



Cancer progression and tumor hypercoagulability: a platelet perspective

Yifan Zhang^{1,2} · Jingtong Zeng^{1,2} · Shihao Bao^{1,2} · Bo Zhang^{1,2} · Xianjie Li^{1,2} · Hanqing Wang^{1,2} · Yuan Cheng^{1,2} · Hao Zhang^{1,2} · Lingling Zu^{1,2} · Xiaohong Xu³ · Song Xu^{1,2} · Zuoqing Song^{1,2}

Accepted: 26 April 2024 / Published online: 17 May 2024

© The Author(s), under exclusive licence to Springer Science+Business Media, LLC, part of Springer Nature 2024

Abstract

Venous thromboembolism, which is common in cancer patients and accompanies or even precedes malignant tumors, is known as cancer-related thrombosis and is an important cause of cancer-associated death. At present, the exact etiology of the elevated incidence of venous thrombosis in cancer patients remains elusive. Platelets play a crucial role in blood coagulation, which is intimately linked to the development of arterial thrombosis. Additionally, platelets contribute to tumor progression and facilitate immune evasion by tumors. Tumor cells can interact with the coagulation system through various mechanisms, such as producing hemostatic proteins, activating platelets, and directly adhering to normal cells. The relationship between platelets and malignant tumors is also significant. In this review article, we will explore these connections.

Keywords Cancer · Platelets · Coagulation · VTE · PD-L1

Introduction

Cancer and cardiovascular disease are diseases with intricate pathophysiology and the most common causes of death; prevalence is steadily increasing worldwide [1, 2]. Although venous thromboembolism (VTE), including deep venous thrombosis (DVT) and pulmonary embolism (PE), is common in the general population, it is especially common in cancer patients, and is accompanied by or can occur prior to malignant tumors [3]. Research indicates that the annual incidence rate of venous thromboembolism (VTE)

in cancer patients is 0.5%, compared to 0.1% in the general population [4]. Active cancer accounts for 20% of the total VTE incidence [5, 6]. Furthermore, cancer-associated thrombosis (CAT) is a significant contributor to mortality in cancer patients [7, 8]. Compared with non-malignant tumor patients, cancer patients have a nine-fold increased risk of developing VTE [9]. The high incidence of VTE in cancer patients arises from a complex interplay between acquired and genetic factors that perturb the delicate balance of hemostasis, ultimately culminating in thrombosis [10]. In the general population, numerous genetic factors associated with VTE have been identified, including mutations in genes related to anticoagulation (SERPINC1, PROC, and PROS1), as well as various genetic polymorphisms [11]. Common factors contributing to the high incidence of VTE in cancer patients include advanced age, reduced mobility, and various cancer treatments such as surgery, radiotherapy, and chemotherapy. During the development of cancer, tumor cells release inflammatory cytokines and pro-angiogenic/procoagulant factors that stimulate stromal cells to express prothrombotic components, thereby promoting blood clotting [12, 13]. Table 1 provides a comprehensive overview of the factors that predisposed patients to thrombosis. Furthermore, cancer patients undergoing immune checkpoint therapy are highly susceptible to developing CAT and exhibit elevated mortality rates [14–16]. The inherent heterogeneity

Authors Yifan Zhang, Jingtong Zeng, and Shihao Bao denotes as the first authors or first co-authors.

✉ Song Xu
xusong28@jd.com

✉ Zuoqing Song
thoracic_expert@aliyun.com

¹ Department of Lung Cancer Surgery, Tianjin Medical University General Hospital, Tianjin, China

² Tianjin Key Laboratory of Lung Cancer Metastasis and Tumor Microenvironment, Lung Cancer Institute, Tianjin Medical University General Hospital, Tianjin, China

³ Colleges of Nursing, Tianjin Medical University, Tianjin, China

Table 1 Cancer and thrombosis risk factor overview [22–32]

Tumor related factors	
Cancer Type and Location	Certain cancers like pancreatic, stomach, and brain are more thrombogenic
Histological stage	Advanced stages often carry a higher risk
Stage of cancer	Patients of distant metastases usually carries a higher risk
Tumor Secretions	Some tumors release substances that increase coagulability
Patient-related factors	
Comorbidity	Conditions like obesity, cardiovascular disease, or previous thrombosis
Genetic Predispositions	Certain genetic mutations increase thrombosis risk
Age	Older patients generally have a higher risk
Varicose Veins	Can predispose to superficial venous thrombosis which may lead to deep vein thrombosis
Treatment related factors	
Surgical treatment	Surgical treatments increase the risk of thrombosis
Central Venous Catheters	Used for treatment can increase risk
Adjunct Therapies	Erythropoietin and Transfusions
Immobility	Due to hospitalization or illness can lead to stasis and clot formation
Pharmacological Interventions	Chemotherapy (e.g., thalidomide, platinum-based drugs)
Hormone Therapy	Altering blood clotting and hormone levels(e.g., tamoxifen)
Lifestyle and External Factors	
Smoking	Increases the risk of thrombosis
Nutrition	Poor diet may contribute to higher risk
Physical Activity	Sedentary lifestyle can increase risk

and biological characteristics of tumors may also contribute to an increased susceptibility to VTE [17], such as glioblastoma multiforme (GBM), pancreatic cancer, gastric cancer, and lung cancer carry a significantly higher risk of VTE due to the unique tumor mechanism that induces a hypercoagulable state in the body [18]. VTE is an independent predictor of reduced survival in cancer patients, meaning that the presence of VTE may indicate tumor progression, tumor treatment failure, and some underlying cancers [14, 19–21]. Gene mutations in tumor cells, particularly those affecting TP53, KRAS, EGFR, PTEN and IDH1 genes, can have an impact on hemostasis by inducing the expression of tumor tissue factor (TF) and vascular endothelial growth factor (VEGF), as well as promoting thrombosis through the release of pro-inflammatory cytokines and extracellular vesicles [13].

Platelets are enucleated cells released from the membrane processes of mature megakaryocytes (pro-platelets), which contribute to the formation of clots and, if maladjusted, may lead to thrombosis [33]. With the development of platelet biology, we have a new understanding of platelet formation, function, and signaling (Fig. 1). Despite numerous hypotheses, the etiology underlying the high prevalence of venous thrombosis in cancer patients remains uncertain. Tumor cells can interact with the hemostatic system in a myriad of ways, including the production of hemostatic proteins such as TF and thrombin, activation of platelets, and direct adhesion to normal cells like endothelial cells, monocytes, and platelets

[34, 35]. In this review, our objective is to provide a concise overview of the pivotal role that platelets play in hemostasis and thrombosis, as well as to examine the intricate interplay between tumors and platelets, the current state of research, and the possibility of future treatment of tumors.

Tumor progression with platelet aggregation and activation

While platelets can have a positive impact on the behavior of cancer cells, the physiology and phenotype of platelets are also subject to influence by tumor cells [36]. In fact, several studies have revealed that tumor cells can regulate the RNA profile, number, and functionality of platelets. Cancer patients often exhibit activated clotting pathways that lead to a four-fold increased risk of thrombosis [37]. Professors Levin and Conley found that at least 40% of their hospitalized cancer patients had thrombocytosis [38]. Since then, a growing number of studies have reported a significant link between thrombocytosis and solid tumors, with a prevalence of 4–55% at the time of initial diagnosis [39–42]. As described in a review study of 3,654 patients with stage I-III breast cancer, 6.5% of these patients were diagnosed with inflammatory breast cancer (IBC), this study concluded that thrombocytosis, which is more prevalent in IBC patients, was associated with lower overall survival in these subjects, but not in non-IBC patients

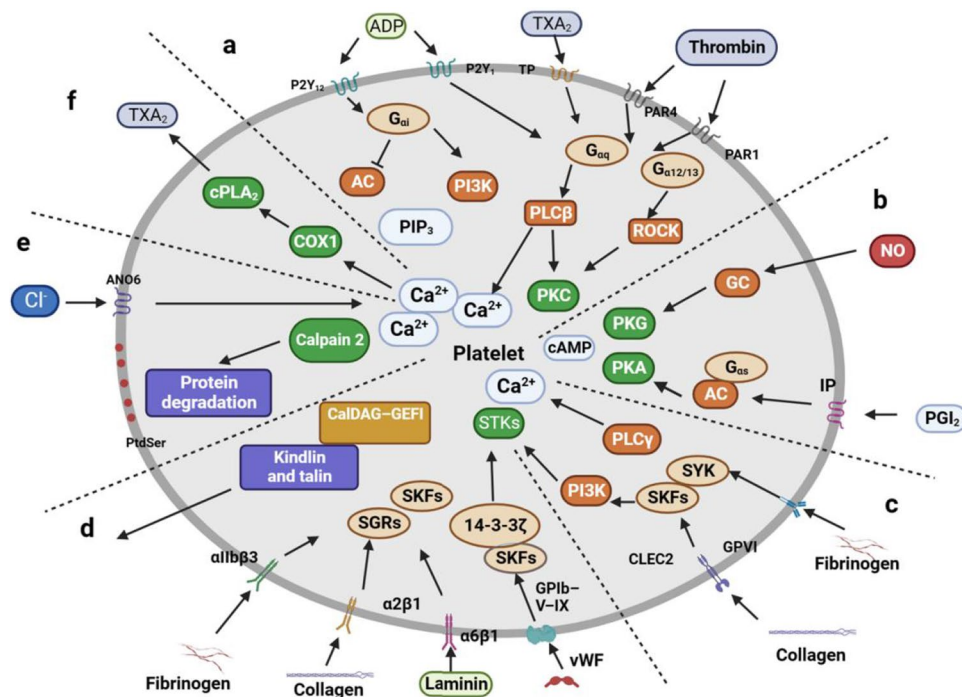


Fig. 1 Main signal events and responses in the process of platelet activation (**a**) The release of ADP and thromboxane A₂(TXA₂) activates purine receptors P2Y₁₂, P2Y₁, TXA₂ receptors (TP) and platelet activation. P2Y₁₂ inhibits AC but stimulates phosphoinositide 3 kinase (PI3Ks) via G_{ai} protein. The P2Y₁ and TP signals activate the G_q protein, which stimulates phospholipase C β (PLC β) to release Ca²⁺ into the cytoplasm, leading to protein kinase C (PKC) activation and downstream signaling events. The TP receptor also activates G₁₂ and 13 proteins, leading to the activation of rho-associated protein kinase (ROCK) and contributing to platelet shape change and diffusion. thrombin activates platelets by binding to G_q-coupled receptors PAR1 and PAR4. **b** Platelet inhibition is achieved through nitric oxide (NO) and prostaglandin I₂ (PGI₂) IP receptor-mediated activation of guanylate cyclase (GC) and adenylate cyclase (AC), leading to protein kinase G (PKG) and protein kinase A (PKA) activation. **c** Enhance platelet activation. GPVI and type c lectin-like receptor 2 (CLEC2) induce strong signals through the protein tyrosine kinase pathway, resulting in the release of Ca²⁺ into the cytoplasm.

d Platelet activation is initiated by the interaction between adhesion receptors (integrin α 6 β 1, α 2 β 1, α IIb β 3 and glycoprotein Ib-V-IX complex) and their ligands, such as collagen and von Willebrand factor. This involves signal transduction through small G protein regulators (SGRs), src family kinases (SKFs) and serine/threonine protein kinases (STKs). The conformational changes of integrins, such as α IIb β 3, involve a pathway downstream of PLC, PKC and PI3K that leads from low to high affinity. This pathway includes CalDAG-GEFI (a guanosine nucleotide exchange factor regulated by Ca²⁺ and diacylglycerol), SGRs (RAS3A and RAPIB), as well as signaling molecules connected to the cytoskeleton (kindlin and talin subtypes). **e** Platelet membrane swelling and exposure of phosphatidylserine (PtdSer) occur through high Ca²⁺ mobilization agonists mediated by the ion channel anoctamin6 (ANO6), while calpain-2 mediates intracellular protein degradation. **f** The activation of Ca²⁺-dependent and protein kinase-dependent cytoplasmic phospholipase A₂ (cPLA₂) and cyclooxygenase 1 (COX1) mediates the release of TXA₂

[43]. Tumor cells have the ability to increase thrombin production through direct and indirect mechanisms. To proliferate and foster metastasis, these cells concurrently manipulate platelet behavior by enhancing the synthesis and/or secretion of various compounds that incite platelet activation and aggregation [44–46]. The initial observation that tumor cells can induce platelet activation is based on a co-culture system, wherein tumor-induced platelet aggregation (TCIPA) is mitigated following the introduction of direct thrombin inhibitors [47]. Despite TCIPA being characterized by a heightened platelet count and conventional platelet activation, contemporary studies have demonstrated alterations in platelet derivatives within cancer patients. It's been discovered that EGFRvIII-mRNA can be isolated from the platelets of glioma patients, and

PCA3-mRNA resides in the platelets of those with prostate cancer [48]. Furthermore, aberrant fusions of specific EML4-ALK genes have been detected in the platelets of patients suffering from non-small cell lung cancer [49]. Dysexpression of platelet protein can also be used as a diagnostic marker and prognostic factor. It has been reported that platelet concentrations of vascular endothelial growth factor (VEGF) are higher in cancer patients than in healthy individuals, including lung cancer patients [50, 51], Liver Cancer [52, 53] and colorectal cancer [54]. Furthermore, malignant tumor cells exhibit heightened thrombin production compared to benign tumors, underscoring thrombin's role as a potent platelet activator with significant pro-coagulant attributes [55]. Previous research has also indicated that the rise in thrombin levels is related

to the tumor's location, with lung cancer demonstrating a more significant increase compared to brain and pancreatic cancers [56]. Previous research has demonstrated that TF plays a crucial role in initiating platelet activation and aggregation, with TF being expressed on the cell membranes of various cancer cells [57]. Extracellular vesicles (EVs), a diverse collection of cell-derived membrane structures including exosomes, microvesicles, and apoptotic bodies, are released by cells and significantly contribute to blood coagulation, thereby facilitating the direct generation of thrombin in cancer patients. Numerous studies have identified that EVs can express TF, linking it to thrombin production across different types of cancer [58]. Furthermore, activated host cells, such as monocytes and cancer cells, can release EVs that bear transferrin on their surface, further enhancing platelet activation [59–61]. Infact, it is reported that transferrin potentiates thrombin/FXIIa and blocks AT's inactivation effect on coagulation proteases inducing activation coagulation and platelet[62]. EVs are also exposed to negatively charged phospholipids, such as phosphatidyl serine (PS), on adventitia lobules [63], is an effective substance to promote coagulation. Circulating TF and EVs are good biomarkers of thrombosis and disseminated intravascular coagulation (DIC) in different diseases [64, 65].

Mechanisms of cancer-induced coagulation

Platelets are closely related to the blood hypercoagulable state, which is an important cause of cancer-related venous thrombosis and tumors may affect the blood hypercoagulable state in many ways [66]. Existing studies have found that, tumor-derived G-CSF induces neutrophilia, triggering NET release and promoting thrombosis in murine model[67] and tumor-derived IL-6 induces thrombopoietin (TPO) expression in hepatocytes, and cancer cells have the ability to generate thrombin as well, promoting thrombosis and increasing the risk of thrombotic events in cancer patients[68–70]. Studies have found that about 5% of patients with idiopathic venous thromboembolism show latent malignant tumors, which are found within one year after the diagnosis of venous thromboembolism [71]. Previous studies have shown that 10%–57% of cancer patients have a high platelet count, which is associated with a poor prognosis [72]. Many cancer types, such as adenocarcinoma, ovarian cancer, brain cancer, gastric cancer, colon cancer, and lung cancer, have a higher risk of developing VTE, while in cancer types such as breast and prostate cancer, the risk of VTE is lower, indicating that there may be specific VTE pathways in different cancer types (Table 2). Additionally, cancer patients often exhibit a hypercoagulable state at the time of diagnosis, with a higher prevalence

Table 2 Primary Thrombogenic Mechanisms in various cancer types

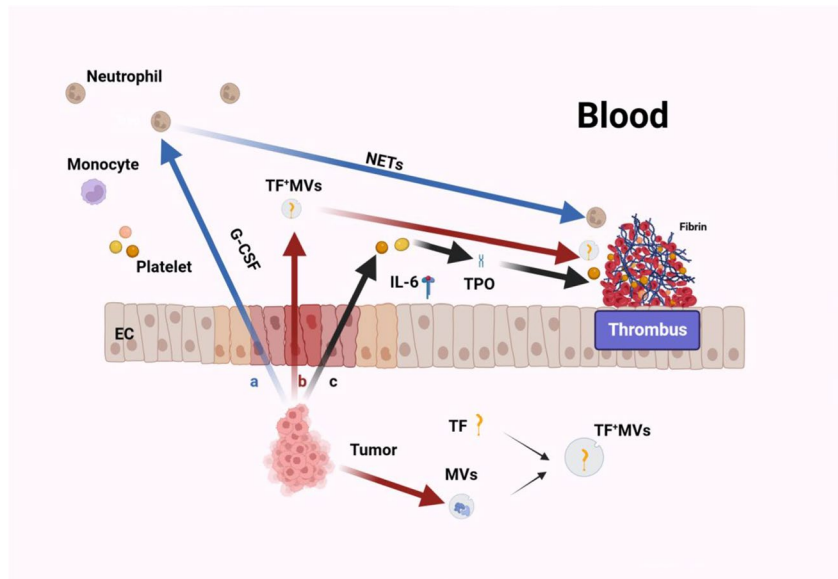
Tumor type	Primary Thrombogenic Mechanisms	Reference
Lung	Neutrophils and neutrophil extracellular traps (NETs), Leukocytosis	[80–82]
Ovary	Thrombocytopenia	[70, 83–85]
Brain	PDPN1 ⁺ MVs	[86, 87]
Colorectum	Leukocytosis	[88–93]
Pancreas	TF1 ⁺ MVs	[61, 94–96]

of certain prothrombotic gene mutations compared to the general population [73, 74]. This suggests a strong correlation between blood status and tumor growth, with potential implications for the role of blood coagulation gene polymorphisms in cancer development. Upon activation of prothrombin, thrombin is generated, which subsequently triggers fibrinogenesis and platelet activation to maintain hemostasis. In addition to its hemostatic function, thrombin, especially through its interaction with the receptor PAR1, is also thought to affect key mechanisms of tumor initiation, including cell survival, proliferation, and adhesion [75, 76]. Previous studies have found that F7rs510317A alleles are associated with an increased risk of breast cancer [77]. This allele is also linked to an elevation in the levels of FVII, the activating enzyme in the exogenous coagulation pathway, within circulation [78, 79].

As mentioned earlier, cancer patients with VTE have a more aggressive underlying cancer phenotype, disease progression, and treatment failure, and tumors may affect the blood hypercoagulable state in many ways (Fig. 2). Platelets play a crucial role in coagulation, indicating their close association with tumor progression. Markers of platelet activation and coagulation have been identified in primary tumor tissues [97].

FVII is structurally expressed in cancer cells under hypoxic conditions, and forms a complex with TF (TF/FVIIa) to enhance blood clotting activity. FX plays a pivotal role in both endogenous and exogenous coagulation pathways. Tumor cells produce a thrombin called a cancer coagulant, which activates FX [98]. Tissue factor pathway inhibitor (TFPI) is known to modulate the activity of transferrin, which triggers the exogenous coagulation pathway and represents the most extensively studied coagulation factor in the context of malignant tumors [99, 100]. TF is the center of current thrombosis models: exposed when injured. Following oncogene induction, expression of TF in tumor tissue is related to cell survival, proliferation, invasion, angiogenesis, and metastasis, and generally related to poor prognosis in different tumor types. As previously mentioned, TF interacts with plasma coagulation factors to facilitate the generation of thrombin, which exerts pleiotropic effects on various cellular processes. Thrombin

Fig. 2 Partial pathways of tumor affecting thrombosis **(a)** Tumor-derived G-CSF induces neutrophilia, triggering NET release and promoting thrombosis. **(b)** Tumor cells release TF1⁺MVs, causing thrombosis in tumor patients through circulation. **(c)** Tumor-derived IL-6 induces TPO expression in hepatocytes, promoting thrombosis and increasing the risk of thrombotic events in cancer patients



contributes to thrombosis by cleaving fibrinogen and activating protease activated receptors (PARs), which mediate a variety of cellular effects. However, when considering the biological function of the thrombin-PAR axis, it remains challenging to determine how this pathway specifically contributes to both blood coagulation and tumor progression. Studies have demonstrated that thrombin activates human platelets via PAR-1 and PAR-4, leading to the secretion of a diverse array of molecules, which facilitates the formation of thrombus [101, 102].

In cancer, transferrin is often structurally overexpressed and can act locally or remotely, and is carried by tumor-derived extracellular vesicles and released into the circulation [18]. In tumor tissue, blood coagulation can be activated by destruction of the blood vessel wall, leading to bleeding and intravascular coagulation, or extravascular coagulation due to increased vascular permeability and plasma extravasation [103]. In addition, the entry of metastatic cancer cells into the circulation and the recruitment/activation of inflammatory cells (immune thrombosis) may further amplify these processes.

Some biomarkers have been shown to be associated with the occurrence of VTE in cancer (Table 3). Parameters such as increased white blood cell and platelet count and decreased hemoglobin have been shown to be good predictors of the risk of venous thromboembolism in cancer patients. In addition, the increased concentrations of prothrombin fragments 1–2, soluble P-selectin, coagulation factor VIII, and D-dimer are closely bound up with an increased incidence of cancer VTE. In an extensive synthesis of eighteen studies incorporating thirty-six biomarkers, D-dimer levels and epidermal growth factor receptor (EGFR) mutations emerged as the paramount predictors for thromboembolic incidents.[104].

Role of platelets in cancer-related thrombosis

The role of platelets in CAT is increasingly recognized. Platelet-cancer cell interactions are involved in the regulation of cancer-associated thrombosis [110]. A noteworthy

Table 3 Selected biomarkers of coagulation / fibrinolysis and their relationship with hemostasis[61, 105–109]

Marker	Link to coagulation
Determination of thrombin production by specific substrate	Thrombin activity
Thrombin-antithrombin (TAT) complex	
Prothrombin 1–2 fragment (F1 + 2)	
TF + ^{ve} microparticles	
D-dimer	Fibrin degradation products
Soluble P-selectin	Biomarkers of platelet activation
Citrulline histone H3	Biomarkers of NETosis

clinical study has shown that patients with high-grade gliomas (HGGs) who exhibit a low platelet count — accounting for 25% of the participants — are at a heightened risk for VTE [86]. This discovery is particularly surprising when contrasted with the commonly observed phenomenon of elevated platelet counts in patients with solid tumors, suggesting a unique aspect of HGG biology. Moreover, the expression of P-selectin on the surface of activated platelets, and its release as soluble P-selectin into the circulation, has been identified as a contributory factor in thrombosis development [111–113]. Elevated levels of P-selectin might result in decreased platelet counts due to its role in augmenting platelet adhesion, potentially leading to enhanced platelet consumption [114, 115]. This theory is corroborated by the observation of increased soluble P-selectin levels in individuals with conditions that lead to platelet depletion before thrombotic events, such as disseminated intravascular coagulation, heparin-induced thrombocytopenia, and thrombotic thrombocytopenic purpura (TTP) [116]. Furthermore, the formation of nodules targeted by prothrombin activity is noted in specific pathological conditions characterized by extensive coagulation activity, including VTE in ovarian cancer [117], highlighting the involvement of podoplanin (PDPN) [87, 118]. PDPN induces platelet aggregation in a CLEC-2-dependent manner within human glioblastoma cells (LN319) [119], and PDPN expression is negatively correlated with platelet count, but positively correlated with D-dimer level [87]. These findings suggest that tumors may release PDPN1-containing microvesicles, which can activate circulating platelets and lead to an increased risk of venous thromboembolism in cancer patients (Fig. 2).

Activated platelets interact with the coagulation system directly or indirectly through MPs or secretory factors. Once activated, expression of anionic phospholipids on the procoagulant surface is involved in thrombin production, fibrin formation, and clotting [120]. In addition, the interaction between platelets and neutrophils leads to the formation of NETs. NETs can be formed without infection, such as in cancer and autoimmunity, and can promote thrombosis [121]. Activated platelets are involved in cancer-related thrombosis by providing procoagulant surfaces, releasing inflammatory molecules, and interacting with monocytes and neutrophils [120].

In cancer patients, platelets play a crucial role not only in the development of arterial thrombosis but also significantly contribute to venous thrombosis [122, 123]. Thrombocytosis is common in cancer patients, especially in gastrointestinal, lung, breast, and ovarian cancer [104]. Previous studies have found that individuals with elevated platelet counts prior to cancer diagnosis have a higher frequency of VTE compared to individuals with lower platelet counts [83], and similar results were observed when measuring platelet counts in cancer patients [84, 85]. The reduction of VTE in patients with multiple myeloma who are treated with thalidomide

or lenalidomide and low-dose aspirin is comparable to that achieved by low molecular weight heparin, indicating the involvement of platelets in VTE development among these patients [124]. In addition, use of aspirin was associated with reduced edge of venous thromboembolism in patients with ovarian cancer, but not in patients with breast cancer [125, 126]. Many studies have measured the biomarkers of platelet activating factor [123]. In general, these biomarkers are increased in cancer patients, but only a limited number of studies have determined whether they can predict venous thromboembolism in this group of patients. A recent study failed to find a link between various platelet activation markers, including PF4 and VTE, in patients with various cancer types [127]. However, a pancreatic cancer study found that increased PF4 levels were associated with an approximately three-fold increase in VTE risk [128]. This disparity highlights the significance of examining distinct cancer categories.

The role of platelets in thrombosis has also been investigated in murine cancer models. A study employing a syngeneic orthotopic model of pancreatic cancer has demonstrated that clopidogrel can mitigate the binding of tumor-derived microvesicles to sites of thrombosis [129]. We have observed that TF1 microvesicles are capable of activating platelets via thrombin, while clopidogrel can effectively mitigate the augmented effect of exogenous TF1 microvesicles on thrombosis in murine models [130]. Collectively, these fundamental and clinical investigations propose that the administration of antiplatelet agents may serve as a preventive measure against venous thromboembolism in select cancer patients [123].

Numerous platelet receptors and signaling pathways, in addition to the release of platelet agonists by malignant cells, are implicated in TCIPA [131–133]. At the same time, the specific mechanism of platelet aggregation in malignant tumor cells is related to the type of cancer cell [132]. Several cancer cell lines, including melanoma, are capable of producing ADP [133], glioblastoma [134], breast cancer [135, 136], lung cancer [137], pancreatic ductal adenocarcinoma [138], and TCIPA induced by fibrosarcoma [137]. The activation of the P2Y12 receptor initiates a positive feedback mechanism in ADP-induced platelet activation, leading to the release of more ADP, ATP, and calcium from dense granules [139]. In human MCF-7 breast cancer cells, the reduction of ADP levels and the inhibition of the ADP receptor P2Y12 were found to diminish platelet activation and aggregation [136].

Platelets in the tumor microenvironment and interaction with PD-L1

Further clinical retrospective studies provide additional evidence that platelets present in the TME contribute to chemotherapy resistance and are associated with early disease

recurrence and poor survival outcomes [140]. GPIIb/IIIa platelet markers were detected in 59% of HER2-negative breast cancer biopsies and were significantly associated with reduced efficacy of neoadjuvant therapy. Circulating platelets are not mere pliable sacs summoned by VWF to impede damaged blood vessels and avert hemorrhage [141]. Platelets indeed express PD-L1, which facilitates direct interaction with T cells and may have an impact on the efficacy of immune checkpoint therapy [142]. Studies have shown that the proliferation and activity of T cells bound to platelets decreased [143]. This may be linked to the significantly reduced efficacy of immune checkpoint therapy utilizing PD-1 antibodies, indicating the direct involvement of platelets in cancer-related immune evasion. In addition to the aforementioned tumor-promoting effects, there is compelling evidence that thrombin cleaves platelet glycoprotein A (GARP) repeats, resulting in the liberation of TGF- β derived from platelets [144, 145]. Activated platelets can also trigger innate immune responses by recruiting neutrophils and inducing reticulocytosis, thereby

unleashing a cascade of physiological events [146]. Neutrophils can block the function of cytotoxic T cells through the expression of inducible nitric oxide synthase (iNOS) and production of nitric oxide (NO) [147]. In addition, the presence of PD-L1 on neutrophils is associated with initiating T cell apoptosis through interference induced activation of T cells and subsequent interaction with PD-1 [148].

Tumor patients exhibiting a procoagulant state, who subsequently develop CAT, manifest significant tumor progression, limited treatment efficacy and poor overall survival. Current evidence from *in vitro* and *in vivo* studies suggests that the procoagulant environment not only facilitates tumor immune evasion but also impedes immunotherapeutic interventions. Revealing the intricate mechanisms by which endothelial cells orchestrate inflammation and thrombosis is pivotal in devising innovative strategies to augment the potency of immune checkpoint inhibition and targeted therapy (Fig. 3). A combined approach targeting both tumor-mediated coagulation and tumor-induced immune evasion

Fig. 3 Interaction between cancer-associated immune thrombosis and immune checkpoint inhibition. Immune thrombosis associated with cancer is formed by the interaction of a variety of cells. Activation of plasma coagulation and complement factor, as well as TF released by endothelial cells, platelets, monocytes and neutrophils can modify T cell responses in relation to immune checkpoint inhibition

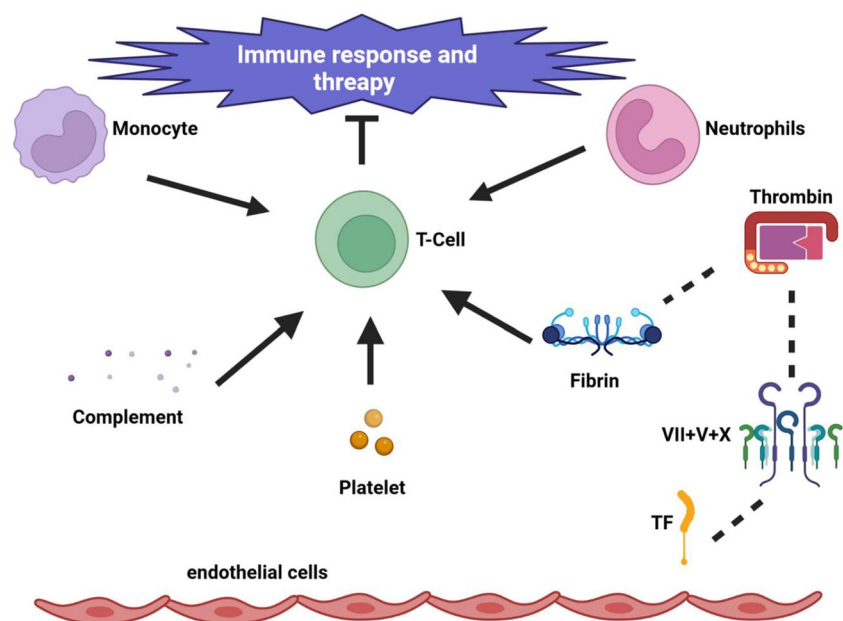


Table 4 Immune thrombosis inhibitors

Medicine	Targets		Reference
M7824	Fusion protein against PD-L1 and TGF- β	Cancer	[151]
Caplacizumab	Nano-body antagonizes VWF A1 domain	Thrombocytopenic purpura (TTP)	[149, 152]
Eculizumab	Anti-C5 monoclonal antibody	Atypical haemolytic uraemic syndrome (aHUS); Paroxysmal nocturnal hemoglobinuria (PNH)	[153, 154]
IPH5401	Anti-C5a receptor antibody	Cancer	[155]
Recombinant ADAMTS13	Cutting and inactivating VWF	TTP	[156]
Low molecular weight heparin (LMWH)	Activate antithrombin III and inhibit thrombin activation	Venous thromboembolism; Cancer	[157]

has been postulated to potentially bestow clinical benefits upon cancer patients. In view of the intricacy of the tumor microenvironment, targeted therapies aimed at specific pathways may augment response rates and ameliorate adverse effects (Table 4). Inhibition of the hypercoagulable state can be accomplished by administering anti-VWF nanoparticles that obstruct the A1 domain, thereby impeding VWF-mediated platelet binding. Alternatively, recombinant disintegrin and metalloproteinase member 13 (ADAMTS13) with thrombin reactive protein type 1 motif can cleave and inactivate VWF to provide a therapeutic option for supporting immune checkpoint inhibition [149]. In addition, the investigation of platelet mRNA analysis or detection of cancer cell activation induced by platelets is also being explored for diagnostic and monitoring purposes related to tumor development [150].

Therapeutic perspectives targeting tumor progression

Platelets are critical not only for hemostasis and coagulation but also, as extensive research has shown, in tumor progression, including aspects like invasion and exosmosis, closely linking them to CAT. This means that this property of platelets could be used for anti-tumor therapy. Integrin α IIb β 3, which is highly expressed in platelets and their progenitors, plays a pivotal role in platelet function, hemostasis, and arterial thrombosis, as well as in tumor progression, including cell proliferation and metastasis [158]. The binding of abciximab to integrin α IIb β 3 creates steric hindrance, preventing fibrinogen and other ligands from interacting with the integrin, thus interfering with platelet aggregation and thrombosis [159]. Caplacizumab, a humanized immunoglobulin fragment, specifically targets the A1 domain of vWF, disrupting the interaction between vWF and the platelet receptor GP1b- α [160]. This action prevents platelet adhesion, a critical step in the formation of microvascular thrombosis, and has been shown to normalize platelet counts in preclinical studies [160, 161]. Currently approved for the treatment of acquired TTP [162–164], but there is no relevant research in the field of cancer. The combination of antitumor drugs with Caplacizumab may be a potential treatment to reduce the occurrence of CAT and improve the prognosis of tumor patients, which needs further research to prove. Whilding et al. constructed α V β 3 specific CAR T cells and evaluated their antitumor function in vitro and in vivo preclinical models. These α V β 3-CAR T cells rapidly and specifically targeted α V β 3-positive tumor cells, secreting IFN- γ and IL-2. In a mouse xenograft model of metastatic A-375 melanoma, the intervention led to the complete eradication of melanoma lesions and long-term tumor-free survival. Integrins are also used to generate specific, controllable, and improved cytotoxicity of CAR-T therapies [165].

These studies illustrate the potential of the integrin family in anti-cancer and prevention of CAT.

Targeted drug delivery has become a new therapeutic strategy for cancer treatment. The tumor microenvironment is highly acidic compared to normal tissues, so Ph-sensitive nanosystems have been developed based on this difference. Drug release is activated in response to the acidic microenvironment and can enhance the therapeutic effects of cancer therapy. Crucially, platelets present promising avenues for targeted drug delivery. Nanocarriers can be engineered with peptides, enzymes, and antibodies [166]. For instance, a dual-targeted delivery system, consisting of paclitaxel-supported PEGylated-poly(lactic acid) nanoparticles and cyclic peptides, is designed to selectively bind to the platelet-derived PDGF/PDG β , offering improved treatment options for multiple myeloma [167].

Liposomes have been extensively studied as drug delivery systems, and many liposomal nanomedicines have been approved for clinical use [168]. Exosomes, which are biological extracellular vesicles, transmit signals through ligands or adhesion molecules on their membranes and have shown significant potential for targeted cancer drug delivery due to their origin from tumor cells [169]. Additionally, virus simulation systems are being developed for cancer diagnosis and targeted therapy. These systems mimic viruses in structure but lack genetic material, allowing them to be ingested by host cells without causing infection. Importantly, nanocarriers can be functionalized with small molecules, aptamers, and antibodies that possess high specificity and affinity [168]. There's growing evidence that these platelet-targeted drug delivery systems can enhance the efficacy of cancer treatments while minimizing potential side effects.

Conclusion and future prospects

Cancer-related venous thromboembolism is a well-known disease, but little is known about the potential biological mechanism of linking tumors to thrombosis [170]. Platelets play an important role in the process of blood coagulation. From a clinical point of view, platelet therapy may reduce the incidence of CAT and the rate of tumor progression. The use of some antiplatelet drugs may have a certain effect on the treatment of tumors (Table 5). Patient-related factors (including acquired and genetic determinants), cancer biology, and antineoplastic therapy are thought to play a synergistic role in venous thrombosis [11]. It is worth noting that the hemostatic system in cancer patients not only causes thrombosis, but also contributes to the growth and spread of tumors, suggesting cancer-related coagulation disorders may not be an accidental phenomenon [171].

In view of the adverse effects of venous thromboembolization on the prognosis of patients, disease prevention in cancer patients is important. It has been difficult to identify

Table 5 Commonly used and potential antiplatelet drugs [172–177]

Target	Drugs	Inhibition mechanism	Platelet function testing
Current target			
COX1	Aspirin and NSAIDs	Block the formation of TXA2 (autocrine)	LTA, PFA-100
P2Y12	Clopidogrel and prasugrel	Irreversible inhibition of ADP receptor (autocrine)	LTA, PAGT, VP
	ticagrelor	Reversible inhibition of ADP receptor (autocrine)	LTA, PAGT, VP
Integrin α IIb β 3	Abciximab, eptifibatide and tirofiban	Inhibition of integrin α IIb β 3 aggregation	LTA, PAGT
PAR1	Vorapaxar	Block thrombin receptor	LTA, PAGT
Potential target			
Factor Xa or thrombin		Inhibition of coagulation	TEM, Blood clotting test
GPIb-V-IX		Hinder vWF adhesion	PFA-100, microfluidics
GPVI and CLEC2	revacept	Block ITAM-like signal receptor	LTA, microfluidics
PI3K β and PDI		Inhibition of integrin α IIb β 3 aggregation	LTA, microfluidics
PAR1 and PAR4	pepducins	Block thrombin receptor	LTA, PAGT

LTA light transmission aggregometry, *PDI* protein disulfide isomerase, *TEM* thrombo-elastometry, *TKI* tyrosine kinase inhibitor, *VP* VASP phosphorylation, *PAGT* platelet aggregation test

coagulation and fibrinolysis biomarkers. There is a strong correlation between TF and primary tumor types, while fibrinolysis is more closely related to the cellular composition of the TME [178], which requires exploration of the kinetics of coagulation and fibrinolysis simultaneously [8]. It was recently reported that longitudinal evaluation of D-dimer significantly improved the estimation of VTE risk in cancer patients [179]. Functional analysis of thrombin production is also used in the early diagnosis of postoperative recurrence of breast cancer [180]. Because studies in the general population have shown that venous thromboembolism is hereditary, genetic markers, especially genetic polymorphisms, are attractive candidates for predicting cancer-related venous thromboembolism. In addition, given the role of hemostatic components in cancer development, these genetic markers may also predict cancer susceptibility and/or progression [11].

Thrombosis in tumor patients may be different from VTE in the general population. According to tumor models, some biological mechanisms are more related than others. In terms of the susceptibility and progression of cancer, the few studies so far have mainly focused on oral squamous cell carcinoma, breast cancer, and gastrointestinal cancer. Twelve and nine genetic polymorphisms related to cancer susceptibility and progression have been reported. It is worth noting that most of these variants have not been confirmed as predictors of cancer-related venous thromboembolism [3]. However, according to rare data, hemostatic gene polymorphisms seem to have an impact on cancer occurrence, which seems to be specific to cancer models, but may also be related to tumor staging. Taking into account thrombus prevention, cancer screening, prognostic assessment, and the development of potential antineoplastic therapy targeting coagulation, the scientific basis of these observations requires further investigation [11].

Blood coagulation is closely related to cancer, and platelets are an important player in blood clotting. The development of strategies that selectively target blood coagulation and simultaneously affect complement factors and the inflammatory response may promote anti-tumor effects. Further research is needed to fully understand the interaction of these pathways in the TME. Targeted coagulation, as a bridge between innate and acquired immune systems, may provide a new strategy to overcome drug resistance to checkpoint inhibition [181, 182]. Specifically, targeted platelets may provide a novel idea for the treatment of tumors and tumor-related embolism in the future.

Author contributions Conceptualization, Y.-F.Z., J.-T.Z., B.Z., S.-H.B., X.-J.L., H.-Q.W., Y.C. and H.Z.; L.-L.Z., X.-H.X., S.X., Z.-Q.S. methodology, Y.-F.Z., J.-T.Z., B.Z., S.-H.B., X.-J.L., H.-Q.W., Y.C. and H.Z.; L.-L.Z., X.-H.X., S.X., Z.-Q.S.; writing—original draft preparation, Y.-F.Z.; writing—review and editing, Y.-F.Z. and J.-T.Z. contributed equally to this study. All authors have read and agreed to the published version of the manuscript.

Funding The present study was funded by the National Natural Science Foundation of China (82172776), Tianjin Science and Technology Plan Project (19ZXDBSY00060), Tianjin Key Medical Discipline (Specialty) Construction Project (TJYXZDXK-061B), and Diversified Input Project of Tianjin National Natural Science Foundation (21JCYBJC01770).

Data availability The authors declare that no new data were created for this narrative review.

Declarations

Ethics statement Not applicable.

Conflict of interest The authors declare that the research was conducted in the absence of any commercial or financial relationship.

References

- Anderson E, Durstine JL (2019) Physical activity, exercise, and chronic diseases: a brief review. *Sports Med Health Sci* 1(1):3–10
- Curigliano G, Lenihan D, Fradley M et al (2020) Management of cardiac disease in cancer patients throughout oncological treatment: ESMO consensus recommendations. *Ann Oncol: Off J Eur Soc Med Oncol* 31(2):171–190
- Tavares V, Neto BV, Vilas-Boas MI et al (2022) Impact of hereditary thrombophilia on cancer-associated thrombosis, tumour susceptibility and progression: a review of existing evidence. *Biochim Biophys Acta* 1877(5):188778
- Elyamany G, Alzaharani AM, Bukhary E (2014) Cancer-associated thrombosis: an overview. *Clin Med Insights Oncol* 8:129–137
- Heit JA, Spencer FA, White RH (2016) The epidemiology of venous thromboembolism. *J Thromb Thrombolysis* 41(1):3
- Laporte S, Mismetti P, Décousus H et al (2008) Clinical predictors for fatal pulmonary embolism in 15,520 patients with venous thromboembolism: findings from the Registro Informatizado de la Enfermedad TromboEmbolica venosa (RIETE) Registry. *Circulation* 117(13):1711–1716
- Farge D, Frere C, Connors JM et al (2019) 2019 international clinical practice guidelines for the treatment and prophylaxis of venous thromboembolism in patients with cancer. *Lancet Oncol* 20(10):e566–e581
- Khorana AA, Cohen AT, Carrier M et al (2020) Prevention of venous thromboembolism in ambulatory patients with cancer. *ESMO Open* 5(6):e000948
- Gran OV, Smith EN, Brækkan SK et al (2016) Joint effects of cancer and variants in the factor 5 gene on the risk of venous thromboembolism. *Haematologica* 101(9):1046–1053
- Reitsma PH, Versteeg HH, Middeldorp S (2012) Mechanistic view of risk factors for venous thromboembolism. *Arterioscler Thromb Vasc Biol* 32(3):563–568
- Tavares V, Pinto R, Assis J et al (2020) Venous thromboembolism GWAS reported genetic makeup and the hallmarks of cancer: linkage to ovarian tumour behaviour. *Biochim Biophys Acta* 1873(1):188331
- Skille H, Paulsen B, Hveem K et al (2021) Prothrombotic genotypes and risk of venous thromboembolism in occult cancer. *Thromb Res* 205:17–23
- Buijs JT, Versteeg HH (2020) Genes and proteins associated with the risk for cancer-associated thrombosis. *Thromb Res* 191:S43–S49
- Moik F, Chan W-SE, Wiedemann S et al (2021) Incidence, risk factors, and outcomes of venous and arterial thromboembolism in immune checkpoint inhibitor therapy. *Blood* 137(12):1669–1678
- Sussman TA, Li H, Hobbs B et al (2021) Incidence of thromboembolism in patients with melanoma on immune checkpoint inhibitor therapy and its adverse association with survival. *J Immunother Cancer* 9(1):e001719
- Roopkumar J, Swaidani S, Kim AS et al (2021) Increased incidence of venous thromboembolism with cancer immunotherapy. *Med (New York, NY)* 2(4):423–434
- Galmiche A, Rak J, Roumenina LT et al (2022) Coagulome and the tumor microenvironment: an actionable interplay. *Trends Cancer* 8(5):369–383
- Falanga A, Schieppati F, Russo L (2019) Pathophysiology 1. Mechanisms of Thrombosis in Cancer Patients. *Cancer Treat Res* 179:11–36
- Khorana AA (2010) Venous thromboembolism and prognosis in cancer. *Thromb Res* 125(6):490–493
- Posch F, Riedl J, Reitter E-M et al (2016) Hypercoagulability, venous thromboembolism, and death in patients with cancer. A multi-state model. *Thromb Haemost* 115(4):817–826
- Riedl JM, Schwarzenbacher E, Moik F et al (2022) Patterns of thromboembolism in patients with advanced pancreatic cancer undergoing first-line chemotherapy with FOL-FIRINOX or gemcitabine/nab-paclitaxel. *Thromb Haemost* 122(4):633–645
- Ay C, Pabinger I, Cohen AT (2017) Cancer-associated venous thromboembolism: Burden, mechanisms, and management. *Thromb Haemost* 117(2):219–230
- Gao S, Ma J-J, Lu C (2014) Venous thromboembolism risk and erythropoiesis-stimulating agents for the treatment of cancer-associated anemia: a meta-analysis. *Tumour Biol* 35(1):603–613
- Ashrani AA, Gullerud RE, Petterson TM et al (2016) Risk factors for incident venous thromboembolism in active cancer patients: a population based case-control study. *Thromb Res* 139:29–37
- Cohen AT, Katholing A, Rietbrock S et al (2017) Epidemiology of first and recurrent venous thromboembolism in patients with active cancer. A population-based cohort study. *Thromb Haemost* 117(1):57–65
- Königsbrügge O, Pabinger I, Ay C (2014) Risk factors for venous thromboembolism in cancer: novel findings from the Vienna cancer and thrombosis study (CATS). *Thromb Res* 133(Suppl 2):S39–S43
- Fiedler T, Rabe M, Mundkowski RG et al (2018) Adipose-derived mesenchymal stem cells release microvesicles with procoagulant activity. *Int J Biochem Cell Biol* 100:49–53
- Farge D, Le Maignan C, Doucet L et al (2019) Women, thrombosis, and cancer. *Thromb Res* 181(Suppl 1):S47–S53
- Würtz M, Grove EL, Corraini P et al (2020) Comorbidity and risk of venous thromboembolism after hospitalization for first-time myocardial infarction: a population-based cohort study. *J Thromb Haemost* 18(8):1974–1985
- Chaturvedi S, Braunstein EM, Yuan X et al (2020) Complement activity and complement regulatory gene mutations are associated with thrombosis in APS and CAPS. *Blood* 135(4):239–251
- Karasu A, Engbers MJ, Cushman M et al (2016) Genetic risk factors for venous thrombosis in the elderly in a case-control study. *J Thromb Haemost* 14(9):1759–1764
- Ghouse J, Tragante V, Ahlberg G et al (2023) Genome-wide meta-analysis identifies 93 risk loci and enables risk prediction equivalent to monogenic forms of venous thromboembolism. *Nat Genet* 55(3):399–409
- Raghunathan S, Rayes J, Sen Gupta A (2022) Platelet-inspired nanomedicine in hemostasis thrombosis and thromboinflammation. *J Thromb Haemost* 20(7):1535–1549
- Levi M (2019) Disseminated intravascular coagulation in cancer: an update. *Semin Thromb Hemost* 45(4):342–347
- Mitrugno A, Tassi Yunga S, Sylman JL et al (2019) The role of coagulation and platelets in colon cancer-associated thrombosis. *Am J Physiol Cell Physiol* 316(2):C264–C273
- Ghoshal K, Bhattacharyya M (2014) Overview of platelet physiology: its hemostatic and nonhemostatic role in disease pathogenesis. *Sci World J* 2014:781857
- Wang S, Li Z, Xu R (2018) Human cancer and platelet interaction, a potential therapeutic target. *Int J Mol Sci* 19(4):1246. <https://doi.org/10.3390/ijms19041246>
- Levin J, Conley CL (1964) Thrombocytosis associated with malignant disease. *Arch Intern Med* 114:497–500
- Wojtukiewicz MZ, Sierko E, Hempel D et al (2017) Platelets and cancer angiogenesis nexus. *Cancer Metastasis Rev* 36(2):249–262

40. Zhang X, Ran Y (2015) Prognostic role of elevated platelet count in patients with lung cancer: a systematic review and meta-analysis. *Int J Clin Exp Med* 8(4):5379–5387
41. Gu D, Szallasi A (2017) Thrombocytosis portends adverse prognosis in colorectal cancer: a meta-analysis of 5,619 patients in 16 individual studies. *Anticancer Res* 37(9):4717–4726
42. Ji Y, Sheng L, Du X et al (2015) Elevated platelet count is a strong predictor of poor prognosis in stage I non-small cell lung cancer patients. *Platelets* 26(2):138–142
43. Harano K, Kogawa T, Wu J et al (2017) Thrombocytosis as a prognostic factor in inflammatory breast cancer. *Breast Cancer Res Treat* 166(3):819–832
44. Zarà M, Canobbio I, Visconte C et al (2018) Molecular mechanisms of platelet activation and aggregation induced by breast cancer cells. *Cell Signal* 48:45–53
45. Reddel CJ, Tan CW, Chen VM (2019) Thrombin generation and cancer: contributors and consequences. *Cancers (Basel)* 11(1):100. <https://doi.org/10.3390/cancers11010100>
46. Chang J, Jiang L, Wang Y et al (2015) 12/15 Lipoxygenase regulation of colorectal tumorigenesis is determined by the relative tumor levels of its metabolite 12-HETE and 13-HODE in animal models. *Oncotarget* 6(5):2879–2888
47. Heinmöller E, Weinel RJ, Heidtmann HH et al (1996) Studies on tumor-cell-induced platelet aggregation in human lung cancer cell lines. *J Cancer Res Clin Oncol* 122(12):735–744
48. Nilsson RJA, Balaj L, Hulleman E et al (2011) Blood platelets contain tumor-derived RNA biomarkers. *Blood* 118(13):3680–3683
49. Nilsson RJA, Karachaliou N, Berenguer J et al (2016) Rearranged EML4-ALK fusion transcripts sequester in circulating blood platelets and enable blood-based crizotinib response monitoring in non-small-cell lung cancer. *Oncotarget* 7(1):1066–1075
50. Sabrkhanly S, Kuijpers MJE, van Kuijk SMJ et al (2017) A combination of platelet features allows detection of early-stage cancer. *Eur J Cancer* 80:5–13. <https://doi.org/10.1016/j.ejca.2017.04.010>
51. Yao L, Dong H, Luo Y et al (2014) Net platelet angiogenic activity (NPAA) correlates with progression and prognosis of non-small cell lung cancer. *PLoS One* 9(4):e96206
52. Alkozai EM, Porte RJ, Adelmeijer J et al (2015) Levels of angiogenic proteins in plasma and platelets are not different between patients with hepatitis B/C-related cirrhosis and patients with cirrhosis and hepatocellular carcinoma. *Platelets* 26(6):577–582
53. Kim SJ, Choi IK, Park KH et al (2004) Serum vascular endothelial growth factor per platelet count in hepatocellular carcinoma: correlations with clinical parameters and survival. *Jpn J Clin Oncol* 34(4):184–190
54. Peterson JE, Zurakowski D, Italiano JE et al (2012) VEGF, PF4 and PDGF are elevated in platelets of colorectal cancer patients. *Angiogenesis* 15(2):265–273
55. Duvernay MT, Temple KJ, Maeng JG et al (2017) Contributions of protease-activated receptors PAR1 and PAR4 to thrombin-induced GPIIb/IIIa activation in human platelets. *Mol Pharmacol* 91(1):39–47
56. Reitter EM, Kaider A, Ay C et al (2016) Longitudinal analysis of hemostasis biomarkers in cancer patients during antitumor treatment. *J Thromb Haemost* 14(2):294–305
57. Arnout J, Hoylaerts MF, Lijnen HR (2006) Haemostasis. *Handb Exp Pharmacol* 176(Pt 2):1–41. https://doi.org/10.1007/3-540-36028-x_1
58. Gardiner C, Harrison P, Belting M et al (2015) Extracellular vesicles, tissue factor, cancer and thrombosis - discussion themes of the ISEV 2014 Educational Day. *J Extracell Vesicles* 4:26901
59. Mackman N, Tilley RE, Key NS (2007) Role of the extrinsic pathway of blood coagulation in hemostasis and thrombosis. *Arterioscler Thromb Vasc Biol* 27(8):1687–1693
60. Gregory SA, Morrissey JH, Edgington TS (1989) Regulation of tissue factor gene expression in the monocyte procoagulant response to endotoxin. *Mol Cell Biol* 9(6):2752–2755
61. Geddings JE, Mackman N (2013) Tumor-derived tissue factor-positive microparticles and venous thrombosis in cancer patients. *Blood* 122(11):1873–1880
62. Tang X, Zhang Z, Fang M et al (2020) Transferrin plays a central role in coagulation balance by interacting with clotting factors. *Cell Res* 30(2):119–132
63. György B, Szabó TG, Pásztói M et al (2011) Membrane vesicles, current state-of-the-art: emerging role of extracellular vesicles. *Cell Mol Life Sci: CMLS* 68(16):2667–2688
64. Reddy EC, Rand ML (2020) Procoagulant phosphatidylserine-exposing platelets and. *Front Cardiovasc Med* 7:15
65. Tesselaar MET, Romijn FPHTM, Van Der Linden IK et al (2007) Microparticle-associated tissue factor activity: a link between cancer and thrombosis? *J Thromb Haemost* 5(3):520–527
66. Mulder FI, Horváth-Puhó E, van Es N et al (2021) Venous thromboembolism in cancer patients: a population-based cohort study. *Blood* 137(14):1959–1969
67. Gomes T, Várady CBS, Lourenço AL et al (2019) IL-1 β blockade attenuates thrombosis in a neutrophil extracellular trap-dependent breast cancer model. *Front Immunol* 10:2088
68. Reusswig F, Fazel Modares N, Brechtenkamp M et al (2021) Efficiently restored thrombopoietin production by Ashwell-Morell receptor and IL-6R induced Janus Kinase 2/Signal transducer and activator of transcription signaling early after partial hepatectomy. *Hepatology* 74(1):411–427
69. Zucchella M, Dezza L, Pacchiarini L et al (1989) Human tumor cells cultured “in vitro” activate platelet function by producing ADP or thrombin. *Haematologica* 74(6):541–545
70. Stone RL, Nick AM, McNeish IA et al (2012) Paraneoplastic thrombocytosis in ovarian cancer. *N Engl J Med* 366(7):610–618
71. van Es N, Le Gal G, Otten H-M et al (2017) Screening for occult cancer in patients with unprovoked venous thromboembolism: a systematic review and meta-analysis of individual patient data. *Ann Intern Med* 167(6):410–417
72. Sierko E, Wojtukiewicz MZ (2004) Platelets and angiogenesis in malignancy. *Semin Thromb Hemost* 30(1):95–108. <https://doi.org/10.1055/s-2004-822974>
73. Pihusch R, Danzl G, Scholz M et al (2002) Impact of thrombophilic gene mutations on thrombosis risk in patients with gastrointestinal carcinoma. *Cancer* 94(12):3120–3126
74. Tinholt M, Viken MK, Dahm AE et al (2014) Increased coagulation activity and genetic polymorphisms in the F5, F10 and EPCR genes are associated with breast cancer: a case-control study. *BMC Cancer* 14:845
75. Maragoudakis ME, Tsopanoglou NE, Andriopoulou P et al (2000) Effects of thrombin/thrombosis in angiogenesis and tumour progression. *Matrix Biol: J Int Soc Matrix Biol* 19(4):345–351
76. Green D (2010) Karpatkin S Role of thrombin as a tumor growth factor. *Cell Cycle (Georgetown, Tex.)* 9(4):656–661
77. Eroğlu A, Öztürk A, Akar N (2011) Association between the -402GA, -401GT, and -323ins10-bp polymorphisms of factor VII gene and breast cancer. *Breast Cancer (Tokyo, Japan)* 18(4):282–285
78. Fang J, Yuan Q, Du Z et al (2021) Contribution of factor VII polymorphisms to coagulopathy in patients with isolated traumatic brain injury. *Clin Neurol Neurosurg* 208:106836
79. Ken-Dror G, Drenos F, Humphries SE et al (2010) Haplotype and genotype effects of the F7 gene on circulating factor VII, coagulation activation markers and incident coronary heart disease in UK men. *J Thromb Haemost* 8(11):2394–2403
80. DuPre SA, Hunter KW (2007) Murine mammary carcinoma 4T1 induces a leukemoid reaction with splenomegaly: association with tumor-derived growth factors. *Exp Mol Pathol* 82(1):12–24

81. Kowanetz M, Wu X, Lee J et al (2010) Granulocyte-colony stimulating factor promotes lung metastasis through mobilization of Ly6G+Ly6C+ granulocytes. *Proc Natl Acad Sci U S A* 107(50):21248–21255
82. Demers M, Krause DS, Schatzberg D et al (2012) Cancers predispose neutrophils to release extracellular DNA traps that contribute to cancer-associated thrombosis. *Proc Natl Acad Sci U S A* 109(32):13076–13081
83. Jensvoll H, Blix K, Brækkan SK et al (2014) Platelet count measured prior to cancer development is a risk factor for future symptomatic venous thromboembolism: the Tromsø Study. *PLoS One* 9(3):e92011
84. Khorana AA, Francis CW, Culakova E et al (2005) Risk factors for chemotherapy-associated venous thromboembolism in a prospective observational study. *Cancer* 104(12):2822–2829
85. Simanek R, Vormittag R, Ay C et al (2010) High platelet count associated with venous thromboembolism in cancer patients: results from the Vienna Cancer and Thrombosis Study (CATS). *J Thromb Haemost* 8(1):114–120
86. Thaler J, Ay C, Kaider A et al (2014) Biomarkers predictive of venous thromboembolism in patients with newly diagnosed high-grade gliomas. *Neuro-oncology* 16(12):1645–1651
87. Riedl J, Preusser M, Nazari PMS et al (2017) Podoplanin expression in primary brain tumors induces platelet aggregation and increases risk of venous thromboembolism. *Blood* 129(13):1831–1839
88. Khorana AA, Kuderer NM, Culakova E et al (2008) Development and validation of a predictive model for chemotherapy-associated thrombosis. *Blood* 111(10):4902–4907
89. Granger JM, Kontoyiannis DP (2009) Etiology and outcome of extreme leukocytosis in 758 nonhematologic cancer patients: a retrospective, single-institution study. *Cancer* 115(17):3919–3923
90. Kasuga I, Makino S, Kiyokawa H et al (2001) Tumor-related leukocytosis is linked with poor prognosis in patients with lung carcinoma. *Cancer* 92(9):2399–2405
91. Ruka W, Rutkowski P, Kaminska J et al (2001) Alterations of routine blood tests in adult patients with soft tissue sarcomas: relationships to cytokine serum levels and prognostic significance. *Ann Oncol: Off J Eur Soc Med Oncol* 12(10):1423–1432
92. Blix K, Jensvoll H, Brækkan SK et al (2013) White blood cell count measured prior to cancer development is associated with future risk of venous thromboembolism—the Tromsø study. *PLoS One* 8(9):e73447
93. Pabinger I, Posch F (2014) Flamethrowers: blood cells and cancer thrombosis risk. *Hematol Am Soc Hematol Educ Program* 2014(1):410–417
94. Khorana AA, Francis CW, Menzies KE et al (2008) Plasma tissue factor may be predictive of venous thromboembolism in pancreatic cancer. *J Thromb Haemost* 6(11):1983–1985
95. Thaler J, Ay C, Mackman N et al (2012) Microparticle-associated tissue factor activity, venous thromboembolism and mortality in pancreatic, gastric, colorectal and brain cancer patients. *J Thromb Haemost* 10(7):1363–1370
96. Bharthuar A, Khorana AA, Hutson A et al (2013) Circulating microparticle tissue factor, thromboembolism and survival in pancreaticobiliary cancers. *Thromb Res* 132(2):180–184
97. Verheul HM, Hoekman K, Lupu F et al (2000) Platelet and coagulation activation with vascular endothelial growth factor generation in soft tissue sarcomas. *Clin Cancer Res: Off J Am Assoc Cancer Res* 6(1):166–171
98. Gordon SG, Mielicki WP (1997) Cancer procoagulant: a factor X activator, tumor marker and growth factor from malignant tissue. *Blood Coagul Fibrinolysis: Int J Haemost Thromb* 8(2):73–86
99. Tinholt M, Vollan HKM, Sahlberg KK et al (2015) Tumor expression, plasma levels and genetic polymorphisms of the coagulation inhibitor TFPI are associated with clinicopathological parameters and survival in breast cancer, in contrast to the coagulation initiator TF. *Breast Cancer Res: BCR* 17(1):44
100. Bazzarelli AK, Scheer AS, Tai LH et al (2016) Tissue factor pathway inhibitor gene polymorphism -33T → C predicts improved disease-free survival in colorectal cancer. *Ann Surg Oncol* 23(7):2274–2280
101. Han N, Jin K, He K et al (2011) Protease-activated receptors in cancer: a systematic review. *Oncol Lett* 2(4):599–608
102. Ruf W (2007) Tissue factor and PAR signaling in tumor progression. *Thromb Res* 120(Suppl 2):S7–12
103. Tumors DHF (2019) Wounds that do not heal—a historical perspective with a focus on the fundamental roles of increased vascular permeability and clotting. *Semin Thromb Hemost* 45(06):576–592
104. Hisada Y, Mackman N (2017) Cancer-associated pathways and biomarkers of venous thrombosis. *Blood* 130(13):1499–1506
105. Lundbeck M, Krag AE, Christensen TD et al (2020) Thrombin generation, thrombin-antithrombin complex, and prothrombin fragment F1+2 as biomarkers for hypercoagulability in cancer patients. *Thromb Res* 186:80–85
106. Orcutt SJ, Pietropaolo C, Krishnaswamy S (2002) Extended interactions with prothrombinase enforce affinity and specificity for its macromolecular substrate. *J Biol Chem* 277(48):46191–46196
107. Cosmi B, Legnani C, Libra A et al (2023) D-Dimers in diagnosis and prevention of venous thrombosis: recent advances and their practical implications. *Pol Arch Intern Med* 133(11):16604. <https://doi.org/10.20452/pamw.16604>
108. Ronchetti L, Terrenato I, Ferretti M et al (2023) Correction: circulating cell free DNA and citrullinated histone H3 as useful biomarkers of NETosis in endometrial cancer. *J Exp Clin Cancer Res* 42(1):278
109. Wurtzel JGT, Lazar S, Askari S et al (2024) Plasma growth factors maintain constitutive translation in platelets to regulate reactivity and thrombotic potential. *Blood Adv* 8(6):1550–1566. <https://doi.org/10.1182/bloodadvances.2023011734>
110. Palacios-Acedo A-L, Langiu M, Crescence L et al (2022) Platelet and cancer-cell interactions modulate cancer-associated thrombosis risk in different cancer types. *Cancers (Basel)* 14(3):730. <https://doi.org/10.3390/cancers14030730>
111. Kansas GS (1996) Selectins and their ligands: current concepts and controversies. *Blood* 88(9):3259–3287
112. Vandendries ER, Furie BC, Furie B (2004) Role of P-selectin and PSGL-1 in coagulation and thrombosis. *Thromb Haemost* 92(3):459–466
113. Michelson AD, Barnard MR, Hechtman HB et al (1996) In vivo tracking of platelets: circulating degranulated platelets rapidly lose surface P-selectin but continue to circulate and function. *Proc Natl Acad Sci U S A* 93(21):11877–11882
114. Palabrica T, Lobb R, Furie BC et al (1992) Leukocyte accumulation promoting fibrin deposition is mediated in vivo by P-selectin on adherent platelets. *Nature* 359(6398):848–851
115. Frenette PS, Johnson RC, Hynes RO et al (1995) Platelets roll on stimulated endothelium in vivo: an interaction mediated by endothelial P-selectin. *Proc Natl Acad Sci U S A* 92(16):7450–7454
116. Chong BH, Murray B, Berndt MC et al (1994) Plasma P-selectin is increased in thrombotic consumptive platelet disorders. *Blood* 83(6):1535–1541
117. Glassman D, Bateman NW, Lee S et al (2022) Molecular Correlates of venous thromboembolism (VTE) in ovarian cancer. *Cancers (Basel)* 14(6):1496. <https://doi.org/10.3390/cancers14061496>

118. Tawil N, Bassawon R, Meehan B et al (2021) Glioblastoma cell populations with distinct oncogenic programs release podoplanin as procoagulant extracellular vesicles. *Blood Adv* 5(6):1682–1694
119. Suzuki-Inoue K, Kato Y, Inoue O et al (2007) Involvement of the snake toxin receptor CLEC-2, in podoplanin-mediated platelet activation, by cancer cells. *J Biol Chem* 282(36):25993–26001
120. Mege D, Aubert M, Lacroix R et al (2019) Involvement of platelets in cancers. *Semin Thromb Hemost* 45(6):569–575
121. Cedervall J, Hamidi A, Olsson A-K (2018) Platelets, NETs and cancer. *Thromb Res* 164(Suppl 1):S148–S152
122. Connolly GC, Phipps RP, Francis CW (2014) Platelets and cancer-associated thrombosis. *Semin Oncol* 41(3):302–310
123. Riedl J, Pabinger I, Ay C (2014) Platelets in cancer and thrombosis. *Hamostaseologie* 34(1):54–62
124. Larocca A, Cavallo F, Bringhen S et al (2012) Aspirin or enoxaparin thromboprophylaxis for patients with newly diagnosed multiple myeloma treated with lenalidomide. *Blood* 119(4):933–939
125. Shai A, Rennert HS, Rennert G et al (2014) Statins, aspirin and risk of thromboembolic events in ovarian cancer patients. *Gynecol Oncol* 133(2):304–308
126. Shai A, Rennert HS, Lavie O et al (2014) Statins, aspirin and risk of venous thromboembolic events in breast cancer patients. *J Thromb Thrombolysis* 38(1):32–38. <https://doi.org/10.1007/s11239-013-1015-8>
127. Riedl J, Hell L, Kaider A et al (2016) Association of platelet activation markers with cancer-associated venous thromboembolism. *Platelets* 27(1):80–85
128. Poruk KE, Firpo MA, Huerter LM et al (2010) Serum platelet factor 4 is an independent predictor of survival and venous thromboembolism in patients with pancreatic adenocarcinoma. *Cancer Epidemiol Biomarkers Prev: Publ Am Assoc Cancer Res Cosponsored Am Soc Prev Oncol* 19(10):2605–2610
129. Mezouar S, Darbousset R, Dignat-George F et al (2015) Inhibition of platelet activation prevents the P-selectin and integrin-dependent accumulation of cancer cell microparticles and reduces tumor growth and metastasis in vivo. *Int J Cancer* 136(2):462–475
130. Geddings JE, Hisada Y, Boulaftali Y et al (2016) Tissue factor-positive tumor microvesicles activate platelets and enhance thrombosis in mice. *J Thromb Haemost* 14(1):153–166
131. Medina C, Harmon S, Inkiewicz I et al (2012) Differential inhibition of tumour cell-induced platelet aggregation by the nicotinate aspirin prodrug (ST0702) and aspirin. *Br J Pharmacol* 166(3):938–949
132. Bastida E, Ordinas A, Jamieson GA (1981) Differing platelet aggregating effects by two tumor cell lines: absence of role for platelet-derived ADP. *Am J Hematol* 11(4):367–378
133. Boukerche H, Berthier-Vergnes O, Penin F et al (1994) Human melanoma cell lines differ in their capacity to release ADP and aggregate platelets. *Br J Haematol* 87(4):763–772
134. Bastida E, Escolar G, Almirall L et al (1986) Platelet activation induced by a human neuroblastoma tumor cell line is reduced by prior administration of ticlopidine. *Thromb Haemost* 55(3):333–337
135. Camez A, Dupuy E, Bellucci S et al (1986) Human platelet-tumor cell interactions vary with the tumor cell lines. *Invasion Metastasis* 6(6):321–334
136. Alonso-Escolano D, Strongin AY, Chung AW et al (2004) Membrane type-1 matrix metalloproteinase stimulates tumour cell-induced platelet aggregation: role of receptor glycoproteins. *Br J Pharmacol* 141(2):241–252
137. Jurasz P, Sawicki G, Duszyk M et al (2001) Matrix metalloproteinase 2 in tumor cell-induced platelet aggregation: regulation by nitric oxide. *Can Res* 61(1):376–382
138. Palacios-Acedo AL, Mezouar S, Mège D et al (2021) P2RY12-inhibitors reduce cancer-associated thrombosis and tumor growth in pancreatic cancers. *Front Oncol* 11:704945
139. Ballerini P, Dovizio M, Bruno A et al (2018) P2Y receptors in tumorigenesis and metastasis. *Front Pharmacol* 9:66. <https://doi.org/10.3389/fphar.2018.00066>
140. Ishikawa S, Miyashita T, Inokuchi M et al (2016) Platelets surrounding primary tumor cells are related to chemoresistance. *Oncol Rep* 36(2):787–794
141. Brass S (2001) Cardiovascular biology. Small cells, big issues. *Nature* 409(6817):145–147
142. Rolfes V, Idel C, Pries R et al (2018) PD-L1 is expressed on human platelets and is affected by immune checkpoint therapy. *Oncotarget* 9(44):27460–27470
143. Zamora C, Cantó E, Nieto JC et al (2017) Binding of platelets to lymphocytes: a potential anti-inflammatory therapy in rheumatoid arthritis. *J Immunol (Baltimore, Md: 1950)* 198(8):3099–3108
144. Metelli A, Wu BX, Riesenberger B et al (2020) Thrombin contributes to cancer immune evasion via proteolysis of platelet-bound GARP to activate LTGF- β . *Sci Transl Med* 12(525):eay4860. <https://doi.org/10.1126/scitranslmed.aay4860>
145. Dahmani A, Delisle J-S (2018) TGF- β in T cell biology: implications for cancer immunotherapy. *Cancers (Basel)* 10(6):194. <https://doi.org/10.3390/cancers10060194>
146. Lisman T (2018) Platelet-neutrophil interactions as drivers of inflammatory and thrombotic disease. *Cell Tissue Res* 371(3):567–576
147. Coffelt SB, Kersten K, Doornebal CW et al (2015) IL-17-producing $\gamma\delta$ T cells and neutrophils conspire to promote breast cancer metastasis. *Nature* 522(7556):345–348
148. de Kleijn S, Langereis JD, Leentjens J et al (2013) IFN- γ -stimulated neutrophils suppress lymphocyte proliferation through expression of PD-L1. *PLoS One* 8(8):e72249
149. Scully M, Cataland SR, Peyvandi F et al (2019) Caplacizumab treatment for acquired thrombotic thrombocytopenic purpura. *N Engl J Med* 380(4):335–346. <https://doi.org/10.1056/NEJMoA1806311>
150. Corvigno S, Johnson AM, Wong K-K et al (2022) Novel markers for liquid biopsies in cancer management: circulating platelets and extracellular vesicles. *Mol Cancer Ther* 21(7):1067–1075
151. Strauss J, Heery CR, Schlom J et al (2018) Phase I trial of M7824 (MSB0011359C), a bifunctional fusion protein targeting PD-L1 and TGF β , in advanced solid tumors. *Clin Cancer Res: Off J Am Assoc Cancer Res* 24(6):1287–1295
152. Goshua G, Sinha P, Hendrickson JE et al (2021) Cost effectiveness of caplacizumab in acquired thrombotic thrombocytopenic purpura. *Blood* 137(7):969–976
153. Loschi M, Porcher R, Barraco F et al (2016) Impact of eculizumab treatment on paroxysmal nocturnal hemoglobinuria: a treatment versus no-treatment study. *Am J Hematol* 91(4):366–370
154. Cofield R, Kukreja A, Bedard K et al (2015) Eculizumab reduces complement activation, inflammation, endothelial damage, thrombosis, and renal injury markers in aHUS. *Blood* 125(21):3253–3262
155. Afshar-Kharghan V (2017) The role of the complement system in cancer. *J Clin Invest* 127(3):780–789. <https://doi.org/10.1172/JCI90962>
156. Scully M, Knöbl P, Kentouche K et al (2017) Recombinant ADAMTS-13: first-in-human pharmacokinetics and safety in congenital thrombotic thrombocytopenic purpura. *Blood* 130(19):2055–2063
157. Wang T-F, Li A, Garcia D (2018) Managing thrombosis in cancer patients. *Res Pract Thromb Haemost* 2(3):429–438

158. Huang J, Li X, Shi X et al (2019) Platelet integrin α Ib β 3: signal transduction, regulation, and its therapeutic targeting. *J Hematol Oncol* 12(1):26
159. Giordano A, Musumeci G, D'Angelillo A et al (2016) Effects of glycoprotein IIb/IIIa antagonists: anti platelet aggregation and beyond. *Curr Drug Metab* 17(2):194–203
160. Ulrichs H, Silence K, Schoolmeester A et al (2011) Antithrombotic drug candidate ALX-0081 shows superior preclinical efficacy and safety compared with currently marketed antiplatelet drugs. *Blood* 118(3):757–765
161. Callewaert F, Roodt J, Ulrichs H et al (2012) Evaluation of efficacy and safety of the anti-VWF Nanobody ALX-0681 in a pre-clinical baboon model of acquired thrombotic thrombocytopenic purpura. *Blood* 120(17):3603–3610
162. Scully M, Cataland SR, Peyvandi F et al (2019) Caplacizumab treatment for acquired thrombotic thrombocytopenic purpura. *N Engl J Med* 380(4):335–346
163. Völker LA, Kaufeld J, Balduin G et al (2023) Impact of first-line use of caplacizumab on treatment outcomes in immune thrombotic thrombocytopenic purpura. *J Thromb Haemost* 21(3):559–572
164. Peyvandi F, Scully M, Kremer Hovinga JA et al (2016) Caplacizumab for acquired thrombotic thrombocytopenic purpura. *N Engl J Med* 374(6):511–522
165. Whilding LM, Halim L, Draper B et al (2019) CAR T-cells targeting the integrin α v β 6 and co-expressing the chemokine receptor CXCR2 demonstrate enhanced homing and efficacy against several solid malignancies. *Cancers (Basel)* 11(5):674. <https://doi.org/10.3390/cancers11050674>
166. Haider T, Sandha KK, Soni V et al (2020) Recent advances in tumor microenvironment associated therapeutic strategies and evaluation models. *Mater Sci Eng C Mater Biol Appl* 116:111229
167. Wang H, Liu H, Sun C et al (2021) Nanoparticles dual targeting both myeloma cells and cancer-associated fibroblasts simultaneously to improve multiple myeloma treatment. *Pharm* 13(2):274. <https://doi.org/10.3390/pharmaceutics13020274>
168. Liu Y, Castro Bravo KM, Liu J (2021) Targeted liposomal drug delivery: a nanoscience and biophysical perspective. *Nanoscale Horiz* 6(2):78–94
169. Shao J, Zaro J, Shen Y (2020) Advances in exosome-based drug delivery and tumor targeting: from tissue distribution to intracellular fate. *Int J Nanomedicine* 15:9355–9371
170. Roopkumar J, Swaidani S, Kim AS et al (2021) Increased incidence of venous thromboembolism with cancer immunotherapy - ScienceDirect. *Med* 2(4):423–434. <https://doi.org/10.1016/j.medj.2021.02.002>
171. Abe K, Yoshimura H, Tanaka H et al (2004) Comparison of conventional and diffusion-weighted MRI and proton MR spectroscopy in patients with mitochondrial encephalomyopathy, lactic acidosis, and stroke-like events. *Neuroradiology* 46(2):113–117
172. Crescente M, Mezzasoma AM, Del Pinto M et al (2011) Incomplete inhibition of platelet function as assessed by the platelet function analyzer (PFA-100) identifies a subset of cardiovascular patients with high residual platelet response while on aspirin. *Platelets* 22(3):179–187
173. Paniccia R, Antonucci E, Gori AM et al (2007) Different methodologies for evaluating the effect of clopidogrel on platelet function in high-risk coronary artery disease patients. *J Thromb Haemost* 5(9):1839–1847
174. Lele M, Sajid M, Wajih N et al (2001) Eptifibatid and 7E3, but not tirofiban, inhibit α (v) β (3) integrin-mediated binding of smooth muscle cells to thrombospondin and prothrombin. *Circulation* 104(5):582–587
175. Kujovich JL Factor V (2011) Leiden thrombophilia. *Genet Med* 13(1):1–16. <https://doi.org/10.1097/GIM.0b013e3181faa0f2>
176. Mori J, Pearce AC, Spalton JC et al (2008) G6b-B inhibits constitutive and agonist-induced signaling by glycoprotein VI and CLEC-2. *J Biol Chem* 283(51):35419–35427
177. Perrault C, Mangin P, Santer M et al (2003) Role of the intracellular domains of GPIb in controlling the adhesive properties of the platelet GPIb/V/IX complex. *Blood* 101(9):3477–3484
178. Saidak Z, Soudet S, Lottin M et al (2021) A pan-cancer analysis of the human tumor coagulome and its link to the tumor immune microenvironment. *Cancer Immunol Immunother: CII* 70(4):923–933
179. Posch F, Riedl J, Reitter E-M et al (2020) Dynamic assessment of venous thromboembolism risk in patients with cancer by longitudinal D-Dimer analysis: a prospective study. *J Thromb Haemost* 18(6):1348–1356
180. Marchetti M, Giaccherini C, Masci G et al (2020) Thrombin generation predicts early recurrence in breast cancer patients. *J Thromb Haemost* 18(9):2220–2231
181. Pantel K, Alix-Panabières C (2019) Liquid biopsy and minimal residual disease - latest advances and implications for cure. *Nat Rev Clin Oncol* 16(7):409–424
182. Keller L, Pantel K (2019) Unravelling tumour heterogeneity by single-cell profiling of circulating tumour cells. *Nat Rev Cancer* 19(10):553–567

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.