Soil temperature

Abstract

In many temperate-zone ecosystems, seasonal changes in environmental and biological factors influence the dynamics and magnitude of surface–atmosphere exchange. Research was conducted between July and October 2001 to measure growing season surface-layer fluxes of $CO₂$ in a *Deveuxia angustifolia* dominated wetland on the Sanjiang Plain in northeastern China. Seasonal fluctuation and daily change in soil-surface $CO₂$ fluxes were measured as well as the edaphic factors controlling $CO₂$ fluxes. Soil-surface $CO₂$ fluxes were measured with a closed-chamber system. The results revealed that there were both seasonal fluctuations and daily change in CO₂ fluxes. The ranges of measured soil-surface CO₂ flux were 0.208–1.265 g CO₂ m⁻² h⁻¹. Soil-surface CO₂ fluxes averaged 0.620 g CO₂ m⁻² h⁻¹. An analysis of several edaphic factors including soil temperature and soil moisture of the *D. angustifolia* wetland showed that there was a significant relationship between flux and temperature ($R^2 = 0.77$).

Introduction

Increasing concentrations of atmospheric $CO₂$ and the potential for climatic change have made the global carbon balance an important scientific and political topic (Tans and Balkwin 1995). Soil $CO₂$ efflux is known to play a major role in the global carbon cycle. At the global scale, the annual flux of $CO₂$ from soil to the atmosphere is estimated to average 68 Pg C y^{-1} (Raich and Schlesinger 1992), roughly 10 times the current annual contribution from fossil fuel combustion (Andres et al. 1996). Soil-surface $CO₂$ flux measurements have been widely used to construct ecosystem C budgets (Singh et al. 1988; Norman et al. 1992; Hungate et al. 1997). At the microcosmic level, in-depth studies have been made on carbon cycles of forest ecosystems, cropland ecosystems, and grassland

ecosystems, but wetland ecosystems have been studied very little. This is the case even more so for China (Lu et al. 1995; Yang and Lu 1999). In this paper, we carried out a field study of soil-surface $CO₂$ fluxes of a *Deyeuxia angustifolia* dominated wetland.

Measurements of soil-surface $CO₂$ flux

Several techniques have been used to measure soilsurface $CO₂$ flux including soil $CO₂$ profile, micrometeorological (including eddy covariance), and static and dynamic chamber methods. Although the soil $CO₂$ profile and micrometeorological techniques avoid confounding chamber effects, their application has generally been limited by a combination of cost and methodological requirements (Jong et al. 1979; Baldocchi and Meyers 1991; Verma et al. 1992). In particular, the soil $CO₂$ profile technique requires the estimation of soil transport coefficients, while eddy covariance typically requires specific atmospheric conditions and an estimation of aboveground plant respiration.

A majority of soil $CO₂$ flux studies have relied on the static chamber technique because it is relatively inexpensive and easy to employ (Gupta and Singh 1981; Bowden et al. 1993; Toland and Zak 1994; Marra and Edmonds 1996). In this method, $CO₂$ diffusing from the soil is absorbed inside a closed chamber using either an alkali solution (e.g., KOH, NaOH) or soda lime (Reinke et al. 1981; Edwards 1982). Although this technique offers some practical advantages, several studies have shown that it tends to overestimate $CO₂$ flux at low flux rates and severely underestimate $CO₂$ flux at high flux rates (Rochette et al. 1992; Nay et al. 1994; Jensen et al. 1996).

Dynamic chamber techniques have also been used to measure soil $CO₂$ efflux in numerous studies (Kucera and Kirkham 1971; Ewel et al. 1987; Hall et al. 1990; Norman et al. 1992; Luo et al. 1996). Dynamic chambers are generally considered more accurate than static methods (Cropper et al. 1985; Rochette et al. 1992; Nay et al. 1994) and can be operated as either closed or open systems. In closed dynamic systems, air is circulated in a closed loop between the chamber head space and an infrared gas analyzer (IRGA). Soil $CO₂$ efflux is then calculated using the difference in $CO₂$ concentration between the beginning and end of the measurement period. In open dynamic systems, ambient air is passed continuously through the chamber head space, and soil $CO₂$ efflux is calculated using the difference in $CO₂$ concentration between air entering and leaving the chamber. Open systems are typically preferred for continuous measurements over periods of hours to days because the flow of outside air into the system maintains near ambient chamber temperature and CO₂ concentration.

Site description

The study site was located in the Sanjiang Plain, a low alluvial plain formed by three rivers: Songhuajiang River, Heilongjiang River, and Wusulijiang River. The Sanjiang Plain is situated in the northeast part of China, with a total area of 5.13×10^6 ha, and it is the most wide spread area of

freshwater wetlands in China with 21% mires (Yang 1989). Freshwater mire wetlands are widespread and quite typical in this area.

Measurements were carried out at the ecological experimental area in the Ecological Experiment Station of Mire-Wetland in the Sanjiang Plain, Chinese Academy of Science, which belongs to one of the fundamental stations of the Chinese Ecosystem Research Network (CERN). The station lies in Honghe Farm, Tongjiang City, Heilongjiang Province, China (47°35'N, 133°31'E) and encompasses 100 ha.

The Ecological Experiment Station of Mire-Wetland is located in the interior of the Sanjiang Plain with a continental monsoon climate of the temperate zone. The mean annual temperature is about 1.9 \degree C and the mean annual precipitation is about 550 mm, 70% of which occur from June to September. The area has a low, smooth topography developed in a meander bend, and various depressions causing poor drainage of the surface. The natural vegetation is dominated by *D. angusti*folia, but meadow and mire plants such as Carex lasiocapa, C. pseudocuraica, and C. meyeriana along with secondary forest consisting of Quercus and Betula stands can also be found. The soil, except meadow soil and brown soil, is dominated by various mire soils, including meadow-, humus-, peat-boggy soils, and peat soil. We studied soilsurface $CO₂$ fluxes of the *D. angustifolia* wetland. At the sampling sites, the soil type is meadow lessive (Yang and Lu 1996). Soil chemical characteristic are given in Table 1.

Methods

Soil-surface $CO₂$ flux was measured with closeddynamic chambers and an IRGA. Flux measurements were made during the growing season, 2001. Collars $(50 \times 50 \text{ cm}$ polyvinyl-chloride (PVC) frame) were placed into the ground to a depth of 2–4 cm. For each flux measurement, a transparent rectangular Plexiglas chamber cover $(50 \times 50 \times 50 \text{ cm})$ was placed over the collar, and a QGS-08B IRGA was used to measure $CO₂$ concentration within the chamber. Air was circulated between the chamber and the IRGA with a small pump, at a rate of 0.5 l min^{-1} . CO₂ concentrations were measured at 1-min intervals.

Table 1. Chemical characteristics of meadow lessive soil type.

		Organic matter content $(\%)$	Entire N $(\%)$	Entire P $(\%)$	Entire K $(\%)$	Mineral composition $(\%)$					
Soil depth (cm)	pH					SiO ₂	Fe ₂ O ₃	Al_2O_3	TiO ₂	CaO	MgO
$0 - 13$	4.94	10.19	0.54	0.12	2.03	62.38	3.97	10.57	0.82	1.86	0.07
$13 - 21$	4.74	3.18	0.24	0.05	2.13	67.47	4.11	12.30	0.86	1.69	0.24
$21 - 40$	5.29	0.57	0.07	0.05	2.32	70.87	4.19	21.21	1.20	1.60	0.07
$40 - 60$	5.59	2.80	0.10	0.04	2.12	59.61	7.15	16.89	0.93	1.74	0.24
$60 - 100$	6.48	0.79	0.08	0.04	2.14	57.96	7.26	17.20	0.91	1.77	0.29

Figure 1. Seasonal pattern of soil $CO₂$ fluxes of D. angustifolia wetland.

Fans mounted inside each chamber ensured wellmixed air during the sampling period. Aboveground vegetation was clipped before the cover was put in place; thus, we did not have to consider the vegetative photosynthesis before we measured the concentration of $CO₂$ in chamber. Three replicate chambers were measured on each sampling day. We measured soil temperature with ageothermometer, and soil moisture was measured with the roast method.

A linear regression was applied to the linear portion of $CO₂$ concentration. The function (Yang and Du 1996) for soil-surface $CO₂$ flux is

$$
F = k \cdot a \cdot \frac{\Delta c}{\Delta t} \cdot h,\tag{1}
$$

where F represents the soil-surface CO_2 flux (g CO_2) $m^{-2} h^{-1}$), \hat{k} is the appropriate coefficient, Δc is the difference in $CO₂$ concentration in the chamber during the Δt minute, and h is the height of the chamber. Fluxes were corrected by the coefficient for atmospheric pressure and air temperature:

$$
k = \left(\frac{P_{\rm c}}{P_{\rm s}}\right) \left[\frac{273.16}{(273.16 + T_{\rm c})}\right],\tag{2}
$$

where k represents the coefficient, P_c is the air pressure in the chamber, P_s is the criterion barometric pressure (1013.25 h Pa), and T_c is the temperature $(^{\circ}C)$ in the chamber.

Results and discussion

Seasonal variations of soil surface $CO₂$ fluxes

Wetland soil-surface $CO₂$ fluxes exhibited a distinct seasonal pattern from July to October in 2001 (Figure 1). From the beginning of July, soilsurface $CO₂$ fluxes increased gradually until the middle of August when it reached the maximum value, and then soil-surface $CO₂$ fluxes decreased gradually until the beginning of October. The

Figure 2. Seasonal pattern of 5 cm soil temperature of D. angustifolia wetland.

Figure 3. Daily variation of soil-surface $CO₂$ fluxes of D. angustifolia wetland.

range of soil-surface $CO₂$ fluxes during the growing season was $0.250-1.265$ g m⁻² h⁻¹, with a mean value of 0.619 g m⁻² h⁻¹. This pattern was similar to the seasonal soil temperature change at 5 cm depth (Figure 2). Soil temperature reached its maximum value in July, and then decreased gradually until the beginning of October. However, soil $CO₂$ exchange is a complicated biological process and is affected by several environmental factors. Soil temperature is only one of these factors, so the seasonal pattern between soil $CO₂$ flux and 5 cm soil temperature have several inconsistent points in Figures 1 and 2.

Daily variation of soil-surface $CO₂$ fluxes

Deyeuxia angustifolia dominated wetland soil-surface $CO₂$ fluxes exhibited a distinct daily pattern (Figure 3). Soil-surface $CO₂$ flux increased from 8:00 until 17:00, decreased gradually until 5:00 the next day, and then increased again. The range

Figure 4. Daily variation of 5 cm soil temperature D. angustifolia wetland.

of soil-surface $CO₂$ fluxes during the day was 0.208–0.329 g m⁻² h⁻¹, and the mean value was 0.271 g m⁻² h⁻¹.

In September, soil-surface $CO₂$ fluxes decreased steadily. The amount of soil-surface $CO₂$ exchange is small, but the atmosphere and soil temperature had a measurable daily variation due to the regional climate condition, so that the soil-surface $CO₂$ flux has a distinct daily pattern. Figure 4 is the daily variation pattern of 5 cm soil temperature on September 28–29. It is clear that the trend seen in Figures 3 and 4 is very similar. This indicates that the soil temperature had a strong effect on soilsurface $CO₂$ fluxes.

Flux–temperature relationships

Soil-surface $CO₂$ flux is a measure of soil respiration. Soil respiration is the sum of root respiration and heterotrophic decomposition of soil organic matter. This biological activity is strongly affected by temperature and moisture (Oberbauer et al. 1992; Zak et al. 1999) and respiration rates are positively correlated with soil temperature (Lloyd and Taylor 1994; Raich and Potter 1995; Davidson et al. 1998). However, temperature responses differ depending on temperature range and type of ecosystem (Lloyd and Taylor 1994; Kirschbaum 1995; Winkler et al. 1996). Root respiration and soil microbial respiration may also respond differently to variation in soil temperature (Boone et al. 1998; Fang and Moncrieff 2001). Some of the variability in apparent responses to temperature may be the result of confounding variation in soil water

	Flux	0 cm	5 cm	10 cm	15 cm	20 cm	40 cm
Flux	1.000	$0.605**$	$0.770**$	$0.812**$	$0.844**$	$0.886**$	$0.877**$
0 cm	0.605	1.000	$0.820**$	$0.782**$	$0.777**$	$0.736**$	$0.684**$
5 cm	0.770	0.820	1.000	$0.995**$	$0.968**$	$0.942**$	$0.944**$
10 cm	0.812	0.782	0.995	1.000	$0.975**$	$0.957**$	$0.963**$
15 cm	0.844	0.777	0.968	0.975	1.000	$0.979**$	$0.969**$
20 cm	0.886	0.736	0.942	0.957	0.979	1.000	$0.964**$
40 cm	0.877	0.684	0.944	0.963	0.969	0.964	1.000

Table 2. Correlation analysis between soil $CO₂$ flux and soil temperature of D. angustifolia wetland.

** Correlation is significant at the 0.01 level (two-tailed), $N = 22$.

Figure 5. Relationship between soil-surface $CO₂$ flux and 5 cm soil temperature.

content (Dorr and Munnich 1987; Bowden et al. 1998; Davidson et al. 1998).

We analyzed the relationship between soil CO2 flux and two important environmental factors, which are soil temperature and soil water content. At the same time we measured wetland soil $CO₂$ exchange, we investigated soil temperature at different depths (0, 5, 10, 15, 20, 40 cm). Because former studies primarily used the 5 cm soil temperature to identify the effects of soil temperature for soil $CO₂$ flux, we also selected 5 cm soil temperature for our analyses. The results of correlation analysis indicated that there are obvious positive correlations between soil temperature and soil $CO₂$ flux (Table 2, Figure 5). $CO₂$ flux increased exponentially with increasing soil temperature. The simulated equation is:

$$
y = 0.1882e^{0.0599x}
$$
 $(R^2 = 0.77),$ (3)

where y represents soil-surface CO_2 flux (g m⁻² h^{-1}), and x is 5 cm soil temperature. Because the measurements were not continuous during the growing season, there is lack of results where soil temperature equals about 15 \degree C at the beginning of July and September. But from the relationship, the positive correlation between soil-surface $CO₂$ flux and soil temperature is obvious. From the temperature versus $CO₂$ flux equation, we calculated a Q_{10} value (1.82).

In addition, soil moisture ranged from 32 to 40.1% with an average of 34.5%. We also calculated the correlation coefficient between soil-surface $CO₂$ flux and soil moisture; the resulting coefficient of 0.306 was not significant. This result is similar to other research results (Reiners 1968; Kucera and Kirkham 1971; Edwards and Harris 1977; Mathes and Schriefer 1985). That is to say, where soil water content is sufficient, it is not the limiting factor.

Conclusions

Data were collected between July and October, 2001 on the Sanjiang Plain in northeastern China. The results revealed that there were both seasonal fluctuations and daily changes in soil-surface $CO₂$ fluxes. The ranges of measured soil-surface $CO₂$ flux were 0.208–1.265 g CO_2 m⁻² h⁻¹ (a positive value represents efflux to the atmosphere). Soilsurface CO_2 fluxes averaged 0.620 g CO_2 m⁻² h⁻¹. Study of edaphic factors such as soil temperature and soil moisture of the *D. angustifolia* wetland showed that the flux-temperature relationships were significant $(R^2 = 0.77)$.

Acknowledgements

This work was carried out with financial assistance provided by the Knowledge Innovation Engineering Project of the Chinese Academy of Sciences (Nos. KZCX2-302 and KZCX2-SW-320). The field observations were done in the Ecological Experiment Station of Mire-Wetland in the Sanjiang Plain, Chinese Academy of Science, which belongs to one of the fundamental stations of the Chinese Ecosystem Research Network (CERN). We give our special appreciation to all those who assisted with field work. The authors would also like to express their gratitude to associate professor Liu Hongyu and anonymous reviewers for criticizing and improving this manuscript. Finally, we sincerely thank Dr John Day and Dr William Conner for their objective criticism and helpful suggestions.

References

- Andres R.J., Marland G. and Fung I. 1996. A 1×1 distribution of carbon dioxide emissions from fossil fuel consumption and cement manufacture, 1950–90. Global Biogeochemical Cycles $10.419 - 429$
- Baldocchi D.D. and Meyers T.P. 1991. Trace gas exchange above the floor of a deciduous forest: evaporation and $CO₂$ efflux. Journal of Geophysical Research 96: 7271–7285.
- Bowden R.D., Nadelhoffer K.J. and Boone R.D. 1993. Contributions of aboveground litter, belowground litter, and root respiration to total soil respiration in a temperate mixed hardwood forest. Canadian Journal of Forest Research 23: 1402–1407.
- Bowden R.D., Newkirk K.M. and Rullo G.M. 1998. Carbon dioxide and methane fluxes by a forest soil under laboratorycontrolled moisture and temperature conditions. Soil Biology and Biochemistry 30: 1591–1597.
- Cropper W.P., Ewel K.C. and Raich J.W. 1985. The measurement of soil $CO₂$ evolution in situ. Pedobiologia 28: 34–40.
- Davidson E.A., Belk E. and Boone R.D. 1998. Soil water content and temperature as independent or confounded factors controlling soil respiration in a temperate mixed hardwood forest. Global Change Biology 4: 217–227.
- Dorr H. and Munnich K.O. 1987. Annual variation in soil respiration in selected areas of the temperate zone. Tellus Series B 39: 114–121.
- Edwards N.T. 1982. The use of soda-lime for measuring respiration rates in terrestrial systems. Pedobiologia 23: 321–330.
- Edwards N.T. and Harris W.F. 1977. Carbon cycling in a mixed deciduous forest floor. Ecology 58: 431–437.
- Ewel K.C., Cropper W.P. and Gholz H.L. 1987. Soil CO₂ evolution in Florida slash pine plantations: importance of root respiration. Canadian Journal of Forest Research 17: 330–333.
- Fang C. and Moncrieff J.B. 2001. The dependence of soil CO₂ flux on temperature. Soil Biology and Biochemistry 33: 155–165.
- Gupta S.R. and Singh J.S. 1981. Soil respiration in a tropical grassland. Soil Biology and Biochemistry 13: 261–268.
- Hall A.J., Connor D.J. and Whitfield D.M. 1990. Root respiration during grain filling in sunflower: the effects of water stress. Plant and Soil 121: 57–66.
- Hungate B.A., Holland E.A. and Jackson R.B. 1997. On the fate of carbon in grasslands under carbon dioxide enrichment. Nature 399: 576–579.
- Jensen L.S., Mueller T. and Tate K.R. 1996. Soil surface $CO₂$ flux as an index of soil respiration in situ: a comparison of two chamber methods. Soil Biology and Biochemistry 28: 1297–1306.
- Jong E., Redmann R.E. and Ripley E.A. 1979. A comparison of methods to measure soil respiration. Soil Science 118: 233–237.
- Kirschbaum M.U. 1995. The temperature dependence of soil organic matter decomposition, and the effect of global warming on soil organic C storage. Soil Biology and Biochemistry 27: 753–760.
- Kucera C.L. and Kirkham D.R. 1971. Soil respiration studies in tallgrass prairie in Missouri. Ecology 52(5): 912–915.
- Lloyd J. and Taylor J.A. 1994. On the temperature dependence of soil respiration. Functional Ecology 8: 315–323.
- Lu X., He Y. and Yang Q. 1995. Wetland carbon cycles and its effect to global change. In: Study on Chinese Wetland, Jilin Science and Technology Press, Changchun, pp. 68–72 (in Chinese).
- Luo Y., Jackson R.B. and Field C.B. 1996. Elevated $CO₂$ increases belowground respiration in California grasslands. Oecologia 108: 130–137.
- Ma X.-H., Lu X.-G. et al. 1996. A brief recount of $CO₂$ flow of mire ecosystems. In: Study on Marsh in the Sanjiang Plain, Science Press, Beijing, China, pp. 146–151 (in Chinese).
- Marra J.L. and Edmonds R.L. 1996. Coarse woody debris and soil respiration in a clearcut on the Olympic Peninsula, Washington, USA. Canadian Journal of Forest Research 26: 1337–1345.
- Mathes K. and Schriefer T. 1985. Soil respiration during secondary succession influence of temperature and moisture. Soil Biology and Biochemistry 17(2): 205–211.
- Nay S.M., Mattson K.G. and Bormann B.T. 1994. Biases of chamber methods for measuring soil $CO₂$ efflux demonstrated with a laboratory apparatus. Ecology 75: 2460–2463.
- Norman J.M., Garcia R. and Verma S.B. 1992. Soil surface $CO₂$ fluxes and the carbon budget of a grassland. Journal of Geophysical Research 97: 18845–18853.
- Oberbauer S.F., Gillespie C.T. and Cheng W. 1992. Environmental effects on $CO₂$ efflux from riparian tundra in the northern foothills of the Brooks Range, Alaska, USA. Oecologia 92: 568–577.
- Raich J.W. and Potter C.S. 1995. Global patterns of carbon dioxide emissions from soils. Global Biogeochemical Cycles $9.23 - 36$
- Raich J.W. and Schlesinger W.H. 1992. The global carbon dioxide flux in soil respiration and its relationship to vegetation and climate. Tellus 44 Series B: 81–99.
- Reiners W.A. 1968. Carbon dioxide evolution from the floor of three Minnesota forests. Ecology 49: 47–483.
- Reinke J.J., Adriano D.C. and McLeod K.W. 1981. Effects of litter alteration on carbon dioxide evolution from a South Carolina pine forest floor. Soil Science Society of America $45.620 - 623$
- Rochette P., Gregorich E.G. and Desjardins R.L. 1992. Comparison of static and dynamic closed chambers for

measurement of soil respiration under field conditions. Canadian Journal of soil Science 72: 605–609.

- Singh S.P., Mer G.S. and Ralhan P.K. 1988. Carbon balance for a central Himalayan cropfield soil. Pedobiologia 32: 187–191.
- Tans P.P. and Balkwin P.S. 1995. Climate change and $CO₂$ forever. Ambio 24: 376–378.
- Toland D.E. and Zak D.R. 1994. Seasonal patterns of soil respiration in intact and clear-cut northern hardwood forests. Canadian Journal of Forest Research 24: 1711–1716.
- Verma S.B.,Kim J. andClementR.J. 1992.Momentum, water vapor and carbon dioxide exchange at a centrally located prairie site during FIFE. Journal of Geophysical Research 97: 18629–18639.
- Winkler J.P., Cherry R.S. and Schlesinger W.H. 1996. The Q_{10} relationship of microbial respiration in a temperate forest soil. Soil Biology and Biochemistry 28: 1067–1072.
- Yang P. and Du B.-H. 1996. Overseas study trends of soil CO2 efflux. Chinese Journal of Agricultural Meteorology 17(1): 8–50 (in Chinese).
- Yang Q. and Lu X. 1996. Kinds of soil and their characteristics in the ecological experimental area of mire-wetlands. In: Chen G. (ed.), Research on Swamp in the Sanjiang Plain, Sciences Press, Beijing, pp. 15–26 (in Chinese).
- Yang Q. and Lu X. 1999. Primary exploration to soil respiration dynamic change of wetland ecosystem in the Sanjiang Plain. Chinese Journal of Soil Science 30(6): 254–256 (in Chinese).
- Yang Y. 1989. An approach to problems of formation and development of marsh in the Sanjiang Plain. In: Study of Mires in China, Science Press, Beijing, pp. 73–80 (in Chinese).
- Zak D.R., Holmes W.E. and MacDonald N.W. 1999. Soil temperature, matric potential, and kinetics of microbial respiration and nitrogen mineralization. Soil Science Society of America Journal 63: 575–584.
- Boone R.D., Nadelhoffer K.J. and Canary J.D. 1998. Roots exert a strong influence on the temperature sensitivity of soil respiration. Nature 396(6711): 570–572.