



Gastrointestinal parasites in red-legged partridges (*Alectoris rufa*) hunted in Spain: a warning to game managers

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Abstract

Red-legged partridge (*Alectoris rufa*) populations are currently declining in the Iberian Peninsula, mainly due to habitat degradation and hunting pressure. In addition, the release of farm-reared partridges may introduce pathogens, including parasites, to wild populations. The presence of digestive parasites in red-legged partridges hunted in fifteen Spanish provinces was studied. Fecal samples and gastrointestinal tracts were collected, analyzed, and the morphometric identification of parasites was carried out. *Eimeria* spp. oocysts, nematode, cestode and trematode eggs were observed in fecal samples. Adult nematodes (*Ascaridia galli*, *Ascaridia compar*, *Heterakis gallinarum*, *Heterakis tenuicauda*, *Trichostrongylus tenuis*, *Subulura* spp., *Cyrtus* spp. and *Aonchotheca caudinflata*), tapeworms (*Raillietina tetragona*, *R. echinobothrida*, *R. micracantha*, *Rhabdometra nigropunctata*, and *Choanotaenia infundibulum*), and trematodes (*Brachylaima* spp., *Brachylecithum* spp., *Dicrocoelium* spp.) were identified in the gastrointestinal tracts. Significant statistical differences were found among climatic regions in the prevalence and intensity of *Eimeria* spp. infection, median intensity and the prevalence of indirect life cycle helminths, with Southern areas always showing higher infection values. The study provides information of the health status of red-legged partridges in Spain, highlighting the risk associated with the release of farm-reared partridges for restocking purposes. This should be taken into account to improve management strategies for the long-term conservation of the species.

Keywords Bioclimatic region · *Eimeria* · Helminths · Management · Parasites · Red-legged partridge

Introduction

The red-legged partridge (*Alectoris rufa*) is a medium-size European galliform, with natural populations reported in Portugal, Andorra, France, Germany, Italy and Spain (Blanco-Aguiar et al. 2003; Birdlife International 2022). This species carries out an important ecological role in Mediterranean ecosystems (Díaz-Fernández et al. 2013), with a wide distribution in the Iberian Peninsula ranging from natural environments to farmland mosaics (Cabodevilla et al. 2021). In addition, the red-legged partridge is the principal prey for a large number of threatened species

(Blanco-Aguiar et al. 2004; Arroyo et al. 2017). Also, the red-legged partridge has a significant socio-economic value as the main game bird species, with around 3 to 5 million of farm-reared partridges annually released in Spain for shooting (Blanco-Aguiar et al. 2003; Sánchez García-Abad et al. 2009).

During the last decade, red-legged partridge populations have declined by 40–45% in Europe (Birdlife International 2022). Agricultural intensification, with negative effects on suitable habitats for this galliform, is one of the main threats to its survival (Delibes-Mateos et al. 2012; Cabodevilla et al. 2021). On the other hand, overhunting, hybridization, predation and diseases are factors that have contributed to the reduction of red-legged partridge populations in the Iberian Peninsula (Calvete et al. 2003; Blanco-Aguiar et al. 2008; Buenestado et al. 2008, 2009; Villanúa et al. 2008; Casas and Viñuela 2010).

The main objective of hunting estate managers is to raise the availability of these birds and, therefore, populations are usually restocked with farm-reared partridges (Díaz-Fernández

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et al. 2013; Casas et al. 2016). Also, predator control or habitat management measures (water points, supplementary feeders or game crops) are frequently used to maintain or increase partridge population density, particularly in areas where hunting provides significant economic benefits (Gortázar et al. 2000; Arroyo et al. 2012). Specifically, water points, supplementary feeders or game crops are some of the measures used to improve the survival of released farm-reared individuals and to achieve their permanent settlement on hunting estates (Gortázar et al. 2000).

Previous studies have pointed out the differences between the parasite community of farm-reared and wild partridges (Millán et al. 2004b). Farm-reared partridges are usually infected by monoxenous nematodes such as *Ascaridia* spp., *Heterakis* spp. or *Aonchoteca caudinflata*. In case of wild individuals, heteroxenous parasites (*Cyrnea* spp., *Subulura* spp., flukes or tapeworms) are the most predominant (Millán 2009). Therefore, one of the negative effects of releasing farm-reared birds is an eventual introduction and spread of diseases to wild red-legged partridge populations. In addition, hybridization with other partridge species represent a threat for the long-term survival of this native species (Villanúa et al. 2007; Jamieson and Lacy 2012; Sánchez-Donoso et al. 2012).

Environment and climatic conditions could influence the life cycle of parasite species, and so, their ecology, survival and spread, impacting also on the host populations (Morand 2015; Brunner and Eizaguirre 2016; Holand et al. 2019). In this sense, the Iberian Peninsula has different bioclimatic zones, being Mediterranean and Atlantic areas the main bioclimatic subdivision. Mediterranean climate covers territories with warm, dry summers and cool, wet winters, while the Atlantic area is wetter and colder (Rey Benayas and Scheiner 2002). This difference in climate defines the host (Sillero et al. 2009) and parasite diversity (Sanchis-Monsonís et al. 2019).

The aim of this study was to characterize the parasites of the gastrointestinal tract of red-legged partridges from several hunting estates distributed along the Iberian Peninsula, as well as to determine the influence of some biotic (sex and age of the host) and abiotic (geographical area of provenance) characteristics on the parasite community. In addition, the relationship between the characteristics of each climatic region and the parasite richness was discussed, as well as the risks related to the release of farm-reared partridges. Our results may help to design new management recommendations to preserve natural red-legged partridge populations.

Material & methods

Study area and animal collection

During the winters of 2010–2013, 934 red-legged partridges shot in 139 hunting estates from 15 out of the 50 Spanish

provinces were collected. The hunting estates sampled regularly restocking with farm-raised red-legged partridges, although it was not possible to obtain detailed information about the frequency of these releases. At the time of sampling, it was not possible to differentiate whether the partridges were wild or farm-reared, because the latter were not ringed. The environmental heterogeneity of the Iberian Peninsula leads to great differences between the northern area and the Mediterranean one, which occupies most of the central, eastern and southern part of the country (Olson et al. 2001). In this sense, the area of study was classified as Humid Temperate Atlantic (average annual precipitation: 1000–1600 mm; average annual temperature: 7–12°C), Subhumid Supramediterranean (600–1000 mm; 8–13°C), Semiarid-Mesomediterranean (350–600 mm; 13–17°C) and Thermomediterranean (200–350 mm; 17–19°C) (Rivas-Martínez et al. 2017).

Red-legged partridges were categorized by sex and age following the descriptions of Sáenz de Buruaga et al. (1991). The distribution of the sample by sex, age and bioclimatic zone is shown in Table 1 and Fig. 1.

The gastrointestinal tract (including proventriculus, gizzard, small intestine, caeca, colon and cloaca) of 547 birds, and only the large intestine and cloaca from the remaining partridges ($n = 387$) were extracted by the hunters, individually refrigerated in plastic labelled bags and submitted to the Department of Animal Health at Murcia University, where they were frozen at -20°C until they were analysed, which was always less than two weeks after receipt of the samples.

Laboratory procedures

Coprological analysis

Of all submitted samples that contained feces in the cloaca ($n = 927$), coprological analysis was performed with both centrifugation-flotation and sedimentation techniques (MAFF 1986), using Sheather's solution ($\rho = 1.27 \text{ g/ml}$) as flotation fluid. Two-chamber McMaster slides were filled in triplicate and the average oocyst/egg count per gram of faeces (OPG and EPG, respectively) was calculated. Oocysts and helminth eggs detected by coprology were morphometrically identified according to Soulsby (1987) and Naciri et al. (2011).

Necropsy

Once faecal samples were collected from the cloaca, the proventriculus, gizzard, small and large intestines of partridges from whom the gastrointestinal tract had been collected ($n = 547$), were longitudinally opened and directly examined under a stereomicroscope for the presence of helminths. When samples included the liver (40/547), this

Table 1 Distribution of red-legged partridges ($n=934$) attending to the sex and age of the host, as well as climate area and province of origin

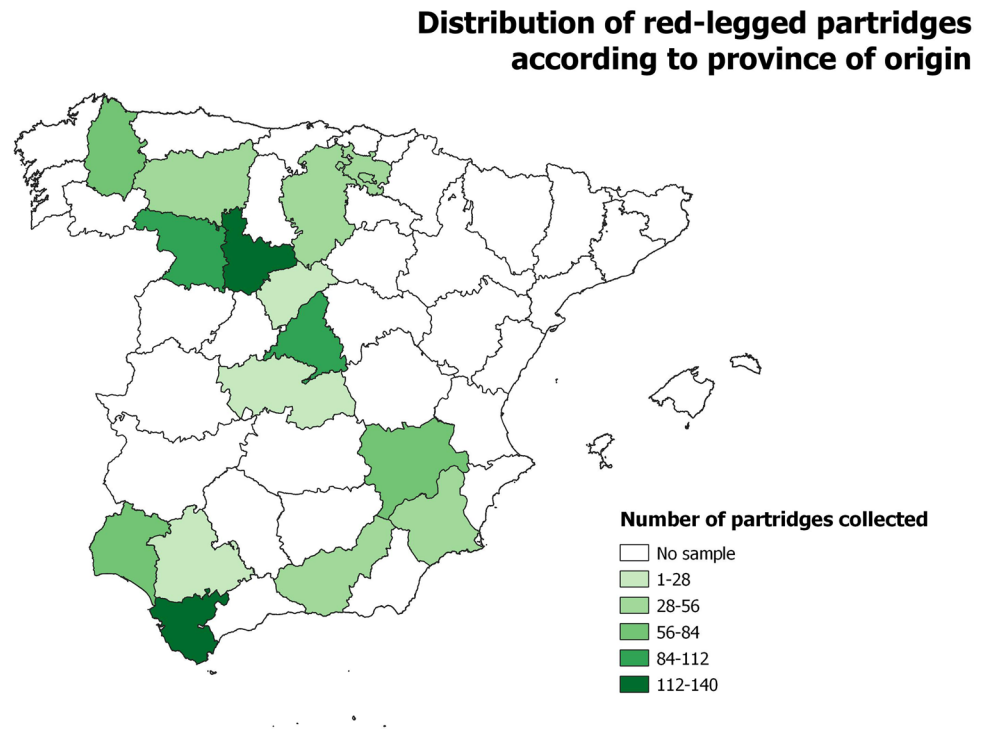
Climate region ^(a)	Province	N ^(c)	Sex			Age category			
			Male	Female	NA ^(b)	Juvenile	Subadult	Adult	NA
HTA	Lugo	63	11	36	16	0	3	44	16
	Álava	40	0	0	40	0	0	0	40
SS	Segovia	6	5	1	0	1	0	5	0
	Burgos	49	1	3	45	3	0	1	45
	Valladolid	140	44	21	75	33	15	17	75
SM	León	53	8	5	40	3	4	6	40
	Toledo	25	6	4	15	4	1	5	15
	Madrid	87	11	13	63	10	2	12	63
	Zamora	91	56	35	0	22	15	54	0
	Albacete	70	27	24	19	21	0	30	19
T	Granada	38	0	3	35	2	0	1	35
	Murcia	56	26	28	2	17	32	5	2
	Huelva	81	0	0	81	0	0	0	81
	Cádiz	118	65	44	9	7	1	102	8
	Sevilla	17	0	0	17	0	0	0	17
Total		934	260	217	457	123	73	282	456

(a) *HTA* Humid Temperate Atlantic, *SS* Subhumid Supramediterranean, *SM* Semiarid-Mesomediterranean, *T* Thermomediterranean

(b) NA: data not available

(c) N: number of red-legged partridges

Fig. 1 Number of red-legged partridges collected and their distribution according to the province of provenance



organ was cut into slices and washed with tap water over a filter to collect any trematodes that might be present. All isolated adult helminths were preserved in 70% ethanol until identification. Adult nematodes were cleared in

lactophenol and morphometrically identified according to Yamaguti (1961) and Anderson et al. (1974). Tapeworms and trematodes were stained using Semichon’s carmine (Schmidt 1986), mounted in DPX and identified according

to the descriptions given by Schmidt (1986) and Yamaguti (1971).

Statistical analysis

Prevalence (P) with 95% confidence intervals (95% CI), median abundance (MA) and median intensity (MI) with the range of detected parasites were determined according to Margolis et al. (1982) and Bush et al. (1997). Median abundance or median intensity data were calculated depending on whether all the sampled animals or only the positive ones were included in the statistical analysis, respectively. Species richness (number of parasite species in each host population) was also determined for the different climatic regions. Shapiro Wilks test was used to determine normality of parasite distributions. Fisher or Chi-square tests and the Kruskal–Wallis test were employed to compare the proportions of parasitised birds, and medians of intensity about parasite communities, respectively, according to host and environmental variables (sex, age category, bioclimatic zone). Additionally, analysis of multivariate abundance was carried out $\left[\log\left(\frac{y}{\min} + 1\right) \text{scale} \right]$ to evaluate the distribution of the parasite species in the different bioclimatic area (Wang et al. 2012). Only nematodes and trematodes were considered, as no data on cestode abundances were calculated, since in many cases the cestode fragments found lacked scolexes, so that the exact number of cestodes could not be calculated with certainty. Significant differences were considered for $p < 0.05$. R software 3.5.2 was used to analyse the data (R Core Team 2018).

Results

Parasites detected by coprological analysis and necropsy

The overall prevalence of infected hosts was 47.8% (446/934). The prevalence of parasites detected by coprological analysis was 44.8% (416/927). *Eimeria* spp. oocysts and helminth eggs shedding records are summarised in Table 2. The most prevalent parasitic forms were *Eimeria* spp. oocysts, followed by nematode eggs belonging to the Ascaridida and Trichurida (*Capillaria*-like) orders.

A total of 1623 helminths were collected by necropsy from 20.1% (110/547) of the red-legged partridges. Fifty-three of 547 (9.6%) partridges presented nematodes belonging to six different species and two genera, with MI (range) = 28.6 (1–239) nematodes and MA (range) = 2.7 (0–239) nematodes. Similarly, 60/547 (11%) partridges had cestodes and only 12 of the 40 individuals whose liver was available (30%) were parasitized by trematodes, with

Table 2 Prevalence (P), 95% confidence intervals (95%CI), median intensity (MI) and range (expressed as oocyst or eggs per gram of faeces) of parasites found by coprological analysis ($n = 927$)

	N	N + ve ^(a)	P (95%CI)	MI (range)
<i>Eimeria</i> oocysts	926	393	42.4 (39.2–45.6)	1970 (10–627,642)
Nematode eggs	927	46	4.9 (3.6–6.4)	518 (1–20,980)
Cestode eggs	570	3	0.53 (-0.07–1.12)	1 (1–1)
Trematode eggs	926	20	2.1 (1.2–3.1)	79 (1–632)

(a) N + ve: number of positive partridges

MI = 9 (1–44) and MA = 0.2 (0–44) trematodes per host. Regarding co-infections, 76.3% (84/110) of the parasitized partridges had only one helminth species, while multiparasitism with two, three and four different species was found in 19 (17.2%), 6 (5.4%) and one (0.9%) bird, respectively. The overall parasite richness found was at least 16 helminth species, considering that some parasite specimens could only be identified down to the genus level because of deterioration or because they were immature stages. All these results are summarized in Table 3.

Biotic and abiotic risk factors

Although sex and age data were not always available, using the existing registries, the influence of different biotic (sex and age) and abiotic (bioclimatic zone of birds' origin) on prevalence and intensity of parasites detected by coprological analysis and necropsy are summarised in Tables 4 and 5, respectively.

Sub-adult partridges showed significantly higher prevalence of trematode eggs than the other age groups. Regarding helminth detection by necropsy, males presented higher intensity of nematodes than females, whereas cestode prevalence was, by contrast, significantly higher in females. Finally, regarding the bioclimatic zones, the prevalence of partridges with *Eimeria* spp. oocyst and median intensity of oocyst, as well as the prevalence of trematode eggs, cestodes and nematodes (except Ascaridida order), were significantly higher in the Thermomediterranean area. On the contrary, the prevalence of Ascaridida helminths found by necropsy, as well as Trichurida and Ascaridida eggs detected by coprological techniques, were significantly higher in the Humid Temperate Atlantic area (Tables 4 and 5), as is the case of *A. galli* abundances. Specifically, 46 birds were collected from this bioclimatic zone, and six of them had these nematodes, so prevalence is higher compared to that found in the Semiarid-Mesomediterranean area (8/243) or in the Thermomediterranean area (14/172). On the other side, regarding Thermomediterranean area, only *T. tenuis* abundances were statistically significant ($p < 0.05$).

Table 3 Parasite prevalence (P), 95% confidence intervals (95%CI), median intensity (MI) and range of helminths isolated from red-legged partridges by necropsy

	N + ve ^(a)	P (95%CI)	MI (range)
Nematodes (N=547)			
<i>Ascaridia galli</i>	11	2 (0.83–3.19)	7 (1–28)
<i>Ascaridia compar</i>	7	1.2 (0.34–2.22)	15 (1–46)
<i>Heterakis tenuicauda</i>	15	2.7 (1.37–4.11)	8 (1–152)
<i>Heterakis gallinarum</i>	8	1.4 (0.46–2.47)	1.5 (1–19)
<i>Trichostrongylus tenuis</i>	18	3.3 (1.79–4.78)	7.5 (1–239)
<i>Aonchotheca caudinflata</i>	2	0.36 (-1.14–1.87)	82 (1–163)
<i>Subulura</i> spp.	8	1.4 (0.46–2.47)	5.5 (1–55)
<i>Cyrnea</i> spp.	3	0.55 (-0.07–1.17)	2 (1–3)
Cestodes (N=537)			
<i>Raillietina</i> spp.	17	3.1 (1.68–4.65)	NA
<i>Raillietina tetragona</i>	19	3.5 (1.97–5.10)	NA
<i>Raillietina echinobotridia</i>	4	0.74 (0.02–1.47)	NA
<i>Raillietina micracantha</i>	15	2.7 (1.40–4.19)	NA
<i>Rhabdometra nigropunctata</i>	3	0.56 (-0.07–1.19)	NA
<i>Choanotaenia infundibulum</i>	1	0.19 (-0.18–0.55)	NA
Trematodes (N=40)			
<i>Dicrocoelium</i> spp.	1	2.5 (-2.34–7.34)	1 (1–1)
<i>Brachylaima</i> spp.	1	2.5 (-2.34–7.34)	18 (18–18)
<i>Brachylecithum</i> spp.	10	25 (11.58–38.42)	4 (1–44)

(a) N + ve: number of positive partridges

In general terms, the mean abundance of nematodes and trematodes was statistically significant higher in southern areas (Semiarid-Mesomediterranean and Thermomediterranean areas) ($p < 0.05$) (Fig. 2). Likewise, the parasite richness was greater in the Semiarid-Mesomediterranean (13 species) and Thermomediterranean (15 species) regions, where the richness of nematodes represents around half of the species. The parasitic species found in all the areas, was *Eimeria* spp. as shown in Table 6.

Discussion

The results of this epidemiological investigation show a rich parasite community in Spanish red-legged populations, with most of the helminth species found having been previously described in this host. Considering the detection of parasites by coprological analysis and/or necropsy, the overall parasite prevalence found was 47.8%.

Focusing on the results obtained by coprological analysis, the overall prevalence was 44.8% (416/927), similar to the

38% previously described by Millán et al. (2003) in captive and wild red-legged partridges. Regarding the prevalence of helminths found by necropsy, it was 20.1% (110/547), a low value when compared to the prevalence of 69% (161/235) reported by Calvete et al. (2004) in red-legged partridges from two provinces of central Spain (Toledo and Ciudad Real), both with an important tradition of game bird hunting. These differences may be due to factors as the density of birds at the sampled site, land use characteristics as suggested by Calvete et al. (2004), or climatic features that influence on the survival of infective parasite stages and/or the distribution of their intermediate and paratenic hosts.

The prevalence of trematode infections was low when compared to the studies previously mentioned (Calvete et al. 2004). In our study, the most prevalent trematode detected by necropsy was *Brachylecithum* spp., a liver fluke that has been previously detected in red-legged partridges (Millán 2009). *Dicrocoelium* spp. was detected in only 2.5% of necropsied livers, while it was the most prevalent genus in hunted wild red-legged partridges studied by Millán et al. (2004a) and Calvete et al. (2003), who described prevalences of 46.8% and 17.3%, respectively. The low *Dicrocoelium* spp. prevalence that we found is probably due to the small number of livers available for our study. In addition, although we detected eggs of trematodes by coprological analysis, we are probably underestimating the prevalence of flukes, since the sensitivity of coprological techniques is lower than the detection of trematode specimens by necropsy, especially in the case of liver flukes (Sithithaworn et al. 1991; Sobral et al. 2019). The prevalence of *Brachylaima* spp. was 2.5%, similar to the findings described by Millán et al. (2004a), who considered these digestive flukes as anecdotal.

Among the helminths detected by necropsy, the prevalence of cestodes was the highest, especially in the case of the genus *Raillietina* spp. (3.1%) and, in particular, to *R. tetragona* and *R. micracantha* (3.5% and 2.7%, respectively). *R. tetragona* was also the most prevalent cestode in the study carried out by Calvete et al. (2003) and Millán et al. (2004a). *Rhabdometra nigropunctata* (4/538) and *Choanotaenia infundibulum* (1/538) prevalence was lower than 1%. These tapeworm species, as well as *R. tetragona*, were detected in wild red-legged partridges from the Iberian Peninsula in previous studies (Calvete et al. 2003; Millán et al. 2004a). Although the prevalence of adult tapeworms by necropsy was 11%, the prevalence of cestodes in partridges was much lower (1%) by coprological analysis, indicating a low diagnostic sensitivity of this last technique, as has been demonstrated by other authors (Zloch et al. 2021).

These results on trematode and cestode prevalence suggest that the parasitized partridges were either wild individuals or, alternatively, farm-reared birds released in the months prior to hunting that, during that time, acquired the infection in the wild (Villanúa et al. 2008). In this regard, it

Table 4 Influence of biotic and abiotic risk factors on prevalence and median of parasites found by coprological techniques

Variable	Level	N ^(a)	Infected	<i>Eimeria</i> spp.			Nematode eggs			Cestode eggs			Trematode eggs		
				P (95%CI) ^a		MI (range) ^(b)	P (95%CI)	MI (range)	P (95%CI)	MI (range)	P (95%CI)	MI (range)	P (95%CI)	MI (range)	
Sex	Female	217	106	49 (42.2–55.5)	45 (38.1–51.3)	1606 (26–627,642)	6 (2.8–9.1)	212 (1–1267)	0.46 (-0.4–1.4)	1 (1–1)	3 (0.6–4.9)	40 (1–632)			
	Male	260	116	45 (38.6–50.6)	42 (35.5–47.5)	1605 (11–550,964)	5 (2.1–7.2)	564 (30–12,281)	0.38 (-0.4–1.1)	1 (1–1)	4 (1.2–5.7)	34 (1–193)			
Age	Juvenile	123	61	50 (40.7–58.4)	50 (39.9–57.6)	1459 (26–550,964)	4 (0.6–7.5)	949 (38–12,281)	0.8 (-0.7–2.4)	1 (1–1)	2 (-0.6–3.9)	108 (23–193)			
	Subadult	73	28	38 (27.2–49.5)	38 (25.9–48.1)	1970 (61–392,733)	3 (-1–6.5)	200 (1–400)	1.4 (-1.3–4)	1 (1–1)	8* (1.9–14.5)	37 (17–632)			
Climate region ^(c)	Adult	282	134	47.5 (41.7–53.3)	43 (36.4–47.9)	1218 (11–627,642)	6 (3.5–9.2)	193 (50–5537)	0.35 (-0.3–1)	1 (1–1)	3 (0.7–4.3)	40 (1–167)			
	HTA	103	37	36 (26.6–45.2)	27 (18.6–35.8)	2692 (42–442,550)	11* (4.7–16.6)	417 (20–2,020,983)	0	0	0	0			
	SS	248	84	34 (27.9–39.7)	32 (26.1–37.6)	2165 (10–550,964)	3 (1–5.4)	471 (1–8942)	0	0	2 (0.3–3.8)	165 (17–188)			
	SM	311	123	39.5 (34.1–44.9)	39 (33.8–44.6)	657 (25–277,333)	2 (0.4–3.4)	558 (278–1111)	0	0	1 (-0.1–2)	1 (1–1)			
	T	272	173	64* (57.9–69.3)	61* (54.5–66.1)	4416 (11–627,642)*	8 (4.5–10.9)	503 (21–12,281)	1 (-0.1–2.3)	1 (1–1)	4* (2–6.8)	113 (21–632)			

(a) N: Total number of partridges belonging to this variable level. Prevalence (P) and 95% confidence intervals

(b) MI (median intensity) and range (minimum and maximum values) of parasites in red-legged partridges

(c) HTA Humid Temperate Atlantic, SS Subhumid Supramediterranean, SM Semiarid-Mesomediterranean, T Thermomediterranean

(*) Significant differences between levels of the variable. Asterisk placed in level with highest value

Table 5 Influence of biotic and abiotic risk factors on prevalence and median of parasites found in necropsies

Variable	Level	N ^(a)	Infected	Nematodes ^(d)			Cestodes			Trematodes ^(e)			Ascaridida		
				P (95%CI) ^(b)	MI (range) ^(b)	P (95%CI)	MI (range)	P (95%CI)	MI (range)	P (95%CI)	MI (range)	P (95%CI)	MI (range)		
Sex	Female	178	38	21 (15.3–27.4)	5 (1–46)	10 (5.2–13.9)	NA	14* (8.9–19.1)	1 (-0.4–2.7)	3 (1–5)	6 (2.6–9.7)	5 (1–46)			
	Male	228	32	14 (9.5–18.5)	16 (1–59)*	9 (5.1–12.4)	NA	7 (3.7–10.3)	1 (-0.2–2.8)	18 (1–18)	5 (2.4–8.2)	22 (1–152)			
Age	Juvenile	101	16	16 (8.7–22.9)	2 (1–188)	9 (3.3–14.7)	NA	12 (5.6–18.2)	0	0	7 (1.9–11.9)	1 (1–25)			
	Subadult	39	5	13 (2.3–23.3)	1 (1–1)	3 (-2.4–7.5)	NA	5 (-1.8–12)	5 (-1.8–12)	11.5 (5–18)	0	0			
Climate region ^(c)	Adult	267	50	19 (14–23.4)	12 (1–152)	10 (6.5–13.7)	NA	11 (6.8–14.2)	1 (-0.1–2.4)	1 (1–18)	6 (3.1–8.8)	14 (1–152)			
	HTA	46	7	15 (4.8–25.6)	9 (1–29)	15 (4.8–25.6)	NA	0	0	0	13* (3.3–22.8)	14 (1–29)			
	SS	86	7	8 (2.4–13.9)	55 (1–56)	3 (-0.4–7.4)	NA	6 (0.9–10.7)	0	0	2 (-0.8–5.5)	28.5 (1–56)			
	SM	243	29	12 (7.8–16)	7 (1–26)	5 (1.9–7.1)	NA	5 (1.9–7.1)	4 (1.3–6)	5 (1–44)	3 (1–5.5)	7.5 (1–22)			
	T	172	63	37* (29.4–43.8)	10 (1–239)	19* (12.8–24.4)	NA	27* (19.1–32.1)	2 (-0.2–3.7)	1 (1–18)	8 (4–12.2)	9.5 (1–152)			

(a) N: Total number of partridges belonging to this variable level. Prevalence (P) and 95% confidence intervals

(b) Median intensity and range (minimum and maximum values) of parasites in red-legged partridges

(c) HTA Humid Temperate Atlantic, SS Subhumid Supramediterranean, SM Semiarid-Mesomediterranean, T Thermomediterranean

(d) Except Ascaridida

(e) These results were obtained from 40 individuals

(*) Significant differences between levels of the variable

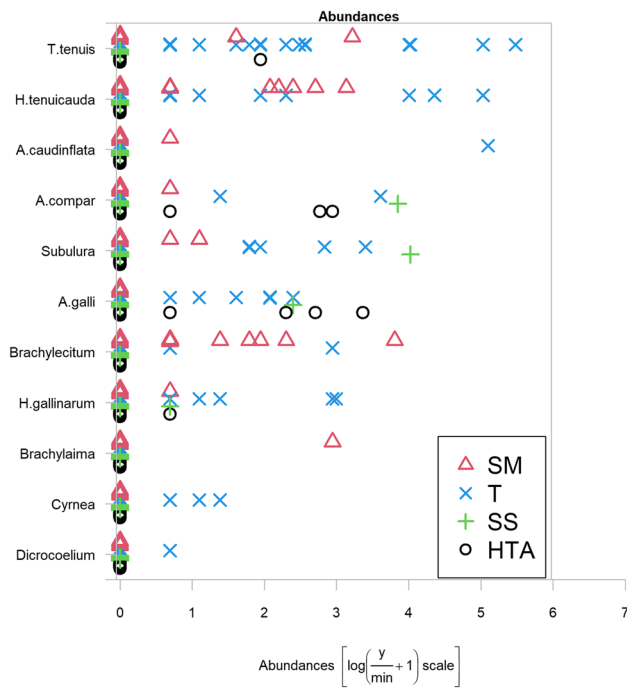


Fig. 2 Abundances of nematode and trematode species found in red-legged partridges according to the climatic region of origin

is noteworthy that 25–34% of the released red-legged partridges died during the first 72 h post-release and that, in general, survival rate is very low due to predation, hunting and starvation (Gortázar et al. 2000; Pérez et al. 2004).

Ascaridida was the most prevalent nematode group (7.5% of animals), including *Heterakis* spp. (4.2%) and *Ascaridia* spp. (3.3%), nematodes that have been described with low prevalences in red-legged partridges from Spain and Italy (Millán 2009; Polello et al. 2021).

Trichostrongylus tenuis was recovered from 3.3% of the red-legged partridges necropsied, while other studies have reported prevalences of 12–13% (Calvete et al. 2004; Millán et al. 2004a). It is a parasite that, despite having a direct life cycle, is considered more prevalent in wild than in farm-reared partridges, because it is unable to complete its life cycle when there is little vegetation in the environment, as happens in breeding farms (Millán et al. 2004a). In case of indirect life cycle nematodes as *Aonchotheca caudinflata*, *Cyrnea* spp. and *Subulura* spp., the prevalence detected was less than 1%. In other studies, similar values of prevalence from wild specimens have been reported: 1.8% (Calvete et al. 2003, 2004; Millan et al. 2004a). Hence, as noted above, these parasites could have been acquired by wild individuals or partridges that had lived in the wild during the first periods of life.

Ecological determinants and anthropogenic factors, such as the type of management implemented, influence the prevalence, intensity and parasite richness in game species

Table 6 Red-legged partridge parasite richness according to climate region provenance

Parasite species	Presence/absence ^(a) of species in each climate region ^(b)			
	HTA	SS	SM	T
<i>Eimeria</i> spp.	+	+	+	+
Nematodes <i>A. galli</i>	+	+	-	+
<i>A. compar</i>	+	+	+	+
<i>H. gallinarum</i>	+	+	+	+
<i>H. tenuicauda</i>	-	-	+	+
<i>T. tenuis</i>	+	-	+	+
<i>A. caudinflata</i>	-	-	+	+
<i>Cyrnea</i> spp.	-	-	-	+
<i>Subulura</i> spp.	-	+	+	+
Richness	4	4	6	8
Cestodes <i>R. tetragona</i>	-	+	+	+
<i>R. echinobotridia</i>	-	+	+	+
<i>R. micracantha</i>	-	+	+	+
<i>R. nigropunctata</i>	-	+	+	-
<i>C. infundibulum</i>	-	-	-	+
Richness	0	4	4	4
Trematodes <i>Dicrocoelium</i> spp.	-	-	-	+
<i>Brachylaima</i> spp.	-	-	+	-
<i>Brachylecithum</i> spp.	-	-	+	+
Richness	0	0	2	2
Total species	5	9	13	15

(a) Presence (+) and absence (-)

(b) *HTA* Humid Temperate Atlantic, *SS* Subhumid Supramediterranean, *SM* Semiarid-Mesomediterranean, *T* Thermomediterranean

(Tompkins et al. 1999; Poulin 2007; González-Quevedo et al. 2014; Morand 2015; Fanelli et al. 2020). In our study, we have included red-legged partridges from 15 provinces, ranging from the north to the south of the Iberian Peninsula, so it can be assumed that the greater heterogeneity of the bioclimatic regions sampled, each with its own characteristics of land use and management of the species, has influenced our results.

Eimeria spp. was the most prevalent parasite in this study, as in previous surveys with other Galliformes species (Santilli and Bagliacca 2012; Globokar et al. 2017; Polello et al. 2021). *Eimeria* spp. and Ascaridida parasites are frequently found in farm-reared partridge populations (Bolognesi et al. 2006; Villanúa et al. 2008; Millán 2009; Naciri et al. 2011; Máca and Pavlásek 2020). In other game birds, such as pheasants, individuals from restocking areas have been found to have twice the prevalence of *Eimeria* spp. as those from non-repopulated areas, 51.3% and 25.6%, respectively (Santilli and Bagliacca 2012). In agreement, Mani et al. (2000) pointed out that wild

pheasant had lower parasite prevalence than those hunted in areas where farm-reared birds are frequently released. In fact, coccidiosis is one of the main problems in partridge farms, where the characteristics of the facilities provide conditions that help to spread infection (Naciri et al. 2011). On the other hand, Albendazole is usually administered in red-legged partridge farms to limit the infection with nematodes, mainly Trichurida and Ascaridida, although results reported by Villanúa et al. (2008) have demonstrated the limited efficacy of this drug, in particular with regard to *Heterakis* spp. This scenario coincides with our results since *Eimeria* spp. was the most prevalent genus detected with coprological techniques, whereas *Ascaridia* spp. and *Heterakis* spp. were the most prevalent helminths detected by necropsy. Moreover, according to the bioclimatic zone of partridge's origin, our results indicate that those birds coming from the Thermomediterranean area showed higher prevalence of *Eimeria* spp. (60.3%; 164/272) and higher number of oocysts shedding (considered as a proxy of median parasite intensity) than partridges from other bioclimatic zones. These results could be the consequence of more frequent restocking with farm-reared birds in hunting estates from the Thermomediterranean area. Hunting is an important tradition in Spain, being especially significant in centre and south areas (Viñuela et al. 2013). In fact, game crops are used as a tool to provide supplementary food, nesting cover or protection for game birds in Mediterranean areas, characterized by dry and hot summers (Reino et al. 2016). Due to the climatic characteristics of the Iberian Peninsula, annual periods of drought are common and, in order to palliate this, the set-up of water points in hunting areas is also a frequently used management measure (Gaudioso Lacasa et al. 2010). These types of practices, which has been widely applied for decades in the game management of partridges, leads to the establishment of hot spots for pathogen transmission, mostly in water stress periods. In this sense, Gaudioso Lacasa et al. (2010) observed that water points were used by both wild and game birds, especially in summer, when chicks need more requirements. As with supplementary feeding, these management measures promote parasite transmission and disease outbreaks due to higher contact among birds (Villanúa et al. 2006; Millán 2009). Therefore, both the frequent release of farm-reared partridges in Southern areas of Spain to cover the needs of the hunters, and the management practices used in these hunting estates, could determine the high *Eimeria* spp. prevalence and the highest presence of Ascaridida among all the helminths found in partridges coming from these areas.

Regarding biotic factors, cestode prevalence was significantly higher in female partridges, whilst the median intensity of nematodes was greater in males. On the other hand,

sub-adult partridges showed a significantly higher prevalence of trematode eggs in faeces. Other studies have shown differences in parasite intensity and prevalence depending on the sex and the age of the host in a broad range of species (Poulin 2007; Martínez-Guijosa et al. 2015). Several reasons could account for gender-related differences in parasite infections. For instance, testosterone production in males is energy demanding and has been linked to immunosuppression and increased probability of becoming parasitized (Klein 2004; Guerra-Silveira and Abad-Franch 2013). Other studies attributed differences in the risk of parasite infection between males and females to food preferences (Provencher et al. 2016). Immunosuppression and food preferences have also been associated to age-related differences in the prevalence of parasite infections (Thieltges et al. 2006).

Finally, considering parasite richness, a total of 16 species of helminths were described by necropsy in this study. These included eight species of nematodes, five of cestodes and three of trematodes (Table 3). The Thermomediterranean area was the one with highest richness of parasite species (15), followed by the Semiarid-Mesomediterranean (14), the Supramediterranean (8), and finally the Humid Temperate Atlantic area (4), which had the lower richness (Table 6). Brown (1984) suggested that abundance and distribution of helminths reflect the abundance and distribution of their hosts, and Calvete et al. (2003) propose that the distribution of parasites, in general, is related to the distribution of their intermediate and definitive hosts. In this sense, socio-economic interest around game birds in the southern Iberian Peninsula, where release of farm-reared partridges as well as management practices (use of game crops or water points) are more frequent than in other territories, could explain the results obtained about *Eimeria* spp. or Ascaridida infections in the Thermomediterranean area, as well as the higher values of the abundance of *T. tenuis* comparing with the other bioclimatic areas.

Moreover, the environmental characteristics of each area may also cause the parasite richness between areas to differ. In fact, results about multivariate abundance indicate statistically significant differences among bioclimatic zones, with Semiarid-Mesomediterranean and Thermomediterranean areas showing the highest rates of trematodes and nematodes, respectively. In this sense, the presence of indirect life cycle parasites and the high rate of parasite richness reached in these areas, could be due to the influence of environmental factors in this bioclimatic zone on parasites using intermediate and paratenic hosts in their life cycles (Krasnov and Poulin 2010; Morand 2015).

Climate has been related to the environmental persistence of parasite free stages, and also determines the abundance and distribution of intermediate and paratenic hosts (Holand et al. 2019). Coleoptera, Diptera, Hymenoptera and other invertebrate parasite hosts constitute an important part of

the partridge's diet (Holland et al. 2006); in particular, dung beetles play an important role in parasite transmission (Nichols and Gómez 2014). Those belonging to the Aphodiidae, Scarabaeidae or Geotrupidae families are present in Iberian Peninsula, and the latter two families comprise dung species that are well adapted to arid environments (Verdú and Galante 2002; Cabrero-Sañudo and Lobo 2003). Additionally, dry lands or calcareous soils typical in Mediterranean ecosystems are favourable for terrestrial snails or ants acting as intermediate host of trematodes (Manga-González et al. 2001; Otranto and Traversa 2003). In fact, fluke species are only present in Semiarid-Mesomediterranean and Thermomediterranean areas. In addition, the environmental characteristics of the Humid Temperate Atlantic area provide adequate requirements for earthworms' development (Edwards and Lofty 1977), which are intermediate or paratenic hosts for Trichurida and Ascaridida nematodes, respectively (Fedynich 2008; Yabsley 2008). Therefore, the presence of earthworms could be helping the maintenance of these parasites in Humid Temperate Atlantic areas.

Conclusions

Our study provides an account of the prevalence, richness, abundance and intensity of parasite infections in red-legged partridges in the main bioclimatic territories in the Iberian Peninsula. This novel information about the geographic distribution of digestive parasites in this game species is necessary to evaluate, in a comprehensive manner, the health status of the Iberian populations. In this way, a deeper knowledge of the distribution and prevalence of these parasites (some of which potentially cause of a negative impact on partridges), is the necessary basis to design management practices that limit the spread of these pathogens and, consequently, to ensure the long-term sustainability of red-legged partridge. Special emphasis should be placed on controlling parasite infections in farm-reared partridges prior to restocking hunting estates, as well as avoiding parasite concentration in areas where supplementary feeding is provided following their introduction.

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Declarations

Ethics approval Not applicable.

Consent to participate Authors have permission to participate.

Consent for publication Authors have permission for publication.

Conflict of interest None.

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References

- Anderson RC, Chabaud AG, Willmott S (1974) CIH Keys to the nematode parasites of vertebrates. Common Health Agricultural Bureau, Farnham Royal, Bucks, England, UK
- Arroyo B, Delibes-Mateos M, Díaz-Fernández S, Viñuela J (2012) Hunting management in relation to profitability aims: red-legged partridge hunting in central Spain. *Eur J Wildlife Res* 58(5):847–855
- Arroyo B, Caro J, Muñoz-Adalla EJ, Díaz-Fernández S, Delibes-Mateos M, Díaz-Fernández M, Viñuela J (2017) Reconciling economic and ecological sustainability: can non-intensive hunting of red-legged partridges be economically profitable? *Eur J Wildl Res* 63:14
- Birdlife International (2022) *Alectoris rufa*. Species factsheet: *Alectoris rufa*. <http://www.birdlife.org>. Accessed 1 May 2022
- Blanco-Aguilar JA, Virgós E, Villafuerte R (2003) Perdiz roja (*Alectoris rufa*). In: Martí R, Del Moral JC (eds) Atlas de las aves reproductoras de España. Dirección General de Conservación de la Naturaleza y Sociedad Española de Ornitología, Madrid, pp 212–213
- Blanco-Aguilar JA, Virgós E, Villafuerte R (2004) La perdiz roja (*Alectoris rufa*). In: Madroño A, González C, Atienza JC (eds) Libro Rojo de las Aves de España. Dirección General para la Biodiversidad-SEO/Birdlife, Madrid, pp 182–185
- Blanco-Aguilar JA, González-Jara P, Ferrero ME, Sánchez-Barbudo I, Virgós E, Villafuerte R, Dávila JA (2008) Assessment of game restocking contributions to anthropogenic hybridization: the case of the Iberian red-legged partridge. *Anim Conserv* 11:535–545
- Bolognesi PG, Galuppi R, Catelli E, Cecchinato M, Frasnelli M, Raffini E, Marzadori F, Tampieri MP (2006) Outbreak of *Eimeria kofoidi* and *E. legionensis* coccidiosis in red-legged partridges (*Alectoris rufa*). *Italian J Animal Sci* 5(3):318–320
- Brown JH (1984) On the relationship between abundance and distribution of species. *Am Nat* 124(2):255–279
- Brunner FS, Eizaguirre C (2016) Can environmental change affect host/parasite-mediated speciation? *Zoology (Jena)* 119(4):384–394

- Buenestado FJ, Ferreras P, Delibes-Mateos M, Tortosa P, Blanco-Aguilar JA, Villafuerte R (2008) Habitat selection and home range size of red-legged partridges in Spain. *Agric Ecosyst Environ* 126(3–4):158–162
- Buenestado FJ, Ferreras P, Blanco-Aguilar JA, Sánchez-Tortosa F, Villafuerte R (2009) Survival and causes of mortality among wild red-legged partridges *Alectoris rufa* in Southern Spain: implications for conservation. *IBIS* 151(4):720–730
- Bush AO, Lafferty D, Lotz JM, Shostak AW (1997) Parasitology meets ecology on own terms: Margolis et al. revisited. *J Parasitol* 83(4):575–583
- Cabodevilla X, Estrada A, Mougeot F, Jimenez J, Arroyo B (2021) Farmland composition and farming practices explain spatio-temporal variations in red-legged partridge density in central Spain. *Sci Total Environ* 799:149406
- Cabrero-Sañudo FJ, Lobo JM (2003) Reconocimiento de los factores determinantes de la riqueza de especies: el caso de los Aphodiinae (Coleoptera, Scarabaeoidea, Aphodiidae) en la Península Ibérica. *Graellsia* 59(2–3):155–177
- Calvete C, Estrada R, Lucientes J, Telletxeta EA, I, (2003) Correlates of helminth community in the red-legged partridge (*Alectoris rufa*) in Spain. *J Parasitol* 89(3):445–451
- Calvete C, Blanco-Aguilar JA, Virgós E, Cabezas-Díaz S, Villafuerte R (2004) Spatial variation in helminth community structure in the red-legged partridge (*Alectoris rufa* L.): effects of definitive host density. *Parasitology* 129:101–113
- Casas F, Viñuela J (2010) Agricultural practices or game management: which is the key to improve red-legged partridge nesting success in agricultural landscapes? *Environ Conserv* 37(2):177–186
- Casas F, Arroyo B, Viñuela J, Guzmán JL, Mougeot F (2016) Are farm-reared red-legged partridge releases increasing hunting pressure on wild breeding partridges in central Spain? *Eur J Wildl Res* 62(1):79–84
- Delibes-Mateos M, Farfán MA, Olivero J, Vargas JM (2012) Impact of land-use changes on red-legged partridge conservation in the Iberian Peninsula. *Environ Conserv* 39(4):337–346
- Díaz-Fernández S, Arroyo B, Casas F, Martínez-Haro M, Viñuela J (2013) Effect of game management on wild red-legged partridge abundance. *PLoS One* 8(6):e66671
- Edwards CA, Lofty JR (1977) Earthworms as pests and benefactors. In: Edwards CA, Lofty JR (eds) *Biology of Earthworms*, 2nd edn. Springer, Boston, pp 222–230
- Fanelli A, Menardi G, Chiodo M, Giordano O, Ficetto G, Bessone M, Lasagna A, Carpinano MG, Molinar Min A, Gugiatti A, Meneguz PG, Tizzani P (2020) Gastroenteric parasite of wild Galliformes in the Italian Alps: implication for conservation management. *Parasitology* 147(4):471–477
- Fedynich AM (2008). Heterakis and Ascaridia. In Atkinson CT, Thomas NJ, Bruce Hunter D. (ed). *Parasitic diseases of wild birds*, Wiley-Blackwell, Ames, Iowa, pp: 388–412
- Gaudioso Lacasa VR, Sánchez García-Abad C, Prieto Martín R, Bartolomé Rodríguez DJ, Pérez Garrido JA, Alonso de La Varga ME (2010) Small game water troughs in a Spanish agrarian pseudo steppe: visits and water site choice by wild fauna. *Eur J Wildlife Res* 56(4):591–599
- Globokar M, Fischer D, Pantchev N (2017) Occurrence of endoparasites in captive birds between 2005 to 2011 as determined by faecal flotation and review of literature. *Berl Munch Tierarztl Wochenschr* 130(11–12):461–473
- González-Quevedo C, Davies RG, Richardson DS (2014) Predictors of malaria infection in a wild bird population: landscape-level analyses reveal climatic and anthropogenic factors. *J Anim Ecol* 83(5):1091–1102
- Gortázar C, Villafuerte R, Martín M (2000) Success of traditional restocking of red-legged partridge for hunting purposes in areas of low density of northeast Spain Aragón. *Z Jagdwiss* 46:23–30
- Guerra-Silveira F, Abad-Franch F (2013) Sex bias in infectious disease epidemiology: patterns and processes. *PLoS One* 8(4):e62390
- Holland JM, Hutchison MAS, Smith B, Aebischer NJ (2006) A review of invertebrates and seed-bearing plants as food for farmland birds in Europe. *Ann Appl Biol* 148(1):49–71
- Holand H, Jensen H, Kvalnes T, Tufto J, Pärn H, Sæther BE, Ringsby TH (2019) Parasite prevalence increases with temperature in an avian metapopulation in northern Norway. *Parasitology* 149(8):1030–1035
- Jamieson IG, Lacy RC (2012) Managing genetic issues in reintroduction biology. In: Ewen JG, Armstrong DP, Parker KA, Seddon PJ (eds) *Reintroduction biology: integrating science and management*. Wiley, Hoboken, pp 441–475
- Klein SL (2004) Hormonal and immunological mechanisms mediating sex differences in parasite infection. *Parasite Immunol* 26(6–7):247–264
- Krasnov BR, Poulin R (2010) Ecological properties of a parasite: species specific stability and geographical variation. In: Morand S, Krasnov BR (eds) *The Biogeography of Host-Parasite Interactions*. Oxford University Press, Oxford, pp 99–114
- Máca O, Pavlásek I (2020) Protozoan and helminth infections of aviary-reared *Alectoris rufa* (Galliformes: Phasianidae) before releasing for hunting in the Czech Republic: infection dynamics and potential risks. *J Parasitol* 106(4):439–443
- MAFF (1986) *Manual of Veterinary Parasitological Laboratory Techniques*. Ministry of Agriculture, Fisheries and Food, HMSO, London
- Manga-González MY, González-Lanza C, Cabanas E, Campo R (2001) Contributions to and review of dicrocoeliosis, with special reference to the intermediate hosts of *Dicrocoelium dendriticum*. *Parasitology* 123:S91–S114
- Mani P, Perrucci S, Bennati L, Bagliacca M, Santilli F, Mazzoni DS, Rossi G (2000) Parasitological control of wild pheasants in managed areas of Siena province. *Ann Fac Med Vet Univ Pisa* 53:35–42
- Margolis L, Esch GW, Holmes JC, Kuris AM, Schad AG (1982) The use of ecological terms in parasitology (Report of an ad hoc committee of the American Society of Parasitologists). *J Parasitol* 68(1):131–133
- Martínez-Guijosa J, Martínez-Carrasco C, López-Olvera JR, Fernández-Aguilar X, Colom-Cadena A, Cabezón O, Mentaberre G, Ferrer D, Velarde R, Gassó D, Garel M, Rossi L, Lavín S, Serrano E (2015) Male-biased gastrointestinal parasitism in a nearly monomorphic mountain ungulate. *Parasit Vectors* 8:165
- Millán J, Gortázar C, Buenestado FJ, Rodríguez P, Tortosa FS, Villafuerte R (2003) Effects of a fiber-rich diet on physiology and survival of farm-reared red-legged partridges (*Alectoris rufa*). *Comp Biochem Physiol A Mol Integr Physiol* 134(1):85–91
- Millán J, Gortázar C, Villafuerte R (2004a) A comparison of the helminth faunas of wild and farm-reared red-legged partridges. *J Wildl Manag* 68(3):701–707
- Millán J, Gortázar C, Martín-Mateo MP, Villafuerte R (2004b) Comparative survey of the ectoparasite fauna of wild and farm-reared red-legged partridges (*Alectoris rufa*), with an ecological study in wild populations. *Parasitol Res* 93(1):79–85
- Millán J (2009) Diseases of the red-legged partridge (*Alectoris rufa* L.): a review. *Wildl Biol Pract* 5(1):70–88
- Morand S (2015) (macro-) Evolutionary ecology of parasite diversity: From determinants of parasite species richness to host diversification. *Int J Parasitol Parasites Wildl* 4(1):80–87
- Naciri M, Repérant JM, Fort G, Crespin J, Duperray J, Benzoni G (2011) *Eimeria* involved in a case of coccidiosis in farmed red-legged partridges (*Alectoris rufa*) in France: oocyst isolation and gross lesion description after experimental infection. *Avian Pathol* 40(5):515–524

- Nichols E, Gómez A (2014) Dung beetles and fecal helminth transmission: patterns, mechanisms and questions. *Parasitology* 141(5):614–623
- Olson DM, Dinerstein E, Wikramanayake ED, Burgess ND, Powell GVN, Underwood EC, D'Amico JA, Itoua I, Strand HE, Morrison JC, Loucks CJ, Allnutt TF, Ricketts TH, Kura Y, Lamoreux JF, Wettengel WW, Hedao P, Kassem KR (2001) Terrestrial Ecoregions of the world: a new map of life on earth. *Bioscience* 51(11):933–938
- Otranto D, Traversa D (2003) Dicrocoeliosis of ruminants: a little known fluke disease. *Trends Parasitol* 19(1):12–15
- Pérez JA, Alonso ME, Gaudioso VR, Olmedo JA, Díez C, Bartolomé D (2004) Use of radiotracking techniques to study a summer repopulation with red-legged partridge (*Alectoris rufa*) chicks. *Poult Sci* 83(6):882–888
- Polello L, Molinar Min AR, Fanelli A, Negri E, Peano A, Meneguz PG, Tizzani P (2021) First data on gastrointestinal parasitic infection in the red-legged partridge (*Alectoris rufa*) in Italy. *Diversity* 13:287
- Poulin R (2007) *Evolutionary Ecology of Parasites*, 2nd edn. Princeton University Press, Princeton
- Provencher JF, Gilchrist HG, Mallory ML, Mitchell GW, Forbes MR (2016) Direct and indirect causes of sex differences in mercury concentrations and parasitic infections in a marine bird. *Sci Total Environ* 551–552:506–512
- R Core Team (2018) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. <http://www.R-project.org/>
- Reino L, Borrallho R, Arroyo B (2016) Influence of game crops on the distribution and productivity of red-legged partridges *Alectoris rufa* in Mediterranean woodlands. *Eur J Wildl Res* 62:609–617
- Rey Benayas MJ, Scheiner SM (2002) Plant diversity, biogeography and environment in Iberia: patterns and possible causal factors. *J Veg Sci* 13(2):245–258
- Rivas-Martínez S, Penas Á, del Río S, Díaz González TE, Rivas-Sáenz S (2017) Bioclimatology of the Iberian Peninsula and the Balearic Islands. In Loidi J (ed) *The Vegetation of the Iberian Peninsula. Plant and Vegetation*, vol 12. Springer, Cham, Netherlands, 29–80
- Sánchez-Donoso I, Vilà C, Puigcerver M, Butkauskas D, De la Calle JRC, Morales-Rodríguez PA, Rodríguez-Teijeiro JD (2012) Are farm-reared quails for game restocking really common quails (*Coturnix coturnix*): a genetic approach. *PLoS One* 7(6):e39031
- Sánchez García-Abad C, Alonso de la Varga ME, Prieto Martín R, González Eguren V, Gaudioso Lacasa VR (2009) Una visión sobre la avicultura para la producción de caza en España. *ITEA* 105(3):169–183
- Sanchis-Monsonís G, Fanelli A, Tizzani P, Martínez-Carrasco C (2019) First epidemiological data on *Spirocerca vulpis* in the red fox: a parasite of clustered geographical distribution. *Vet Parasitol Reg Stud Reports* 18:100338
- Santilli F, Bagliacca M (2012) Occurrence of eggs and oocysts of intestinal parasites of pheasant (*Phasianus colchicus*) in droppings collected in differently managed protected areas of Tuscany (Italy). *Eur J Wildl Res* 58:369–372
- Sáenz De Buruaga M, Lucio AJ, Purroy J (1991) Reconocimiento de sexo y edad en especies cinegéticas. Diputación Foral de Álava, Vitoria
- Schmidt GD (1986) *Handbook of tapeworm identification*. CRC Press Inc., Boca Raton, Florida
- Sillero N, Brito JC, Skidmore AK, Toxopeus AG (2009) Biogeographical patterns derived from remote sensing variables: the amphibians and reptiles of the Iberian Peninsula. *Amphibia-Reptilia* 30(2):185–206
- Sithithaworn P, Tesana S, Pipitgool V, Kaewkes S, Pairojkul C, Sripa B, Paupairoj A, Thaiklar K (1991) Relationship between faecal egg count and worm burden of *Opisthorchis viverrini* in human autopsy cases. *Parasitology* 102(2):277–281
- Sobral M, Sousa S, Ribeiro T, Galvão SR, Santos RM, Silva R, Reis TS, Dias F, Santos HD (2019) Infection by *Platynosomum illiciens* (= *P. fastosum*) in domestic cats of Araguaína, Tocantins, northern Brazil. *Rev Bras Parasitol Vet* 28(4):786–789
- Soulsby EJJ (1987) *Parasitología y enfermedades parasitarias en los animales domésticos*. 7th edn. Nueva Editorial Interamericana, México D.F.
- Thieltges DW, Hüssel B, Baekgaard H (2006) Endoparasites in common eiders *Somateria mollissima* from birds killed by an oil spill in the northern Wadden Sea. *J Sea Res* 55(4):301–308
- Tompkins DM, Dickson G, Hudson PJ (1999) Parasite-mediated competition between pheasant and grey partridge: a preliminary investigation. *Oecologia* 119(3):378–382
- Verdú JR, Galante E (2002) Climatic stress, food availability and human activity as determinants of endemism patterns in the Mediterranean region: the case of dung beetles (Coleoptera, Scarabaeoidea) in the Iberian Peninsula. *Divers Distrib* 8(5):259–274
- Viñuela J, Casas F, Díaz-Fernández S, Delibes-Mateos M, Mougeot F, Arroyo B (2013) The red-legged partridge (*Alectoris rufa*) in Spain: a threatened game species. *Ecosistemas* 22(2):6–12
- Villanúa D, Hofle U, Pérez-Rodríguez L, Gortázar G (2006) *Trichomonas gallinae* in wintering common wood pigeons *Columba palumbus* in Spain. *Ibis* 148(4):641–648
- Villanúa D, Pérez-Rodríguez L, Rodríguez O, Viñuela J, Gortázar C (2007) How effective is pre-release nematode control in farm-reared red-legged partridges *Alectoris rufa*? *J Helminthol* 81(1):101–103
- Villanúa D, Pérez-López L, Casas F, Alzaga V, Acevedo P, Viñuela J, Gortázar C (2008) Sanitary risks of red-legged partridge releases: introduction of parasites. *Eur J Wildl Res* 54:199–204
- Wang Y, Naumann U, Wright ST, Warton DI (2012) Mvabund - an R package for model-based analysis of multivariate abundance data. *Methods Ecol Evol* 3:471–474
- Yamaguti S (1961) Nematodes of birds. In: Yamaguti S (ed) *System Helminthum*. Interscience publishers INC, New York, pp 183–349
- Yamaguti S (1971) *Synopsis of the Digenetic Trematodes of Vertebrates*. Keigaku Publishing Co., Vols I & II, Tokio
- Yabsley MJ (2008) Capillarid nematodes. In: Atkinson CT, Thomas NJ, Bruce Hunter D (eds) *Parasitic Diseases of Wild Birds*. Wiley-Blackwell, Ames, Iowa, pp 463–497
- Zloch A, Kuchling S, Hess M, Hess C (2021) In addition to birds' age and outdoor access, the detection method is of high importance to determine the prevalence of gastrointestinal helminths in laying hens kept in alternative husbandry systems. *Vet Parasitol* 299:109559

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