ORIGINAL ARTICLE



Early detection of graft incompatibility in sweet cherry by internode association and callus fusion techniques

Arezoo Jalali¹ · Ebrahim Ganji Moghaddam² · Ali Marjani¹

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Abstract

The scion-rootstock incompatibility is one of the most common problems in fruit trees. The study evaluated predicted compatibility or incompatibility by internode association and callus fusion techniques. Two factorial experiments were laid out with an incompletely randomized design with three replications. Treatment includes the first factor, four levels of sweet cherry cultivars ('Bing', 'Takdaneh', 'Siyah Mashhad' and 'Adli'), and the second factor, four levels of rootstocks ('Gisela-5', 'Gisela-6', 'Mahaleb' (M-168) and 'GF-305'). Results showed significant differences in total phenolic content, peroxidase and starch content in internode association and callus fusion experiments. 'GF-305' rootstock (incompatible) had the lowest success rate of grafts on all cultivars, while 'Gisela-6' rootstock showed the highest success rate of grafts on 'Siyah Mashhad' and 'Bing' cultivars. Both experiments showed that in incompatible grafts, an increase in total phenol content and peroxidase activity and a decrease in grafts' success rate were observed. The degree of compatibility was significantly and positively correlated with the success rate of grafts while significantly and negatively correlated with total phenol content and peroxidase activity. In both experiments, no clear trend was observed regarding starch content and the success rate of graft's success rate and the degree of compatibility. Our findings lead to the conclusion that phenolic compounds (mostly) and peroxidase activity can be used to pre-screen for incompatible grafts. Moreover, the callus fusion technique can be a quick way to predict (in)compatibility of graft in sweet cherry.

Key Message

Results of this article indicated that phenolic and peroxidase activity can be used to pre-screen for incompatible grafts. The callus fusion technique can be a quick way to predict (in) compatibility of graft in sweet cherry.

Keywords Cherry · Callus · Degree compatibility · Phenol content · Peroxidase activity · Success rate of graft

Abbreviations

- MS Murashige and skoog medium
- 2,4-D 2,4-Dichlorophenoxyacetic acid
- BAP 6-Benzylaminopurine
- NAA 1-Naphthaleneacetic acid
- DC Degree of compatibility

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Ebrahim Ganji Moghaddam eganji31568@gmail.com

² Department of Crop and Horticulture Science Research, Khorasan Razavi, Agricultural and Natural Resources Research and Education Center, AREEO, Mashhad, Iran

Introduction

Graft incompatibility is one of the obstacles and problems in using the rootstock of fruit trees. The genetic difference between rootstock and scion is one of the important factors of graft incompatibility (Gainza et al. 2015). Although a series of biological studies have been performed on woody and herbaceous plants, this area has limited information. Moreover, the molecular and biochemical mechanisms of graft incompatibility are not well understood. The accurate and rapid prediction of graft incompatibility is essential (Hudina et al. 2014).

Global sweet cherry production has slightly increased recently (Lanauskas et al. 2023). Different studies with *Prunus* sp. have shown that rootstocks significantly influence the grafted cultivar. This influence is expressed as differences in

¹ Department of Agriculture, Faculty Agriculture, Bojnourd Branch, Islamic Azad University, Bojnourd, Iran

vigor, yield and fruit characteristics such as firmness, fruit size, sugar and organic acid content (Lopez-Ortega et al. 2016). Sweet cherry rootstocks show different suitability for sweet cherry cultivars. Such a different pattern is highlighted by the nutrient uptake efficiency, leaf area development, chlorophyll content and subsequent photosynthetic activity (Asanica et al. 2015).

The use of new methods, such as tissue culture for the production of plants has received much attention. Tissue culture guarantees production at a high speed and with this method, it is possible to produce uniform plants similar to the mother base and free from disease and contamination (Sarropoulou et al. 2017). Jonard et al. (1990) reported the possibility of predicting graft incompatibility of apricot and lemon trees using in vitro techniques (e.g., internode association and callus fusion). Callus formation at the graft union is the first positive reaction in the graft, while failure to form callus tissue leads to graft failure (Porika et al. 2016).

Recently, different methods, such as anatomical studies (Balbi et al. 2019) and the determination of starch accumulation (Zhou et al. 2020), have been used to predict rootstock and scion incompatibility quickly. These techniques facilitate the early detection of graft incompatibility and provide a model for a more accurate analysis of the grafting phenomenon (Suryadi et al. 2020).

Some compounds are essential in the early stages of growth for the compatibility of the rootstock and scion. Many studies show the role of phenols in lignification. Xylem cell wall cells have a dynamic structure, including polysaccharides, phenolic compounds (such as lignin), minerals and proteins (Tucker et al. 2018; Mnich et al. 2020).

Porika et al.(2016) showed different phenols in the branches of apple, pear, plum, cherry, apricot and peach trees, and the amount of the metabolites depends on the branches' growth degree. Prabpree et al.(2018) have reported phenolic compounds as markers for evaluating graft compatibility between rootstock and scion. Cheng et al. (2017) said the number of phenolic compounds significantly differed at the top and bottom of the graft union. Due to the significant relationship between arbutin accumulation at the top and bottom of the graft union can be used to perform pre-screening graft incompatibility.

Many reports have shown peroxidase's role in inhibiting growth (Guclu and Koyuncu 2012). Researchers reported that peroxidase levels in incompatible grafts were higher than in compatible grafts (Zarrouk et al. 2010). Peroxidase profiles in grafted pears were not similar compatible and incompatible grafts, which caused abnormal lignification at the graft union and incompatibility.

Different opinions exist about starch accumulation at the compatible and incompatible grafts. Studies have shown that starch accumulation above the graft union and its absence or lack at the bottom of the graft union destroys the phloem. However, it should be noted that in some rootstock and scion interactions, signs of graft incompatibility may be observed. Still, there is no accumulation of starch at the top of the graft union, or the opposite may occur (Melnyk et al. 2018).

This study aimed to predict the (in)-compatibility by internode association and callus fusion techniques and compare the amount of total phenol, peroxidase and starch accumulation in compatible and incompatible grafts of sweet cherry cultivars.

Materials and methods

This experiment was carried out on four sweet cherry cultivars ('Bing', 'Takdaneh', 'Siyah Mashhad', and 'Adli'), one of Iran's important sweet cherry cultivars. Four rootstocks ('Gisela-5', 'Gisela-6', 'Mahaleb'(M-168), and 'GF-305') in the tissue culture laboratory of Khorasan Razavi Agricultural and Natural Resources Research and Education Center, Iran.

'Gisela' rootstock from the interspecific *P. cerasus* \times *P. avium* and 'Gisela 6' rootstock interspecific hybridization *P. cerasus* \times *P. Canescens* is compatible with a wide range of cherry cultivars. In this experiment, 'Gisela-6' rootstock was considered a compatible control and 'GF-305' an incompatible control. This study was performed in two independent experiments, internode association and callus fusion, under in vitro conditions.

Internode association experiment

The internode (1.0 cm) was sterilized with 70% ethanol for 1 min, washed three times with distilled water 5% NaClO for 10 min and cleaned more than three times with sterile water. With respected polarity rootstock and scion, sideveneer (spliced-side) grafts joined each other. A sloping cut is made at the rootstock. The scion is cut to be paired with the rootstock, and the graft union site is covered with sterile glue until fusion.

The explants were cultured in Murashige and Skoog (MS) sterilized-medium modified supplemented with 0.2 mg L⁻¹ 6-Benzylaminopurine (BAP) and 1 mg L⁻¹ naphthaleneacetic acids (NAA) (Murashige and Skoog 1962). After culturing in the test glass tube, including 10 ml medium, the samples were transferred to the growth chamber (16/8 photoperiod and temperature 25 ± 1 °C) (Fig. 1).

Approximately four weeks after grafting, grafting's success rate and total phenol, peroxidase, and starch content at the graft union were measured.



Fig. 1 Respected polarity rootstock and scion (A). Covered with sterile glue until fusion (B). The explants were cultured in MS medium (C)



Fig. 2 Internode explant for production callus (A). Production callus (B). Calluses were paired together in MS medium (C)

Callus fusion experiment

The internode explant (1.0 cm) was sterilized like in the previous experiment and cultured in MS medium supplemented with BAP $(0,1, 2, 3 \text{ mg } \text{L}^{-1})$ and 2,4-D $(0, 1, 2, 3 \text{ mg } \text{L}^{-1})$ concentrations. Twenty explants were in each treatment (10 glass Petri dishes included two explants), and ten treatments and 200 explants were used in the callus formation stage. After culturing in a petri dish, the explants were transferred to a growth chamber (16/8 photoperiod and temperature 25 ± 1 °C). After about a week, callus formation started in the sample of different treatments (Fig. 2).

Three weeks later, determined the best treatment, the rootstock produced calluses and the scions were placed next to each other, with side-veneer (spliced-side) grafts joined to each other and in MS medium supplemented with 2 mg L^{-1} BAP and 3 mg L^{-1} 2,4-D to determine the rootstock's compatibility and incompatibility with the scion. After ten days, the compatible calli were grown and merged, and the incompatible calli remained apart (Fig. 3), and the degree of compatibility of the rootstock and scion (DC) was determined in Eq. (1):

$$DC = (GI_{RS}/GI_{Rc}) + GI_{SR}/GI_{SC})/2$$
(1)

 GI_{RS} : the percentage increase in the rootstock's mass from rootstock and sweet cherry scion cultivar from the rootstock's co-culture. GI_{RC} (the rootstock control): the percentage increase in the group of the rootstock. GI_{SR} : the percentage increase in the mass of the scion from the co-culture of sweet cherry cultivar and rootstock and GIsc (the scion control): the percentage increase in the scion's mass cultivar (Pedersen 2006).

Approximately 4 weeks after grafting, grafting's success rate and the amount of total phenol, peroxidase and starch content at the graft union were measured.

Measurement of total phenol

The measurement of total phenolic compounds was determined according to the Folin-Ciocalteu method (Singleton et al. 1999). Finally, the absorption of samples was calculated using the standard curve for gallic acid (Guimaraes et al. 2020).



Fig. 3 Callus fusion experiment, Compatible (A-E) and incompatible (F-J) grafting in different sweet cherry cultivars and rootstocks callus

Extraction of enzymatic antioxidant activity

Antioxidant enzyme extraction was defined following Sunohara and Matsumoto's (2004) method. The filtrate was collected to determine the peroxidase activity (EC 1.11.1). Peroxidase activity was assayed using a method proposed by Polle et al. (1997).

Measurement of starch content

To extract the starch, the method of Whistler (1964) was used with some modification; the absorbance of all samples was measured at 630 nm (max-plu 5384 UV–VIS). Antron reagent was used as a control.

Statistical analysis

After checking the data distribution normality (Kolmogorov–Smirnov and Shapiro–Wilk test) assumption, the statistical analyses to determine the individual effect were conducted using Statistical Analysis System software (SAS Institute, Cary, NC, USA, *ver.* 9.2). The least significant difference (LSD) test was applied to compare the means (at the 0.05 probability level). Simple correlations (Pearson) and regression analysis between traits were performed using SAS and Minitab (v. 19) software.

Results

First experiment: internode association

As shown in Table 1, rootstocks, cultivars and interaction effects of rootstocks and cultivars were significant on the rate of success grafting, phenol content, peroxidase activity and starch content. The highest success rate of grafts was related to the 'Bing' cultivars on the 'Gisela-6' (compatible control) rootstock. The lowest success rate of grafts was related to the 'GF-305' (incompatible control) on all cultivars (Fig. 4).

The highest amount of total phenol was achieved in union's grafting 'Takdaneh' and 'Siyah Mashhad' cultivars on 'GF-305' rootstock with an average of 6.81 and 6.67 mg g⁻¹ FW. There was no significant difference in the amount of total phenol in the grafting 'Bing' cultivar on the 'GF-305' rootstock (6.42 mg g⁻¹ FW). The lowest total phenol was found in the grafting 'Bing' cultivar on the

Table 1Variance analysis of theeffect of cultivar and rootstockon the success rate of grafting,phenol content, peroxidaseactivity and starch accumulationin the internode associationexperiment

S.O.V	df	Mean square (MS)						
		Success rate of grafts	Phenol	Peroxidase	Starch			
Rootstock (R)	3	15,178.3**	36.61**	0.312**	0.0026**			
Cultivar (C)	4	299.6**	5.40**	0.047**	0.0005**			
R×C	12	3169.2**	1.98**	0.018**	0.0004**			
Error	40	36.20	0.11	0.002	0.00005			
CV (%)	-	9.7	8.03	13.87	9.97			

**Significant at 1% probably level



Fig. 4 Interaction effects of sweet cherry cultivars and rootstocks on the success rate of grafts in the internode association experiment 'Gisela-6' rootstock with an average of 2.34 mg g⁻¹ FW. The results showed that grafting 'Mahaleb' and 'GF-305' root-stocks on all cultivars increased the phenol content (Fig. 5).

The lowest peroxidase activity was observed in the 'Bing' cultivar on 'Gisela-5'. Moreover, the highest peroxidase activity was related to the grafting 'Siyah Mashhad' cultivar on 'GF-305' rootstock (Fig. 6). In general, the highest level of peroxidase was obtained in grafting sweet cherry cultivars on 'GF-305' incompatible control.

The highest starch accumulation was observed in the 'Takdaneh' cultivar union's grafting on the 'GF-305' (0.12%). The lowest accumulation of starch was observed in the 'Adli' cultivar grafting union on the 'Gisela-6' (0.06%) (Fig. 7). Despite the significant difference in starch accumulation at the unions grafts between cultivars on rootstocks,

there was no significant relationship between compatible and incompatible graft in starch accumulation. In other words, starch accumulation in all 'GF-305' grafts (incompatible control) was not higher than in other grafts.

Table 2 shows significant negative and positive correlations among grafts' success rate, phenol content, peroxidase activity and starch content. For example, the success rate of grafts was significantly and negatively correlated with phenol content, peroxidase activity, and starch content. Phenol content was highly and significantly correlated.

Regression analysis between graft success rate and other traits was performed separately to understand the relationships between characteristics better. The results showed that phenol content, peroxidase activity and starch content harmed grafts' success rate and the highest R^2 (0.34) was



Fig. 6 Interaction effects of sweet cherry cultivars and rootstocks on peroxidase activity in the internode association experiment

Fig. 5 Interaction effects of

sweet cherry cultivars and rootstocks on phenol content

in the internode association

experiment



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Fig. 7 Interaction effects of sweet cherry cultivars and rootstocks on starch content in the internode association experiment



 Table 2
 Simple correlation coefficients (Pearson) between the success rate of grafts with phenol content, peroxidase activity and starch content in the internode association experiment

	Success rate of grafts	Phenol	Peroxidase	Starch
Success rate of grafts	1			
Phenol content	- 0.58**	1		
Peroxidase activity	- 0.31*	0.74**	1	
Starch content	- 0.45*	0.43*	0.20 ^{ns}	1

ns non-significant

*,**Significant at the 5 and 1% probability level, respectively

related to phenol content (Fig. 8). These results were in line with the correlation analysis findings between traits because the highest negative correlation coefficient was observed between phenol content and grafts' success rate.

Second experiment: callus fusion

The mean comparison of callus formation in sweet cherry explant in MS medium under the effect of 2,4-D and BAP in different concentrations showed a statistically significant difference. The highest percentage of callus formation was treating 3 mg L⁻¹ 2,4-D and 2 mg L⁻¹ BAP in all cultivars and rootstocks (Table 3). Callus production time was 6 to 8 days after planting, depending on the type of hormone. 'Gisela-6' rootstock, 'Siyah Mashhad' and 'Takdaneh'



Fig. 8 Regression changes in phenol content, peroxidase activity, and starch content with changes in the success rate of grafts in the internode association experiment

Treatment	BAP (mgl ⁻¹)	2,4-D (mgl ⁻¹)	Rootstocks			Cultivars				
			'Gisela-6'	'Gisela-5'	'GF-305'	'Mahaleb'	'Bing'	'Adli'	Siyahmashhad	'Takdaneh'
1	0	0	_	_	_	_	_	_	_	_
2	1	1	33.3 ^{de}	25.0 ^{cde}	25.0 ^{cd}	25.0 ^{cd}	16.6 ^e	33.3d ^e	8.33 ^{cd}	25.0 ^c
3	2	1	25.0 ^e	8.33 ^e	25.0 ^{cd}	25.0 ^{cd}	25.0 ^d	25.0 ^e	16.6 ^{cd}	8.33 ^d
4	3	1	16.67 ^f	16.67 ^{de}	8.33 ^d	16.6 ^d	16.6 ^e	25.0 ^e	16.6 ^{cd}	8.33 ^d
5	1	2	75.0 ^{ab}	58.3 ^b	50.0 ^{bc}	58.3 ^b	66.6 ^b	75.0 ^b	50.0 ^{ab}	50.0 ^b
6	2	2	50.0 ^{cd}	41.6 ^{bcd}	50.0 ^{bc}	33.3 ^c	58.3 ^{bc}	58.3°	33.3 ^{bcd}	25.0 ^c
7	3	2	58.3 ^{bc}	41.6 ^{bcd}	58.33 ^{ab}	41.6 ^{bc}	41.6 ^{cd}	50.0 ^{cd}	33.3 ^{bcd}	25.0 ^c
8	1	3	66.6 ^{bc}	25.0 ^{cde}	50.0 ^{bc}	50.0 ^b	58.3 ^{bc}	75.0 ^b	41.67 ^{bc}	41.67 ^{bc}
9	2	3	91.67 ^a	91.6 ^a	83.3 ^a	75.0 ^a	91.6 ^a	91.67 ^a	75.0 ^a	83.3 ^a
10	3	3	25.0 ^e	50.0 ^{bc}	16.67 ^d	41.6 ^{bc}	50.0 ^{bcd}	41.6 ^d	25.0 ^{cd}	25.0 ^c

Table 3 The effects of different concentrations of 2,4-D and BAP in MS medium on callus formation in the internode explants, It showed that treatment 9 is the best treatment and is bold

The means of each cultivar and rootstock followed by the superscript same letter in each column are not significantly different according to LSD test

cultivars started callus formation after six days, and 'GF-305' started callus formation after eight days.

As shown in Table 4, rootstocks, cultivars, and interaction effects of rootstocks and cultivars were significant on the degree of compatibility, grafts' success rate, phenol content, peroxidase activity and starch content ($p \le 0.01$).

In the interaction effects of cultivars and rootstocks, the highest degree of compatibility (1.74) was related to the grafting of 'Takdaneh' on 'Gisela-5'. Moreover, 'Bing' on 'GF-305' grafting showed the lowest degree of compatibility (0.50). The results showed that 'Gisela-5' and -6 rootstocks increased the degree of compatibility in each cultivar, while 'GF-305' rootstocks decreased the degree of compatibility (Fig. 9).

Results indicated that the highest success rate was the grafting of 'Adli', 'Bing', 'Siyah Mashhad', and 'Takdaneh' cultivars on 'Gisela-6' rootstocks. The calluse fusion of the 'Bing' cultivar on 'Gisela-6' had the highest success rate (83.33%). On the other hand, 'GF-305' rootstocks' application reduced the success rate of calluse fusion, which was very noticeable in applying 'GF-305' rootstocks on cultivars (Fig. 10). Predicting the possibility of incompatibility

between rootstock and scion in fruit trees in orchards may take a long time. Still, by using the tissue culture technique, graft incompatibility can be predicted in a shorter period. The callus fusion technique is a quick way to predict graft compatibility or incompatibility.

As shown in Fig. 11, the interaction of cultivars and rootstocks showed that the phenol content in the 'Takdaneh' cultivar was much higher than in other cultivars. The highest phenol content was related to the 'Takdaneh' cultivar's grafting on the 'GF-305' rootstock by 28.76 mg g⁻¹ FW. In all cultivars, the application of 'Gisela-6' compatible rootstock decreased phenol content. The grafting of 'Adli' on 'Gisela-6' and 'Mahaleb' (M-168) rootstocks had the lowest phenol content (2.05 and 2.5 mg g⁻¹ FW, respectively).

Peroxidase activity in 'GF-305' incompatible rootstocks showed a significant increase compared to other rootstocks. The highest peroxidase activity was related to 'Takdaneh' cultivar grafting on 'GF-305' rootstocks with an average of 0.754 U mg protein⁻¹ min. On the other hand, application of 'Gisela-6' (compatible rootstock) resulted in a significant reduction in the activity of this enzyme, so that the lowest peroxidase activity was achieved in the

Table 4Variance analysisof the effects of cultivarsand rootstocks in the graftbinding area on the degreeof compatibility, success rateof grafts, phenol content,peroxidase activity, and starchcontent in the callus fusionexperiment

S.O.V	df	Mean square (MS)						
		Degree of compatibility	Success rate of grafts	Phenol	Peroxidase	Starch		
Rootstock (R)	3	0.930**	8096.52**	233.5**	0.068**	0.00066**		
Cultivar (C)	4	0.120**	329.86*	493.8**	0.372**	0.00071**		
R×C	12	0.119**	1728.75**	31.19**	0.014**	0.00048**		
Error	40	0.0046	108.3	1.65	0.0002	0.000008		
CV (%)	-	5.97	18.8	14.96	6.30	10.90		

**Significant at 1% probably level

Fig. 9 Interaction effects of sweet cherry cultivars and rootstocks on the degree of compatibility in the callus fusion experiment



Fig. 10 Interaction effects of sweet cherry cultivars and rootstocks on the success rate of grafts in the callus fusion experiment





Cultivars





'Adli' cultivar on 'Gisela-6' rootstock with an average of $0.105 \text{ U mg protein}^{-1} \min (\text{Fig. 12})$.

The highest starch content was observed in the grafting of 'Siyah Mashhad' on Gisela- 5 (0.1%) and 'Takdaneh' on Gisela- 6 (0.9%), respectively. The lowest starch accumulation was in 'Adli' on 'GF-305' (0.06%) (Fig. 13). Despite the significant difference in starch accumulation at the unions grafts between cultivars on rootstocks. Still, there was no significant relationship between compatible and incompatible grafts in starch accumulation. In other words, starch accumulation in all 'GF-305' grafts (incompatible control) was not higher than other grafts.

As shown in Table 5, the degree of compatibility was significantly and positively correlated with the success rate of grafts (0.64^{**}) while significantly and negatively

correlated with phenol content (-0.34^*) and peroxidase activity (-0.33^*) . The degree of compatibility showed a non-significant correlation with starch content.

As shown in Fig. 14, the regression changes in the success rate of grafts, phenol content, peroxidase activity and starch content with differences in the degree of compatibility under the callus fusion experiment showed that the characteristics such as phenol content and peroxidase activity were negatively related with the degree of compatibility. In other words, by increasing the range of these compounds, the degree of graft compatibility decreased. However, the R^2 obtained from this analysis had a very low coefficient compared to the first experiment. On the other side, the relationship between the degree of compatibility and the success rate of the positive relation ($R^2 = 0.45$).

Fig. 13 Interaction effects of sweet cherry cultivars and root-stocks on starch content in the callus fusion experiment



Table 5Simple correlationcoefficients (Pearson) betweenthe degree of compatibilitywith the success rate of grafts,phenol content, peroxidaseactivity and starch content in thecallus fusion experiment

	Degree of compatibility	Success rate of grafts	Phenol	Peroxidase	Starch
Degree of compatibility	1				
Success rate of grafts	0.64**	1			
Phenol content	- 0.34*	- 0.51**	1		
Peroxidase activity	- 0.33*	- 0.47*	0.93**	1	
Starch content	0.14 ns	0.05 ns	0.18 ns	0.34*	1

*,**Significant at 5 and 1% probably level, respectively



Fig. 14 Regression changes in the success rate of grafts, phenol content, peroxidase activity, and starch content with differences in the degree of compatibility in the callus fusion experiment

Discussion

Not many studies were performed on the graft incompatibility mechanism and its prediction methods in sweet cherry trees. Sweet cherries are mainly grown by amateur growers and are rarely planted in commercial orchards (Lanauskas et al. 2023). This study aimed to investigate the changes in some metabolites and techniques needed to understand this mechanism better. Callus tissue is formed at the junction of the rootstock and scion to the graft 4 to 5 days after grafting, while the lack of callus tissue formation at the intersection leads to graft failure (Porika et al. 2016). Using tissue culture tests helps us detect transplant compatibility in a shorter period and prevent damage to gardeners.

The grafts' degree of compatibility showed that the grafts are incompatible when the degree of compatibility is less than 1. If the degree of compatibility is more than 1 (Fig. 9), the grafts are compatible. The highest degree of compatibility was recorded in 'Takdaneh' and 'Siyah Mashhad' cultivars on the 'Gisela-6' rootstock. The degree of compatibility in 'Gisela-5' rootstocks was less than 'Gisela-6', which indicates that the 'Takdaneh' and 'Siyah Mashhad' cultivars are more compatible with 'Gisela-6'(Fig. 9).

Plants rapidly increase their antioxidant capacity when stress occurs to increase resistance and tolerance to the conditions created. It can be said that the plant's ability to disperse excess energy and neutralize free radicals is impaired, thus increasing the antioxidant capacity to increase stress resistance, so incompatible grafts increase phenolic compounds and peroxidase to minimize oxidative damage compared to compatible grafts (Zarrouk et al. 2010; Baron et al. 2019). Since the 'GF-305' rootstock was an incompatible control in this experiment, higher phenol content was observed in grafting sweet cherry cultivars on the 'GF-305' rootstock. Moreover, the total phenol content in sweet cherry cultivars on the 'Gisela-6' rootstock, which was a compatible control, was lower than that of the incompatible control. The amount of total phenol in the transplants performed on the 'Mahaleb' (M-168) rootstock was higher than the compatible control and less than the incompatible control. Generally, the total phenol in the grafts on 'Gisela-6' (compatible control) and 'Gisela-5' rootstocks was less than 'GF-305' (incompatible control).

The lowest phenol in the first experiment in 'Bing/Gisela 6' (Fig. 5) and the second experiment was 'Adli/Gisela 6' (Fig. 11). 'GF-305' is incompatible with sweet cherry cultivars and it seems that phenolic compounds can be used to pre-screen incompatible grafts. According to high storage theories, phenolic compounds such as catechins above the graft union can be used as a biochemical marker in diagnosing graft incompatibility (Canas et al. 2015; Baron et al. 2019). Moreover, Hudina et al. (2014) have reported phenolic compounds as markers to evaluate the grafts' compatibility between the rootstocks and scion.

Peroxidase levels in incompatible grafts were higher than in compatible grafts. The peroxidase enzyme can be used to predict incompatible grafts quickly. Peroxidases are enzymes that play essential biochemical and physiological roles in plants, including plant growth, differentiation and development, auxin catabolism, ethylene biosynthesis, plasma membrane regeneration, cell wall development, ligninification and response to pathogens (Pandey et al. 2017). Preliminary analyses of peroxidase indicate that this enzyme is essential in forming cell wall constituents and ligninification. By creating cross-linking between cell wall phenolic polymers, the next step reduces cell wall flexibility. This process causes irreversible hardening of the cell wall (Suchy et al. 2010).

The result of present study showed that the peroxidase enzyme activity in grafting cultivars on 'GF-305' rootstocks that were incompatible was higher than grafting cultivars on 'Gisela-6' (compatible control), 'Gisela-5' and Mahaleb. The lowest peroxidase in the first experiment was in 'Bing/ Gisela 5' (Fig. 6), and in the second experiment was 'Adli/ Gisela 6' (Fig. 12).

Many studies have shown the role of peroxidase in inhibiting growth and reported that peroxidase levels in incompatible grafts are higher than incompatible grafts (Zarrouk et al. 2010; Baron et al. 2019). The researchers noted that the lack of similarity in the isoperoxidase composition between the rootstock and scion could lead to abnormal ligninification and the absence of vascular attachment at the graft union, leading to incompatible grafts (Zarrouk et al. 2010).

Some research findings on the relationships between peroxidase isoenzyme patterns and graft incompatibility in different plants suggest that the matching of isoperoxidase grafts between the rootstock and the scion can be used as an indicator of graft proliferation (Dogra et al. 2018). In the present study, starch accumulation at the union's grafting did not perform well in predicting the graft's incompatibility. Different results have been reported for starch accumulation at the union's grafting (Deng et al. 2019).

In some of them, starch accumulation occurred in incompatible grafting. Due to genetic differences and cultivar type, starch accumulation was not a good sign of graft incompatibility as in the present study. Starch accumulation above the graft union and its absence or lack at the bottom of the graft union cause damage to the phloem vessel, resulting from the starch collection in the phloem vessel (Deng et al. 2019). Researchers have shown that the grafts union's total starch content changes dramatically (Hudina et al. 2014). The relative starch content of rootstock and scion tissues is affected by the type of graft composition. In incompatible compounds, the relative starch content of scion wood is higher than rootstock wood. Accumulation of starch and soluble sugars in scion has also been observed in transmitted incompatibility. Low accumulation of starch at the top and bottom of the graft causes better boiling of the graft and adequate transfer of nutrients to the roots and vice versa (Karimi and Hassanpour 2017). However, in the grafting of cherries on the 'Mahaleb' (M-168) rootstock, although signs of incompatibility were seen, no difference was found between the starch at the graft union's top and bottom.

The lowest starch in the first experiment was in 'Siyah Mashhad/6' (Fig. 7). The second experiment was the lowest Starch, 'Adli/ Mahaleb' (Fig. 13).

The callus fusion experiment showed that hormones (2,4-D and BAP) are necessary for callus production, and callus was not produced in the culture medium without hormones.

The callus fusion technique is a quick way to predict graft compatibility or incompatibility. The rootstock and scion callus are fully integrated with the compatible grafts after about ten days, but the calluses do not merge if the graft is incompatible. The present results showed that the 'Gisela-6' rootstock grows more than other rootstocks with scion in the culture medium. It seems that in compatible sweet cherry grafts, a growth-promoting substance is secreted to which the compatible rootstock reacts. Determination of graft incompatibility using callus culture in some plants was reported by Gainza et al. (2015). Callus tissue formation at the graft union is the first positive reaction in the graft (Porika et al. 2016). Irisarri et al. (2015) used callus fusion to predict early graft compatibility in apricot and lemon trees. Early and accurate prediction of transplant incompatibility is essential because of the need to avoid incompatible compounds and select compatible ones (Zarrouk et al. 2010). However, the metabolic and physiological mechanisms involved in maladaptive responses are not well understood and require further research.

The highest total phenol and peroxidase levels at the graft union were higher on the 'GF-305' incompatible rootstock than on the 'Gisela-6' compatible rootstock. Therefore, our finding leads to the conclusion that phenolic compounds and peroxidase activity can be used to pre-screen for incompatible grafts.

Conclusion

The finding of this study leads to the following conclusions: The rootstock and scion degree of compatibility increased with graft success rate and decreased with phenol content and peroxidase activity. Increases in phenol content and peroxidase activity were observed in incompatible grafts.

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Data availability All data generated or analyzed during this study are included in this published article.

Declarations

Conflict of interest The authors declare that there are no conflicts of interest related to this article.

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