# REVIEW PAPER



# **Comprehensive insights into sustainable conversion of agricultural and food waste into microbial protein for animal feed production**

**Kashif Rasoo[l](http://orcid.org/0000-0002-7759-8448) · Sabir Hussain · Asif Shahzad · Waheed Miran · Khaled A. Mahmoud · Nisar Ali · Fares Almomani**

Received: 29 December 2022 / Accepted: 3 April 2023 / Published online: 15 April 2023 © The Author(s) 2023

**Abstract** The growing global population and higher living standards instantly demand the transition in the direction of a sustainable food system. A substantial section of means and agricultural lands are presently committed to protein-rich feed production to rear livestock for human consumption. Conversely, accelerated farming activities and the food industry have rendered a drastic increase in waste which impair the economic and environmental sustainability of the ecosystem. This situation emerges the need for developing an integrated technology for waste management and to improve sustainability footprints. Microbial protein (MP) production based on renewable electron and carbon sources has the potential as a substitute protein source. MP production for animal feed use is growing fast and is derived from bacteria, algae, and fungi including yeast. MP produced from

all types of microbes is currently commercialized and in use. However, novel methods and processes are also under investigation to make MP production more economical and sustainable. Current research on MP has concentrated on the valorization of waste materials by using high protein content-containing microorganisms, which can then be used in animal feed. Using such kind of integrated approach, the agroindustry waste resources upcycling can contribute towards fnding sustainable, cheaper, and environment-friendly protein sources. This review frst describes the potential waste feedstock for MP production and summarizes the recent progress in the application of MP-producing microorganisms including fungus, yeast, bacteria, and phototrophic

Kashif Rasool, Sabir Hussain have contributed equally.

K. Rasool  $(\boxtimes) \cdot$  K. A. Mahmoud Qatar Environment and Energy Research Institute (QEERI), Qatar Foundation, Hamad Bin Khalifa University, P.O. Box 5825, Doha, Qatar e-mail: krasool@hbku.edu.qa

#### S. Hussain

Department of Environmental Science, Institute of Space Technology, Islamabad, Pakistan

#### A. Shahzad

Department of Materials Science and Engineering, Uppsala University, Box 534, 75121 Uppsala, Sweden

#### W. Miran

School of Chemical and Materials Engineering, National University of Sciences and Technology, Islamabad 44000, Pakistan

#### N. Ali

Key LaboratoHry for Palygorskite Science and Applied Technology of Jiangsu Province, National and Local Joint Engineering Research Center for Mineral Salt Deep Utilization, Huaiyin Institute of Technology, Huai'an 223003, China

F. Almomani

Department of Chemical Engineering, Qatar University, P. O. Box 2713, Doha, Qatar

microbes. Bioprocesses, and production technology advances for MP production have been explored and discussed in detail. Finally, the MP application as animal feed, its challenges, and future perspectives in research have been evaluated.

**Keywords** Agro-food waste · Microbial protein · Fermentation · Sustainability · Animal feed

## **1 Introduction**

The growing population and increasing living standards in developed and developing countries globally are anticipated to generate 1,250 million tonnes per year of worldwide meat demand by 2050 (Ritala et al. [2017\)](#page-32-0). Livestock farming is a signifcant constituent of the agricultural economies of several countries. However, fulflling the demand for increased meat and dairy products from conventional sources will not be sustainable due to the inferior conversion productivity of feed to protein (Aiking [2011;](#page-25-0) Alberti et al. [2022\)](#page-26-0). Meat and dairy production is ever-increasing to meet the global demand for animal-derived protein. Livestock farming has long played an important role in sustaining the nutritional requirements of the world's population. Rapid industrial and population development over the last few decades has led to the development of an emerging livestock industry in most agricultural countries (Mugagga and Nabaasa [2016\)](#page-31-0). Traditional sources such as green fodder, and silage obtained from a variety of plants are not considered sustainable to meet the protein-rich feed requirement for animal farming (Newman et al. [2023\)](#page-31-1). Therefore, new solutions are needed for a sustainable protein supply.

On the other hand, accelerated agriculture farming activities and food industry have rendered a drastic increase in agricultural waste (Garrity et al. [2010](#page-28-0); Kesavan and Swaminathan [2008\)](#page-29-0). About 4532 trillion British thermal units (TBtu) of biomass were generated only in the United States (USA), making up about 4.9% of the total primary energy con-sumption of the USA in 2020 (Dey et al. [2021](#page-27-0)). Agriculture-based industries also produced a large number of waste materials (Newman et al. [2023](#page-31-1)). Agriculture waste is originated from diverse waste stream sources across the farms including farms and agro-industries like animal manure, vegetable, and farm waste generated from production to packing activities (Ahmad Khorairi et al. [2021;](#page-25-1) Asiri and Chu [2022;](#page-26-1) Díaz-Vázquez et al. [2021](#page-27-1)). These wastes impair the economic and environmental sustainability of the ecosystem. For public health and wellbeing, valuable and state-of-the-art recycling methods are needed. This situation emerges the need for developing an integrated technology for waste management and to improve sustainability footprints. Currently, utilizing agriculture industry and food waste through recycling for resource recovery and manufacturing of value-added products has shown a great potential. An assimilated bio-refnery concept is steadily developing into an optimistic resolution with various products developments such as biofuels, biomaterials, and other bioactive compounds.

Ding et al. [2023;](#page-27-2) Gervasi et al. [2018](#page-28-1); Jones et al. [2020;](#page-29-1) Nyyssölä et al. [2022](#page-31-2); Raziq et al. [2020](#page-32-1); Sharif et al. [2021](#page-32-2); Zeng et al. [2022;](#page-35-0) Zhou et al. [2022a;](#page-35-1) Zhu et al. [2022\)](#page-35-2). The protein produced in microbial cells also known as MP is an option with the potential to address the issue simultaneously. Microbial strains which are characterized by protein contents higher than 30% in their biomass and essential amino acids are considered more suitable for this purpose (Bourdichon et al. [2012;](#page-26-2) Jiang et al. [2022;](#page-29-2) Raziq et al. [2020;](#page-32-1) Yang et al. [2022](#page-34-0); Zhou et al. [2022a\)](#page-35-1). Microorganisms have been traditionally used for centuries in fermented foods. Microalgae and fungal-derived commercial products are already available in the market for human consumption with various commercial names such as Spirulina, Chlorella, Dunaliella salina, Aphanizomenon, QuornTM, and, Algaeon, primarily as health supplements (Ritala et al. [2017](#page-32-0)). Some strict regulatory frameworks in place are relevant for MP human consumption (Bourdichon et al. [2012](#page-26-2)). The microbial sources for animal feed production are generally wider compared to those permitted for human consumption. This is because the safety and quality standards for animal feed are typically diferent from those for human consumption. MP production for animal feed use is growing fast and is derived from bacteria, algae, and fungi (including yeast) (Asiri and Chu [2022](#page-26-1); Khoshnevisan et al. [2022;](#page-29-3) Woolley et al. [2023](#page-34-1); Zheng et al. [2023\)](#page-35-3). Animal feed from sugar fermentation was produced in Finland in 1974 and the process was registered with the commercial name "PEKILO®" to be used in the European Union countries. MP produced from microbes is currently commercialized and in use as animal feed from companies like Uniprotein®, and Unibio A/S, Denmark, known as FeedKind® (Alloul et al. [2022](#page-26-3)). Very recently, Gulf Biotech Qatar, and Unibio, have partnered to establish a production facility in Qatar, the gulf's frst natural gas to protein plant, to produce initially 6,000 tonnes of UniProtein® as a feed for aquaculture livestock (Unibio [2022](#page-33-0)). While several companies are working on natural-methane-fed and other edible food substrates MP production worldwide, yet, the production processes are expensive and not environment friendly due to the use of fossil and edible resources (Kalyuzhnaya et al. [2013\)](#page-29-4). Therefore, cheaper and sustainable substrates are required resulting in the bacterial protein production roar.

Current research on MP has concentrated on the valorization of organic waste materials by using high protein content containing microorganisms including fungus, yeast, algae, and bacteria, which can then be used in animal feed (Chandra et al. [2021](#page-27-3); Hülsen et al. [2022](#page-29-5)a; Xu et al. [2020;](#page-34-2) Yang et al. [2022;](#page-34-0) Zhu et al. [2022\)](#page-35-2). Fig. [1](#page-2-0) presents a schematic of MP production from agroindustrial waste using diferent microorganisms. Microorganisms use inexpensive and plentiful agro-waste for metabolism and to produce biomass, which may decrease the environmental impact. Start-up company eniferBio has recently updated the PEKILO® process using diferent industrial by-products as a substrate to produce the MP biomass validated for aquafeed and aiming to extend it further for human consumption (eniferBio [2022\)](#page-28-2). The study of optimal fermentation conditions, cheap substrates, and various microorganisms is on the rise. The objective of this review is to perform a comprehensive analysis of the applications and research being carried out in the feld of MP production from waste substrates for animal feed production in one place. The potential waste substrates for MP production, suitable microorganisms, bioprocesses, and production technology



tion of microbial protein production from agroindustrial waste using diferent microbes including bacteria, fungi, yeast, algae, and phototrophic bacteria in the context of circular economy and sustainability

<span id="page-2-0"></span>**Fig. 1** Schematic depic-

advances for MP production have been explored and discussed in detail. Finally, the MP application as animal feed, its challenges, and future perspectives in research in MP have been evaluated.

# **2 Agriculture and food waste as feedstock for MP production**

Carbon and nitrogen are required with proper carbonto-nitrogen ratios for the growth of microorganisms and MP production (Ugalde and Castrillo [2002](#page-33-1)). Nitrogen from diferent sources in form of ammonia, ammonium salt, nitrite, and/or nitrate is used by microbes. Typically 45–75% of the total MP production cost comes from the carbon feedstock and 7–15% of the total cost is from nitrogen sources (Nyyssölä et al. [2022\)](#page-31-2). In the case of algal MP, carbon in the atmosphere is free, however, the cost of agitation to dissolve it into dense algal culture is high (Wang et al. [2022;](#page-34-3) Xu et al. [2021b\)](#page-34-4). Globally a large number of wastes rich in organics and nutrients are produced from agriculture forms, food processing units, restaurants, supermarkets, and consumers including lignocellulose wastes such as corn, rice, and wheat (Capanoglu et al. [2022](#page-26-4); Chandra et al. [2021](#page-27-3); Rosenboom et al. [2022;](#page-32-3) Talan et al. [2022;](#page-33-2) Türker et al. [2022\)](#page-33-3). These waste materials are potential raw feedstock for the production of MP (Yang et al. [2022\)](#page-34-0). Industrial and agricultural wastes have high biological oxygen demand (BOD) which can cause signifcant environmental pollution (Alloul et al. [2019;](#page-26-5) Capson-Tojo et al. [2020](#page-27-4); Tropea et al. [2022](#page-33-4)). Although so far, commercially available MP is mostly produced from edible agriculture substrates and carbohydrate-derived materials with carbon as necessary constituents are usually employed for this objective, the current focus is on utilizing waste materials for MP synthesis for a sustainable environment and on making the process cost-efective (Thiviya et al. [2022b;](#page-33-5) Türker et al. [2022](#page-33-3); Yang et al. [2022](#page-34-0); Zeng et al. [2023](#page-35-4)). Waste feedstock selection is done based on their availability in the vicinity of the production site. A variety of agriculture and food-oriented solid wastes including rice straw, corncob starch waste, wheat bran, banana peel, pineapple waste, watermelon, potato bagasse, fermented grass, tofu and cheese wastes, sugarcane bagasse, banana, coconut, grape, and mango waste have been studied and reported as a feedstock for MP production (Asiri and Chu [2022](#page-26-1); Chandra et al. [2021;](#page-27-3) Farhan et al. [2021;](#page-28-3) Kurcz et al. [2018;](#page-30-0) Nascimento et al. [2022;](#page-27-5) Sakarika et al. [2022;](#page-32-4) Thiviya et al. [2022a;](#page-33-6) Thiviya et al. [2022b;](#page-33-5) Tropea et al. [2022](#page-33-4); Voutilainen et al. [2021;](#page-34-5) Zhu et al. [2022\)](#page-35-2). Several liquid substrates are also reported for MP production such as waste milk, non-dairy creamer wastewater, sugar beet pulp, cheese whey, sugar refnery wastewater, pineapple peel juice, pumpkin, farm manure, biogas slurry, rice washing water, latex rubber sheet wastewater, olive mill wastewater, sugarcane molasses, soybean molasses, food waste-derived volatile fatty acids, and municipal wastewater treat-ment effluent (Acosta et al. [2020;](#page-25-2) Bertasini et al. [2022;](#page-26-6) Cao et al. [2021;](#page-26-7) Ding et al. [2023;](#page-27-2) Pillaca-Pullo et al. [2023;](#page-32-5) Yang et al. [2017](#page-34-6); Zha et al. [2021](#page-35-5)).

Annually millions of tons of agricultural and food waste is generated globally and its inadequate management and dumping pose adverse impacts on the environment and are detrimental to the ecosystem (Nyyssölä et al. [2022\)](#page-31-2). Consequently, sustainable and environment-friendly conversion of organic and nutrient-rich waste to valuable products has become an important objective worldwide (Chandra et al. [2021;](#page-27-3) Dey et al. [2021](#page-27-0); Nyyssölä et al. [2022](#page-31-2)). With proper management and processing, agro-food waste can play a vital role in the sustainability of the ecosystem and energy security. Overall, the choice of waste feedstock for MP production depends on factors such as availability, cost, and suitability for the production process (Awad Saad Allah [2021;](#page-26-8) Capanoglu et al. [2022](#page-26-4); Kumar et al. [2021](#page-30-1); Leite et al. [2021;](#page-30-2) Reihani and Khosravi-Darani [2019\)](#page-32-6). It is important to consider the potential impacts of using agricultural waste feedstocks for MP production on the production process and the environmental impacts of the production process itself. Table [1](#page-4-0) presents a list of the most frequently reported agro-food wastes used for MP production along with the type of microbes used. Diferent waste substrates that can be used for MP production have their advantages and disadvantages. For instance, wheat bran, sugarcane bagasse, and sugar beet pulp are the byproduct of wheat processing and sugar processing, respectively, and can be used as a feedstock for MP production (Aker and Robinson [1987;](#page-25-3) Puligundla and Mok [2021;](#page-32-7) Saejung and Salasook [2020](#page-32-8); Yunus et al. [2015](#page-34-7)). These waste feedstocks are relatively inexpensive and abundant, and their use does not compete with their use as a

<span id="page-4-0"></span>



food or feed crop. However, their low protein content may limit their suitability for some MP production processes. Similarly, corn stover and rice straw are byproducts of harvesting that can be used as feedstock for MP production (Voutilainen et al. [2021](#page-34-5)). Both are relatively abundant and low-cost feedstock. For efective biomass valorization into MP, biodegradation and depolymerization of lignin is required which can be done only by selected microbes capable of producing lignin-degrading enzymes such as *Bacillus sp. LD2*, *Aneurinibacillus sp*. and *Trichoderma harzianum (*Sharma et al. [2022](#page-33-7)*)*. It is important to consider the potential impacts of using agricultural wastes for MP production on waste reduction and environmental

sustainability, as well as the quality and safety of the resulting MP product.

The use of agro-food waste for MP production must comply with relevant regulatory standards, including those related to food and feed safety, environmental protection, and animal feed which depend on several factors, including the type of waste, the source of waste, and the intended use of the MP (Janssen et al. [2022](#page-29-10); Nyyssölä et al. [2022](#page-31-2); Vethathirri et al. [2021;](#page-34-12) Voutilainen et al. [2021](#page-34-5); Wadhwa and Bakshi [2016\)](#page-34-10). Regulatory standards may difer between countries or regions, and it is important to consult local regulations and guidelines to determine the suitability of specifc types of agricultural waste. For instance, in the United States, the Food and Drug Administration (FDA) regulates animal feed and sets standards for animal food ingredients, including MP (Jonaitis et al. [2022\)](#page-29-11). The FDA generally permits the use of MP in animal feed, but it must meet certain requirements for safety and nutrition. In Europe, MP production from organic waste for animal feed must comply with regulations set forth by the European Union (EU)'s regulations which include the General Food Law Regulation (EC) No. 178/2002, which sets out the general principles of food safety, and the Novel Food Regulation (EC) No. 2015/2283, which establishes requirements for novel foods, including MP (EFSA Panel on Dietetic Products et al. [2016](#page-27-9); EFSA Panel on Nutrition et al. [2019;](#page-27-10) Lähteenmäki-Uutela et al. [2021\)](#page-30-5). Similarly in Japan, MP production from agro-food waste and municipal waste must comply with regulations set forth by the Ministry of Agriculture, Forestry, and Fisheries (MAFF) and the Ministry of Health, Labour, and Welfare (MHLW). MAFF regulates animal feed, including MP, and requires that it be manufactured following Good Feed Manufacturing Practices (GFMP) (Kondo and Taguchi [2022](#page-29-12)). MHLW regulates food safety and requires that MP intended for human consumption meet certain safety standards. Globally, the fnal product MP as animal feed should be tested for purity, safety, and nutritional value, and it should be labeled and marketed according to applicable regulations and guidelines (Pereira et al. [2022\)](#page-32-10). Overall, the suitability of agricultural waste for animal feed production will depend on a range of factors, and compliance with regulatory standards will be essential to ensure the safety and quality of the feed.

### **3 Microorganisms for MP production**

A wide variety of microorganisms have shown the potential to produce MP including heterotrophic bacteria, fungi, microalgae, chemoautotrophs, and methylotrophs (Nyyssölä et al. [2022](#page-31-2)). A detailed description of the microbes used for protein-rich biomass production is provided below.

## 3.1 Yeast and fungi

Yeast and fungi have been exclusively used to carry out the commercial-scale industrial synthesis of MP in the past due to their promising balanced amino acid profle making it a complete protein source. Yeast has been used historically as animal and human food and is an aspiring MP candidate (Nyyssölä et al. [2022\)](#page-31-2). Many fungal species are being used as MP and are available in the market with commercial brand names. For instance, QuornTM extracted from the flamentous fungus *F. venenatum* was launched decades ago by Marlow Foods (UK) and is utilized for human consumption extensively (Wiebe [2004](#page-34-13)). The fungal MPs have normally a balanced composition of lipids, protein, fber, and amino acids meeting the FAO guidelines for food supplements (Groenewald et al. [2014\)](#page-28-7). Yeast and fungal MP can be applied to not only enhance the nutritional quality of food products but can also improve the functional properties such as texture, and emulsifying capability (Barzee et al. [2021;](#page-26-10) Sharif et al. [2021](#page-32-2); Wiebe [2004](#page-34-13)). Nevertheless, there is a need to consider the possible production of mycotoxins while working with a few species like *Fusarium* and *Aspergillus (*Barzee et al. [2021](#page-26-10)*)*. Furthermore, yeast MP can be utilized as probiotics, has higher vitamin B. composition, and comprises relatively lower nucleic acid (5–12%) as compared to bacterial MP (8–14%) which reduces health dangers and limits the downstream treatment cost (Alkalbani et al. [2022](#page-26-11); Sen and Mansell [2020;](#page-32-11) Yao et al. [2020](#page-34-14)). *Saccharomyces cerevisiae* (*S. cerevisiae)* species have been extensively investigated for diferent applications, including MP production (Abdelwahab et al. [2020;](#page-25-4) Dunuweera et al. [2021](#page-27-11); Gunun et al. [2022](#page-28-8); Li et al. [2022](#page-30-6)a; Sen and Mansell [2020;](#page-32-11) Tropea et al. [2022\)](#page-33-4). *S. cerevisiae,* also known as Brewer's yeast, is traditionally used for the production of yeast extracts (Dunuweera et al. [2021](#page-27-11); Farhan et al. [2021](#page-28-3)). *Yarrowia lipolytica* (*Y. lipolytica)* is another yeast specie that is being applied in the biotechnology industry owing to its ability to utilize numerous carbon substrates and synthesize MP with high-quality lipids (Groenewald et al. [2014](#page-28-7); Li et al. [2022](#page-30-6)a; Yang et al. [2022](#page-34-0)). European Food Safety Authority (EFSA), consequently, granted and accepted *Y. lipolytica* MP as a food as requested by the European Commission (EFSA Panel on Nutrition et al. [2019](#page-27-10)). The flamentous fungus *Paecilomyces varioti* has long been utilized for a registered PEKILO® process to produce MP for animal feed production (eniferBio [2022](#page-28-2)). *Aspergillus oryzae* (*A.oryzae*) is another flamentous fungus that is most studied and applied at the industrial scale, particularly in Asian countries like Korea, Japan, and China for the production of several fermented products (Ferreira et al. [2016](#page-28-9)). Yeast *Candida utilis* (*C. utilis*) has been approved as safe food and feed supplement by the China Food and Drug Administration and is included in the Generally Recognized as Safe (GRAS) list by the United States (Kurcz et al. [2018](#page-30-0)). *C. utilis* is currently being studied a lot to produce MP for safe and robust production of MP and has shown a great perspective for the synthesis of edible MP for human and animal consumption because of its higher yield, protein contents, growth rate, and capability to utilize diferent substrates as nutrients source (Carranza-Méndez et al. [2022](#page-27-12); Ding et al. [2023;](#page-27-2) Kurcz et al. [2018;](#page-30-0) Li et al. [2022a](#page-30-6); Yang et al. [2021c](#page-34-15)). *C. utilis* yeasts have a vitamin B complex and can produce a steroid compound called ergosterol which can immediately be converted to vitamin  $D_2$ (Dunuweera et al. [2021\)](#page-27-11). *Candida spp.* are among the most studied microbes for MP production because of their high growth rates and low energy metabolism at room temperature. *C. sorboxylosa, C. Lipolytica,* and *C. parapsilosis* have been recently reported to produce MP using diferent waste materials as feedstock (Coimbra et al. [2021](#page-27-13); Pillaca-Pullo et al. [2023](#page-32-5); Rages et al. [2021\)](#page-32-12). Several other yeast strains are under investigation and have shown potential to produce MP for dietary applications such as *Galactomyces geotrichum*, *Candida tropicalis, Debaryomyces hansenii, Pichia guilliermondii, Pichia kudriavzevii, Wickerhamomyces anomalus, Pichia jadinii, Nectaromyces rattus (*Dias et al. [2021](#page-27-14)*;* Su et al. [2021](#page-33-9)*;* Zhang et al. [2021](#page-35-6)*;* Zhou et al. [2022a](#page-35-1)*).* Hashem et. al. has recently investigated few non-conventional yeast strains like *Hanseniaspora guilliermondii*, *Hanseniaspora uvarum*, *Issatchenkia orientalis*, and *Cyberlindnera*  *fabianii* for MP production and reported that these newly isolated strains could be promising candidate for MP synthesis (Hashem et al. [2022](#page-28-10)).

Furthermore, the interest in the use of mixed yeast cultures for MP production is growing (Areniello et al. [2023](#page-26-12)). The use of mixed culture is advantageous due to the wider range of hydrolytic activities performed by the diverse culture, enhancing the utilization of complex substrates. Such as lignocellulosic raw materials require several enzymes for their efficient hydrolysis and a mixed fungal culture can address this by producing diferent enzymes (Salazar-López et al. [2022](#page-32-13)). Moreover, the metabolic products of one species may work synergistically by catabolizing various feedstocks (Vethathirri et al. [2021](#page-34-12)). Mixed microbial cultures have been reported to increase the nutritional content, by balancing the composition of proteins, vitamins, and lipids. However, controlling and optimizing the operational parameters for MP production in mixed culture may be more complicated and needs careful design and biochemical reaction control (Hashempour-Baltork et al. [2022](#page-28-6)). Moreover, the production of toxic secondary metabolites and process inhibitors is another challenge that needs to be addressed in the mixed culture production process (Nyyssölä et al. [2022](#page-31-2)).

#### 3.2 Microalgae and photosynthetic bacteria

Microalgae are considered a great source of MP due to their high protein and amino acid contents. Microalgae can utilize various waste substrates as nutrient sources to produce MP as an alternative to soy feed for protein-rich animal feed (Janssen et al. [2022](#page-29-10)). Some of the studies of algae biomass for MP production include *Chlorella (C.) Vulgaris*, *C. pyrenoidosa, C. luteoviris, C. robusta, Arthrospira (A.) maxima, A. platensis*, *Tetraselmis chui, Odontella aurita, Nannochloropsis oceanica, Nannochloropsis gaditana, Dunaliella salina*, *Euglena gracilis* and *Galdieria sulphuraria* (Abiusi et al. [2022;](#page-25-5) Muys et al. [2019](#page-31-7); Nascimento et al. [2022;](#page-27-5) Sui and Vlaeminck [2020\)](#page-33-10). Microalgal biomass protein content ranges from 30–80%, for diferent strains (Sui and Vlaeminck [2020\)](#page-33-10). Microalgae can fx carbon dioxide through photosynthesis (Almomani et al. [2019b\)](#page-26-13). Microalgae like *Scenedesmus obliquus* (*S. obliquus*) have the potential to fix higher concentrations of  $CO<sub>2</sub>$  as shown in a recent study (Molitor et al. [2019\)](#page-31-8). In this study, the growth rate of *S. obliquus* at  $2.5\%$  CO<sub>2</sub> surpassed all earlier reported values and growth was not too much afected even at  $35\%$  CO<sub>2</sub> with amino acid content comparable to that of soy. Cyanobacteria also known as blue-green algae are usually categorized together with microalgae due to their photosynthetic ability and phenotype. *Spirulina* and *Chlorella* species are the most famous cyanobacteria having protein contents of up to 50–70% and are currently commercially marketed as human and animal food. Presently, the projected production of *Chlorella* and *Spirulina* is about 6600 and 12,000 tons *d.m*/year worldwide, respectively (García et al. [2017\)](#page-28-11). Cyanobacteria are not rich only in protein but also have the valuable composition of other essential food elements such as small peptides,  $B_{12}$ ,  $B_{2}$ ,  $B_{1}$ ,  $B_{3}$ , E-vitamins, lipids, and pigments. Nevertheless, *Spirulina* MP has lower concentrations of amino acids like cysteine, methionine, and lysine which make it not a favorable protein for human food (Abiusi et al. [2022;](#page-25-5) Muys et al. [2019](#page-31-7)). Because of this, most of the current commercial companies' focus is to use *Spirulina* as a feed product with advantageous features for animal feed and aquaculture (Vethathirri et al. [2021\)](#page-34-12).

Photosynthetic bacteria (PSB) are another group of protein-rich microorganisms that have shown the potential to produce MP with a variety of metabolic abilities (Zhu et al. [2022\)](#page-35-2). PSB can not only produce MP but also other high-quality products including hydrogen and biodiesel (Li et al. [2019\)](#page-30-7). Some of the most studies of PSB biomass for MP production include *Rhodobacter capsulatus*, *Ectothiorhodospira*, *Rhodopseudomonas faecalis*, *Rhodobacter sphaeroides, Rhodopseudomonas palustris (*Cao et al. [2021;](#page-26-7) Deseure et al. [2021;](#page-27-15) Saejung and Chanthakhot [2021;](#page-32-14) Saejung and Sanusan [2021](#page-32-15); Yu et al. [2021,](#page-34-16) [2022](#page-34-17)*)*. Although there are many types of PSB microbes based on the metabolism and operational parameters requirements, it comprises four key families including purple non-sulfur bacteria (PNSB) such as *Rhodospirillaceae*, purple sulfur bacteria (PB) like *Chromatiaceae,* green sulfur bacteria (GSB) like *Chlorobiaceae* and gliding flamentous green sulfur bacteria (GFB) such as *Chlorofexaceae (*Lu et al. [2019](#page-30-8)*)*. PSB has demonstrated signifcant MP production efficiency with high protein productivity (about 150 mg/L·d) (Cao et al. [2021\)](#page-26-7). PSB has shown potential as an additive for the synthesis of value added-products because of the presence of several important components like coenzyme Q10, nicotinic acid, pantothenic acid, 5-aminolevulinic acid, carotenoids, and pigments like carotenoid and bacteriochlorin. These pigments can produce red, purple, or orange colors and can be used as natural color additives for dairy and bakery products (Lu et al. [2019](#page-30-8)).

#### 3.3 Chemoautotrophs and methylotrophs

Although various bacterial species are being studied currently for MP production by heterotrophic bacteria by fermentation, commercial production of bacterial MP has mainly focused on the use of chemoautotrophs and methanotrophic species using gaseous substrates for growth (Martin et al. [2013;](#page-31-9) Sakarika et al. [2022;](#page-32-4) Woolley et al. [2023\)](#page-34-1). Methanotrophic species oxidize methane to formaldehyde through ribulose monophosphate or serine pathways. MP production by methanotrophs was long ago started on a commercial scale in the late 1970s nevertheless manufacturing was stopped due to economic issues. However, methanotrophic MP has again come to attention for the production of microbial feed, and currently, various commercial products are marketed with brand names like KnipBio (USA), UniProtein® (Unibio, Denmark), and FeedKind® (Calysta, USA) (Khoshnevisan et al. [2022;](#page-29-3) Sakarika et al. [2022](#page-32-4); Woolley et al. [2023](#page-34-1); Xu et al. [2021b\)](#page-34-4). KnipBio is the frst genetically modifed MP product that has achieved GRAS status from the United States Food and Drug Administration (Nyyssölä et al. [2022](#page-31-2)). Several methanotrophic microbes are prevalently reported for MP synthesis from biogas including *Methylococcales, Methylococcus capsulatus, Methylocystis, Methylophilus, Rurimicrobium, Comamonadaceae, Methylophilale, Methylomicrobium buryatense, Methylocystis parvus, Methylomonas, Methylocapsa acidiphila, Methylomonas methanica*, and *Methylomicrobium alcaliphilum (*Jintasataporn et al. [2021;](#page-29-13) Woolley et al. [2023;](#page-34-1) Xu et al. [2021a](#page-34-18), [2020;](#page-34-2) Zha et al. [2021](#page-35-5)*)*. Moreover, several methanotrophs such as *Methyloferula, Methylococcus*, and *Methylocaldum* can fix  $CO<sub>2</sub>$ , making them advantageous to grow on biogas utilizing both  $CH_4$  and  $CO_2$  (Kim et al. [2022;](#page-29-14) Kulkarni et al. [2021;](#page-29-15) Salehi and Chaiprapat [2022](#page-32-16)). Autotrophically growing hydrogen oxidizing bacteria (HOB) also known as Knallgas bacteria energize their metabolism using hydrogen as the electron donor in presence of oxygen while fixing  $CO<sub>2</sub>$  to biomass (Jiang et al. [2022\)](#page-29-2). The common HOB employed for biogas-based MP production are *Alcaligenes eutrophus Z, Hydrogenomonas, Pseudomonas, Aquaspirillum, Paracoccus denitrifcans Y5, Ralstonia eutropha B5786*, *Cupriavidus necator* H16, and *Paracoccus versutus D6* (Jiang et al. [2022](#page-29-2)). Hydrogen is oxidized both by aerobic and anaerobic bacteria. In anaerobic oxidation,  $CO<sub>2</sub>$  is fixed through the reductive acetyl-CoA pathway, in which carbon is mainly converted to organic co-products such as ethanol and acetate rather than cell biomass. Whereas, in aerobic hydrogen oxidation, carbon dioxide is assimilated through the reverse tricarboxylic acid cycle to produce the cell mass (Muñoz et al. [2015;](#page-31-10) Pander et al. [2020;](#page-32-17) Sakarika et al. [2022](#page-32-4)). Therefore, the current industrial process is focused on the synthesis of value-added chemicals such as ethanol and acetate instead of cell biomass for MP production (Pander et al. [2020\)](#page-32-17). A two-step process has been suggested for MP production by acetogens in which frstly carbon dioxide is reduced to acetate by acetogenic bacteria, followed by the cultivation of heterotrophic yeast or fungal species using produced acetate as the carbon source (Bolognesi et al. [2022\)](#page-26-14).

#### **4 MP production processes**

MP is generally produced by the fermentation process using particular microorganisms cultivated on appropriate substrates (Ding et al. [2023](#page-27-2)). For this purpose, suitable microorganisms are isolated from diferent sources including water, air, and soil samples, and then growth conditions are optimized for maximum product yields (Kurcz et al. [2018\)](#page-30-0). A detail description is provided below.

4.1 Direct feedstock fermentation by bacteria and yeast

The fermentation process can be divided into submerged, semisolid, and solid-state fermentation. In the submerged fermentation process, the substrate with the required nutrients for microbial growth is always in liquid form (Ding et al. [2023\)](#page-27-2). Submerged fermentation can be carried out in batch mode, fedbatch reactor, and/or in a continuous operating reactor with continuous harvesting of the biomass followed by separation and biomass drying. The fed-batch fermentation process is advantageous as compared to the batch system, for instance, it ofers a higher biomass growth rate which can increase the fermentation performance. However, it is important to optimize the different factors affecting the fermentation efficiency, such as pH, temperature, oxygen level, nutrients concentration (*e.g.* nitrogen and phosphorus), and C/N ratios (Ding et al. [2023](#page-27-2); Gervasi et al. [2018;](#page-28-1) Nyyssölä et al. [2022](#page-31-2); Ugalde and Castrillo [2002;](#page-33-1) Zeng et al. [2022\)](#page-35-0). Heat is generated in this process and aeration is continuously performed to properly cool down the bioreactor system. So usually, the fermenter setup is provided with sensors and regulators for agitation, pH, dissolved oxygen, and temperature. Several organic wastes rich in nutrients are being explored for fungal and yeast biomass production. *Candida utilis* MP was produced using biogas slurry as ammonia nitrogen and acetate, lactate, and sugar as carbon sources with maximum biomass production of 14.8 g dried biomass/L containing 46.5% protein content. The C/N ratio showed a signifcant impact on the biomass yield where the increase in C/N ratio from 3 to 15, resulted in the reduction of biomass yield depicting that a controlled C/N ratio of 3, is benefcial (Ding et al. [2023\)](#page-27-2). Controlling the pH is an important factor as it can afect biomass growth by changing enzyme action, and membrane permeability. However, diferent yeast strains have diferent pH ranges for optimal microbial growth. For instance, *C. utilis* can grow in a wide-ranging pH of 3.5–8.0 (Ding et al. [2023\)](#page-27-2), whereas, the optimum pH for *Galactomyces candidum* and *Nectaromyces rattus* growth is pH 5.5 (Zhou et al. [2022a](#page-35-1)).

Another factor that can signifcantly impact MP production is phosphorus concentration. Phosphorus is an essential nutrient for microorganisms, and it plays a crucial role in various cellular processes, including energy metabolism, nucleic acid, and protein synthesis. However, excess phosphorus in the growth medium can lead to the formation of insoluble phosphates, which can reduce the availability of phosphorus for microbial growth and protein synthesis (Goonesekera et al. [2022](#page-28-12)). This can negatively afect the yield and quality of the MP produced. On the other hand, inadequate phosphorus levels can also limit MP production by slowing down cell growth and reducing protein synthesis (Goonesekera et al. [2022;](#page-28-12) Quan et al. [2001;](#page-32-18) Stern and Hoover [1979;](#page-33-11) Zhu et al. [2020\)](#page-35-7). Therefore, it is crucial to maintain an optimal phosphorus concentration in the growth medium to maximize MP production. Furthermore, excess phosphorus can lead to environmental pollution, as it can be discharged into water bodies, causing eutrophication and algal blooms (Van Heyst et al. [2022;](#page-33-12) Zhou et al. [2022b\)](#page-35-8). Therefore, it is essential to develop sustainable methods for MP production that take into account the optimal use of nutrients and minimize environmental impacts.

The digestate from anaerobic digestion (AD) of numerous organic wastes is usually rich in acetate and  $NH<sub>4</sub>-N$  and in a recent study pretreated digestate was investigated as an alternate substrate to produce fungal/yeast MP by fermentation (Zeng et al. [2023](#page-35-4)). A hybrid electrochemical-membrane fermentation process was employed for the recovery of ammonium and acetate from digestate, followed by fermentation processes using *S. cerevisiae* yeast. Acetate and ammonium as feedstock were recovered by electrodialysis (ED) and fed to *S. cerevisiae* for MP productions of about  $0.76-0.86$  g/L (Fig. [2\)](#page-10-0). The separation process to recover the biomass is chosen based on the type of microbes used for MP production. For instance, flamentous fungi are usually separated by fltration whereas bacteria are collected by the centrifugation process. Moreover, it is important to recover the maximum quantity of nutrient-rich water contents which can be obtained after drying.

In the semisolid fermentation process, the substrate is used in the solid state. The MP production through this process involves the selection of suitable media preparation with an explicit carbon source, accurate media decontamination, careful selection of microbial strain, biomass separation, and fnal product processing (Ritala et al. [2017](#page-32-0); Thiviya et al. [2022b](#page-33-5)). Various substrates as a carbon source are employed for this objective such as diferent fruit and vegetable wastes, food industries waste, carbon oxide, polysaccharides, several gaseous hydrocarbons, ethanol, methanol, effluents of different industrial effluents including breweries and other solid organic waste materials (Areniello et al. [2022;](#page-26-12) Puligundla and Mok [2021;](#page-32-7) Thiviya et al. [2022b](#page-33-5); Zhang et al. [2018](#page-35-9)). This type of fermentation requires a specifc type of bioreactor carefully designed to identify mass and energy conversion and transportation (Selvaraj et al. [2021](#page-32-19); Sharif et al. [2021](#page-32-2)). Biomass cultivation involves several operations such as multiphase mixing using stirring, oxygen transport to microbes from the gaseous bubble phase to the liquid phase, and eventually heat transfer from the aqueous phase to the environs (Molfetta et al. [2022;](#page-31-11) Sharif et al. [2021](#page-32-2); Śliżewska and Chlebicz-Wójcik [2020\)](#page-33-13). This fermentation process needs higher initial capital investment and operating costs.

While fungal MP is usually synthesized in submerged fermentations there is a growing focus on employing solid-state fermentation to provide physical support for the microbial culture and deliver nutrients for the production of various value-added products (Cerda et al. [2019](#page-27-16); Kumar et al. [2021](#page-30-1); Leite et al. [2021](#page-30-2); Melnichuk et al. [2020;](#page-31-12) Sharif et al. [2021;](#page-32-2) Vauris et al. [2022\)](#page-34-19). Solid-state fermentation offers the advantages of higher nutritional value, less effluent generation, and lower energy costs (Kumar et al. [2021\)](#page-30-1). MP production in solid-state fermentation can be carried out using various microbes under diferent environmental conditions in a variety of reactor designs (Godoy et al. [2018;](#page-28-13) Vauris et al. [2022](#page-34-19)). The process is not only suitable for the synthesis of MP but also capable of producing various other products such as ethanol, enzymes, organic acids, peptides, several vitamins, and favors (Aita et al. [2019;](#page-25-6) Cerda et al. [2019;](#page-27-16) Godoy et al. [2018;](#page-28-13) Hashempour-Baltork et al. [2022](#page-28-6); Li et al. [2022a](#page-30-6); Liu et al. [2018](#page-30-9); Melnichuk et al. [2020](#page-31-12); Vauris et al. [2022](#page-34-19); Zhang et al. [2018\)](#page-35-9). As evident by the name, the process needs feedstock in pure solid form such as agriculture form waste like wheat bran and/or rice bran, orange pulp, molasses, brewer's spent grain, poultry litter; and waste capsicum powder, etc*.* (Aita et al. [2019;](#page-25-6) Olukomaiya et al. [2019;](#page-31-13) Sharif et al. [2021](#page-32-2)). A feedstock is inoculated with the selected microbes and spread on the fatbeds with controlled operating parameters like proper moisture level (60–65%), continuous oxygen supply, controlled temperature, essential nutrient concentration, and pH (Aita et al. [2019](#page-25-6); Kumar et al. [2021](#page-30-1)). Though, the formation of unexpected toxic secondary byproducts poses a signifcant risk in using such complex waste materials. Therefore, the recent research focus is to develop novel synthesis processes to improve the utilization efficiency of waste byproducts from further complex waste substrates. In this effort, an integrated approach to employ both solid-state and submerged fermentations have been proposed (Khonngam and Salakkam [2019;](#page-29-16) Liu et al. [2020;](#page-30-10)



<span id="page-10-0"></span>**Fig. 2 A** Graphical depiction of acetate and ammonium recovery from anaerobic digestate for MP synthesis by an integrated electrochemical-membrane fermentation system. **B** Acetate

and **C** ammonia recovery, **D** & **E** yeast biomass growth using integrated process (Reproduced from (Zeng et al.) with permission from Elsevier 2023)

Martău et al. [2021;](#page-31-14) Olukomaiya et al. [2019;](#page-31-13) Premalatha et al.; Villegas-Méndez et al. [2022](#page-34-20)).

Substrate composition, concentration, presence of anions and cations, and operational parameters like reaction time, dissolved oxygen, pH, and temperature strongly infuence MP production through the fermentation process. The substrate cost for bioprocessing makes up half of the total production cost (Padoan et al. [2022](#page-31-15); Zeng et al. [2022\)](#page-35-0). Current research on MP production is focused on fnding less expensive feedstock at large-scale. The cost of the substrate and strict sterilization are two major issues when utilizing pure culture fermentation. Currently, research on the valorization of agriculture and FW is gaining growing interest, and researchers all over the world are working on it by proposing numerous strategies to reuse food waste (Zeng et al. [2022\)](#page-35-0). Organic waste components from the agriculture and food industry have been recognized as low-cost feedstock, to produce several biologically driven value-added products by fermentation. Agriculture residues including rice and wheat straw, cassava waste, orange peel, sugarcane, paper mill waste, sugar industry wastewater (Saejung and Salasook [2020\)](#page-32-8), sawdust, corn cobs, sugar beet pulp, coconut waste, grape waste, mango waste, etc. have been studied as substrates for MP production employing various microorganisms (Cao et al.  $2021$ ). Sugar industry effluents are highly polluted and generated from diferent processes including sugarcane washing, crushing, evaporation, crystallization, molasses preparation, and end products refning. Sugar-industry wastewater is largely composed of organic carbon with high chemical oxygen demand (COD) and therefore can be a good feedstock for MP production by cultivating microbes (Kushwaha [2015\)](#page-30-11). Chewapat *et. al.* studied MP production using sugar industry wastewater as a carbon source and reported *R. faecalis* biomass with more than 50% protein contents which are suitable to be used as animal feed while simultaneously reducing 80% of the COD (Saejung and Salasook [2020](#page-32-8)). The experimental setup involved batch cultivation of *R. faecalis* in a 3 L photo-bioreactor using sugar industry waste as a co-substrate. The biomass produced was reported to have high concentrations of essential amino acids like leucine and lysine.

Waste generated from agroindustry and some other waste streams are rich in short-chain and longchain organic acids, however, the utilization of these organic acids by diferent microbes to produce MP is still not well understood. There is a need to understand the biomass composition and protein contents of the MP produced by waste feeds tock rich in organic acids content. Recently, Zeng et al. employed diferent organic acids to produce S. cerevisiae MP production and reported 0.94 g/L MP at 20 g/L of acetate COD (Zeng et al. [2022\)](#page-35-0). *S. cerevisiae* biomass and protein concentration were found to increase with increasing acetate concentration. Moreover, acetic acid and lactic acid were described as suitable shortchain organic acids for *S. cerevisiae* MP production. In another set of experiments, it was found that a mixture of acetate with lactate, oleate, or linoleate enhanced MP production. Long-chain fatty acids linoleate and oleate are commonly found in the anaerobic digestate of food waste and can be a good cosubstrate for MP production as shown in this study.

While these waste materials are excellent raw feedstock for the synthesis of MP, there is a need to pretreat it to improve its physicochemical and biological characteristics (Chandra et al. [2021;](#page-27-3) Dey et al. [2021;](#page-27-0) Hashem et al. [2022;](#page-28-10) Hashempour-Baltork et al. [2022;](#page-28-6) Ugalde and Castrillo [2002](#page-33-1); Yang et al. [2022](#page-34-0)). Several physicochemical and biological processes are employed for the treatment. It is important to fnd out the optimal conversion process that can liberate maximum monomers from the organic waste to get the highest availability of secondary metabolites like carbon sources, nutrients, phenols, starch and cellulose, proteins, and lipids (Godoy et al. [2018;](#page-28-13) Hashempour-Baltork et al. [2022](#page-28-6); Sakarika et al. [2022](#page-32-4); Suriyapha et al. [2020](#page-33-14); Vauris et al. [2022](#page-34-19); Yang et al. [2022](#page-34-0); Zeng et al. [2023](#page-35-4)). These compounds are the secondary substrate for enzymatic hydrolysis and fermentation in the later stages of the MP production process. To achieve this, several approaches are being applied including combined physical/chemical and biological treatment of the substrate (Khonngam and Salakkam [2019;](#page-29-16) Li et al. [2022](#page-30-6)a; Newman et al. [2023](#page-31-1); Xu et al. [2021a\)](#page-34-18). Mechanical grinding, microwave, steam explosion, and hot water treatments are the method being used for physical pretreatments (Leite et al. [2021;](#page-30-2) Molfetta et al. [2022;](#page-31-11) Zhu et al. [2022\)](#page-35-2). Whereas chemicals including strong acids are used to hydrolyze the biomass structure in chemical treatment. Biological pretreatment is carried out by employing commercial enzymes and/or selected microbes. However, recently researchers are working to use organic waste substrates directly for fermentation without pretreatments to reduce the costs (Gervasi et al. [2018](#page-28-1)). For instance, Teresa et al*.* (Gervasi et al. [2018](#page-28-1)) used *S. cerevisiae* to produce MP by aerobic fermentation from mixed fruit and vegetable FW without any pretreatments. The MP produced from the process has a protein content of 39.8% which was higher than many earlier reported studies. Another approach to avoid costly pretreatment is to use the digestate obtained after the digestion of waste materials. The anaerobic digestate comprising organic acids like acetate, lactate, and other long-term VFAs can be directly consumed by several fungal and bacterial strains such as *S. cerevisiae* with little pretreatment (Zeng et al. [2022\)](#page-35-0).

#### 4.2 Phototrophic processes

Microalgae, photosynthetic bacteria (PSB), and cyanobacteria are grown in open ponds or photobioreactors under control environment and operational parameters (Capson-Tojo et al. [2020](#page-27-4); Janssen et al. [2022\)](#page-29-10). High-intensity light exposure is the major requirement to cultivate microalgae and cyanobacteria in any kind of production setup. Algae cultivation in open ponds is generally considered suitable and economical, however, it has some drawbacks including low productivity and potential contamination issues. The growth of microbes in a photobioreactor can be afected by several factors including carbon and nitrogen sources, temperature, pH value, the chemical composition of the feedstock, light intensity, oxygen saturation level, and hydraulic retention time (Alloul et al. [2019](#page-26-5); Almomani et al. [2019a](#page-26-15); Capson-Tojo et al. [2020;](#page-27-4) Laskowska et al. [2017\)](#page-30-12). The light-oxygen state is the most important operational parameter for phototrophic microbes. It is reported that a high  $NH_4^+$ <sub>-</sub>N concentration (2000 mg)  $NH_4^+$ <sub>-</sub>N/L) improved the protein contents of PSB to 65.0% because it supplied suitable nitrogen resources for cell growth and synthesis of protein (Yang et al. [2017\)](#page-34-6). Numerous agri-industrial wastewaters have been used for MP production in photobioreactors with reasonable protein contents (Alloul et al. [2019](#page-26-5); Capson-Tojo et al. [2020](#page-27-4); Hülsen et al. [2018a\)](#page-29-17). The objective of current research is to produce microalgae biomass with high protein contents and recently few studies have investigated microalgae biomass even in

extreme growing environments such as acidic pH 0 to 4 and temperature above 40 °C (Montenegro-Herrera et al. [2022](#page-31-16)). In another study, a combined microalgae *Chlorella vulgaris* and the methanotrophic system was investigated to convert biogas into MP (Wang et al. [2022\)](#page-34-3).

In laboratory-scale investigations, fasks and small glass reactors (transparent acrylic material) are typically used for microbial growth experiments with magnetic stirring, artifcial light, pumps for influent and effluent flows, pH, and thermometers for temperature monitoring (Liu et al. [2016;](#page-30-13) Meng et al. [2017;](#page-31-17) Yang et al. [2018\)](#page-34-21). Several photobioreactor designs and operations strategies are proposed to enhance biomass yields. Flat panel photobioreactors with transparent flat vessels for suspended biomass growth have been used for both PBS and microalgae cultivation (Carone et al. [2022;](#page-27-17) Chen et al. [2011;](#page-27-18) Gabrielyan et al. [2022;](#page-28-14) Hülsen et al. [2022b](#page-29-18); Lim et al. [2023](#page-30-14); Maia et al. [2022](#page-30-15); Ravi Kiran and Venkata Mohan [2022\)](#page-32-20). A fat-panel photobioreactor was tested for growing *Tetradesmus sp. SVMIICT4* algae biomass using diary wastewater as feedstock (Ravi Kiran and Venkata Mohan [2022](#page-32-20)). The bioreactor showed substantial carbon and nutrient utilization for biomass assimilation with protein contents of 19.52 mg/g. Figure [3A](#page-14-0) shows the graphical presentation of the fat panel photobioreactor utilized for the selected algae biomass growth. A pilot scale fat-panel vertical photobioreactor (70 L) has been employed for the cultivation of *Chlorella sorokiniana* algae biomass in semi-continuous mode with a biomass growth of 2.8 g dw/L (Fig. [3](#page-14-0)B and [C](#page-14-0)) (Gabrielyan et al. [2022](#page-28-14)). Microalgae-based biotechnological processes are gaining importance for direct  $CO<sub>2</sub>$  capture and highquality biomass production for numerous industrial applications (Montenegro-Herrera et al. [2022](#page-31-16)). To capture atmospheric  $CO<sub>2</sub>$  and improve microalgae growth,  $CO<sub>2</sub>$  gas with controlled bubble size is fed to photobioreactors (Carone et al. [2022;](#page-27-17) Lim et al. [2023\)](#page-30-14). Carone et. al. designed a new 1.3 cm thick alveolar flat panel photobioreactor to fix the  $CO<sub>2</sub>$  and produce algae biomass of *Acutodesmus obliquus* aiming to increase the gas-liquid mass transfer coefficient (Fig. [3](#page-14-0)D) (Carone et al. [2022](#page-27-17)). The designed reactor achieved  $64\%$  CO<sub>2</sub> fixation efficiency and biomass growth of 1.9 g dw/L under a controlled environment (Fig. [3E](#page-14-0)).



Bio-electrochemical systems (BESs) have been applied for various applications including water/ wastewater treatment and resource recovery. The application of BESs for microalgae growth has also gained attention in the last few years. The membranes provide an access to migrate the nutrients <span id="page-14-0"></span>**Fig. 3** Diferent fat panel photobioreactors design and per-◂formance for algae biomass production. **A** Graphical presentation of fat panel photobioreactor using dairy wastewater for algal biomass production (Reproduced from (Ravi Kiran and Venkata Mohan [2022](#page-32-20)) with permission from Elsevier 2023) **B** General scheme of A fat-panel vertical photobioreactor for Cultivation of *Chlorella sorokiniana* and **C** photo of the main reactor units consisting of a reservoir, bottom, lid, Suspension, LED module, LED, power supply, platform, temperature indicator & sensor, supply tube, and sprayer (Reproduced from (Gabrielyan et al. [2022](#page-28-14)) with permission from the MDPI 2022). **D** Process diagram of the fat panel photobioreactor with microbubble for CO<sub>2</sub> fixation **E** Algal biomass concentration in the fat panel photobioreactor (Reproduced from (Carone et al. [2022\)](#page-27-17) with permission from Elsevier 2022)

and can separate microalgae from aqueous phase solutions. BESs studies usually focus on single- and two-chamber microbial fuel cells and microbial electrochemical systems with high efficiency in organics utilization and nutrient recovery. Few researchers have investigated the use of the BESs system for protein-rich microalgae cultivation to produce MP (Pan et al. [2021](#page-31-18)). For instance, Pan et. al. reported the cultivation of *Chlorella vulgaris* in a three-chamber microalgal BES while using agro-industry wastewater for MP production and obtained a biomass growth of 0.87–1.11 g dw/L under an optimal operating environment. Similarly, several other studies have reported the potential of diferent BESs for microalgae growth with high protein contents (Bolognesi et al. [2022;](#page-26-14) Cevik et al. [2020;](#page-27-19) Elmaadawy et al. [2022](#page-28-15); Elshobary et al. [2021](#page-28-16); Jadhav et al. [2019;](#page-29-19) Sharma et al. [2020;](#page-32-21) Yang et al. [2021b](#page-34-22)).

Higher economic costs, light penetration, long microbial acclimatization period, and seasonal weather variations are among some of the obstacles inhibiting extensive MP generation by phototrophs from waste feedstock (Ayre et al. [2017\)](#page-26-16). Because of the recent developments in PBS growth technology, MP production by PSB can address some of the issues that come upon in phototrophic systems. PSB are photo heterotrophs, using organic carbon as an anabolic substrate and infrared (IR) light spectrum for energy generation (Hülsen et al. [2018b](#page-29-20)). PSB has a faster growth rate as compared to algae and can utilize nutrients and organics from waste feedstock, simultaneously (Capson-Tojo et al. [2020](#page-27-4)). PSB has a unique metabolism pathway as it can execute photosynthesis, aerobic, and anaerobic fermentation depending upon the growth environment. Generally, dark-aerobic and light-anaerobic environment are considered suitable light-oxygen conditions for PSB growth. PSB can be produced in small households or large plants by maintaining the temperature around  $23-39$  °C, pH range 6.0–9.0, and light at about  $100-200 \mu \text{mol/m}^2/\text{s}$ in the bioreactors constructed of transparent glass containers, plastic bags, and/or ponds. PSB is produced by two diferent processes. The frst is closed light-anaerobic cultivation mode and the second is known as the open light-micro aerobic process. In the anaerobic process, the medium is sterilized and inoculated with 20–50% PSB culture in the reactor containers where PSB is grown for 5–10 days under continuous stirring to keep the biomass fowing up to obtain light. In the open light-micro aerobic process, air stones are applied to provide a suitable micro aerobic environment under controlled light intensity, temperature, and pH like the anaerobic process. The oxygen concentration needs to be carefully controlled in this process as a higher oxygen level may impair the PSB photosynthesis leading to a decrease in biomass quality and quantity (Laskowska et al. [2017](#page-30-12)). The focus of most of the earlier studies was on the lab-scale operation to optimize the reactor design and operational parameters, however, there is a need to upscale the designed technology in an outdoor environment and demonstrate the technical performance in terms of biomass production and substrate utilization. In a recent study, a horizontal fat plate photobioreactor (10 m long) with  $0.95 \text{ m}^3$  volume was supplied with poultry processing wastewater for an extended period of 253 days under diferent operational schemes to grow PSB (Fig. [4](#page-15-0)A) (Hülsen et al. [2022b](#page-29-18)). The reactor was operated in microaerobic, and anaerobic modes and it was found that the anaerobic process showed higher performance in terms of PSB growth, whereas carbon and nutrient utilization was recorded maximum in anaerobic/aerobic integrated performance (Fig. [4](#page-15-0)B). The total relative abundance of PPB in the mixed microbial culture was over 56.0% when the reactor was operated under an optimal working environment (Fig. [4](#page-15-0)C).

PSB can also grow at low light intensities and can acclimatize to low temperatures under anaerobic conditions. PSB tolerance towards high ammonia concentration and capacity to grow in outdoor systems make PSB a potential microbe for agri-industrial waste to produce MP. Illumination is the critical factor that enhances the overall phototrophic system cost,



<span id="page-15-0"></span>**Fig. 4 A** Schematic depiction of the photobioreactor for PSB growth on poultry wastewater (top) and pictures of the bioreactor with and without the UV–VIS absorbing foil (bottom). **B** biomass production during the whole operation period **C** Microbial community structure in the bioreactor during the

prohibiting its use for MP production. Recent studies are addressing this factor by employing sunlight as an energy source to reduce MP production costs by PSB (Capson-Tojo et al. [2020](#page-27-4)). Though, there are limited studies on actually using the mixed PSB culture in an outdoor environment for MP production (Hülsen et al. [2022](#page-29-5)a). Most of the experiments done are labs scale under controlled environmental conditions and the results from these studies cannot be directly generalized to real MP production systems from organic waste substrates. The use of organic waste as substrate for mixed PSB culture may not only impact biomass growth but also can signifcantly afect the produced biomass quality to be utilized as MP-based animal feed (Capson-Tojo et al. [2020\)](#page-27-4).

## 4.3 Gas-based processes

Although direct fermentation is a well-established technology for MP production, there is a risk of



diferent phases fed on poultry wastewater (Reproduced from (Hülsen et al. [2022b](#page-29-18)) with permission from Elsevier 2022). **D** Relative abundances based on 16S analysis for the 100 L photobioreactor fed with piggery wastewater (Reproduced from (Hülsen et al. [2022](#page-29-5)a) with permission from Elsevier 2022)

potential bioaccumulation of heavy metals, pesticides, and chlorinated hydrocarbons present in agroindustrial waste (Luo et al. [2012;](#page-30-16) Martin et al. [2013;](#page-31-9) Puligundla and Mok [2021](#page-32-7); Sakarika et al. [2022;](#page-32-4) Salehi and Chaiprapat [2022\)](#page-32-16). This issue associated with the toxicity of these contaminants can be addressed by converting the wastes into a non-toxic substrate that can be easily used by microorganisms for example biogas production by AD of organic biomass (Acosta et al. [2020](#page-25-2); Kim et al. [2022](#page-29-14); Pander et al. [2020;](#page-32-17) van der Ha et al. [2012](#page-33-15); Verbeeck et al. [2019\)](#page-34-23). This approach is commercially more attractive as it can utilize diferent waste streams together for MP production. Figure [5](#page-16-0) depicts the process integration of biogas production and liquid streams of the AD process to synthesize MP for animal feed production. The frst step in this process is AD (codigestion in case of more than one substrate) of the organic waste and subsequently MP production by fermentation of biogas with selected methanotrophs



<span id="page-16-0"></span>**Fig. 5** Process integration of anaerobic digestion of farm organic waste and fermentation of produced biogas to produce MP for animal feed. The combination of both processes refects the integrated valorization of agriculture waste streams

towards renewable feed/food products. The process includes the demonstration and analysis that the produced feed is both safe and benefcial compared with currently used animal feed

in a closed bioreactor (Bertasini et al. [2022;](#page-26-6) Tsapekos et al. [2021;](#page-33-16) Zha et al. [2021](#page-35-5)). Biogas fermentation is performed in a sterile medium comprising the necessary nutrient for microbial growth. The optimization of the production process is controlled by several factors including bio-methane to oxygen ratio, AD digestate nutrient concentration, temperature, reaction time, biogas concentration, moisture, and pH (Martin et al. [2013;](#page-31-9) Puligundla and Mok [2021;](#page-32-7) Sakarika et al. [2022;](#page-32-4) Tsapekos et al. [2021\)](#page-33-16). In this process, biogas composition can considerably afect the growth of methanotroph microbial cultures (Cantera et al. [2016](#page-26-17)). Diferent microbial strains have diferent growth and substrate utilization and it is important to understand the conditions necessary to select the most suitable microbial culture and waste streams to be utilized as substrate (Areniello et al. [2023](#page-26-12); Banks et al. [2022;](#page-26-18) Li et al. [2022](#page-30-17)b). The generated biomass is then processed for protein extraction which requires biomass separation and drying. Finding suitable separation techniques for cost-efectively harvesting biomass, as well as drying studies is a key process for preparing the fnal product (Lee and Stuckey [2022;](#page-30-18) Sakarika et al. [2022](#page-32-4)). Several separation techniques have been reported including focculation with gravity settling. Spray and solar drying are being studied currently as sustainable and cost-efective techniques (Hu et al. [2022;](#page-29-21) Lee and Stuckey [2022\)](#page-30-18). The dried product will is fnally analyzed to fnd out the content of total proteins, amino acids, vitamins, salts, and other parameters of interest (Bonan et al. [2022;](#page-26-19) Lee and Stuckey [2022;](#page-30-18) Salehi and Chaiprapat [2022\)](#page-32-16).

To warrant the sustainability and economic viability of MP as animal feed, bio-methane as a feedstock could be an attractive alternative. Bio-methane is a viable substrate, as it is a major by-product of the AD of organic wastes (farm manure, organic agriculture waste, landflls, food waste, etc.) (Acosta et al. [2020;](#page-25-2)

Muñoz et al. [2015](#page-31-10); Puligundla and Mok [2021](#page-32-7); Sakarika et al. [2022;](#page-32-4) Woolley et al. [2023](#page-34-1); Xu et al. [2021a](#page-34-18)). Current research is focused on innovating the technology for coupling the biogas generation with gas fermentation using the selected methanotrophs such as *Methylocystis parvus* and *Methylococcus capsulatus* (Comesaña-Gándara et al. [2022](#page-27-20); Gęsicka et al. [2021](#page-28-17); Salehi and Chaiprapat [2022](#page-32-16); Tsapekos et al. [2021](#page-33-16); Zha et al. [2021](#page-35-5)). Moreover, nutrient-rich digestate, a product of anaerobic digestion, can provide nitrogen and phosphorus instead of using expensive chemical nutrients. Using such kind of integrated approach, organic waste resources upcycling can considerably contribute to fnding sustainable, cheaper, and environment-friendly protein sources (Tsapekos et al. [2021\)](#page-33-16). However, there is a lack of research on the industrial-scaled production of MP from AD streams as most of the work earlier was focused on the use of natural gas, and conventional nutrient sources. Upcycling of biogas and nutrient recovery from agricultural organic waste is the only viable method for large-scale MP production without requiring an associated increase in energy consumption.

The MP produced by aerobic fermentation of methane gas has distinctive benefts. Methanotrophic microbes are protein-rich biomass with greater than 75% protein content and have the potential to replace conventional protein sources (Angelidaki et al. [2018](#page-26-20); Puligundla and Mok [2021](#page-32-7); van der Ha et al. [2012](#page-33-15); Verbeeck et al. [2019;](#page-34-23) Wang et al. [2022;](#page-34-3) Xu et al. [2021a](#page-34-18)). The ability to use a variety of wastes with a wide range of organic compounds, under diferent environmental conditions of temperature, humidity, etc*.* is the key advantage of this process (Li et al. [2022b](#page-30-17); Martin et al. [2013](#page-31-9); Puligundla and Mok [2021](#page-32-7); Sakarika et al. [2022](#page-32-4); Tsapekos et al. [2021](#page-33-16)). However, this process emits  $CO<sub>2</sub>$  from the biogas into the air which is a shortcoming because  $CO<sub>2</sub>$  is the main contributor to climate change. Additionally, there is a risk of explosion because of the formation of a mixture of biogas and  $O<sub>2</sub>$  directly in the bioreactor culture medium (Molitor et al. [2019;](#page-31-8) Muñoz et al. [2015;](#page-31-10) Van Peteghem et al. [2022](#page-33-17); Yang et al. [2021a](#page-34-24)).

To overcome the issue of  $CO<sub>2</sub>$  emissions, several alternative strategies are proposed as a possible solution including the co-cultivation of autotrophic HOB with methanotrophs in a single-stage process (Areniello et al. [2023;](#page-26-12) Chen et al. [2022](#page-27-21); Hu et al. [2022](#page-29-21); Lin et al. [2022](#page-30-19)). However, the application of HOB requires  $H_2$  gas which is generally produced by an energy-demanding water electrolysis method with a risk of an explosion raising safety issues as well. Another promising replacement process is the integration of methanotrophic and algal biomass production in a phototrophic bioreactor in which the  $CO<sub>2</sub>$ generated is fixed by algae while  $O_2$  produced by algae is utilized as an electron acceptor by methanotrophs (Areniello et al. [2023;](#page-26-12) Azarpour et al. [2022;](#page-26-21) Balagurunathan et al. [2022](#page-26-22); Wada et al. [2022](#page-34-25)). This process is advantageous over the methanotrophic and methanotrophs-HOB processes as it can all together convert  $CH_4$  and  $CO_2$  in the biogas without the requirement of any external  $O_2$ . Hydrogen and methane gases need to be dissolved in the aqueous media for their availability to microbes, however, poor solubility of these gases is a bottleneck in their application for MP production (Areniello et al. [2023\)](#page-26-12). Several bioreactor confgurations such as with controlled stirring systems and shapes like U-loop have been designed as an alternative to conventional reactors to address the solubility issues (Nizovtseva et al. [2022;](#page-31-19) Nyyssölä et al. [2022;](#page-31-2) Tyagi et al. [2022\)](#page-33-18). Natural gas is generally used as a source of methane gas for MP production. However, in the AD process, hydrogen is produced in the acidogenesis stage as an intermediate product which is rapidly transformed into methane by methanogenic archaea. Therefore, biogas-based MP production by HOB is not considered a suitable process unless the methane formation is inhibited intentionally to produce bio-hydrogen which is not viable commercially so far (Khoshnevisan et al. [2022;](#page-29-3) Nyyssölä et al. [2022](#page-31-2)).

An alternative strategy to address the issues related to the use of biogas is to upgrade it to get the needed characteristics (Khoshnevisan et al. [2022](#page-29-3)). Biogas upgradation can be done by several physicochemical and biological processes such as absorption and adsorption which are already commercially available and can be utilized for MP production (Luo et al. [2012\)](#page-30-16). It can also be done using biological processes which involve electrochemical means to perform the in situ or *ex-situ* reduction of the substrates for instance conversion of  $CO<sub>2</sub>$  to  $CH<sub>4</sub>$  in presence of methanogens (Martin et al. [2013;](#page-31-9) Muñoz et al. [2015;](#page-31-10) Verbeeck et al. [2019](#page-34-23)). For instance, the production of MP from upgraded biogas was reported by Acosta et. al., (Acosta et al. [2020\)](#page-25-2) in which an electrochemical process was applied to separate  $CO<sub>2</sub>$  and



<span id="page-18-0"></span>**Fig. 6** Overview of the integrated process scheme involving anaerobic digestion and electrochemical biogas upgrade **A** Schematic of the MP production process by electrochemically upgraded biogas  $BCO<sub>2</sub>$  flux in the biogas stream with the corresponding removal and current efficiencies (Acosta et al. [2020\)](#page-25-2). PV-driven MP production **C** schematic depiction of energy transfer of MP from solar energy. Each conversion step is associated with an energetic efficiency corresponding to

 $CH<sub>4</sub>$  from the biogas stream produced from anaerobic digestion. The electrochemically upgraded biogas having a blend of  $CH_4$  with  $H_2$  exhibited the high quality and quantity of MP production demonstrating the electrochemical separation as a viable option for biogas upgradation to produce MP as shown in Fig. [6.](#page-18-0)  $CO<sub>2</sub>$  in the biogas can be efficiently separated from  $CH<sub>4</sub>$  by solar energy driven electrochemical process while generating  $H_2$  and  $O_2$ , simultaneously (Kumar et al. [2016](#page-29-22); Matassa et al. [2016](#page-31-20)). A quantitative analysis on the potential of renewable photovoltaic-driven biomass production showed that MP produced using such renewable resources has production efficiently higher than that of conventional crops Fig. [6](#page-18-0)C (Leger et al.  $2021$ ). In this study, the efficiency of applying solar power for conversion of  $CO<sub>2</sub>$  in the air into microbial biomass was investigated and it was showed that the MP based food production leave behind the cultivation of staple crops like soybean in terms of protein yields and caloric value per unit land area (Fig. [6](#page-18-0)D). The study concluded that MP food

the electricity fraction used for electrosynthesis of the carbonsubstrate, **D** Protein yield of PV-driven MP production as a function of irradiance. Diferent electron donors and assimilation pathways were analyzed in comparison with conventional crops like soybean the highest protein-yielding staple food (Reproduced from (Leger et al. [2021](#page-30-20)) with permission from Elsevier 2021)

could considerably contribute to the growing food demand and can help in sharing future inadequate land resources.

The application of organic wastes as carbon and nitrogen source is considered a viable option. The supernatant of the AD process is considered a rich source of nitrogen for the subsequent methanotrophs to produce MP. However, supernatant from the AD process has a high ammonium concentration ranging from 1,000–3,000 mg-NH<sub>4</sub>  $+$ <sub>-</sub>N/L, which needs to be diluted to meet the desired ammonium concentration (Acosta et al. [2020\)](#page-25-2). The protein contents of the biomass produced in the methanotrophic bioreactor are reported in the range of 40–60% on a dry cell weight (DCW) basis, which is higher than the protein contents of commonly used soybean meal for animal feed. Hence, MP-based feed can be an alternative to conventional protein sources for animal feed additives.

 $H<sub>2</sub>S$  is usually a component of the biogas produced from AD and its concentration can vary depending on

the substrates used (Angelidaki et al. [2018](#page-26-20); Miltner et al. [2017](#page-31-21); Salehi and Chaiprapat [2019\)](#page-32-22). Recently, several studies have reported the detrimental effects of  $H_2S$  on the protein contents of the MP (Xu et al. [2020\)](#page-34-2). It is reported that biomass production with natural gas is signifcantly higher than that of using raw biogas containing  $H_2S$ . Xu et. al. reported the growth inhibition of *M. acidiphila* and a decrease in amino acid contents in the produced biomass at approximately 1000 ppm of  $H_2S$  in crude biogas as shown in Fig. [7](#page-19-0) (Xu et al. [2020](#page-34-2)). Moreover, the quality of MP produced is also affected by the  $H_2S$  in the biogas. So, it is very important to desulfurize raw biogas before using it for MP production. In MP production, the source and concentration of nitrogen have an important role in synthesizing protein by microbes. Expensive synthetic nitrogen-rich media like ammonium mineral salts (AMS), nitrate mineral salts (NMS), etc*.* are generally used to grow MP-producing microbes including algae, methanotrophs, and HOB. Organic waste has nitrogen in diferent forms such as ammonia, ammonium salts, nitrite, and nitrate, and can be used as a source of nitrogen and carbon, simultaneously. In anaerobic digestion, ammonia nitrogen  $(NH_4^+$ -N) rich biogas slurry is produced which becomes a grave environmental and economic issue

given that about 1 billion tons of biogas slurry per year are only produced in China (Ding et al. [2023](#page-27-2)). Therefore, using waste streams as a nitrogen source for the growth of MP-producing microbes has shown the potential to minimize the cost. pH has a signifcant impact generally on microbial growth. Regulation of pH in MP production is crucial to get the maximum yield (Zeng et al. [2022](#page-35-0)). It was observed that keeping an acidic environment in the bioreactor was essential to get higher biomass and MP synthesis. Biogas and nutrient-rich liquid products of AD of organic wastes can serve as a solution. Traditionally, biogas is used for combined heat and power generation and the nutrients rich digestate of the AD process is supplied to plants as fertilizer, however, it is not safe and its storage could result in fugitive emissions of greenhouse gases including methane and ammonia (Haraldsen et al. [2011;](#page-28-18) Matassa et al. [2015;](#page-31-22) Styles et al. [2018\)](#page-33-19). The assimilation of nutrients by methanotrophs to produce protein-rich biomass seems to be a favorable substitute (Khoshnevisan et al. [2020;](#page-29-23) Matassa et al. [2015](#page-31-22)). Therefore, effective biotechnology for protein-rich microbial biomass production using both liquid and gas streams of farm organic waste AD process will improve the economic value of the overall process. Yet, this 2nd generation's

<span id="page-19-0"></span>**Fig. 7** Growth inhibition of methanotrophs by  $H_2S$ : **a** OD<sub>410</sub> with reaction time; **b** CH<sub>4</sub> profile over time; **c** sulfde profle with reaction time; **d** amino acid profle analysis as a percentage of dry biomass (Reproduced from (Xu et al. [2020](#page-34-2)) with permission from Elsevier 2020)



concept of MP production from biogas is not established. There are questions about the efficiency of the AD process, biogas fermentation, and quality of the MP produced. Also, it is a subject of research whether biogas-fed MP can substitute traditional animal feed sources such as soybean and fshmeal (Suriyapha et al. [2020\)](#page-33-14). MP can be produced locally from organic-rich food waste, is easy to trace source and quality, and can be capable of providing all necessary amino acid and vitamin requirements for animal growth (Alloul et al.). Furthermore, compared to other sources of protein, the environmental burdens in terms of energy, greenhouse gases, water footprint, and land area are claimed to be much lower for MP, although limited scientifc literature is available in this area (Sillman et al. [2020\)](#page-33-20). However, the selection of bacteria with comparatively higher growth rates, protein content, and healthy amino acid composition are the most important factors to produce high-quality cell biomass for animal feed production.

While the concept of direct biogas fermentation to MP has many potential benefts, several challenges must be overcome to make this process viable, efficient, and safe. Diferent challenges are associated with the direct conversion of gases into MP including mass transfer limitations, quality of gas feed gas, the need for microbial catalysts, and safety(Khoshnevisan et al. [2022](#page-29-3); Salehi and Chaiprapat [2022](#page-32-16); Wang et al. [2022;](#page-34-3) Xu et al. [2021a](#page-34-18); Zha et al. [2021\)](#page-35-5). Limiting gas-to-liquid mass transfer is still regarded as one of the main obstacles to the commercialization of gasbased fermentation systems (Sakarika et al. [2022](#page-32-4)). A useful parameter for comparing the mass transfer capacities of various reactor layouts is the volumetric mass transfer coefficient, which is a direct evaluation of a reactor's hydrodynamic state (Li et al. [2021\)](#page-30-21). Improvements in impeller designs, fuid fow patterns, aerated power efficiency, mixing time, baffle design, and the utilization of microbubble dispersers are some of the traditional methods investigated in literature to overcome mass transfer constraints (Dupnock and Deshusses [2019;](#page-27-22) Lai et al. [2021](#page-30-22); Rodríguez et al. [2020](#page-32-23); Soto et al. [2021\)](#page-33-21). Before being added to the fermentation process, the feed gas needs to be clean. To guarantee optimal production, appropriate gas cleanup techniques should be utilized before gas-based fermentation processes (Acosta et al. [2020;](#page-25-2) Golmakani et al. [2022](#page-28-19); Naquash et al. [2022](#page-31-23)). Another crucial step in commercializing the process is the discovery of anaerobic bacteria that can convert biogas into MP with greater product yields (Leu et al. [2020;](#page-30-23) Thamdrup et al. [2019\)](#page-33-22). Several safety challenges must be considered and addressed to minimize the risks associated with biogas fermentation to MP. Some of the key safety challenges include explosion hazards, toxic gas emissions, and fre hazards (Salehi and Chaiprapat [2022](#page-32-16); Xu et al. [2021a;](#page-34-18) Zha et al. [2021\)](#page-35-5). Biogas is a combustible gas that can pose an explosion hazard if it is not handled properly. Bioreactors and other equipment used in the fermentation process must be designed and operated to minimize the risk of explosion. Appropriate safety measures, such as gas detection systems, explosion-proof equipment, and adequate ventilation, must be in place to prevent accidents (Stolecka and Rusin [2021\)](#page-33-23). Also, biogas fermentation can produce toxic gases such as hydrogen sulfde and carbon monoxide, which can pose health risks to workers and the environment (Tayou et al. [2022;](#page-33-24) Xu et al. [2020](#page-34-2)). Proper ventilation, gas detection systems, and personal protective equipment must be used to prevent exposure to these gases.

Another issue related to the conversion of biogas to MP is its competition with other biogas end uses. Biogas is a versatile fuel that can be used for electricity generation, heating, and transportation (Abanades et al. [2022](#page-25-7)). As such, the biogas market is highly competitive, and the cost of producing biogas for use in MP production would need to be competitive with other uses of biogas. Moreover, the production of MP requires a signifcant amount of energy, including the energy needed to maintain the bioreactor temperature and supply nutrients to the microorganisms. This energy must be supplied from renewable sources to ensure that the process has a low carbon footprint. The MP production requires a high level of nutrients, including nitrogen and phosphorus, which may not be readily available in the substrate used for biogas fermentation. So additional nutrient supplementation may be required to achieve high yields of MP. The production of MP from biogas must be economically viable to compete with other sources of protein. The cost of production must be low enough to make the fnal product competitive with other protein sources on the market.

## **5 MP in animal feed**

MP produced from microbes is currently commercialized and in use as animal feed from companies like Uniprotein®, and Unibio A/S, Denmark, known as FeedKind® (Alloul et al.). These commercial products are produced by aerobic fermentation of methane in natural gas. In this process, the fermentation of methane is carried out by methanotrophs while assimilating nitrogen compounds such as ammonium, ammonia, and nitrate in their proteinaceous cells. MP produced by methanotrophic fermentation has distinctive benefts and can be utilized as cattle feed as an alternative to plant-based protein due to its higher contents of protein  $(>75\%$  dry weight) (Ritala et al. [2017\)](#page-32-0). Few selected species of bacteria and fungus have been employed to produce MP as animal feed for the last two decades. Recently, the production of MP for animal feed applications has gained momentum. Animal feed from sugar fermentation was produced in Finland in 1974 and the process was registered with the commercial name "PEKILO®" to be used in the European Union countries. This process utilizes the flamentous fungus *Paecilomyces varioti* using sugar as a substrate to produce MP (Ritala et al. [2017\)](#page-32-0). Start-up company eniferBio has recently updated the PEKILO® process using diferent industrial by-products as a substrate to produce the MP biomass validated for aquafeed and aiming to extend it further for human consumption (eniferBio [2022](#page-28-2)). Imperial Chemical Industries UK, developed an MP using methanol as a substrate for animal feed with the brand name Pruteen, by employing *Methylophilus methylotrophus* bacteria up to 70% protein (Johnson [2013](#page-29-24)). Nevertheless, Pruteen production became commercially unviable because of the availability of other inexpensive protein sources such as soybean, and Pruteen production was discontinued in 1970. Pruteen was produced from methanol, but lately, methane gained interest as a substrate for MP production. Several multinational companies including UniBio A/S and Calysta Inc. have developed a U-loop fermenter technology employing fermentative methanotroph bacteria to convert natural gas to MP. UniBio commercialized the MP by the brand name UniProtein® with~70% protein, which is already approved for use in animal feed (Petersen et al. [2017](#page-32-24)). Similarly, Calysta Inc. introduced its MP with the commercial name FeedKind®, in the UK market in 2016

as animal feed has partnered with Cargill USA to build a larger production facility. The biotechnology companies in Qatar have already shown interest and ambition to adopt new technologies. Very recently, Gulf Biotech Qatar, and Unibio, have partnered to establish a production facility in Qatar, the gulf's frst natural gas to protein plant, to produce initially 6,000 tonnes of UniProtein® as a feed for aquaculture livestock (Unibio [2022\)](#page-33-0).

The biotechnology companies like Calysta Inc. and UNIBIO are producing MP by employing pure microbial cultures that necessitate comparatively pure sources of co-substrates and nutrients to avoid contamination risks. However, the supply of chemical grade nutrients (micro and macronutrients) and natural gas as methane sources increase the operational costs and are the bottlenecks to decreasing the production costs of the MP. Furthermore, the usage of natural gas is not sustainable and environmentally friendly. Therefore, the sustainability of MP production by natural gas-fed fermentation process is under question (Kalyuzhnaya et al. [2013](#page-29-4)). To address the above-mentioned issues, cheaper and sustainable sources of both substrates (methane and nutrients), are required resulting in the bacterial protein production roar.

Yeast and methanotrophs-based MP produced using waste substrates have been tested as animal, fish, and shrimp meal (Sharif et al. [2021;](#page-32-2) Woolley et al. [2023](#page-34-1); Zheng et al. [2023](#page-35-3)). The yeast C. *utilis* MP was used as a feed supplement for weaned piglets in a study by Yang et. al (Yang et al. [2021](#page-34-15)c). It was observed that *C. utilise-*based MP has the potential to substitute the use of antibiotics in the growth of weaned piglets. Additionally, MP enhanced the overall growth, and intestinal health with a reduction in diarrhea, and enhanced cecal microfora diversity and richness in piglets. MP produced by PSB has been applied for many years as the poultry and pig feed to enhance the egg laying rate, yolk pigment content, chicken health fneness, and animal weight (Lu et al. [2019\)](#page-30-8).

MP-based aquaculture feed is being considered the next generation of unconventional proteins as MP has shown promising results in research studies on aquaculture (Glencross et al. [2020](#page-28-20)). Several research trails exploring the application of fungal, algal, and methanotrophic MP as aquafeeds in shrimp, Japanese yellowtail, salmon, trout,

barramundi, turbot juveniles, largemouth bass, and kingfsh have reported higher growth rates compared to conventional feed with good digestibility (Biswas et al. [2020;](#page-26-23) Glencross et al. [2020;](#page-28-20) Pilmer et al. [2022](#page-32-25); Woolley et al. [2023](#page-34-1); Zamani et al. [2020](#page-35-10); Zhang et al. [2022](#page-35-11); Zheng et al. [2023\)](#page-35-3). *S. cerevisiae* MP in diferent formulations ranging from 15 to 24%, has been studied as an alternative to traditional fshmeal or soybean meal without any negative impact on the growth rate (Øvrum Hansen et al. [2019](#page-31-24)). Methanotroph-assisted MP meals containing 36% MP showed a higher growth rate of Atlantic salmon as compared to that of control, however, a reduction in the digestibility of nutrients was observed. MP produced from methanotrophic *M. capsulatus* biomass showed the optimum growth of trout when fed with 38% MP of the food protein and 52% in salmon rations. Further, it was reported that using a blend of *M. capsulatus* feed in soybean meal prohibited the development of enteritis in salmon fish which is usually assigned to the use of soybean meal, signifying additional advantages of MP feed. Another study found that KnipBio Meal based on *Methylobacterium extorquens* can be used at 55% MP instead of fshmeal in salmon rations with a comparable growth rate to that of soybean meal (Hardy et al. [2018\)](#page-28-21). Production of MP from purple non-sulfur bacteria biomass is currently on the rise for aquaculture feed. MP produced from a combination of two purple non-sulfur bacteria with 1% in the aquaculture feed improved the growth of shrimp to that of the control feed (Alloul et al. [2021](#page-26-24)). MP feed usually leads to palatability issues in aquaculture and several research trials have suggested the use of garlic (*Allium sativum*) and fsh hydrolysates as palatability and digestion enhancers by exciting the enzymes responsible for performing digestion (Abdelwahab et al. [2020;](#page-25-4) Esmaeili et al. [2017](#page-28-22); Jones et al. [2020](#page-29-1); Tola et al. [2022](#page-33-25); Woolley et al. [2023](#page-34-1)). Methanotrophic MP feed used in barramundi trials showed substantial enhancement in palatability when fed with additives, increase in growth and feed utilization efficiency without any decrease in feed intake compared to conventional fsh feed. Moreover, triglyceride and histopathology results depicted that MP-fed barramundi showed healthier livers as compared to that of the control feed (Fig. [8\)](#page-23-0). The studies above presented the MP derived from diferent waste streams as an efective

alternative next-generation feed for aquaculture, however, it is imperative to carefully control the substitution level of MP with the traditional fsh meal as a higher percentage of MP can have detri-mental effects on aquaculture (Zhang et al. [2022](#page-35-11)). For instance, a recent study reported that methanotroph MP can substitute 30% fsh meal without afecting the growth rate and health of urbot juveniles (*Scophthalmus Maximus L.*) (Zheng et al. [2023](#page-35-3)). Nevertheless, fshmeal's higher substitution levels with MP showed detrimental impacts on antioxidant capacity, liver health, and metabolism (Zhang et al. [2022\)](#page-35-11).

High-value MP and additives produced by PSB can be used in aquaculture for plankton, shrimp, and fsh feed. PSB-based aquaculture feed is reported to enhance fsh and shrimp growth, and suppress some diseases such as fn rot (Lu et al. [2019](#page-30-8)). PSBs are one of the earlier microorganisms developed in China for aquaculture and are now widely being used for animal feed (Laskowska et al. [2017](#page-30-12)) and being sold on the largest electronic business platform, Alibaba. The products can be liquid or powder, and the prices range from 600 to 1200 US dollars/ton. About 103.6 million tons of liquid and dry powder products of PSB with 621.6 million to 1.2 billion US dollars market per year are being produced (Lu et al. [2019](#page-30-8)).

# **6 Environmental impact, perspectives, and challenges**

Sustainable environment and food security demand circular management of carbon and nutrients rich organic wastes and industrial side streams. The utilization of organic liquid and solid wastes for MP production can be a better approach to waste management as compared to conventional technologies such as anaerobic digestion, landflling, composting, etc*.* being employed for diferent waste streams (Pereira et al. [2022](#page-32-10)). Nonetheless, a vigilant assessment of the individual case and process development is a prerequisite to attain sustainability, for instance, the integration of AD and gas fermentation with a clean and safe MP production process (van der Ha et al. [2012](#page-33-15)). Furthermore, MP production using waste feedstock can neutralize harmful substances and reduce environmental pollution converting waste to value-added products. Another aspect is the  $CO<sub>2</sub>$  capture by the <span id="page-23-0"></span>**Fig. 8** (I) Graphical presentation of methanotrophic MP evaluation as fshmeal in aquaculture (II) Representative histological microphotographs of liver and spleen sections of barramundi fed diets (**A** & **C**) control diet (**B** & **D**) 30% MP (Reproduced from (Woolley et al. [2023\)](#page-34-1) with permission from Elsevier 2023)



autotrophic MP-producing microbes. According to a study, the synthesis of MP and biochar can lead to an annual carbon sequestration of up to 50% of the Paris Agreement target (Ngoc-Dan Cao et al. [2022](#page-31-25)). The environmental benefts of MP production along with sustainable high-quality edible protein production have led to enhanced interest in this process. Besides, MP-based food and feed production can become an alternative and reduce the production of traditional protein sources which are unsustainable due to their low conversion efficiencies. The application of MP biomass for animal feed is limited by

 $V$  and  $V$  and  $V$  $\bigcircled{2}$  Springer

the higher cost of the current substrates and competition from conventional feed sources. Climate change is a serious threat to the global food production system and MP-based diets can address the several environmental boundaries currently limiting food production in addition to sinking its environmental footprint. MP food and feed can be produced from photosynthetic microbes like microalgae, purple sulfur bacteria, and cyanobacteria without the need for arable land. In addition,  $CO<sub>2</sub>$  capture, storage, and conversion to organic compounds have opened up a new niche for the synthesis of MP biomass for food and feed exclusively free of photosynthesis. Another approach to assess the environmental benefts of MP is its comparison to livestock-based protein production. Although ruminant meat offers valued protein to humans and some animals, the production of livestock has several diverse implications on the environment including but not limited to greenhouse gas emissions, deforestation, excessive land, and water use, and eutrophication. MP has shown the potential to address the issues associated with livestock production by performing a life cycle assessment (LCA) of MP production in bioreactors (Finnigan et al. [2019;](#page-28-23) Hashempour-Baltork et al. [2020](#page-28-24); Linder [2019](#page-30-24); Sillman et al. [2020](#page-33-20); Stephens et al. [2018](#page-33-26)). A recent model-based study projected that globally replacing 20% of traditional protein consumption with MP by 2050 can decrease the required global pasture area, deforestation, and  $CO<sub>2</sub>$  emissions by approximately half (Humpenöder et al. [2022\)](#page-29-25).

The 1st generation of MP feed for livestock and aquaculture is produced using natural gas and synthetic chemicals and therefore may be considered a less sustainable alternative to conventional feeds. These challenges have resulted in the development of 2nd generation MP where scientists have integrated the use of cost-efective feedstock with sustainable technologies such as carbon capture and reduction with renewable energies, anaerobic digestion, nutrient recovery, biogas upgrading, and fermentation. The fermentation of two protein-rich microbes MOB and HOB has shown a great potential to upcycle effluents from AD into protein-rich biomass integrated with renewable energy power under the model of Powerto-food. Scaled-up production of MOB-based MP has been commercialized long ago and its environmental impact has been studied and reported (Khoshnevisan et al. [2022](#page-29-3)). On the other hand, the production of 2nd generation MP is still under investigation and has been not scaled up therefore its industrial environmental impacts have not been studied to this point. There is a lack of detailed analysis on the sustainability characteristics of  $2<sup>nd</sup>$  generation MP synthesis facilities under diferent feeding strategies and technological applications, particularly, its integration with renewable energy resources.

Although MP has very attractive properties, there are some anti-nutritional factors involved that raise some concerns about its use in feed. The major issue is the presence of nucleic acid in MP which is higher as compared to other conventional protein sources. The presence of nucleic acid in a protein diet is related to an increase in the serum's uric acid which results in the development of kidney stones. Amino acids form about 70–80% of the nitrogen composition whereas the remaining nitrogen is in the form of nucleic acids which support microbial growth. Several physicochemical treatment processes have been proposed to remove nucleic acids during the MP production process (Dantas Jr et al. [2016\)](#page-27-23). Another challenge that needs to be addressed is the presence of microbial cell walls which creates some digestion issues for birds and some animals with simple digestive systems. Moreover, viable microbial cells need to be inactivated to avoid skin and gastrointestinal infections and unpleasant color and taste. Filamentous fungi MP is considered a higher yield process because of its high growth rates as compared to yeasts; however, it posed higher contamination risks as compared to any other microbial MP. Bacterial MP has also some limiting factors including high RNA content, the presence of endotoxins, and the risk of contamination.

Conversion of organic waste into high-quality food/feed has great potential, but many challenges need to be addressed before a broader application. MP is presently not commonly recognized by the market as animal feed, and it is a question of whether farmers are willing to use MP in place of conventional animal feed. Accordingly, a widespread campaign among different stakeholders including legal entities is required to get a prevalent public acceptance of MP biomassbased feed and food products. Careful selection of the MP synthesis process, suitable feedstock, and microorganisms are the key elements to avoid all of the above-mentioned issues associated with MP food and feed production and get the maximum benefts of MP.

Finally, there is a need to perform up-scale outdoor research for all the microbial systems including fungal, yeast, algae, PSB, and methanotrophs to progress and implement the MP production technology in the real world. Upscaling of the process would facilitate to determination of various factors involved in microbial growth, waste feedstock utilization, and fnally MP product quality continuously. To do so, detailed cultivation strategies need to be designed and optimized for each bioprocess involved in the production scheme including comprehensive economic and environmental evaluations. A scaled system can provide more information about biomass retention for diferent microorganisms, bioreactor design based on the feedstock and local environmental conditions, and downstream processing such as sterilization, dewatering, and drying. Further applied research is required on diferent components of the MP produced, together with validating their impact on aquaculture, poultry, and other livestock through enthusiastic feed trials.

Conversion of agroindustrial waste to MP also poses several challenges that must be addressed to ensure the safety of the production process. Safety challenges in biogas fermentation include explosion hazards, toxic gas emissions, and fre hazards. Several microorganisms can produce some toxic substances such as cyanotoxins and mycotoxins during the MP manufacturing process and there is a need to control these toxins production by carefully choosing the microorganism for MP processing (Areniello et al. [2023;](#page-26-12) Xu et al. [2021a\)](#page-34-18). Additionally, apart from toxic substances, mutation of the microbes during MP processing may also result in the formation of a few toxic carcinogenic constituents (Areniello et al. [2023\)](#page-26-12). The fermentation process can produce toxic gases which can pose health risks to workers and the environ-ment (Kerckhof et al. [2021\)](#page-29-9). Also, there are several challenges associated with the conversion of biogas into MP including mass transfer limitations, quality of gas feed gas, the need for microbial catalysts, and safety. Furthermore, the production of MP-based food and feed products is subject to regulation by various national and international agencies. The regulatory framework for MP-based feed production processes typically requires safety assessment to ensure that they do not pose a risk to human or animal health. The safety assessment typically includes an evaluation of the production process, the source of the MP, and any potential risks associated with the use of the product.

**Acknowledgements** This work was made possible by QNRF grant No. MME03-1101-210007 from the Qatar National Research Fund. The statements made herein are solely the responsibility of the authors.

**Funding** Open Access funding provided by the Qatar National Library.

#### **Declarations**

**Confict of interest** The authors have no relevant fnancial or non-fnancial interests to disclose.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit [http://creativecommons.org/licenses/by/4.0/.](http://creativecommons.org/licenses/by/4.0/)

#### **References**

- <span id="page-25-7"></span>Abanades S et al (2022) A critical review of biogas production and usage with legislations framework across the globe. Int J Environ Sci Technol 19:3377–3400. [https://doi.org/](https://doi.org/10.1007/s13762-021-03301-6) [10.1007/s13762-021-03301-6](https://doi.org/10.1007/s13762-021-03301-6)
- <span id="page-25-4"></span>Abdelwahab AM, El-Bahr SM, Al-Khamees S (2020) Infuence of dietary garlic (Allium sativum) and/or ascorbic acid on performance, feed utilization, body composition and hemato-biochemical parameters of juvenile Asian Sea Bass (Lates calcarifer). Animals 10:2396
- <span id="page-25-5"></span>Abiusi F, Moñino Fernández P, Canziani S, Janssen M, Wijffels RH, Barbosa M (2022) Mixotrophic cultivation of Galdieria sulphuraria for C-phycocyanin and protein production. Algal Res 61:102603. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.algal.2021.102603) [algal.2021.102603](https://doi.org/10.1016/j.algal.2021.102603)
- <span id="page-25-2"></span>Acosta N, Sakarika M, Kerckhof F-M, Law CKY, De Vrieze J, Rabaey K (2020) Microbial protein production from methane via electrochemical biogas upgrading. Chem Eng J 391:123625. [https://doi.org/10.1016/j.cej.2019.](https://doi.org/10.1016/j.cej.2019.123625) [123625](https://doi.org/10.1016/j.cej.2019.123625)
- <span id="page-25-1"></span>Ahmad Khorairi ANS, Sofan-Seng N-S, Othaman R, Abdul Rahman H, Mohd Razali NS, Lim SJ, Wan Mustapha WA (2021) A review on agro-industrial waste as cellulose and nanocellulose source and their potentials in food applications. Food Rev Int. [https://doi.org/10.1080/](https://doi.org/10.1080/87559129.2021.1926478) [87559129.2021.1926478](https://doi.org/10.1080/87559129.2021.1926478)
- <span id="page-25-0"></span>Aiking H (2011) Future protein supply. Trends Food Sci Technol 22:112–120. [https://doi.org/10.1016/j.tifs.2010.04.](https://doi.org/10.1016/j.tifs.2010.04.005) [005](https://doi.org/10.1016/j.tifs.2010.04.005)
- <span id="page-25-6"></span>Aita BC, Spannemberg SS, Schmaltz S, Zabot GL, Tres MV, Kuhn RC, Mazutti MA (2019) Production of cell-wall degrading enzymes by solid-state fermentation using agroindustrial residues as substrates. J Environ Chem<br>Eng 7:103193. https://doi.org/10.1016/j.jece.2019. 7:103193. [https://doi.org/10.1016/j.jece.2019.](https://doi.org/10.1016/j.jece.2019.103193) [103193](https://doi.org/10.1016/j.jece.2019.103193)
- <span id="page-25-3"></span>Aker KC, Robinson CW (1987) Growth of Candida utils on single-and multicomponent-sugar substrates and on waste banana pulp liquors for single-cell protein

production. MIRCEN Journal of Applied Microbiology and Biotechnology 3:255–274. [https://doi.org/10.1007/](https://doi.org/10.1007/BF00933579) [BF00933579](https://doi.org/10.1007/BF00933579)

- <span id="page-26-0"></span>Alberti MA, Blanco I, Vox G, Scarascia-Mugnozza G, Schettini E, Pimentel da Silva L (2022) The challenge of urban food production and sustainable water use: current situation and future perspectives of the urban agriculture in Brazil and Italy. Sustain Cities Soc 83:103961. [https://](https://doi.org/10.1016/j.scs.2022.103961) [doi.org/10.1016/j.scs.2022.103961](https://doi.org/10.1016/j.scs.2022.103961)
- <span id="page-26-11"></span>Alkalbani NS et al (2022) Assessment of yeasts as potential probiotics: a review of gastrointestinal tract conditions and investigation methods. J Fungi 8:365
- <span id="page-26-5"></span>Alloul A, Wuyts S, Lebeer S, Vlaeminck SE (2019) Volatile fatty acids impacting phototrophic growth kinetics of purple bacteria: Paving the way for protein production on fermented wastewater. Water Res 152:138–147. [https://](https://doi.org/10.1016/j.watres.2018.12.025) [doi.org/10.1016/j.watres.2018.12.025](https://doi.org/10.1016/j.watres.2018.12.025)
- <span id="page-26-24"></span>Alloul A, Wille M, Lucenti P, Bossier P, Van Stappen G, Vlaeminck SE (2021) Purple bacteria as added-value protein ingredient in shrimp feed: Penaeus vannamei growth performance and tolerance against vibrio and ammonia stress. Aquaculture 530:735788. [https://doi.org/10.](https://doi.org/10.1016/j.aquaculture.2020.735788) [1016/j.aquaculture.2020.735788](https://doi.org/10.1016/j.aquaculture.2020.735788)
- <span id="page-26-3"></span>Alloul A, Spanoghe J, Machado D, Vlaeminck SE (2022) Unlocking the genomic potential of aerobes and phototrophs for the production of nutritious and palatable microbial food without arable land or fossil fuels. Microbial Biotechnol. [https://doi.org/10.1111/1751-7915.](https://doi.org/10.1111/1751-7915.13747) [13747](https://doi.org/10.1111/1751-7915.13747)
- <span id="page-26-15"></span>Almomani F, Al Ketife A, Judd S, Shurair M, Bhosale RR, Znad H, Tawalbeh M (2019) Impact of  $CO<sub>2</sub>$  concentration and ambient conditions on microalgal growth and nutrient removal from wastewater by a photobioreactor. Sci Total Environ 662:662–671. [https://doi.org/10.](https://doi.org/10.1016/j.scitotenv.2019.01.144) [1016/j.scitotenv.2019.01.144](https://doi.org/10.1016/j.scitotenv.2019.01.144)
- <span id="page-26-13"></span>Almomani F, Judd S, Bhosale RR, Shurair M, Aljaml K, Khraisheh M (2019b) Intergraded wastewater treatment and carbon bio-fxation from fue gases using Spirulina platensis and mixed algal culture. Process Saf Environ Prot 124:240–250. [https://doi.org/10.1016/j.psep.2019.](https://doi.org/10.1016/j.psep.2019.02.009) [02.009](https://doi.org/10.1016/j.psep.2019.02.009)
- <span id="page-26-20"></span>Angelidaki I, Treu L, Tsapekos P, Luo G, Campanaro S, Wenzel H, Kougias PG (2018) Biogas upgrading and utilization: Current status and perspectives. Biotechnol Adv 36:452–466. [https://doi.org/10.1016/j.biotechadv.2018.](https://doi.org/10.1016/j.biotechadv.2018.01.011) [01.011](https://doi.org/10.1016/j.biotechadv.2018.01.011)
- <span id="page-26-12"></span>Areniello M, Matassa S, Esposito G, Lens PNL (2023) Biowaste upcycling into second-generation microbial protein through mixed-culture fermentation. Trends Biotechnol 41:197–213. [https://doi.org/10.1016/j.tibtech.2022.07.](https://doi.org/10.1016/j.tibtech.2022.07.008) [008](https://doi.org/10.1016/j.tibtech.2022.07.008)
- <span id="page-26-1"></span>Asiri F, Chu K-H (2022) Valorization of agro-industrial wastes into polyhydroxyalkanoates-rich single-cell proteins to enable a circular waste-to-feed economy. Chemosphere 309:136660. [https://doi.org/10.1016/j.chemosphere.](https://doi.org/10.1016/j.chemosphere.2022.136660) [2022.136660](https://doi.org/10.1016/j.chemosphere.2022.136660)
- <span id="page-26-8"></span>Awad Saad Allah AS (2021) Production of single-cell protein from some fruit peels annals of agricultural science. Moshtohor 59:755–762. [https://doi.org/10.21608/assjm.](https://doi.org/10.21608/assjm.2021.207308) [2021.207308](https://doi.org/10.21608/assjm.2021.207308)
- <span id="page-26-16"></span>Ayre JM, Moheimani NR, Borowitzka MA (2017) Growth of microalgae on undiluted anaerobic digestate of piggery effluent with high ammonium concentrations. Algal Res 24:218–226.<https://doi.org/10.1016/j.algal.2017.03.023>
- <span id="page-26-21"></span>Azarpour A, Zendehboudi S, Mohammadzadeh O, Rajabzadeh AR, Chatzis I (2022) A review on microalgal biomass and biodiesel production through co-cultivation strategy. Energy Convers Manag 267:115757. [https://doi.org/10.](https://doi.org/10.1016/j.enconman.2022.115757) [1016/j.enconman.2022.115757](https://doi.org/10.1016/j.enconman.2022.115757)
- <span id="page-26-22"></span>Balagurunathan B, Ling H, Choi WJ, Chang MW (2022) Potential use of microbial engineering in single-cell protein production. Curr Opin Biotechnol 76:102740. <https://doi.org/10.1016/j.copbio.2022.102740>
- <span id="page-26-18"></span>Banks M, Johnson R, Giver L, Bryant G, Guo M (2022) Industrial production of microbial protein products. Curr Opin Biotechnol 75:102707. [https://doi.org/10.1016/j.copbio.](https://doi.org/10.1016/j.copbio.2022.102707) [2022.102707](https://doi.org/10.1016/j.copbio.2022.102707)
- <span id="page-26-10"></span>Barzee TJ, Cao L, Pan Z, Zhang R (2021) Fungi for future foods. Journal of Future Foods 1:25–37. [https://doi.org/](https://doi.org/10.1016/j.jfutfo.2021.09.002) [10.1016/j.jfutfo.2021.09.002](https://doi.org/10.1016/j.jfutfo.2021.09.002)
- <span id="page-26-9"></span>Bayat Kohsar J, Rezaii F, Mahmoudnia N, Ghanbari F (2021) The effect of fermentation by Bacillus subtilis and Aspergillus niger on the nutritional value of date palm kernels. J Livestock Sci Technol 9:41–50. [https://doi.org/10.](https://doi.org/10.22103/jlst.2021.17236.1359) [22103/jlst.2021.17236.1359](https://doi.org/10.22103/jlst.2021.17236.1359)
- <span id="page-26-6"></span>Bertasini D, Binati RL, Bolzonella D, Battista F (2022) Single cell proteins production from food processing effluents and digestate. Chemosphere 296:134076. [https://doi.org/](https://doi.org/10.1016/j.chemosphere.2022.134076) [10.1016/j.chemosphere.2022.134076](https://doi.org/10.1016/j.chemosphere.2022.134076)
- <span id="page-26-23"></span>Biswas A et al (2020) Methanotroph (Methylococcus capsulatus, Bath) bacteria meal as an alternative protein source for Japanese yellowtail. Seriola Quinqueradiata Aquaculture 529:735700. [https://doi.org/10.1016/j.aquaculture.](https://doi.org/10.1016/j.aquaculture.2020.735700) [2020.735700](https://doi.org/10.1016/j.aquaculture.2020.735700)
- <span id="page-26-14"></span>Bolognesi S, Bañeras L, Perona-Vico E, Capodaglio AG, Balaguer MD, Puig S (2022) Carbon dioxide to bio-oil in a bioelectrochemical system-assisted microalgae biorefnery process Sustainable. Energy Fuels 6:150–161. [https://](https://doi.org/10.1039/D1SE01701B) [doi.org/10.1039/D1SE01701B](https://doi.org/10.1039/D1SE01701B)
- <span id="page-26-19"></span>Bonan CIDG et al (2022) Biorefnery platform for spathaspora passalidarum NRRL Y-27907 in the production of ethanol xylitol, and single cell protein from sugarcane bagasse. BioEnergy Res 15:1169–1181. [https://doi.org/](https://doi.org/10.1007/s12155-021-10255-7) [10.1007/s12155-021-10255-7](https://doi.org/10.1007/s12155-021-10255-7)
- <span id="page-26-2"></span>Bourdichon F et al (2012) Food fermentations: microorganisms with technological beneficial use. Int J Food Microbiol 154:87–97. [https://doi.org/10.1016/j.ijfoodmicro.2011.](https://doi.org/10.1016/j.ijfoodmicro.2011.12.030) [12.030](https://doi.org/10.1016/j.ijfoodmicro.2011.12.030)
- <span id="page-26-17"></span>Cantera S, Lebrero R, García-Encina PA, Muñoz R (2016) Evaluation of the infuence of methane and copper concentration and methane mass transport on the community structure and biodegradation kinetics of methanotrophic cultures. J Environ Manag 171:11–20. [https://doi.org/10.](https://doi.org/10.1016/j.jenvman.2016.02.002) [1016/j.jenvman.2016.02.002](https://doi.org/10.1016/j.jenvman.2016.02.002)
- <span id="page-26-7"></span>Cao K, Zhi R, Li Q, Zhang G, Wang H (2021) Photosynthetic bacterial protein production from wastewater: effects of C/N and light-oxygen condition. J Water Process Eng 44:102361. <https://doi.org/10.1016/j.jwpe.2021.102361>
- <span id="page-26-4"></span>Capanoglu E, Nemli E, Tomas-Barberan F (2022) Novel approaches in the valorization of agricultural wastes and

their applications. J Agric Food Chem 70:6787–6804. <https://doi.org/10.1021/acs.jafc.1c07104>

- <span id="page-27-4"></span>Capson-Tojo G et al (2020) Purple phototrophic bacteria for resource recovery: challenges and opportunities. Biotechnol Adv 43:107567. [https://doi.org/10.1016/j.biotechadv.](https://doi.org/10.1016/j.biotechadv.2020.107567) [2020.107567](https://doi.org/10.1016/j.biotechadv.2020.107567)
- <span id="page-27-17"></span>Carone M, Alpe D, Costantino V, Derossi C, Occhipinti A, Zanetti M, Riggio VA (2022) Design and characterization of a new pressurized fat panel photobioreactor for microalgae cultivation and  $CO<sub>2</sub>$  bio-fixation. Chemosphere 307:135755. [https://doi.org/10.1016/j.chemo](https://doi.org/10.1016/j.chemosphere.2022.135755) [sphere.2022.135755](https://doi.org/10.1016/j.chemosphere.2022.135755)
- <span id="page-27-12"></span>Carranza-Méndez RC, Chávez-González ML, Sepúlveda-Torre L, Aguilar CN, Govea-Salas M, Ramos-González R (2022) Production of single cell protein from orange peel residues by Candida utilis. Biocatal Agricul Biotechnol 40:102298. <https://doi.org/10.1016/j.bcab.2022.102298>
- <span id="page-27-16"></span>Cerda A, Artola A, Barrena R, Font X, Gea T, Sánchez A (2019) Innovative production of bioproducts from organic waste through solid-state fermentation. Front Sustain Food Syst. [https://doi.org/10.3389/fsufs.2019.](https://doi.org/10.3389/fsufs.2019.00063) [00063](https://doi.org/10.3389/fsufs.2019.00063)
- <span id="page-27-19"></span>Cevik E et al (2020) Direct electricity production from microalgae Choricystis sp. and investigation of the boron to enhance the electrogenic activity. Int J Hydrogen Energy 45:11330-11340. https://doi.org/10.1016/i.iihydene. [https://doi.org/10.1016/j.ijhydene.](https://doi.org/10.1016/j.ijhydene.2020.02.077) [2020.02.077](https://doi.org/10.1016/j.ijhydene.2020.02.077)
- <span id="page-27-8"></span>Chanda S, Chakrabarti S (1996) Plant origin liquid waste: a resource for singlecell protein production by yeast. Biores Technol 57:51–54. [https://doi.org/10.1016/0960-](https://doi.org/10.1016/0960-8524(96)00053-3) [8524\(96\)00053-3](https://doi.org/10.1016/0960-8524(96)00053-3)
- <span id="page-27-3"></span>Chandra MS, Srinivasulu M, Yadav PS, Ramesh B, Narasimha G, Chandrasekhar T (2021) Value added products from agriculture, paper and food waste: a source of bioenergy production. In: Srivastava M, Srivastava N, Singh R (eds) Bioenergy research: commercial opportunities & challenges. Springer Singapore, Singapore, pp 91–126. [https://doi.org/10.1007/](https://doi.org/10.1007/978-981-16-1190-2_3) [978-981-16-1190-2\\_3](https://doi.org/10.1007/978-981-16-1190-2_3)
- <span id="page-27-6"></span>Chee JY, Lakshmanan M, Jeepery IF, Mohamad Hairudin NH, Sudesh K (2019) The potential application of cupriavidus necator as polyhydroxyalkanoates producer and single cell protein: a review on scientifc. Cult Religious Perspect Appl Food Biotechnol 6:19-34. [https://doi.org/10.](https://doi.org/10.22037/afb.v6i1.22234) [22037/afb.v6i1.22234](https://doi.org/10.22037/afb.v6i1.22234)
- <span id="page-27-18"></span>Chen C-Y, Liu C-H, Lo Y-C, Chang J-S (2011) Perspectives on cultivation strategies and photobioreactor designs for photo-fermentative hydrogen production. Biores Technol 102:8484–8492. [https://doi.org/10.1016/j.biortech.2011.](https://doi.org/10.1016/j.biortech.2011.05.082) [05.082](https://doi.org/10.1016/j.biortech.2011.05.082)
- <span id="page-27-21"></span>Chen Y-Z et al (2022) Sustainable treatment of nitrate-containing wastewater by an autotrophic hydrogen-oxidizing bacterium. Environ Sci Ecotechnol 9:100146. [https://doi.](https://doi.org/10.1016/j.ese.2022.100146) [org/10.1016/j.ese.2022.100146](https://doi.org/10.1016/j.ese.2022.100146)
- <span id="page-27-7"></span>Chowdhary P, Gupta A, Gnansounou E, Pandey A, Chaturvedi P (2021) Current trends and possibilities for exploitation of Grape pomace as a potential source for value addition. Environ Pollut 278:116796. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.envpol.2021.116796) [envpol.2021.116796](https://doi.org/10.1016/j.envpol.2021.116796)
- <span id="page-27-13"></span>Coimbra JM, dos Cristina Reis K, Schwan RF, Silva CF (2021) Efect of the strategy of molasses supplementation in

vinasse to high SCP production and rose favor compound. Waste Biomass Valorization 12:359–369. [https://](https://doi.org/10.1007/s12649-020-00961-2) [doi.org/10.1007/s12649-020-00961-2](https://doi.org/10.1007/s12649-020-00961-2)

- <span id="page-27-20"></span>Comesaña-Gándara B, García-Depraect O, Santos-Beneit F, Bordel S, Lebrero R, Muñoz R (2022) Recent trends and advances in biogas upgrading and methanotrophs-based valorization. Chem Eng J Adv 11:100325. [https://doi.](https://doi.org/10.1016/j.ceja.2022.100325) [org/10.1016/j.ceja.2022.100325](https://doi.org/10.1016/j.ceja.2022.100325)
- <span id="page-27-23"></span>Dantas EM Jr, Valle BCS, Brito CMS, Calazans NKF, Peixoto SRM, Soares RB (2016) Partial replacement of fshmeal with biofoc meal in the diet of postlarvae of the Pacifc white shrimp Litopenaeus vannamei. Aquac Nutr 22:335–342.<https://doi.org/10.1111/anu.12249>
- <span id="page-27-5"></span>de Nascimento JC et al (2022) Single cell protein production by Candida robusta isolated from sugar cane (Saccharum sp.) for animal feed Produção de Biomassa por candida robusta isolada da Cana-de-açúcar (Saccharum sp.) para alimentação animal. J Develop 8:56092–56105. [https://](https://doi.org/10.34117/bjdv8n8-081) [doi.org/10.34117/bjdv8n8-081](https://doi.org/10.34117/bjdv8n8-081)
- <span id="page-27-15"></span>Deseure J, Obeid J, Willison JC, Magnin J-P (2021) Reliable determination of the growth and hydrogen production parameters of the photosynthetic bacterium rhodobacter capsulatus in fed batch culture using a combination of the Gompertz function and the luedeking-piret model. Heliyon 7:e07394. [https://doi.org/10.1016/j.heliyon.2021.](https://doi.org/10.1016/j.heliyon.2021.e07394) [e07394](https://doi.org/10.1016/j.heliyon.2021.e07394)
- <span id="page-27-0"></span>Dey T, Bhattacharjee T, Nag P, Ghati A, Kuila A (2021) Valorization of agro-waste into value added products for sustainable development. Bioresour Technol Rep 16:100834. <https://doi.org/10.1016/j.biteb.2021.100834>
- <span id="page-27-14"></span>Dias B, Lopes M, Ramôa R, Pereira AS, Belo I (2021) Candida tropicalis as a promising oleaginous yeast for olive mill wastewater bioconversion. Energies 14:640
- <span id="page-27-1"></span>Díaz-Vázquez D, Carrillo-Nieves D, Orozco-Nunnelly DA, Senés-Guerrero C, Gradilla-Hernández MS (2021) An integrated approach for the assessment of environmental sustainability in Agro-industrial waste management practices: the case of the Tequila industry. Front Environ Sci. <https://doi.org/10.3389/fenvs.2021.682093>
- <span id="page-27-2"></span>Ding H et al (2023) Ammonia nitrogen recovery from biogas slurry by SCP production using Candida utilis. J Environ Manag 325:116657. [https://doi.org/10.1016/j.jenvman.](https://doi.org/10.1016/j.jenvman.2022.116657) [2022.116657](https://doi.org/10.1016/j.jenvman.2022.116657)
- <span id="page-27-11"></span>Dunuweera AN, Nikagolla DN, Ranganathan K (2021) Fruit waste substrates to produce single-cell proteins as alternative human food supplements and animal feeds using baker's yeast (Saccharomyces cerevisiae). J Food Quality 2021:9932762. <https://doi.org/10.1155/2021/9932762>
- <span id="page-27-22"></span>Dupnock TL, Deshusses MA (2019) Detailed investigations of dissolved hydrogen and hydrogen mass transfer in a biotrickling flter for upgrading biogas. Bioresource Technol 290:121780. [https://doi.org/10.1016/j.biortech.](https://doi.org/10.1016/j.biortech.2019.121780) [2019.121780](https://doi.org/10.1016/j.biortech.2019.121780)
- <span id="page-27-9"></span>EFSA Panel on Dietetic Products N et al. (2016) Guidance on the preparation and presentation of an application for authorisation of a novel food in the context of regulation (EU) 2015/2283 EFSA Journal 14: e04594 [https://doi.](https://doi.org/10.2903/j.efsa.2016.4594) [org/10.2903/j.efsa.2016.4594](https://doi.org/10.2903/j.efsa.2016.4594)
- <span id="page-27-10"></span>EFSA Panel on Nutrition NF et al. (2019) Safety of Yarrowia lipolytica yeast biomass as a novel food pursuant to

regulation (EU) 2015/2283 EFSA Journal 17: e05594 <https://doi.org/10.2903/j.efsa.2019.5594>

- <span id="page-28-15"></span>Elmaadawy K et al (2022) Microalgae-assisted fxed-flm activated sludge MFC for landfll leachate treatment and energy recovery. Process Saf Environ Prot 160:221–231. <https://doi.org/10.1016/j.psep.2022.02.021>
- <span id="page-28-16"></span>Elshobary ME, Zabed HM, Yun J, Zhang G, Qi X (2021) Recent insights into microalgae-assisted microbial fuel cells for generating sustainable bioelectricity. Int J Hydrogen Energy 46:3135–3159. [https://doi.org/10.](https://doi.org/10.1016/j.ijhydene.2020.06.251) [1016/j.ijhydene.2020.06.251](https://doi.org/10.1016/j.ijhydene.2020.06.251)
- <span id="page-28-2"></span>EniferBio (2022) PEKILO® the all-round protein of the future. eniferBio. [https://www.eniferbio.f/product/.](https://www.eniferbio.fi/product/) Accessed 09 Nov 2022 2022
- <span id="page-28-22"></span>Esmaeili M, Abedian Kenari A, Rombenso AN (2017) Efects of fsh meal replacement with meat and bone meal using garlic (Allium sativum) powder on growth, feeding, digestive enzymes and apparent digestibility of nutrients and fatty acids in juvenile rainbow trout (Oncorhynchus mykiss Walbaum, 1792). Aquacult Nutr 23:1225–1234. <https://doi.org/10.1111/anu.12491>
- <span id="page-28-3"></span>Farhan SN, Mahmood WA, Ali Godeib AH (2021) Pre-evaporation of ethanol as an efective method to improve single cell protein (SCP) production from date palm residue by saccharomyces cerevisiae. IOP Conf Ser Mater Sci Eng 1076:012029. [https://doi.org/10.1088/1757-899X/](https://doi.org/10.1088/1757-899X/1076/1/012029) [1076/1/012029](https://doi.org/10.1088/1757-899X/1076/1/012029)
- <span id="page-28-9"></span>Ferreira JA, Mahboubi A, Lennartsson PR, Taherzadeh MJ (2016) Waste biorefneries using flamentous ascomycetes fungi: Present status and future prospects. Biores Technol 215:334–345. [https://doi.org/10.1016/j.biortech.](https://doi.org/10.1016/j.biortech.2016.03.018) [2016.03.018](https://doi.org/10.1016/j.biortech.2016.03.018)
- <span id="page-28-23"></span>Finnigan TJA, Wall BT, Wilde PJ, Stephens FB, Taylor SL, Freedman MR (2019) Mycoprotein: the future of nutritious nonmeat protein, a symposium review. Curr Develop Nutr.<https://doi.org/10.1093/cdn/nzz021>
- <span id="page-28-14"></span>Gabrielyan DA et al (2022) Cultivation of Chlorella sorokiniana IPPAS C-1 in fat-panel photobioreactors: from a laboratory to a pilot scale. Life 12:1309
- <span id="page-28-4"></span>Gao Y, Li D, Liu Y (2012) Production of single cell protein from soy molasses using Candida tropicalis. Ann<br>Microbiol 62:1165-1172. https://doi.org/10.1007/ [https://doi.org/10.1007/](https://doi.org/10.1007/s13213-011-0356-9) [s13213-011-0356-9](https://doi.org/10.1007/s13213-011-0356-9)
- <span id="page-28-11"></span>García JL, de Vicente M, Galán B (2017) Microalgae, old sustainable food and fashion nutraceuticals. Microbial Biotechnol 10:1017–1024. [https://doi.org/10.1111/1751-](https://doi.org/10.1111/1751-7915.12800) [7915.12800](https://doi.org/10.1111/1751-7915.12800)
- <span id="page-28-0"></span>Garrity DP et al (2010) Evergreen agriculture: a robust approach to sustainable food security in Africa. Food Security 2:197–214. [https://doi.org/10.1007/](https://doi.org/10.1007/s12571-010-0070-7) [s12571-010-0070-7](https://doi.org/10.1007/s12571-010-0070-7)
- <span id="page-28-1"></span>Gervasi T, Pellizzeri V, Calabrese G, Di Bella G, Cicero N, Dugo G (2018) Production of single cell protein (SCP) from food and agricultural waste by using Saccharomyces cerevisiae. Nat Prod Res 32:648–653. [https://doi.org/](https://doi.org/10.1080/14786419.2017.1332617) [10.1080/14786419.2017.1332617](https://doi.org/10.1080/14786419.2017.1332617)
- <span id="page-28-17"></span>Gęsicka A, Oleskowicz-Popiel P, Łężyk M (2021) Recent trends in methane to bioproduct conversion by methanotrophs. Biotechnol Adv 53:107861. [https://doi.org/10.](https://doi.org/10.1016/j.biotechadv.2021.107861) [1016/j.biotechadv.2021.107861](https://doi.org/10.1016/j.biotechadv.2021.107861)
- <span id="page-28-5"></span>Ghaly AE, Kamal M, Correia LR (2005) Kinetic modelling of
- continuous submerged fermentation of cheese whey for single cell protein production. Biores Technol 96:1143– 1152. <https://doi.org/10.1016/j.biortech.2004.09.027>
- <span id="page-28-20"></span>Glencross BD, Huyben D, Schrama JW (2020) The application of single-cell ingredients in aquaculture feeds—a review. Fishes 5:22
- <span id="page-28-13"></span>Godoy MG, Amorim GM, Barreto MS, Freire DMG (2018) Chapter 12-agricultural residues as animal feed: protein enrichment and detoxifcation using solid-state fermentation. In: Pandey A, Larroche C, Soccol CR (eds) Current developments in biotechnology and bioengineering. Elsevier, pp 235–256. [https://doi.org/10.1016/B978-0-444-](https://doi.org/10.1016/B978-0-444-63990-5.00012-8) [63990-5.00012-8](https://doi.org/10.1016/B978-0-444-63990-5.00012-8)
- <span id="page-28-19"></span>Golmakani A, Ali Nabavi S, Wadi B, Manovic V (2022) Advances, challenges, and perspectives of biogas cleaning, upgrading, and utilisation. Fuel 317:123085. [https://](https://doi.org/10.1016/j.fuel.2021.123085) [doi.org/10.1016/j.fuel.2021.123085](https://doi.org/10.1016/j.fuel.2021.123085)
- <span id="page-28-12"></span>Goonesekera EM, Tsapekos P, Angelidaki I, Valverde-Pérez B (2022) Impact of recovered phosphorus supply on methanotrophic cultivation and microbial protein production. J Environ Manag 322:115820. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jenvman.2022.115820) [jenvman.2022.115820](https://doi.org/10.1016/j.jenvman.2022.115820)
- <span id="page-28-7"></span>Groenewald M, Boekhout T, Neuvéglise C, Gaillardin C, van Dijck PWM, Wyss M (2014) Yarrowia lipolytica: safety assessment of an oleaginous yeast with a great industrial potential. Crit Rev Microbiol 40:187–206. [https://doi.](https://doi.org/10.3109/1040841X.2013.770386) [org/10.3109/1040841X.2013.770386](https://doi.org/10.3109/1040841X.2013.770386)
- <span id="page-28-8"></span>Gunun N et al (2022) Efects of rubber seed kernel fermented with yeast on feed utilization, rumen fermentation and microbial protein synthesis in dairy heifers. Fermentation 8:288
- <span id="page-28-18"></span>Haraldsen TK, Andersen U, Krogstad T, Sørheim R (2011) Liquid digestate from anaerobic treatment of sourceseparated household waste as fertilizer to barley. Waste Manag Res 29:1271–1276. [https://doi.org/10.1177/](https://doi.org/10.1177/0734242X11411975) [0734242X11411975](https://doi.org/10.1177/0734242X11411975)
- <span id="page-28-21"></span>Hardy RW, Patro B, Pujol-Baxley C, Marx CJ, Feinberg L (2018) Partial replacement of soybean meal with methylobacterium extorquens single-cell protein in feeds for rainbow trout (Oncorhynchus mykiss Walbaum). Aquacult Res 49:2218–2224. [https://doi.org/10.1111/](https://doi.org/10.1111/are.13678) [are.13678](https://doi.org/10.1111/are.13678)
- <span id="page-28-10"></span>Hashem M, Al-Qahtani MS, Alamri SA, Moustafa YS, Lyberatos G, Ntaikou I (2022) Valorizing food wastes: assessment of novel yeast strains for enhanced production of single-cell protein from wasted date molasses. Biomass Convers Biorefnery 12:4491–4502. [https://doi.org/10.](https://doi.org/10.1007/s13399-022-02415-2) [1007/s13399-022-02415-2](https://doi.org/10.1007/s13399-022-02415-2)
- <span id="page-28-24"></span>Hashempour-Baltork F, Khosravi-Darani K, Hosseini H, Farshi P, Reihani SFS (2020) Mycoproteins as safe meat substitutes. J Clean Prod 253:119958. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jclepro.2020.119958) [jclepro.2020.119958](https://doi.org/10.1016/j.jclepro.2020.119958)
- <span id="page-28-6"></span>Hashempour-Baltork F, Farshi P, Khosravi-Darani K (2022) Vegetable and fruit wastes as substrate for production of single-cell protein and aquafeed meal. In: Ray RC (ed) Fruits and vegetable wastes : valorization to bioproducts and platform chemicals. Springer Nature Singapore, Singapore, pp 169–187. [https://doi.org/10.1007/](https://doi.org/10.1007/978-981-16-9527-8_7) [978-981-16-9527-8\\_7](https://doi.org/10.1007/978-981-16-9527-8_7)
- <span id="page-29-6"></span>He J, Zhang G, Lu H (2010) Treatment of soybean wastewater by a wild strain rhodobacter sphaeroides and to produce protein under natural conditions. Front Environ Sci Eng China 4:334–339. [https://doi.org/10.1007/](https://doi.org/10.1007/s11783-010-0239-5) [s11783-010-0239-5](https://doi.org/10.1007/s11783-010-0239-5)
- <span id="page-29-21"></span>Hu X, Vandamme P, Boon N (2022) Co-cultivation enhanced microbial protein production based on autotrophic nitrogen-fxing hydrogen-oxidizing bacteria. Chem Eng J 429:132535.<https://doi.org/10.1016/j.cej.2021.132535>
- <span id="page-29-17"></span>Hülsen T, Hsieh K, Lu Y, Tait S, Batstone DJ (2018) Simultaneous treatment and single cell protein production from agri-industrial wastewaters using purple phototrophic bacteria or microalgae–A comparison. Bioresource Technol 254:214–223. [https://doi.org/10.1016/j.biortech.](https://doi.org/10.1016/j.biortech.2018.01.032) [2018.01.032](https://doi.org/10.1016/j.biortech.2018.01.032)
- <span id="page-29-20"></span>Hülsen T, Hsieh K, Tait S, Barry EM, Puyol D, Batstone DJ (2018) White and infrared light continuous photobioreactors for resource recovery from poultry processing wastewater–a comparison. Water Res 144:665–676. [https://](https://doi.org/10.1016/j.watres.2018.07.040) [doi.org/10.1016/j.watres.2018.07.040](https://doi.org/10.1016/j.watres.2018.07.040)
- <span id="page-29-18"></span>Hülsen T et al (2022) Outdoor demonstration-scale flat plate photobioreactor for resource recovery with purple phototrophic bacteria. Water Res 216:118327. [https://doi.org/](https://doi.org/10.1016/j.watres.2022.118327) [10.1016/j.watres.2022.118327](https://doi.org/10.1016/j.watres.2022.118327)
- <span id="page-29-5"></span>Hülsen T, Stegman S, Batstone DJ, Capson-Tojo G (2022) Naturally illuminated photobioreactors for resource recovery from piggery and chicken-processing wastewaters utilising purple phototrophic bacteria. Water Res 214:118194. <https://doi.org/10.1016/j.watres.2022.118194>
- <span id="page-29-25"></span>Humpenöder F, Bodirsky BL, Weindl I, Lotze-Campen H, Linder T, Popp A (2022) Projected environmental benefts of replacing beef with microbial protein. Nature 605:90–96. <https://doi.org/10.1038/s41586-022-04629-w>
- <span id="page-29-7"></span>Ibrahim Rajoka M, Tariq Kiani MA, Khan S, Awan MS, Hashmi A-S (2004) Production of single cell protein from rice polishings using Candida utilis. World J Microbiol Biotechnol 20:297–301. [https://doi.org/10.1023/B:](https://doi.org/10.1023/B:WIBI.0000023845.96123.dd) [WIBI.0000023845.96123.dd](https://doi.org/10.1023/B:WIBI.0000023845.96123.dd)
- <span id="page-29-19"></span>Jadhav DA, Neethu B, Ghangrekar MM (2019) Microbial carbon capture cell: advanced bio-electrochemical system for wastewater treatment, electricity generation and algal biomass production. In: Gupta SK, Bux F (eds) Application of microalgae in wastewater treatment: Biorefnery approaches of wastewater treatment, vol 2. Springer International Publishing, Cham, pp 317–338. [https://doi.](https://doi.org/10.1007/978-3-030-13909-4_14) [org/10.1007/978-3-030-13909-4\\_14](https://doi.org/10.1007/978-3-030-13909-4_14)
- <span id="page-29-10"></span>Janssen M, Wijfels RH, Barbosa MJ (2022) Microalgae based production of single-cell protein. Curr Opin Biotechnol 75:102705. <https://doi.org/10.1016/j.copbio.2022.102705>
- <span id="page-29-2"></span>Jiang Y, Yang X, Zeng D, Su Y, Zhang Y (2022) Microbial conversion of syngas to single cell protein: The role of carbon monoxide. Chem Eng J 450:138041. [https://doi.](https://doi.org/10.1016/j.cej.2022.138041) [org/10.1016/j.cej.2022.138041](https://doi.org/10.1016/j.cej.2022.138041)
- <span id="page-29-13"></span>Jintasataporn O, Chumkam S, Triwutanon S, LeBlanc A, Sawanboonchun J (2021) Efects of a single cell protein (Methylococcus capsulatus, bath) in Pacifc white shrimp (Penaeus vannamei) diet on growth performance, survival rate and resistance to vibrio parahaemolyticus, the causative agent of acute hepatopancreatic necrosis disease. Front Marine Sci. [https://doi.org/10.3389/fmars.](https://doi.org/10.3389/fmars.2021.764042) [2021.764042](https://doi.org/10.3389/fmars.2021.764042)
- <span id="page-29-24"></span>Johnson EA (2013) Biotechnology of non-saccharomyces yeasts—the ascomycetes. Appl Microbiol Biotechnol 97:503–517.<https://doi.org/10.1007/s00253-012-4497-y>
- <span id="page-29-11"></span>Jonaitis T et al (2022) Subchronic feeding allergenicity and genotoxicity safety evaluations of single strain bacterial protein. Food Chem Toxicol 162:112878. [https://doi.org/](https://doi.org/10.1016/j.fct.2022.112878) [10.1016/j.fct.2022.112878](https://doi.org/10.1016/j.fct.2022.112878)
- <span id="page-29-1"></span>Jones SW, Karpol A, Friedman S, Maru BT, Tracy BP (2020) Recent advances in single cell protein use as a feed ingredient in aquaculture. Curr Opin Biotechnol 61:189–197. <https://doi.org/10.1016/j.copbio.2019.12.026>
- <span id="page-29-4"></span>Kalyuzhnaya MG et al (2013) Highly efficient methane biocatalysis revealed in a methanotrophic bacterium Nature. Communications 4:2785. [https://doi.org/10.1038/ncomm](https://doi.org/10.1038/ncomms3785) [s3785](https://doi.org/10.1038/ncomms3785)
- <span id="page-29-9"></span>Kerckhof F-M et al (2021) From biogas and hydrogen to microbial protein through co-cultivation of methane and hydrogen oxidizing bacteria. Front Bioeng Biotechnol. <https://doi.org/10.3389/fbioe.2021.733753>
- <span id="page-29-0"></span>Kesavan PC, Swaminathan MS (2008) Strategies and models for agricultural sustainability in developing Asian countries. Philosoph Trans Royal Soc B Biol Sci 363:877– 891.<https://doi.org/10.1098/rstb.2007.2189>
- <span id="page-29-8"></span>Khan MKI, Asif M, Razzaq ZU, Nazir A, Maan AA (2022) Sustainable food industrial waste management through single cell protein production and characterization of protein enriched bread. Food Biosci 46:101406. [https://](https://doi.org/10.1016/j.fbio.2021.101406) [doi.org/10.1016/j.fbio.2021.101406](https://doi.org/10.1016/j.fbio.2021.101406)
- <span id="page-29-16"></span>Khonngam T, Salakkam A (2019) Bioconversion of sugarcane bagasse and dry spent yeast to ethanol through a sequential process consisting of solid-state fermentation, hydrolysis, and submerged fermentation. Biochem Eng J 150:107284.<https://doi.org/10.1016/j.bej.2019.107284>
- <span id="page-29-23"></span>Khoshnevisan B et al (2020) Coupling electrochemical ammonia extraction and cultivation of methane oxidizing bacteria for production of microbial protein. J Environ Manag 265:110560. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jenvman.2020.110560) [jenvman.2020.110560](https://doi.org/10.1016/j.jenvman.2020.110560)
- <span id="page-29-3"></span>Khoshnevisan B et al (2022) From renewable energy to sustainable protein sources: advancement, challenges, and future roadmaps. Renew Sustain Energy Rev 157:112041. <https://doi.org/10.1016/j.rser.2021.112041>
- <span id="page-29-14"></span>Kim M, Li S, Song YE, Lee D-Y, Kim JR (2022) Electrodeattached cell-driven biogas upgrading of anaerobic digestion effluent  $CO<sub>2</sub>$  to  $CH<sub>4</sub>$  using a microbial electrosynthesis cell. Chem Eng J 446:137079. [https://doi.](https://doi.org/10.1016/j.cej.2022.137079) [org/10.1016/j.cej.2022.137079](https://doi.org/10.1016/j.cej.2022.137079)
- <span id="page-29-12"></span>Kondo K, Taguchi C (2022) Japanese regulatory framework and approach for genome-edited foods based on latest scientifc fndings. Food Saf 10:113–128. [https://doi.](https://doi.org/10.14252/foodsafetyfscj.D-21-00016) [org/10.14252/foodsafetyfscj.D-21-00016](https://doi.org/10.14252/foodsafetyfscj.D-21-00016)
- <span id="page-29-15"></span>Kulkarni PP, Khonde VK, Deshpande MS, Sabale TR, Kumbhar PS, Ghosalkar AR (2021) Selection of methanotrophic platform for methanol production using methane and biogas. J Biosci Bioeng 132:460–468. <https://doi.org/10.1016/j.jbiosc.2021.07.007>
- <span id="page-29-22"></span>Kumar R, Singh L, Zularisam AW (2016) Exoelectrogens: recent advances in molecular drivers involved in extracellular electron transfer and strategies used to improve it for microbial fuel cell applications. Renew Sustain

Energy Rev 56:1322–1336. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.rser.2015.12.029) [rser.2015.12.029](https://doi.org/10.1016/j.rser.2015.12.029)

- <span id="page-30-1"></span>Kumar V, Ahluwalia V, Saran S, Kumar J, Patel AK, Singhania RR (2021) Recent developments on solid-state fermentation for production of microbial secondary metabolites: challenges and solutions. Bioresour Technol 323:124566. https://doi.org/10.1016/i.biortech. 323:124566. [https://doi.org/10.1016/j.biortech.](https://doi.org/10.1016/j.biortech.2020.124566) [2020.124566](https://doi.org/10.1016/j.biortech.2020.124566)
- <span id="page-30-0"></span>Kurcz A, Błażejak S, Kot AM, Bzducha-Wróbel A, Kieliszek M (2018) Application of industrial wastes for the production of microbial single-cell protein by fodder yeast candida utilis. Waste Biomass Valorization 9:57–64. [https://](https://doi.org/10.1007/s12649-016-9782-z) [doi.org/10.1007/s12649-016-9782-z](https://doi.org/10.1007/s12649-016-9782-z)
- <span id="page-30-11"></span>Kushwaha JP (2015) A review on sugar industry wastewater: sources, treatment technologies, and reuse. Desalin Water Treat 53:309–318. [https://doi.org/10.1080/19443](https://doi.org/10.1080/19443994.2013.838526) [994.2013.838526](https://doi.org/10.1080/19443994.2013.838526)
- <span id="page-30-5"></span>Lähteenmäki-Uutela A, Rahikainen M, Lonkila A, Yang B (2021) Alternative proteins and EU food law. Food Control 130:108336. [https://doi.org/10.1016/j.foodcont.2021.](https://doi.org/10.1016/j.foodcont.2021.108336) [108336](https://doi.org/10.1016/j.foodcont.2021.108336)
- <span id="page-30-22"></span>Lai C-Y, Zhou L, Yuan Z, Guo J (2021) Hydrogen-driven microbial biogas upgrading: advances, challenges and solutions. Water Res 197:117120. [https://doi.org/10.](https://doi.org/10.1016/j.watres.2021.117120) [1016/j.watres.2021.117120](https://doi.org/10.1016/j.watres.2021.117120)
- <span id="page-30-12"></span>Laskowska E, Jarosz Ł, Grądzki Z (2017) The efect of feed supplementation with effective microorganisms (EM) on pro- and anti-infammatory cytokine concentrations in pigs. Res Veterin Sci 115:244–249. [https://doi.org/10.](https://doi.org/10.1016/j.rvsc.2017.03.008) [1016/j.rvsc.2017.03.008](https://doi.org/10.1016/j.rvsc.2017.03.008)
- <span id="page-30-18"></span>Lee SY, Stuckey DC (2022) Separation and biosynthesis of value-added compounds from food-processing wastewater: towards sustainable wastewater resource recovery. J Clean Prod 357:131975. [https://doi.org/10.1016/j.jclep](https://doi.org/10.1016/j.jclepro.2022.131975) [ro.2022.131975](https://doi.org/10.1016/j.jclepro.2022.131975)
- <span id="page-30-4"></span>Lee JZ, Logan A, Terry S, Spear JR (2015) Microbial response to single-cell protein production and brewery wastewater treatment. Microb Biotechnol 8:65–76. [https://doi.org/](https://doi.org/10.1111/1751-7915.12128) [10.1111/1751-7915.12128](https://doi.org/10.1111/1751-7915.12128)
- <span id="page-30-20"></span>Leger D, Matassa S, Noor E, Shepon A, Milo R, Bar-Even A (2021) Photovoltaic-driven microbial protein production can use land and sunlight more efficiently than conventional crops. Proc Natl Acad Sci 118:e2015025118. <https://doi.org/10.1073/pnas.2015025118>
- <span id="page-30-2"></span>Leite P et al (2021) Recent advances in production of lignocellulolytic enzymes by solid-state fermentation of agro-industrial wastes. Curr Opin Green Sustain Chem 27:100407. <https://doi.org/10.1016/j.cogsc.2020.100407>
- <span id="page-30-23"></span>Leu AO et al (2020) Anaerobic methane oxidation coupled to manganese reduction by members of the Methanoperedenaceae. ISME J 14:1030–1041. [https://doi.org/10.](https://doi.org/10.1038/s41396-020-0590-x) [1038/s41396-020-0590-x](https://doi.org/10.1038/s41396-020-0590-x)
- <span id="page-30-7"></span>Li C, Li X, Qin L, Wu W, Meng Q, Shen C, Zhang G (2019) Membrane photo-bioreactor coupled with heterogeneous Fenton fuidized bed for high salinity wastewater treatment: Pollutant removal, photosynthetic bacteria harvest and membrane anti-fouling analysis. Sci Total Environ 696:133953. [https://doi.org/10.1016/j.scitotenv.2019.](https://doi.org/10.1016/j.scitotenv.2019.133953) [133953](https://doi.org/10.1016/j.scitotenv.2019.133953)
- <span id="page-30-21"></span>Li L, Zhao Y, Lian W, Han C, Zhang Q, Huang W (2021) Review on the effect of heat exchanger tubes on flow

behavior and heat/mass transfer of the bubble/slurry reactors. Chin J Chem Eng 35:44–61. [https://doi.org/10.](https://doi.org/10.1016/j.cjche.2021.03.017) [1016/j.cjche.2021.03.017](https://doi.org/10.1016/j.cjche.2021.03.017)

- <span id="page-30-17"></span>Li Y et al (2022) Multi-omics joint analysis of the effect of temperature on microbial communities, metabolism, and genetics in full-scale biogas reactors with food waste. Renew Sustain Energy Rev 160:112261. [https://doi.org/](https://doi.org/10.1016/j.rser.2022.112261) [10.1016/j.rser.2022.112261](https://doi.org/10.1016/j.rser.2022.112261)
- <span id="page-30-6"></span>Li Q et al (2022) Bioconversion of food waste to crayfsh feed using solid-state fermentation with yeast. Environ Sci Pollut Res. <https://doi.org/10.1007/s11356-022-23100-x>
- <span id="page-30-14"></span>Lim YA, Ilankoon IMSK, Chong MN, Foo SC (2023) Improving microalgae growth and carbon capture through micro-size bubbles generation in fat-panel photobioreactors: Impacts of diferent gas sparger designs on mixing performance. Renew Sustain Energy Rev 171:113001. <https://doi.org/10.1016/j.rser.2022.113001>
- <span id="page-30-19"></span>Lin L, Huang H, Zhang X, Dong L, Chen Y (2022) Hydrogenoxidizing bacteria and their applications in resource recovery and pollutant removal. Sci Total Environ 835:155559. https://doi.org/10.1016/j.scitotenv.2022. [https://doi.org/10.1016/j.scitotenv.2022.](https://doi.org/10.1016/j.scitotenv.2022.155559) [155559](https://doi.org/10.1016/j.scitotenv.2022.155559)
- <span id="page-30-24"></span>Linder T (2019) Making the case for edible microorganisms as an integral part of a more sustainable and resilient food production system. Food Security 11:265–278. [https://](https://doi.org/10.1007/s12571-019-00912-3) [doi.org/10.1007/s12571-019-00912-3](https://doi.org/10.1007/s12571-019-00912-3)
- <span id="page-30-3"></span>Liu B, Li Y, Song J, Zhang L, Dong J, Yang Q (2014) Production of single-cell protein with two-step fermentation for treatment of potato starch processing waste.<br>Cellulose 21:3637-3645. https://doi.org/10.1007/ [https://doi.org/10.1007/](https://doi.org/10.1007/s10570-014-0400-6) [s10570-014-0400-6](https://doi.org/10.1007/s10570-014-0400-6)
- <span id="page-30-13"></span>Liu S, Zhang G, Zhang J, Li X, Li J (2016) Performance, carotenoids yield and microbial population dynamics in a photobioreactor system treating acidic wastewater: effect of hydraulic retention time (HRT) and organic loading rate (OLR). Bioresource Technol 200:245–252. [https://](https://doi.org/10.1016/j.biortech.2015.10.044) [doi.org/10.1016/j.biortech.2015.10.044](https://doi.org/10.1016/j.biortech.2015.10.044)
- <span id="page-30-9"></span>Liu J, Sun H, Nie C, Ge W, Wang Y, Zhang W (2018) Oligopeptide derived from solid-state fermented cottonseed meal signifcantly afect the immunomodulatory in BALB/c mice treated with cyclophosphamide. Food Sci Biotechnol 27:1791–1799. [https://doi.org/10.1007/](https://doi.org/10.1007/s10068-018-0414-1) [s10068-018-0414-1](https://doi.org/10.1007/s10068-018-0414-1)
- <span id="page-30-10"></span>Liu J et al (2020) Comparative characterization of extracellular enzymes secreted by Phanerochaete chrysosporium during solid-state and submerged fermentation. Int J Biol Macromol 152:288–294. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.ijbiomac.2020.02.256) [ijbiomac.2020.02.256](https://doi.org/10.1016/j.ijbiomac.2020.02.256)
- <span id="page-30-8"></span>Lu H, Zhang G, Zheng Z, Meng F, Du T, He S (2019) Bioconversion of photosynthetic bacteria from non-toxic wastewater to realize wastewater treatment and bioresource recovery: a review. Biores Technol 278:383– 399. <https://doi.org/10.1016/j.biortech.2019.01.070>
- <span id="page-30-16"></span>Luo G, Johansson S, Boe K, Xie L, Zhou Q, Angelidaki I (2012) Simultaneous hydrogen utilization and in situ biogas upgrading in an anaerobic reactor. Biotechnol Bioeng 109:1088–1094. [https://doi.org/10.1002/bit.](https://doi.org/10.1002/bit.24360) [24360](https://doi.org/10.1002/bit.24360)
- <span id="page-30-15"></span>Maia IB et al (2022) Diel biochemical and photosynthetic monitorization of Skeletonema costatum and Phaeodactylum tricornutum grown in outdoor pilot-scale fat

panel photobioreactors. J Biotechnol 343:110–119. <https://doi.org/10.1016/j.jbiotec.2021.11.008>

- <span id="page-31-14"></span>Martău G-A, Unger P, Schneider R, Venus J, Vodnar DC, López-Gómez JP (2021) Integration of solid state and submerged fermentations for the valorization of organic municipal solid waste. J Fungi 7:766
- <span id="page-31-9"></span>Martin MR, Fornero JJ, Stark R, Mets L, Angenent LT (2013) A single-culture bioprocess of methanothermobacter thermautotrophicus to upgrade digester biogas by  $CO_2$ -to-CH<sub>4</sub> conversion with H<sub>2</sub>. Archaea 2013:157529. <https://doi.org/10.1155/2013/157529>
- <span id="page-31-22"></span>Matassa S, Batstone DJ, Hülsen T, Schnoor J, Verstraete W (2015) Can direct conversion of used nitrogen to new feed and protein help feed the world? Environ Sci Technol 49:5247–5254. <https://doi.org/10.1021/es505432w>
- <span id="page-31-20"></span>Matassa S, Verstraete W, Pikaar I, Boon N (2016) Autotrophic nitrogen assimilation and carbon capture for microbial protein production by a novel enrichment of hydrogen-oxidizing bacteria. Water Res 101:137–146. <https://doi.org/10.1016/j.watres.2016.05.077>
- <span id="page-31-3"></span>Matassa S, Pelagalli V, Papirio S, Zamalloa C, Verstraete W, Esposito G, Pirozzi F (2022) Direct nitrogen stripping and upcycling from anaerobic digestate during conversion of cheese whey into single cell protein. Bioresour Technol 358:127308. [https://doi.org/10.1016/j.biort](https://doi.org/10.1016/j.biortech.2022.127308) [ech.2022.127308](https://doi.org/10.1016/j.biortech.2022.127308)
- <span id="page-31-12"></span>Melnichuk N, Braia MJ, Anselmi PA, Meini M-R, Romanini D (2020) Valorization of two agroindustrial wastes to produce alpha-amylase enzyme from Aspergillus oryzae by solid-state fermentation. Waste Manag 106:155– 161. <https://doi.org/10.1016/j.wasman.2020.03.025>
- <span id="page-31-17"></span>Meng F, Yang A, Zhang G, Wang H (2017) Effects of dissolved oxygen concentration on photosynthetic bacteria wastewater treatment: pollutants removal, cell growth and pigments production. Bioresour Technol 241:993– 997. <https://doi.org/10.1016/j.biortech.2017.05.183>
- <span id="page-31-5"></span>Mensah JKM, Twumasi P (2017) Use of pineapple waste for single cell protein (SCP) production and the efect of substrate concentration on the yield. J Food Process Eng 40:e12478. <https://doi.org/10.1111/jfpe.12478>
- <span id="page-31-21"></span>Miltner M, Makaruk A, Harasek M (2017) Review on available biogas upgrading technologies and innovations towards advanced solutions. J Clean Prod 161:1329– 1337.<https://doi.org/10.1016/j.jclepro.2017.06.045>
- <span id="page-31-11"></span>Molfetta M et al (2022) Protein sources alternative to meat: state of the art and involvement of fermentation. Foods 11:2065
- <span id="page-31-8"></span>Molitor HR, Moore EJ, Schnoor JL  $(2019)$  Maximum CO<sub>2</sub> utilization by nutritious microalgae. ACS Sustain Chem Eng 7:9474–9479. [https://doi.org/10.1021/acssu](https://doi.org/10.1021/acssuschemeng.9b00656) [schemeng.9b00656](https://doi.org/10.1021/acssuschemeng.9b00656)
- <span id="page-31-16"></span>Montenegro-Herrera CA, Vera-López Portillo F, Hernández-Chávez GT, Martinez A (2022) Single-cell protein production potential with the extremophilic red microalgae Galdieria sulphuraria: growth and biochemical characterization. J Appl Phycol 34:1341–1352. [https://doi.](https://doi.org/10.1007/s10811-022-02733-y) [org/10.1007/s10811-022-02733-y](https://doi.org/10.1007/s10811-022-02733-y)
- <span id="page-31-4"></span>Moon NJ, Hammond EG, Glatz BA (1978) Conversion of cheese whey and whey permeate to oil and single-cell protein1. J Dairy Sci 61:1537–1547. [https://doi.org/10.](https://doi.org/10.3168/jds.S0022-0302(78)83762-X) [3168/jds.S0022-0302\(78\)83762-X](https://doi.org/10.3168/jds.S0022-0302(78)83762-X)
- <span id="page-31-0"></span>Mugagga F, Nabaasa BB (2016) The centrality of water resources to the realization of sustainable development goals (SDG) a review of potentials and constraints on the african continent. Int Soil Water Conserv Res 4:215–223.<https://doi.org/10.1016/j.iswcr.2016.05.004>
- <span id="page-31-10"></span>Muñoz R, Meier L, Diaz I, Jeison D (2015) A review on the state-of-the-art of physical/chemical and biological technologies for biogas upgrading. Rev Environ Sci Bio/Technology 14:727–759. [https://doi.org/10.1007/](https://doi.org/10.1007/s11157-015-9379-1) [s11157-015-9379-1](https://doi.org/10.1007/s11157-015-9379-1)
- <span id="page-31-7"></span>Muys M, Sui Y, Schwaiger B, Lesueur C, Vandenheuvel D, Vermeir P, Vlaeminck SE (2019) High variability in nutritional value and safety of commercially available Chlorella and Spirulina biomass indicates the need for smart production strategies. Biores Technol 275:247– 257. <https://doi.org/10.1016/j.biortech.2018.12.059>
- <span id="page-31-23"></span>Naquash A, Qyyum MA, Haider J, Bokhari A, Lim H, Lee M (2022) State-of-the-art assessment of cryogenic technologies for biogas upgrading: Energy, economic, and environmental perspectives. Renew Sustain Energy Rev 154:111826. [https://doi.org/10.1016/j.rser.2021.](https://doi.org/10.1016/j.rser.2021.111826) [111826](https://doi.org/10.1016/j.rser.2021.111826)
- <span id="page-31-1"></span>Newman L, Fraser E, Newell R, Bowness E, Newman K, Glaros A (2023) Chapter 1-cellular agriculture and the sustainable development goals. In: Lopez-Correa C, Suarez-Gonzalez A (eds) Genomics and the global bioeconomy. Academic Press, Cambridge, Massachusetts, pp 3–23. <https://doi.org/10.1016/B978-0-323-91601-1.00010-9>
- <span id="page-31-25"></span>Ngoc-Dan Cao T, Mukhtar H, Yu C-P, Bui X-T, Pan S-Y (2022) Agricultural waste-derived biochar in microbial fuel cells towards a carbon-negative circular economy. Renew Sustain Energy Rev 170:112965. [https://doi.org/](https://doi.org/10.1016/j.rser.2022.112965) [10.1016/j.rser.2022.112965](https://doi.org/10.1016/j.rser.2022.112965)
- <span id="page-31-6"></span>Nigam JN (1998) Single cell protein from pineapple cannery effluent. World J Microbiol Biotechnol 14:693-696. <https://doi.org/10.1023/A:1008853303596>
- <span id="page-31-19"></span>Nizovtseva IG et al (2022) Simulation of two-phase air–liquid flows in a closed bioreactor loop: Numerical modeling, experiments, and verifcation. Math Methods Appl Sci 45:8216–8229. <https://doi.org/10.1002/mma.8132>
- <span id="page-31-2"></span>Nyyssölä A, Suhonen A, Ritala A, Oksman-Caldentey K-M (2022) The role of single cell protein in cellular agriculture. Curr Opin Biotechnol 75:102686. [https://doi.org/10.](https://doi.org/10.1016/j.copbio.2022.102686) [1016/j.copbio.2022.102686](https://doi.org/10.1016/j.copbio.2022.102686)
- <span id="page-31-13"></span>Olukomaiya O, Fernando C, Mereddy R, Li X, Sultanbawa Y (2019) Solid-state fermented plant protein sources in the diets of broiler chickens: a review Animal. Nutrition 5:319–330. <https://doi.org/10.1016/j.aninu.2019.05.005>
- <span id="page-31-24"></span>Øvrum Hansen J et al (2019) Efect of Candida utilis on growth and intestinal health of Atlantic salmon (Salmo salar) parr. Aquaculture 511:734239. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.aquaculture.2019.734239) [aquaculture.2019.734239](https://doi.org/10.1016/j.aquaculture.2019.734239)
- <span id="page-31-15"></span>Padoan E et al (2022) Waste biopolymers for eco-friendly agriculture and safe food production. Coatings 12:239
- <span id="page-31-18"></span>Pan M, Su Y, Zhu X, Pan G, Zhang Y, Angelidaki I (2021) Bioelectrochemically assisted sustainable conversion of industrial organic wastewater and clean production of microalgal protein. Resour Conserv Recycling 168:105441. [https://doi.org/10.1016/j.resconrec.2021.](https://doi.org/10.1016/j.resconrec.2021.105441) [105441](https://doi.org/10.1016/j.resconrec.2021.105441)
- <span id="page-32-17"></span>Pander B et al (2020) Hydrogen oxidising bacteria for production of single-cell protein and other food and feed ingredients. Eng Biol 4:21–24. [https://doi.org/10.1049/enb.](https://doi.org/10.1049/enb.2020.0005) [2020.0005](https://doi.org/10.1049/enb.2020.0005)
- <span id="page-32-10"></span>Pereira AG et al (2022) Single-cell proteins obtained by circular economy intended as a feed ingredient in aquaculture. Foods 11:2831
- <span id="page-32-24"></span>Petersen LAH, Villadsen J, Jørgensen SB, Gernaey KV (2017) Mixing and mass transfer in a pilot scale U-loop bioreactor. Biotechnol Bioeng 114:344–354. [https://doi.org/10.](https://doi.org/10.1002/bit.26084) [1002/bit.26084](https://doi.org/10.1002/bit.26084)
- <span id="page-32-5"></span>Pillaca-Pullo OS, Lopes AM, Rodriguez-Portilla LMI, Estela-Escalante W (2023) Optimizing medium composition with wastewater from Coffea arabica processing to produce single-cell protein using Candida sorboxylosa. J Chem Technol Biotechnol 98:106–116. [https://doi.org/](https://doi.org/10.1002/jctb.7219) [10.1002/jctb.7219](https://doi.org/10.1002/jctb.7219)
- <span id="page-32-25"></span>Pilmer LW, Woolley LD, Lymbery AJ, Salini M, Partridge GJ (2022) Using dietary additives to improve palatability of diets containing single-cell protein from methanotrophic bacteria in yellowtail kingfsh (Seriola lalandi) diets. Aquaculture Res 53:5006–5017. [https://doi.org/10.1111/](https://doi.org/10.1111/are.15986) [are.15986](https://doi.org/10.1111/are.15986)
- <span id="page-32-7"></span>Puligundla P, Mok C (2021) Valorization of sugar beet pulp through biotechnological approaches: recent developments. Biotech Lett 43:1253–1263. [https://doi.org/10.](https://doi.org/10.1007/s10529-021-03146-6) [1007/s10529-021-03146-6](https://doi.org/10.1007/s10529-021-03146-6)
- <span id="page-32-18"></span>Quan C, Zhang L, Wang Y, Ohta Y (2001) Production of phytase in a low phosphate medium by a novel yeast Candida krusei. J Biosci Bioeng 92:154–160. [https://doi.](https://doi.org/10.1016/S1389-1723(01)80217-6) [org/10.1016/S1389-1723\(01\)80217-6](https://doi.org/10.1016/S1389-1723(01)80217-6)
- <span id="page-32-12"></span>Rages AA, Haider MM, Aydin M (2021) Alkaline hydrolysis of olive fruits wastes for the production of single cell protein by Candida lipolytica. Biocatal Agricul Biotechnol 33:101999. <https://doi.org/10.1016/j.bcab.2021.101999>
- <span id="page-32-20"></span>Ravi Kiran B, Venkata Mohan S (2022) Phycoremediation potential ofTetradesmus spSVMIICT4in treating dairy wastewaterusingFlat-Panel photobioreactor. Bioresource Technol 345:126446. [https://doi.org/10.1016/j.biortech.](https://doi.org/10.1016/j.biortech.2021.126446) [2021.126446](https://doi.org/10.1016/j.biortech.2021.126446)
- <span id="page-32-1"></span>Raziq A, Lateef M, Ullah A, Ullah H, Khan MW (2020) 02. Single cell protein (SCP) production and potential substrates: a comprehensive review. Pure Appl Biol (PAB) 9(3):1743–1754
- <span id="page-32-9"></span>Razzaq ZU, Maan AA, Nazir A, Hafeez MA, Khan MKI (2022) Characterizing the single cell protein enriched noodles for nutritional and organoleptic attributes. J Food Meas Charact 16:1725–1732. [https://doi.org/10.1007/](https://doi.org/10.1007/s11694-022-01300-w) [s11694-022-01300-w](https://doi.org/10.1007/s11694-022-01300-w)
- <span id="page-32-6"></span>Reihani SFS, Khosravi-Darani K (2019) Infuencing factors on single-cell protein production by submerged fermentation: a review. Electron J Biotechnol 37:34–40. [https://](https://doi.org/10.1016/j.ejbt.2018.11.005) [doi.org/10.1016/j.ejbt.2018.11.005](https://doi.org/10.1016/j.ejbt.2018.11.005)
- <span id="page-32-0"></span>Ritala A, Häkkinen ST, Toivari M, Wiebe MG (2017) Single cell protein—state-of-the-art industrial landscape and patents 2001–2016. Front Microbiol. [https://doi.org/10.](https://doi.org/10.3389/fmicb.2017.02009) [3389/fmicb.2017.02009](https://doi.org/10.3389/fmicb.2017.02009)
- <span id="page-32-23"></span>Rodríguez Y, Firmino PIM, Pérez V, Lebrero R, Muñoz R (2020) Biogas valorization via continuous polyhydroxybutyrate production by Methylocystis hirsuta in a bubble

column bioreactor. Waste Manag 113:395–403. [https://](https://doi.org/10.1016/j.wasman.2020.06.009) [doi.org/10.1016/j.wasman.2020.06.009](https://doi.org/10.1016/j.wasman.2020.06.009)

- <span id="page-32-3"></span>Rosenboom J-G, Langer R, Traverso G (2022) Bioplastics for a circular economy. Nat Rev Mater 7:117–137. [https://doi.](https://doi.org/10.1038/s41578-021-00407-8) [org/10.1038/s41578-021-00407-8](https://doi.org/10.1038/s41578-021-00407-8)
- <span id="page-32-14"></span>Saejung C, Chanthakhot T (2021) Single-phase and two-phase cultivations using diferent light regimes to improve production of valuable substances in the anoxygenic photosynthetic bacterium Rhodopseudomonas faecalis PA2. Bioresource Technol 328:124855. [https://doi.org/10.](https://doi.org/10.1016/j.biortech.2021.124855) [1016/j.biortech.2021.124855](https://doi.org/10.1016/j.biortech.2021.124855)
- <span id="page-32-8"></span>Saejung C, Salasook P (2020) Recycling of sugar industry wastewater for single-cell protein production with supplemental carotenoids. Environ Technol 41:59–70. <https://doi.org/10.1080/09593330.2018.1491633>
- <span id="page-32-15"></span>Saejung C, Sanusan W (2021) Valorization of lignocellulosic wastes and nutrient recovery by anoxygenic photosynthetic bacteria. Waste Biomass Valorization 12:4835– 4844. <https://doi.org/10.1007/s12649-021-01351-y>
- <span id="page-32-4"></span>Sakarika M, Delmoitié B, Ntagia E, Chatzigiannidou I, Gabet X, Ganigué R, Rabaey K (2022) Production of microbial protein from fermented grass. Chem Eng J 433:133631. <https://doi.org/10.1016/j.cej.2021.133631>
- <span id="page-32-13"></span>Salazar-López NJ, Barco-Mendoza GA, Zuñiga-Martínez BS, Domínguez-Avila JA, Robles-Sánchez RM, Ochoa MAV, González-Aguilar GA (2022) Single-cell protein production as a strategy to reincorporate food waste and agro by-products back into the processing chain. Bioengineering 9:623
- <span id="page-32-22"></span>Salehi R, Chaiprapat S (2019) Single/triple-stage biotrickling flter treating a H2S-rich biogas stream: statistical analysis of the efect of empty bed retention time and liquid recirculation velocity. J Air Waste Manag Assoc 69:1429–1437. [https://doi.org/10.1080/10962247.2019.](https://doi.org/10.1080/10962247.2019.1645761) [1645761](https://doi.org/10.1080/10962247.2019.1645761)
- <span id="page-32-16"></span>Salehi R, Chaiprapat S (2022) Conversion of biogas from anaerobic digestion to single cell protein and bio-methanol: mechanism, microorganisms and key factors. A Rev Environ Eng Res 27:210109–210100. [https://doi.org/10.](https://doi.org/10.4491/eer.2021.109) [4491/eer.2021.109](https://doi.org/10.4491/eer.2021.109)
- <span id="page-32-19"></span>Selvaraj S, Natarajan K, Nowak A, Murty VR (2021) Mathematical modeling and simulation of newly isolated bacillus cereus M1GT for tannase production through semisolid state fermentation with agriculture residue triphala. South Afr J Chem Eng 35:89–97. [https://doi.org/10.](https://doi.org/10.1016/j.sajce.2020.10.001) [1016/j.sajce.2020.10.001](https://doi.org/10.1016/j.sajce.2020.10.001)
- <span id="page-32-11"></span>Sen S, Mansell TJ (2020) Yeasts as probiotics: mechanisms outcomes and future potential. Fungal Gen Biol 137:103333.<https://doi.org/10.1016/j.fgb.2020.103333>
- <span id="page-32-2"></span>Sharif M, Zafar MH, Aqib AI, Saeed M, Farag MR, Alagawany M (2021) Single cell protein: sources, mechanism of production, nutritional value and its uses in aquaculture nutrition. Aquaculture 531:735885. [https://doi.org/10.](https://doi.org/10.1016/j.aquaculture.2020.735885) [1016/j.aquaculture.2020.735885](https://doi.org/10.1016/j.aquaculture.2020.735885)
- <span id="page-32-21"></span>Sharma S, Sevda S, Garlapati VK (2020) Chapter 19-microalgae in bioelectrochemical systems: technologic interventions. In: Krishnaraj Rathinam N, Sani RK (eds) Biovalorisation of wastes to renewable chemicals and biofuels. Elsevier, Amsterdam, Netherlands, pp 361–371. [https://](https://doi.org/10.1016/B978-0-12-817951-2.00019-5) [doi.org/10.1016/B978-0-12-817951-2.00019-5](https://doi.org/10.1016/B978-0-12-817951-2.00019-5)
- <span id="page-33-7"></span>Sharma V, Tsai M-L, Nargotra P, Chen C-W, Kuo C-H, Sun P-P, Dong C-D (2022) Agro-industrial food waste as a low-cost substrate for sustainable production of industrial enzymes: a critical review. Catalysts 12:1373
- <span id="page-33-20"></span>Sillman J, Uusitalo V, Ruuskanen V, Ojala L, Kahiluoto H, Soukka R, Ahola J (2020) A life cycle environmental sustainability analysis of microbial protein production via power-to-food approaches The. Int J Life Cycle Assess 25:2190–2203. [https://doi.org/10.1007/](https://doi.org/10.1007/s11367-020-01771-3) [s11367-020-01771-3](https://doi.org/10.1007/s11367-020-01771-3)
- <span id="page-33-13"></span>Śliżewska K, Chlebicz-Wójcik A (2020) Growth kinetics of probiotic lactobacillus strains in the alternative cost-efficient semi-solid fermentation medium. Biology 9:423
- <span id="page-33-21"></span>Soto V, Ulloa C, Garcia X (2021) A CFD design approach for industrial size tubular reactors for SNG production from biogas (CO<sub>2</sub> Methanation). Energies  $14:6175$
- <span id="page-33-26"></span>Stephens N, Di Silvio L, Dunsford I, Ellis M, Glencross A, Sexton A (2018) Bringing cultured meat to market: technical, socio-political, and regulatory challenges in cellular agriculture. Trends Food Sci Technol 78:155–166. <https://doi.org/10.1016/j.tifs.2018.04.010>
- <span id="page-33-11"></span>Stern MD, Hoover WH (1979) Methods for determining and factors afecting rumen microbial protein synthesis: a review. J Anim Sci 49:1590–1603. [https://doi.org/10.](https://doi.org/10.2527/jas1979.4961590x) [2527/jas1979.4961590x](https://doi.org/10.2527/jas1979.4961590x)
- <span id="page-33-23"></span>Stolecka K, Rusin A (2021) Potential hazards posed by biogas plants. Renew Sustain Energy Rev 135:110225. [https://](https://doi.org/10.1016/j.rser.2020.110225) [doi.org/10.1016/j.rser.2020.110225](https://doi.org/10.1016/j.rser.2020.110225)
- <span id="page-33-19"></span>Styles D, Adams P, Thelin G, Vaneeckhaute C, Chadwick D, Withers PJA (2018) Life cycle assessment of biofertilizer production and use compared with conventional liquid digestate management. Environ Sci Technol 52:7468– 7476. <https://doi.org/10.1021/acs.est.8b01619>
- <span id="page-33-9"></span>Su W et al (2021) Variations of soybean meal and corn mixed substrates in physicochemical characteristics and microbiota during two-stage solid-state fermentation. Front Microbiol 12:688839. [https://doi.org/10.3389/fmicb.](https://doi.org/10.3389/fmicb.2021.688839) [2021.688839](https://doi.org/10.3389/fmicb.2021.688839)
- <span id="page-33-10"></span>Sui Y, Vlaeminck SE (2020) Dunaliella microalgae for nutritional protein: an undervalued asset. Trends Biotechnol 38:10–12. <https://doi.org/10.1016/j.tibtech.2019.07.011>
- <span id="page-33-14"></span>Suriyapha C, Ampapon T, Viennasay B, Matra M, Wann C, Wanapat M (2020) Manipulating rumen fermentation, microbial protein synthesis, and mitigating methane production using bamboo grass pellet in swamp buffaloes. Trop Animal Health Prod 52:1609–1615. [https://doi.org/](https://doi.org/10.1007/s11250-019-02163-y) [10.1007/s11250-019-02163-y](https://doi.org/10.1007/s11250-019-02163-y)
- <span id="page-33-2"></span>Talan A, Pokhrel S, Tyagi RD, Drogui P (2022) Biorefnery strategies for microbial bioplastics production: Sustainable pathway towards circular bioeconomy. Bioresource Technol Rep 17:100875. [https://doi.org/10.1016/j.biteb.](https://doi.org/10.1016/j.biteb.2021.100875) [2021.100875](https://doi.org/10.1016/j.biteb.2021.100875)
- <span id="page-33-24"></span>Tayou LN et al (2022) Acidogenic fermentation of food waste and sewage sludge mixture: effect of operating parameters on process performance and safety aspects. Process Saf Environ Prot 163:158–166. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.psep.2022.05.011) [psep.2022.05.011](https://doi.org/10.1016/j.psep.2022.05.011)
- <span id="page-33-22"></span>Thamdrup B et al (2019) Anaerobic methane oxidation is an important sink for methane in the ocean's largest oxygen minimum zone. Limnol Oceanogr 64:2569-2585. [https://](https://doi.org/10.1002/lno.11235) [doi.org/10.1002/lno.11235](https://doi.org/10.1002/lno.11235)
- <span id="page-33-6"></span>Thiviya P, Gamage A, Kapilan R, Merah O, Madhujith T (2022a) Production of single-cell protein from fruit peel wastes using palmyrah toddy yeast. Fermentation 8:355
- <span id="page-33-5"></span>Thiviya P, Gamage A, Kapilan R, Merah O, Madhujith T (2022b) Single cell protein production using diferent fruit waste: a review. Separations 9:178
- <span id="page-33-25"></span>Tola S et al (2022) Effects of dietary tuna hydrolysate supplementation on feed intake, growth performance, feed utilization and health status of Asian sea bass (Lates calcarifer) fed a low fsh meal soybean meal-based diet. Aquacult Res 53:3898–3912. [https://doi.org/10.1111/are.](https://doi.org/10.1111/are.15894) [15894](https://doi.org/10.1111/are.15894)
- <span id="page-33-8"></span>Tomlinson EJ (1976) The production of single-cell protein from strong organic waste waters from the food and drink processing industries—I. Lab Cult Water Res 10:367– 371. [https://doi.org/10.1016/0043-1354\(76\)90053-1](https://doi.org/10.1016/0043-1354(76)90053-1)
- <span id="page-33-4"></span>Tropea A, Ferracane A, Albergamo A, Potortì AG, Lo Turco V, Di Bella G (2022) Single cell protein production through multi food-waste substrate fermentation. Fermentation 8:91
- <span id="page-33-16"></span>Tsapekos P, Khoshnevisan B, Alvarado-Morales M, Zhu X, Pan J, Tian H, Angelidaki I (2021) Upcycling the anaerobic digestion streams in a bioeconomy approach: a review. Renew Sustain Energy Rev 151:111635. [https://](https://doi.org/10.1016/j.rser.2021.111635) [doi.org/10.1016/j.rser.2021.111635](https://doi.org/10.1016/j.rser.2021.111635)
- <span id="page-33-3"></span>Türker M, Selimoğlu SM, Taşpınar-Demir H (2022) Chapter 12-waste(water) to feed protein: effluent characteristics, protein recovery, and single-cell protein production from food industry waste streams. In: An A, Tyagi V, Kumar M, Cetecioglu Z (eds) Clean energy and resource recovery. Elsevier, Amsterdam, Netherlands, pp 201–244. [https://doi.org/10.1016/B978-0-323-90178-9.](https://doi.org/10.1016/B978-0-323-90178-9.00017-2) [00017-2](https://doi.org/10.1016/B978-0-323-90178-9.00017-2)
- <span id="page-33-18"></span>Tyagi B, Sahota S, Thakur IS, Ghosh P (2022) Chapter 9-Microbial transformation of methane to biofuels and biomaterials. In: Thakur IS, Pandey A, Ngo HH, Soccol CR, Larroche C (eds) Biomass, biofuels, biochemicals. Elsevier, Amsterdam, Netherlands, pp 203–230. [https://](https://doi.org/10.1016/B978-0-12-823500-3.00020-0) [doi.org/10.1016/B978-0-12-823500-3.00020-0](https://doi.org/10.1016/B978-0-12-823500-3.00020-0)
- <span id="page-33-1"></span>Ugalde UO, Castrillo JI (2002) Single cell proteins from fungi and yeasts. In: Khachatourians GG, Arora DK (eds) Applied mycology and biotechnology, vol 2. Elsevier, Amsterdam Netherlands, pp 123–149. [https://doi.org/10.](https://doi.org/10.1016/S1874-5334(02)80008-9) [1016/S1874-5334\(02\)80008-9](https://doi.org/10.1016/S1874-5334(02)80008-9)
- <span id="page-33-0"></span>Unibio (2022) Atar-based gulf biotech buys license from unibio to turn natural gas into sustainable protein. [https://](https://www.unibio.dk/press-release-qatar-based-gulf-biotech-buys-license-from-unibio-to-turn-natural-gas-into-sustainable-protein/) [www.unibio.dk/press-release-qatar-based-gulf-biotech](https://www.unibio.dk/press-release-qatar-based-gulf-biotech-buys-license-from-unibio-to-turn-natural-gas-into-sustainable-protein/)[buys-license-from-unibio-to-turn-natural-gas-into-susta](https://www.unibio.dk/press-release-qatar-based-gulf-biotech-buys-license-from-unibio-to-turn-natural-gas-into-sustainable-protein/) [inable-protein/](https://www.unibio.dk/press-release-qatar-based-gulf-biotech-buys-license-from-unibio-to-turn-natural-gas-into-sustainable-protein/)
- <span id="page-33-15"></span>van der Ha D, Nachtergaele L, Kerckhof F-M, Rameiyanti D, Bossier P, Verstraete W, Boon N (2012) Conversion of biogas to bioproducts by algae and methane oxidizing bacteria. Environ Sci Technol 46:13425–13431. [https://](https://doi.org/10.1021/es303929s) [doi.org/10.1021/es303929s](https://doi.org/10.1021/es303929s)
- <span id="page-33-12"></span>Van Heyst A, Sinclair A, Jamieson R (2022) Application of phosphorus loading models to understand drivers of eutrophication in a complex rural lake-watershed system. J Environ Manag 302:114010. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jenvman.2021.114010) [jenvman.2021.114010](https://doi.org/10.1016/j.jenvman.2021.114010)
- <span id="page-33-17"></span>Van Peteghem L, Sakarika M, Matassa S, Pikaar I, Ganigué R, Rabaey K (2022) Towards new carbon–neutral food

systems: combining carbon capture and utilization with microbial protein production. Bioresource Technol 349:126853. https://doi.org/10.1016/j.biortech.2022. [https://doi.org/10.1016/j.biortech.2022.](https://doi.org/10.1016/j.biortech.2022.126853) [126853](https://doi.org/10.1016/j.biortech.2022.126853)

- <span id="page-34-19"></span>Vauris A, Valcauda S, Husson F, Coninck JD (2022) A novel method to assess heat transfer and impact of relevant physicochemical parameters for the scaling up of solid state fermentation systems. Biotechnol Rep 36:e00764. <https://doi.org/10.1016/j.btre.2022.e00764>
- <span id="page-34-9"></span>Vendruscolo F, Albuquerque PM, Streit F, Esposito E, Ninow JL (2008) Apple pomace: a versatile substrate for biotechnological applications. Crit Rev Biotechnol 28:1–12. <https://doi.org/10.1080/07388550801913840>
- <span id="page-34-23"></span>Verbeeck K, De Vrieze J, Biesemans M, Rabaey K (2019) Membrane electrolysis-assisted  $CO<sub>2</sub>$  and  $H<sub>2</sub>S$  extraction as innovative pretreatment method for biological biogas upgrading. Chem Eng J 361:1479–1486. [https://doi.org/](https://doi.org/10.1016/j.cej.2018.09.120) [10.1016/j.cej.2018.09.120](https://doi.org/10.1016/j.cej.2018.09.120)
- <span id="page-34-11"></span>Verbeeck K, De Vrieze J, Pikaar I, Verstraete W, Rabaey K (2021) Assessing the potential for up-cycling recovered resources from anaerobic digestion through microbial protein production. Microb Biotechnol 14:897–910. <https://doi.org/10.1111/1751-7915.13600>
- <span id="page-34-12"></span>Vethathirri RS, Santillan E, Wuertz S (2021) Microbial community-based protein production from wastewater for animal feed applications. Bioresource Technol 341:125723. [https://doi.org/10.1016/j.biortech.2021.](https://doi.org/10.1016/j.biortech.2021.125723) [125723](https://doi.org/10.1016/j.biortech.2021.125723)
- <span id="page-34-20"></span>Villegas-Méndez MÁ, Montañez J, Contreras-Esquivel JC, Salmerón I, Koutinas A, Morales-Oyervides L (2022) Coproduction of microbial oil and carotenoids within the circular bioeconomy concept: a sequential solid-state and submerged fermentation approach. Fermentation 8:258
- <span id="page-34-5"></span>Voutilainen E, Pihlajaniemi V, Parviainen T (2021) Economic comparison of food protein production with single-cell organisms from lignocellulose side-streams. Bioresour Technol Rep 14:100683. [https://doi.org/10.1016/j.biteb.](https://doi.org/10.1016/j.biteb.2021.100683) [2021.100683](https://doi.org/10.1016/j.biteb.2021.100683)
- <span id="page-34-25"></span>Wada OZ, Vincent AS, Mackey HR (2022) Single-cell protein production from purple non-sulphur bacteria-based wastewater treatment. Rev Environ Sci<br>Bio/technol 21:931-956. https://doi.org/10.1007/ [https://doi.org/10.1007/](https://doi.org/10.1007/s11157-022-09635-y) [s11157-022-09635-y](https://doi.org/10.1007/s11157-022-09635-y)
- <span id="page-34-10"></span>Wadhwa M, Bakshi MPS (2016) Chapter 10-application of waste-derived proteins in the animal feed industry. In: Singh Dhillon G (ed) Protein byproducts. Academic Press, Cambridge, Massachusetts, pp 161–192. [https://](https://doi.org/10.1016/B978-0-12-802391-4.00010-0) [doi.org/10.1016/B978-0-12-802391-4.00010-0](https://doi.org/10.1016/B978-0-12-802391-4.00010-0)
- <span id="page-34-3"></span>Wang D-H, Zhu M-Y, Lian S-J, Zou H, Fu S-F, Guo R-B (2022) Conversion of renewable biogas into single-cell protein using a combined microalga- and methane-oxidizing bacterial system ACS ES&T. Engineering. [https://](https://doi.org/10.1021/acsestengg.2c00237) [doi.org/10.1021/acsestengg.2c00237](https://doi.org/10.1021/acsestengg.2c00237)
- <span id="page-34-13"></span>Wiebe MG (2004) Quorn™ myco-protein—overview of a successful fungal product. Mycologist 18:17–20
- <span id="page-34-1"></span>Woolley L, Chaklader MR, Pilmer L, Stephens F, Wingate C, Salini M, Partridge G (2023) Gas to protein: microbial single cell protein is an alternative to fshmeal in aquaculture. Sci Total Environ 859:160141. [https://doi.org/](https://doi.org/10.1016/j.scitotenv.2022.160141) [10.1016/j.scitotenv.2022.160141](https://doi.org/10.1016/j.scitotenv.2022.160141)
- <span id="page-34-2"></span>Xu M, Zhou H, Yang X, Angelidaki I, Zhang Y (2020) Sulfde restrains the growth of methylocapsa acidiphila converting renewable biogas to single cell protein. Water Res 184:116138. [https://doi.org/10.1016/j.watres.2020.](https://doi.org/10.1016/j.watres.2020.116138) [116138](https://doi.org/10.1016/j.watres.2020.116138)
- <span id="page-34-18"></span>Xu M, Zhao D, Zhu X, Su Y, Angelidaki I, Zhang Y (2021) Biogas upgrading and valorization to single-cell protein in a bioinorganic electrosynthesis system. Chem Eng J 426:131837.<https://doi.org/10.1016/j.cej.2021.131837>
- <span id="page-34-4"></span>Xu M, Zhou H, Zou R, Yang X, Su Y, Angelidaki I, Zhang Y (2021) Beyond the farm: making edible protein from CO<sub>2</sub> via hybrid bioinorganic electrosynthesis. One Earth 4:868–878. <https://doi.org/10.1016/j.oneear.2021.05.007>
- <span id="page-34-8"></span>Yan J et al (2018) Engineering yarrowia lipolytica to simultaneously produce lipase and single cell protein from agroindustrial wastes for feed. Sci Rep 8:758. [https://doi.org/](https://doi.org/10.1038/s41598-018-19238-9) [10.1038/s41598-018-19238-9](https://doi.org/10.1038/s41598-018-19238-9)
- <span id="page-34-6"></span>Yang A, Zhang G, Meng F, Lu P, Wang X, Peng M (2017) Enhancing protein to extremely high content in photosynthetic bacteria during biogas slurry treatment. Biores Technol 245:1277–1281. [https://doi.org/10.1016/j.biort](https://doi.org/10.1016/j.biortech.2017.08.109) [ech.2017.08.109](https://doi.org/10.1016/j.biortech.2017.08.109)
- <span id="page-34-21"></span>Yang A, Zhang G, Meng F, Zhang P, Chen Y (2018) Membrane concentrate treatment by photosynthetic bacteria: feasibility and tolerance mechanism analysis. Biores Technol 253:378–381. [https://doi.org/10.1016/j.biortech.](https://doi.org/10.1016/j.biortech.2018.01.034) [2018.01.034](https://doi.org/10.1016/j.biortech.2018.01.034)
- <span id="page-34-15"></span>Yang Z et al (2021) Effects of dietary yucca schidigera extract and oral candida utilis on growth performance and intestinal health of weaned piglets. Front Nutr. [https://doi.org/](https://doi.org/10.3389/fnut.2021.685540) [10.3389/fnut.2021.685540](https://doi.org/10.3389/fnut.2021.685540)
- <span id="page-34-24"></span>Yang X, Xu M, Zou R, Angelidaki I, Zhang Y (2021) Microbial protein production from  $CO_2$ ,  $H_2$ , and recycled nitrogen: Focusing on ammonia toxicity and nitrogen sources. J Clean Prod 291:125921. [https://doi.org/10.1016/j.jclep](https://doi.org/10.1016/j.jclepro.2021.125921) [ro.2021.125921](https://doi.org/10.1016/j.jclepro.2021.125921)
- <span id="page-34-22"></span>Yang Z, Tsapekos P, Zhang Y, Zhang Y, Angelidaki I, Wang W (2021) Bio-electrochemically extracted nitrogen from residual resources for microbial protein production. Bioresource Technol 337:125353. [https://doi.org/10.](https://doi.org/10.1016/j.biortech.2021.125353) [1016/j.biortech.2021.125353](https://doi.org/10.1016/j.biortech.2021.125353)
- <span id="page-34-0"></span>Yang R, Chen Z, Hu P, Zhang S, Luo G (2022) Two-stage fermentation enhanced single-cell protein production by Yarrowia lipolytica from food waste. Bioresource Technol 361:127677. [https://doi.org/10.1016/j.biortech.2022.](https://doi.org/10.1016/j.biortech.2022.127677) [127677](https://doi.org/10.1016/j.biortech.2022.127677)
- <span id="page-34-14"></span>Yao Y-Y et al (2020) Surface display system for probiotics and its application in aquaculture. Rev Aquac 12:2333–2350. <https://doi.org/10.1111/raq.12437>
- <span id="page-34-16"></span>Yu S, Peng L, Xu Y, Song S, Xie G-J, Liu Y, Ni B-J (2021) Optimizing light sources for selective growth of purple bacteria and efficient formation of value-added products. J Clean Prod 280:124493. [https://doi.org/10.1016/j.jclep](https://doi.org/10.1016/j.jclepro.2020.124493) [ro.2020.124493](https://doi.org/10.1016/j.jclepro.2020.124493)
- <span id="page-34-17"></span>Yu S, Xu Y, Liang C, Lou W, Peng L (2022) Spectral bands of incandescent lamp leading to variable productivity of purple bacteria biomass and microbial protein: Full is better than segmented. Sci Total Environ 823:153736. <https://doi.org/10.1016/j.scitotenv.2022.153736>
- <span id="page-34-7"></span>Yunus FN, Nadeem M, Rashid F (2015) Single-cell protein production through microbial conversion of lignocellulosic

residue (wheat bran) for animal feed. J Inst Brewing 121:553–557.<https://doi.org/10.1002/jib.251>

- <span id="page-35-10"></span>Zamani A, Khajavi M, Nazarpak MH, Gisbert E (2020) Evaluation of a bacterial single-cell protein in compound diets for rainbow trout (Oncorhynchus mykiss) fry as an alternative protein source. Animals 10:1676
- <span id="page-35-0"></span>Zeng D, Jiang Y, Su Y, Zhang Y (2022) Upcycling waste organic acids and nitrogen into single cell protein via brewer's yeast. J Clean Prod 369:133279. [https://doi.org/](https://doi.org/10.1016/j.jclepro.2022.133279) [10.1016/j.jclepro.2022.133279](https://doi.org/10.1016/j.jclepro.2022.133279)
- <span id="page-35-4"></span>Zeng D, Jiang Y, Schneider C, Su Y, Hélix-Nielsen C, Zhang Y (2023) Recycling of acetate and ammonium from digestate for single cell protein production by a hybrid electrochemical-membrane fermentation process. Resour Conserv Recycling 188:106705. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.resconrec.2022.106705) [resconrec.2022.106705](https://doi.org/10.1016/j.resconrec.2022.106705)
- <span id="page-35-5"></span>Zha X, Tsapekos P, Zhu X, Khoshnevisan B, Lu X, Angelidaki I (2021) Bioconversion of wastewater to single cell protein by methanotrophic bacteria. Bioresource Technol 320:124351. https://doi.org/10.1016/j.biortech.2020. [https://doi.org/10.1016/j.biortech.2020.](https://doi.org/10.1016/j.biortech.2020.124351) [124351](https://doi.org/10.1016/j.biortech.2020.124351)
- <span id="page-35-9"></span>Zhang H, Zhang X, Cao XR, Iftikhar M, Wang J (2018) Semi-solid state fermentation and enzymatic hydrolysis impeded the destroy of wheat bran on gluten polymerization. LWT 98:306–313. [https://doi.org/10.1016/j.lwt.](https://doi.org/10.1016/j.lwt.2018.08.047) [2018.08.047](https://doi.org/10.1016/j.lwt.2018.08.047)
- <span id="page-35-6"></span>Zhang L, Zhou P, Chen YC, Cao Q, Liu XF, Li D (2021) The production of single cell protein from biogas slurry with high ammonia-nitrogen content by screened nectaromyces rattus. Poultry Sci 100:101334. [https://doi.org/10.](https://doi.org/10.1016/j.psj.2021.101334) [1016/j.psj.2021.101334](https://doi.org/10.1016/j.psj.2021.101334)
- <span id="page-35-11"></span>Zhang Q et al (2022) Effects of replacing fishmeal with methanotroph (Methylococcus capsulatus, Bath) bacteria meal (FeedKind®) on growth and intestinal health status of juvenile largemouth bass (Micropterus salmoides). Fish Shellfsh Immunol 122:298–305. [https://doi.org/10.](https://doi.org/10.1016/j.fsi.2022.02.008) [1016/j.fsi.2022.02.008](https://doi.org/10.1016/j.fsi.2022.02.008)
- <span id="page-35-3"></span>Zheng J et al (2023) Effects of fish meal replaced by methanotroph bacteria meal (Methylococcus capsulatus) on growth, body composition, antioxidant capacity, amino acids transporters and protein metabolism of turbot juveniles (Scophthalmus maximus L.). Aquaculture 562:738782. [https://doi.org/10.1016/j.aquaculture.2022.](https://doi.org/10.1016/j.aquaculture.2022.738782) [738782](https://doi.org/10.1016/j.aquaculture.2022.738782)
- <span id="page-35-1"></span>Zhou P, Zhang L, Ding H, Gao X, Chen Y, Li D (2022a) Optimization of culture conditions of screened Galactomyces candidum for the production of single cell protein from biogas slurry. Electron J Biotechnol 55:47–54. [https://](https://doi.org/10.1016/j.ejbt.2021.11.006) [doi.org/10.1016/j.ejbt.2021.11.006](https://doi.org/10.1016/j.ejbt.2021.11.006)
- <span id="page-35-8"></span>Zhou Z-X, Yu R-C, Zhou M-J (2022) Evolution of harmful algal blooms in the East China Sea under eutrophication and warming scenarios. Water Res 221:118807. [https://](https://doi.org/10.1016/j.watres.2022.118807) [doi.org/10.1016/j.watres.2022.118807](https://doi.org/10.1016/j.watres.2022.118807)
- <span id="page-35-7"></span>Zhu W, He Q, Gao H, Nitayavardhana S, Khanal SK, Xie L (2020) Bioconversion of yellow wine wastes into microbial protein via mixed yeast-fungus cultures. Bioresource Technol 299:122565. [https://doi.org/10.1016/j.biortech.](https://doi.org/10.1016/j.biortech.2019.122565) [2019.122565](https://doi.org/10.1016/j.biortech.2019.122565)
- <span id="page-35-2"></span>Zhu Z, Wu Y, Hu W, Zheng X, Chen Y (2022) Valorization of food waste fermentation liquid into single cell protein by photosynthetic bacteria via stimulating carbon metabolic pathway and environmental behaviour. Bioresource Technol 361:127704. [https://doi.org/10.1016/j.biortech.](https://doi.org/10.1016/j.biortech.2022.127704) [2022.127704](https://doi.org/10.1016/j.biortech.2022.127704)

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional afliations.