

Seed predators can increase nectar volumes in an alpine daisy: but do the insects benefit?

Junpeng Mu · Yan Chen · Yulian Yang · Ronghua Fu · Hui Wang · Stephen G. Compton

Received: 15 May 2016 / Accepted: 19 August 2016 / Published online: 25 August 2016
© Springer Science+Business Media Dordrecht 2016

Abstract Pollinators and pre-dispersal seed predators can interact via the plants they share. We examined how pre-dispersal seed predators modify nectar quality and quantity and thereby influence pollinator behaviour. Working in a Tibetan alpine meadow, we hypothesized that increasing levels of pre-dispersal seed predation by larvae of tephritid flies would reduce nectar quantity and quality in *Saussurea nigrescens* (Asteraceae), and that this would make the flowers less attractive to honeybees, the most frequent floral visitors. Our field experiments showed that floret nectar volumes responded differently to high and low densities of fly larvae, with significant increases when there was one larva present, but decreases when two or more larvae were present in a capitulum. Experimental

manipulations of fly larvae yielded the same result. The increases in nectar volume generated by a single larva are likely to be beneficial for these insects in locations where pollinators preferentially visit those *S. nigrescens* that produce more nectar. At our study sites, honeybees were the main pollinators and visitation rates were unaffected by the changes in nectar volumes, but they are introduced to the area, and native pollinators may be more selective.

Keywords *Apis mellifera* · Asteraceae · Insect–plant interactions · Nectar · *Saussurea* · *Tephritis* · Tibetan Plateau

J. Mu (✉) · Y. Chen · Y. Yang · H. Wang
Ecological Security and Protection Key Laboratory of
Sichuan Province, Mianyang Normal University,
Mianyang 621000, China
e-mail: gbmujp@163.com

R. Fu
Key Laboratory of Southwest China Wildlife Resources
Conservation (Ministry of Education), China West
Normal University, Nanchong 637002, China

S. G. Compton
School of Biology, University of Leeds, Leeds LS2 9JT,
UK

S. G. Compton
Department of Zoology and Entomology, Rhodes
University, Grahamstown 6140, South Africa

Introduction

Pollinators and flowering plants have contributed significantly to our understanding of the evolution of mutualisms involving both generalist and specialist pollination systems (Kjellberg et al. 2001; Lunau 2004; Mitchell et al. 2009; Bronstein 2015). Co-evolution is most readily apparent in relation to the accessibility of rewards, with pollinators exerting selective pressures on floral traits such as spur length (Nilsson 1988) and plants selecting for pollinator tongue length (Whittall and Hodges 2007; Miller-Struttman et al. 2015). The quantity of accessible rewards thereby influences which pollinators are attracted, and of which species, and can also influence

how long individual pollinators remain on each flower (Kalinganire et al. 2001).

Plants and pollinators rarely if ever interact in isolation from other organisms, many of which are likely to have antagonistic rather than mutualistic relationships with the plants (Galen 1999). They include antagonists that can directly or indirectly influence various traits through consumption of plant parts, such as decrease flower size (Barber et al. 2012), modify nectar odour (Press and Phoenix 2005), reduce the quality of floral displays (McCall and Irwin 2006), and reduce nectar production (McDade and Kinsman 1980), all of which may influence pollinator behaviour (Rodríguez- Rodríguez et al. 2015).

Seed predatory insects often have a strong influence on plant reproductive success and can generate significant selection pressures on their host plants (Kolb et al. 2007). Their impact on their hosts varies in relation to plant reproductive traits such as flowering phenology, flower number and seed size, and these attributes are also significant for the insects' own reproductive success, thereby facilitating co-adaptation and co-evolution (Thompson 2005). Pre-dispersal seed predators require seeds to feed their larvae, but often oviposit early in floral development, before ovules have been pollinated. Their reproductive success is therefore influenced by the quality and quantity of subsequent pollination events (Strauss and Irwin 2004). The adults of some seed predators such as fig wasps and yucca moths ensure that seeds are available for their larvae by routinely pollinating the flowers themselves (Pellmyr and Huth 1994), but for the majority of species, this is achieved by the selection of oviposition sites in flowers or inflorescences that have a high probability of being pollinated. Many pre-dispersal seed predators have a limited range of host plants and often just a single host species (Collin and Shykoff 2010). Within each plant species, the likelihood and quality of pollination can depend on a wide range of environmental and biological variables, but phenotypic characteristics of individual plants are also significant. For insect-pollinated species, the size and quality of floral displays and the quality of rewards they offer influence the extent of seed set (Thomson 1988; Vaughton and Ramsey 1998). Consequently, the flowers that are most rewarding for oviposition by seed predators are likely to be those that are also most favourable to the plant's pollinators (Cariveau et al. 2004).

Seeds are nitrogen-rich and often contain high concentrations of energetically expensive defensive compounds (Janzen et al. 1977; Birch et al. 1986). Damage generated by seed predators during oviposition or early larval feeding, together with adaptive responses by the plants to the presence of the insects, can result in flowers containing seed predators receiving less investment, with resources being switched to undamaged flowers or retained elsewhere (Kudoh and Whigham 1998; Westerbergh and Westerbergh 2001; Cariveau et al. 2004). Consequently, flowers containing seed predators may be less attractive to pollinators and set fewer seeds (Cariveau et al. 2004).

Floral nectar is the most widely provided reward for insect visitors to flowers (Scaven and Rafferty 2013), and plants with more and higher quality nectar can attract more pollinators (Mitchell et al. 2004; Larsson and Franzen 2007; Wallis de Vries et al. 2012). Nectar is composed mainly of sugars derived from photosynthesis and forms part of a plant's overall carbohydrate content. Carbohydrates are stored in both reproductive and vegetative organs (Pacini and Nepi 2007), and linkage has been recorded between carbohydrate storage and nectar production, with more nectar produced at times of day when overall carbohydrate content is higher (Mu et al. 2015). Herbivory often reduces the quantity of reserves stored within plants (Machado et al. 2013), which suggests it may also reduce nectar production, and indirectly influence pollinator behaviour to the further detriment of the plant.

Here, we describe the interplay between a pre-dispersal seed predator, the quantity and quality of nectar in flowers of its host plant, the carbohydrates stored throughout the plant and the willingness of bees to visit its flowers. We hypothesized that

1. Pre-dispersal seed predators reduce the quantity and quality of nectar in the flowers where they were feeding,
2. Increasing numbers of seed predators have a progressively greater impact on nectar production,
3. Seed predators reduce the proportion of the plant's carbohydrates stored in the flowers, relative to other parts of the plant, and
4. Any changes in nectar rewards as a result of the seed predators reduce flower visitation by the pollinators of the plant.

Methods

Natural history

Saussurea nigrescens (Asteraceae section Compositae, ECCAS 1999) is a widespread high-altitude perennial distributed 2000–4300 m on the Tibetan Plateau. Its growing season is short, with plants reviving in mid-May and senescing by mid-September. Mature plants vary in height about 15–45 cm. Flowering occurs from July to August and seeds mature and disperse in late-September. Each plant produces 2–5 dull-purple capitula, and each of which contains 20–55 florets. The florets have an annular bowl-shaped nectary between the ovary and anthers (bowl diameter is 1.5–2.0 mm). The flowers are monoclinal (with stamens and pistils in the same flower) but outcrossing is favoured by protandry (stamens mature before the ovaries), see Mu et al. (2014, 2015).

Saussurea nigrescens is the most economically important nectar-producing plant in parts of the Tibetan plateau and contributes approximately 80 % of the annual honey harvest in Hongyuan County, China (Mu et al. 2014). The plant is allogamous and requires cross-pollination, which is usually by honeybees (Mu et al. 2014).

The only recorded pre-dispersal seed predators of *S. nigrescens* are larvae of several tephritid flies belonging to the genera *Campiglossa*, *Tephritis* and *Urophora* (Xi et al. 2016). Tephritid females oviposit into the flower buds of the host plant in the middle of July. Their larvae develop within the capitula, where they consume developing seeds and damage the receptacles (Xi et al. 2016). Between one and three, larvae share a capitulum. No obvious gall formation takes place. The tephritid larvae do not feed on floral nectar, nor do they directly damage the nectaries. They pupate inside the capitula, where they remain until the adults emerge the following spring.

Study sites

The study was conducted from 2014 to 2015 at the Hongyuan Alpine Meadow Ecosystem Research Station of the Chinese Academy of Sciences, located in Hongyuan County, Sichuan Province, China (32°48′–32°52′N, 102°01′–102°33′E). This area of the eastern Qinghai-Tibetan Plateau is at an altitude of about 3500 m, with short and cool spring, summer and

autumn seasons and a long cold winter. Liu et al. (2012) and Mu et al. (2015) provided details of the local climate, soil conditions and vegetation. The meadows are sometimes used for cattle grazing during the winter but are otherwise undisturbed. *Tephritis femoralis* Chen is the common tephritid seed predator of *S. nigrescens* locally.

In late July 2014, we selected three sites (sites 1–3) located about 5 km apart that had similar soil moisture and physical and chemical properties (Mu et al. 2014). Their plant communities were also similar. *Saussurea nigrescens* was the most abundant species at the sites, contributing 33–46 % of total plant cover during its flowering period. *Elymus nutans* Griseb, *Anemone rivularis* Buch.-Ham. ex DC. and *Potentilla anserina* L. were also common. The three sites were at roughly similar distances from the nearest apiaries and similar numbers of bees and adult tephritid flies were recorded visiting the flowers of *S. nigrescens* (Mu et al. 2014). *Saussurea nigrescens* was the major source of nectar for bees at all three study sites (Mu et al. 2014). Native pollinators of *S. nigrescens* include an Asian honeybee (*Apis cerana*) and the bumblebees *Bombus filchnerae*, *B. humilis* and *B. supremus* (Macior et al. 2001). The introduced *Apis mellifera* has been tended by beekeepers in the area since 1981 (Sun et al. 2013).

Field records of tephritid numbers and nectar variables

In July 2014, we tagged 50 healthy *S. nigrescens* with undamaged leaves at each of the three sites. Afterwards, the tephritids had an opportunity to oviposit, but before the flowers had opened, we enclosed all the capitula on each plant within fine mesh netting to exclude further insect visitors (Real and Rathcke 1991). This plant produces its maximum nectar volumes when the anthers are white and emerging from the capitula (Mu et al. 2014). When they reached this stage, we selected five florets at random from each of the capitula, and on sunny days at 10:00–15:00 h, we measured their nectar volumes and concentrations. Nectar volumes were measured using 1 or 5 µl micropipettes (Hirschmann Laborgeräte, Germany). Nectar concentrations were measured concurrently with a hand-held refractometer (Eclipse, Stanley Company, England Bellingham, UK) following the methods of Johnson et al. (2006). The numbers of tephritid larvae inside each capitulum were then

counted using a binocular microscope. A total of 472 capitula were selected to monitor nectar volume and concentration.

Plant responses to experimental manipulations

In July 2015, we selected 120 vigorous and undamaged *S. nigrescens* growing at site 2 and enclosed groups of six plants within cylindrical steel netting enclosures (100 cm diameter × 100 cm high, mesh size 0.1 mm × 0.1 mm). When flower buds were at the stage suitable for tephritid oviposition, all but one randomly chosen capitulum on each plant were removed. One, two or three *T. femoralis* (Tephritidae) larvae (about 2 mm in length) were then inserted into the remaining capitula. The control group was physically manipulated in the same way as the other groups, but no larvae were inserted. The larvae were obtained as described by Xi et al. (2016). The netting remained in place until the capitula reached peak nectar production, when floret nectar volumes and concentrations were recorded as before. A total of 120 capitula were selected to monitor nectar volume and concentration.

After nectar production had been monitored, each plant was removed and the lengths and fresh weights of capitula, leaves, stems and roots were weighed in the field using a 0.001 g capacity balance. The plant components were then stored immediately in a portable icebox cooled with dry ice, and their soluble sugar and starch contents were measured later following the procedures of Hansen and Møller (1975) and Yoshida et al. (1976). The relative amounts of carbohydrates (soluble sugars and starch, mg/g) in the capitula, leaves, stems and roots were expressed as a percentage of the total carbohydrates in each plant. Carbohydrate contents were calculated as in Rivera-Solís et al. (2012). A total of 120 capitula were selected to monitor the carbohydrate contents.

Honeybee responses to experimental manipulations

Twelve 2 × 2 m × 1 m high enclosures covered in the steel netting described above were erected at site 2 to enclose groups of *S. nigrescens* with recently developed flower buds. Once they reached the stage suitable for tephritid oviposition 30 vigorous, undamaged plants in each plot had all but one of their capitula removed and zero, one, two or three tephritid larvae

were inserted as before. Any additional plants in the enclosures had all their capitula removed, leaving a consistent 30 capitula within each 4 m² plot. To monitor honeybee visitation, we first removed the netting screens. Six operatives then simultaneously recorded honeybee visits to individual capitula in pairs of adjacent plots every hour (each plot was observed for 30 min per hour) at 9:00–17:00 on three sunny days in July 2015. Observers were located about 3 m from each plot, which allowed for clear observation of pollinator behaviour. Honeybee visitation rates to each capitulum per hour were calculated following the protocol of Arroyo et al. (1985). We then harvested the plants and measured fresh weights as before. The plant parts were then dried to constant mass and re-weighed to the nearest 0.001 g.

Statistical analyses

All statistical analyses were performed in R (R Development Core Team 2013, URL <http://www.R-project.org>). Plant traits were first tested for normality using the Shapiro–Wilk test and for homogeneity of variance using Levene’s test. Floret nectar volume was log₁₀-transformed, and the proportion of flowering plants per subplot was arcsine-transformed to achieve normality. Correlation analyses were used to determine the relationship between floret nectar volume and capitula carbohydrate contents. Pollinator visitation rates were assessed using one-way ANOVA followed by Tukey’s test.

We used general linear-mixed models (GLMMs) to assess the effects of pre-dispersal seed predator numbers and capitula mass on variation in floret nectar volume and concentration, capitulum carbohydrates and total carbohydrates storage as percentages of dry and wet weights. To account for interaction effects between pre-dispersal seed predator and capitulum size, we used a model with fly numbers and capitulum mass as fixed factors, and site and above-ground vegetative mass as random factors. For the mixed models, we used the *lme* function in the *lme4* package in R (Bates et al. 2011).

Results

Saussurea nigrescens colonized naturally by tephritid larvae produced different volumes of nectar

depending on how many larvae were present, but there was no simple relationship between numbers of these seed predators and nectar volume (Fig. 1, Table 1, “Appendix 1”). Capitula containing a single tephritid larva produced more nectar than controls, whereas capitula containing two or three larvae produced significantly less nectar. The presence of one larva increase nectar volumes by an average of 15.1 %, but two or three larvae led to decreases of 76.9 and 83.1 %, respectively (Fig. 1). In contrast to nectar volume, the tephritids had no effect on nectar concentration (“Appendix 1”). Very similar results were obtained when tephritid numbers were manipulated experimentally (Fig. 1; Table 1). With one larva added, nectar increased by 17.0 % relative to controls, and when two or three larvae were added nectar volumes fell by 77.8 and 79.4 %, respectively. Nectar concentrations were again unchanged (“Appendix 1”).

The starch and soluble sugar contents of *S. nigrescens* capitula showed the same response to tephritid larvae as nectar volume, with elevated carbohydrates compared with controls in capitula containing one larva and significantly lower concentrations of carbohydrates when two or three larvae were present (Fig. 2). Across treatments there was a significant positive correlation between capitulum carbohydrate concentrations and average nectar volume, with indications that the same pattern was also present within capitula that contained one larva (Fig. 3). The changes in capitulum carbohydrate concentrations in response to the tephritids reflected changes in the relative distributions of carbohydrates within the plants (Fig. 4). A single larva was associated with an increase in the percentage of the plant’s total carbohydrates stored within the capitula, relative to controls, but two or three larvae were associated with declines in the capitula carbohydrates (Fig. 4;

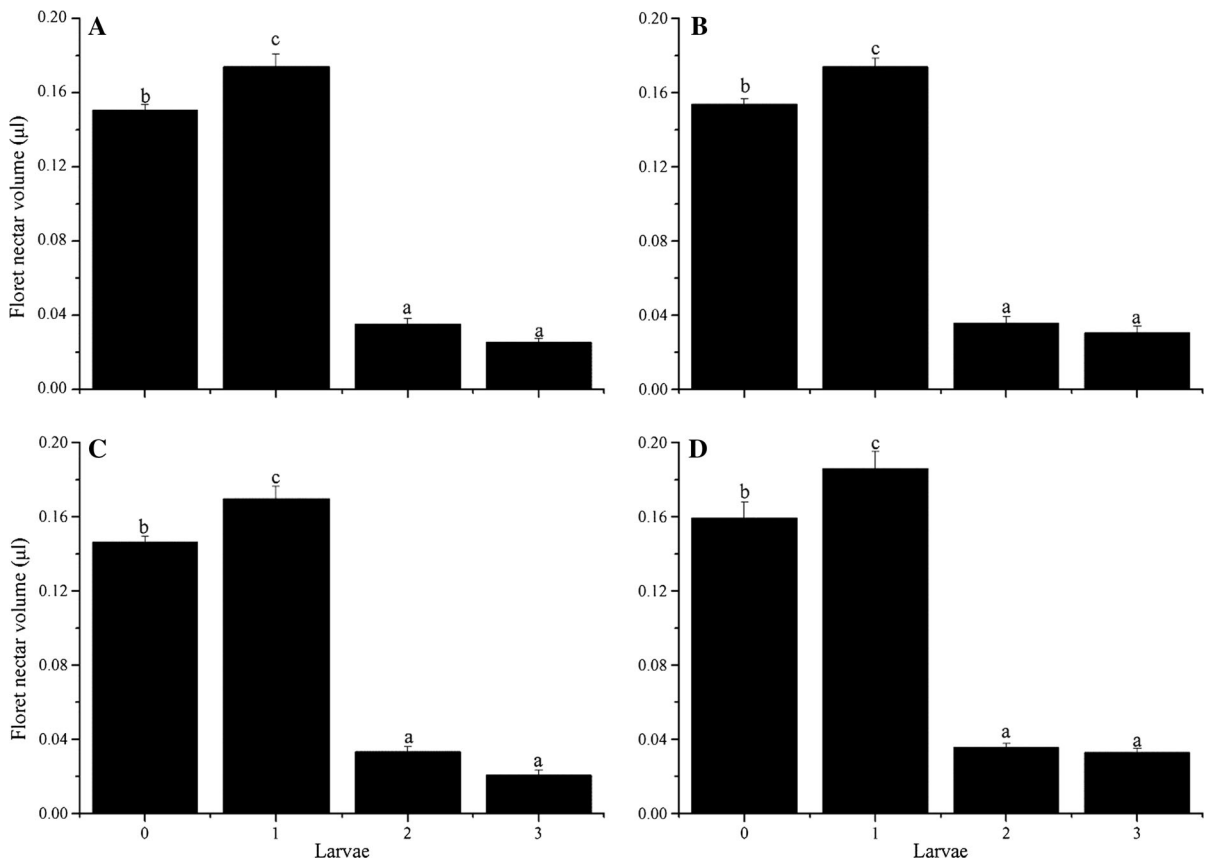


Fig. 1 Variation in *S. nigrescens* nectar volumes in relation to the numbers of tephritid larvae in their capitula (means ± SEs per floret). **a–c** plants growing at sites 1–3 with natural variation

in larval numbers, **d** experimentally introduced tephritid larvae (site 2 only). Different letters above columns indicate differences within sites at $P < 0.05$ (one-way analysis of variance)

Table 1 Summary of linear mixed effect models comparing the effects of pre-dispersal seed predators and capitulum size on floret nectar volume, nectar concentration and capitulum carbohydrate concentrations

Variables/AIC/BIC	Fixed effects			
	Value	SE	<i>t</i> value	<i>p</i> value
Field conditions				
Floret nectar volume/−1539.99/−1515.10				
Intercept	0.159	0.009	18.119	<0.001
Pre-dispersal seed predators (<i>P</i>)	−0.028	0.011	−2.441	0.015
Capitulum mass (<i>C</i>)	−0.050	0.054	0.929	0.353
<i>P</i> × <i>C</i>	−0.097	0.069	−1.403	0.161
Nectar concentration/3449.41/3474.30				
Intercept	39.977	2.000	19.984	<0.001
Pre-dispersal seed predators(<i>P</i>)	−1.568	2.388	−0.657	0.512
Capitula mass (<i>C</i>)	10.411	12.789	0.814	0.416
<i>P</i> × <i>C</i>	8.517	14.612	0.583	0.560
Experimental manipulations				
Floret nectar volume/−338.37/−321.85				
Intercept	0.159	0.018	8.983	<0.001
Pre-dispersal seed predators (<i>P</i>)	−0.046	0.010	−4.587	<0.001
Capitula mass (<i>C</i>)	0.158	0.106	1.495	0.138
<i>P</i> × <i>C</i>	−0.046	0.061	−0.759	0.449
Nectar concentration/747.25/763.77				
Intercept	41.630	1.909	21.802	<0.001
Pre-dispersal seed predators (<i>P</i>)	−0.055	1.083	−0.051	0.959
Capitulum mass (<i>C</i>)	−2.152	11.400	−0.189	0.851
<i>P</i> × <i>C</i>	−1.901	6.548	−0.290	0.772
Capitulum carbohydrate concentrations/1016.50/1033.02				
Intercept	187.545	46.583	4.026	<0.001
Pre-dispersal seed predators (<i>P</i>)	−8.602	3.921	−2.194	0.030
Capitulum mass (<i>C</i>)	16.649	30.306	0.549	0.584
<i>P</i> × <i>C</i>	−1.391	2.544	−0.545	0.587

The numbers after floret nectar volume and nectar concentration are AIC and BIC, respectively

AIC Akaike Information Criterion, BIC Bayesian Information Criterion

P values at <0.05 level are in bold

$t = -2.44$, $P < 0.05$). Corresponding changes were present in the proportion of carbohydrates stored in the stems, with declines when one larva was present, and increases when there were more larvae (Fig. 4, $t = 1.99$, $P < 0.05$). There were no treatment effects on the proportion of the plants' carbohydrates in the leaves and roots (Fig. 4, $t = -0.56$, $P = 0.58$ and $t = 0.86$, $P = 0.39$, respectively).

Despite the changes in nectar volumes associated with the tephritid larvae, they had no significant impact on the visitation rates of honeybees to the capitula ($F = 0.903$, $P = 0.388$, “Appendix 2”).

Discussion

We hypothesized that pre-dispersal seed predators would reduce floret nectar production and nectar concentrations in proportion to their densities, and that this would lead to a decline in pollinator visitation rates. These hypotheses were not supported by the data from both field records and experimental manipulations. Floret nectar volumes did decline in capitula that contained two or three tephritid larvae, but volumes increased significantly in capitula with one larva. Furthermore, nectar concentrations were

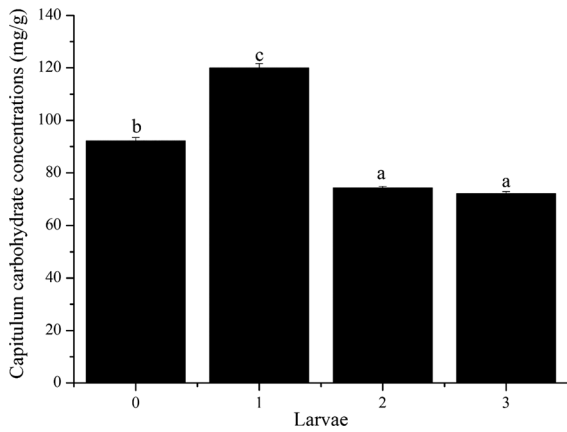


Fig. 2 Capitulum carbohydrate concentrations of *S. nigrescens* that had contained varying numbers of tephritid larvae (means \pm 1 SE). Different letters above columns indicate differences at $P < 0.05$ (one-way analysis of variance)

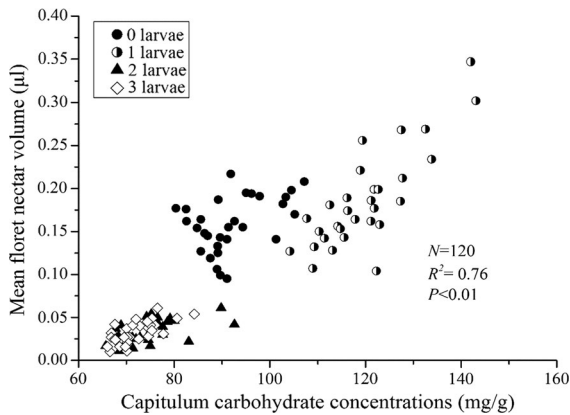


Fig. 3 The relationship between *S. nigrescens* capitulum carbohydrate concentrations and mean floret nectar volume in capitula where different numbers of tephritid larvae had been introduced. $N = 30$ florets for each larval density

consistently independent of seed predator numbers, as were visitation rates by honeybees. Although carbohydrate storage in the capitula was influenced by the presence of the tephritid larvae, it was a non-linear response that mirrored nectar production, with capitula containing a single larva having a higher proportion of the plant’s carbohydrates than controls with no seed predators.

Rates of nectar production and secretion are related to the quantities of soluble sugars and starch available and thereby to carbohydrate reserves and rates of photosynthesis (Búrquez and Corbet 1991;

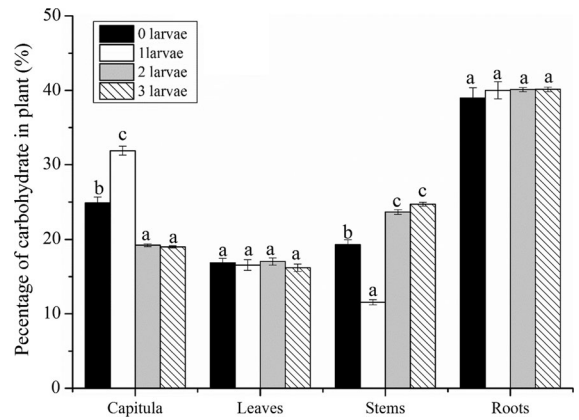


Fig. 4 Percentage of total carbohydrates (soluble sugars and starch, mg/g) in capitula, leaves, stems and roots of *S. nigrescens* (means \pm 1 SE). Different letters above columns indicate differences between parts of the plants at $P < 0.05$ (one-way analysis of variance)

Pacini et al. 2003). Physical conditions such as elevated night-time temperatures and damage to leaves and flowers can all reduce nectar production (Wäckers et al. 2001; Mu et al. 2015). In our study system, the positive relationship between local carbohydrate storage and floret nectar volume was particularly clear, because they increased and decreased together in response to varying densities of tephritid larvae. Relative to un-occupied capitula, carbohydrate storage increased by 19.4 % and floret nectar volumes increase by 17.0 % in capitula with one larva but were reduced by 30.3 and 32.6 %, respectively, when more larvae were present. Carbohydrate storage elsewhere on the plants reflected these changes.

The non-linear carbohydrate storage and nectar volume responses by the plants to the presence of insects feeding in their capitula can be interpreted in terms of adaptive responses by the plants in the face of varying levels of damage, and also as reflecting manipulations by the insects. The increase in capitula carbohydrate storage and nectar volumes when a single larva is present could be a plant compensation effect (Rivera-Solís et al. 2012). Plants attacked by herbivores often allocate more resources to their reproductive organs (reviewed by Trumble 1993), but increased allocation to reproduction, including elevation of nectar quality, is most evident when vegetative structures are damaged (Inouye 1982; Lanza 1988; Smith et al. 1990).

An alternative explanation for the increase in nectar volumes produced by capitula occupied by a single tephritid larva is that the insects are actively inducing this effect, and that increased nectar volumes favour their own reproductive success, rather than that of their host plants. The tephritid larvae feed on developing seeds and depend on capitula being visited by pollinators after their eggs have been laid. Any increase in the likelihood of those capitula being adequately pollinated, such as might be the case by increasing nectar volumes, would therefore be to the advantage of the insect. Many of the species of tephritids with larvae that develop in the capitula of Asteraceae are gall formers (Varley 1947; Straw 1989). This includes congeners of the *Tephritis* species from *S. nigrescens* (Goeden 1988). Gall forming insects routinely generate nutrient sinks that draw in resources from elsewhere on their host plants (Price et al. 1987). Although *T. femoralis* does not generate clearly defined galls in the capitula of *S. nigrescens*, its larvae may be capable of initiating some of the plant physiological effects associated with galling, including stimulation of local carbohydrate storage. This may have direct beneficial effects in terms of food quality for developing larvae and indirect benefits via larger nectar volumes leading to increased numbers of seed in the capitula it occupies.

Quite different carbohydrate storage and nectar volume responses were recorded in capitula where more than one tephritid larva was present. The declines seen in capitula containing multiple larvae are likely to reflect the damage they cause to the receptacles, through which nutrients and metabolites are transported (Teuber et al. 1983; Rivera-Solís et al. 2012). Damage to the receptacles was more frequent and much more extensive when two or more larvae were sharing a capitulum, with more than 90 % of the receptacles damaged in capitula with two or three larvae, compared with only around 10 % of the receptacles when only a single larva was present (J Mu unpublished data). The increased feeding on the receptacles in capitula with several larvae may be the result of antagonistic behaviour among the fly larvae and a shortage of seeds remaining to be eaten (Averill and Prokopy 1987).

Seed predators are expected to preferentially oviposit on flowers that are more attractive to pollinators, because these flowers are most likely to set seed (Strauss and Irwin 2004). Competition is often

intense among seed predators sharing what are usually finite and spatially limited resources. Only 15 % of the *S. nigrescens* capitula at our study site that contained tephritid larvae had more than one larva present (X XI unpublished data), suggesting that females typically lay a single egg on each plant. This oviposition pattern not only reduces the likelihood of competition, but also means that most of the larvae are developing in capitula that are producing more nectar than adjacent capitula that lack the insects.

Contrary to expectations that the insects would gain from developing in capitula with higher nectar volumes, capitula with one larva present did not attract more honeybees than un-occupied capitula. This result was contrary to much of the literature, which suggests that floral nectar volumes have a positive relationship with pollinator visitation (Schemske and Bradshaw 1999; Pyke 1982, 2016). Different bee species may respond differently to aspects of nectar rewards, such as nectar volume and concentration. For example, honeybees have been shown to prefer flowers that have high sugar concentrations (Scheiner et al. 1999, 2001; Vaudo et al. 2015), whereas bumblebees are sensitive to nectar volume (Harder and Real 1987). In this study, we found no changes in nectar concentration (sugar content) resulting from the pre-dispersal seed predators, and this may have led to the similar honeybee visitation rates. We did not examine visitation rates by native bees such as bumblebees and they may be more responsive to nectar volumes than honeybees. Our future investigations will examine whether pre-dispersal seed predators have different effects on the behaviour of honeybees and native bees.

Acknowledgments We thank Xinqiang Xi, Xinwei Wu, Jie Xiong, Hongli Chen, Yongping Li, Xu Luo, Xue Wei, Rui Cao, Hanxiang Chen and Kai He for their field and laboratory assistance. This study was funded by National Science Foundation of China (31270513, 31270387), Innovative Research Team by Sichuan Provincial Department of Education (13TD0015) and Project of Introduced Excellent Talents by Mianyang Normal University (QD2012A07).

Appendix 1

S. nigrescens nectar concentrations with natural variation in numbers of tephritid larvae at sites 1–3 (a–c) and in the larval-addition experiment at site 2 (d). Similar letters above columns indicate no differences within sites at $P < 0.05$ (one-way analysis of variance) (Fig. 5).

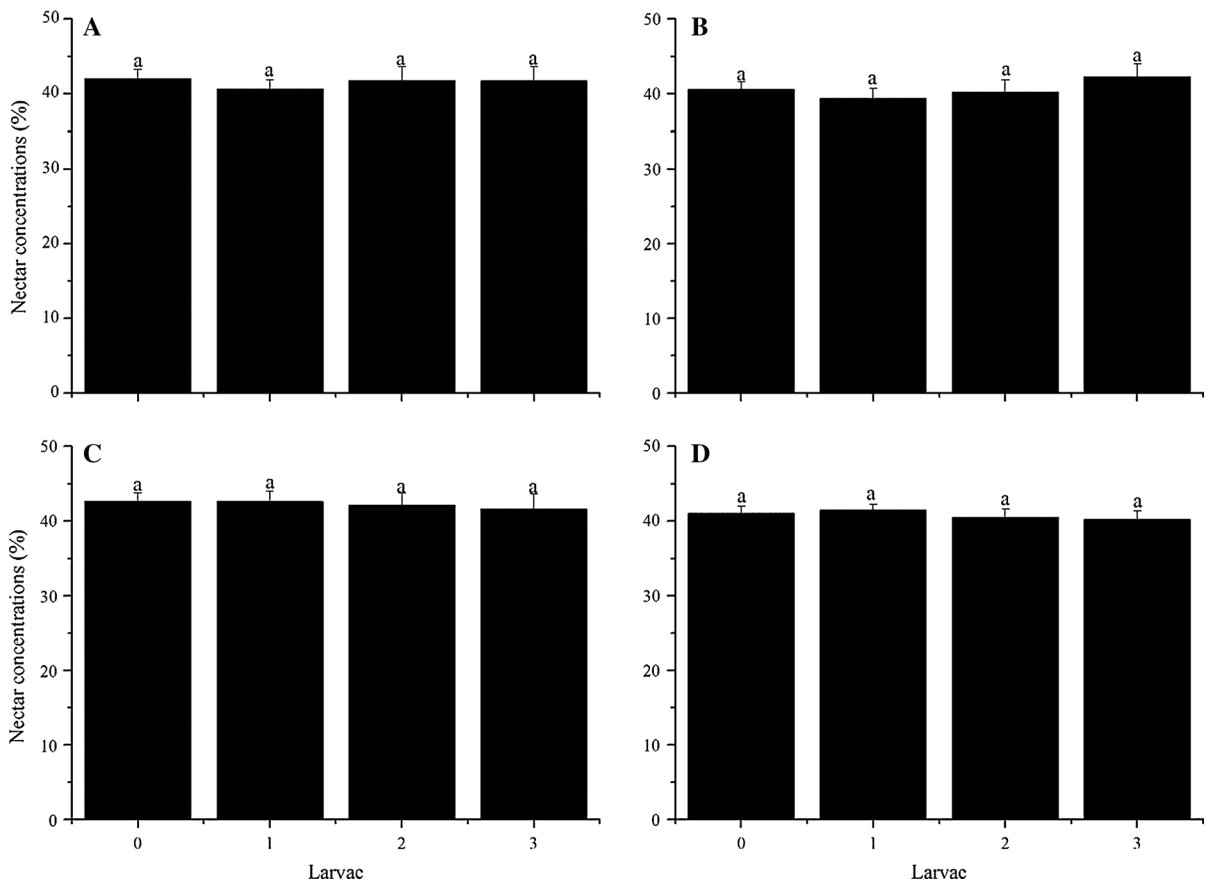


Fig. 5 *S. nigrescens* nectar concentrations at sites 1 (a), 2 (b) and 3 (c) under natural condition and after experimental introductions (d)

Appendix 2

Apis mellifera visits to capitula of *Saussurea nigrescens* that contained different numbers of tephritid

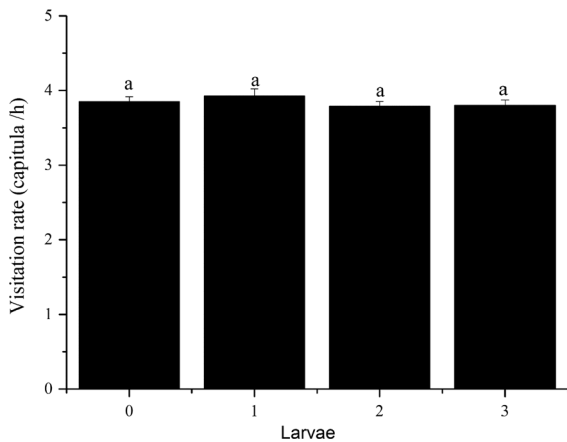


Fig. 6 *Apis mellifera* visitation rates per capitulum of *S. nigrescens* in relation to tephritid larval numbers

larvae (means \pm 1 SE). Capitulum visitation rates were compared by one-way ANOVAs followed by Tukey’s tests. Similar letters above columns indicate no difference in visitation rates at $P < 0.05$ (one-way analysis of variance) (Fig. 6).

References

Arroyo MTK, Armesto JJ, Primack RB (1985) Community studies in pollination ecology in the high temperate Andes of central Chile. II. Effect of temperature on visitation rates and pollination possibilities. *Plant Syst Evol* 149:187–203

Averill AL, Prokopy RJ (1987) Intraspecific competition in the tephritid fruit fly *Rhagoletis pomonella*. *Ecology* 68:878–886

Barber NA, Adler LS, Theis N, Hazzard RV, Kiers ET (2012) Herbivory reduces plant interactions with above- and belowground antagonists and mutualists. *Ecology* 93:1560–1570

Bates D, Maechler M, Bolker B (2011) Lme 4: linear mixed-effects models using S4 classes. R package version 0.999375-38

- Birch ANE, Fellows LE, Evans SV, Doherty K (1986) Para-aminophenylalanine in *Vigna*: possible taxonomic and ecological significance as a seed defence against bruchids. *Phytochemistry* 25:2745–2749
- Bronstein JL (ed) (2015) *Mutualism*. Oxford University Press, Oxford, 297 pp
- Búrquez A, Corbet SA (1991) Do flowers reabsorb nectar? *Funct Ecol* 5:369–379
- Cariveau D et al (2004) Direct and indirect effects of pollinators and seed predators to selection on plant and floral traits. *Oikos* 104:15–26
- Collin CL, Shykoff JA (2010) Flowering phenology and female fitness: impact of a pre-dispersal seed predator on a sexually polymorphic species. *Plant Ecol* 206:1–13
- ECCAS (Editor Committee of the Chinese Academy of Sciences for Flora of China) (1999) *Flora of China*. Beijing: Science Press (in Chinese) 78: 38
- Galen C (1999) Why do flowers vary? The functional ecology of variation in flower size and form within natural plant population. *Bioscience* 49:631–640
- Goeden RD (1988) Gall formation by the capitulum-infesting fruit fly, *Tephritis stigmatica* (Diptera: Tephritidae). *Proc Entomol Soc Wash* 90:37–43
- Hansen J, Møller IB (1975) Percolation of starch and soluble carbohydrates from plant tissue for quantitative determination with anthrone. *Anal Biochem* 68:87–94
- Harder LD, Real LA (1987) Why are bumble bees risk averse? *Ecology* 68:1104–1108
- Janzen DH, Juster HB, Bell EA (1977) Toxicity of secondary compounds to the seed-eating larvae of the bruchid beetle *Callosobruchus maculatus*. *Phytochemistry* 16:223–227
- Johnson SD, Hargreaves AL, Brown M (2006) Dark, bitter-tasting nectar functions as filter of flower visitors in a bird-pollinated plant. *Ecology* 87:2706–2716
- Kaliganire A, Harwood CE, Slee MU, Simons AJ (2001) Pollination and fruit-set of *Grevillea robusta* in western Kenya. *Aust Ecol* 26:637–648
- Kjellberg F, Jouselin E, Bronstein JL, Patel AJ, Yokoyama J, Rasplus JY (2001) Pollination mode in fig wasps: The predictive power of correlated traits. *Proc Roy Soc B Biol* 268:1113–1121
- Kolb A, Ehrlén J, Eriksson O (2007) Ecological and evolutionary consequences of spatial and temporal variation in pre-dispersal seed predation. *Perspect Plant Ecol* 9:79–100
- Kudoh H, Whigham DF (1998) The effect of petal size manipulation on pollinator/seed-predator mediated female reproductive success of *Hibiscus moscheutos*. *Oecologia* 117:70–79
- Lanza J (1988) Ant preferences for *Passiflora* nectar mimics that contain amino acids. *Biotropica* 20:341–344
- Larsson M, Franzen M (2007) Critical resource levels of pollen for the declining bee *Andrena hattorfiana* (Hymenoptera, Andrenidae). *Biol Conserv* 134:405–414
- Liu YZ, Mu JP, Li GY, Sun SC (2012) Global warming may reduce plant reproductive effort for temperate multi-flowered species. *New Phytol* 195:427–436
- Lunau K (2004) Adaptive radiation and coevolution—pollination biology case studies. *Org Divers Evol* 4:207–224
- Machado RAR, Ferrieri AP, Robert CA, Glauser G, Kallenbach M, Baldwin IT, Erb M (2013) Leaf-herbivore attack reduces carbon reserves and regrowth from the roots via jasmonate and auxin signaling. *New Phytol* 200:1234–1246
- Macior LW, Tang Y, Zhang JC (2001) Reproductive biology of *Pedicularis* (Scrophulariaceae) in the Sichuan Himalaya. *Plant Spec Biol* 16:83–89
- McCall AC, Irwin RE (2006) Florivory: the intersection of pollination and herbivory. *Ecol Lett* 9:1351–1365
- McDade LA, Kinsman S (1980) The impact of floral parasitism in two Neotropical hummingbird—pollinated plant species. *Evolution* 34:944–958
- Miller-Struttman NE, Geib JC, Frankin JD, Kevan PG, Holdo RM et al (2015) Functional mismatch in a bumble bee pollination mutualism under climate change. *Science* 349:1541–1544
- Mitchell RJ, Karron JD, Holmquist KJ, Bell JM (2004) The influence of *Mimulus ringens* floral display size on pollinator visitation patterns. *Funct Ecol* 18:116–124
- Mitchell RJ, Irwin RE, Flanagan RJ, Karron JD (2009) Ecology and evolution of plant-pollinator interactions. *Ann Bot-Lond* 103:1355–1364
- Mu JP, Peng YH, Xi XQ, Wu XW, Griffin JN, Niklas KJ, Su SC (2014) Domesticated honeybees evolutionarily reduce flower nectar volume in a Tibetan Asteraceae. *Ecology* 95:3161–3172
- Mu JP, Peng YH, Xi XQ, Wu XW, Griffin JN, Niklas KJ, Sun SC (2015) Artificial asymmetric warming dramatically reduces nectar production in a Tibetan lotus. *Ann Bot-Lond* 116:899–906
- Nilsson LA (1988) The evolution of flowers with deep corolla tubes. *Nature* 334:147–149
- Pacini E, Nepi M (2007) Nectar production and presentation. In: Nicolson SW, Nepi M, Pacini E (eds) *Nectaries and nectar*. Springer, Dordrecht, pp 167–214
- Pacini E, Nepi M, Vesprini JL (2003) Nectar biodiversity: a short review. *Plant Syst Evol* 238:7–21
- Pellmyr O, Huth CJ (1994) Evolutionary stability of mutualism between yuccas and yucca moths. *Nature* 372:257–260
- Press CA, Phoenix GK (2005) Impacts of parasitic plants on natural communities. *New Phytol* 166:737–751
- Price PW, Fernandes GW, Waring GL (1987) Adaptive Nature of insect galls. *Environ Entomol* 16:15–24
- Pyke GH (1982) Foraging in bumblebees: rule of departure from an inflorescence. *Can J Zool* 60:417–428
- Pyke GH (2016) Floral nectar: pollinator attraction or manipulation. *Trends Ecol Evol* 31:339–341
- Real LA, Rathcke BJ (1991) Individual variation in nectar production and its effects on fitness in *Kalmia latifolia*. *Ecology* 72:149–155
- Rivera-Solís G, Abdala-Roberts L, Cervera JC, Parra-Tabla V, Ruiz-Ruiz J, Betancur-Ancona D (2012) Mechanisms and traits associated with compensation for defoliation in *Ruellia nudiflora*. *Plant Ecol* 213:303–314
- Rodríguez- Rodríguez MC, Jordano P, Valido A (2015) Hotspots of damage by antagonists shape the spatial structure of plant-pollinator interactions. *Ecology* 96:2181–2191
- Scaven V, Rafferty NE (2013) Physiological effects of climate warming on flowering plants and insect pollinator and potential consequences for their interaction. *Curr Zool* 59:418–426

- Scheiner R, Erber J, Page RE (1999) Tactile learning and the individual evaluation of the reward in honeybees (*Apis mellifera* L.). *J Comp Physiol A* 185:1–10
- Scheiner R, Page RE, Erber J (2001) Responsiveness to sucrose affects tactile and olfactory learning in preforaging honeybees of two genetic strains. *Behav Brain Res* 120:67–73
- Schemske DW, Bradshaw HD (1999) Pollinator preference and the evolution of floral traits in monkeyflowers (*Mimulus*). *Proc Natl Acad Sci USA* 96:11910–11915
- Smith LL, Lanza J, Smith GC (1990) Amino acid concentrations in extrafloral nectar of *Impatiens sultani* increase after stimulated herbivory. *Ecology* 71:107–115
- Strauss SY, Irwin RE (2004) Ecological and evolutionary consequences of multispecies plant–animal interactions. *Annu Rev Ecol Evol S* 35:435–466
- Straw NA (1989) Taxonomy, attack strategies and host relations in flowerhead Tephritidae: a review. *Ecol Entomol* 14:455–462
- Sun SG, Huang SQ, Guo YH (2013) Pollinator shift to managed honeybees enhances reproductive output in a bumblebee-pollinated plant. *Plant Syst Evol* 299:139–150
- Teuber LR, Barnes DK, Rincker CM (1983) Effectiveness of selection for nectar volume, receptacle diameter, and seed yield characteristics in Alfalfa. *Crop Sci* 23:283–289
- Thompson JN (2005) The geographic mosaic of coevolution. University of Chicago Press, Chicago
- Thomson JD (1988) Effects of variation in inflorescence size and floral rewards on the visitation rates of traplining pollinators of *Aralia hispida*. *Evol Ecol* 2:65–76
- Trumble JT (1993) Plant compensation for arthropod herbivory. *Annu Rev Entomol* 38:93–119
- Varley GC (1947) The natural control of population balance in the Knapweed Gall-Fly (*Urophora jaceana*). *J Anim Ecol* 16:139–187
- Vaudo AD, Tooker JF, Grozinger CM, Patch HM (2015) Bee nutrition and floral resource restoration. *Curr Opin Insect Sci* 10:133–141
- Vaughton G, Ramsey M (1998) Floral display, pollinator visitation and reproductive success in the dioecious perennial herb *Wurmbea dioica* (Liliaceae). *Oecologia* 115:93–101
- Wallis de Vries MF, van Swaay CAM, Plate CL (2012) Changes in nectar supply: a possible cause of widespread butterfly decline. *Curr Zool* 58:384–391
- Westerbergh A, Westerbergh J (2001) Interactions between seed predators/pollinators and their host plants: a first step towards mutualism. *Oikos* 95:324–334
- Whittall JB, Hodges SA (2007) Pollinator shifts drive increasingly long nectar spurs in columbine flowers. *Nature* 447:706–709
- Wäckers FL, Zuber D, Wunderlin R, Keller F (2001) The effect of herbivory on temporal and spatial dynamics of foliar nectar production in cotton and castor. *Ann Bot-Lond* 87:365–370
- Xi XQ, Wu XW, Nylin S, Sun SC (2016) Body size response to warming: time of the season matters in tephritid fly. *Oikos* 125:386–394
- Yoshida S, Forno D, Cock J, Gomez KA (1976) Laboratory manual for physiological studies of rice. International Rice Reserch Institute, Los Baños, Laguna, pp 46