



Chemometric modeling of PET imaging agents for diagnosis of Parkinson's disease: a QSAR approach

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Abstract

Recently, adenosine A_{2A} receptor antagonists have been identified as an interesting drug target for the treatment of Parkinson's disease (PD). Radiolabelled molecular imaging technologies such as positron emission tomography (PET) have emerged in the research field of medicinal chemistry as a diagnostic tool for PD. In the current study, we have performed quantitative structure–activity relationship (QSAR) analysis of 35 xanthine ligand PET tracers as A_{2A} R (adenosine receptors) antagonists in order to determine their structural features required to have binding affinity and selectivity towards A_{2A} R. The division of the dataset into training and test sets was done using a random method, while the feature selection for the binding affinity was done using Genetic Algorithm (GA). The best model with five descriptors was obtained using the spline option in the GA run. QSAR models with four descriptors were also developed for A_{2A} R selectivity, where significant descriptors were selected from the large pool of descriptors using stepwise regression method followed by Best Subset Selection (BSS) method. Furthermore, to improve the quality of the external predictions, we used the “Intelligent Consensus Predictor” tool (http://teqip.jdvu.ac.in/QSAR_Tools/DTCLab/). Both the models showed robustness in terms of statistical parameters. Molecular docking studies have been carried out to understand the molecular interactions between the ligand and receptor, and the results are then correlated with the structural features obtained from the QSAR models. Furthermore, the information derived from the newly found descriptors gives an insight for the development of new candidate PET tracers for the use in PD.

Keywords Parkinson disease (PD) · Positron emission tomography (PET) · QSAR

Introduction

Parkinson's disease (PD) is a progressive neurodegenerative disorder of the central nervous system characterized by muscle rigidity, bradykinesia, and tremor. The disease affects older people, and it is known that 2–3% of the population \geq 65 years of age are more prone towards this disease [1]. It is also associated with loss of dopaminergic neurons in the

substantia nigra, lewy body generation, and abnormal clustering of α -synuclein protein, which is directly connected to expectancy of long life. Hence, effective research for neurodegenerative disease treatment is one of the vital clinical needs of today's life. The current therapy of PD includes restoration of dopamine with levodopa in the striatum of the brain. However, to maintain the therapeutic level, the dosage has to be increased which does not prevent the underlying neuronal loss [2, 3]. On the other hand, such long-term treatment in addition may cause adverse effects which include levodopa-induced dyskinesia and behavioral disturbances in the individuals [4, 5].

Adenosine enzyme inhibitors can be considered an alternative medication in the treatment of PD having less degree of adverse effects. Adenosine is an endogenous modulator of different physiological functions in the peripheral tissues in addition to the central nervous system (CNS). It is a purine nucleoside having four varieties of subtypes consisting of A_1 , A_{2A} , A_{2B} , and A_3 . A_{2A} receptors are highly expressed in striatum (dopamine-rich areas of the CNS) where it is almost co-

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located with dopamine D₂ receptor on GABAergic striatopallidal neurons [6]. A_{2A} antagonistically interferes with the D₂ receptor and as a result decreases the affinity of the D₂ receptor for dopamine upon stimulation and show opposite effect on motor function [7]. Thus, adenosine A_{2A} receptor blockade may show motoric improvement as proven by many animal models [8–11].

Positron emission tomography (PET) [12] and single-photon emission computed tomography (SPECT) [13] are non-invasive methodologies which make use of the dynamic distribution of the radiotracers and provide 3D map of the brain quantifying the biological processes. These imaging agents help in the detection and quantification of dopamine and adenosine receptors in the brain thereby creating a path for early detection of the disease. PET studies are superior to SPECT in terms of accurate results and in determining the temporal measurements of radioactivity with their regional distributions. Agonists and antagonists containing positron-emitting radioisotopes can be introduced in vivo to get 3D image of the receptors which have been helpful in CNS diagnosis. The PET tracers can be used as in vivo-imaging agents in order to improve the pharmacokinetics, physicochemical properties, and mapping of the receptor as per interest. As search of new compounds with desired activity is time-consuming and expensive, pharmaceutical companies have a great interest upon theoretical approaches to design compounds with desired activity.

Quantitative structure–activity relationships (QSAR) have gained a lot of attention in molecular modeling field and are beneficial due to less involvement of human resource and cost-effectiveness [14, 15]. It attempts to develop a correlation between the chemical structures with a well-defined activity. It expresses chemical structures and physiological property in the numerical form and develops a mathematical correlation between them. Furthermore, this relationship can be used to predict the biological response of other existing chemical structures. QSAR-based studies have shown useful applications in drug discovery, molecular modeling, pharmaceutical toxicity modeling, pharmacokinetics/toxicokinetics, data mining, environmental toxicity (ecotoxicity), chemical or drug property modeling, food science, agricultural sciences, pesticide toxicity, fragrance, nanoscience (Nano-QSAR), and many other fields [16–24]. QSAR is also used to predict the absorption, distribution, metabolism, excretion, and toxicological (ADMET) of drug like compounds [25, 26]. QSAR has widespread applications in drug design, medicinal chemistry, and predictive toxicology. It has also become an effective tool in understanding and determining the major biochemical features associated with the Parkinson's disease [27, 28].

In the present study, we have tried to develop QSAR models with PET tracers of xanthine ligands as A_{2A}R (adenosine receptors) antagonist using only 2D descriptors to

explore the structural features required for binding affinity towards A_{2A}R and selectivity of the tracers between A_{2B} and A_{2A} receptors.

Materials and methods

Dataset

The experimental binding affinity and selectivity data of 35 xanthine ligand-based PET tracers were taken from a previously published literature [29] and applied for QSAR modeling to determine the essential structural features needed for binding affinity and explore the structural requirements necessary to be present in the antagonists for selectivity towards A_{2A} adenosine receptors. The experimental values of selectivity and binding affinity (K_i) ranged from 0.1–20 nM and 7.84–16,500 nM respectively, and the details are provided in Supplementary Material I (Table S1). The experimental values were converted into negative logarithm scale during modeling and were used as independent values. No compounds with binding affinity data were removed during modeling but some compounds (**14**, **32**, **33**, and **34**) with no experimental selectivity values were eliminated during modeling. Here, the binding affinity and selectivity were separately used as endpoints or independent variables in modeling. The compounds for both the dataset were represented in the MarvinSketch software [30] with proper aromatization and addition of hydrogen bond as necessary.

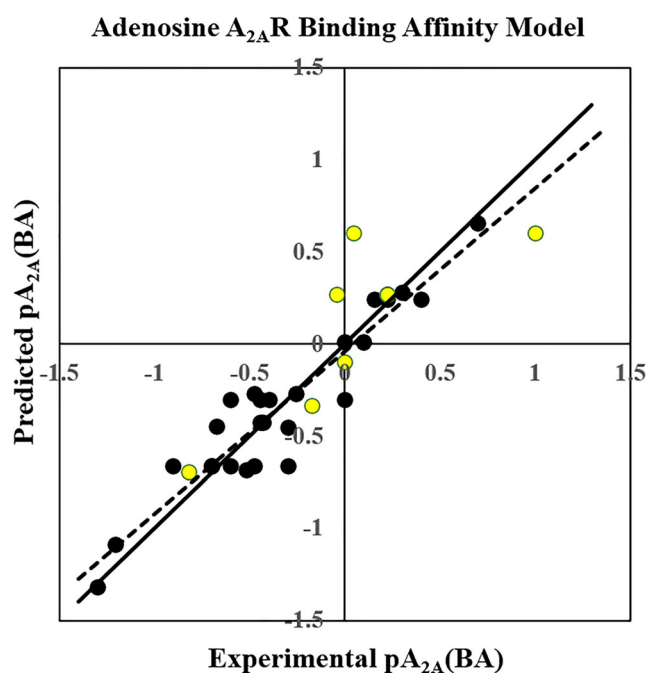


Fig. 1 Observed vs predicted A_{2A}R binding affinity scatter plot

Molecular descriptors

In the present study, QSAR models were developed using a selected class of two-dimensional molecular descriptors involving E-state indices, connectivity, constitutional, functional, 2D atom pairs, ring, atom-centered fragments, molecular property descriptors, and extended topochemical atom (ETA) indices. The ETA descriptors were calculated using the PaDel-Descriptor software [31], whereas the non-ETA descriptors were calculated using the Dragon 7 software [32]. Intercorrelated ($|r| > .95$), constant (variance < 0.0001), and other incompetent and redundant data was removed using an in-house software available at <http://dtclab.webs.com/software-tools> before model development.

Dataset division

Dataset division is a crucial part of QSAR modeling in order to develop a properly validated and robust model. Rational data division ensures an unbiased external validation along with uniform data distribution [33]. The division of the dataset into training set ($\sim 70\%$) and test set ($\sim 30\%$) was performed employing random dataset division method [34] for both binding affinity and selectivity end points. The training set was used for model development, and the test set was used for model validation.

Variable selection and model development

Prior to model development, we have performed variable selection strategies such as Genetic Algorithm (GA) [35, 36] and stepwise regression [35, 37] for binding affinity and selectivity, respectively, to extract the important and influential descriptors and created a reduced pool of descriptors. After obtaining the important descriptors, we went for model development. The best model with five descriptors was obtained using the spline option in the GA run on Discovery Studio version 4.1 for the binding affinity. On the other hand, for A_{2A} R selectivity, four models with four descriptors were selected from the Best Subset Selection (BSS) method based on MAE criteria [38]. Furthermore, to improve the quality of the external prediction via “intelligent” selection of multiple models, we have applied an “Intelligent consensus predictor” tool [39] developed in our laboratory [40].

Statistical validation metrics

The statistical quality of the models developed in the present study was rigorously examined using multiple approaches to check the robustness and predictivity of the developed models. All the models were validated both externally and internally. Various parameters like determination coefficient R^2 , explained variance R^2_a , variance ratio (F), and standard

error of estimate (s) were computed. Internal predictivity parameters such as predicted residual sum of squares (PRESS) and leave-one-out cross-validated correlation coefficient (Q^2_{LOO}) were also calculated along with external predictivity parameters like R^2_{pred} or Q^2_{F1} , Q^2_{F2} , and concordance correlation coefficient (CCC) [41]. It has been reported that consensus models are better in performance in comparison with an individual model [41]. Therefore, we have also performed “Intelligent Consensus Prediction (ICP)” using multiple models to see whether the quality of predictions can be increased through an intelligent selection.

Applicability domain

Applicability domain (AD) [42] is a theoretical region in the chemical space developed based on modeled descriptors and modeled response of the training set, where the developed model could make predictions basing on some logical reliability. Here, we have checked AD using standardization approach using the tool developed in our laboratory [40].

Molecular docking

Molecular docking analysis has been implemented in the present work that helps in understanding the intermolecular interactions taking place between the PET tracer antagonists and the A_{2A} receptor. The protein structure for adenosine A_{2A} receptor is retrieved from the protein data bank with PDB ID:3UZA [43]. The X-ray crystal structure of the protein consists of a bound ligand T4G commonly known as 6-(2,6-dimethylpyridin-4-yl)-5-phenyl-1,2,4-triazin-3-amine (formula: $C_{16}H_{15}N_5$). Before docking the target PET tracers, protein preparation was done by cleaning the protein for any missing residues, explicit hydrogen addition, and generation of the docking site. The generation of active docking site was done in the BIOVIA Discovery Studio platform from the ligand-binding domain of the bound ligand T4G by the selection of the ligand and generating the site “from current selection” program in receptor-ligand interaction module of the software. After the generation of the active ligand-binding domain, the bound ligand was removed for new molecule docking. For ligand preparation, the PET tracers were put through small molecule module in the Discovery Studio platform where a series of ligand conformers were generated. Each of these generated conformers was then used in the CDOCKER module energy for molecular docking involving CHARMM interaction [44]. The CDOCKER interaction energy parameter (kJ/mol) was checked for all the receptor ligand complexes, and the top scoring (most negative, thus favorable to binding) poses were kept.

Results and discussion

Based on the binding affinity and selectivity endpoints of 35 xanthine PET tracer antagonists of adenosine A_{2A} receptor, we have developed one model for the binding affinity ($Q^2 = 0.85$, $R^2 = 0.90$, $Q^2_{F1} = 0.80$) and 4 models ($Q^2 = 0.80$ – 0.87 , $R^2 = 0.87$ – 0.91 , $Q^2_{F1} = 0.84$ – 0.85) for selectivity. All the models were externally and internally validated which showed model robustness and good predictivity in terms of the statistical results. We have also checked the r_m^2 parameters for both internal sets ($\overline{r_{m(loo)}^2}$, $\Delta r_{m(loo)}^2$) and external sets ($r_{m(test)}^2$ and $\Delta r_{m(test)}^2$), and the statistical results were above the critical point justifying the reliability of the models. To improve the quality of the external prediction for selectivity, we also performed “*Intelligent Consensus Prediction*” of the multiple MLR models using the ICP tool [39], and found that the consensus predictions were better than the individual MLR model-derived predictions. The winner model was consensus model 0 (CM0).

Modeling binding affinity of PET tracers towards adenosine (A_{2A}) receptor

The model for binding affinity consists of five descriptors: C-025, F09 [N-O], nBnz, NRS, and nCIR which significantly influence the binding of the antagonists to the adenosine (A_{2A}) receptor. The 5 descriptor MLR

model (Eq. 1) developed using Genetic Function Algorithm (GFA) could predict 85.0% variance of the training set and 80.0% of the test set. The values of all descriptors appearing in the model for training and test set compounds are given in Supplementary Material II (Excel file) and the scatter plot of the observed vs. predicted binding affinity is shown in Fig. 1.

$$pKi(A_{2A}R) = -0.849(\pm 0.2167) \\ -0.36271(\pm 0.06190) C-025 \\ + 0.17693(\pm 0.05895) F09[N-O] \\ -0.52109(\pm 0.07616) NRS \\ + 0.81699(\pm 0.09908) nBnz \\ + 0.3024(\pm 0.03363) nCIR$$

$$n_{\text{training}} = 25, R^2 = 0.901, R^2_{\text{adj}} = 0.875, Q^2 = 0.850, S \\ = 0.170027, F = 34.62, \text{PRESS} \\ = 0.833306, \overline{r_{m(\text{LOO})}^2} = 0.790, \Delta r_{m(\text{LOO})}^2 \\ = 0.072, \text{MAE-based criteria} = \text{Moderate}$$

$$n_{\text{test}} = 10, Q^2_{F1} = 0.80, Q^2_{F2} = 0.681, \overline{r_{m(\text{test})}^2} \\ = 0.54, \Delta r_{m(\text{test})}^2 = 0.23, \text{MAE-based criteria} = \text{Good}$$

Table 1 Definition and contribution of all the descriptors obtained from the MLR models (models developed by using binding affinity)

Sl. no.	Name of descriptors	Descriptor type	Contribution	Discussion	Probable mechanism of binding
1	C-025	Atom-centered fragment descriptor	-ve	C-025 can be depicted as R-CR-R , where ‘R’ can be any group linked to carbon and ‘-’ is any aromatic bond. It is the number of fragments in which a C (sp ²) aromatic atom is bound to three carbon atoms, two of them by an “aromatic bond” and the third by a simple single bond	Flexibility which helps in accommodating the antagonist well in the receptor pocket
2	nBnz	Ring descriptor	+ve	Indicates number of benzene-like rings	π - π Stacking interaction
3	F09 [N-O]	2D atom pair descriptor	+ve	Frequency of N-O fragment at the topological distance 9	Hydrogen bonding
4	NRS	Ring descriptor	-ve	A ring descriptor indicates number of ring systems within a molecule	-
5	nCIR	Ring descriptor	+ve	Number of circuits, i.e., larger loops around two or more rings in a molecule	Hydrophobic interaction/ π - π stacking interaction

Essential features required for binding and receptor interaction

The descriptors obtained in the QSAR model (Table 1) give an insight regarding the mechanism of interaction occurring during binding of the xanthine PET tracer antagonists to adenosine A_{2A} receptor. Unsaturation and aromaticity play a dominating role in regulating the receptor binding affinity which is evident from the occurrence of descriptors such as **C-025**, **nBnz**, **NRS**, and **nCIR**. Descriptors like nBnz and nCIR have positive influences on the adenosine A_{2A} receptor binding (Fig. 2). But on the other hand, descriptors like C-025 and NRS have negative effects on the binding affinity of the PET tracers (Fig. 3). The occurrence of these similar types of descriptors with opposite influence is contradictory and leads to a conclusion that aromaticity provided by benzene nucleus (as seen in compounds like **A-32** and **A-23**) is more important for binding. On the other hand, the presence of heterocyclic aromatic rings and fused-ring systems decrease the overall binding affinity of the radiotracer molecule (found in compounds **A-1**, **A-2**, and **A-20**).

The 2D atom pair descriptor **F09 [N-O]** gives information about the electronegativity of the compounds, and the positive coefficient of the descriptor suggests that higher occurrence of nitrogen and oxygen at topological distance 9 would enhance binding affinity of the compounds as seen in compounds **A-4** and **A-32**. It is found that the presence of electronegative atoms in the compounds or chemical structures can influence the binding to the receptor through hydrogen bonding [45].

Molecular docking Molecular docking helped in understanding the optimized conformation of the complex between the imaging agent and A_{2A} receptor and gave evidences related to the orientation of the imaging agents at the binding zone of the receptor. The major goal was to understand the molecular interactions taking place during radiotracer binding and correlate these findings with QSAR analysis. The docking analysis showed the predominance of different types of π bonding interactions and hydrogen-bonding interactions. In higher active compounds (Fig. 4) like **A-4**, **A-8**, and **A-25** ($pA_{2A}R(BA) = 0.699$, 1.000 , and 0.398 respectively), the interaction forces include mainly hydrogen-bonding interactions (conventional hydrogen bond and carbon-hydrogen bond interaction), π interactions (π -cation, π -donor hydrogen, π - π stacked, π - π T-

shaped, and π -alkyl). Other interactions include halogen and alkyl interaction in compound **A-4** and salt bridge formation in compound **A-8**. Higher number of interacting residues supports the fact that these compounds have higher binding affinity. Compounds having binding affinity in the medium range (Fig. 5) like compound numbers **A-14** and **A-27** ($pA_{2A}R(BA) = -0.301$ and -0.255 respectively) make less number of interactions with the adenosine receptor, but the type of interactions remains similar, i.e., π interactions and hydrogen-bonding interactions. The lowest active compounds (Fig. 5) like compound numbers **A-20** and **A-35** ($pA_{2A}R(BA) = -1.301$ and -1.204 respectively) show the least number of interactions. All the details of binding including interacting residues and type of binding interactions are given in Table 2.

Relationship with QSAR models The docking study shows different types of π interactions occurring between the PET radiotracer molecules and adenosine A_{2A} receptor. This observation supports the occurrence of **nBnz** and **nCIR** descriptors obtained in the QSAR models. The presence of aromatic rings like benzene can enhance binding with the receptor through aromatic π - π stacking interaction with the phenyl/imidazole residue of the receptor [46]. The interaction of these antagonists through π - π stacking interaction eventually blocks the receptor in the indirect pathway thus blocking the activity of GABA-mediated influence in the globus pallidus pars externa (GPe). This helps the PD patients to gain the motor function again by regaining the balance between direct and indirect pathway. Nitrogen and oxygen are capable of hydrogen bond formation and various types of hydrogen bonding as observed in both higher active and lower active compounds, and this can be also correlated to the **F09 [N-O]** descriptor which gives an idea about the electronegativity of the molecule.

Modeling selectivity of PET tracers towards adenosine (A_{2A}) receptor

In the current work, we have developed four MLR models to understand the selectivity of the PET tracer molecules towards adenosine A_{2A} receptor. A single QSAR model may not be efficient enough for the prediction of activity since the property of molecules cannot be understood by a limited number of

Fig. 2 Features increasing the binding affinity (pKi) value

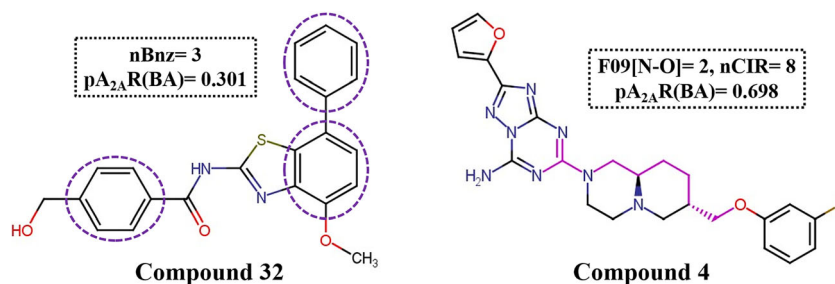
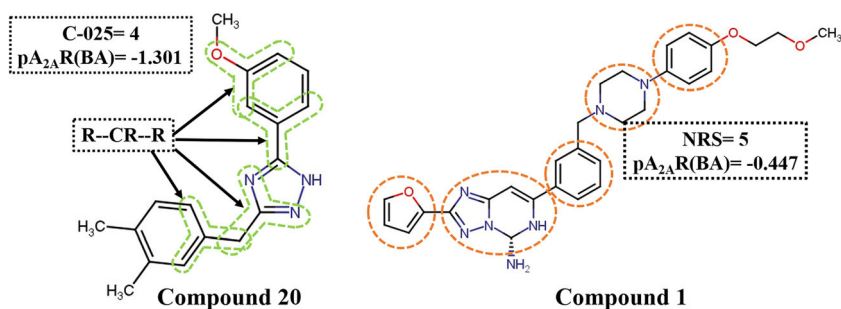


Fig. 3 Features decreasing the binding affinity (pKi) value



features. The use of multiple models for prediction using consensus approach helps in reducing model uncertainty by enhancing the prediction quality of the external set and also in reducing the prediction errors [38]. The four MLR models are given below:

Model 1

$$\begin{aligned} \log A_{2A}R(SeI) &= 0.5875(\pm 0.4130) \\ &+ 0.4643(\pm 0.1574) C-027 \\ &- 0.8679(\pm 0.1797) C-040 \\ &+ 0.7245(\pm 0.1006) F09[N-O] \\ &+ 0.8382(\pm 0.01749) ETA_Beta.s \\ n_{\text{training}} &= 21, R^2 = 0.915, R^2_{\text{adj}} = 0.893, Q^2 = 0.867, S \\ &= 0.234982, F = 42.88, \\ \text{PRESS} &= 1.37546, \overline{r^2}_{m(\text{LOO})} = 0.81227, \Delta r^2_{m(\text{LOO})} \\ &= 0.07373, \text{MAE-based criteria} = \text{Moderate} \end{aligned}$$

$$\begin{aligned} n_{\text{test}} &= 10, Q^2_{F1} = 0.84, Q^2_{F2} = 0.81, \overline{r^2}_{m(\text{test})} \\ &= 0.7682, \Delta r^2_{m(\text{test})} = 0.11949, \text{MAE-based criteria} \\ &= \text{Good} \end{aligned}$$

Model 2

$$\begin{aligned} \log A_{2A}R(SeI) &= 0.36359(\pm 0.43605) - 0.76227(\pm 0.18863) C \\ &- 0.040 - 0.05224(\pm 0.02421) T(F..Cl) \\ &+ 0.71046(\pm 0.11057) F09[N-O] \\ &+ 0.09777(\pm 0.01808) ETA_Beta.s \\ n_{\text{training}} &= 21, R^2 = 0.90, R^2_{\text{adj}} = 0.87, Q^2 = 0.82, S \\ &= 0.274853, F = 35.21, \\ \text{PRESS} &= 1.05627, \overline{r^2}_{m(\text{LOO})} = 0.7526, \Delta r^2_{m(\text{LOO})} \\ &= 0.05874, \text{MAE-based criteria} = \text{Moderate} \end{aligned}$$

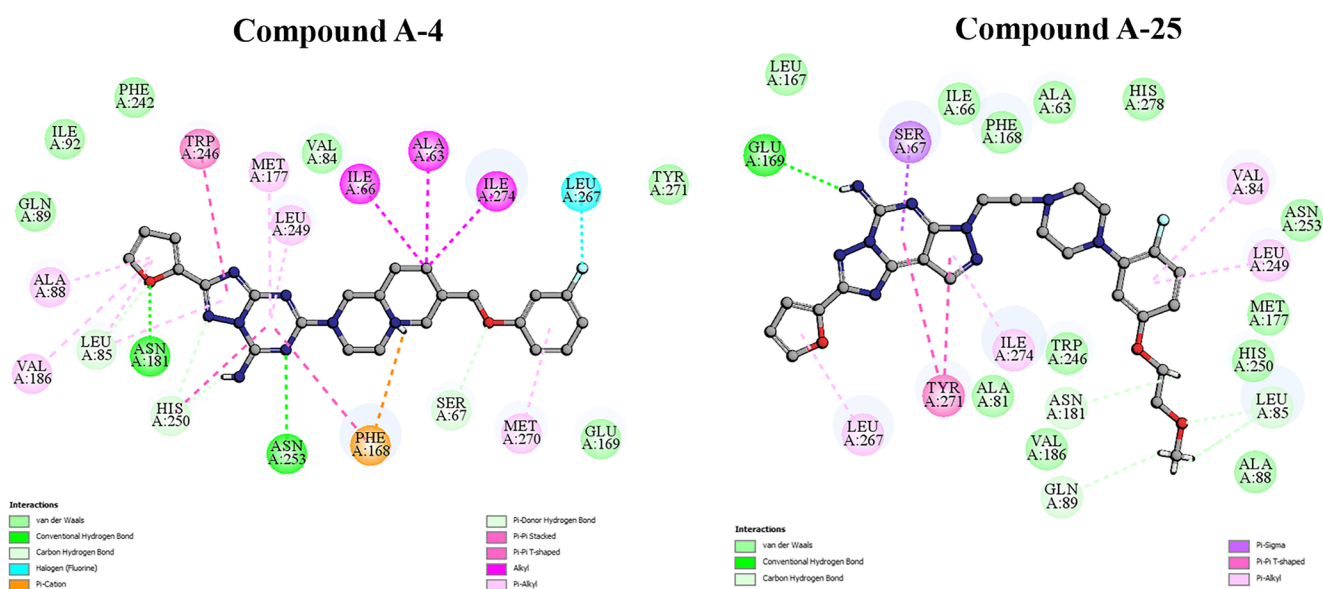


Fig. 4 Docking interactions for compounds having higher binding affinity (pKi)

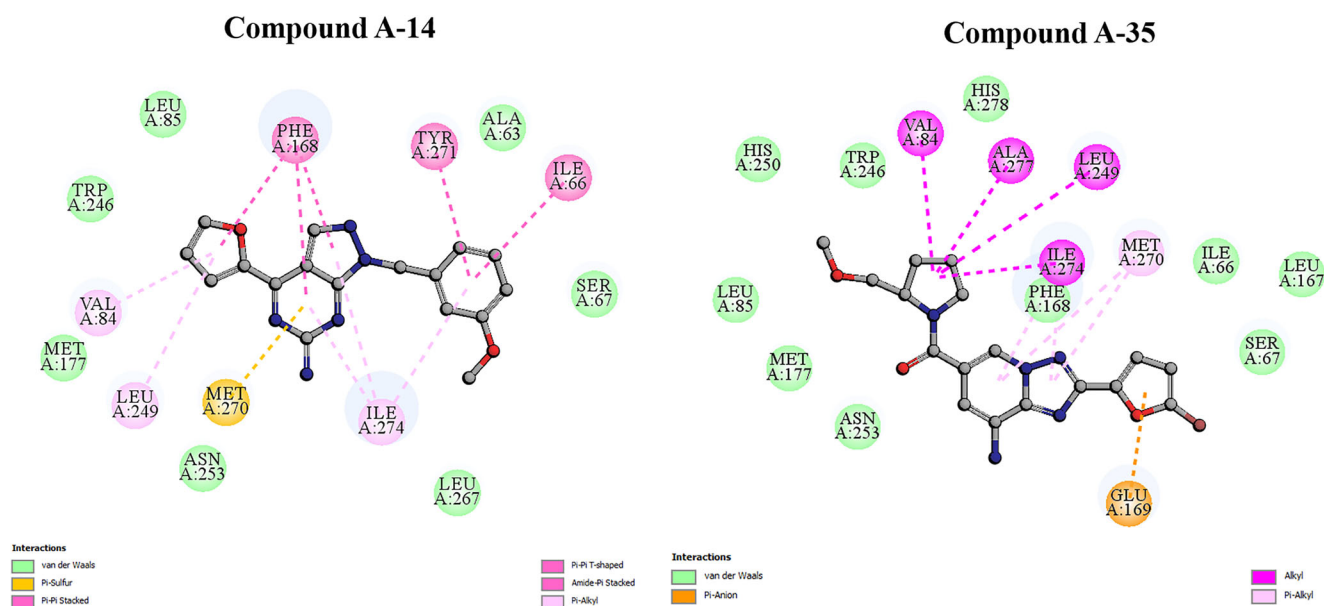


Fig. 5 Docking interactions for compounds having medium (A-14) and low (A-35) binding affinity (pKi)

$$\begin{aligned}
 n_{\text{test}} &= 10, Q_{F1}^2 = 0.84, Q_{F2}^2 = 0.82, \overline{r_{m(\text{test})}^2} \\
 &= 0.7737, \Delta r_{m(\text{test})}^2 = 0.04197, \text{MAE-based criteria} \\
 &= \text{Good}
 \end{aligned}$$

Model 3

$$\begin{aligned}
 \log A_{2A}R(\text{Sel}) &= 0.9642(\pm 0.4535) \\
 &+ 0.31245(\pm 0.08846) \text{ nCIC} \\
 &+ 0.4848(\pm 0.1856) \text{ C-027} \\
 &- 0.9394(\pm 0.2114) \text{ C-040} \\
 &+ 0.6662(\pm 0.1184) \text{ F09[N-O]} \\
 n_{\text{training}} &= 21, R^2 = 0.883, R_{\text{adj}}^2 = 0.854, S = 0.274853, F \\
 &= 30.27, \\
 \text{PRESS} &= 1.72765, Q^2 = 0.833, \overline{r_{m(\text{LOO})}^2} \\
 &= 0.76, \Delta r_{m(\text{LOO})}^2 = 0.12, \text{MAE-based criteria} \\
 &= \text{Moderate}, \\
 n_{\text{test}} &= 10, Q_{F1}^2 = 0.84, Q_{F2}^2 = 0.82, \overline{r_{m(\text{test})}^2} \\
 &= 0.77, \Delta r_{m(\text{test})}^2 = 0.13, \text{MAE-based criteria} \\
 &= \text{Good}
 \end{aligned}$$

Model 4

$$\begin{aligned}
 \log A_{2A}R(\text{Sel}) &= 1.3245(\pm 0.2988) - 0.6702(\pm 0.2119) \text{ C-040} \\
 &+ 0.10445(\pm 0.04427) \text{ SsssN} \\
 &+ 0.05519(\pm 0.01932) \text{ F07[C-C]} \\
 &+ 0.5954(\pm 0.1263) \text{ F09[N-O]} \\
 n_{\text{training}} &= 21, R^2 = 0.872, R_{\text{adj}}^2 = 0.84, S = 0.287861, F \\
 &= 27.24, \\
 \text{PRESS} &= 2.09555, Q^2 = 0.827, \overline{r_{m(\text{LOO})}^2} = 0.717, \Delta r_{m(\text{LOO})}^2 \\
 &= 0.131, \text{MAE-based criteria} = \text{Moderate}, \\
 n_{\text{test}} &= 10, Q_{F1}^2 = 0.85, Q_{F2}^2 = 0.83, \overline{r_{m(\text{test})}^2} = 0.78, \Delta r_{m(\text{test})}^2 \\
 &= 0.07, \text{MAE-based criteria} = \text{Good}
 \end{aligned}$$

The significant descriptors obtained from the four MLR models (M1–M4) contributing to A_{2A} receptor selectivity are C-040, C-027, F09 [N-O], ETA_Beta_s, nCIC, T (F..Cl), SsssN, and F07[C-C]. All the descriptors positively contribute to the A_{2A} receptor selectivity, except C-040, as identified from the regression coefficients of the descriptors and summarized in Table 3. We have also checked the applicability domain of the developed MLR models. The models showed good predictive ability as per the statistical results. The details of the descriptors, their contribution, and frequency of appearance in all the four models are explained elaborately in

Table 2 Details of interacting residues and different types of binding interaction occurring between the PET imaging agents and the target protein (adenosine A_{2A} receptor)

Compound no.	Activity	Binding affinity [pA _{2A} R(BA)]	Interacting residues	Binding interactions
A-4_6	High	0.699	Ala A:88, Val A:186, Leu A:85, Asn A:181, His A:250, Asn A:253, Phe A:168, Ser A:67, Met A:270, Leu A:267, Ile A:274, Ala A:63, Ile A:66, Leu A:249, Met A:177, Trp A:246	Conventional hydrogen bond, carbon hydrogen bond, halogen (fluorine), π -cation, π -donor hydrogen bond, π - π stacked, π - π T-shaped, alkyl, π -alkyl
A-8	High	1.000	Met A:270, Asn A:253, Leu A:249, Phe A:168, Ala A:81, Ile A:66, Glu A:169	Conventional hydrogen bond, carbon hydrogen bond, π - π stacked, π -alkyl, salt bridge
A-25	High	0.398	Leu A:267, Tyr A:271, Ile A:274, Asn A:181, Gln A:89, Leu A:85, Leu A:249, Val A:84, Ser A:67, Glu A:169	Conventional hydrogen bond, carbon hydrogen bond, π -sigma, π - π T-shaped, π -alkyl
A-14	Medium	-0.301	Val A:84, Leu A:249, Met A:270, Ile A:274, Ile A:66, Tyr A:271, Phe A:168	π -sulfur, π - π T-shaped, π - π stacked, amide- π stacked, π -alkyl
A-27	Medium	-0.255	Asn A:253, Ser A:67, Ile A:274, Leu A:167, Glu A:169, Ala A:63, Ile A:66, Leu A:249, Val A:84	Conventional hydrogen bond, carbon hydrogen bond, π -anion, π -alkyl
A-20	Low	-1.301	Val A:84, Leu A:249, Leu A:267, Tyr A:271, Ser A:67, Ile A:274, Asn A:253	Conventional hydrogen bond, π - π T-shaped, π -sigma, π -alkyl, alkyl
A-35	Low	-1.204	Val A:84, Ala A:277, Leu A:249, Ile A:274, Met A:270, Glu A:169	π -alkyl, alkyl, π -anion

Table 3. The values of all descriptors appearing in the models for training and test set compounds are given in the Supplementary Material II (Excel file) and the scatter plots of the observed vs. predicted selectivity values are given in Figure 6.

Mechanistic interpretation All the descriptors obtained in the four models and their frequency give an idea about their importance in modeling the selectivity of the PET tracers towards adenosine A_{2A} receptor. The descriptors like C-027, F09[N-O], SsssN, T(F..Cl), and ETA_Beta_s appearing in the models give information about the electronic feature of the compounds and are essential when the selectivity of receptor is considered (Fig. 7). Electronegativity is a chemical property that describes the tendency of an atom to draw electron towards itself. If a compound contains higher number of electronegative atoms in its structure, then the selectivity of the A_{2A} receptor for that compound also increases.

The presence of atom-centered fragments like C-027 (R-CH-X) in compounds like A-23 and A-25 increase the antagonist selectivity of the PET compounds. Since 'X' represents any electronegative atom like O, N, S, P, Se, and halogens, the presence of heteroatoms increases the selectivity of the compounds towards A_{2A} receptor. The descriptor F09[N-O] explains the frequency of presence of nitrogen and oxygen at the topological distance 9, and its positive regression coefficient indicates its influential activity on the antagonistic behavior of the imaging agents (as seen in compounds A-4 and A-27). Another similar kind of descriptor is T (F..Cl), explaining the information about sum of topological distances

between F and Cl atoms in the chemical structure. These descriptors give information about the electronegative atoms, i.e., nitrogen and oxygen in F09[N-O] and fluorine and chlorine in T(F..Cl). ETA_Beta_s ($\Sigma\beta_s$) is an extended topochemical atom (ETA) descriptor, which can be represented as sum of β_s values of all non-hydrogen vertices divided by 2. The term ' β_s ' can be denoted as

$$\Sigma\beta_s = \Sigma x\sigma$$

Here, x represents contribution of sigma bonds and σ signifies parameters related to sigma bonds. During the computation of β values, the sigma bond value for two similar types of electronegative atoms should be considered 0.5, and dissimilar electronegative atoms should be considered 0.75. This suggests that compounds bearing dissimilar heteroatoms will have greater selectivity to A_{2A} receptor as seen in compounds A-25, A-23, and A-4. Sigma bonds connected with different heteroatoms will have higher descriptor values indicating that the presence of dissimilar heteroatoms is more favorable for selectivity than similar heteroatoms. E-state descriptor SsssN (>N—) encodes the intrinsic electronic state of the nitrogen atom as perturbed by the electronic influence of other molecules with the context of topological character within the molecule. The electronegative contribution of nitrogen is well-depicted in this descriptor, and the positive regression coefficient shows that an increase in the number of tertiary nitrogen benefits in receptor selectivity as seen in compounds A-30 and A-4.

Other descriptors which significantly contribute to A_{2A} receptor selectivity are nCIC, F07[C-C], and C-040. These

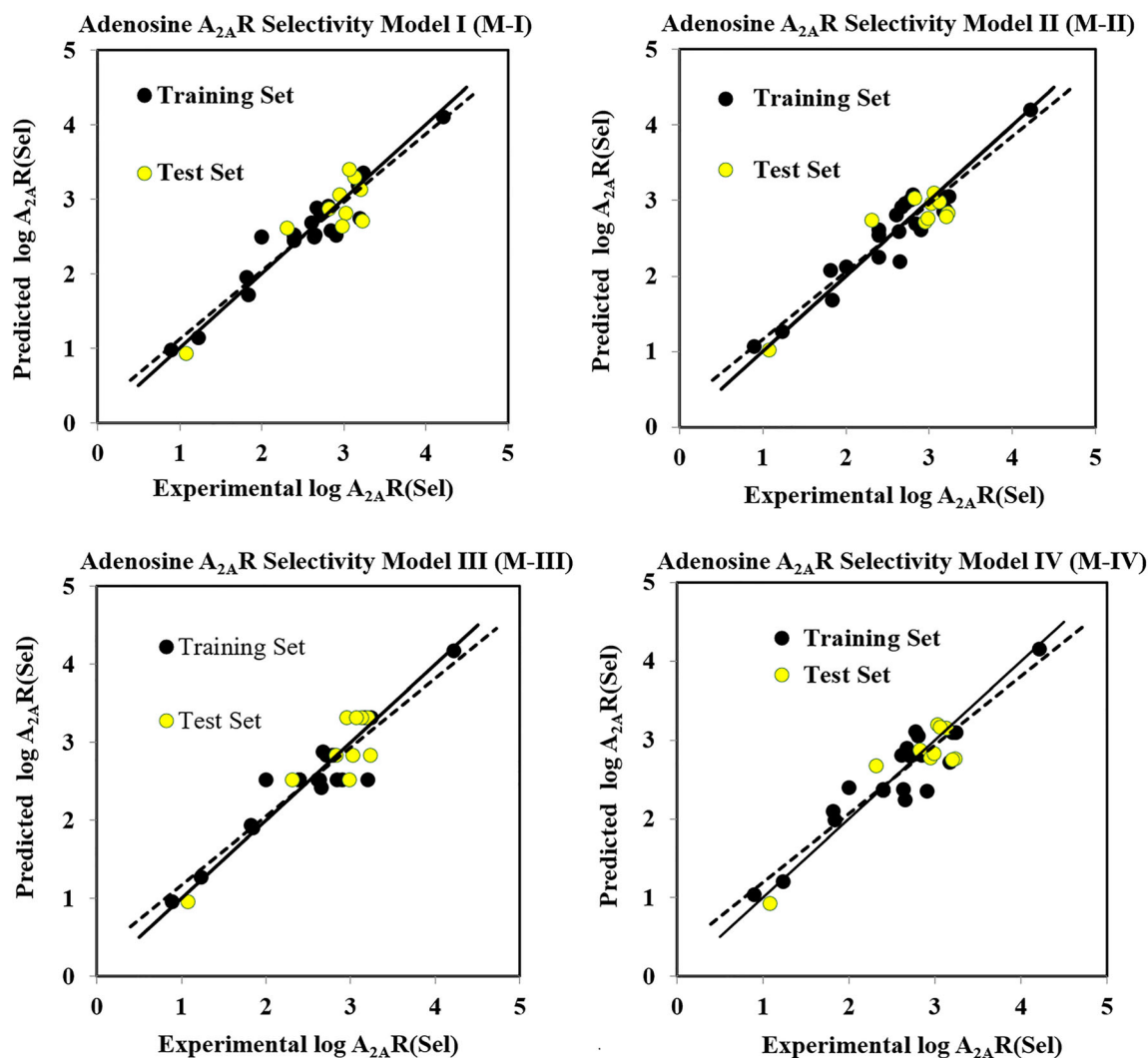


Fig. 6 Observed vs predicted $A_{2A}R$ selectivity plots for all four MLR models

descriptors give information about the number of rings present, type of bonds, and size of the antagonists showing selectivity towards the receptor. The number of rings (cyclomatic number) in the structure is indicated by **nCIC** descriptor. The positive regression coefficient of the descriptor suggests that the presence of high number of rings increases the selectivity towards the A_{2A} receptor as observed in compounds **A-25** and **A-4**. **F07[C-C]**, a 2D atom pair stands for frequency of C–C fragment at the topological distance 7. It provides information about the size (chain length) of the molecule. This means that with an increase in the number of this fragment, i.e., carbon chain, the selectivity towards the A_{2A} receptor increases (as in compounds **A-4** and **A-25**). The atom-centered fragment descriptor, **C-040** (Table 3) gives information about the number of carbon atoms that are attached to heteroatoms by single/double or triple bonds in the straight chain length. The negative regression coefficient suggests that an increase in the number of such fragments decreases the selectivity of the compound towards the A_{2A} receptor as seen in compounds

A-6, **A-7**, and **A-35**. As this fragment suggests high number of double and triple bonds attached with the carbon, it can be concluded that unsaturation in the straight chain of the antagonists is unfavorable for the receptor selectivity.

Intelligent consensus predictions For further refinement of the predictions obtained from the individual models, we have applied intelligent consensus modeling methods. Consensus modeling helps in enhancing the prediction performance of the models and also reduces the test set errors. It was observed that consensus prediction of the test set compounds (Table 4) is better in terms of both MAE-based criteria and predicted R^2 parameter. Four different consensus approaches were used employing “Intelligent Consensus Prediction” tool [39]: CM0 (simple average of predictions), CM1 (average of predictions from the ‘qualified’ individual models), CM2 (weighted average predictions (WAPs) from ‘qualified’ individual models), and CM3 (best selection of predictions (compound-wise) from ‘qualified’ individual models). From

Table 3 Definition, frequency, and contribution of all the descriptors obtained from the MLR models

Sl. no.	Name of descriptors	Type of descriptor	Contribution	Discussion	Frequency of descriptors
1	C-027	Atom-centered fragment	+ve	Counts for certain structural fragment (R--CH--X) in the antagonist, where 'R' can be any group linked to carbon and '--' is any aromatic bond. X can be any electronegative atom (O, N, S, P, Se, halogens)	3
2	ETA_Beta_s	ETA indices	+ve	Sum of all sigma bond contributions considering non-hydrogen vertices divided by 2. The descriptor deals with the presence of dissimilar heteroatoms.	1
3	F09 [N-O]	2D atom pairs	+ve	Frequency of the N-O fragment at the topological distance 9	4
4	SsssN	Atom-type E-state indices	+ve	E-state of ssssN which encodes the intrinsic electronic state of the nitrogen atom as perturbed by the electronic influence of other molecules with the context of topological character within the molecule. SsssN is the atom-type E-state of all tertiary nitrogen in molecules.	1
5	nCIC	Ring descriptors	+ve	Number of rings (cyclomatic number) present in the antagonist	2
6	C-040	Atom-centered fragment	-ve	Represented as R-C(=X)-X/R-C#X/X = C = X fragments where number of carbon atoms are attached to heteroatoms by single/double or triple bonds	4
7	F07[C-C]	2D atom pairs	+ve	Frequency of C-C at topological distance 7	1

the four consensus model obtained, CM0 was found to be the best.

Applicability domain Applicability domain (AD) is an important tool for reliable application of QSAR models. It can be considered a “theoretical region in chemical space defined by the respective model descriptors and responses in which the predictions are reliable” [42, 47]. We have checked the AD of all the models using standardization approach [48] to check whether any molecule in the test set lies outside the AD of a model. From the domain of

applicability analysis, it was found that there were no test set compounds outside the AD, and no compound in the training set came as an outlier (see Supplementary II Excel file).

Comparison with a previously published model A direct comparison between the current and a previously published model [29] is infeasible due to the differences in the composition of training and test sets. However, the current model can be considered more advantageous since it has been developed using simple and easily interpretable two-dimensional descriptors

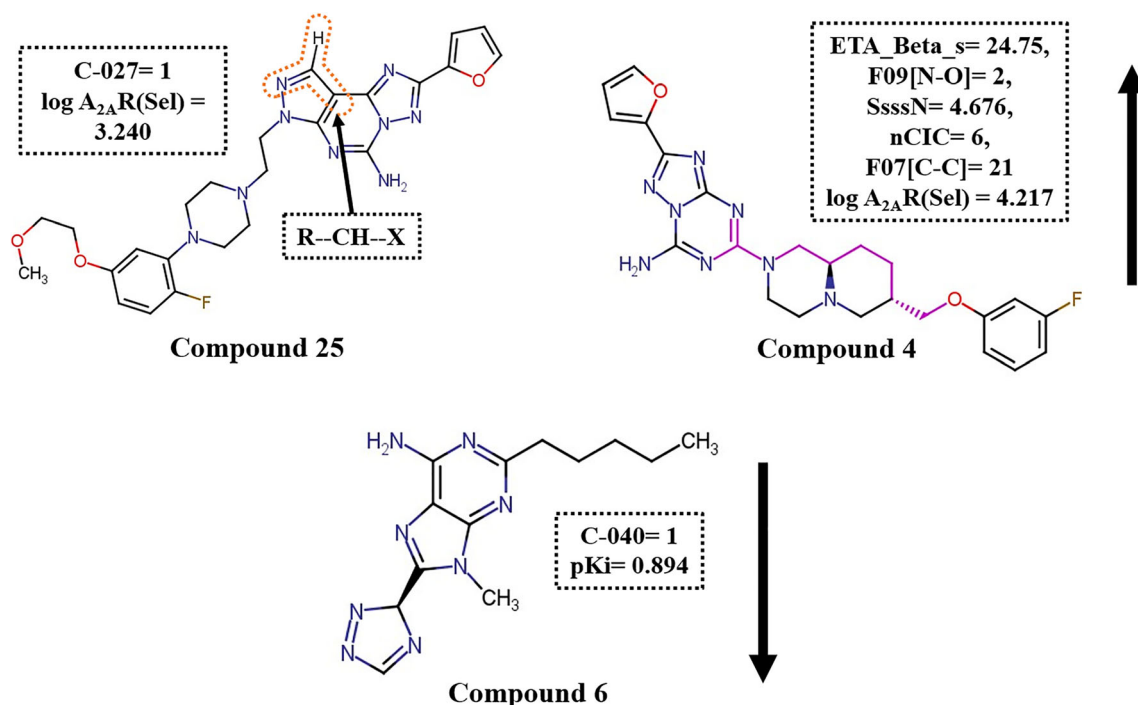
**Fig. 7** Features affecting the adenosine A_{2A} selectivity

Table 4 Detailed summary of the QSAR models and consensus models obtained for selectivity PET tracer compounds for adenosine A_{2A} selectivity (the quality of the best model CM0 is shown in italics)

Dataset	Type of model	Training set statistics					Test set statistics						
		Model R ²	Model Q ² _(LOO)	MAE_train	r ² _{m(LOO)}	Δr ² _{m(LOO)}	R ² _{pred} or Q ² _{F1}	Q ² _{F2}	CCC	r ² _{m(test)}	Δr ² _{m(test)}	MAE (95%)	MAE
Individual models (N1–N5)	IM1	0.92	0.87	Good	0.81	0.07	0.84	0.81	0.77	0.12	0.18	Good	
	IM2	0.90	0.82	Moderate	0.75	0.06	0.84	0.82	0.77	0.04	0.24	Good	
	IM3	0.88	0.83	Moderate	0.76	0.12	0.84	0.82	0.77	0.13	0.18	Good	
	IM4	0.87	0.83	Moderate	0.72	0.13	0.85	0.83	0.78	0.07	0.22	Good	
Consensus models	CM0	-	-	-	-	-	<i>0.88</i>	<i>0.86</i>	<i>0.93</i>	<i>0.82</i>	<i>0.10</i>	<i>0.16</i>	<i>Good</i>
	CM1	-	-	-	-	-	0.86	0.84	0.92	0.82	0.11	0.18	Good
	CM2	-	-	-	-	-	0.85	0.83	0.92	0.82	0.11	0.18	Good
	CM3	-	-	-	-	-	0.83	0.81	0.90	0.80	0.10	0.19	Good

The quality of the best model CM0 is provided in italics

which does not require any conformational analysis or energy minimization before their calculation.

Conclusion

Parkinson's disease is a neurodegenerative disease affecting the elderly person around the world. An important target for its treatment is blocking adenosine A_{2A} receptor which is co-located with the D₂ receptor and is pharmacologically opposite in motor function. Many studies hint that blocking A_{2A} receptor would be a beneficial strategy in the treatment of PD. Thus, this work endeavors exploring QSAR analysis to correlate the chemical structures with their biological activity with the aim to filter the essential chemical features of an antagonist for selectivity and binding affinity to A_{2A} receptor. The computational approach used in this work consists firstly the calculation of the molecular descriptors, and secondly, correlating these descriptors with the binding affinity and selectivity using different chemometric tools such as Genetic Function Algorithm (GFA), Best Subset Selection (BSS) method, and Intelligent consensus predictor (ICP) tools. The statistical quality of the models was checked using traditional metrics both internally and externally. We have also discussed about the contributions of the descriptors in the light of known binding mechanisms such as π-π stacking interaction, hydrophobic interaction, and hydrogen bonding with the different protein residues present in the receptor binding sites. From the insights obtained from such mechanism, we found that electronegative atoms and presence of aromatic ring like benzene are favorable for enhancing the binding affinity to the A_{2A} receptor. Furthermore, the docking studies supported the conclusions derived from the QSAR studies. In conclusion, the study highlights the pharmacophoric features mainly responsible for antagonizing adenosine receptors that can be further

modified for better binding and selectivity to A_{2A} receptor. In case of selectivity also, electronegativity and aromaticity of the compounds play essential and influential roles. The simple two-dimensional (2D) descriptors appearing in all the models are easier to compute requiring no conformation analysis or energy minimization process. Thus, this information would help in the future development and synthesis of newer PET tracer targeted towards adenosine receptor.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

References

- Poewe W, Seppi K, Tanner CM, Halliday GM, Brundin P, Volkman J, Schrag AE, Lang AE (2017) Parkinson disease. Nat Rev Dis Primers 3:1–21
- Voss T, Ravina B (2008) Neuroprotection in Parkinson's disease: myth or reality? Curr Neurol Neurosci Rep 8:304–309
- Ahmed SS, Ahameethunisa A, Santosh W (2010) QSAR and pharmacophore modeling of 4-arylthieno [3, 2-d] pyrimidine derivatives against adenosine receptor of Parkinson's disease. J Theor Comput Chem 9:975–991
- Chen JJ, Swope DM (2007) Pharmacotherapy for Parkinson's disease. Pharmacotherapy: The Journal of Human Pharmacology and Drug Therapy 27:161S–173S
- Jankovic J, Stacy M (2007) Medical management of levodopa-associated motor complications in patients with Parkinson's disease. CNS Drugs 21:677–692

6. Fredholm BB, IJzerman AP, Jacobson KA, Klotz KN, Linden J (2001) International Union of Pharmacology. XXV. Nomenclature and classification of adenosine receptors. *Pharmacol Rev* 53:527–552
7. Fuxe K, Ferré S, Genedani S, Franco R, Agnati LF (2007) Adenosine receptor–dopamine receptor interactions in the basal ganglia and their relevance for brain function. *Physiol Behav* 92: 210–217
8. Chen JF, Xu K, Petzer JP, Staal R, Xu YH, Beilstein M, Sonsalla PK, Castagnoli K, Castagnoli N, Schwarzschild MA (2001) Neuroprotection by caffeine and A2A adenosine receptor inactivation in a model of Parkinson's disease. *J Neurosci* 21:RC143–RC143
9. Grondin R, Bedard PJ, Tahar AH, Gregoire L, Mori A, Kase H (1999) Antiparkinsonian effect of a new selective adenosine A2A receptor antagonist in MPTP-treated monkeys. *Neurology* 52: 1673–1673
10. Ongini E, Monopoli A, Impagnatiello F, Fredduzzi S, Schwarzschild M, Chen JF (2001) Dual actions of A2A adenosine receptor antagonists on motor dysfunction and neurodegenerative processes. *Drug Dev Res* 52:379–386
11. Ikeda K, Kurokawa M, Aoyama S, Kuwana Y (2002) Neuroprotection by adenosine A2A receptor blockade in experimental models of Parkinson's disease. *J Neurochem* 80:262–270
12. Pike VW (2009) PET radiotracers: crossing the blood–brain barrier and surviving metabolism. *Trends Pharmacol Sci* 30:431–440
13. Rahmim A, Zaidi H (2008) PET versus SPECT: strengths, limitations and challenges. *Nucl Med Commun* 29:193–207
14. Roy K (2018) Quantitative structure–activity relationships (QSARs): a few validation methods and software tools developed at the DTC laboratory. *J Indian Chem Soc* 95:1497–1502
15. Gramatica P (2020) Principles of QSAR modeling: comments and suggestions from personal experience. *IJQSPR* 5:1–37. <https://doi.org/10.4018/IJQSPR.20200701.oa1>
16. Puzyn T, Leszczynski J, Cronin MT, eds. (2010) Recent advances in QSAR studies: methods and applications, Vol. 8 Springer Science & Business Media, Berlin, Germany
17. Gao DW, Wang P, Yang L, Peng YZ, Liang H (2002) Study on the screening of molecular structure parameter in QSAR model. *J Environ Sci Heal A* 37:601–609
18. Tropsha A (2004) Application of predictive QSAR models to database mining. *Chemoinformatics Drug Discov* 23:437–455
19. Roy K (2020) Ecotoxicological QSARs. Springer, New York
20. Kar S, Roy K, Leszczynski J (2017) In: Roy K. (eds) On applications of QSARs in food and agricultural sciences: history and critical review of recent developments. *Advances in QSAR Modeling*, Springer, Cham, Switzerland
21. Ojha PK, Roy K (2018) Chemometric modeling of odor threshold property of diverse aroma components of wine. *RSC Adv* 8:4750–4760
22. Tantra R, Oksel C, Puzyn T, Wang J, Robinson KN, Wang XZ, Ma CY, Wilkins T (2015) Nano (Q) SAR: Challenges, pitfalls and perspectives. *Nanotoxicology* 9:636–642
23. Mikolajczyk A, Gajewicz A, Mulkiewicz E, Rasulev B, Marchelek M, Diak M, Hirano S, Zaleska-Medynska A, Puzyn T (2018) Nano-QSAR modeling for ecosafe design of heterogeneous TiO₂-based nano-photocatalysts. *Environ Sci Nano* 5:1150–1160
24. Mikolajczyk A, Sizochenko N, Mulkiewicz E, Malankowska A, Rasulev B, Puzyn T (2019) A chemoinformatics approach for the characterization of hybrid nanomaterials: safer and efficient design perspective. *Nanoscale* 11:11808–11818
25. Hoekman D (1996) Exploring QSAR fundamentals and applications in chemistry and biology, volume 1. hydrophobic, electronic and steric constants, Volume 2. *J. Am. Chem. Soc.* 1995, 117, 9782. *J Am Chem Soc* 118:10678–10678
26. Klein C, Kaiser D, Kopp S, Chiba P, Ecker GF (2002) Similarity based SAR (SIBAR) as tool for early ADME profiling. *J Comput Aided Mol Des* 16:785–793
27. Sebastián-Pérez V, Martínez MJ, Gil C, Campillo NE, Martínez A, Ponzoni I (2019) QSAR Modelling to identify LRRK2 inhibitors for Parkinson's disease. *J Integr Bioinform* 16
28. Khanfar MA, Al-Qtaishat S, Habash M, Taha MO (2016) Discovery of potent adenosine A2a antagonists as potential anti-Parkinson disease agents. Non-linear QSAR analyses integrated with pharmacophore modeling. *Chem Biol Interact* 254:93–101
29. Tamiji Z, Salahinejad M, Niazi A (2018) Molecular modeling of potential PET imaging agents for adenosine receptor in Parkinson's disease. *Struct Chem* 29:467–479
30. MarvinSketch software, <https://www.chemaxon.com>. Accessed on 05 Jan 2020
31. Yap CW (2011) PaDEL-descriptor: an open source software to calculate molecular descriptors and fingerprints. *J Comput Chem* 32:1466–1474
32. Dragon version 7, Kodesrl, Milan, Italy, 2016; software available at <http://www.taletе.mi.it/index.htm>. Accessed 07 Jan 2020
33. Golbraikh A, Shen M, Xiao Z, Xiao Y-D, Lee K-H, Tropsha A (2003) Rational selection of training and test sets for the development of validated QSAR models. *J Comput Aided Mol Des* 17: 241–253
34. Golbraikh A, Tropsha A (2000) Predictive QSAR modeling based on diversity sampling of experimental datasets for the training and test set selection. *Mol Divers* 5:231–243
35. Khan PM, Roy K (2018) Current approaches for choosing feature selection and learning algorithms in quantitative structure–activity relationships (QSAR). *Expert Opin Drug Discovery* 13:1075–1089
36. Devillers J (1996) Genetic algorithms in molecular modeling. Academic Press, Cornwall, Great Britain
37. Pope PT, Webster JT (1972) The use of an F-statistic in stepwise regression procedures. *Technometrics* 14:327–340
38. Roy K, Das RN, Ambure P, Aher RB (2016) Be aware of error measures. Further studies on validation of predictive QSAR models. *Chemom Intell Lab Syst* 152:18–33
39. Roy K, Ambure P, Kar S, Ojha PK (2018) Is it possible to improve the quality of predictions from an “intelligent” use of multiple QSAR/QSPR/QSTR models? *J Chemom* 32:e2992
40. DTC Lab QSAR Tools http://teqip.jdvu.ac.in/QSAR_Tools/DTCLab
41. Roy K, Mitra I (2011) On various metrics used for validation of predictive QSAR models with applications in virtual screening and focused library design. *Comb Chem High Throughput Screen* 14: 450–474
42. Gadaleta D, Mangiatordi GF, Catto M, Carotti A, Nicolotti O (2016) Applicability domain for QSAR models: where theory meets reality. *IJQSPR* 1:45–63
43. Congreve M, Andrews SP, Doré AS, Hollenstein K, Hurrell E, Langmead CJ, Mason JS, Ng IW, Tehan B, Zhukov A, Weir M (2012) Discovery of 1, 2, 4-triazine derivatives as adenosine A2A antagonists using structure based drug design. *J Med Chem* 55: 1898–1903
44. Wu G, Robertson DH, Brooks Iii CL, Vieth M (2003) Detailed analysis of grid-based molecular docking: a case study of CDOCKER—a CHARMM-based MD docking algorithm. *J Comput Chem* 24:1549–1562
45. Pan AC, Borhani DW, Dror RO, Shaw DE (2013) Molecular determinants of drug–receptor binding kinetics. *Drug Discov Today* 18:667–673
46. Jaakola VP, Griffith MT, Hanson MA, Cherezov V, Chien EY, Lane JR, Ijzerman AP, Stevens RC (2008) The 2.6 angstrom crystal structure of a human A2A adenosine receptor bound to an antagonist. *Science* 322:1211–1217

47. Yun YH, Wu DM, Li GY, Zhang QY, Yang X, Li QF, Cao DS, Xu QS (2017) A strategy on the definition of applicability domain of model based on population analysis. *Chemom Intell Lab Syst* 170: 77–83
48. Roy K, Kar S, Ambure P (2015) On a simple approach for determining applicability domain of QSAR models. *Chemom Intell Lab Syst* 145:22–29

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