Cyclothiomethylation of carboxylic acid hydrazides with aldehydes and H₂S

V. R. Akhmetova,^{a*} R. R. Khairullina,^a T. V. Tyumkina,^a Yu. V. Nelyubina,^b A. F. Smol 'yakov,^b I. S. Bushmarinov,^b Z. A. Starikova,^b M. F. Abdullin,^c and R. V. Kunakova^d

^aInstitute of Petrochemistry and Catalysis, Russian Academy of Sciences, 141 prosp. Oktyabrya, 450075 Ufa, Russian Federation. Fax: +7 (347 2) 84 2750. E-mail: ink@anrb.ru
^bA. N. Nesmeyanov Institute of Organoelement Compounds, Russian Academy of Sciences, 28 ul. Vavilova, 119991 Moscow, Russian Federation. Fax: +7 (495) 135 9271. E-mail: star@xray.ineos.ac.ru
^cInstitute of Organic Chemistry, Ufa Research Center of the Russian Academy of Sciences, 71 prosp. Oktyabrya, 450054 Ufa, Russian Federation. Fax: +7 (347 2) 35 6066. E-mail: elmolek@anrb.ru ^dUfa State Academy of Economy and Service, 145 ul. Chernyshevskogo, 450077 Ufa, Russian Federation. Fax: +7 (347 2) 52 0806. E-mail: nis utis@bashnet.ru

The cyclothiomethylation of carboxylic acid hydrazides RCONHNH₂ ($R = C_5H_4N$, Ph, 2-MeOC₆H₄, or 4-HOC₆H₄CH₂) with formaldehyde and H₂S at 70 °C affords predominantly the corresponding *N*-(1,3,5-dithiazinan-5-yl)amides, whereas this reaction at 0—50 °C gives a mixture of the latter compounds with 3-acyl-1,3,4-thiadiazolidines. *N*-(1,3,5-Dithiazinan-5-yl)amides were selectively synthesized by the reaction of carboxylic acid hydrazides with formal-dehyde and H₂S in the presence of BuONa in BuOH.

Key words: cyclothiomethylation, carboxylic acid hydrazides, 1,3,5-dithiazinanes, 1,3,4-thiadiazolidines, formaldehyde, acetaldehyde, hydrogen sulfide, X-ray diffraction study.

Cyclic acid amides and hydrazides, including cyclic peptides, have found use as antibiotics.^{1,2} Sulfur-containing cyclic amides exhibit also antituberculosis, radio-protective, antidepressant, diuretic,³ and antibacterial properties.²

1,3,4-Thiadiazolidine derivatives have antifungal activity against *Candida spp.*,⁴ and substituted 1,3,5-dithiazinanes are effective sorbents for platinum, gold, and silver.⁵

Previously,^{6,7} we have described the selective synthesis of 5-acyl-1,3,5-dithiazinanes by the cyclothiomethylation of carboxamides with CH_2O and H_2S in aqueous butanol in the presence of BuONa. By contrast, the cyclothiomethylation of *p*-toluenesulfonic acid hydrazide with CH_2O and H_2S affords a mixture of 1,3,4-thiadiazolidine, 1,3,5-dithiazinane, and 1,5,3,7-dithiadiazaoctane regardless of the reaction conditions used (pH, the temperature mode).⁸ Before we started our research, carboxylic acid hydrazides have not been subjected to cyclothiomethylation with aldehydes and H_2S .

In the present study, with the aim of performing the selective synthesis of *N*-acyl-1,3,4-thiadiazolidines and

N-(1,3,5-dithiazinan-5-yl)amides, we investigated the condensation of carboxylic acid hydrazides, *viz.*, isonicotinic acid hydrazide (1a), benzhydrazide (1b), *o*-methoxybenzhydrazide (1c), and *p*-hydroxybenzhydrazide (1d), with the CH₂O-H₂S thiomethylating mixture(see Ref. 9) under different conditions, and also investigated the cyclothioalkylation with acetaldehyde and propionaldehyde for isonicotinic acid hydrazide 1a.

Results and Discussion

By analogy with sulfonic acid hydrazides,⁸ carboxylic acid hydrazides would be expected to undergo heterocyclization with CH₂O and H₂S at the β -nitrogen atom, as well as at both the α - and β -nitrogen atoms of hydrazides. Actually, the cyclothiomethylation of isonicotinic acid hydrazide (**1a**) affords a mixture of 4-(isonicotinoyl)-1,3,4-thiadiazolidine (**2a**) and *N*-(1,3,5-dithiazinan-5yl)isonicotinamide (**3a**) (Scheme 1, Table 1). It should be noted that at 0, -10, and -50 °C, the yield of 3-acyl-1,3,4-thiadiazolidine **2a** increases, whereas at 70 °C the reaction produces predominantly *N*-(1,3,5-dithiazin-

Published in Russian in Izvestiya Akademii Nauk. Seriya Khimicheskaya, No. 2, pp. 416-424, February, 2010.

1066-5285/10/5902-0425 © 2010 Springer Science+Business Media, Inc.



Scheme 1

 $R = C_5 H_4 N(a)$, Ph (b), 2-MeOC₆H₄ (c), 4-HOC₆H₄CH₂ (d)

anyl)amide **3a**. However, the cyclothiomethylation of benzhydrazide (**1b**) in the temperature range of 0-70 °C gives N-(1,3,5-dithiazinan-5-yl)benzamide (**3b**) as the major product, whereas the cyclothiomethylation of *o*-methoxybenzhydrazide (**1c**) and *p*-hydroxybenzhydrazide (**1d**) yields mixtures of the corresponding 3-acyl-1,3,4-thiadiazolidines **2c,d** and N-(1,3,5-dithiazinanyl)amides **3c,d** (see Scheme 1 and Table 1). A change in the order of mixing of the reagents (the reaction of **1a** with CH₂O) results in the formation of imine **4** (90%), and the subsequent bubbling of H₂S (20 °C) through a mixture of imine **4** and CH₂O affords a mixture of compounds **2a** and **3a** in a ratio of 2 : 3.

It should be noted that, in some cases, this reaction under reflux conditions in ethanol (70-80 °C) occurs with a higher selectivity. Thus, isonicotinic acid hydrazide (1a) and benzhydrazide (1b) were selectively transformed into the corresponding dithiazinanes 3a,b (see Table 1).

Table 1. Influence of the temperature and the nature of the reaction medium on the yield and the composition of the cyclothiomethylation products of acid hydrazides 1a-d with CH₂O and H₂S

| Hydr- | <i>T</i> /°C | Medium | Yield (%) | | |
|-------|--------------|-----------------------|-----------|----|--|
| azide | | | 2 | 3 | |
| 1a | -50 | H ₂ O | 56 | 22 | |
| | -10 | H ₂ O | 53 | 23 | |
| | 0 | H ₂ O | 44 | 35 | |
| | 70 | H ₂ O | 22 | 58 | |
| | 70 | EtOH-H ₂ O | _ | 62 | |
| 1b | -10 | H ₂ O | 21 | 46 | |
| | 0 | H ₂ O | 18 | 55 | |
| | 70 | H ₂ O | 16 | 60 | |
| | 70 | EtOH-H ₂ O | _ | 57 | |
| 1c | -10 | H ₂ O | 34 | 47 | |
| | 0 | H ₂ O | 39 | 51 | |
| | 70 | H ₂ O | 18 | 55 | |
| | 70 | EtOH-H ₂ O | 35 | 48 | |
| 1d | -10 | H ₂ O | 36 | 49 | |
| | 0 | H ₂ O | 37 | 51 | |
| | 70 | H ₂ O | 10 | 61 | |
| | 70 | EtOH-H ₂ O | 31 | 47 | |

Previously, the reaction temperature, 10 as well as the nature and the arrangement of the substituents in the aromatic rings of anilines, 11,12 have been found to have a similar effect on the cyclothiomethylation pathway.

Therefore, the heterocyclization of carboxylic acid hydrazides 1a-d with CH₂O and H₂S involves two competitive reactions. One reaction occurs at both N atoms of the hydrazide group and affords 1,3,4-thiadiazolidines 2, whereas another reaction proceeds with the involvement of the NH₂ group of the hydrazide moiety and gives 1,3,5-dithiazinanes 3, the pathway of cyclothiomethylation being dependent on the nature of the solvent and the reaction temperature.

Compounds **2a**—**d** and **3a**—**d** were separated by column chromatography on silica gel.

In the ¹³C NMR spectra of thiadiazolidines **2a,c,d**, the signals for the carbon atoms of the rings are magnetically nonequivalent. The 2D HMBC spectra show cross-peaks corresponding to the C(2)–H(5) (C(5)–H(2)) coupling. The ¹H NMR spectra have, in addition to the signals of the rings and the aromatic substituent, signals for the protons of the secondary amino group of 1,3,4-thiadiazo-lidines (δ 9.33–10.54).

The mass spectra of compounds 2a-d and 3a-d contain the corresponding molecular ion peaks $[M]^+$ and peaks of fragmentation ions produced by the successive elimination of CH₂S and CH₂SCH₂S fragments from $[M]^+$.

The structures of 1,3,5-dithiazinanes 3a-d were established by ¹H and ¹³C NMR spectroscopy and X-ray diffraction. Thus, the signals for the carbon atoms of the methylene fragments S-CH₂-S and S-CH₂-N are observed at δ 30.90-33.57 and 53.30-57.86, respectively. The chemical shifts are similar to those described in the literature¹³ for other 1,3,5-dithiazinane derivatives. Analogously, the ¹H NMR spectra of compounds 3a-d show signals for the methylene groups as broadened singlets due to the rapid heterocyclic ring inversion on the NMR time scale. The crystal structures of compounds 3a-c were studied by X-ray diffraction (Figs 1-5).

In dithiazinane **3a**, the Py–CO–NH fragment is nonplanar. The dihedral angle between the planes of the pyridine ring and CONH is 35.4° . The dithiazinane ring is twisted around the N(1)–N(2) bond by 87.5° , *i.e.*, the



Fig. 1. Molecular structures of N-(1,3,5-dithiazinan-5-yl)isonicotinamide (**3a**) (*a*) and N-(1,3,5-dithiazinan-5-yl)-*o*-methoxybenzamide (**3c**) (*b*) in the crystals (displacement ellipsoids are drawn at the 50% probability level).

aromatic substituent is in the axial position. The ring adopts the classical chair conformation slightly flattened at the N(1) atom (the deviations of the C(2) and N(1) atoms from the mean plane of the ring are 0.90 and 0.64 Å, respectively). The dihedral angles between the bottom of the chair (C(1)–S(1)–S(2)–C(3)) and the N(1)C(1)C(3) and C(2)S(1)S(2) planes are 55.2 and 63.7°, respectively (see Fig. 1). In the crystal structure, the molecules are linked through the N(2)–H(2N)...N(1) (H(2N)...N(1), 2.18 Å) and S(2)...H(3)–C(3)



Fig. 2. Double layers in the crystal packing of 3a.



Fig. 3. Molecular chains in the crystal structure of 3c.

(S(2)...H(3), 2.93 Å) hydrogen bonds to form double layers parallel to the *ac* plane (see Fig. 2).

In molecule **3c**, the Ph–CO–NH fragment is strongly flattened, as opposed to molecule **3a** (the flattening is associated with the formation of the strong intramolecular N(1)–H(1N)...O(2)Me hydrogen bond with the methoxy group (H(1N)...O(2), 2.003 Å)). The similar bond was found in 18 structures of hydrazones containing the *o*-methoxy group¹⁴ (H...N, 1.79–2.02 Å). The dithiazinane ring is twisted around the N(1)–N(2) bond line by 92.6° and adopts the conformation identical to that observed in **3a**. The angles between the bottom of the chair



Fig. 4. Molecular structure of N-(1,3,5-dithiazinan-5-yl)-p-hydroxybenzamide (3d) in the crystal.



Fig. 5. Hydrogen bonds in the crystal structure of 3d.

(C(1)-S(1)-S(2)-C(3)) and the N(2)C(11)C(9) and C(10)S(1)S(2) planes are 56.0 and 62.3°, respectively (the deviations of the C(10) and N(2) atoms from the mean plane are 0.89 and 0.65 Å, respectively) (see Fig. 1).

In the crystal structure, molecules 3c are linked to each other through the weak C(9)—H(9a)...O(1) hydrogen bonds (H(9a)...O(1), 2.36 Å) to form zigzag chains running along the *a* axis (see Fig. 3).

The NH–CO group in molecule **3d**, unlike those in **3a,c**, has a fixed *cis* conformation (see Fig. 4). This is associated with the formation of the strong O(2)–H(2O)...O(1) (H(2O)...O(1), 1.90 Å) and N(1)–H(1N)...S(2) (H(1N)...S(2), 2.52 Å) hydrogen bonds (see Fig. 5). The N(1)N(2)C(4)O(1) group is planar and orthogonal to the dithiazinane ring (the dihedral angle is 88.7°). The conformation of the dithiazinane rings is identical to that in compounds **3a,c**. The angles between the bottom of the chair (C(1)–S(1)–S(2)–C(3)) and the N(1)C(1)C(3) and C(2)S(1)S(2) planes are 55.4 and 63.8°, respectively (the deviations of the C(2) and N(1) atoms from the mean plane are 0.90 and 0.64 Å, respectively).

The bond lengths are given in Table 2. As can be seen from the above-considered data, the molecules have similar geometric characteristics, which are close to the standard values.

We found that the reaction of hydrazide 1a with CH₂O and H₂S in the presence of HCl (-50-20 °C) occurs with the involvement of the NH₂ group of protonated form 1a. As a result, 1,3,5-dithiazinane 3a and linear oligomer 5 were obtained after the neutralization of the reaction mixture (NaOH) (Scheme 2, Table 3). Compounds 3a and 5 were separated by column chromatography.

Scheme 2



The HMBC spectrum of linear product 5 contains cross-peaks between the C(10) (C(10')) atoms and the

Table 2. Selected bond lengths (*d*) and torsion angles (τ) in molecules **3a,c,d** and **8**

| Parameter | 3a | 3c | 3d | 8 | |
|-----------|-----------------------------------|-----------------------------------|-----------------------------------|---|--|
| Bond | d/Å | | | | |
| N—N | 1.410(3) | 1.406(3) | 1.402(2) | 1.389(3), 1.384(4), 1.389(3), | |
| N-C(=0) | 1.351(4) | 1.341(3) | 1.341(3) | 1.397(3) 1.333(4), 1.342(4), 1.326(4), | |
| C=O | 1.220(3) | 1.217(3) | 1.232(3) | 1.331(4) 1.235(3), 1.231(3), 1.241(3), 1.234(4) | |
| C-S* | 1.833(2), 1.829(3) | 1.826(3), 1.824(3) | 1.828(2), 1.827(2) | - | |
| C—S** | 1.829(3) 1.820(3), 1.829(4) | 1.824(3) 1.812(3), 1.800(3) | 1.827(2) 1.812(2), 1.809(2) | _ | |
| Angle | | τ/deg | 5 | | |
| N-N-C-O | 0.0(4) | 2.5(4) | -5.9(2) | -4.2(5), -3.9(4), -1.0(4), -4.1(4) | |

* The bottom of the chair.

** The SCH₂S fragment.

protons of the S–CH₂–S fragment. The mass spectrum of compound 5 shows a molecular ion peak $[M]^-$ at m/z 378 and a characteristic fragmentation pattern corresponding to the successive elimination of the CH₂CH₂NNHCOC₅H₄N (m/z 184) and CH₂NNHCOC₅H₄N (m/z 148) fragments.

Liquid chromatography mass spectrometry (LCMS) of a mixture of reaction products **3a** and **5** also revealed the cyclic oligomer of formaldehyde and CH₂S, *viz.*, 1,3,5,7,9-oxatetrathiecane **6** (~10%).

The selective cyclothiomethylation of hydrazides 1a-d to dithiazinylamides 3a-d was carried out with the use of the CH_2O-H_2S thiomethylating mixture in the presence of BuONa in BuOH (see Table 3) by analogy with the cyclothiome-



thylation of carboxamides.^{6,7} The activation of hydrazide **1a** with sodium metal in refluxing benzene¹⁵ followed by the thiomethylation with the CH_2O-H_2S system affords the mixture **2a** + **3a**.

Alkyl-substituted nitrogen- and sulfur-containing heterocyclic systems were synthesized by the cyclothioalkylation of hydrazide **1a** with acetaldehyde or propionaldehyde and H₂S. Unlike the reactions with CH₂O and H₂S, the reaction of **1a** with these aldehydes under usual conditions (20 °C in ethanol) produces exceptionally Schiff bases 7 and 8 in 72 and 50% yields, respectively (Scheme 3).

Table 3. Influence of the reaction temperature and the pH of the reaction mixture on the yield and the composition of the cyclothioalkylation products of carboxylic acid hydrazides 1a-d

| Hyd | r- <i>T</i> | Alde- hyde | Medium | Reagent | Yield (%) | | | |
|------|-------------|-------------------|-------------|---------|-----------|----|----|----|
| azid | e ∕°C | | | ratio | 2a | 3a | 5 | 9 |
| 1a | 20 | CH ₂ O | HCl | 1:2:1 | _ | 39 | 11 | _ |
| | 20 | $CH_{2}O$ | HCl | 1:2:3 | _ | 56 | 14 | _ |
| | -50 | CH_2O | HCl | 1:2:3 | _ | 40 | 12 | _ |
| | 40 | $CH_{2}O$ | BuONa/BuOH | 1:3:3 | _ | 68 | _ | _ |
| | 40 | MeĈHO | BuONa/BuOH | 1:3:3 | _ | _ | _ | 41 |
| | 20 | CH ₂ O | Na/C_6H_6 | 1:3:1 | 24 | 35 | _ | _ |
| 1b | 40 | CH_2O | BuONa/BuOH | 1:3:3 | _ | 57 | _ | _ |
| 1c | 40 | $CH_{2}O$ | BuONa/BuOH | 1:3:3 | _ | 59 | _ | _ |
| 1d | 40 | CH ₂ O | BuONa/BuOH | 1:3:3 | — | 55 | — | _ |

Imines 7 and 8 were identified by 1 H and 13 C NMR spectroscopy. The structure of compound 8 was established also by X-ray diffraction (Fig. 6).

It should be noted that the Py—CO—NH fragment in molecule **8** is structurally similar to that in dithiazinane **3a**. This fragment is nonplanar; the dihedral angle between the planes of the pyridine ring and CONH is 32.4°.

The reaction of imine **7** with MeCHO and H_2S affords a mixture of the heterocycles *N*-(2,4,6-trimethyl-1,3,5dithiazinan-5-yl)isonicotinamide (**9**) and 1,1-bis[4-(isonicotinoyl-2,5-dimethyl-1,3,4-thiadiazolidin-3-yl)ethylsulfanyl]ethane (**10**) in a ratio of ~5 : 1 both upon refluxing in ethanol and in the presence of BuONa/BuOH (see Scheme 3).

The liquid chromatography mass spectra show molecular ion peaks $[M - H]^-$ at m/z 283 and 591 for compounds 9 and 10, respectively, and fragmentation ion peaks. For molecule 10, the characteristic elimina-



Fig. 6. Molecular structure of N'-(propylidene)isonicotinic acid hydrazide (8) (displacement ellipsoids are drawn at the 50% probability level).

tion of two CHCH₃SCHCH₃ fragments from $[M - H]^-$ (*m*/*z* 413) and the successive fragments C₅H₄NCONCH-CH₃SCHCH₃NCHCH₃SCHCH₃S (*m*/*z* 281) and C₅H₄NCONCHCH₃SCHCH₃N (*m*/*z* 223) were observed.

The stereoselective synthesis of (*cis*,*cis*-2,4,6-trimethyl-1,3,5-dithiazinan-5-yl)isonicotinamide (**9**) was carried out in the presence of BuONa/BuOH (pH 11.25) at 40 °C (see Scheme 3 and Table 3).

The ¹³C and ¹H NMR spectra of compound **9** contain one set of signals. A comparison of the ¹³C NMR spectra of compound **9** with the corresponding data for two stereoisomers of 2-methyl-1,3-dithiane¹⁶ showed that the methyl group at the C(2) atom in the dithiazinane ring is in the equatorial position, because its chemical shift (δ 19.72) agrees well with the corresponding chemical shift (δ 20.80) of the equatorial methyl group of 2-methyl-1,3-dithiane, whereas the corresponding resonance of the axial isomer should appear at much lower field (δ 25.4). It is also known¹⁷ that under these reaction conditions, the condensation of *N*-nitrosodimethylamine, acetaldehyde, and NaSH affords the 2,4,6-*cis,cis* stereoisomer. Hence, it can be said with certainty that heterocycle **9** is the stereo-





R = Me (7, 9, 10), Et (8)

Reagents and conditions: *i*. RCHO $-H_2S$ (3 : 2), 20 °C, EtOH. *ii*. RCHO $-H_2S$ (3 : 2), EtOH, refluxing. *iii*. RCHO $-H_2S$ (3 : 2), 1) BuONa/BuOH, 2) HCl.

isomer with the 2,4,6-*cis*,*cis* configuration of the methyl substituents.

To sum up, the cyclothiomethylation of carboxylic acid hydrazides with aldehydes (CH₂O, MeCHO) and H₂S in the presence of BuONa/BuOH (pH 11.25) selectively gives N-(1,3,5-dithiazin-5-yl)amides of carboxylic acids in 55–68% yields. 3-Acyl-1,3,4-thiadiazolidines were synthesized by the reactions of carboxylic acid hydrazides with formaldehyde and H₂S at 0––50 °C in 21–56% yields.

Experimental

Experiments were carried out with the use of the starting compounds with a purity of 95%. The solvents were purified, dried, and distilled according to known procedures.¹⁸

The ¹H and ¹³C NMR spectra were recorded on a Bruker Avance-400 spectrometer operating at 400.13 and 100.62 MHz, respectively, in DMSO-d₆ (δ_C , 39.50) and CDCl₃ (δ_C , 77.10). The IR spectra were measured on a Specord 75 IR spectrometer as Nujol mulls. The LC-mass-spectrometric analysis of compounds 2b and 3b was carried out on a Finnigan 4021 instrument (50000×0.25 mm glass capillary column, HP-5 stationary phase, helium as the carrier gas, temperature-programmed from 50 to $300 \,^{\circ}\text{C}$ at a rate of 5 deg min⁻¹, the temperature of the vaporizer was 280 °C, the temperature of the ion source was 250 °C). Compounds 2a,c,d, 3a,c,d, and 4-10 were analyzed on a Shimadzu LCMS-2010 EV instrument by atmospheric pressure chemical ionization (APCI) mass spectrometry; the maximum temperature of the APCI probe was 500 °C; the temperature of the heating source was 200 °C, the temperature of the vaporizer was 250 °C; nitrogen that was produced by an NM18L ultra-high purity nitrogen generator was used as the nebulizing gas; the liquid flow rate was 0.05 mL min⁻¹, the nebulizing gas flow rate was 2.5 mL min⁻¹; the ion source voltage was as follows: (+), 4.5 kV; (-), 3 kV.

The elemental analysis was carried out on a Carlo Erba (model 1106) elemental analyzer. The melting points were determined on a PHMK 80/2617 instrument; the refractive index of compound **9** was measured on an IRF-22 refractometer; the pH values of solutions were measured and adjusted using a pH-340 pH-meter.

Cyclothioalkylation of acid hydrazides with aldehydes and H₂S (general procedure). A solution of aldehyde (2 or 3 mol) was saturated with H₂S by bubbling for 30 min until the required aldehyde : H_2S ratio (2 : 1 or 3 : 2) was achieved. Then a solution of isonicotinic acid hydrazide (1 mol) in water (method A), in EtOH (method B), in aqueous 37% HCl (1 : HCl = 1 : 3) (method C), or in BuONa/BuOH (1 : BuONa = 1 : 3) (method D) was added dropwise to the thiomethylating mixture, and the reaction solution was stirred for 5 h at a specified temperature (-50, -10, -10)0, 20, or 70 °C). In the method C, the mixture was neutralized with NaOH; in the method D, with HCl. The reaction products were extracted with chloroform and separated by column chromatography on silica gel (CHCl₃-AcOEt-EtOH, 5:1:1 (2a, 3a), diethyl ether—AcOEt, 5:1 (2b, 3b), CHCl₃—AcOEt—acetone, 10 : 1 : 1 (2c, 3c), and CHCl₃-AcOEt, 10 : 1 (2d, 3d) as the eluents).

Cyclothiomethylation of isonicotinic acid hydrazide (1a) with H_2S and CH_2O by the reverse mixing procedure. Paraformaldehyde (1 mol), isonicotinic acid hydrazide (1a) (1 mol), and benzene (15 mL) were placed into a reactor at room temperature. The reaction mixture was refluxed with stirring for 2 h and then divided into two aliquots. Imine **4** was isolated from one aliquot. Another aliquot was saturated with hydrogen sulfide for 5 h. After bubbling with H_2S , the products were extracted with chloroform and separated by column chromatography on silica gel (CHCl₃-AcOEt-EtOH, 5:1:1, as the eluent; compounds **2a** and **3a**).

Cyclothiomethylation of isonicotinic acid hydrazide (1a) with H_2S and CH_2O in the presence of the Na/C₆H₆ system. Isonicotinic acid hydrazide (1a) (1 mol), benzene (20 mL), and sodium metal (1 mol) were mechanically stirred in a reactor until a homogeneous cream-like substance was obtained. Then paraformaldehyde (1 mol) was added. The mixture was stirred at 70 °C for 3 h, and then hydrogen sulfide was bubbled through the reaction mixture for 4 h. The benzene was decanted, and a mixture of heterocycles 2a and 3a was isolated from the residue by extraction with CHCl₃.

4-(Isonicotinoyl)-1,3,4-thiadiazolidine (2a). The yield was 56% (method *A*), m.p. 185–187 °C, $R_{\rm f}$ 0.35 (CHCl₃–AcOEt–EtOH, 5 : 1 : 1). IR, v/cm⁻¹: 690, 900, 1080, 1270, 1440–1520, 1660, 2900, 3385. ¹H NMR (DMSO-d₆), δ : 4.35 (s, 2 H, C(2)H₂); 4.53 (s, 2 H, C(5)H₂); 7.64–7.70 (d, 2 H, C(10)H, C(12)H, *J* = 5.6 Hz); 8.68–8.75 (d, 2 H, C(9)H, C(13)H, *J* = 5.6 Hz); 10.09 (s, 1 H, N(3)H). ¹³C NMR (DMSO-d₆), δ : 55.5 (t, C(2)); 74.0 (t, C(5)); 121.6 (d, C(9), C(11)); 140.8 (d, C(7)); 150.2 (d, C(8), C(12)); 163.7 (s, C(6)). MS, *m/z* ($I_{\rm rel}$ (%)): 196 [M + 1]⁺ (10); 151 [M – CH₂SCH₂NHN]⁺ (12). Found (%): C, 49.83; H, 4.64; N, 21.87; S, 16.67. C₈H₉N₃OS. Calculated (%): C, 49.23; H, 4.62; N, 21.54; S, 16.41.

4-Benzoyl-1,3,4-thiadiazolidine (2b). The yield was 21% (method *A*), m.p. 223–225 °C, R_f 0.40 (diethyl ether—AcOEt, 5 : 1, as the eluent). IR, v/cm⁻¹: 670, 1100, 1265, 1360–1450, 1620, 1670, 2900, 3315. ¹H NMR (DMSO-d₆), δ : 4.63 (s, 2 H, C(5)H₂); 5.05 (s, 2 H, C(2)H₂); 7.60–7.80 (m, 4 H, Ph); 9.83 (s, 1 H, N(3)H). ¹³C NMR (DMSO-d₆), δ : 55.0 (t, C(5)); 73.5 (t, C(2)); 127.7 (d, C(8), C(12)); 128.6 (d, C(9), C(11)); 132.0 (d, C(10)); 132.8 (d, C(7)); 166.2 (s, C(6)). MS, *m/z* (I_{rel} (%)): 194 [M]⁺ (2); 149 [M – CH₂S]⁺ (43); 119 [M – CH₂SCH₂NH]⁺ (61); 105 [PhCO]⁺ (100); 45 [CH₂S]⁺ (17). Found (%): C, 56.48; H, 5.74; N, 14.87; S, 15.97. C₉H₁₀N₂OS. Calculated (%): C, 55.65; H, 5.19; N, 14.42; S, 16.51.

4-(o-Methoxybenzoyl)-1,3,4-thiadiazolidine (2c). The yield was 34% (method *A*), m.p. 110–112 °C, $R_{\rm f}$ 0.09 (CHCl₃–AcOEt– –acetone, 10 : 1 : 1, as the eluent). IR, v/cm⁻¹: 740, 1005, 1165, 1280, 1470–1480, 1595, 1640, 2840, 2950, 3295. ¹H NMR (DMSO-d₆), δ : 3.65 (br.s, 3 H, C(13)H₃); 3.92 (s, 2 H, C(5)H₂); 4.25 (s, 2 H, C(2)H₂); 7.10–7.95 (m, 4 H, Ph); 9.33 (s, 1 H, N(3)H). ¹³C NMR (DMSO-d₆), δ : 56.0 (s, C(13)); 56.5 (t, C(5)); 74.0 (t, C(2)); 112.1 (d, C(12)); 120.8 (d, C(11)); 121.7 (d, C(10)); 131.2 (d, C(9)); 133.5 (d, C(8)); 157.5 (d, C(7)); 164.4 (s, C(8)). MS, *m/z* ($I_{\rm rel}$ (%)): 225 [M + 1]⁺ (50); 193 [M – S]⁺ (10); 180 [M – SCH₂]⁺ (100); 165 [M – SCH₂N]⁺ (20); 136 [M – CH₂SCH₂NNH]⁺ (10); 93 [M – CH₂SCH₂NNHCO,Me]⁺ (15). Found (%): C, 53.58; H, 5.29; N, 14.87; S, 15.18. C₁₀H₁₂N₂O₂S. Calculated (%): C, 53.57; H, 5.36; N, 12.50; S, 14.29.

4-(4-Hydroxybenzylcarbonyl)-1,3,4-thiadiazolidine (2d). The yield was 36% (method *A*), m.p. 183–185 °C, $R_{\rm f}$ 0.15 (CHCl₃–AcOEt, 10 : 1, as the eluent). IR, v/cm⁻¹: 670, 1100, 1265, 1360–1450, 1620, 1670, 2900, 3315. ¹H NMR (DMSO-d₆), δ : 4.56 (s, 1 H, OH(13)); 4.15 (s, 2 H, C(5)H₂); 4.81 (s, 2 H, C(2)H₂); 6.71–7.00 (m, 4 H, Ph); 9.35 (s, 1 H, N(3)H). ¹³C NMR (DMSO-d₆), δ : 55.9 (t, C(5)); 70.9 (t, C(2)); 115.1 (s, C(9), C(11)); 126.5 (d, C(10)); 130.0 (d, C(8), C(12)); 155.9 (d, C(7)); 169.9 (s, C(6)). MS, *m/z* (*I*_{rel} (%)): 224 [M]⁺ (15); 180 [M - SCH₂]⁺ (100); 150 [M - CH₂SCH₂NH]⁺ (11); 135 [M - CH₂SCH₂NNH]⁺ (5). Found (%): C, 53.47; H, 5.29. C₁₀H₁₂N₂O₂S. Calculated (%): C, 53.55; H, 5.39.

N-(1,3,5-Dithiazinan-5-yl)isonicotinamide (3a). The yield was 62% (method *B*), 68% (method *D*), m.p. 200–202 °C, $R_{\rm f}$ 0.35 (CHCl₃-AcOEt-EtOH, 5 : 1 : 1). IR, v/cm⁻¹: 700, 1070, 1270, 1460–1540, 1650, 2905, 3390. ¹H NMR (DMSO-d₆), δ : 4.59 (s, 2 H, C(2)H₂); 5.06 (s, 4 H, C(4)H₂, C(6)H₂); 8.12–8.17 (d, 2 H, C(11)H, C(13)H, *J* = 4.7 Hz); 9.14–9.19 (d, 2 H, C(10)H, C(14)H, *J* = 4.7 Hz); 9.74 (s, 1 H, N(7)H). ¹³C NMR (DMSO-d₆), δ : 30.9 (t, C(2)); 57.7 (t, C(4), C(6)); 121.5 (d, C(11), C(13)); 141.2 (d, C(9)); 150.3 (d, C(10), C(14)); 163.2 (s, C(8)). MS, *m/z*($I_{\rm rel}$ (%)): 242 [M]⁺(5); 150 [M – SCH₂SCH₂]⁺ (100); 122 [M – CH₂SCH₂SCH₂N]⁺ (13); 108 [M – CH₂SCH₂SCH₂NHN]⁺ (6). Found (%): C, 46.18; H, 4.74; N, 17.97; S, 25.97. C₉H₁₁N₃OS₂. Calculated (%): C, 44.81; H, 4.56; N, 17.43; S, 26.55.

N-(1,3,5-Dithiazinan-5-yl)benzamide (3b). The yield was 57% (method *B*), 57% (method *D*), m.p. 240–242 °C, $R_{\rm f}$ 0.43 (diethyl ether—AcOEt, 5 : 1, as the eluent). IR, v/cm⁻¹: 700, 1070, 1250, 1360–1450, 1615, 1670, 2900, 3310. ¹H NMR (DMSO-d₆), & 4.20 (s, 2 H, C(2)H₂); 4.66 (s, 4 H, C(4)H₂, C(6)H₂); 7.60–7.80 (m, 4 H, Ph); 8.86 (s, 1 H, N(7)H). ¹³C NMR (DMSO-d₆), & 30.9 (t, C(2)); 58.1 (t, C(4), C(6)); 127.3 (d, C(11), C(13)); 128.8 (d, C(10), C(14)); 131.9 (d, C(12)); 148.5 (d, C(9)); 155.6 (s, C(8)). MS, m/z ($I_{\rm rel}$ %): 240 [M]⁺ (25); 162 [M – SCH₂S]⁺ (5); 121 [M – PhCONH]⁺ (6); 105 [PhCO]⁺ (100); 45 [CH₂S]⁺ (10). Found (%): C, 49.47; H, 4.99; N, 11.84; S, 25.37. C₁₀H₁₂N₂OS₂. Calculated (%): C, 49.97; H, 5.03; N, 11.66; S, 26.68.

N-(1,3,5-Dithiazinan-5-yl)-*o*-methoxybenzamide (3c). The yield was 59% (method D), m.p. 105-107 °C, R_f 0.50 (CHCl₃-AcOEt-acetone, 10:1:1, as the eluent). IR, v/cm⁻¹: 740, 1005, 1100, 1265, 1370-1445, 1595, 1640, 2840, 2900, 3340. ¹H NMR (DMSO-d₆), δ: 3.51 (br.s, 3 H, C(15)H₃); 4.05 (s, 2 H, C(2)H₂); 4.54 (s, 4 H, C(4)H₂, C(6)H₂); 7.11 (t, 1 H, C(11)H, J = 7.6 Hz; 7.15 (d, 1 H, C(10)H, J = 8.6 Hz); 7.57 (t, 1 H, C(12)H, J = 7.4 Hz); 7.99 (d, 1 H, C(13)H, J = 7.4 Hz);10.23 (s, 1 H, N(7)H). ¹³C NMR (DMSO-d₆), δ: 30.9 (t, C(2)); 56.7 (t, C(4), C(6)); 58.1 (s, C(15)); 112.6 (d, C(13)); 120.5 (d, C(12)); 121.1 (d, C(11)); 131.2 (d, C(14)); 136.1 (d, C(10)); 157.1 (d, C(9)); 164.4 (s, C(8)). MS, $m/z (I_{rel} (\%))$: 271 [M + 1]⁺ (100); 179 [M - SCH₂SCH₂]⁺ (72); 151 [M - SCH₂SCH₂N]⁺ (8); 135 [M - SCH₂SCH₂NNH]⁺ (13). Found (%): C, 49.42; H, 5.57; N, 10.37; S, 24.28. $C_{11}H_{14}N_2O_2S_2$. Calculated (%): C, 48.88; H, 5.19; N, 10.37; S, 23.70.

N-(1,3,5-Dithiazinan-5-yl)-2-(hydroxyphenyl)acetamide (3d). The yield was 55% (method *D*), m.p. 209–210 °C, R_f 0.52 (CHCl₃–AcOEt, 10 : 1, as the eluent). IR, v/cm⁻¹: 700, 1070, 1250, 1360–1450, 1615, 1670, 2900, 3310. ¹H NMR (DMSO-d₆), δ: 3.89 (s, 2 H, C(2)H₂); 4.44 (s, 4 H, C(4)H₂, C(6)H₂); 6.63–6.73 (d, 2 H, C(12)H, C(14)H, *J* = 8.5 Hz); 7.04–7.14 (d, 2 H, C(11)H, C(15)H, *J* = 8.5 Hz); 8.94 (s, 1 H, N(7)H); 9.26 (s, 1 H, OH(15)). ¹³C NMR (DMSO-d₆), δ: 31.2 (t, C(2)); 57.9 (t, C(4), C(6)); 115.2 (d, C(11), C(13)); 126.0 (d, C(12)); 130.2 (d, C(10), C(14)); 156.1 (d, C(9)); 168.4 (s, C(8)). MS, $m/z (I_{rel}(\%))$: 271 [M + 1]⁺ (49); 179 [M - SCH₂SCH₂]⁺ (100); 151 [M - SCH₂SCH₂N]⁺ (9); 107 [M - CH₂SCH₂SNNCH₂]⁺ (12). Found (%): C, 49.12; H, 5.47; N, 10.39; S, 24.28. C₁₁H₁₄N₂O₂S₂. Calculated (%): C, 48.86; H, 5.22; N, 10.36; S, 23.72.

N[']-(Methylidene)isonicotinic acid hydrazide (4). The yield was 90% (method *A*), m.p. 156–158 °C, $R_{\rm f}$ 0.58 (CHCl₃–EtOH, 5 : 1, as the eluent). IR, v/cm⁻¹: 1060, 1160, 1300, 1530, 1650, 2905, 3150, 3250. ¹H NMR (DMSO-d₆), δ : 4.20 (br.s, 2 H, C(10)H); 7.67 (d, 2 H, C(2)H, C(6)H, *J* = 5.6 Hz); 8.72 (d, 2 H, C(3)H, C(5)H, *J* = 5.6 Hz); 10.33 (s, 1 H, N(8)H). ¹³C NMR (DMSO-d₆), δ : 121.6 (d, C(2), C(6)); 140.60 (d, C(4)); 149.7 (d, C(10)); 150.4 (d, C(3), C(5)); 163.9 (s, C(7)). MS, m/z ($I_{\rm rel}$ (%)): 150 [M]⁺ (100); 138 [M – CH₂]⁺ (20); 123 [M – CH₂N]⁺ (10). Found (%): C, 56.11; H, 4.71; N, 27.85. C₇H₇N₃O. Calculated (%): C, 56.37; H, 4.73; N, 28.17.

Bis[4-(isonicotinoylaminohydrazino)methylsulfanyl]methane (5a). The yield was 14% (method C), $R_{\rm f} 0.05$ (CHCl₃-AcOEt--EtOH, 10 : 1 : 1, as the eluent). IR, v/cm^{-1} : 740, 1020, 1190, 1280, 1370–1410, 1500, 1590, 1650, 2905, 3280. ¹H NMR (DMSO-d₆), δ: 3.97 (s, 2 H, C(12)H₂); 4.27 (s, 4 H, C(10)H₂, C(10')H₂); 7.49 (s, 1 H, N(9)H, N(9')H); 7.78 (d, 4 H, C(2)H, C(2')H, C(6)H, C(6')H, J = 4.4 Hz; 8.83 (d, 4 H, C(3)H, C(3')H, C(5)H, C(5')H), J = 4.4 Hz); 9.29 (s, 2 H, N(8)H,N(8')H). ¹³C NMR (DMSO- d_6), δ : 33.7 (t, C(12)); 56.6 (t, C(10), C(10')); 120.6 (d, C(2), C(2'), C(6), C(6')); 140.9 (d, C(4), C(4')); 151.5 (d, C(3), C(3'), C(5), C(5')); 162.9 (s, C(7), C(7')). MS, *m/z* (*I*_{rel} (%)): 378 [M]⁻ (50); 184 [M - CH₂SCH₂NHNHCOC₅H₄N]⁻ (100); 148 [M -- CH₂SCH₂SCH₂NHNHCOC₅H₄N]⁻ (37). Found (%): C, 48.58; H, 4.54; N, 20.17; S, 17.23. C₁₅H₁₈N₆O₂S₂. Calculated (%): C, 47.60; H, 4.79; N, 22.21; S, 16.94.

1,3,5,7,9-Oxatetrathiecane (6). The yield was 10%. MS, $m/z (I_{rel} (\%)): 217 [M]^+ (87); 201 [M - CH_2]^+ (100); 185 [M - CH_2O]^+ (44); 182 [M - S]^+ (87); 138 [M - CH_2SCH_2O]^+ (87).$

N'-(Ethylidene)isonicotinic acid hydrazide (7). The yield was 72% (method *A*), m.p. 150–151 °C, $R_{\rm f}$ 0.69 (CHCl₃–EtOH, 5 : 1, as the eluent). IR, v/cm⁻¹: 1020, 1120, 1290, 1455–1540, 1610, 1650, 2900, 3050, 3210. ¹H NMR (CDCl₃), δ : 1.94 (m, 3 H, C(11)H₃); 7.67 (d, 2 H, C(2)H, C(6)H, *J* = 6.0 Hz); 7.78 (q, 1 H, C(10)H, *J* = 5.2 Hz); 8.61 (d, 2 H, C(3)H, C(5)H, *J* = 6.0 Hz); 10.97 (s, 1 H, N(8)H). ¹³C NMR (CDCl₃), δ : 18.6 (s, C(11)); 121.0 (d, C(2), C(6)); 140.4 (d, C(4)); 150.3 (d, C(3), C(5)); 150.9 (t, C(10)); 162.7 (s, C(7)). MS, *m/z* (*I*_{rel} (%)): 164 [M]⁺ (100); 138 [M – CH₃CH]⁺ (20); 121 [M – CH₃CHN]⁺ (17); 78 [M – CH₃CHNHNCO]⁺ (8). Found (%): C, 58.18; H, 5.71; N, 25.97. C₈H₉N₃O. Calculated (%): C, 58.88; H, 5.56; N, 25.75.

N - (**Propylidene**)isonicotinic acid hydrazide (8). The yield was 50% (method *A*), m.p. 134–135 °C, $R_{\rm f}$ 0.73 (CHCl₃–EtOH, 5 : 1, as the eluent). IR, v/cm⁻¹: 1000, 1130, 1295, 1450–1530, 1615, 1650, 2905, 3035, 3190. ¹H NMR (CDCl₃), δ : 1.04 (m, 3 H, C(12)H₃); 2.27 (m, 2 H, C(11)H₂); 7.70 (d, 2 H, C(2)H, C(6)H, *J* = 4.4 Hz); 7.77 (t, 1 H, C(10)H, *J* = 5.2 Hz); 8.61 (d, 2 H, C(3)H, C(5)H, *J* = 4.4 Hz); 10.06 (s, 1 H, N(8)H). ¹³C NMR (CDCl₃), δ : 10.6 (s, C(12)); 26.0 (s, C(11)); 121.5 (d, C(2), C(6)); 140.4 (d, C(4)); 150.3 (d, C(3), C(5)); 156.0 (t, C(10)); 162.8 (s, C(7)). MS, *m/z* (*I*_{rel} (%)): 178 [M]⁺ (100); 150 [M - CH₃CH₂]⁺ (6); 135 [M - CH₃CH₂CH]⁺ (79); 121 [M - CH₃CH₂CHN]⁺ (6). Found (%): C, 61.18; H, 6.34; N, 24.07. C₉H₁₁N₃O. Calculated (%): C, 61.00; H, 6.26; N, 23.71.

N-(2,4,6-Trimethyl-1,3,5-dithiazinan-5-yl)isonicotinamide (9). The yield was 32% (method C), 41% (method D), n_D^{20} 1.4990, $R_{\rm f}$ 0.78 (CHCl₃-AcOEt-EtOH, 5:1:1, as the eluent). IR, v/cm⁻¹: 700, 1050, 1255, 1450–1510, 1600, 1695, 2935, 3370. ¹H NMR (CDCl₃), δ: 1.39 (m, 3 H, C(15)H₃); 1.64 (m, 6 H, C(16)H₃, C(17)H₃); 3.54 (s, 2 H, C(2)H₂); 4.28 (s, 4 H, C(4)H₂, $C(6)H_2$; 7.30 (d, 2 H, C(11)H, C(13)H), J = 4.2 Hz); 7.66 (d, 2 H, C(10)H, C(14)H, J = 4.2 Hz); 8.71 (s, 1 H, N(7)H). ¹³C NMR (CDCl₃), δ: 19.5 (s, C(15)); 23.6 (s, C(16), C(17)); 44.2 (s, C(2)); 68.9 (s, C(4), C(6)); 121.1 (d, C(11), C(13)); 140.2 (d, C(9)); 150.6 (d, C(10), C(14)); 164.9 (s, C(8)). MS, $m/z (I_{rel} (\%)): 284 [M + 1]^+ (100); 224 [M - CH_3 CHSCHCH_3]^+$ (5); 164 [M - SCHCH₃SCHCH₃]⁺ (25); 107 [M -- CH₃CHSCHCH₃SCHCH₃]⁺ (7). Found (%): C, 50.52; H, 6.09; N, 14.57; S, 22.78. C₁₂H₁₇N₃OS₂. Calculated (%): C, 50.85; H, 6.05; N, 14.83; S, 22.63.

1,1-Bis[4-(isonicotinoyl-2,5-dimethyl-1,3,4-thiadiazolidin-3-yl)ethylsulfanyl]ethane (10). The yield was 6% (method *C*). MS, $m/z (I_{rel}(\%))$: 591 [M – H]⁻ (53); 531 [M – SCHCH₃]⁻ (100); 515 [M – CHCH₃SCHCH₃]⁻ (58); 413 [M – 2 CHCH₃SCHCH₃]⁻ (41); 281 [M – C₅H₄NCONCHCH₃SCHCH₃NCH-CH₃]⁻ (29); 223 [M – C₅H₄NCONCHCH₃SCHCH₃NCH-

CH₃SCHCH₃SCHCH₃]⁻ (24); 190 [M - C₅H₄NCONCH-CH₃SCHCH₃NCHCH₃SCHCH₃SCHCH₃,S]⁻ (29).

X-ray diffraction study of compounds 3a,c,d and 8. Crystals of 3a were grown by crystallization from a 5:1:1 CHCl₃—AcOEt— —EtOH mixture; crystals of 3c, by crystallization from a 10:1:1 CHCl₃—AcOEt—acetone mixture; crystals of 3d, by crystallization from a 10:1 CHCl₃—AcOEt mixture; crystals of 8, by crystallization from a 5:1 CHCl₃—EtOH mixture.

The X-ray diffraction data sets for compounds **3a,c,d** and **8** were collected on Bruker SMART CCD Area Detector (**3c,d, 8**) and Bruker APEX 2 CCD Area Detector (**3a**) diffractometers (Mo-K α radiation) at 100, 295, 100, and 120 K for compounds **3a,c,d** and **8**, respectively. For compounds **3a,c,d**, semiempirical absorption corrections were applied; the transmission coefficients T_{max} and T_{min} were determined using the SADABS program.¹⁹ The structures were solved by direct methods. All nonhydrogen atoms were located in different electron density maps and refined based on F^2_{hkl} with anisotropic displacement parameters. The hydrogen atoms of the NH and OH groups were located in difference electron density maps. The H(C) atoms were positioned geometrically. All hydrogen atoms were refined using a riding model with U(H) = 1.2U(Ci), where U(X) are the

Table 4. Crystallographic characteristics and the structure refinement statistics for 3a,c,d and 8

| Parameter | 3a | 3c | 3d | 8 |
|--|---|--|--|---|
| Molecular formula | $C_9H_{11}N_3OS_2$ | C ₁₁ H ₁₄ N ₂ O ₂ S ₂ | C ₁₁ H ₁₄ N ₂ O ₂ S ₂ | C ₉ H ₁₁ N ₃ O |
| М | 241.33 | 270.36 | 270.36 | 177.21 |
| Space group | Pccn | Pbca | $P2_1/c$ | $P\overline{1}$ |
| a/Å | 12.083(1) | 8.778(1) | 12.754(3) | 9.893(2) |
| b/Å | 20.398(2) | 12.143(1) | 7.864(2) | 14.219(2) |
| c/Å | 9.0115(9) | 23.483(3) | 13.496(3) | 15.123(2) |
| α/deg | 90 | 90 | 90 | 93.022(4) |
| β/deg | 90 | 90 | 111.675(6) | 106.224(3) |
| γ/deg | 90 | 90 | 90 | 109.655(3) |
| $V/Å^3$ | 2221.1(4) | 2502.9(5) | 1258.0(5) | 1898.1(5) |
| Ζ | 8 | 8 | 4 | 8 |
| $d_{\rm calc}/{\rm g}~{\rm cm}^{-3}$ | 1.443 | 1.435 | 1.428 | 1.240 |
| Crystal color and shape Colorless r | | | ess prism | |
| Crystal dimensions/mm | $0.25 \times 0.20 \times 0.20$ | $0.40 \times 0.30 \times 0.20$ | 0.20×0.20×0.15 | $0.25 \times 0.21 \times 0.18$ |
| μ/mm^{-1} | 0.456 | 0.417 | 0.414 | 0.085 |
| Absorption correction | | SADABS | | _ |
| $T_{\rm min}/T_{\rm max}$ | 0.896/0.914 | 0.851/0.921 | 0.923/0.940 | _ |
| $2\theta_{\rm max}/{\rm deg}$ | 56 | 58 | 58 | 56 |
| Number of reflections | 23456 | 14083 | 7927 | 19578 |
| Number of independent reflections (R_{int}) | 2658 (0.0803) | 3284 (0.0397) | 3333 (0.0559) | 9133 (0.0582) |
| R_1 (based on <i>F</i> for reflections with $I > 2\sigma(I)$) | 0.0509 (1932) | 0.0525 (2143) | 0.0485 (2186) | 0.0671 (3829) |
| wR_2 (based on F^2 for all reflections) | 0.1341 | 0.1555 | 0.1048 | 0.2206 |
| Number of refined parameters | 136 | 155 | 154 | 490 |
| Weighting scheme | $w^{-1} = \sigma^2(F_0^2) + (aP)^2 + bP, P = 1/3(F_0^2 + 2F_0^2)$ | | | |
| A | 0.0533 | 0.0313 | 0.0350 | 0.0918 |
| В | 4.8523 | 2.5802 | 0.2000 | 0.2650 |
| GOOF | 1.000 | 1.023 | 1.000 | 1.001 |
| <i>F</i> (000) | 1008 | 1136 | 568 | 752 |

equivalent thermal parameters of the corresponding parent atoms; for the hydrogen atoms of the methyl groups, U(H) = 1.5U(Ci). All calculations were carried out with the use of the SHELXTL PLUS 5.10 program package.¹⁴

Principal X-ray data collection and refinement statistics are given in Table 4. The atomic coordinates, bond lengths, bond angles, and thermal parameters were deposited with the Cambridge Crystallographic Data Centre (CCDC 716620–716623) and can be obtained, free of charge, on application to www.ccdc.cam.uk/conts/retrieving.html (or CCDC, 12 Union Road, Cambridge CB2 1EZ; fax: +44 1223 335 033; or deposit@ccdc.cam.ac.uk).

We thank Corresponding Member of the Russian Academy of Sciences U. M. Dzhemilev for helpful discussion.

This study was financially supported by the Council on Grants of the President of the Russian Federation (Program for State Support of Leading Scientific Schools of the Russian Federation, Grant NSh-3019.2008.3).

References

- A. S. Zasedatelev, A. L. Zhuze, K. Tsimmer, S. L. Grokhovskii, V. G. Tumanyan, G. V. Gurskii, B. P. Gottikh, *Dokl. Akad. Nauk SSSR*, 1976, 4, 1006 [*Dokl. Chem. (Engl. Transl.*), 1976].
- 2. M. D. Mashkovskii, *Lekarstvennye sredstva* [*Drugs*], Meditsina, Moscow, 1993, **1**, 322 pp. (in Russian).
- R. B. Pawar, V. V. Mulwad, *Khim. Geterotsikl. Soedin.*, 2004, 257 [Chem. Heterocycl. Compd. (Engl. Transl.), 2004, 40, 257].
- J. Matysiak, Z. Malinski, Bioorg. Khim., 2007, 33, 640 [Russ. J. Bioorg. Chem. (Engl. Transl.), 2007, 33, 640].
- 5. V. Roessler, Fr. Pat. 1341792; Chem. Abstrs, 1965, 65, 14529.
- V. R. Akhmetova, R. R. Khairullina, S. R. Khafizova, R. V. Kunakova, U. M. Dzhemilev, RF Pat. 2291150; *Byul. Izobr.* [*Inventor Bull.*], 2007, No. 1 (in Russian).

- V. R. Akhmetova, R. R. Khairullina, G. R. Nadyrgulova, R. V. Kunakova, U. M. Dzhemilev, *Zh. Org. Khim.*, 2008, 200 [*Russ. J. Org. Chem. (Engl. Transl.)*, 2008, **44**].
- V. R. Akhmetova, G. R. Nadyrgulova, T. V. Tyumkina, Z. A. Starikova, D. G. Golovanov, M. Yu. Antipin, R. V. Kunakova, U. M. Dzhemilev, *Izv. Akad. Nauk, Ser. Khim.*, 2006, 1758 [*Russ. Chem. Bull., Int. Ed.*, 2006, 55, 1824].
- 9. A. Wohl, Ber., 1886, 19, 2344.
- V. R. Akhmetova, R. A. Vagapov, G. R. Nadyrgulova, T. V. Tyumkina, Z. A. Starikova, M. Y. Antipin, R. V. Kunakova, U. M. Dzhemilev, *Tetrahedron*, 2007, **47**, 11702.
- V. R. Akhmetova, G. R. Nadyrgulova, Z. T. Niatshina, R. R. Khairullina, Z. A. Starikova, A. O. Borisova, M. Yu. Antipin, R. V. Kunakova, U. M. Dzhemilev, *Heterocycles*, 2009, 78, 45.
- S. R. Khafizova, V. R. Akhmetova, R. V. Kunakova, U. M. Dzhemilev, *Izv. Akad. Nauk, Ser. Khim.*, 2003, 1722 [*Russ. Chem. Bull., Int. Ed.*, 2003, **52**, 1817].
- S. R. Khafizova, V. R. Akhmetova, L. F. Korzhova, T. V. Khakimova, G. R. Nadyrgulova, R. V. Kunakova, E. A. Kruglov, U. M. Dzhemilev, *Izv. Akad. Nauk, Ser. Khim.*, 2005, 423 [*Russ. Chem. Bull., Int. Ed.*, 2005, **54**, 432].
- G. M. Sheldrick, SHELXTL v. 5.10, Structure Determination Software Suite, Bruker AXS, Madison, Wisconsin, USA, 1998.
- 15. R. L. Hinman, M. C. Flores, J. Org. Chem., 1958, 23, 660.
- R. G. Buttery, I. G. Turnbaugh, L. C. Ling, J. Agric. Food Chem., 1988, 36, 1006.
- Th. J. Hansen, R. M. Angeles, L. K. Keefer, *Tetrahedron*, 1981, 37, 4143.
- A. J. Gordon, R. A. Ford, *The Chemist's Companion*, New York–London–Sydney–Toronto, 1972.
- SADABS, Version 2004/1, Bruker AXS Inc., Madison, WI, USA, 2005.

Received May 5, 2009; in revised form August 5, 2009