

Neoskrjabinolepis nuda n. sp. from shrews on Sakhalin Island, Russia, with a taxonomic review of *Neoskrjabinolepis* Spasskii, 1947 (Cestoda: Cyclophyllidea: Hymenolepididae)

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Abstract *Neoskrjabinolepis* (*Neoskrjabinolepidoides*) *nuda* n. sp. is described from the shrews *Sorex unguiculatus* (type-host), *S. gracillimus*, *S. isodon* and *S. caecutiens* on Sakhalin Island, Russia. The new species is characterised by: rostellar hooks 40–44 µm long and provided with small epiphyseal thickening of the handle; a long (95–100 µm) cirrus consisting of basal region with claw-shaped spines, a parbasal region with thin needle-shaped spines and an unarmed distal region; a cirrus-sac extending well into the median field; and 15–22 eggs per gravid uterus. A review of the species of *Neoskrjabinolepis* Spasskii, 1947 is presented. Currently, this genus includes nine species and is divided in two subgenera on the basis of strobilar development, which is gradual in the subgenus *Neoskrjabinolepis* (four species) and serial in the subgenus *Neoskrjabinolepidoides* Kornienko, Gulyaev & Mel'nikova, 2006

(five species). An amended generic diagnosis and an identification key to *Neoskrjabinolepis* spp. are presented.

Introduction

Neoskrjabinolepis Spasskii, 1947 includes parasites specific to shrews of the genus *Sorex* L. (Insectivora, Soricidae) (Spasskii, 1947, 1954; Gulyaev, 1991; Czaplinski & Vaucher, 1994). For many years, only two species were recognised, *N. schaldybini* Spasskii, 1947 and *N. singularis* (Cholodkowsky, 1912) Spasskii, 1954 (see Spasskii, 1954; Genov, 1984; Czaplinski & Vaucher, 1994). Sometimes, *N. schaldybini* was considered a junior synonym of *N. singularis* (see Kobuley, 1953; Zarnowski, 1955; Kisieleska, 1958; Prokopič, 1956, 1958; Pojmanska, 1957; Rybicka, 1959), but its validity was supported by Spasskii (1954), Vaucher (1971) and Genov (1984). In order to assess the validity of *N. schaldybini* and to examine the variability of the species of *Neoskrjabinolepis*, recent studies have been undertaken on cestodes belonging to this genus from *Sorex* spp. originating from various parts of the Palaearctic Region. These studies have resulted in redescriptions of the two known forms and the description of a further six species (Kornienko et al., 2006, 2007). These taxa were divided into two subgenera, *Neoskrjabinolepis* (*Neoskrjabinolepis*) Spasskii, 1947, with gradual

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strobilar development (type-species *N. schaldybini*), and *Neoskrjabinolepis* (*Neoskrjabinolepidoides*) Kornienko, Gulyaev & Mel'nikova, 2006, with strobila consisting of series of proglottides at the same developmental stage (type-species *N. (Neoskrjabinolepidoides) singularis*) (see Kornienko et al., 2006). Therefore, the current species diversity of *Neoskrjabinolepis* is much greater than was believed a few years ago. However, much taxonomic work remains in order to achieve a comprehensive alphataxonomy of *Neoskrjabinolepis* in view of the recently proposed criteria for distinguishing species (Kornienko et al., 2006, 2007).

Recent examinations of cestodes from shrews on Sakhalin Island revealed the occurrence of yet another new species of *Neoskrjabinolepis*. The aim of the present article is to describe this new species. We also provide a taxonomic review and an identification key to the species of this genus.

Materials and methods

In June, 2005, one of us (VDG) collected cestodes from shrews at three localities on Sakhalin Island. These were at Sokol Biological Station, the suburbs of the town of Poronaysk and the village of Ozerskiy. Four species of *Sorex* were sampled: *S. unguiculatus* Dobson (130 specimens), *S. gracillimus* Thomas (10 specimens), *S. isodon* Turov (5 specimens) and *S. caecutiens* Laxmann (16 specimens).

Host specimens were dissected immediately after their death. Cestodes were isolated, washed and relaxed in water, and then fixed in 70% ethanol. They were stained in Ehrlich's haematoxylin, differentiated in a 3% aqueous solution of ferric ammonium sulphate 12-hydrate, dehydrated in an ethanol series, cleared in clove oil and mounted in Canada balsam. Some specimens were mounted in Berlese's medium to facilitate the examination of the rostellar hooks and copulatory apparatus.

Type-specimens are deposited in the collections of the Natural History Museum, Geneva, Switzerland (MHNG) and the Zoological Museum at the Institute of Systematics and Ecology of Animals, Novosibirsk (ISEZH).

Measurements are given in micrometres except where otherwise stated. Metrical and meristic data are

presented as the range followed by the mean, with the number of the measurements taken (n) in parentheses.

Neoskrjabinolepis (Neoskrjabinolepidoides) nuda n. sp.

Type-host: *Sorex unguiculatus* Dobson (Insectivora: Soricidae).

Other host: *Sorex gracillimus* Thomas, *S. isodon* Turov and *S. caecutiens* Laxmann.

Type-locality: Sokol Biological Station (SBS), Sakhalin Island, Russia.

Other localities: Village of Ozerskiy and the suburbs of the town of Poronaysk, Sakhalin Island, Russia.

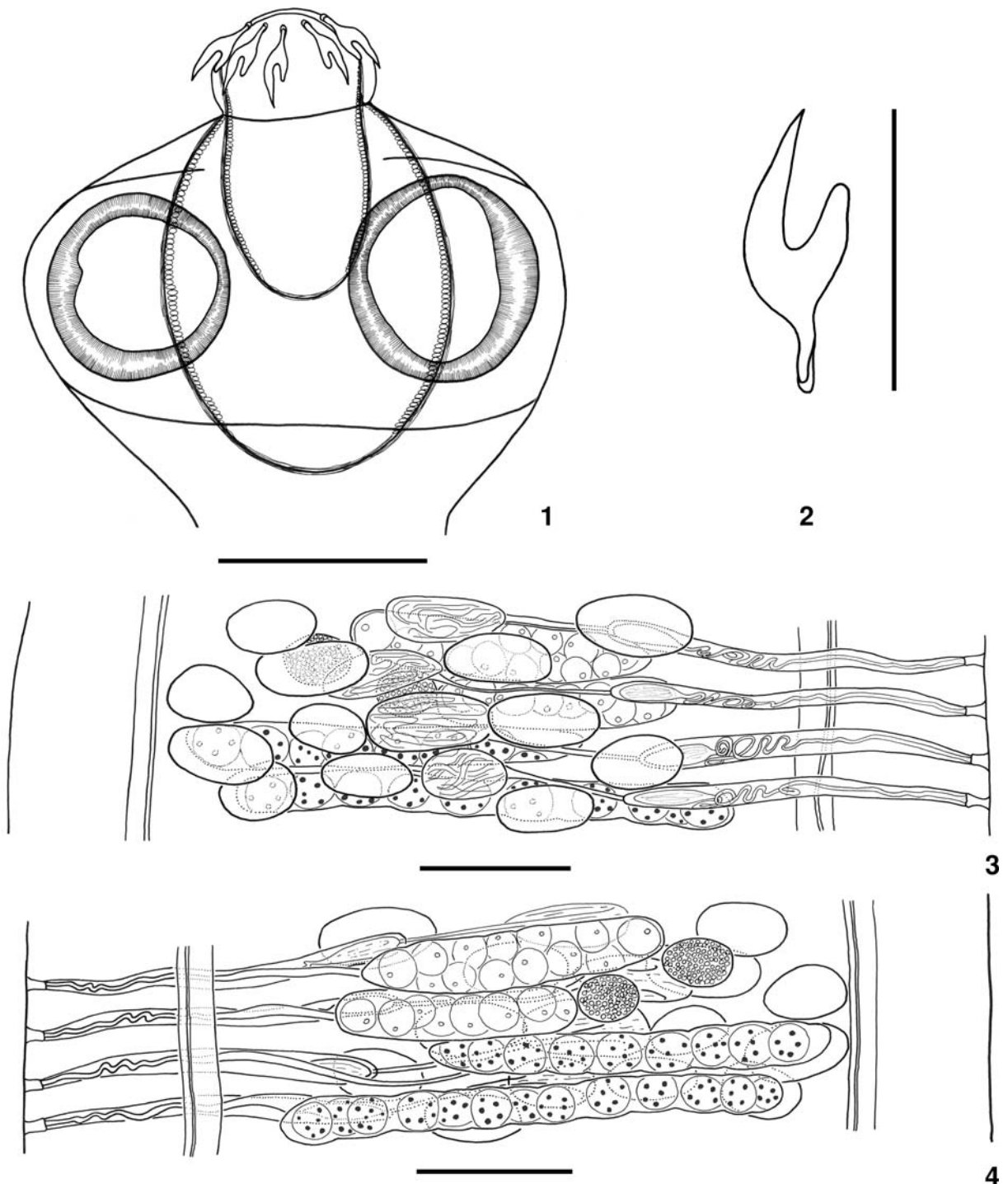
Type-material: Holotype: MHNG INVE 49755, 23.06.2005. Paratypes: MHNG INVE 49756 (2 slides), *S. unguiculatus*, SBS, 28.06.2005; ISEZH No. 1641–1644, *S. unguiculatus*, SBS, 9–14.06.2005; ISEZH No. 1645, *S. gracillimus*, SBS, 16.06.2005; ISEZH No. 1646, *S. caecutiens*, Poronaysk, 08.07.2005; ISEZH No. 1647–1649, *S. unguiculatus*, SBS, 19–20.07.2005, 05.07.2005; ISEZH No. 1650–1652, *S. unguiculatus*, SBS, 01.06.2005, 21.06.2005, 08.09.2005; ISEZH No. 1653, *S. unguiculatus*, Ozerskiy, 9.07.2005; ISEZH No. 1655, *S. isodon*, SBS, 12.06.2005.

Prevalence and intensity: 33.8% and 1–32 (av. 7.5) in *S. unguiculatus*; 20.0% and 1 in *S. gracillimus*; 20.0% and 2 in *S. isodon*, 50.0% and 1–20 (av. 9.8) in *S. caecutiens*.

Etymology: The species name *nuda* (bare) refers to the lack of armament on the distal part of the cirrus, which is a unique character among its congeners.

Description (Figs. 1–7)

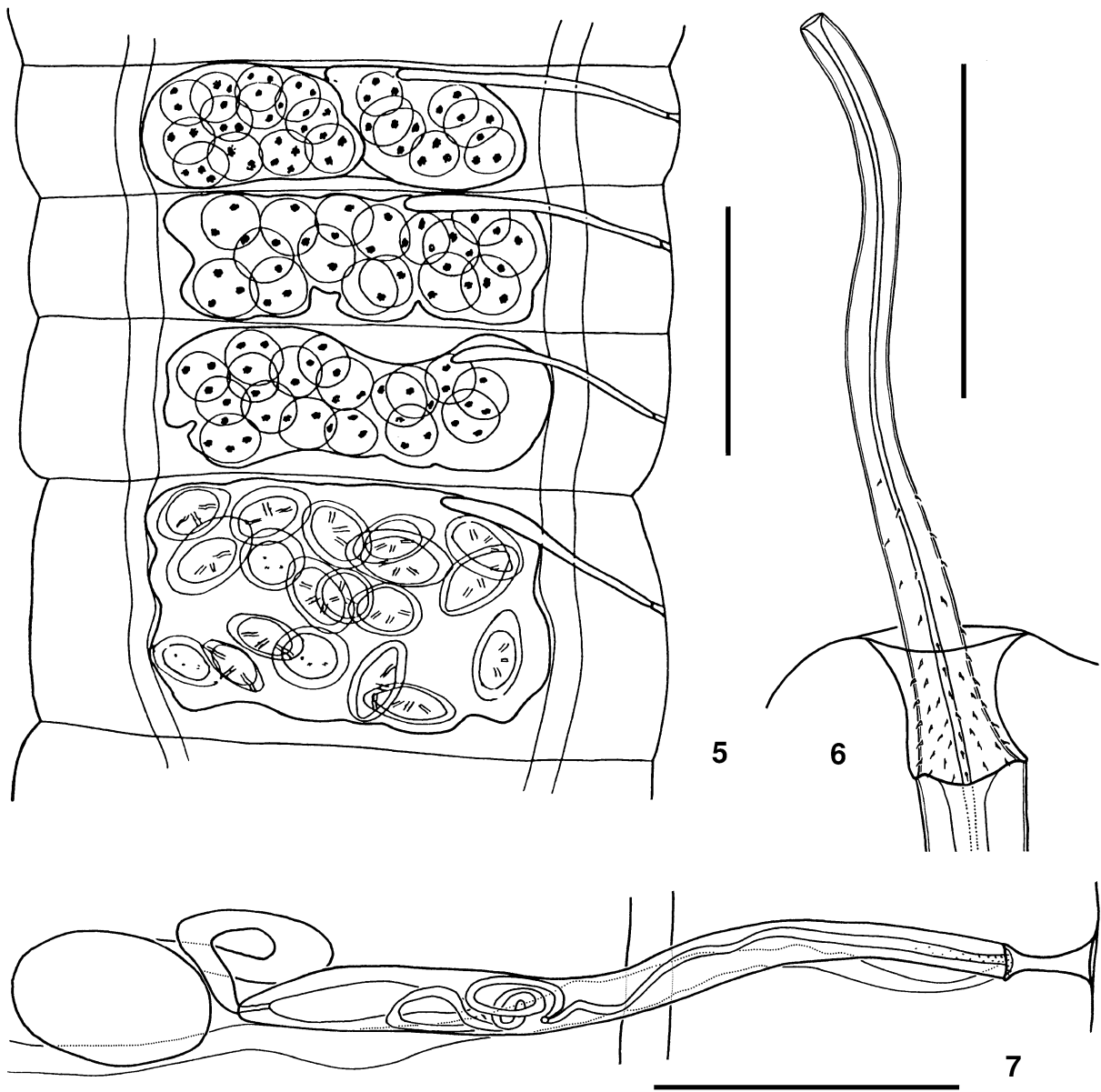
[Based on specimens from the type-host.] Pregravid specimens 5.0–14.0 mm (8.2 mm, n = 10) long, with maximum width at level of postmature proglottides, 280–350 (313, n = 7); strobila flat, consisting of 400–560 (470, n = 7) proglottides. Strobilation serial; pregravid or gravid strobila usually consisting of 4 series of proglottides, each containing proglottides at same developmental stage (first series of juvenile or premature proglottides; second section of hermaphroditic mature proglottides; third series of postmature proglottides; fourth series of pregravid



Figs. 1–4 *Neoskrjabinolepis (Neoskrjabinolepidoides) nuda* n. sp. 1. Scolex. 2. Rostellar hook. 3. Mature proglottides, dorsal view. 4. Transition between the series of mature and postmature proglottides, dorsal (3) and ventral (4) views. *Scale-bars*: 1, 100 μm ; 2, 40 μm ; 3, 4, 50 μm

or gravid proglottides); each series consists of *c.* 100–140 proglottides. Strobilar portions containing juvenile, premature or mature proglottides without

external segmentation; proglottides externally distinct at level of postmature part of strobila. Scolex 250–270 wide (260, $n = 7$), clearly wider than neck



Figs 5–7 *Neoskrjabinolepis (Neoskrjabinolepidoides) nuda* n. sp. 5. Pregravid and gravid proglottides. 6. Cirrus. 7. Terminal genital ducts. Scale-bars: 5, 100 μ m; 6, 20 μ m; 7, 50 μ m

(Fig. 1). Suckers round, 90–100 \times 88–91 (95 \times 90, $n = 7$), with well-developed musculature. Rostellar apparatus complex. Rhynchus short, 60–65 long, 80–85 wide. Rostellum sac-like, 100–140 \times 70–75 (121 \times 72, $n = 7$); its walls consist of external layer of longitudinal muscular fibres and internal layer of circular muscular bundles. Rostellar hooks 10 in number, arranged in single row, 40–44 (42, $n = 10$) long, with characteristic pincers-like shape: axis of

blade almost parallel to axis of guard; blade twice length of handle (Fig. 2); handle provided with small epiphyseal thickening. When rostellar apparatus retracted, rostellar hooks with blades directed anteriorly. Rostellar pouch voluminous, 170–230 \times 140–180 (204 \times 159, $n = 5$), reaches beyond level of posterior margins of suckers; its wall consists of longitudinal muscular fibres and circular muscular bundles. Neck 110–150 (130, $n = 5$) wide.

Proglottides acraspedote, transversely elongate. Mature proglottides 12–15 × 260–330 (14 × 296, $n = 6$) (Fig. 3), with length/width ratio 1:20–22; lateral fields 39–60 wide; gonads densely situated in median field. Postmature proglottides 17–26 × 280–350 (22 × 313, $n = 7$) (length of proglottides measured as distance between genital atria of adjacent proglottides). Pregravid proglottides 95–110 × 280–300 (102 × 290, $n = 5$), with length/width ratio c.1:3. Osmoregulatory canals 2 pairs, without transverse anastomoses; ventral canals 7–10 wide; dorsal canals 3–4 wide. Genital pores unilateral. Genital atrium simple, 2–5 deep, 3–6 in diameter.

Testes 3, of almost equal size, 18–25 × 28–41 (20 × 35, $n = 8$), situated in one row; poral testis separated from two antiporal testes by external seminal vesicle (Fig. 3). Diameter of testes larger than proglottis length and, consequently, dense dorsal testicular field is formed in mature strobilar portion. Degenerating testes persist in postmature proglottides (Fig. 4). Cirrus-sac claviform, 110–120 × 9–10 (113 × 9.4, $n = 5$), slightly winding, passes deeply into median field but does not reach median line of proglottis (Fig. 7). Cirrus long, 95–100 (97, $n = 5$), whip-shaped; its basal region armed with small claw-shaped spines; parabasal region provided with sparsely distributed, thin, needle-shaped spines; distal region of cirrus unarmed (Fig. 6). Ductus ejaculatorius forms several coils. Internal seminal vesicle small, 20–25 × 7–9 (22 × 8, $n = 5$), occupies less than quarter of cirrus-sac length even when filled. External seminal vesicle 28–34 × 15–19 (30 × 17, $n = 5$), connected to cirrus-sac by narrow duct.

Vitellarium subspherical, 13–18 × 18–22 (15 × 20, $n = 5$), situated antiporally to ovary. Ovary transversely elongate, compact, 19–27 × 65–98 (23 × 83, $n = 7$), medial or may be in poral half of median field; in latter case, ovary overlaps cirrus-sac, external seminal vesicle and testes ventrally (Fig. 4); oocytes large. Vagina thin-walled, passes ventally to cirrus-sac.

Uterus not extending into lateral fields of both postmature and gravid proglottides. Uterine wall thin, membranous during all morphogenesis of organ. Number of eggs in uterus 15–22 (Fig. 5). Ripe eggs 49–56 × 32–41; embryophore 29–32 × 17–20; oncosphere 22–26 × 14–15. Embryonic hooks: lateral pairs 10–11 long; medial pair 13–14 long.

Remarks

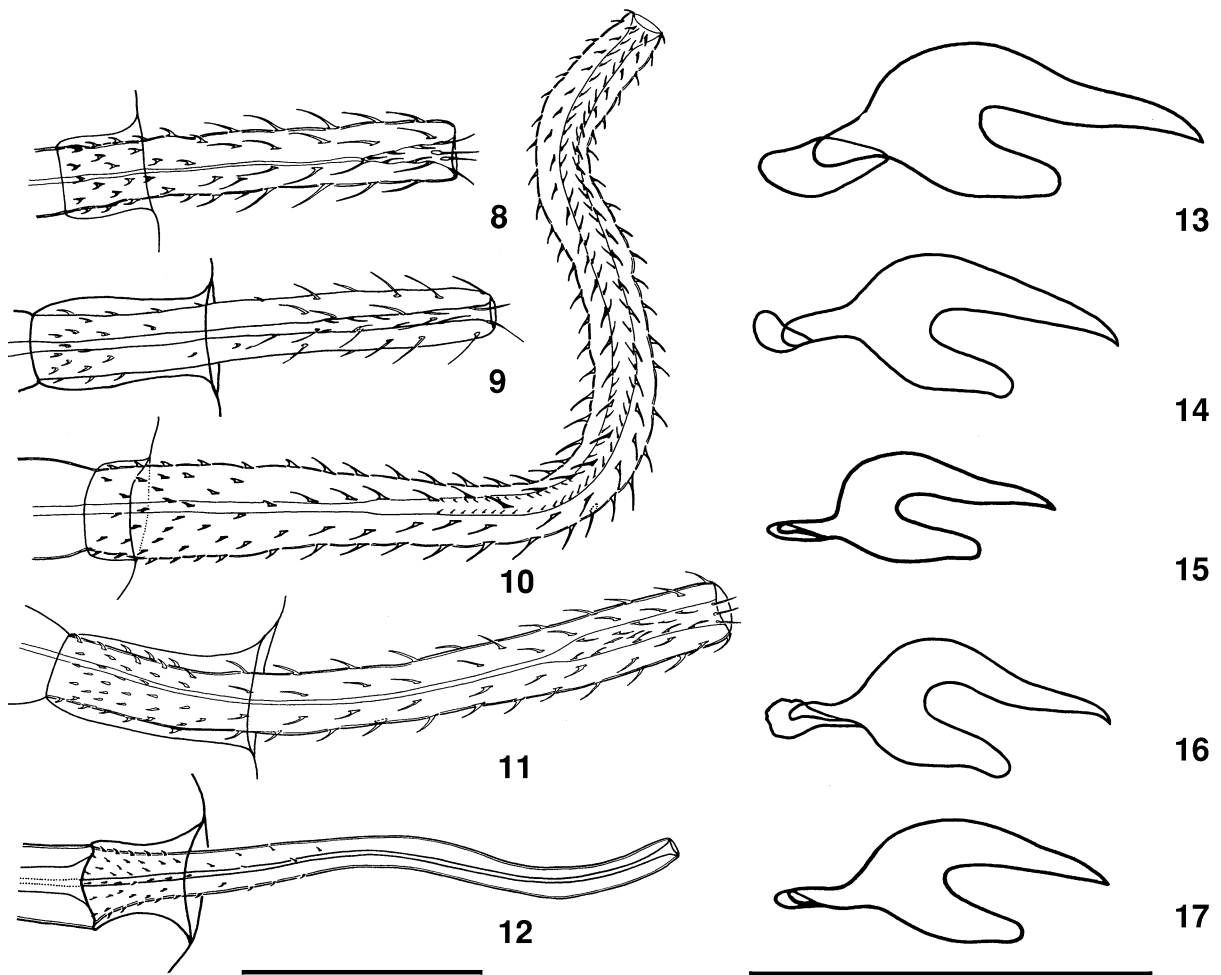
N. nuda n. sp. is characterised by a serial strobilar development. Therefore, it belongs to the subgenus *Neoskrjabinolepis* (*Neoskrjabinolepidoides*) Kornienko, Gulyaev & Mel'nikova, 2006. Currently, this subgenus includes *N. singularis* (Cholodkowsky, 1912), *N. nadtochijae* Kornienko, Gulyaev & Mel'nikova, 2006, *N. corticirrosa* Kornienko, Gulyaev & Mel'nikova, 2007 and *N. kedrovensis* Kornienko, Gulyaev & Mel'nikova, 2007. The new species can be distinguished from all of these on the basis of the characteristics of the rostellar hooks, the structure of the male copulatory apparatus, the armament and size of the cirrus, and the number of eggs per uterus.

N. nuda differs from *N. singularis* by having shorter rostellar hooks (40–44 versus 56–65 μm), the small epiphyseal thickening on the handle and the relatively long handle; in contrast, the rostellar hooks of *N. singularis* possess a short handle and a relatively large epiphyseal thickening (Fig. 13). The new species differs from *N. nadtochijae* by the shape of the blade and the small epiphyseal thickening (Fig. 16). Its rostellar hooks are intermediate in length between *N. corticirrosa* (48–53 μm) and *N. kedrovensis* (36–38 μm).

The new species differs from all the other species of this subgenus in the morphology of the male copulatory apparatus. Compared to *N. corticirrosa* and *N. singularis*, which have short cirrus-sacs (70–90 and 90–95 μm long, respectively) that only just cross the poral osmoregulatory canals, those of *N. nuda* are 100–130 μm long and extend deeply into the median field. In terms of the size of the cirrus-sac, *N. nuda* is similar to *N. nadtochijae* (160–180 μm in length) and *N. kedrovensis* (110–120 μm in length).

The species of this subgenus also differ from each other in their cirral armament (Figs. 8–12), and *N. nuda* is unique in lacking any armament throughout the entire distal region of the evaginated cirrus. Furthermore, the species of this subgenus have differences in the number of the eggs per gravid uterus: 15–22 in *N. nuda*; 10–20 in *N. corticirrosa*; 10–15 in *N. kedrovensis*; 34–43 in *N. singularis*; and 20–46 in *N. nadtochijae*.

On the basis of these differences, we recognise the specimens studied as a species new to science. In



Figs. 8–17 Cirri (8–12) and rostellar hooks (13–17) of *Neoskrjabinolepis* (*Neoskrjabinolepidoides*) spp. (partly after Kornienko et al., 2006, 2007). 8, 13. *N. singularis*. 9, 14. *N. corticirrosa*. 10, 15. *N. kedrovensis*. 11, 16. *N. nadochijae*. 12, 17. *N. nuda* n. sp. Scale-bars: 8–12, 20 μ m; 13–17, 50 μ m

view of the results of this study, the genus is redefined below.

Neoskrjabinolepis Spasskii, 1947

Generic diagnosis [modified after Kornienko et al., 2006]. Cestodes of small body size, consisting of numerous acraspedote proglottides. Anterior portion of strobila (to level of postmature proglottides) without external segmentation. Both gradual and serial patterns of strobilar development occur. Mature proglottides considerably wider than long; gravid proglottides almost as long as wide or longer than wide. Scolex relatively large, provided with complex

rostellar apparatus with invaginable rostellum. Rostellar hooks 10 in number, pincer-shaped, with epiphyseal thickening of handle. Male and female genital systems with simultaneous development. Testes three, arranged in transverse row situated dorsally to female gonads. Cirrus-sac short or long, from just crossing poral osmoregulatory canals to reaching median line of proglottis. Cirrus armed with spines, often of various shapes. Vitellarium compact, rounded, situated in antiporal half of median field. Ovary oval, transversely elongate, poral to vitellarium. Uterus sac-like, situated in median field during its entire development. Gravid proglottis with strong persisting walls, functioning as oöphore and enabling group dispersion of eggs. Parasites of shrews of genus

Sorex (Insectivora: Soricidae) in Palaearctic Region.
Type-species: *N. schaldybini* Spasskii, 1947.

Review of species

Subgenus *Neoskrjabinolepis* (*Neoskrjabinolepis*) Spasskii, 1947

Neoskrjabinolepis schaldybini Spasskii, 1947 (type-species)

Syn. *Hymenolepis schaldybini* (Spasskii, 1947) Vaucher, 1971.

Type-host and locality: “*Sorex* sp.”, vicinities of the town of Sudzhenka (currently Anzhero-Sudzhensk), Kemerovskaya Oblast’, West Siberia, Russia (Spasskii, 1947).

Other hosts: *Sorex araneus* (see Genov, 1984; Kornienko et al., 2006; Binkienė, 2006), *S. minutus* L. (see Kornienko et al., 2006, Genov, 1984; Binkienė, 2006), *S. isodon* (see Kornienko et al., 2006), *S. caecutiens* (see Kornienko et al., 2006), *S. roboratus* Hollister (see Kornienko et al., 2006).

Distribution: West Siberia (Spasskii, 1947; Kornienko et al., 2006), Zabaykalye (Eltyshev, 1975), Bulgaria (Genov, 1984), Lithuania (Binkienė, 2006), Switzerland (Vaucher, 1971), Moldova (Spasskii & Andreyko, 1970), Germany (Vaucher, 1971), United Kingdom (Vaucher, 1971), Finland (Vaucher, 1971; Haukisalmi, 1989), France (Jourdane, 1971; Vaucher, 1971), Netherlands (Vaucher, 1971), Belgium (Vaucher, 1971), Sweden (Vaucher, 1971), Norway (Vaucher, 1971), Poland (Vaucher, 1971), Czech Republic (Vaucher, 1971), Slovakia (Vaucher, 1971; Murai & Mészáros, 1984).

Remarks: Examinations of collections of cestodes of the genus *Neoskrjabinolepis* from West and East Siberia, Bulgaria and Lithuania revealed the presence of *N. schaldybini*. The records from the remaining above-mentioned countries are either accompanied with descriptions not corresponding to the current diagnostic characters of the species or no morphological data have been published. Therefore, the geographical distribution of *N. schaldybini* requires further study.

Sato et al. (1988) reported *N. schaldybini* from *S. unguiculatus*, *S. gracillimus* and *S. caecutiens saevus* Thomas on the island of Hokkaido, Japan. According to their illustrated description, the rostellar hooks are 38–42 µm long. In our opinion, the shape

of these hooks corresponds with that of *N. nuda* n. sp.: the axis of the blade is almost parallel to the axis of the guard and the blade is twice as long as the handle. On this basis, we consider that the material of Sato et al. (1988) belongs to *N. nuda*.

Neoskrjabinolepis longicirrosa Kornienko, Gulyaev & Mel’nikova, 2006 (emend.)

Type-host and locality: *Sorex araneus*, village of Artybash, Altay Mts, Russia.

Other hosts: *Sorex minutus*, *S. isodon*, *S. caecutiens* (see Kornienko et al., 2006).

Distribution: West Siberia (Russia) – Altay Mts, Barabinsk Lowlands, Kuznetskiy Alatau (Kornienko et al., 2006).

Remark: Kornienko et al. (2006) spelled the specific name “*longicirrosus*”. Since the generic name is feminine in gender, this is hereby emended here to “*longicirrosa*”.

Neoskrjabinolepis pilosa Kornienko, Gulyaev & Mel’nikova, 2007 (emend.)

Type-host and locality: *Sorex araneus*, village of Artybash, Altay Mts, Russia.

Other hosts: *Sorex isodon* (see Kornienko et al., 2007).

Distribution: Altay Mts, West Siberia (Kornienko et al., 2007).

Remark: Kornienko et al. (2007) spelled the specific name “*pilosus*”. Since the generic name is feminine in gender, this is hereby emended here to “*pilosa*”.

Neoskrjabinolepis plagis Kornienko, Gulyaev & Mel’nikova, 2007

Type-host and locality: *Sorex caecutiens*, village of Shakhterskiy, Chukotka, Russia.

Distribution: The coast of the Anadyr Gulf, Chukotka (Kornienko et al., 2007).

Subgenus *Neoskrjabinolepis* (*Neoskrjabinolepidoides*) Kornienko, Gulyaev & Mel’nikova, 2006

Neoskrjabinolepis singularis (Cholodkowsky, 1912) Spasskii, 1954 (type-species)

Syn. *Hymenolepis singularis* Cholodkowsky, 1912

Type-host and locality: “*Sorex* sp.”, vicinities of the city of Novgorod, Russia (Cholodkowsky, 1912).

Other hosts: *Sorex minutus* (see Kornienko et al., 2006), *S. araneus* (see Binkienė, 2006; Kornienko et al., 2006), *S. caecutiens* (see Kornienko et al., 2006), *S. isodon* (see Kornienko et al., 2006).

Distribution: West Siberia (Russia) (Kornienko et al., 2006), Lithuania (Binkienė, 2006), Poland (Zarnowski, 1955; Soltys, 1952; Pojmanska, 1957; Rybicka, 1959; Vaucher, 1971), Hungary (Kobuley, 1953), Moldova (Spasskii & Andreyko, 1970), France (Vaucher, 1971), Switzerland (Vaucher, 1971), Netherlands (Vaucher, 1971), Germany (Vaucher, 1971), Denmark (Vaucher, 1971), Norway (Vaucher, 1971), Sweden (Vaucher, 1971), Finland (Vaucher, 1971; Haukisalmi, 1989).

Remarks: We found *N. singularis* in collections from West Siberia, Bulgaria and Lithuania. This species has been recorded from many European countries and from Northern Asia. However, the records have been presented either without morphological data or with data that do not correspond with the current diagnosis of this species. Therefore, its geographical distribution requires further study.

Sato et al. (1988) identified *N. singularis* from *S. unguiculatus*, *S. gracillimus* and *S. caecutiens saevus* on Hokkaido, Japan. However, their description and drawing of rostellar hooks correspond with none of the known species of *Neoskrjabinolepis*. The status of this species (probably new) requires further study.

***Neoskrjabinolepis nadtochijae* Kornienko, Gulyaev & Mel'nikova, 2006**

Type-host and locality: *Sorex isodon*, Bolshekhkhtsirskiy Nature Reserve, Primorye, Russia.

Other hosts: *Sorex caecutiens* (see Kornienko et al., 2006), *S. unguiculatus* (see Kornienko et al., 2006).

Distribution: Kedrovaya Pad Nature Reserve, Lazovsky Nature Reserve (Sikhote-Alin Mts), Primorye; Bolshekhkhtsirskiy Nature Reserve, Primorye, Russia (Kornienko et al., 2006).

***Neoskrjabinolepis kedrovensis* Kornienko, Gulyaev & Mel'nikova, 2007**

Type-host and locality: *Sorex unguiculatus*, Kedrovaya Pad Nature Reserve, Primorye, Russia.

Other hosts: *Sorex isodon* (see Kornienko et al., 2007), *S. caecutiens* (see Kornienko et al., 2007).

Distribution: Primorye, Chernye Gory Mts, Sikhote-Alin Mts, Kamchatka, Russia (Kornienko et al., 2007).

***Neoskrjabinolepis corticirrosa* Kornienko, Gulyaev & Mel'nikova, 2007 (emend.)**

Type-host and locality: *Sorex caecutiens*, village of Shakhtersky, the coast of Anadyr Gulf, Chukotka, Russia.

Other hosts: *Sorex tundrensis* (see Kornienko et al., 2007), *S. portenkoi* Stroganov (see Kornienko et al., 2007).

Distribution: Village of Ezzo, Kamchatka; village of Shakhterskiy on the coast of the Anadyr Gulf, Chukotka; Bolshekhkhtsirskiy Nature Reserve, Primorye, Russia (Kornienko et al., 2007).

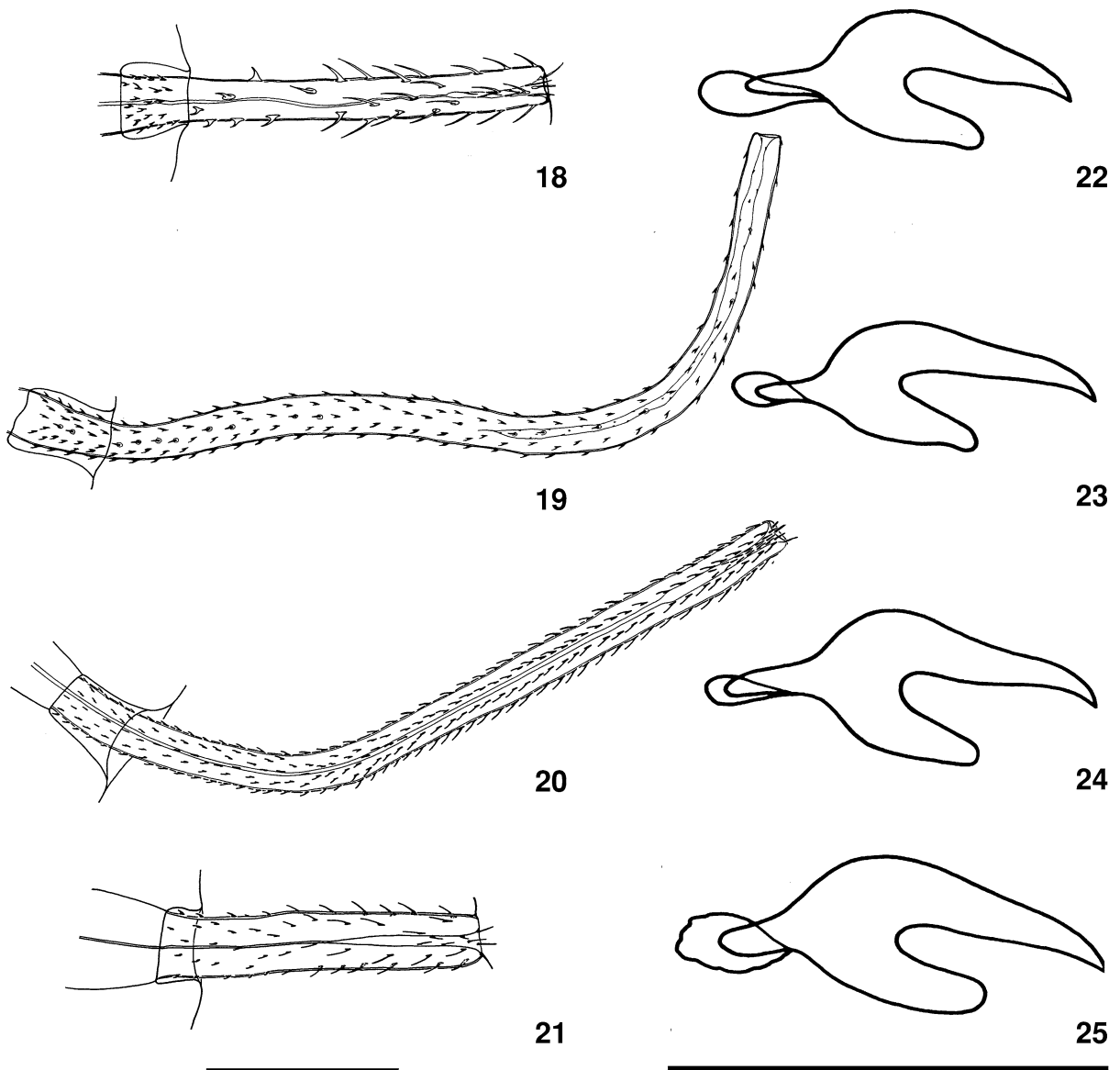
Remark: The original spelling of the specific name was “*corticirrosus*” (see Kornienko et al., 2007). Since the generic name is feminine in gender, this is hereby emended here to “*corticirrosa*”.

***Neoskrjabinolepis nuda* n. sp.**

See above.

Key to the species of *Neoskrjabinolepis*

- 1a. Strobilar development gradual [subgenus *Neoskrjabinolepis* (*Neoskrjabinolepis*)] 2
- 1b. Strobilar development serial [subgenus *Neoskrjabinolepis* (*Neoskrjabinolepidoides*)] ... 5
- 2a. Fully-everted cirrus short, cylindrical 3
- 2b. Fully-everted cirrus long, whip-shaped 4
- 3a. Fully-everted cirrus 40–42 µm long, its parabaasal region armed with several large, claw-shaped spines; middle and distal regions of cirrus armed with sparsely distributed, sabre-shaped spines. Rostellar hooks 38–43 µm long ... *N. schaldybini*
- 3b. Fully-everted cirrus 45–50 µm long, its median region armed with thin, sabre-shaped spines, with size decreasing in distal direction; distal region of cirrus unarmed. Rostellar hooks 52–55 µm long; handle short; epiphyseal thickening large *N. plagis*
- 4a. Fully-everted cirrus 120–125 µm long, armed with small, relatively scarce spines whose size decreases in distal direction, becoming indistinct on distalmost region of cirrus. Rostellar hooks 41–45 µm long; axes of blade and guard form acute angle. Eggs per proglottis 16–20 in number *N. longicirrosa*
- 4b. Fully-everted cirrus 100–110 µm long, armed of small, dense spines along its entire length. Rostellar hooks 45–49 µm long; axes of blade



Figs. 18–25 Cirri (18–21) and rostellar hooks (22–25) of *Neoskrjabinolepis* (*Neoskrjabinolepis*) spp. (partly after Kornienko et al., 2006, 2007). 18, 22. *N. schaldybini*. 19, 23. *N. longicirrosa*. 20, 24. *N. pilosa*. 21, 25. *N. plagis*. Scale-bars: 18–21, 20 μm; 22–25, 50 μm

- | | |
|--|------------------------|
| and guard almost parallel. Eggs per proglottis 35–47 in number | <i>N. pilosa</i> |
| 5a. Fully-everted cirrus short, cylindrical | 6 |
| 5b. Fully-everted cirrus long, whip-shaped | 8 |
| 6a. Handle of rostellar hooks about half length of epiphyseal thickening. Rostellar hooks 56–65 μm long | <i>N. singularis</i> |
| 6b. Handle of rostellar hooks longer than epiphyseal thickening or of comparable length. Rostellar hooks < 56 μm | 7 |
| 7a. Rostellar hooks 40–45 μm long. Cirrus 71–74 μm long. Eggs per proglottis 20–46 in number | <i>N. nadtochijae</i> |
| 7b. Rostellar hooks 48–53 μm long. Cirrus 60–65 μm long. Eggs per proglottis 10–20 in number | <i>N. corticirrosa</i> |
| 8a. Fully-everted cirrus 90–110 μm long; its distal region with scarce, sabre-shaped spines. Rostellar hooks 36–38 μm long | <i>N. kedrovensis</i> |
| 8b. Fully-everted cirrus 95–100 μm long; its | |

distal region smooth. Rostellar hooks 40–44 μm long *N. nuda* n. sp.

Discussion

The validity of the two species, *N. singularis* and *N. schaldybini*, corresponding to the generic diagnosis of *Neoskrjabinolepis* was widely recognised for many years (see Spasskii, 1954; Vaucher, 1971; Genov, 1984). An attempt to verify morphological criteria useful for distinguishing these two species showed that, in Russia at least, each of these names had been used for a complex of species (Kornienko et al., 2006, 2007). The same is probably true for the majority of the records of these two species from European countries and Japan (Soltys, 1952; Kobuley, 1953; Zarnowski, 1955; Pojmanska, 1957; Kisielewska, 1958; Prokopič, 1956, 1958; Rybicka, 1959; Shaldybin, 1964; Spasskii & Andreyko, 1970; Vaucher, 1971; Genov, 1984; Sato et al., 1988; Haukialmi, 1989). Currently, nine species of *Neoskrjabinolepis* are recognised (see Kornienko et al., 2006, 2007; present study), but there are likely still undescribed species belonging to this group.

Traditionally, the species within *Neoskrjabinolepis* were differentiated on the basis of the length of the rostellar hooks and the size of the scolex and the strobila (Spasskii, 1954; Vaucher, 1971). In addition, our study has revealed that the pattern of strobilar development (serial versus gradual), the size and

the armament of the cirrus, the morphology of the rostellar hooks (Figs. 8–25), the position of the cirrus-sac in relation to the poral osmoregulatory canals and the number of eggs per gravid proglottis (Tables 1 and 2) as reliable differentiating characters (Kornienko et al., 2006, 2007; present study).

There are records of four sympatric species of *Neoskrjabinolepis* in the Altay Mountains (Kornienko et al., 2006, 2007), three species in Primorye (Kornienko et al., 2006, 2007) and four species in Bulgaria (Kornienko, unpublished data) and Lithuania (Kornienko & Binkienė, unpublished data). On the basis of the examination of our samples and a reconsideration of the published descriptions, we suggest that in many areas of the Palaearctic Region, between three and five sympatric species of *Neoskrjabinolepis* occur in *Sorex* spp. For example, Kobuley (1953) described as *N. singularis* cestodes from shrews in Hungary, with illustrations better fitting *N. schaldybini*. However, the rostellar hooks were longer (42–46 μm) than those of *N. schaldybini* (38–43 μm), which indicates that his material was heterogeneous. According to Karpenko (1989), the length of the rostellar hooks of *N. schaldybini* ranges between 31 and 53 μm . The proposed synonymy of *N. schaldybini* and *N. singularis* (see Zarnowski, 1955; Prokopič, 1956, 1958; Rybicka 1959) resulted in the loss of useful criteria for distinguishing species within *Neoskrjabinolepis*. For this reason, it is currently difficult to characterise the geographical and

Table 1 Differential characteristics of the species of the subgenus *Neoskrjabinolepis* (*Neoskrjabinolepis*)

	<i>N. schaldybini</i>	<i>N. pilosa</i>	<i>N. longicirrosa</i>	<i>N. plagis</i>
Number of proglottides	>300	c.250	c.450	c.200
Length of rostellar hooks (μm)	38–43	45–49	41–45	52–55
Size of cirrus-sac (μm)	73–90 \times 8–11	120–140 \times 12–19	150–165 \times 12–15	77–93 \times 10–12
Length of cirrus (μm)	40–42	100–110	120–125	45–50
Cirral armament	Parabasal region armed with several large, claw-shaped spines; remainder armed with scarce, sabre-shaped spines	Armed with small, dense spines along entire length	Armed with small, relatively sparsely distributed spines; their size decreasing in distal direction, becoming indistinct on the most distal region	Median region armed with thin, sabre-shaped spines, whose size decreases in distal direction; distal region unarmed
Number of eggs in uterus	20–30	35–47	16–20	25–40

Table 2 Differential characteristics of the species of the subgenus *Neoskrjabinolepis* (*Neoskrjabinolepidoides*)

	<i>N. singularis</i>	<i>N. nadtochijae</i>	<i>N. kedrovensis</i>	<i>N. corticirrosa</i>	<i>N. nuda</i> n. sp.
Number of proglottides	>350	c.200	c.300	c.250	>420
Length of rostellar hooks (μm)	56–65	40–45	36–38	48–53	40–44
Size of cirrus-sac (μm)	70–90 \times 10–12	160–180 \times 11–14	100–130 \times 9–11	90–95 \times 11–15	110–120 \times 9–10
Length of cirrus (μm)	40–47	71–74	90–110	50–55	95–100
Cirral armament	Sparsely distributed, sabre-shaped spines along entire length	Sparsely distributed, sabre-shaped spines along entire length	Distal region with dense, sabre-shaped spines; their size decreasing in distal direction	Parabasal region with several small, claw-shaped spines; remainder with scarce, sabre-shaped spines	Distal region smooth
Vitellarium in relation to ovary	Dorsal	Antiporal	Antiporal	Antiporal	Antiporal
Uterus	Sac-like	Horseshoe-shaped in transverse section	Sac-like	Sac-like	Sac-like
Number of eggs in uterus	34–43	20–46	10–15	10–20	15–22

host ranges of *N. schaldybini* and *N. singularis* on the basis of the published descriptions only.

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References

- Binkienė, R. (2006). Helminth fauna of shrews (*Sorex* spp.) in Lithuania. *Acta Zoologica Lituanica*, 16, 241–245.
- Cholodkowsky, N. (1912). Cestodes nouveaux ou peu connus. Deuxième ser. *Annales Musée Zoologique de l'Académie Impériale des Sciences de St. Petersburg*, 18, 221–232.
- Czaplinski, B., & Vaucher, C. (1994). Family Hymenolepididae Ariola, 1899. In L. F. Khalil, A. Jones, & R. A. Bray (Eds.), *Keys to the cestode parasites of vertebrates* (pp. 595–663). Wallingford, UK: CAB International.
- Eltyshev, Yu. A. (1975). [Helminth fauna in mammals from Barguzinskaya Lowland and an attempt for its geographical analysis. 1. Systematic survey of helminths.] In V. L. Kontrimavichus & A. V. Roytman (Eds.), *Paraziticheskie Organizmy Severo-Vostoka Azii* (pp. 135–167). Vladivostok: Dal'nevostochniy Nauchnyy Tsentr (In Russian).
- Genov, T. (1984). [*Helminths of insectivores and rodents in Bulgaria*.] Sofia: Izdatelstvo na Bulgarskata akademiya na naukite, pp. 348 (In Bulgarian).
- Gulyaev, V. D. (1991). [Morphology and taxonomy of the Ditestolepidini–cestodes (Cyclophyllidea) of shrews with a serial metameric structure of the strobila.] *Zoologicheskij Zhurnal*, 70, 44–53 (In Russian).
- Haukisalmi, V. (1989). Intestinal helminth communities of *Sorex* shrews in Finland. *Annales Zoologici Fennici*, 26, 401–409.
- Jourdane, J. (1971). Helminthes parasites des Micromammifères des Pyrénées-Orientales. II Les Plathelminthes de Soricinae. *Annales de Parasitologie Humaine et Comparée*, 46, 553–574.
- Karpenko, S. V. (1989). [Ecology and morphology of the cestode *Neoskrjabinolepis schaldybini* Spassky, 1947 (Hymenolepididae).] In K. P. Fedorov (Ed.), *Ekologiya gel'mintov pozvonochnykh Sibiri* (pp. 27–44). Novosibirsk: Nauka (Siberian Branch) (In Russian).
- Kisielewska, K. (1958). Cysticercoid of the tapeworm *Neoskrjabinolepis singularis* (Cholodkowsky 1912) Spassky, 1954 in a beetle of the family Catopidae. *Bulletin de l'Académie Polonaise de Sciences (Cl. 2)*, 6, 206–208.
- Kobuley, T. (1953). On the anatomy and systematics of poorly known cestodes from shrews. *Acta Veterinaria Academiae Scientiarum Hungaricae*, 3, 431–438 (In Russian).
- Kornienko, S. A., Gulyaev, V. D., & Mel'nikova, Yu. A. (2006). [On the morphology and systematics of cestodes of the genus *Neoskrjabinolepis* Spassky, 1947 (Cyclophyllidea, Hymenolepididae).] *Zoologicheskij Zhurnal*, 85, 134–145 (In Russian).
- Kornienko, S. A., Gulyaev, V. D., & Mel'nikova, Yu. A. (2007). [New species of cestodes of the genus *Neoskrjabinolepis* Spassky, 1947 (Cyclophyllidea, Hymenolepididae) from shrews of Russia.] *Zoologicheskij Zhurnal*, 86, 259–269 (In Russian).
- Murai, É., & Mészáros, F. (1984). Helminths from small mammals in the Čergov Mountains (Western Carpathians, Czechoslovakia). *Miscellanea Zoologica Hungarica*, 2, 17–28.

- Pojmanska, T. (1957). Pasożyty wewnętrzne (Cestoda, Trematoda) drobnych ssaków polnych z okolic Turwi koło Poznania. *Acta Parasitologica Polonica*, 5, 117–161.
- Prokopič, J. (1956). Helminthofauna rejska obecnego (*Sorex araneus*) v ČSSR. *Československa Parasitologie*, 3, 109–131.
- Prokopič, J. (1958). [On the helminth fauna of the genus *Sorex* in Czechoslovakia.] *Zoologicheskij Zhurnal*, 38(2), 174–183 (In Russian).
- Rybicka, K. (1959). Tapeworms of forest micromammals (Rodentia and Insectivora) from Kampinos Wilderness. *Acta Parasitologica Polonica*, 7, 393–420.
- Sato, H., Kamiya, H., & Ohbayashi, M. (1988). Hymenolepidid and dilepidid cestodes with armed rostellum in shrews, *Sorex* spp., from Hokkaido, Japan. *Japanese Journal of Veterinary Research*, 36, 119–131.
- Shaldybin, L. S. (1964). [Helminth fauna of mammals of Mordovsky State Nature Reserve.] *Uchenye Zapiski Gor'kovskogo Gosudarstvenogo Pedagogicheskogo Instituta, Seriya Zoologicheskaya*, 48, 50–81 (In Russian).
- Soltys, A. (1952). Pasożyty wewnętrzne ryjowki aksamitnej (*Sorex araneus* L.) Białowieskiego Parku Narodowego. *Annales de l'Université Marie Curie-Sklodowska*, 6, 165–209.
- Spasskii, A. A. (1947). [The phenomenon of confluence of proglottides and uteri in cestodes.] *Doklady Akademii Nauk SSSR*, 58, 723–724 (In Russian).
- Spasskii, A. A. (1954). [Classification of hymenolepidids of mammals.] *Trudy Gel'mintologicheskoi Laboratorii*, 7, 120–167 (In Russian).
- Spasskii, A. A., & Andreyko, O. F. (1970). [Cestodes of Insectivora of Moldavia.] *Parazity Zhivotnykh i Rastenii*, 5, 44–59 (In Russian).
- Vaucher, C. (1971). Les cestodes parasites des Soricidae d'Europe. Étude anatomique, révision taxonomique et biologie. *Revue Suisse de Zoologie*, 78, 1–113.
- Zamowski, E. (1955). Robaki pasożytnicze drobnych ssaków leśnych (Rodentia i Insectivora) okolicy Pulaw (woj. Lubelskie). I. Cestoda. *Acta Parasitologica Polonica*, 3, 279–368.