



Plant-based therapies for urolithiasis: a systematic review of clinical and preclinical studies

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Abstract

Purpose Urolithiasis, the formation of kidney stones, is a common and severe condition. Despite advances in understanding its pathophysiology, affordable treatment options are needed worldwide. Hence, the interest is in herbal medicines as alternative or supplementary therapy for urinary stone disease. This review explores the use of plant extracts and phytochemicals in preventing and treating urolithiasis.

Methods Following PRISMA standards, we systematically reviewed the literature on PubMed/Medline, focusing on herbal items evaluated in in vivo models, in vitro studies, and clinical trials related to nephrolithiasis/urolithiasis. We searched English language publications from January 2021 to December 2023. Studies assessing plant extracts and phytochemicals' therapeutic potential in urolithiasis were included. Data extracted included study design, stone type, plant type, part of plant used, solvent type, main findings, and study references.

Results A total of 64 studies were included. Most studies used ethylene glycol to induce hyperoxaluria and nephrolithiasis in rat models. Various extraction methods were used to extract bioactive compounds from different plant parts. Several plants and phytochemicals, including *Alhagi maurorum*, *Aerva lanata*, *Dolichos biflorus*, *Cucumis melo*, and quercetin, demonstrated potential effectiveness in reducing stone formation, size, and number.

Conclusions Natural substances offer an alternative or supplementary approach to current treatments, potentially reducing pain and improving the quality of life for urolithiasis patients. However, further research is needed to clarify their mechanisms of action and optimize their therapeutic use. The potential of plant-based therapies in treating urolithiasis is promising, and ongoing research is expected to lead to treatment advancements benefiting patients globally.

Keywords Nephrolithiasis · Kidney stones · Calcium oxalate · Phytochemicals · Urinary calculi

Introduction

Kidney stones or urolithiasis results from the buildup of solid mineral and salt deposits in the urinary system or kidneys. These deposits can induce severe pain and discomfort; potential problems, such as infection or urinary tract blockage, could arise if they are not addressed. The incidence and the prevalence of urolithiasis exhibit significant variation across different populations. According to some

estimations, this condition might impact as much as 10–15% of the world's population [1].

The burden of urolithiasis is substantial in terms of its impact on individual patients and its broader economic and healthcare costs. Kidney stones can cause severe pain and discomfort, often necessitating hospitalization or surgical intervention for removal. Beyond the direct costs of treatment, urolithiasis can also result in lost productivity and diminished quality of life for affected individuals. Moreover, emerging evidence links nephrolithiasis to an increased risk of chronic kidney disease (CKD) [2].

The etiology of urolithiasis is sophisticated and remains incompletely understood. The production of kidney stones is believed to be motivated by a mixture of factors, such as nutrition, genetics, underlying medical conditions, and specific drugs. Stones frequently result when urine gets concentrated, facilitating the crystallization and adhesion of

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minerals. After their formation, these crystals can undergo further growth, eventually forming more giant stones that can provoke pain and other symptoms [3].

Also, present therapies for urolithiasis incorporate both medicinal and surgical methods. Petite stones may pass naturally with pain medication and increased fluid consumption in certain circumstances. Nonetheless, larger stones or those causing troubles may demand more intrusive therapies such as shock wave lithotripsy or ureteroscopy for removal. While these therapies can effectively remove existing stones, recurrence is expected, and they do not state the fundamental causes of stone formation.

There are drawbacks to current treatments for urolithiasis. For instance, surgical interventions carry risks such as infection or bleeding and may not be appropriate for all patients. Medical treatments such as thiazide diuretics or potassium citrate can help counteract stone recurrence in some cases but may not be helpful for all types of stones. Furthermore, these treatments need continuing checking and may have side effects [4].

Given these limitations, there is considerable interest in using plant extracts and phytochemicals to prevent and treat urolithiasis. Many herbs have been traditionally utilized for their diuretic or antispasmodic qualities, which may help facilitate the transit of kidney stones. Additionally, several plant chemicals have been proven to decrease the crystallization or aggregation of minerals in the urine, potentially reducing the likelihood of stone formation [5]. Urolithiasis has traditionally been treated using a variety of herbal remedies, including *Crataeva magna*, *Aerva javanica*, *Ipomoea eriocarpa*, *Peperomia tetraphylla*, *Punica granatum*, *Terminalia bellirica*, *Hibiscus rosa-sinensis*, *Moringa oleifera*, and *Costus spiralis* [6]. To dissolve kidney stones and stop them from coming back, people have turned to anti-urolithiatic herbs in various forms, such as decoction, infusion, or juice. There are fewer side effects and lower costs when using medicinal plants, but their efficacy is lower, and the period of therapy is longer. Additional scientific investigations are required to investigate safe and natural anti-urolithiatic substances based on the ethnopharmacological data that is now accessible [7]. Phytochemicals contain complex molecular structures that work across various metabolic pathways to deliver desired medicinal effects. Some of these secondary metabolites are bioactive, with high selectivity for cellular targets. In contrast, some metabolites have several cellular targets that may cooperate to produce a specific biological activity. Also, phytochemicals can create biological activity through synergistic processes [8].

Current evidence-based guidelines for managing urolithiasis, such as those from the European Association of Urology (EAU), recommend increasing fluid intake, maintaining a balanced diet, and engaging in regular physical

activity to prevent stone formation [9]. In addition, weight management is emphasized as a crucial factor in reducing the risk of stone recurrence [10]. While these lifestyle modifications are effective, they may only be sufficient for some patients, particularly those with recurrent stones or underlying metabolic disorders.

Herbal treatments can provide specific bioactive compounds that inhibit stone formation, reduce oxidative stress, and improve renal function. By integrating herbal treatments with conventional recommendations, we can offer a more comprehensive approach to managing urolithiasis. This combined strategy may enhance patient outcomes, particularly for those who do not respond adequately to lifestyle modifications alone.

In conclusion, urolithiasis is a frequent and significant disorder that can cause considerable pain and discomfort for affected individuals. While current therapies can help relieve existing stones, there are limits to these techniques, and recurrence is likely. This systematic review outlines the utilization of plant extracts and phytochemicals as a potential field of research for the prevention and treatment of urolithiasis to give specific references for further study.

Methodology

Search strategy

A comprehensive literature search was conducted using five databases: PubMed, Web of Science, Cochrane Database of Systematic Reviews, Medline, and Scopus. The search was limited to articles published in English between January 2021 and December 2023, as shown in Fig. 1. The search strategy aimed to include all research articles, whether *in vivo*, *in vitro*, or clinical trial studies. Plant names were verified using the Plant List and Royal Botanical Garden, Kew databases. The search included the following keywords: Plants OR Phytotherapy OR Pharmacognosy OR Ethnopharmacology OR Dietary Phytochemical OR Plant Bioactive Compound OR Plant-Derived Chemical OR Bioactive Compounds OR Plant OR Phytonutrient OR extract OR leaves OR seeds AND Nephrolithiasis OR Urolithiasis OR Kidney Calculi OR Urinary Lithiasis OR Ureterolithiasis OR Urinary Calculi OR Ureteral Calculi OR Urinary Bladder Calculi OR Kidney stone AND Treatment OR Therapeutic OR Therapy OR Therapies OR Prophylaxis OR Preventive therapy OR Prevention OR Control.

Study selection

Two authors independently screened the titles and abstracts of all articles identified from the search. Full-text articles were then assessed for eligibility based on the inclusion

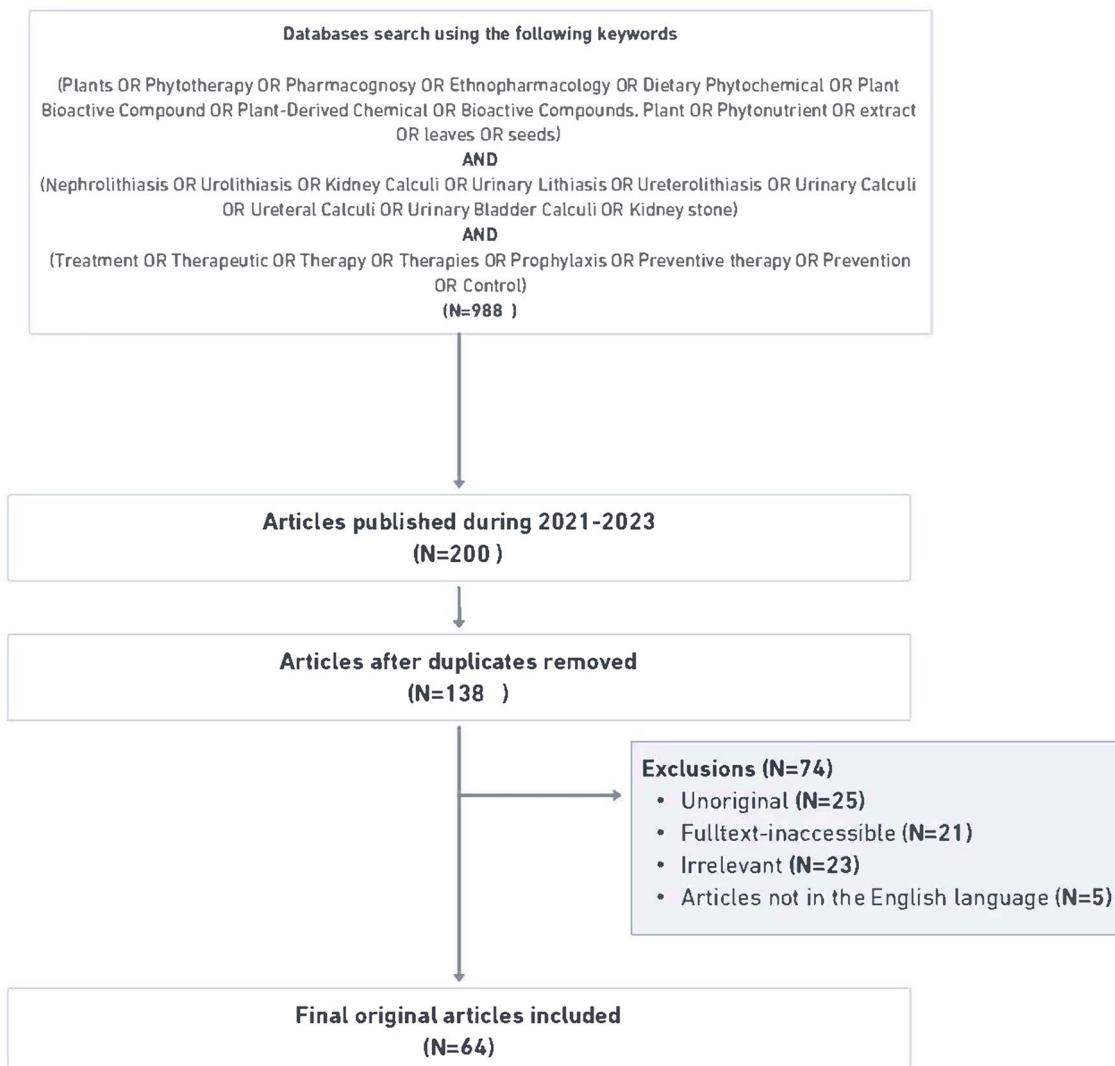


Fig. 1 Search parameters and criteria. The schematic illustration summarizes all keywords used for database searches and criteria to retrieve articles for discussion in this review

and exclusion criteria. Any disagreements were resolved through discussion. The inclusion criteria were studies that investigated the use of plants, dietary phytochemicals, phytotherapy, or plant bioactive compounds for the prevention or treatment of urolithiasis in *in vivo*, *in vitro*, or clinical trials. The exclusion criteria were non-original articles, duplicate publications, articles with inaccessible full text, irrelevant articles, studies focused on risk factors and mechanisms of urolithiasis without investigating anti-urolithic effects, and articles not in English.

Data extraction

Data extraction was performed independently by two authors using a standardized form. The extracted data included study design, type of stone, plant species, plant part used, solvent

type, main findings, and study reference. Discrepancies were resolved through discussion. Data were analyzed using Microsoft Excel to summarize and synthesize the findings.

Data synthesis

Data were analyzed using the Excel program to summarize the data.

Results

The initial literature search used specified terms to identify 200 articles. After removing 62 duplicates, 138 articles remained. Upon further screening and reading, 21 articles were excluded due to the unavailability of full

text. Additionally, 53 articles were excluded as they were either review articles, not written in English, or focused on mechanisms of urolithiasis risk factors without investigating anti-urolithic effects. Ultimately, 64 publications were included in this systematic review.

Our review identified several studies that focused on different types of kidney stones, including calcium oxalate, uric acid, and infectious stones (struvite stones). The majority of studies investigated the effects of herbal treatments on calcium oxalate stones, which are the most common type. However, we also found evidence supporting the efficacy of herbal remedies in managing other stone types.

For instance, uric acid stones, which form in acidic urine, may be prevented by plant extracts that alkalinize the urine. Plants like *Cucumis melo var. inodorus* have shown potential in increasing urinary pH, thereby reducing the risk of uric acid stone formation [16]. Similarly, infectious stones, often associated with urinary tract infections, may benefit from the antimicrobial properties of certain herbs. For example, *Mentha piperita* has demonstrated antibacterial activity against common uropathogens, which can help prevent the formation of struvite stones [75].

The relevant information from all suitable articles was extracted and organized into tables and figures. The retrieved data encompassed details such as the study's methodology, the composition of the stone, the specific plant utilized, the plant part employed, the solvent type used for extraction, the nature of the study (in vivo, in vitro, clinical trial), the primary findings, and the study's reference.

Tables 1 and 2 demonstrate experimental signs on plants that prevent and treat urolithiasis in in vivo and in vitro studies, respectively. Table 3 exhibits clinical evidence of plants used to prevent and treat urolithiasis, while Table 4 summarizes the phytochemicals of different plants used for the same purpose.

Tables 1, 2, 3, and 4 present a comprehensive collection of pharmacological investigations on different types of extracts and formulations of medicinal plants and phytochemical substances, including aqueous, hydroalcoholic, alcoholic, and other varieties. The tables contain data on the cited study, the plant's scientific name, the specific plant part employed, the kind of extract, the type of stones, the crystal-inducing agent/model, and the anti-urolithic activity/mechanism. The herbal extracts were found to exhibit anti-urolithic actions, as evidenced by the following effects: decreased crystal deposition, reduced oxidative stress, improved renal morphology, changes in urinary pH, decreased levels of lithogenic factors in urine, such as oxalate, calcium, and phosphate, improved renal function, increased urinary citrate, and altered protein expression.

The majority of studies used 0.75–1% ethylene glycol (EG) in drinking water alone ($n=16$, 25%) [14, 16, 19,

22, 34] or in combination with ammonium chloride (AC) ($n=15$, 23.43%) [11–13, 18, 26] (Fig. 2) to provoke calcium oxalate (CaOx) nephrolithiasis. A small number of studies retained other methods, such as intraperitoneal injection of sodium oxalate (NaOx) in Wistar rats or *Drosophila* [21, 24], implantation of zinc disks into the bladder of rats [25], or feeding rats with 3% glycolic acid mixed with food for seven days [27]. Additionally, certain plants were the subject of multiple studies, in contrast to other plants, while the majority of stones utilized were CaOx ($n=54$, 84.4%) (Figs. 3 and 4).

The most common studies used aqueous extracts ($n=20$, 31.25%) [21–23, 42] and leaves ($n=23$, 35.9%) [12, 18, 20, 37, 44] were the widely utilized herbal preparations, as shown in Figs. 5 and 6. Other botanical parts utilized involved fruits ($n=6$, 9.4%) [13, 15, 22, 28], roots ($n=4$, 6.25%) [31, 57], tea preparation [46], peel and pulp [40], pits [42], citrus waste peel [49], seeds ($n=12$, 18.75%) [16, 19, 67], rhizomes [65], stems [14], and flowers ($n=4$, 6.25%) [29]. Some studies utilized whole plants ($n=18$, 28.1%) [21, 23, 24, 33, 35], herbal medications (Ningmitai capsule) [68], Daidzin (isoflavone compound) [70], and poly-herbal formulations such as Safoof-e-Pathar Phori [11].

Other extraction methods included hydroalcoholic ($n=7$, 10.9%) [15, 28, 30], methanolic ($n=10$, 15.6%) [17, 19, 20], or ethanolic solvents ($n=18$, 28.125%) [12, 13, 16, 18, 24, 29], although some studies applied formulations/constituents like triterpenoids extracted from plants [74] (Fig. 5). Numerous clinical trials have been conducted to explore the efficacy of various herbal treatments for urolithiasis. This systematic review inspected three clinical trials. In a randomized, single-blind clinical trial by Aryaeefar et al. (2022), a total of 126 patients with ureteral stones (0–10 mm) were randomly split into a control group and an intervention group that administered whole plant distillate of *Alhagi maurorum* for four weeks. Even though an insignificant difference in stone size or placement was detected between the groups, the time essential for stone removal was markedly shorter in the intervention group. Furthermore, a randomized, single-blinded study by Shakeri et al. (2022) evaluated the effects of *Nigella sativa* seeds and tamsulosin in 80 patients with kidney and ureteral calculi (4–10 mm). The two groups displayed a reduction in stone size and number, with a more considerable decrease in pain score noted in the *Nigella sativa* group. Wang et al. (2022) conducted a randomized clinical trial with 123 patients diagnosed with urinary stones (10–20 mm), where the Ningmitai capsule (a herbal formulation) group displayed significantly higher stone expulsion rates, stone-free rates, and shorter duration to complete a stone-free state compared to the control group.

The review systematically investigated six studies that searched the effects of plant-based compounds on kidney

Table 1 In vivo investigations on the therapeutic benefits of Medicinal Plants against urolithiasis

In vivo		Plant	Part of the plant used	Study type	Type of stones	Study design	Main results	References
In vivo	In vitro							
Safoof-e-Pathar Phori consisted of (<i>Didymocarpous pedicellatus</i> R.Br (Gesneriaceae), <i>Macroyloma biflorum</i> var. <i>biflorum</i> (Leguminosae), <i>Rheum webbianum</i> Royle (Polygonaceae), <i>Hordeum vulgare</i> Linn. (Poaceae), <i>Raphanus raphanistrum</i> subsp. <i>sativus</i> (L.) Domínguez (Brassicaceae), and <i>potassium nitrate</i>)	<i>Didymocarpous</i> leaves, <i>Macrolyloma</i> seeds, <i>Rheum</i> rhizome, <i>Hordeum</i> whole plant, and <i>Raphanus</i> whole plant finely powdered	In vivo	CaOx crystal	EG + AC-induced UL in male Wistar albino Rats	The application of treatment at dosages of 700 mg/kg led to a notable reduction in Ca ²⁺ , serum creatinine (SCr), blood urea nitrogen (BUN) levels, and lipid peroxidation (LPO)	[11]		
<i>Platanus orientalis</i> (Platanaceae)	Ethanolic extract of leaves	In vivo	CaOx crystal	EG + AC-induced UL in rats	Effectively, the extract reinstated the levels of Ca ²⁺ , citrate in the urine sample, superoxide dismutase (SOD) activity, and glutathione (GSH) levels in the kidney sample of the EG + AC group to their normal state. Simultaneously, it rectified the heightened levels of oxalate, myeloperoxidase, caspase-3, N-acetyl-β-D-glycosaminidase (NAG) activities, malondialdehyde (MDA), 8-hydroxy-2'-deoxyguanosine (8-OHDG), Tumor necrosis factor-α (TNF-α), and interleukine-1-beta (IL-1β) in the kidney sample of the same group. Remarkably, a histo-pathological examination further demonstrated the extract's efficacy by indicating a regression of tubule expansion in both the cortex and medulla	[12]		
<i>Prunus cerasoides</i> (Rosaceae)	The ethanolic extract of dried fruits	In vivo	CaOx crystal	EG + AC-induced UL in rats	The study observed ↑ in relative body weight, accompanied by a significant prevention of elevated urinary levels of Ca ²⁺ , phosphate, oxalate, uric acid, and protein. Simultaneously, there were ↓ in the levels of Mg ²⁺ and citrate. Interestingly, the study also found a mitigated ↑ in BUN, Cr, and uric acid serum levels. In addition, there was ↓ in kidney weight, tissue damage, inflammation, and tubular dilation	[13]		
		In vitro	CaOx crystal in aqueous solution		The study observed a substantial ↓ in CaOx crystal size. Interestingly, crystals that were initially hexagonal or monoclinic calcium oxalate monohydrate (COM) transformed into octahedral-shaped calcium oxalate dihydrate (COD) crystals			

Table 1 (continued)

Plant	In vivo	Part of the plant used	Study type	Type of stones	Study design	Main results	References
<i>Ficus tikoua Bur. (Moraceae)</i>	The stem extract	In vivo	CaOx crystal	EG-induced UL in rats		The study demonstrated restoring lymphocyte % in the blood and the concentration of salt, chlorine, and inorganic phosphates back to normal levels. Diuresis was maintained at the control level of the animals. Interestingly, no effect was observed on the urine urobilogen concentration or ↑ in erythrocytes. However, the histo-pathological investigation did not indicate the efficacy of the treatment	[14]
<i>Cucumis callousus (Rottl.) Cogn (Cucurbitaceae)</i>	Hydro-ethanolic extract of fruits	In vivo	CaOx crystal	EG+AC-induced UL in rats		The study noticed ↓ urine oxalate levels, volume, and urinary urea nitrogen (UUN). Conversely, there was ↑ urine Ca ²⁺ and pH. Furthermore, there was ↓ in BUN, Cr, and total protein. The study also observed ↓ the oxidant enzyme, lipid peroxidase, and ↑ GSH and catalase (CAT) levels. Furthermore, ↓ CaOx crystal deposition led to ↓ inflammation and renal damage in the hyperoxaluric rat kidney. Lastly, the study found ↓ the expression of osteopontin (OPN)	[15]
<i>Cucumis melo var. inodorus (Cucurbitaceae)</i>	Ethanolic extract of seeds	In vivo	CaOx crystal	EG-induced UL in rats		The study reported ↓ the kidney index, urinary Ca ²⁺ and oxalate levels, CaOx deposits number and score, and the extent of histo-pathological damages. It was also observed ↓ in the inflammation score in the kidney sections. On the other hand, there were ↑ urinary pH, Mg ²⁺ , and citrate levels. Furthermore, the expression of the spp1, UMOD, and reg1 genes in the kidneys of the treated animals was found to be elevated	[16]

Table 1 (continued)

Plant	In vivo	Part of the plant used	Study type	Type of stones	Study design	Main results	References
<i>Trachyspermum ammi</i> (L.) (Apiaceae)		Methanolic extract of seeds	In vivo	CaOx crystal	EG + AC-induced UL in rats	The extract led to ↑ urine excretion of Na^+ , although no substantial elevation was detected in the K^+ excretion. It halted the net loss in body weight, and both 24-h urine volume and water consumption were elevated. The extract considerably ↓ the urinary oxalate and restored the urinary Ca^{2+} to the average level. Interestingly, the urinary contents of citrate, phosphate, uric acid, and Mg^{2+} remained constant	[17]
			In vitro	CaOx crystal	CaOx crystal in aqueous solution	The study observed ↓ both slopes of nucleation and a concentration-dependent inhibition of crystal aggregation. There was also ↓ in CaOx crystal formation and size of COM. The study further noted an inhibition of lipid peroxidation, lactate dehydrogenase (LDH) release and DPPH-free radicals. Importantly, no toxic effects were observed on 2,2-diphenyl-1-picrylhydrazyl (MDCK) cells	[18]
<i>Calotropis procera</i> (Apocynaceae)	Ethanolic extract of leaves		In vivo	CaOx crystal	EG + AC-induced UL in rats	The n-hexane fraction was used for in vivo research. The extract led to ↓ in blood Mg^{2+} levels and Mg^{2+} , Ca^{2+} urine levels compared to the control group. There was a considerable rise in Cr concentration but no noticeable variation in urea or uric acid quantities. A considerable elevation in the activities of alanine aminotransferase (ALT) and Aspartate aminotransferase (AST) was noted, along with ↑ the kidney GSH and SOD activity and ↓ kidney MDA	[18]
			In vitro	CaOx crystal	CaOx crystal in aqueous solution	The study observed a gradual ↑ in the inhibition of DPPH, with the highest values recorded in the n-hexane fraction. Notably, the n-hexane fraction of the extract demonstrated the most potent inhibitory effect on stone nucleation	

Table 1 (continued)

Plant	In vivo	Part of the plant used	Study type	Type of stones	Study design	Main results	References
<i>Peganum harmala</i> L. (Nitrariaceae)	Methanolic extraction of seeds	In vivo	CaOx crystal	EG-induced UL in rats	In the treatment groups, there was a notable ↓ in serum toxicity indicators, such as BUN, Cr, urea, uric acid, Kidney Injury Molecule-1 (KIM-1), phosphate, Ca ²⁺ , MDA, Mg ²⁺ , and oxate levels. Similarly, inflammatory markers, including TNF- α and nuclear factor kappa B (NF- κ B), were downregulated in the kidney homogenate. Concurrently, urine production and urine pH were significantly ↑. The treatment facilitated a gradual recovery in the injured glomeruli, interstitial spaces, medulla, and tubules, accompanied by ↓ in brown calculi materials	[19]	
<i>Myrtus communis</i> L. (Myrtaceae)	70% methanolic extract of leaves	In vivo	CaOx crystal	EG-induced UL in rats	The study found that the levels of various chemicals in the urine, including Ca ²⁺ , Cr, Mg ²⁺ , uric acid, citrate, and oxalate, were more aligned with those of the control group. Concurrently, water intake and urine output were ↓ while there was an ↑ in the body weights of rats when compared to the EG group. Despite noticeable tubular dilatation in the collecting tubules, as observed in hematoxylin-eosin staining, the sectional images bore more resemblance to the control group. In Pizzolato's staining, both the quantity and density of the crystals in the sections were ↓ in the extract group	[20]	
<i>Salvia miltiorrhiza</i> (danshen) (Lamiaceae), <i>Astragalus</i> (Huang qi) (Fabaceae), and <i>Carthami flos</i> (HongHua) (Asteraceae)	Aqueous extracts of whole plant	In vivo	CaOx crystal	NaOx-induced UL in Drosophila model	The study demonstrated that the extracts indeed possess therapeutic benefits, which were notably more potent than those of nilutamide, luteolin, and quercetin	[21]	
<i>Ziziphus lotus</i> (Rhamnaceae)	The aqueous fruit extract	In vivo	CaOx crystal	EG-induced UL in rats	The extract ↓ oxalate and Ca ²⁺ in the urine primarily expels tiny COD crystals. This was accompanied by an ↓ in urine output. Notably, the crystalluria consisted of minimal COM particles. The dimensions of these crystals were ↓, leading to an ↓ in renal weight relative to the EG group	[22]	

Table 1 (continued)

In vivo	Plant	Part of the plant used	Study type	Type of stones	Study design	Main results	References
	<i>Astragalus membranaceus</i> (Fisch.) (Leguminosae)	The aqueous extract of whole plant	In vivo	CaOx crystal	EG-induced UL in Drosophila model	The extract significantly ↓ the formation of CaOx crystals induced by EG, demonstrating a protective effect that extended the average survival days of <i>Drosophila</i> . However, in ex vivo tests, the extract did not impact the dissolution of CaOx crystals formed in the Malpighian tubule of <i>Drosophila</i>	[23]
<i>Cyperus rotundus L.</i> (Cyperaceae)	Ethanolic extract of the whole plant	In vivo	CaOx crystal	NaOx-induced UL in rats	The extract showed a significant ↓ in BUN, Cr, uric acid, urinary Na+, and chloride levels	[24]	
		In vitro	CaOx crystal	CaOx crystal in aqueous solution	The research observed that ↑ the concentration of the extract was directly correlated with the inhibition of both crystal formation and aggregation		
	<i>Mimosa malacophylla</i> (Fabaceae)	Methanolic extract of the aerial part	In vivo	CaOx crystal	Zinc disks-induced UL in rats	The extract did not induce any changes in the bladder, renal cortex, or medulla. Additionally, it did not trigger any cytotoxic activity or lead to morphological abnormalities in the nucleus of standard kidney cell lines	[25]
	<i>Mentha piperita L.</i> (peppermint) (Lamiaceae)	The aqueous methanolic crude extract of fresh aerial parts	In vivo	CaOx crystal	EG+AC-induced Urolithiasis in rats	The extract ↑ urine output, Na ⁺ and K ⁺ excretion, and body weight. It also demonstrated a significant ↓ in the number of crystals and neutralized the acidic urine pH. The extract was significantly ↑ urinary Mg ²⁺ , and exhibited a notable ↓ in phosphate, uric acid, and total protein levels. It normalized Cr and BUN levels in a dose-dependent manner and restored MDA, GSH, and SOD levels. Moreover, the extract ↓ inflammation improved the integrity of the kidney epithelial membrane and normalized the interstitial gaps between cells	[26]
			In vitro	CaOx crystal	CaOx crystal in aqueous solution	The extract displayed antioxidant activity in a dose-dependent manner. Furthermore, the extract was observed to inhibit crystal nucleation, aggregation, and growth	

Table 1 (continued)

Plant	In vivo	Part of the plant used	Study type	Type of stones	Study design	Main results	References
<i>Caesalpinia bonducuella</i> (Caesalpiniaceae)	Ethanolic extract of seed	In vivo	CaOx crystal	EG, glycolic acid, and NaOx-induced different UL model		The extract treatment resulted in a significant ↑ in the body weight of the groups. Concurrently, there was a ↓ in the levels of urinary Ca^{2+} and phosphorus, while urinary Mg^{2+} levels saw an ↑. There was also an ↑ in urine volume, acidity, and pH levels. Blood biochemical markers, including uric acid, urea, Cr, BUN, and ALP, exhibited a notable ↓. The extract, supplied at dosages of 200 and 400 mg/kg, demonstrated that glomerular hypercellularity, casts and tubular hydropic degeneration were ameliorated in a dose-dependent manner in the kidney sections of the treated rats	[27]
<i>Musa balbisiana</i> (Musaceae)	Hydro-ethanolic extract from fruits	In vivo	CaOx crystal	EG-induced UL in rats		Histopathological evaluations revealed that the hydroethanolic extract treatment effectively counteracted the presence of CaOx crystal deposits in the renal tubules, as well as the congestion and dilation of these tubules. The extract also displayed diuretic properties, ↓ the growth of urinary stones, and led to an ↑ in the urinary excretion of Na^+ , K^+ , and Cl^- . At a high dosage of 1.6 g/kg, there was a ↓ in urine Ca^{2+} levels and an ↑ in Mg^{2+} concentration. The high dosage also resulted in ↓ serum phosphate, Cr, and urea levels	[28]
<i>Hibiscus rosa-sinensis</i> (Malvaceae)	Standardized ethanolic extract of flowers	In vitro	CaOx crystal	CaOx crystal in aqueous solution		All extracts derived from the fruits of <i>M. balbisiana</i> demonstrated in vitro properties that ↓ the formation and aggregation of uric crystals, ↓ inflammation, combat microbes, and act as antioxidants. Among these extracts, the hydroethanolic extract obtained via heat extraction exhibited the most potent effects	[29]

Table 1 (continued)

In vivo	Plant	Part of the plant used	Study type	Type of stones	Study design	Main results	References
	<i>Citrus medica, citrus limon and citrus Aurantium L.</i> (Rutaceae)	Hydroalcoholic extract of leaves of all three plants	In vivo	CaOx crystal	EG-induced UL in rats	The extracts showed a ↓ in urinary concentrations of Ca ²⁺ , phosphate, Mg ²⁺ , Cr, uric acid, protein, UUN, and oxalate, along with an ↑ in urinary output. A combined extract of all three plants, administered at a dose of 300 mg/kg, demonstrated a more substantial effect compared to the response from individual plant extracts	[30]
	<i>Aerva lanata (L.)</i> (Amaranthaceae)	Ethanolic extract of roots	In vivo	CaOx crystal	EG-induced UL in rats	The extract returned body weight to normal levels, ↑ urine volume and pH, and ↓ the excretion of urinary components, such as Ca ²⁺ , phosphorus, and oxalate. It also ↓ serum uric acid and Cr levels. Additionally, it ↓ crystal formation and size. Mild chronic interstitial inflammation was observed in both the conventional and extract therapy groups, with no signs of acute tubular damage. The group treated with a low dosage of the extract showed minimal signs of interstitial inflammation with lymphocytic infiltration. In contrast, the renal parenchyma of the group treated with a high dosage of the extract showed no significant pathological abnormalities, indicating that the 800 mg/kg extract mitigated kidney damage, thereby managing or preventing the disease	[31]
	<i>Persea Americana</i> (Lauraceae)	Methanolic seed extracts	In vitro	CaOx crystal	CaOx crystal in aqueous solution	The extract, at varying concentrations, significantly ↓ stone formation, as evidenced by an ↑ in the rate of absorbance. It also inhibited COM aggregation. Notably, a concentration of 100 µg/mL produced the lowest turbidity, greatest absorbance, and inhibited crystal formation	[32]

Table 1 (continued)

In vivo	Plant	Part of the plant used	Study type	Type of stones	Study design	Main results	References
	<i>Pyrrosia lingua</i> (Polypodiaceae)	Ethanolic extract of the whole plant	In vivo	CaOx crystal	EG + AC-induced UL in rats	In the group treated with a high dose of the extract, blood Ca ²⁺ and urine oxalic acid levels were ↓, while urine Ca ²⁺ levels were ↑. In the moderate and low-dose extract groups, only urine Ca ²⁺ and urine oxalate exhibited a ↓. The extract groups revealed a ↓ in the level of CaOx rat urine, indicating a dose-dependent impact. Nitric oxide levels were ↓, and SOD activity was ↑ in the renal tissues of rats in the low-dose extract group. In contrast, the levels of nitric oxide and MDA were ↓, while SOD activity was ↑ in the renal tissues of rats in the moderate and high-dose extract groups. The crystal deposits and degenerative scores were ↓ in the renal tissue of the rats in the extract groups in a dose-dependent manner. The expression of OPN was decreased in the kidney tissue and urine of rats in the extract groups, indicating a dose-dependent impact, with the high-dose extract group presenting the most significant changes. Furthermore, levels of <i>Bacteroides</i> , <i>Oxalobacter formigenes</i> , <i>Faecalibacterium</i> , and <i>Bifidobacterium</i> were noticeable ↓ following treatment with the extracts	[33]
	<i>Glechoma Herba</i> (Lamiaceae) from 10 different regions	Ethanolic extract of the whole plant	In vivo	CaOx crystal	EG-induced UL in rats	The blood levels of Cr and BUN in rats administered the extract from various regions were significantly ↓ than those in the diseased group. The MDA concentration in the blood of all the extract groups was definitely ↓, while the CAT and SOD levels ↑ dramatically. The extract was effective in alleviating the dilatation of renal tubules and inhibiting the infiltration of inflammatory cells. It also had a lower stone sedimentation score, indicating that the extract has an anti-inflammatory effect	[34]

Table 1 (continued)

In vivo		Plant	Part of the plant used	Study type	Type of stones	Study design	Main results	References
		<i>Pyrrosia perfoliata</i> (Polypodiaceae)	Ethanolic extract of the whole plant	In vivo	CaOx crystal	EG-induced UL in rats	The extract was found to significantly ↓ the levels of BUN, Cr, and Na ⁺ in serum, as well as 24-h oxalate, urinary protein, uric acid, Cr, Ca ²⁺ , and phosphorus in the urine. It also ↑ the urine volume in rats in a dose-dependent manner, although it did not affect the urine pH. Additionally, the extract considerably mitigated EG-induced damage to renal tissue. The extract notably ↑ the levels of SOD and GSH, while ↓ the MDA level and the expression of NADPH oxidases 2 (NOX2) and NOX4 in kidney tissue. Additionally, in EG-stimulated kidney tissue, the extract dramatically decreased the levels of IL-1 β , IL-6, TNF- α , and monocyte chemoattractant protein-1 (MCP-1). Additionally, in a concentration-dependent way, it suppressed the protein levels of EG-induced transforming growth factor- β 1 (TGF- β 1), p-Smad3, and p-Smad2.	[35]
		<i>Gomphocarpus fruticosus</i> (Asclepiadaceae)	Different solvents of the whole plant extract	In vivo	CaOx crystal	EG + AC-induced UL in rats	The ethyl acetate extracts significantly ↓ the level of Na ⁺ , but it ↑ the levels of Mg ²⁺ and citrate, compared to the lithiasic control group. The butanol extracts ↓ K ⁺ , Ca ²⁺ , and phosphate levels in urolithiasic rats. It was also found that the ethyl acetate extract ↓ the amount of oxalate in the urine, whereas the butanol extract ↑ the levels of Mg ²⁺ and citrate in serum tests. CaOx crystal formations were remarkably ↓ by the ethyl acetate extract in the kidneys. Treatment with ethyl acetate and butanol extracts resulted in large ↓ urine protein excretions. The petroleum ether extracts ↑ the serum AST level considerably compared to the healthy control. The butanol extracts greatly ↓ the level of AST compared to the lithiasic control group	[36]

Table 1 (continued)

In vivo	Plant	Part of the plant used	Study type	Type of stones	Study design	Main results	References
<i>Origanum vulgare (OV) Linn</i> (Lamiaceae)		Ethanol, aqueous and hexane extract of leaves	In vivo	CaOx crystal	EG + AC-induced UL in rats	In contrast to the untreated group, the treated group recovered from the extract right away. This was notably evidenced by weight gain and a significant ↓ in urinary oxalate, serum uric acid, urea, Cr, and renal crystal deposition. There was also an improvement in renal functions relative to the lithogenic group. Interestingly, the study indicated significant ↑ in blood pressure and heart rate in the EG + AC-treated group, but no significant changes were observed in both the extract and control groups	[37]
<i>Dolichos biflorus</i> (Fabaceae) and <i>Crataeva nurvala</i> (Capparidaceae)		<i>Dolichos biflorus</i> (D.b) (hydroalcoholic seed extract) and <i>Crataeva nurvala</i> (C.n) (aqueous bark extract) in two ratios 1:1 and 3:1	In vivo	CaOx crystal	EG + AC-induced UL in rats	The combination extracts significantly ↑ urine production compared to the UL group. When the urinary oxalate levels of rats treated with the combination extract were evaluated, in comparison to the values of the disease control group, there was no discernible difference. In the UL group, a higher Cr level was observed. Although in the standard and extract groups, it was elevated in the initial first two weeks, the findings were inconsequential ↓ in the last 2 weeks of the experiment. All the combination extracts appreciably ↓ the serum Ca ²⁺ levels. In a 3:1 ratio, the glomeruli maintained the complete morphology of the glomerulus, and there was less injury to the tubules and other areas of the tissue	[38]
<i>Argemone mexicana L.</i> (Papaveraceae)		Methanol extract from leaves	In vivo	CaOx crystal	EG-induced UL in rats	Rats administered the extract showed steady ↑ body weight from the second week onwards, and their urine color transitioned from red to yellow. The extract-treated rats exhibited an ↑ in urine volume and pH. Additionally, there was a ↓ in serum Ca ²⁺ , urea, and Cr levels in both male and female rats. The extract demonstrated a ↓ in crystal formation. The renal weight and size of the rats were ↓, and accumulated stones were eliminated with an ↑ in diuresis following exposure to the extract. The extract also repaired basophilic cells, degeneration cells, and regenerative cells	[39]

Table 1 (continued)

In vivo	Plant	Part of the plant used	Study type	Type of stones	Study design	Main results	References
	<i>Cucumis melo</i> L. (Cucurbitaceae)	Different solvent extracts of peel and pulp	In vivo	CaOx crystal	EG-induced UL in rats	Among all the peel extracts, the study discovered that even in comparison to the positive control, the chloroform and methanol extracts demonstrated a superior ability to ↓ renal calculi. Levels of serum Cr, uric acid, and BUN evidenced this. Conversely, only the chloroform extract showed a significant improvement in blood Cr and uric acid levels among all the pulp extracts	[40]
	<i>Cassia auriculata</i> (Fabaceae)	Ethanolic extract of the whole plant	In vivo	CaOx crystal	EG+AC-induced UL in rats	Treatment with the extract was reported to provide a dosage-dependent anti-urolithic effect. This was evidenced by ↓ serum Ca ²⁺ , Cr, and uric acid levels, along with an ↑ in urine volume. Additionally, there was a ↓ in urine Ca ²⁺ , phosphate, and oxalate levels. SOD and CAT levels were ↑, coupled with a considerable ↓ in MDA levels	[41]
	<i>Date Palm</i> (<i>Phoenix dactylifera</i> L.) (Arecaceae)	Aqueous extract of pits	In vivo	CaOx crystal	EG-induced UL in rats	The administration of date palm pit extracts in both the treatment and preventive groups at 300 mg/kg significantly ↓ the levels of BUN, uric acid, Ca ²⁺ , Cr, and phosphorus. The urine data findings indicate that the extract administration led to a substantial ↓ in Cr, uric acid, and Ca ²⁺ in the preventative group and a significant ↓ in Cr and uric acid in the therapy group with a dose of 300 mg/kg. The pathological findings indicate a dose-dependent decrease in the incidence and size of CaOx crystals in renal tubules in both the preventive and treated groups	[42]

LDH: lactate dehydrogenase, *LPO*: lipid peroxidation, *MCP-1*: chemoattractant protein-1 (MCP-1), *MDA*: malondialdehyde, *NAC*: *N*-acetyl-β-D-glycosaminidase, *NF-κB*: nuclear factor kappa B, *OPN*: osteopontin, *SCr*: serum creatinine, *SOD*: superoxide dismutase, *TGF-β1*: transforming growth factor-β1, *TNF-α*: tumor necrosis factor-α, *UL*: urolithiasis, *UUN*: urinary urea nitrogen

↑: increased; ↓: decreased; 8-OHdG: 8-hydroxy-2'-deoxyguanosine, AC: ammonium chloride ACP: acid phosphatase ALP: alkaline phosphatase AST: aspartate aminotransferase, BUN: blood urea nitrogen, CaOx: Calcium oxalate, CAT: catalase, COD: calcium oxalate monohydrate, COM: calcium oxalate dihydrate, Cr: creatinine, EG: ethylene glycol, GSII: reduced glutathione, IL-1β: interleukine-1 beta, KIM-1: Kidney Injury Molecule-1

Table 2 In vitro studies on the therapeutic effects of Medicinal Plants against urolithiasis

In vitro	Plant	Part of the plant used	Study type	Type of stones	Study design	Main results	References
	<i>Paronychia argentea Lam</i> (Rejel El-Hamama) (Caryophyllaceae), <i>Tecurium polium L.</i> (Jaa'deh)	Aqueous extract of aerial parts of <i>P. argentea</i> , <i>T. polium</i> , and leaves of <i>C. aronia</i> .	In vitro	CaOx	CaOx crystal in aqueous solution	The study found that all plant extracts could inhibit nucleation, regardless of the dosage. Interestingly, the extract from <i>V. iphionoides</i> demonstrated the most potent aggregation inhibition at a low concentration	[43]
	<i>Altagi maurorum Medik.</i> (Aqool" or "Shook El-Jamal) (Fabaceae), <i>Varrhemia iphionoides</i> A.P. (Kitilh" or "Shtilh)	"Shook El-Jamal" (Fabaceae), <i>Varrhemia iphionoides</i> A.P. (Kitilh" or "Shtilh)					
	<i>Aronia L.</i> (Rosaceae)	Leaves aqueous extract	In vitro	CaOx	CaOx crystal in aqueous solution	The extract dissolved kidney stones, reduced pH to 5–6, and lessened stone mass	[44]
	<i>Altagi maurorum</i> (Boiss.), (Leguminosae)	The ethanolic extract of whole plant	In vitro	CaOx	CaOx crystal in aqueous solution	The extract efficiently catalyzed the mineralization of CaOx in a dose-dependent manner	[45]
	<i>Alternanthera sessilis</i> (Amaranthaceae)	The root is used as a crumbled herb for tea preparation	In vitro	Various kidney stones from patients	Various kidney stones in <i>O. spinosa</i> extract solution	Spectrophotometric analysis revealed a statistically significant passage of Ca ²⁺ , phosphate, and uric acid through the tea preparation solution	[46]
	<i>Ononis spinosa</i> L. (Fabaceae)						
	<i>Enhydra fluctuans</i> Lour. (Asteraceae)	The aqueous extract of whole plant	In vitro	Calcium phosphate crystals	Ca phosphate crystal in aqueous solution	The extract exhibited inhibitory effects on the development of brushite crystals, ↓ the average length of deposited brushite crystals and demonstrated antibacterial and antioxidant properties	[47]
	<i>Melia azedarach</i> L. (Meliaceae)	Chloroform, Methanolic and aqueous extract of fruit-seeds	In vitro	CaOx	CaOx crystal in aqueous solution	All the extracts exhibited an ↑ in value for the incubation time ↑. The chloroform extract was more potent than methanol and aqueous extracts in dissolving Ca ²⁺ crystals and nucleation assay	[48]

Table 2 (continued)

In vitro	Plant	Part of the plant used	Study type	Type of stones	Study design	Main results	References
<i>Citrus limetta</i> , <i>Citrus limon</i> and <i>Citrus sinensis</i> (Rutaceae)		Hexane, aqueous and ethanol extract of citrus waste peel	In vitro	CaOx	CaOx crystal in aqueous solution	The ethanolic extract of C. sinensis peels significantly suppressed CaOx nucleation, growth, and aggregation at a concentration of 1000 µg/mL, demonstrating superior efficacy compared to other solvents and plants	[49]
<i>Bryophyllum pinnatum</i> (BPE) (Crassulaceae) and <i>Macrotyloma uniflorum</i> (MUE) (Fabaceae)		Hydroalcoholic extracts of leaves and seeds of BPE and MUE, respectively	In vitro	COM crystal	COM crystal-injured cells	The extracts significantly ↑ Vero cells viability that were exposed to COM crystal in a dose-dependent manner, showing maximal efficacy at 200 µg/ml. Macrotyloma uniflorum exhibited superior wound healing efficacy than BPE. Additionally, the extracts demonstrated antioxidant activity	[50]
<i>Drymoglossum piloselloides</i> (Polypodiaceae), <i>Aegle marmelos</i> (Rutaceae) and <i>Kalanchoe laciniata</i> Crassulaceae)		The aqueous, ethanol, and hexane extracts of <i>Drymoglossum piloselloides</i> leaves, <i>Kalanchoe laciniata</i> leaves, and <i>Aegle marmelos</i> flowers	In vitro	CaOx crystal	CaOx crystal in aqueous solution	All the extracts exhibited a dosage-dependent ability to inhibit the nucleation, growth, and aggregation of CaOx crystals	[51]
<i>Basella rubra</i> (Basellaceae)		The hydroalcoholic extracts of leaves and stem pod	In vitro	CaOx crystal	CaOx crystal in aqueous solution	The extracts demonstrated a significant ↓ in the weight of the CaOx tablet, and the stem pod extract has the highest percentage solubility of the tablet	[52]
The tea bag was composed of (<i>Dolichos biflorus</i> (Fabaceae), <i>Phyllanthus emblica</i> L. (Euphorbiaceae), <i>Ocimum tenuiflorum</i> L. (Lamiaceae), <i>Green Tea</i> , <i>Withania somnifera</i> (Solanaceae), <i>Foeniculum vulgare</i> Mill. (Apiaceae) and <i>Stevia</i> (Asteraceae))		Aqueous extract of Seeds, fruit, leaves, leaves, root powder, Seed powder, and leaves, respectively	In vitro	CaOx crystal	CaOx crystal in the urine of healthy subjects	Smaller crystals and a decrease in crystal density were the results of increasing dosages of the study formulation. A dose-dependent % inhibition was detected in the DPPH assay, indicating antioxidant activity in both the reducing power assay and hydrogen peroxide scavenging activity	[53]

Table 2 (continued)

In vitro	Plant	Part of the plant used	Study type	Type of stones	Study design	Main results	References
	<i>Bryophyllum pinnatum</i> (Crassulaceae) and <i>Aerva lanata</i> (Amaranthaceae)	Ethyl acetate extract of Fresh leaves of <i>Bryophyllum pinnatum</i> and flowers of <i>Aerva lanata</i>	In vitro	CaOx crystal	CaOx crystal in aqueous solution	The maximum dissolution of CaOx was observed in the extract of <i>Bryophyllum pinnatum</i> at a concentration of 10 mg, which may be attributed to Bufadienolides rather than <i>Aerva lanata</i>	[54]
	<i>Saussurea costus (Falc) Lipsch</i> (Asteraceae)	Aqueous and Ethanolic extract of roots	In vitro	Cystine stone and CaOx crystal in aqueous solution	Cystine stone and CaOx crystal in aqueous solution	<p>The dissolution of cystine stones ↑ over time, from the first to the last week, with the aqueous extract showing the most significant effect.</p> <p>The dissolution process was independent of the cystine calculi's pH for the ethanolic extract. This plant's ethanolic extract significantly inhibited the crystallization of CaOx.</p> <p>The ethanolic extract had the greatest IC50, according to the DPPH technique, with a value of $IC50 = 0.12325$ ng/ml. However, the FRAP method showed that the aqueous extract, with a 300 µg/ml concentration, possessed the most excellent reducing power</p>	[55]
	<i>Garcinia humilis</i> (Clusiaceae)	Methanolic, dichloromethane, and ethyl acetate fractions obtained from the leaves	In vitro	CaOx crystal	CaOx crystal in urine	All the different preparations could produce an anti-urolithic action in vitro and ↓ the quantity of COM and COD crystals	[56]

Table 2 (continued)

In vitro	Plant	Part of the plant used	Study type	Type of stones	Study design	Main results	References
<i>Pleurolobus gangeticus (L.) J. St.- Hil. ex H. Ohashi & K. Ohasi</i> (Fabaceae)		Chloroform extract of fresh roots	In vitro	CaOx crystal	In vitro CaOx crystal	The findings showed that the production of CaOx crystals was impacted in a dose-dependent manner by the chloroform fraction. The maximum dose of 10 mg/mL in both nucleation and aggregation studies demonstrated superior results. This concentration resulted in considerable ↑ in CaOx crystal nucleation, a reduction in crystal size and the suppression of crystal aggregation	[57]
<i>Sida acuta Burm. F.</i> (Malvaceae)		Ethanolic extract of leaves	In vitro	Struvite crystal	Struvite crystal hydrogel medium	The extract, at various dosages, significantly ↓ the average weights of struvite crystals in a dose-dependent manner	[58]
<i>Orthosiphon stamineus</i> (Lamiaceae)		The standardized water extract of the whole plant in different concentration	In vitro	CaOx and uric acid stones	Stones from patients immersed in the plant extract solution	The maximum percentage weight decreases of CaOx stone occurred at a dosage of 4 mg/ml. The <i>O. stamineus</i> extract at 4 mg/ml demonstrated a more significant chemolytic activity on CaOx stone than the potassium citrate solution, with 70% effectiveness at pH 5, 48% at pH 7, and less than 10% at pH 8. The percentage weight decrease of uric acid stones was found to be 47%, 11%, and 14% at pH 5, 7, and 8, respectively. The data analysis demonstrated that the % weight reductions of combination stones differed considerably across acidic, neutral, and alkaline conditions	[59]

Table 2 (continued)

In vitro	Plant	Part of the plant used	Study type	Type of stones	Study design	Main results	References
	<i>Herniaria hirsuta L.</i> (Caryophyllaceae), <i>Opuntia ficus-indica</i> (Cactaceae), <i>Zea mays</i> (Poaceae) and <i>Ammi visnaga L</i> (Apiaceae)	Aqueous extract of <i>H. hirsuta</i> L., fully used (leaves and stems); <i>O. ficus-indica</i> flowers, <i>A. visnaga L.</i> seeds and very fine filaments that come from the outer shell corn cobs <i>Z. mays</i> styles	In vitro	CaOx crystal	In vitro CaOx crystal in extract solution	The comparison of the stone dissolution rates for the four plants suggests that <i>Zea mays</i> may exert a slightly more pronounced effect than the others	[60]
	<i>Ocimum sanctum</i> (Lamiaceae)	Hydroalcoholic extract of leaves	In vitro	CaOx crystal	CaOx crystal solution	A 40.32% suppression of CaOx nucleation was reported with the extract's maximal concentration (8 mg/ml). Microscopic analysis showed that the extract decreased the number and size of crystal nuclei as concentration increased. The highest aggregation inhibition (33.9%) was seen at a dosage of 1 mg/ml. Significant suppression of crystal growth was reported at 1 mg/ml and 2 mg/ml of the extract, with 94.59% and 97.21%, respectively	[61]
	<i>Phyllanthus niruri L.</i> (Phyllanthaceae)	Different solvent leaves extract	In vitro	CaOx crystal	CaOx crystal solution	The methanolic extract exhibited the highest inhibition against the aggregation of CaOx crystals. The aqueous extract was shown to be more effective in the dissolution of CaOx. The nucleation rate, aggregation of CaOx crystallization, and crystal density were all markedly inhibited by <i>P. niruri</i>	[62]

Table 2 (continued)

In vitro	Plant	Part of the plant used	Study type	Type of stones	Study design	Main results	References
	<i>Rhus chinensis Mill.</i> (Anacardiaceae)	Aqueous extract of leaves	In vitro	CaOx crystal	CaOx crystal solution	The water extract greatly ↓ the rate of nucleation and aggregation of CaOx crystallization and ↓ the crystal density in a way dependent on both time and concentration	[63]
	<i>Costus spicatus (Jacq.) Sw.</i> (Costaceae)	Ethanoic extract of leaves at different concentrations	In vitro	CaOx crystal	CaOx crystal in human urine	The extract had a concentration-dependent effect on urinary crystallization, reducing the size and percentage of monohydrated crystals	[64]
	<i>Bergenia ligulata (Wall.)</i> (Saxifragaceae)	Different solvent extracts of dried rhizome	In vitro	CaOx crystal	CaOx crystal solution	The ethanolic extract (200 µg/mL) demonstrated the maximum suppression in nucleation and aggregation experiments. This extract also enhanced cell vitality in a dosage-dependent manner, with the highest viability observed in cells treated with 200 µg/mL. Notably, there was a considerable change from thermodynamically stable COM crystals to less damaging COD crystals	[65]

↑: increase/d; ↓: decrease/d; CaOx: Calcium oxalate, COD: calcium oxalate dihydrate, COM: calcium oxalate monohydrate

Table 3 Clinical trial research on the therapeutic efficacy of Medicinal Plants on urolithiasis

Clinical trial						Main results	References
Plant	Part of the plant used	Study type	Type of stones	Study design			
<i>Alhagi maurorum</i> (Leguminosae)	Whole plant distillate	A randomized, single-blind, clinical trial	Not identified	In the study, 65 patients were randomized to the control group, while 61 were assigned to the intervention group. These individuals, all aged over 18 years, were presented with symptoms of renal colic due to ureteral stones measuring 0–10 mm. The patients were administered the distillate for four weeks	No significant difference was observed in the intervention and control groups ^[66]		
<i>Nigella sativa</i> (Ranunculaceae)	Intact seeds capsule	Randomized single-blinded clinical trial study	Not identified	A study was conducted on 80 adult patients, each presenting with kidney and ureteral calculi measuring 4–10 mm. These individuals, who exhibited no symptoms necessitating immediate intervention or significant discomfort, were recruited from a urology clinic. The patients were subsequently divided into two distinct groups. The first group was administered a daily dosage of 0.4 mg of tamsulosin. Conversely, the second group received 2g of encapsulated <i>Nigella sativa</i> seeds daily	Before treatment, no significant difference was observed between the two groups. However, post-treatment, both groups exhibited a reduction in the size and number of stones per patient. The average pain score before therapy was comparable between the two groups. Following the intervention, a considerable ↓ in pain score was noted in both groups, with a more significant reduction in the <i>Nigella sativa</i> group	In terms of treatment efficacy, the two subjects' combined whole and fractional responses approached 60%. Nevertheless, substantial differences were observed between the two groups ^[67]	

Table 3 (continued)

Clinical trial	Plant	Part of the plant used	Study type	Type of stones	Study design	Main results	References
	Ningmitai capsule composed of (<i>Herba Polygoni Capitati</i> (Polygonaceae), <i>Rhizoma Imperatae</i> (Gramineae), <i>Radix Cocculi Trilobi</i> (Menispermaceae), <i>Fructus Forsythiae suspensa</i> (Oleaceae), <i>Berberidis radix</i> (Berberidaceae), <i>Herba Agrimoniae</i> (Rosaceae) and, <i>Folium Hibisci Mutabilis</i> (Malvaceae))	Ningmitai capsule	Randomized clinical trial study	Not identified	Patients within the age range of 18–60 years were diagnosed with upper urinary tract stones, with sizes varying from 10 to 20 mm. The diagnostic methods employed were Intravenous Pyelography/Computed Tomography Urography (IVP/CTU) and non-contrast Computed Tomography (CT). One hundred twenty-three patients were randomly assigned to two groups: the Ningmitai capsule group (63 patients) and the control group (60 patients)	The cumulative expulsion rates of stones on the 3rd, 7th, 14th, and 28th days were significantly elevated in the group treated with Ningmitai capsules compared to the control group. Furthermore, the stone-free rates on the 14th and 28th days were markedly higher in the Ningmitai capsule group. The average duration to achieve a stone-free state was shorter in the Ningmitai capsule group relative to the control group. On day 14, the urine white blood cell (WBC) counts in the Ningmitai capsule group were significantly lower than in the control group	[68]

↓: decrease/d

Table 4 Therapeutic Effects of Phytochemicals on Urolithiasis

Phytochemical	Part of the plant used	Study type	Type of stones	Study design	Main results	References
Quercetin Betulin	Flowers of <i>Aerva lanata</i> (Amaranthaceae)	In vivo	CaOx crystal	EG-induced UL in rats	The administered components significantly ↑ both the weight of the rats and their urine output. Concurrently, a ↓ in the formation of calculi was observed in the renal tissue. Serum analysis revealed significant ↓ in levels of BUN, uric acid, and Cr. Histopathological evaluations indicated an ↑ in the anatomical structure of the renal tissue. Animals treated with the test drug in combination with piperine exhibited substantial ↓ in levels of Ca^{2+} , phosphate, and oxalate crystals compared to both the diseased animals and those treated with the test drug alone	[69]
Daidzin	Not identified	In vivo	CaOx crystal	EG-induced UL in rats	Daidzin therapy effectively ↓ the urine pH and protein levels, while ↑ the urine volume in the UL rats. This was accompanied by a significant ↓ in the urine crystal score. Daidzin also ↓ the levels of Ca^{2+} , oxalate, uric acid, urea, Cr, and BUN, while ↑ the levels of Mg^{2+} and phosphorus in the UL rats. The activities of ALT, AST, ALP, gamma-glutamyl transferase (GGT), and LDH in serum and renal tissue were successfully ↓ by Daidzin. Additionally, Daidzin ↓ the levels of TNF- α and adiponectin, ↑ the antioxidant levels (SOD, Glutathione Peroxidase (GPx), GSH), and ↓ LPO. Microscopic examination revealed relatively few calcified structures and no degeneration of glomeruli in the kidney	[70]
Trigonelline	Not identified	In vitro	COM crystal	COM crystal in solution, Madin-Darby canine kidney, the renal tubular epithelial cell line	The findings indicated that trigonelline significantly ↓ the number, size, and mass of COM crystals during crystallization. Trigonelline reduced crystal growth and cell adhesion in a dose-dependent manner but did not influence crystal aggregation. Trigonelline treatment led to a decrease in the amount of COM crystal receptors on the apical membranes, as confirmed by mass spectrometry protein identification. Trigonelline therapy led to decreased levels of some crystal receptors, as validated by Western blotting analysis	[71]

Table 4 (continued)

Phytochemical	Part of the plant used	Study type	Type of stones	Study design	Main results	References
Medicagenic acid	An aqueous extract of the aerial parts of <i>Herニアria hisrsuta L.</i> (Caryophyllaceae)	In vitro	COM crystal	COM crystal in solution, Madin-Darby canine kidney, the renal tubular epithelial cell line	The results from the crystallization assay revealed that there was no significant impact on either the nucleation or aggregation phase. However, the crystal-cell interaction experiment showed considerable ↓ differences in crystal binding	[72]
Methyl gallate (MG) and gallic acid (GA)	From <i>Mimosa bimucronata</i> leaves (Fabaceae)	In vitro	CaOx crystal	CaOx crystal in solution	The GA compound suppressed around 44–57% of the overall production of CaOx crystals, whereas the MG compound inhibited around 48–35%. Exposure to both GA and MG hindered COM formation. Additionally, these chemicals ↓ the absorbance in urine specimens, which is associated with a reduction in CaOx aggregation and precipitation	[73]
Pentacyclic triterpenoids (Lupeol and Ursolic acid)	Fresh leaves of <i>Alstonia scholaris</i> (Apocynaceae)	In vivo	CaOx crystal	EG + AC-induced UL in rats	The components ↓ nitrogenous wastes, such as Cr, uric acid, BUN, TNF- α , and IL-6. Additionally, they ↓ levels of crystallization promoters like Na^+ , K^+ , Cl^- , Ca^{2+} , oxalate, and phosphorus. The treatment ↑ urine volume and ↓ kidney weight. Microscopic examination of urine revealed the absence of stag horn calculi. MDA levels were ↓, while SOD, GSH, and CAT levels were ↑. Kidneys from the treatment group exhibited normal morphology with no crystal accumulation	[74]

↑: increase/d, ↓: decrease/d, AC: ammonium chloride, ALT: aminotransferase, AST: Aspartate aminotransferase, BUN: blood urea nitrogen, CaOx: Calcium oxalate, CAT: catalase, COM: calcium oxalate monohydrate, Cr: creatinine, EG: creatinine, GPx: Glutathione Peroxidase, GSH: reduced glutathione, IL-6: interleukine-6, LDH: lactate dehydrogenase, LPO: lipid peroxidation, MDA: malondialdehyde, SOD: superoxide dismutase TNF- α : Tumor necrosis factor- α , UL: urolithiasis

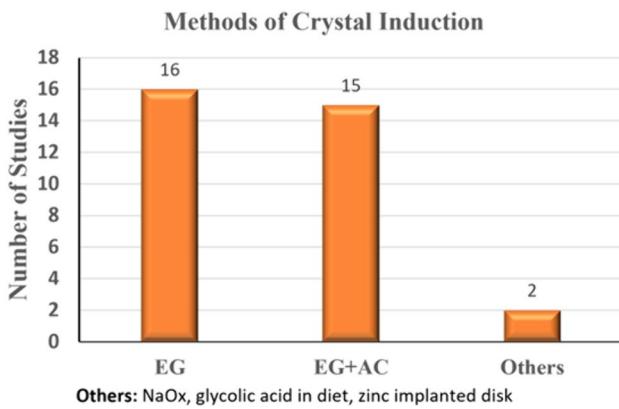


Fig. 2 Number of studies based on the methods used for crystal induction. *NaOx* sodium oxalate, *EG* ethylene glycol, *AC* ammonium chloride

stones. These studies used *in vitro* and *in vivo* methods to investigate the impact on CaOx and calcium oxalate monohydrate (COM) crystals. The compounds examined included Quercetin, Daidzin, Trigonelline, Medicagenic Acid, Methyl Gallate, Gallic Acid, and Pentacyclic Triterpenoids (Lupeol and Ursolic acid). The studies' results showed the effects of these substances on the development of stones, including alterations in urine characteristics, such as urine output volume, pH levels, protein concentrations, crystal features, such as size, number, and mass, as well as changes in levels of other biochemical markers. In rat models with CaOx nephrolithiasis circumstances were associated with modifications in components like oxalate and citrate levels together with variations in pH balance, oxidative stress markers, production of crystallization modulators and inflammatory molecules, Crystal formations in urine and deposition within the

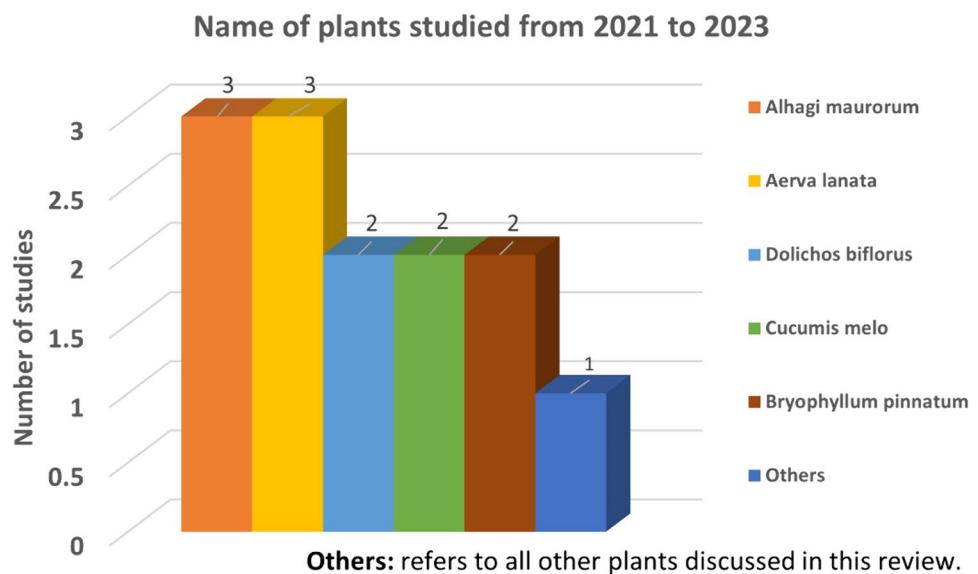
kidneys [12, 15]. Most studies have revealed enhancements in renal structure and function following the administration of these compounds. Moreover, it was frequently noted in the research that there were changes in the excretion levels of CaOx, magnesium, and phosphate.

Discussion

Nephrolithiasis, or the development of kidney stones, is a complex and multifactorial process that involves several steps and physicochemical changes in the urine environment. These changes result in the production of crystals, their growth, aggregation, and subsequent retention inside the kidneys [76, 77]. This process concerns interactions between many urinary ions and a range of crystallization modulatory macromolecules. Most idiopathic CaOx stones develop on a base of biological apatite called Randall's plaque (RP). This plaque starts in the renal papillary interstitium and travels outward to the papillary surface. When the surface epithelium breaks down, the plaque becomes exposed to the urine in the pelvic area. Furthermore, some stones form joined to tubular crystal deposits, struggling the terminal collecting ducts [78, 79].

Herbal treatments can complement lifestyle changes by targeting specific pathways involved in stone formation. For example, many phytochemicals have anti-inflammatory and antioxidant properties that can mitigate the renal damage caused by oxidative stress, which is not directly addressed by lifestyle modifications [80]. Furthermore, some herbal remedies have diuretic and antispasmodic effects, which can aid in the expulsion of stones and provide symptomatic relief [81].

Fig. 3 Number of studies discussed a certain plant name from 2021 to 2023



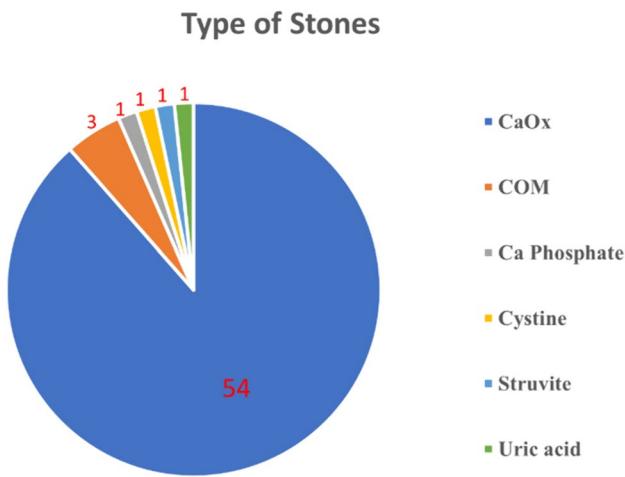


Fig. 4 Number of studies based on the types of stones studied. *CaOx* calcium oxalate, *COM* calcium monohydrate

To investigate the pathogenesis of CaOx nephrolithiasis and develop therapeutic agents, various in vitro and in vivo models have been established [82, 83]. CaOx crystal nucleation, growth, and aggregation were investigated in vitro crystallization studies with and without crystallization modulators [51]. These methods afford a preliminary evaluation of crystallization modifying activity, probable modes of action, and anti-urolithic potential. Nonetheless, the biological system and pathogenesis of urolithiasis are complicated, and these in vitro results cannot be extrapolated to therapeutic effects [84].

As a result, in vivo animal models of CaOx nephrolithiasis have been established to understand the pathophysiology better and examine the anti-urolithic activities and potential

of various medicines [29, 30]. Experimental nephrolithiasis is induced by administering hyperoxaluria-inducing agents through drinking water, diet, or injection [85, 86]. These *in vivo* models have considerably contributed to our understanding of human illnesses and remain a key tool for researchers to examine numerous physiological processes, biochemical events, and test novel pharmaco-therapeutic drugs [87].

The majority of the studies reviewed here have utilized the well-established and relatively economical rat model of nephrolithiasis by administering EG in drinking water, either alone [19, 20] or in combination with AC [15, 18]. EG, a precursor of oxalic acid, is quickly absorbed from the gastrointestinal system and converted to oxalic acid by hepatic enzymes. EG predominantly affects the kidneys, with substantial variations in sensitivity among strains, species, and sexes. In comparison to mice, rats are more sensitive, and male rats are more sensitive than female ones. While EG (0.75–1%) alone can induce CaOx deposition, its effects are variable [88]. In order to decrease the amount of time needed and attain a consistently high rate of renal crystal deposition, hypercalciuric, nephrotoxic, or pH-reducing procedures, such as AC [89], gentamicin [90], or a diet lacking in magnesium, has been combined with EG.

When rats are given EG at a concentration of 0.75% or more in drinking water, they develop hyperoxaluria, which leads to crystalluria and CaOx crystal deposition in the renal tubules [28]. The incidence of crystal deposition in the kidney varies from 80 to 100%, depending on the co-administered medicine, and nephrolithiasis develops in around 1–3 weeks [91]. Oxidative stress in the kidneys, increased water intake and polyuria, lower urinary pH, decreased urinary Ca²⁺, Mg²⁺, and citrate contents,

Fig. 5 Number of studies based on parts of the plants used

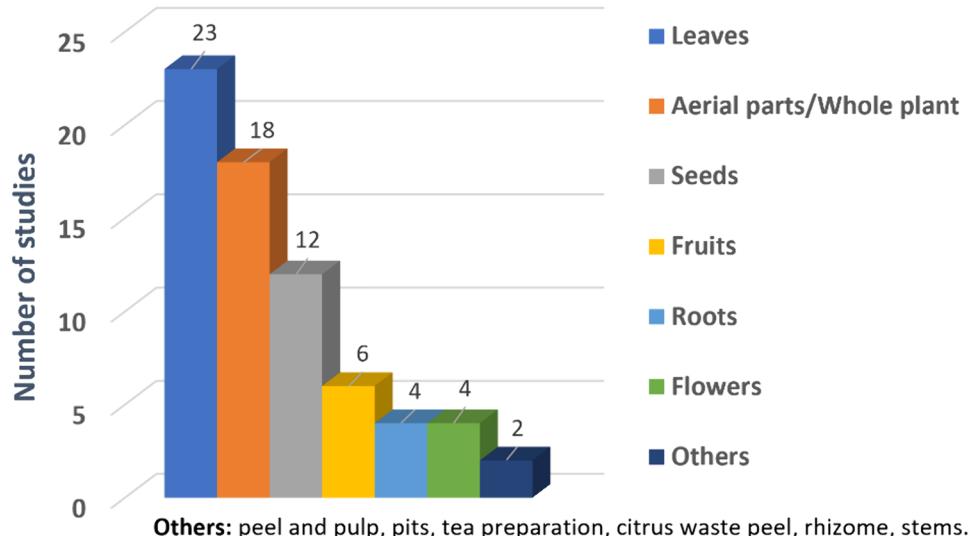
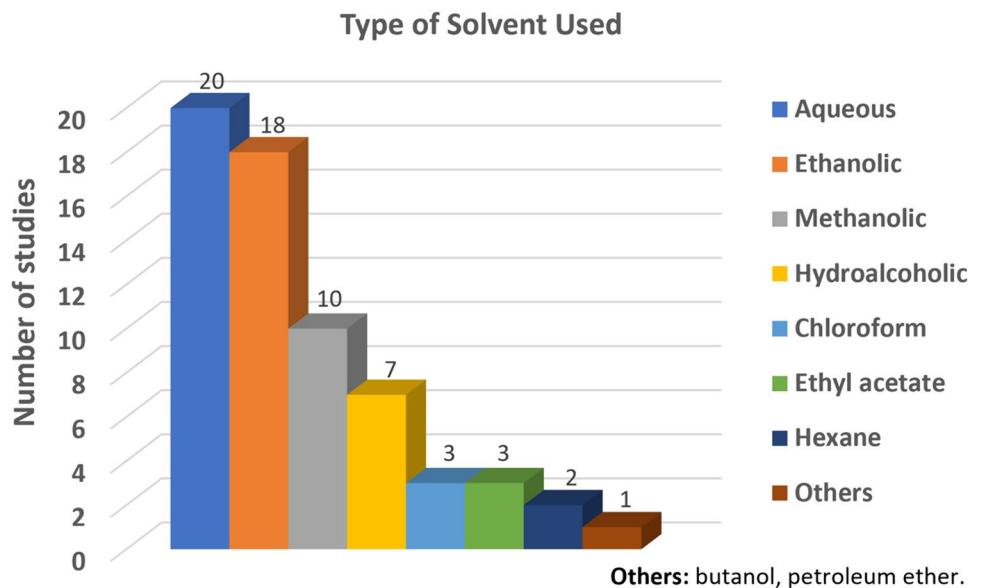


Fig. 6 Number of studies based on the types of solvents used in the extraction



increased CaOx crystalluria, phosphate excretion, renal hypertrophy, and weight loss are the main characteristics of hyperoxaluria-induced nephrolithiasis [35, 39]. Increased loss of urine protein, decreased clearance of creatinine, and raised levels of blood urea nitrogen (BUN) and creatinine in the serum are further indicators of renal impairment [30, 34].

Herbal therapies were prepared for assessment using a variety of plant parts in the examined research, including flowers, seeds, fruits, leaves, stems, roots, and rhizomes. The most popular component was leaves, which were extracted using alcoholic, hydroalcoholic, and aqueous solvents. The effectiveness of herbal administration was evaluated in relation to common biomarkers of nephrolithiasis, including renal CaOx crystal deposits, citrate, pH, oxalate, oxidative stress markers, urine calcium, phosphate, and improved renal structure and function. While some studies did not investigate every facet of nephrolithiasis, every therapy decreased the amount of CaOx crystals that were deposited in the kidneys. Most studies indicated that using herbal remedies improved the kidneys' structure and function. Furthermore, research on oxidative stress showed that herbal remedies have antioxidant qualities.

The clinical trials in this review suggest potential benefits of herbal treatments like *Alhagi maurorum*, *Nigella sativa*, and the Ningmitai capsule (a Chinese herbal formulation) in facilitating stone passage, alleviating pain, promoting stone expulsion, and increasing stone-free rates in patients with urolithiasis. While these findings corroborate the traditional use of these herbs and provide preliminary evidence for their anti-urolithic properties, the number of trials is limited. Larger, well-designed clinical studies are warranted to further evaluate the efficacy and safety of these

herbal treatments, including their potential interactions with conventional therapies, before recommending their use in clinical practice.

The studies investigating various phytochemicals highlight their potential as therapeutic agents for urolithiasis. Quercetin, Daidzin, trigonelline, medicagenic acid, methyl gallate, gallic acid, lupeol, and ursolic acid exhibited anti-urolithic effects by reducing crystal formation, adhesion, aggregation, and oxidative stress while improving renal function in experimental models. These findings suggest phytochemicals may target multiple pathways involved in stone formation and associated renal dysfunction. However, further research is warranted to elucidate their mechanisms of action, optimize dosing and formulations, evaluate safety profiles, and translate these findings into clinical applications to prevent and manage urolithiasis.

Differentiating between stone types is crucial for effective management. Herbal treatments can be tailored to target the specific pathophysiological mechanisms of different stones. For calcium oxalate stones, phytochemicals, such as quercetin and gallic acid, can inhibit crystal formation and aggregation. For infectious stones, the antimicrobial properties of herbs like *Mentha piperita* can help prevent stone formation by controlling urinary infections. This targeted approach allows for a more personalized treatment plan, potentially improving outcomes for patients with different types of kidney stones.

In conclusion, this systematic review critically evaluates the use of various phytochemical and natural herbal treatments in experimental models of nephrolithiasis, highlighting their potential as therapeutic agents for managing this complex condition. The findings underscore

the need for further research to elucidate the underlying mechanisms and translate these findings into clinical practice.

Conclusion

This systematic review has provided a comprehensive overview of the current state of research on the use of plants in treating and preventing urolithiasis. The findings suggest that various plants and their components have significant potential in managing this condition. They reduce the size and number of stones and alter the levels of urinary oxalate, calcium, phosphate, and citrate, which are critical factors in stone formation. However, further well-designed clinical trials are needed to validate these findings and establish these plants' optimal use in clinical practice. This research opens new avenues for developing safe and effective phytotherapeutic strategies for urolithiasis, and ongoing research is essential to translate these findings into clinical applications.

Author contributions E.A.H.A was responsible for the conceptualization of the study, developing the methodology, and overseeing the data curation process. Essmat conducted the formal analysis and investigation, prepared the original draft of the manuscript, and contributed significantly to the review and editing process. Additionally, Essmat handled the visualization of data, supervised the project, and managed the overall project administration. M.S. contributed to the data curation and formal analysis, participated in the investigation, and assisted in the review and editing of the manuscript. Mahmoud also contributed to the visualization of the data.

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Data availability The data supporting the findings of this study are available from the corresponding author upon reasonable request.

Declarations

Conflict of interest The authors declare that they have no conflict of interest.

Ethical approval Not applicable.

Consent for publication Not applicable.

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References

- Zhu C, Wang D-Q, Zi H, Huang Q, Gu J-M, Li L-Y et al (2021) Epidemiological trends of urinary tract infections, urolithiasis and benign prostatic hyperplasia in 203 countries and territories from 1990 to 2019. *Mil Med Res* 8:1–12
- Sigurjonsdottir VK, Runolfsdottir HL, Indridason OS, Palsson R, Edvardsson VO (2015) Impact of nephrolithiasis on kidney function. *BMC Nephrol* 16:149. <https://doi.org/10.1186/s12882-015-0126-1>
- Zeng G, Zhu W, Robertson WG, Penniston KL, Smith D, Pozdzik A et al (2022) International Alliance of Urolithiasis (IAU) guidelines on the metabolic evaluation and medical management of urolithiasis. *Urolithiasis* 51:4
- Zumstein V, Betschart P, Abt D, Schmid H-P, Panje CM, Putora PM (2018) Surgical management of urolithiasis—a systematic analysis of available guidelines. *BMC Urol* 18:25. <https://doi.org/10.1186/s12894-018-0332-9>
- Abu Zarin M, Tan JS, Murugan P, Ahmad R (2020) Investigation of potential anti-urolithiatic activity from different types of *Musa* pseudo-stem extracts in inhibition of calcium oxalate crystallization. *BMC Complement Med Therap* 20:1–12
- Sansores-España D, Pech-Aguilar AG, Cua-Pech KG, Medina-Vera I, Guevara-Cruz M, Gutiérrez-Solis AL et al (2022) Plants used in Mexican traditional medicine for the management of urolithiasis: a review of preclinical evidence, bioactive compounds, and molecular mechanisms. *Molecules* 27:2008
- Ahmed S, Hasan MM, Alam Mahmood Z (2016) Antiurolithiatic plants: formulations used in different countries and cultures. *Pak J Pharm Sci* 29:2129–2139
- Ahmed S, Hasan MM, Khan H, Mahmood ZA, Patel S (2018) The mechanistic insight of polyphenols in calcium oxalate urolithiasis mitigation. *Biomed Pharmacother* 106:1292–1299
- Kiremit MC, Boyuk A, Petkova K (2023) Fluid intake recommendations in urolithiasis and general advice to patients without metabolic risk factors. *World J Urol* 41:1251–1259. <https://doi.org/10.1007/s00345-023-04285-3>
- Taylor EN, Curhan GC (2006) Diet and fluid prescription in stone disease. *Kidney Int* 70:835–839. <https://doi.org/10.1038/sj.ki.5001656>
- Ahmad W, Khan MA, Ashraf K, Ahmad A, Daud Ali M, Ansari MN et al (2021) Pharmacological evaluation of Safoof-e-Pathar Phori- A polyherbal unani formulation for urolithiasis. *Front Pharmacol* 12:597990. <https://doi.org/10.3389/fphar.2021.597990>
- Ayaz Adakul B, ŞEn A, ŞEner TE, ErdoĞAn Ö, ÇEvİK Ö, Eker P, et al (2022) *Platanus orientalis* (plane tree) extract protects against hyperoxaluria induced kidney damage. *Jrp.* 26(2):311–324. <https://doi.org/10.29228/jrp.129>
- Bawari S, Sah AN, Tewari D (2022) Discovering the antiurolithiatic potential of wild Himalayan cherry through in vitro and preclinical investigations. *S Afr J Bot* 145:218–227
- Bervinova AV, Palikov VA, Mikhailov ES, Palikova YA, Borozdina NA, Kazakov VA et al (2022) Efficacy of *Ficus tikoua* Bur. extract in ethylene glycol-induced urolithiasis model in SD rats. *Front Pharmacol* 13:974947. <https://doi.org/10.3389/fphar.2022.974947>
- Choudhary SS, Panigrahi PN, Dhara SK, Sahoo M, Dan A, Thakur N et al (2023) *Cucumis callosus* (Rottl.) Cogn. fruit extract ameliorates calcium oxalate urolithiasis in ethylene glycol induced hyperoxaluric Rat model. *Heliyon*. 9:e14043. <https://doi.org/10.1016/j.heliyon.2023.e14043>
- Eidi M, Ashjazadeh L (2023) Anti-urolithiatic effect of *Cucumis melo* L. var inodorous in male rats with kidney stones. *Urolithiasis*. 51:45. <https://doi.org/10.1007/s00240-023-01418-6>
- Khan A, Gilani AH (2023) An insight investigation to the anti-urolithic activity of *Trachyspermum ammi* using the in vitro and in vivo experiments. *Urolithiasis* 51:43. <https://doi.org/10.1007/s00240-023-01415-9>

18. Nafiu MO, Ogunsola IJ (2023) Anti-nephrolithiatic evaluation of partitioned ethanol extract of *Calotropis procera* Leaf in wistar rats. *Jordan J Biol Sci* 16:105–115
19. Rashid S, Sameti M, Alqarni MH, Abdel Bar FM (2023) In vivo investigation of the inhibitory effect of *Peganum harmala* L. and its major alkaloids on ethylene glycol-induced urolithiasis in rats. *J Ethnopharmacol* 300:115752. <https://doi.org/10.1016/j.jep.2022.115752>
20. Yilmaz H, Ekinci N, Ömerli A, Nisari M, Yay AH, Ülger H et al (2023) The protective effect of *Myrtus communis* L. against experimental kidney stone in rats. *Adv Trad Med* 23:241–249. <https://doi.org/10.1007/s13596-021-00620-4>
21. Zhang S, Zhu J, Ju Y, Lv M, Yang R, Li Y et al (2023) Drosophila model and network pharmacology to explore novel targets and novel active components of Chinese traditional medications for treating kidney stones. *Pharmacol Res Modern Chin Med* 6:100220. <https://doi.org/10.1016/j.prmcm.2023.100220>
22. Chakit M, Boussekkour R, Hessni AE, Bahbiti Y, Nakache R, Mustaphi HE et al (2022) Antiurolithiatic activity of aqueous extract of *Ziziphus lotus* on ethylene glycol-induced lithiasis in rats. *Pharmacogn J* 14:596–602. <https://doi.org/10.5530/pj.2022.14.141>
23. Chen S-J, Dalanbaatar S, Chen H-Y, Wang S-J, Lin W-Y, Liu P-L et al (2022) *Astragalus membranaceus* extract prevents calcium oxalate crystallization and extends lifespan in a drosophila urolithiasis model. *Life* 12(8):1250
24. Golla S, Pasala PK, Sura S, Nainita K, Katabathina D (2022) Anti urolithiatic activity of *Cyperus rotundus* tubers: in silico in vitro and in vivo approaches. *Braz J Pharm Sci*. <https://doi.org/10.1590/s2175-97902022e181009>
25. Guillén-Meléndez GA, Soto-Domínguez A, Loera-Arias MdJ, Castillo-Velázquez U, Villa-Cedillo SA, Piña-Mendoza EI et al (2022) Effect of methanolic extract of *Mimosa malacophylla* A Gray in vero and HEK-293 cell lines, and in the morphology of kidney and bladder of rats with induced urolithiasis. *J Ethnopharmacol* 297:115552. <https://doi.org/10.1016/j.jep.2022.115552>
26. Jamshed A, Jabeen Q (2022) Pharmacological evaluation of *Mentha piperita* against urolithiasis: an in vitro and in vivo study. *Dose Response* 20:1–15. <https://doi.org/10.1177/15593258211073087>
27. Ajay Kumar MKN, Kumar B, Kumar A, Kumar R, Kailashya V, Singh AK (2022) Toxicity (acute and subacute) assessment and in-vivo antiurolithiatic activity of ethanolic extract of *Caesalpinia bonducuella* seed in albino Wistar rat. *J Appl Pharm Sci* 12:187–197. <https://doi.org/10.7324/JAPS.2022.120220>
28. Ly HT, Le TKO, Nguyen MK, Le VM (2022) Diuretic efficacy and prophylactic effects of hydroethanolic extract from *Musa balbisiana* fruits against urolithiasis. *Adv Trad Med* 22:823–836. <https://doi.org/10.1007/s13596-022-00629-3>
29. Sahu MK, Singh G (2022) Structural identification through GC mass spectrophotometer and determine anti lithiotic activity of hibiscus rosa sinensis by using ethylene glycol induced method. *JMPAS* 11:4244–4249. <https://doi.org/10.55522/jmpas.v11i1.1575>
30. Yogesh Kumar S, Umesh Kumar G (2022) Effect of citrus limon (L.), citrus aurantium and citrus medica on ethylene glycol induced urolithiasis in rats. *J Pharm Neg Results* 13:601–608. <https://doi.org/10.47750/pnr.2022.13.s06.086>
31. Singh SA, Vellapandian C, Krishna G (2022) Preventive and therapeutic effects of *Aerva lanata* (L.) extract on ethylene glycol-induced nephrolithiasis in male Wistar albino rats. *Dig Chin Med* 5:199–209. <https://doi.org/10.1016/j.dcm.2022.06.009>
32. Smitha Grace SR, Manasa BY, Jyoti Bala C (2022) Evaluation Of antiurolithiatic activity of methanolic seed extracts of *Persea americana* against calcium oxalate induced urolithiasis in rats. *J Pharm Neg Results*. <https://doi.org/10.47750/pnr.2022.13.s05.101>
33. Xu X, Chen J, Lv H, Xi Y, Ying A, Hu X (2022) Molecular mechanism of *Pyrrosia lingua* in the treatment of nephrolithiasis: network pharmacology analysis and in vivo experimental verification. *Phytomedicine* 98:153929. <https://doi.org/10.1016/j.phymed.2022.153929>
34. Zhang J, Hou A, Dong J, Zheng S, Yu H, Wang X et al (2022) Screening out key compounds of *Glechoma Herba* for anti-urolithic activity and quality control based on spectrum-effect relationships coupled with UPLC-QDA. *Biomed Pharmacother* 149:112829. <https://doi.org/10.1016/j.bioph.2022.112829>
35. Zhou F, Wang X (2022) Pyrrhoa petiolosa extract ameliorates ethylene glycol-induced *Urolith iasis* in rats by inhibiting oxidative stress and inflammatory response. *Dis Markers* 2022:1913067. <https://doi.org/10.1155/2022/1913067>
36. Alelign T, Tessema TS, Debella A, Petros B (2021) Evaluations of the curative efficacy of *G fruticosus* solvent extracts in experimentally induced nephrolithiatic Wistar male rats. *BMC Complement Med Ther* 21:145. <https://doi.org/10.1186/s12906-021-03320-3>
37. ElSawy NA, Mosa OF (2021) The antiurolithic activity of *Organum vulgare* on rats treated with ethylene glycol and ammonium chloride: possible pharmaco-biochemical and ultrastructure effects. *Curr Urol* 15:119–125. <https://doi.org/10.1097/cu9.00000000000000017>
38. Kaushik S, Choudhary M, Rajpal S (2021) Antiurolithiatic efficacy of combination preparations of *Dolichos biflorus* and *Crataeva nurvala*: folk medicines used in Indian traditional medicine. *Future J Pharm Sci* 7:21. <https://doi.org/10.1186/s43094-020-00170-7>
39. Ravi Kiran C, Rajkuberan C, Sangilimuthu AY, Hakkim FL, Bakshi H, Manoharan SP (2021) Intrinsic evaluation of antiurolithiatic capacity of *Argemone mexicana* L. in wistar albino rats. *J Herbs Spices Med Plants* 27:289–304. <https://doi.org/10.1080/1049475.2021.1891181>
40. Saleem A, Islam M, Saeed H, Iqtedar M (2021) In-vivo evaluation of anti-urolithiatic activity of different extracts of peel and pulp of *Cucumis melo* L in mice model of kidney stone formation. *PJZ*. <https://doi.org/10.17582/journal.pjz/20190418170457>
41. Sumanjali C, Shashidhar M, Sravani M, Babu KR, Tejeswarudu B, Kalyani CD (2021) Anti-urolithiatic activity of the ethanolic extract of *Cassia auriculata* against ethylene glycol induced urolithiasis in experimental rats. *RJPT*. <https://doi.org/10.52711/0974-360x.2021.00906>
42. Tabas PM, Aramjoo H, Yousefinia A, Zardast M, Abedini MR, Malekanbeh M (2021) Therapeutic and preventive effects of aqueous extract of date palm (*Phoenix dactylifera* L.) pits on ethylene glycol-induced kidney calculi in rats. *Urol J* 18:612–617
43. Al-Mamoori F, Aburjai T, Al-Tawalbe DM (2022) In-vitro anti-nephrolithiatic activity of selected medicinal plants. *Trop J Nat Prod Res*. 6:1426–1429
44. Ammar RB, Khalifa A, Alamer SA, Hussain SG, Hafez AM, Rajendran P (2022) Investigation of the potential anti-urolithiatic activity of *Alhagi maurorum* (Boiss) grown wild in Al-Ahsa (Eastern Province) Saudi Arabia. *Braz J Biol* 84:e259100
45. Babu M, Uma KH, Joseph S, Sree A, Scariya S, Shubina KA (2021) In-vitro evaluation of anti-urolithiatic and larvicidal activity of *alternanthera sessilis*. *Biomed Pharmacol J* 14:671–680
46. Bashan I, Bozlu M (2020) The possible litholytic effect of *Ononis spinosa* L. on various human kidney stones—an in vitro experimental evaluation. *J Herb Med*. 22:100345
47. Chattaraj B, Nandi A, Das A, Baidya A, Mahata S, Chowdhury A et al (2023) *Enhydra fluctuans* Lour aqueous extract inhibited the growth of calcium phosphate crystals: an in vitro study. *Food Chem Adv*. 2:100287. <https://doi.org/10.1016/j.focha.2023.100287>
48. Latif A, Azhar F, Rafay MZ, Iqbal A, Anwar I (2023) Phytochemical screening and in vitro anti-urolithiatic activity of fruit-seed

- extracts of *Melia azedarach*. Jordan J Pharm Sci 16:137–147. <https://doi.org/10.35516/jjps.v16i1.1074>
49. Pushparani VP, Baskar G (2023) Synthesize and characterization of CaOx crystals against various citrus waste peel extracts: an in vitro study. Prep Biochem Biotechnol 53:353–365. <https://doi.org/10.1080/10826068.2022.2090003>
 50. Chetna F, Priyadarshini P (2022) Comparative study of hydroalcoholic extracts of *Bryophyllum pinnatum* and *Macrostyloma uniflorum* for their antioxidant, antiurolithiatic, and wound healing potential. J Appl Biol Biotech. <https://doi.org/10.7324/jabb.2021.100124>
 51. Hewagama SP, Hewawasam RP (2022) Antiurolithiatic potential of three sri lankan medicinal plants by the inhibition of nucleation, growth, and aggregation of calcium oxalate crystals in vitro. ScientificWorldJournal 2022:8657249. <https://doi.org/10.1155/2022/8657249>
 52. Huy RNA, Govindan R, Sivaramakumar N, Raman R, Jayaraman S, Basavan D et al (2022) Inhibition of calculi forming oxalate by dietary *Basella rubra* organs: litholytic activity. Braz J Pharm Sci. <https://doi.org/10.1590/s2175-97902022e20582>
 53. Kant R, Singh TG, (2021). Effect of *Dolichos biflorus* seeds based functional beverage on in vitro calcium oxalate crystallization in human urine. 12:5836–44. <https://doi.org/10.33263/BRIAC125.58365844>
 54. Kaviraj M, Andrew Pradeep M, Satheesh D (2022) In-vitro investigation on antiurolithiatic activity and phytochemical examination of *Aerva lanata* and *Bryophyllum pinnatum*: a comparative study. J Indian Chem Soc 99:100487. <https://doi.org/10.1016/j.jics.2022.100487>
 55. Mammate N, El Oumari FE, Imtara H, Belchkar S, Lahrichi A, Alqahtani AS et al (2022) Antioxidant and anti-urolithiatic activity of aqueous and ethanolic extracts from *Saussurea costus* (Falc) lispitch using scanning electron microscopy. Life (Basel) 12:1026. <https://doi.org/10.3390/life12071026>
 56. Mariano LNB, Pontioli DA, da Silva AA, Niero R, Cechinel-Filho V, de Souza P (2022) Diuretic and antiurolithic effect of *Garcinia humilis* (Vahl) CD Adams leaves, a medicinal plant native to South American countries. Chem Biodivers 19:e202200022. <https://doi.org/10.1002/cbdv.202200022>
 57. Mohan PK, Krishna TPA, Thirumurugan A, Kumar TS, Kumari BDR (2022) Chemical profiling and in vitro antiurolithiatic activity of *Pleurolobus gangeticus* (L.) J. St.- Hil. ex H. Ohashi & K. Ohashi along with its antioxidant and antibacterial properties. Appl Biochem Biotechnol 194:5037–5059. <https://doi.org/10.1007/s12010-022-04017-0>
 58. Smanthong N, Tavichakorntrakool R, Tippayawat P, Lulitanond A, Pinlaor P, Daduang J et al (2022) Anti-proteus activity, anti-struvite crystal, and phytochemical analysis of *Sida acuta* Burm F ethanolic leaf extract. Molecules 27:1092. <https://doi.org/10.3390/molecules27031092>
 59. Ambursa MB, Rahman MNG, Sulaiman SA, Zakaria AD, Mohamed Daud MA, Zakaria Z et al (2021) An in vitro study of *Orthosiphon stamineus* (Misai Kucing) standardized water extract as a chemolytic agent in urolithiasis. J Pharm Bioallied Sci 13:373–379. https://doi.org/10.4103/jpbs.jpbs_526_21
 60. El Habbani R, Lahrichi A, Sqalli Houssaini T, Kachkoul R, Mohim M, Chouhani BA et al (2021) In vitro mass reduction of calcium oxalate urinary calculi by some medicinal plants. Afr J Urol 27:28. <https://doi.org/10.1186/s12301-021-00132-2>
 61. Faujdar C (2021) Investigating the effect of hydroalcoholic extract of *Ocimum sanctum* on in-vitro calcium oxalate crystallization. Curr Trends Biotechnol Pharm 15:47–52
 62. Gul MT, Muhammad N, Pauzi AN, Bakar MFA, Talip BA, Abdullah N et al (2021) Evaluation of *Phyllanthus niruri* L. from Malaysia for in-vitro anti-urolithiatic properties by different solvent extraction. Pak J Sci Ind Res 64:81–86. <https://doi.org/10.52763/pjisr.biol.sci.64.1.2021.81.86>
 63. Heirangkhongjam MD, Ngaseppam IS (2021) *Rhus chinensis* Mill: a medicinal plant with promising inhibition of calcium oxalate crystallization, an in-vitro study. J Herb Med. 29:100489. <https://doi.org/10.1016/j.hermed.2021.100489>
 64. Moreno KGT, Gasparotto Junior A, Dos Santos AC, Palozi RAC, Guarner LP, Marques AAM et al (2021) Nephroprotective and antilithiatic activities of *Costus spicatus* (Jacq) Sw: ethnopharmacological investigation of a species from the Dourados region, Mato Grosso do Sul State Brazil. J Ethnopharmacol 266:113409. <https://doi.org/10.1016/j.jep.2020.113409>
 65. Singh A, Tandon S, Nandi SP, Kaur T, Tandon C (2021) Downregulation of inflammatory mediators by ethanolic extract of *Bergenia ligulata* (Wall) in oxalate injured renal epithelial cells. J Ethnopharmacol 275:114104. <https://doi.org/10.1016/j.jep.2021.114104>
 66. Aryaeefar MR, Khakbaz A, Akbari S, Movahedi A, Gazerani A, Bidkhorri M et al (2022) Effect of *Alhagi maurorum* distillate on ureteral stone expulsion: a single-blind randomized trial. J Herb Med 34:100567. <https://doi.org/10.1016/j.hermed.2022.100567>
 67. Shakeri N, Mehrabi S, Paymand A (2022) Comparison efficacy of oral *Nigella sativa* seeds and tamsulosin on pain relief and passage of 4–10 mm stones of kidney and ureter; a randomized clinical trial. J Nephropharmacol 11:e08. <https://doi.org/10.34172/npj.2022.08>
 68. Wang R, Qiao Q, Yang D, Zhang J, Zhu C, Sun J et al (2022) Ningmitai capsule promotes calculi expulsion after RIRS for 10–20-mm upper urinary stones: a multicenter, prospective, randomized controlled trial. Urolithiasis 50:205–214. <https://doi.org/10.1007/s00240-021-01296-w>
 69. Nagula S, Subhashini NJP, Bhikshapathi DVRN, Mamatha P, Rao PS (2023) Anti-urolithiatic and nephroprotective activity of quercetin and betulin in conjunction with a bio enhancer—an in vivo study. Biomed Pharmacol J 16:847–862. <https://doi.org/10.13005/bpj/2667>
 70. Yuan S, Ibrahim IAA, Ren R (2023) Anti-urolithiatic activity of daidzin in ethylene glycol-induced urolithiasis in rats. Appl Biochem Biotechnol 195:905–918
 71. Peerapen P, Boonmark W, Thongboonkerd V (2022) Trigonelline prevents kidney stone formation processes by inhibiting calcium oxalate crystallization, growth and crystal-cell adhesion, and downregulating crystal receptors. Biomed Pharmacother 149:112876. <https://doi.org/10.1016/j.biopha.2022.112876>
 72. Peeters L, Fouquet K, Breynaert A, Schreurs G, Verhulst A, Pieters L et al (2022) Effects of medicagenic acid metabolites, originating from biotransformation of an *Herniaria hirsuta* extract, on calcium oxalate crystallization in vitro. J Ethnopharmacol 285:114860. <https://doi.org/10.1016/j.jep.2021.114860>
 73. Cechinel-Zanchett CC, Bolda Mariano LN, Schlickmann F, Cechinel-Filho V, de Souza P (2021) In vitro effects of two bioactive compounds, gallic acid and methyl gallate, on urolithiasis. Actas Urol Esp (Engl Ed) 45:604–608. <https://doi.org/10.1016/j.acuroe.2020.09.010>
 74. Zehra S, Sanaye MM (2021) Evaluation of anti-urolithiatic potential of leaves of *Alstonia scholaris* and its isolated pentacyclic triterpenoids in ethylene glycol-induced renal calculi rat model. Indian J Pharm Educ 55:232–239
 75. Hussaini IM, Ahmed HS, Ahmad Hu, Mamunu, Sulaiman AM, Usman A, editors (2023) Preliminary screening for antibacterial activity of endophytic fungi isolated from *Azadirachta indica* and *Mentha piperita* Phyllosphere against *Staphylococcus aureus*, *Escherichia coli* and *Pseudomonas aeruginosa*
 76. Shastri S, Patel J, Sambandam KK, Lederer ED (2023) Kidney stone pathophysiology, evaluation and management: core

- curriculum 2023. Am J Kidney Dis 82:617–634. <https://doi.org/10.1053/j.ajkd.2023.03.017>
77. Wang Z, Zhang Y, Zhang J, Deng Q, Liang H (2021) Recent advances on the mechanisms of kidney stone formation (Review). Int J Mol Med 48:149. <https://doi.org/10.3892/ijmm.2021.4982>
 78. Khan SR, Canales BK, Dominguez-Gutierrez PR (2021) Randall's plaque and calcium oxalate stone formation: role for immunity and inflammation. Nat Rev Nephrol 17:417–433. <https://doi.org/10.1038/s41581-020-00392-1>
 79. Daudon M, Bazin D, Letavernier E (2015) Randall's plaque as the origin of calcium oxalate kidney stones. Urolithiasis 43(Suppl 1):5–11. <https://doi.org/10.1007/s00240-014-0703-y>
 80. Niu Y, Na L, Feng R, Gong L, Zhao Y, Li Q et al (2013) The phytochemical EGCG, extends lifespan by reducing liver and kidney function damage and improving age-associated inflammation and oxidative stress in healthy rats. Aging Cell 12:1041–1049
 81. Yadav RD, Jain S, Alok S, Mahor A, Bharti JP, Jaiswal M (2011) Herbal plants used in the treatment of urolithiasis: a review. Int J Pharm Sci Res 2:1412
 82. Khan A, Bashir S, Khan SR (2021) Antiurolithic effects of medicinal plants: results of in vivo studies in rat models of calcium oxalate nephrolithiasis-a systematic review. Urolithiasis 49:95–122. <https://doi.org/10.1007/s00240-020-01236-0>
 83. Khan SR (1997) Animal models of kidney stone formation: an analysis. World J Urol 15:236–243. <https://doi.org/10.1007/BF01367661>
 84. Kavanagh JP (2006) In vitro calcium oxalate crystallisation methods. Urol Res 34:139–145. <https://doi.org/10.1007/s00240-005-0027-z>
 85. Varalakshmi P, Shamila Y, Latha E (1990) Effect of *Crataeva nurvala* in experimental urolithiasis. J Ethnopharmacol 28:313–321. [https://doi.org/10.1016/0378-8741\(90\)90082-5](https://doi.org/10.1016/0378-8741(90)90082-5)
 86. Atmani F, Slimani Y, Mimouni M, Hacht B (2003) Prophylaxis of calcium oxalate stones by *Herniaria hirsuta* on experimentally induced nephrolithiasis in rats. BJU Int 92:137–140. <https://doi.org/10.1046/j.1464-410x.2003.04289.x>
 87. Davis CM (2013) Animal models of drug abuse: place and taste conditioning. Curr Trends Biotechnol Pharm, pp 681–707
 88. Fan J, Glass MA, Chandhoke PS (1999) Impact of ammonium chloride administration on a rat ethylene glycol urolithiasis model. Scan Microsc 13:299–306
 89. Khan SR, Glenton PA (1995) Investigative urology: deposition of calcium phosphate and calcium oxalate crystals in the kidneys. J Urol 153:811–817
 90. Hackett RL, Shevock PN, Khan SR (1990) Cell injury associated calcium oxalate crystalluria. J Urol 144:1535–1538. [https://doi.org/10.1016/s0022-5347\(17\)39793-8](https://doi.org/10.1016/s0022-5347(17)39793-8)
 91. Okada Y, Kawamura J, Nonomura M, Kuo YJ, Yoshida O (1985) Experimental and clinical studies on calcium urolithiasis: (I) animal model for calcium oxalate urolithiasis using ethylene glycol and 1-alph a (OH) D3 Hinyokika kiyo. Acta Urol Japonica 31:565–577

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