



Role of eating disorders-related polymorphisms in obesity pathophysiology

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Abstract

Human biological system provides innumerable neuroendocrine inputs for food intake control, with effects on appetite's modulation and the satiety signs. Its regulation is very complex, engaging several molecular interactions with many tissues, hormones, and neural circuits. Thus, signaling molecules that control food intake are critical for normal energy homeostasis and a deregulation of these pathways can lead to eating disorders and obesity. In line of this, genetic factors have a significantly influence of the regulation of neural circuits controlling the appetite and satiety pathways, as well as the regulation of brain reward systems. Single Nucleotide Polymorphisms (SNPs) in genes related to hypothalamic appetite and satiety mechanisms, further in multiple neurotransmitter systems may contribute to the development of major Eating Disorders (EDs) related to obesity, among them Binge Eating Disorder (BED) and Bulimia Nervosa (BN), which are discussed in this review.

Keywords Obesity · Food intake · Eating disorders · Binge eating disorder (BED) · Bulimia nervosa (BN) · Polymorphism

Abbreviations

SNP	Single Nucleotide Polymorphisms	CCK	cholecystokinin
ED	Eating Disorders	ARC	arcuate nucleus
BED	Binge Eating Disorder	PVN	paraventricular nucleus
BN	Bulimia Nervosa	DMN	dorsomedial nucleus
<i>MC4R</i>	Melanocortin 4 receptor	VMN	ventromedial nucleus
<i>OPRD1</i>	Opioid Receptor delta 1	LHA	lateral hypothalamic area
<i>BDNF</i>	Brain derived neurotrophic factor	NPY	neuropeptide Y
<i>FTO</i>	Fat mass and obesity associated	AgRP	agouti related peptide
VTA	ventral tegmental area VTA	POMC	pro-opiomelanocortin
NAc	nucleus accumbens	CART	cocaine- and amphetamine-regulated transcript
PYY	peptide tyrosine tyrosine	α -MSH	α -melanocyte-stimulating hormone
		MC-Rs	G protein coupled melanocortin receptors
		CRH	corticotropin-releasing hormone
		TRH	thyrotropin-releasing hormone
		CSF	lumbar cerebrospinal fluid
		DSR	dopamine 2 receptors
		<i>DAT</i>	dopamine transporter genes
		<i>DRD2</i>	dopamine receptor gene
		<i>5-HTTLPR</i>	serotonin transporter gene
		<i>BDNF</i>	brain derived neurotrophic factor
		ESR1	estrogen receptor 1
		ESR2	estrogen receptor 2
		CB1	cannabinoid receptor 1
		DAT	dopamine transporter
		<i>COMT</i>	catechol-O-methyltransferase

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1 Introduction

Obesity is a complex and chronic medical condition with a larger negative impact on human health [1], which has been associated with increased morbidity and considered a risk factor for cardiovascular disease, type 2 diabetes mellitus, hypertension, dyslipidemia and multiple cancers [2]. Obesity is often labeled and stigmatized, however there is a wide literature based on evidence that this is a complex disease caused by the interaction of multiple genetic, environmental, metabolic, psychological and behavioral factors [1].

Among genetic factors, single nucleotide polymorphisms (SNPs) can influence obesity risk; as well both gain and weight maintenance. Studies with twins and relatives have shown that the heritability of obesity ranges from 25 to 90%, confirming that a portion of the risk for the disease may be due to hereditary factors [3]. Also, genetic polymorphism is associated with different weight loss outcomes in patients submitted to diverse strategies for weight loss (clinical /or surgical) [4].

Obesity is closely associated with eating disorders (ED) and it shares several psychological and genetic factors [5]. According to a gene-environment approach, psychopathological differences can be influenced by genetic variability, which would explain the variance of susceptibility to ED [6]. The literature has shown the existence of neurobiological mechanisms common to these diseases that involve the regulation of food intake and emotion control [7]. An imbalance between appetite and satiety control associated with the rewarding aspect of food is possibly linked to genetic predisposition [8]. Thus, several confirmed genetic loci for obesity are expressed in regions of the brain that regulate energy consumption and reward seeking behavior [9].

In this case, the literature highlights the genes *MC4R* (Melanocortin 4 receptor), *OPRD1* (Opioid Receptor delta 1), *BDNF* (Brain derived neurotropic factor), *FTO* (Fat mass and obesity associated) and others, which have also been associated with mechanisms related to ED [9–12]. Thus, individuals with ED may be prone to food-related hedonic responses through genetic, dopaminergic, and opioidergic influences in reward-related processes [9, 13].

In this context, this review shows the role of polymorphisms involved with different eating disorders and its association with the pathophysiology of obesity (Fig. 1).

2 Obesity and food intake control

In the context of obesogenic environment, it is important to understand the mechanisms that control human appetite and satiety, which leads to the balance or imbalance between energy intake and energy expenditure, and consequently to overweight and obesity [14, 15]. Satiety can be defined as the

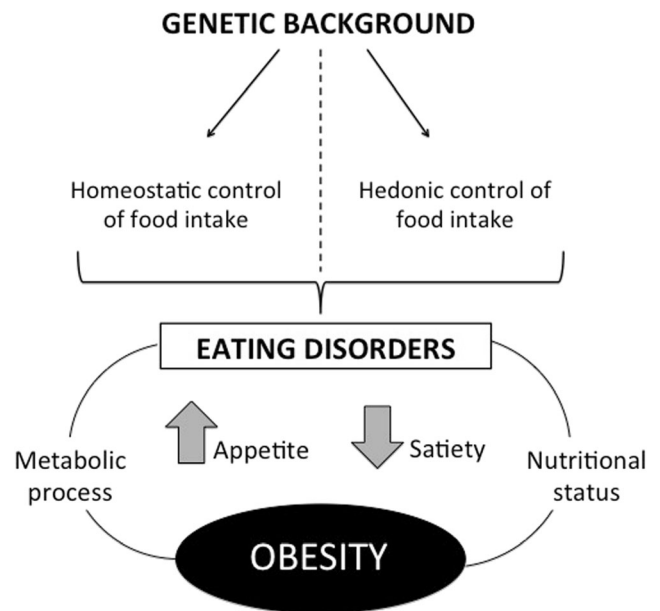


Fig. 1 Genetic polymorphism can affect appetite and satiety control, leading to eating disorders and consequently to obesity

feeling of fullness that persists after eating and, also is one important psycho-biological mechanism whose function is to inhibit intake following food ingestion, being able to suppress overconsumption [14]. In addition, satiety is modulated by macronutrient intake, as well as micronutrients, non-nutrients, bioactive food compounds and gut microbiota [15]. On the other hand, appetite is defined as the process that leads to the qualitative selection and quantitative ingestion of specific foods [16].

Briefly, gastric and intestinal signals interact with central nervous system pathways to terminate food intake [17]. The first phase of this control is the cephalic phase that denotes to the physiological preprandial responses due to olfaction and cognizance [18]. In post-prandial phase, food contact and digested food components in gastrointestinal track promotes a physical distension of stomach and hormones secretion that increase satiety levels. After absorption and digestion process, food metabolites promote secretion of hormones from adipose tissue (leptin) and pancreas (insulin), which suppress appetite through the hypothalamic circuits [19].

Thus, food intake is regulated by peripheral nutritional signals, through hormonal and neuronal pathways (Table 1) that are carried to specific brain areas, hypothalamic nuclei and the brainstem, both involved energy balance regulation (homeostatic regulation). However, the control of food intake is also performed by a hedonic behavior in which food ingestion occurs for pleasure regardless of the need for energy (non-homeostatic regulation). In this way, feeding is based on food reward properties and involves brain central areas such as the mesolimbic reward system, ventral tegmental area (VTA), nucleus accumbens (NAc), and the opioid, endocannabinoid and dopamine systems [20, 21].

Table 1 Summary of orexigenic and anorexigenic hormones/ peptides of hedonic food intake control

Tissue	Orexigenic factor	Anorexigenic factors
Gut	Ghrelin	CCK PYY PP Enterostatin GLP-1 Glucagon OXM
Neurons	NPY AgRP Orexin A MCH	POMC CART ACTH α -MSH
Adipose tissue		Leptin Insulin

CCK cholecystokinin, *PYY* peptide tyrosine tyrosine, *PP* pancreatic polypeptide, *GLP-1* glucagon-like peptide-1, *OXM* oxyntomodulin, *NPY* co-expressing neuropeptide Y, *AgRP* agouti related peptide, *MCH* melanin-concentrating hormone, *POMC* co-expressing pro-opiomelanocortin, *CART* cocaine- and amphetamine-regulated transcript, *ACTH* α -MSH: α -melanocyte-stimulating hormone

2.1 Homeostatic food intake control

2.1.1 Gut hormone and peptides

Gastrointestinal tract secretes hormones and peptides responsible for peripheral control of the food intake by controlling the initiation and termination of the meal. The biggest signal for the secretion of these peptides is gastric distension [22]. Among gut hormone and peptides, we highlight the peptide tyrosine tyrosine (PYY), cholecystokinin (CCK) and ghrelin.

CCK is secreted by I cells of the gastrointestinal tract, in response to the presence of lipids and protein, promotes prandial satiety and induces the secretion pancreatic, biliary secretion and vesicular contraction [23]. PYY peptide is expressed by the gut endocrine cells and acts as an inhibitor of food intake. On the other hand, ghrelin is secreted by gastric mucosal cells and is one of the most important factors in promote food intake, by increasing the appetite, and stimulates digestive secretions [23].

2.1.2 Peripheral adiposity signals

Leptin and insulin act as peripheral adiposity signals for informing the brain about the adipose tissue mass. Leptin, a hormone produced in white adipose tissue, promotes satiety and regulate energy balance for acting on its receptors expressed in the hypothalamus [24]. In arcuate nucleus (ARC), leptin exerts anorexigenic effect via inhibition of neuropeptide Y (NPY)/ agouti related peptide (AgRP) neurons

and activation of co-expressing pro-opiomelanocortin (POMC)/cocaine- and amphetamine-regulated transcript (CART) neurons [25]. Insulin is produced by the pancreatic β -cells and has an important function in the central nervous system to increase satiety, energy expenditure and regulate action of leptin [26].

2.1.3 Neuronal signals

The hypothalamus plays a major role in the control of the appetite by processing afferent signals from the gut and brainstem. Specific neuronal regions take part in the regulation of food intake such as ARC, the paraventricular nucleus (PVN), the dorsomedial nucleus (DMN), the ventromedial nucleus (VMN) and the lateral hypothalamic area (LHA) [27].

In the ARC nucleus, different neurons with opposing effects act in food intake control: 1. co-expressing NPY and AgRP neurons, which stimulate appetite and 3. POMC and 4. CART neurons, that suppress the feeding (stimulate satiety) [25]. Also, in ARC, the cleavage of POMC results in the production of α -melanocyte-stimulating hormone (α -MSH) that exert their effects by binding to G protein coupled melanocortin receptors (MC-Rs). On the other hand, PVN nucleus contains neurons that secrete peptides responsible to reduce food consumption (i.e. corticotropin-releasing hormone (CRH), oxytocin, and thyrotropin-releasing hormone (TRH)).

2.2 Brain reward systems and the hedonic food intake control

Brain reward systems involved in eating behavior have two mechanisms, one related with motivating the desire for food, referred to as ‘wanting’, and another mechanism related with the hedonic properties of the food, referred to as ‘liking’. Since dopamine (DA) is predominantly involved in ‘wanting’, opioids and cannabinoids are predominantly involved in food ‘liking’ [28].

The dopaminergic system is a central component of the hedonic feeding. DA is a key neurotransmitter able to modulate reward, which does mainly through its projections from the VTA into the NAc [29]. DA receptors are expressed in specific hypothalamic nucleus and its effects depend on the specific region of its actions [30, 31]. The DA acts in two major regions, LHA and VMH that have opposed effects on dietary intake. In the LHA area, dopamine levels increased immediately with the meal, remain high during the meal consumption, and normalize when it ends [32, 33]. In contrast, in the VMH region, dopamine levels decrease after food ingestion and increase during fast [32].

Conversely, the role of DA in mesolimbic/mesocortical has been associated with reward processing in reaction to feeding stimuli [34]. Environmental stimulus induces burst-firing of

dopamine neurons with subsequent phasic DA release. This increase in DA signaling will motivate specific behaviors, such as ‘wanting’ food [28, 35]. On the other hand, the activation of the opioid and cannabinoid systems seems to stimulate in part by enhancing the ‘liking’ of the food. Although the brain reward systems involved in eating behavior are separate, they act together to regulating food intake [28].

Also, serotonin (5-HT) has been implicated in the control of food intake [36]. Brain 5-HT interacts within the hypothalamus and acts on central melanocortin neurons to promote satiety. Activation of 5-HT_{2c} receptor increases the activity of anorexigenic POMC neurons and the activation of 5-HT_{1b} receptor inhibits the orexigenic NPY/AgRP neurons [37]. In the nucleus of the solitary tract serotonin integrates peripheral satiety signals [36].

2.3 Cross-talk between homeostatic and hedonic systems

There is a crosstalk between the neurochemical substrates of the two systems [38] in which food deprivation or satiation affects attraction for food (Fig. 2). Interestingly, this cross-talk is mediated by the direct action of leptin, insulin, GLP-1 and ghrelin, on the mesocorticolimbic dopamine system [39]. In

addition to acting through peripheral pathways by stimulating energy storage, insulin can also excite specific brain receptors (negative feedback), reducing activity in mesolimbic dopamine circuits and consequently food reward [40, 41]. In the same direction, GLP-1 signaling in the mesolimbic system can have a selective effect to reduce the rewarding value for tasty food [42]. In addition, ghrelin has an important role in increasing the neural response to food photos in reward-related circuits [43]. In line of this, the increase of signalling by the stomach-derived orexigenic hormone, ghrelin, can activate the mesoaccumbal dopamine system and the reward from palatable food [44]. Besides these direct mechanisms described between the two systems, there are indirect effects mediated by heterogeneous projections from the LH to the VTA, including neurons expressing orexin [45], gamma-aminobutyric acid (GABA) and glutamate [46, 47].

In this context, human biological system provides innumerable neuroendocrine inputs for food intake control, exerting effects on modulation of appetite and the satiety signs. As above, its regulation is very complex, engaging several molecular interactions with many tissues, hormones, and neural circuits. Thus, signaling molecules that affect food intake are critical for normal energy homeostasis [26, 48]. In addition, the homeostatic and hedonic circuits act together to promote

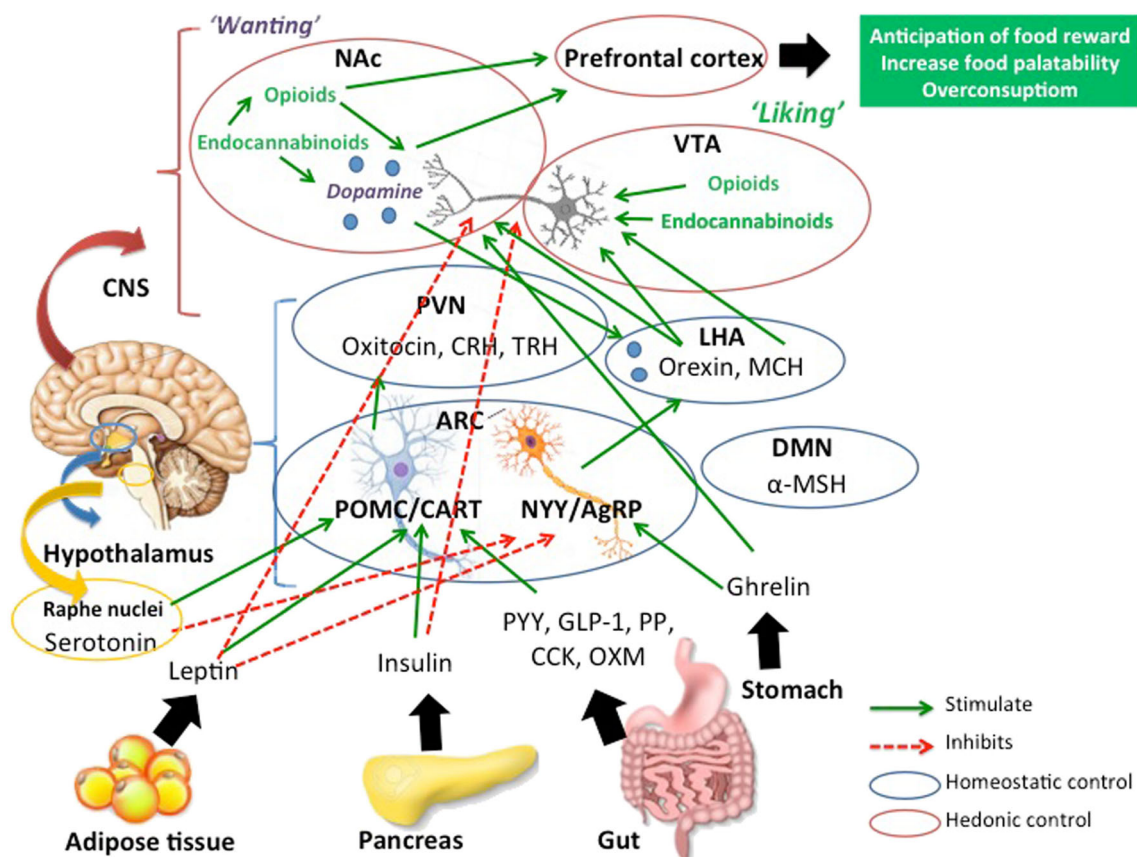


Fig. 2 Cross-talk between homeostatic and hedonic regulation of food intake. The action of gut hormones, neural and hormonal signals (peripheral signals), dopamine and serotonin in hypothalamus and central nervous system

food intake in times of deprivation and to inhibit food intake in satiety conditions. Interruption of the interaction between these circuits may promote the development of EDs such as Bulimia Nervosa (BN) and Binge Eating Disorder (BED) and contribute to the development and/or aggravation of obesity [28]. An increase in EDs and in atypical eating patterns such as binge eating and weight control behaviors associated with EDs and obesity have been reported globally [49], and the next chapter addresses EDs main characteristics.

3 Eating disorders associated with obesity: Binge eating disorder (BED) and bulimia nervosa (BN)

EDs are characterized by regular and persistent disturbances in eating or eating behavior, culminating in the consumption and altered absorption of food, significantly compromising physical health and psychosocial functioning, thus generating a reduction in quality of life [50]. The etiology of EDs is multifactorial, involving biological, psychological and socio-cultural factors [51]. Among the biological aspects, the genetic factors have a significantly influence of the regulation of neural circuits controlling the appetite and satiety pathways, as well as the regulation of brain reward systems [52, 53].

Several studies have shown a significant association between EDs and obesity, and the most prevalent EDs in obese subjects are BED and BN [54]. In the adult population the prevalence is about 2–5% for BED and 1% for BN, being more frequent in females [55–57]. Recurrent episodes of binge eating are the main feature of both conditions and are characterized by the consumption of an excessive amount of food associated with the feeling of loss of control over food. This feeling of loss of control of the amount of food intake during a binge eating episode is one of the most important clinical features in this population and contributes to the feeling of guilt, disappointment, sadness and depression [54, 58, 59]. The Diagnostic and Statistical Manual of Mental Disorders (DSM-5) classifies and establishes some criteria for the diagnosis of these eating disorders.

3.1 Binge eating disorder (BED)

BED is characterized by recurrent episodes of binge eating that occur at least once a week over a three-month period associated with lack of control and pronounced distress. Episodes of binge eating are usually associated with eating faster than normal, eating a quantity that exceeds gastric capacity, eating in large quantities in the absence of physical hunger, eating alone because of the amount of food consumed, and the feeling of guilt after episodes of compulsion. However, it is not associated with inappropriate compensatory

strategies after compulsion. Therefore BED is often associated with obesity [50, 55, 60].

3.2 Bulimia nervosa (BN)

BN is characterized by self-assessment and excessive preoccupation with body image, recurrent eating compulsion and inappropriate compensatory behaviors to avoid weight gain, such as self-induced vomiting, misuse of laxatives, diuretics, fasting and excessive physical exercise. According to DSM-5, binge eating episodes and compensatory behaviors should occur at least once a week for three months for the diagnosis of bulimia [50, 60].

3.3 Physiopathology of EDs

The homeostatic control of body energy balance is exerted by specific populations of neurons located, for the most part, in the hypothalamic nuclei, which through a neuroendocrine pathway respond to fluctuations in the energetic state, altering the expression of neuropeptides, resulting in changes in ingestion and energy expenditure [61, 62].

In addition to homeostatic regulation, hedonic controls of appetite exert a great influence on food intake, being related to feelings of pleasure and reward. Thus, there is a complex interaction between hedonic controls and homeostatic controls for regulating food intake as well as regulating body weight [38].

Brain reward systems mechanisms are closely involved in the pathophysiology of binge eating behavior, which is a symptom present in EDs (BN and BED) [63]. Multiple neurotransmitter systems (dopaminergic, serotonergic, opioidergic, GABAergic and glutamatergic) contribute to binge eating episodes, but dopaminergic, serotonergic and opioidergic neurotransmission are essential for the consumption of reward-related foods [64, 65].

DA is a key neurotransmitter involved in the reward and induction aspects of feeding. Some studies have shown that subjects who presented BN have lower levels of dopamine metabolites in lumbar cerebrospinal fluid (CSF) when compared to healthy individuals. Some studies have provided evidence that the decrease in DA signaling in striatal regions, either by reduction in the action or number of dopamine 2 receptors (D2R) or in DA release, is related to a reduction in sensitivity to natural rewards, in which obese individuals with EDs can compensate by eating more, especially high-fat foods [66–68]. Based on this, some studies have observed that individuals with polymorphisms in dopamine transporter genes (*DAT*), as well as in the dopamine receptor gene (*DRD2*) showed a change in the turnover of DA. These findings support the fact that there is a change in DA metabolism in BN, affecting the hedonic processes associated with food induction and reward mechanisms [52, 69].

5-HT also plays an important role, related to satiety. In animals and in humans, manipulations that increase 5-HT neurotransmission lead to reduced food intake, whereas those that reduce 5-HT activity lead to binge eating [70]. These tendencies lead to the expectation that EDs characterized by binge eating (such as BED and BN) should be related to reduced 5-HT activity [71] and it is evident that the deregulation of 5-HT system, like polymorphisms related to the serotonin transporter gene (*5-HTTLPR*), is associated with EDs, such as BED and BN [65].

4 Genetic polymorphisms and eating disorders associated with obesity

Genetic factors also contribute to changes in eating behavior and to the development of EDs [72]. Despite the mechanisms mentioned above in the appetite and satiety control, individual genetic variability (characterized by SNP) in the predisposition for obesity development and in the biological response to weight loss makes this regulation so complex [48]. In line of this, there are evidences among interactions of common genetic variants, altered circulating gut hormone levels and obesity-risk effects [73, 74]. These SNPs, or other variants located in hypothalamus appetite and satiety related genes, may be involved in the develop unusual food intake behavior/patterns that favor the establishment of EDs. In addition to SNPs in genes related to hypothalamic appetite and satiety mechanisms, some SNPs in genes involved in multiple neurotransmitter systems (dopaminergic, serotonergic, opioidergic, GABAergic and glutamatergic) contribute to the development of major EDs related to obesity, which will be discussed in the next subtopics [75].

4.1 Genetic polymorphisms and BED

With the advancement of studies in genomic medicine, estimates of heritability have been demonstrated for BED with variation from 41 to 57% [76–78], in addition to the association of BED with some SNPs [79–86].

In relation to SNPs in genes involved with the mechanisms of appetite and satiety, one is related to the hunger hormone, ghrelin, which stimulates the release of orexigenic neuropeptides NPY and AgRP, increasing food intake, was identified that carriers of the Leu72Met variant of the ghrelin gene (*GHRL*), that is possibly involved with some physiological effects by bidding to ghrelin receptors, was associated with BED in preliminary results in a pilot study [87]. A recent systematic review and meta-analysis that evaluated a possible association between coding variants in the melanocortin 4 receptor gene (*MC4R*), which is the α -MSH receptor and is involved in inhibiting food intake and increasing the energy expenditure, with BED in obese individuals, showed a

positive association between variants of gain-of-function (GOF) in *MC4R* and risk of BED [88].

Dopamine is an important neurotransmitter that is involved in a wide variety of brain functions, including eating behavior and reward mechanisms. Based on this, dopaminergic SNPs have been studied and some have been associated with BED, such as dopamine D2 receptor (*DRD2*) and neighboring ankyrin repeat and kinase domain containing 1 (*ANKK1*) [13, 89, 90]. The most prevalent variations in individuals with BED are *Taq1A*⁺ allele (A1/A1 and A1/A2 genotypes), which are associated with a reduced DA function, due to a 30–40% reduction in *DRD2* density in the striated and diminished glucose metabolism in brains of these individuals [13, 91, 92]. The *VNTR* polymorphisms of the human dopamine transporter (*DAT1*) are also related to BED [93].

5-HT is another neurotransmitter that is involved in eating behavior, and serotonergic genes and their relation in EDs have been studied extensively [94]. In relation to the SNPs involved with the serotonergic system and BED, one study observed a significantly higher frequency of LL genotype and L allele for polymorphism in the *5-HTTLPR* of the serotonin transporter gene (*5HTT*) in individuals with BED ($n = 77$), compared to individuals without BED ($n = 61$) [95].

The endogenous opioid system in reward processes presents a major role in μ -opioid receptors, which activation enhances positive hedonic aspects to eat more sweet and fatty foods [96] and one study revealed that individuals with obesity and BED ($n = 66$) had a higher “Gain of function” G allele prevalence of rs1799971 polymorphism for the *OPRM1* gene (μ 1 opioid receptor gene), compared to individuals with obesity without BED ($n = 70$) [13].

The brain derived neurotrophic factor (BDNF) and its tyrosine kinase receptor are expressed in various hypothalamic nuclei and when active decrease food ingestion and increase energy expenditure [97]. One cohort study in women with obesity identified a greater association between the 66met allele of the *BDNF* rs6265 and BED [82], and a higher frequency and more severe episodes of binge eating in those carrying the 196G/A polymorphism (val66met) for the *BDNF* gene [95] thus suggesting a relationship of these SNPs with the BED. These polymorphisms related to BED are presented in Table 2.

4.2 Genetic polymorphisms and BN

BN is also familial and individuals who have a relative with BN are at elevated risk for developing this ED [98, 99]. Although few studies show a direct association between SNPs and BN [100–102], others associate some SNPs with the aggravation of psychological symptoms of this disorder [102, 103] and we will mention those of greater relevance in the current literature.

Table 2 Overview of SNPs findings in eating disorders

Gene	Description	Polymorphism	ED	References
<i>GHRL</i>	Ghrelin	rs696217	BED	Monteleone et al. [87]
<i>MC4R</i>	Melanocortin 4 receptor	rs2229616 (Val103Ile) rs52820871 (Ile251Leu)	BED	Qasin et al. [115] Potoczna et al. [116]
<i>DRD2</i>	Dopamine receptor D2	rs6277 (957 C/T)	BED	Davis et al. [13, 89, 90]
<i>ANKK1</i>	Ankyrin repeat and kinase domain containing I	rs1800497 (Taq1A)	BED	Davis et al. [13, 89, 90]
<i>DAT1</i>	Dopamine transporter 1	rs2270912 rs28363130	BED	Hirata et al. [117]
<i>SLC6A4</i>	Serotonin transporter	5-HTTLPR	BED	Monteleone et al. [95]
<i>OPRM1</i>	Mu I opioid receptor	rs1799971 (118A/G)	BED	Davis et al. [13]
<i>BDNF</i>	Brain derived neurotrophic factor	rs6265 (Val66Met)	BED	Beckers et al. [82]
<i>ESR1</i>	Estrogen receptor 1	rs928554	BN	Nilsson et al. [118]
<i>CNR1</i>	Cannabinoid receptor 1	rs1049353	BN	Monteleone et al. [100]
<i>SLC6A4</i>	Serotonin transporter	5-HTTLPR	BN	Steiger et al. [103, 107]; Thaler et al. [119]
<i>DRD2</i>	Dopamine receptor D2	rs1800497 (Taq1A)	BN	Thaler et al. [102]
<i>COMT</i>	Catechol-O-methyltransferase	rs4680 (Val158Met)	BN	Thaler et al. [102]
<i>FTO</i>	Fat mass and obesity associated	rs9939609	BN	Müller et al. [101]

The role of sex hormones in the development or aggravation of EDs has recently gained research attention [104] and as estrogen has also been implicated in feeding behavior, the focus has been on the estrogen receptor 1 (ESR1) and estrogen receptor 2 (ESR2 or ER β). One study showed a positive association between ER β rs928554 SNP and BN in 76 women, which may be related to a reduction in the ER β function [105].

The endocannabinoid system, with activation of cannabinoid receptor 1 (CB1), is involved in the modulation of energy balance by controlling food intake, possibly with the mesolimbic pathway involved in reward mechanisms and by the hypothalamus, interacting with other orexigenic and anorexigenic mediators to modulate appetite and satiety [105, 106]. One study identified that the *CNR1* (gene coding the CB1 receptor) rs1049353 SNP is significantly associated with BN patients ($n = 149$), which may have functional effects by altering mRNA stability or translation of CB1 [100].

5-HT has an important role in food intake and is involved in BN [94]. Based on this, genetic factors associated with reduced 5-HT neurotransmission, like low-function alleles of the serotonin transporter polymorphism (5-HTTLPR) have been linked to BN in some studies [102, 103, 107].

The dopaminergic system is a system of interest in BN, due to demonstrated associations between dopamine (DA) activity and food intake [108], and the fact that people with BN display decrease DA metabolites [109] and decreased DA transporter (DAT) availability [110]. Some associations between polymorphisms acting upon postsynaptic receptors like dopamine D2 receptor (*DRD2*) TaqA1 rs1800497 and dopamine D4 receptor (*DRD4*) 7R and dopamine regulators catechol-O-

methyltransferase (*COMT*) rs4680 and DA transporter 1 (*DAT1*) are related to increased psychopathology and body mass in women with BN [102]. One study shows that the hypofunctional 7R allele of *DRD4* contributes to weight gain in Caucasian women with BN and that the Met66 allele of *BDNF* gene acts with *DRD4* to influence weight regulation in these women [111].

Several studies have demonstrated a strong association between SNPs in the fat mass and obesity associated gene (*FTO*) and obesity in different age groups [112–114]. The *FTO* SNP rs9939609 which is strongly associated with obesity, also was associated with BN in 477 patients in a German study [100]. These polymorphisms related to BN are presented in Table 2.

5 Conclusion

This review manuscript highlights that genetic and psychological factors are closely associated with eating disorders, including BED and BN, and to obesity development. Neurobiological mechanisms in common to these diseases are involved in food intake and emotion control, and to an imbalance between appetite and satiety associated with the rewarding aspect of food, which is possibly linked to genetic predisposition. Given the evidence found in current literature, it is safe to say that there is a clear link between these diseases that needs to be explored in the field of molecular biology. Therefore, it is important to highlight the relevance in the identification of genetic risk factors for a possible personalized treatment and consequently more effective for these

patients. However, it is also important to point out that obesity treatment success remains a challenge, partly since obesity pathophysiology is orchestrated by complex interactions among environment, genetic background and behaviors. Thus, the analysis of polymorphisms can be a useful tool in obesity treatment, mainly in the early diagnosis of eating disorders, therefore it should be used by trained professionals with caution.

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Compliance with ethical standards

Conflicts of interest The authors declare no conflicts of interest.

References

- Upadhyay J, Farr O, Perakakis N, Ghaly W, Mantzoros C. Obesity as a disease. *Med Clin N Am*. 2018;102:13–33.
- Flegal KM, Kit BK, Orpana H, Graubard BI. Association of all-cause mortality with overweight and obesity using standard body mass index categories: a systematic review and meta-analysis. *JAMA*. 2013;309:71–82.
- Ng M, Fleming T, Robinson M, Thomson B, Graetz N, Margono C, et al. Global, regional, and national prevalence of overweight and obesity in children and adults during 1980–2013: a systematic analysis for the global burden of disease study 2013. *Lancet*. 2014;384:766–81.
- Sarzynski MA, Jacobson P, Rankinen T, Carlsson B, Sjostrom L, Bouchard C, et al. Associations of markers in 11 obesity candidate genes with maximal weight loss and weight regain in the SOS bariatric surgery cases. *Int J Obes*. 2011;35:676–83.
- Peckmezian T, Hay P. A systematic review and narrative synthesis of interventions for uncomplicated obesity: weight loss, well-being and impact on eating disorders. *J Eat Disord*. 2017;5:15.
- Steiger H, Richardson J, Schmitz N, Joober R, Israel M, Bruce KR, et al. Association of trait-defined, eating-disorder sub-phenotypes with (biallelic and triallelic) 5HTTLPR variations. *J Psychiatr Res*. 2009;43:1086–94.
- Gorwood P, Blanchet-Collet C, Chartrel N, Duclos J, Dechelotte P, Hanachi M, et al. New insights in anorexia nervosa. *Front Neurosci*. 2016;10:256.
- Sardahaee FS, Holmen TL, Micali N, Kvaloy K. Effects of single genetic variants and polygenic obesity risk scores on disordered eating in adolescents - the HUNT study. *Appetite*. 2017;118:8–16.
- Castellini G, Franzago M, Bagnoli S, Lelli L, Balsamo M, Mancini M, et al. Fat mass and obesity-associated gene (FTO) is associated to eating disorders susceptibility and moderates the expression of psychopathological traits. *PLoS One*. 2017;12:e0173560.
- Hinney A, Volckmar AL. Genetics of eating disorders. *Curr Psychiatry Rep*. 2013;15(12):423. <https://doi.org/10.1007/s11920-013-0423-y>.
- Micali N, Field AE, Treasure JL, Evans DM. Are obesity risk genes associated with binge eating in adolescence? *Obesity (Silver Spring)* 2015; 23: 1729–1736.
- Locke AE, Kahali B, Berndt SI, Justice AE, Pers TH, Day FR, et al. Genetic studies of body mass index yield new insights for obesity biology. *Nature*. 2015;518:197–206.
- Davis CA, Levitan RD, Reid C, Carter JC, Kaplan AS, Patte KA, et al. Dopamine for "wanting" and opioids for "liking": a comparison of obese adults with and without binge eating. *Obesity (Silver Spring)*. 2009;17:1220–5.
- Njike VY, Smith TM, Shuval O, Shuval K, Edshteyn I, Kalantari V, et al. Snack food, satiety, and weight. *Adv Nutr*. 2016;7:866–78.
- Tremblay A, Bellisle F. Nutrients, satiety, and control of energy intake. *Appl Physiol Nutr Metab*. 2015;40:971–9.
- Blundell JE, Halford JCG. Appetite: Physiological and Neurobiological Aspects. *Encyclopedia of Human Nutrition*. London: Academic Press, 1998, p. 121–126.
- Chambers AP, Sandoval DA, Seeley RJ. Integration of satiety signals by the central nervous system. *Curr Biol*. 2013;23:R379–88.
- Korner J, Leibel RL. To eat or not to eat - how the gut talks to the brain. *N Engl J Med*. 2003;349:926–8.
- Rui L. Brain regulation of energy balance and body weight. *Rev Endocr Metab Disord*. 2013;14:387–407.
- Matafome P, Seiça R. The role of brain in energy balance. *Adv Neurobiol*. 2017;19:33–48. https://doi.org/10.1007/978-3-319-63260-5_2.
- Yu YH, Vasselli JR, Zhang Y, Mechanick JI, Korner J, Peterli R. Metabolic vs. hedonic obesity: a conceptual distinction and its clinical implications. *Obes Rev*. 2015;16:234–47.
- Martin CK, Bellanger DE, Rau KK, Coulon S, Greenway FL. Safety of the Ullorex oral intragastric balloon for the treatment of obesity. *J Diabetes Sci Technol*. 2007;1:574–81.
- Konturek SJ, Konturek JW, Pawlik T, Brzozowski T. Brain-gut axis and its role in the control of food intake. *J Physiol Pharmacol*. 2004;55:137–54.
- Trayhum P, Bing C. Appetite and energy balance signals from adipocytes. *Philos Trans R Soc Lond Ser B Biol Sci*. 2006;361:1237–49.
- Suzuki K, Simpson KA, Minnion JS, Shillito JC, Bloom SR. The role of gut hormones and the hypothalamus in appetite regulation. *Endocr J*. 2010;57:359–72.
- Schwartz MW, Woods SC, Porte D Jr, Seeley RJ, Baskin DG. Central nervous system control of food intake. *Nature*. 2000;404:661–71.
- Stanley S, Wynne K, McGowan B, Bloom S. Hormonal regulation of food intake. *Physiol Rev*. 2005;85:1131–58.
- Volkow ND, Wang GJ, Baler RD. Reward, dopamine and the control of food intake: implications for obesity. *Trends Cogn Sci*. 2011;15:37–46.
- Wise RA. Role of brain dopamine in food reward and reinforcement. *Philos Trans R Soc Lond Ser B Biol Sci*. 2006;361:1149–58.
- Ramos EJ, Meguid MM, Campos AC, Coelho JC. Neuropeptide Y, alpha-melanocyte-stimulating hormone, and monoamines in food intake regulation. *Nutrition*. 2005;21:269–79.
- Hernandez L, Hoebel BG. Food intake and lateral hypothalamic self-stimulation covary after medial hypothalamic lesions or ventral midbrain 6-hydroxydopamine injections that cause obesity. *Behav Neurosci*. 1989;103:412–22.
- Meguid MM, Yang ZJ, Koseki M. Eating induced rise in LHA-dopamine correlates with meal size in normal and bullectomized rats. *Brain Res Bull*. 1995;36:487–90.
- Yang ZJ, Koseki M, Meguid MM, Laviano A. Eating-related increase of dopamine concentration in the LHA with oronasal stimulation. *Am J Phys*. 1996;270:R315–8.
- Berridge KC, Robinson TE, Aldridge JW. Dissecting components of reward: 'liking', 'wanting', and learning. *Curr Opin Pharmacol*. 2009;9:65–73.
- Berridge KC. The debate over dopamine's role in reward: the case for incentive salience. *Psychopharmacology*. 2007;191:391–431.

36. Voigt JP, Fink H. Serotonin controlling feeding and satiety. *Behav Brain Res*. 2015;277:14–31.
37. Wyler SC, Lord CC, Lee S, Elmquist JK, Liu C. Serotonergic control of metabolic homeostasis. *Front Cell Neurosci*. 2017;11:277.
38. Berthoud HR, Munzberg H, Morrison CD. Blaming the brain for obesity: integration of hedonic and homeostatic mechanisms. *Gastroenterology*. 2017;152:1728–38.
39. Jerlhag E, Janson AC, Waters S, Engel JA. Concomitant release of ventral tegmental acetylcholine and accumbal dopamine by ghrelin in rats. *PLoS One*. 2012;7:e49557.
40. Figlewicz DP. Adiposity signals and food reward: expanding the CNS roles of insulin and leptin. *Am J Phys Regul Integr Comp Phys*. 2003;284:R882–92.
41. Mebel DM, Wong JC, Dong YJ, Borgland SL. Insulin in the ventral tegmental area reduces hedonic feeding and suppresses dopamine concentration via increased reuptake. *Eur J Neurosci*. 2012;36:2336–46.
42. Farr OM, Sofopoulos M, Tsoukas MA, Dincer F, Thakkar B, Sahin-Efe A, et al. GLP-1 receptors exist in the parietal cortex, hypothalamus and medulla of human brains and the GLP-1 analogue liraglutide alters brain activity related to highly desirable food cues in individuals with diabetes: a crossover, randomised, placebo-controlled trial. *Diabetologia*. 2016;59:954–65.
43. Malik S, McGlone F, Bedrossian D, Dagher A. Ghrelin modulates brain activity in areas that control appetitive behavior. *Cell Metab*. 2008;7:400–9.
44. Egecioglu E, Skibicka KP, Hansson C, Alvarez-Crespo M, Friberg PA, Jerlhag E. Hedonic and incentive signals for body weight control. *Rev Endocr Metab Disord*. 2011;12:141–51.
45. Harris GC, Wimmer M, Aston-Jones G. A role for lateral hypothalamic orexin neurons in reward seeking. *Nature*. 2005;437:556–9.
46. Nieh EH, Matthews GA, Allsop SA, Presbrey KN, Leppla CA, Wichmann R, et al. Decoding neural circuits that control compulsive sucrose seeking. *Cell*. 2015;160:528–41.
47. Nieh EH, Vander Weele CM, Matthews GA, Presbrey KN, Wichmann R, Leppla CA, et al. Inhibitory input from the lateral hypothalamus to the ventral tegmental area disinhibits dopamine neurons and promotes behavioral activation. *Neuron*. 2016;90:1286–98.
48. MacLean PS, Blundell JE, Mennella JA, Batterham RL. Biological control of appetite: a daunting complexity. *Obesity (Silver Spring)*. 2017;25(Suppl 1):S8–S16.
49. Pike KM, Dunne PE. The rise of eating disorders in Asia: a review. *J Eat Disord*. 2015;3:33.
50. Association AP. *Diagnostic and Statistical Manual of Mental Disorders: DSM-5*. 5th ed ed. Arlington: American Psychiatric Association; 2013.
51. Culbert KM, Racine SE, Klump KL. Research review: what we have learned about the causes of eating disorders - a synthesis of sociocultural, psychological, and biological research. *J Child Psychol Psychiatry*. 2015;56:1141–64.
52. Mitchison D, Hay PJ. The epidemiology of eating disorders: genetic, environmental, and societal factors. *Clin Epidemiol*. 2014;6:89–97.
53. Marcus MD. Obesity and eating disorders: articles from the international journal of eating disorders 2017-2018. *Int J Eat Disord*. 2018;51:1296–9.
54. Palavras MA, Hay P, Filho CA, Claudino A. The efficacy of psychological therapies in reducing weight and binge eating in people with bulimia nervosa and binge eating disorder who are overweight or obese—a critical synthesis and meta-analyses. *Nutrients*. 2017;9.
55. Treasure J, Claudino AM, Zucker N. Eating disorders. *Lancet*. 2010;375:583–93.
56. Smink FR, van Hoeken D, Hoek HW. Epidemiology of eating disorders: incidence, prevalence and mortality rates. *Curr Psychiatry Rep*. 2012;14:406–14.
57. Himmerich H, Treasure J. Psychopharmacological advances in eating disorders. *Expert Rev Clin Pharmacol*. 2018;11:95–108.
58. Conceicao E, Bastos AP, Brandao I, Vaz AR, Ramalho S, Arrojado F, et al. Loss of control eating and weight outcomes after bariatric surgery: a study with a Portuguese sample. *Eat Weight Disord*. 2014;19:103–9.
59. Meany G, Conceicao E, Mitchell JE. Binge eating, binge eating disorder and loss of control eating: effects on weight outcomes after bariatric surgery. *Eur Eat Disord Rev*. 2014;22:87–91.
60. da Luz FQ, Hay P, Touyz S, Sainsbury A. Obesity with comorbid eating disorders: associated health risks and treatment approaches. *Nutrients*. 2018;10.
61. Velloso LA, Schwartz MW. Altered hypothalamic function in diet-induced obesity. *Int J Obes*. 2011;35:1455–65.
62. Wilson JL, Enriori PJ. A talk between fat tissue, gut, pancreas and brain to control body weight. *Mol Cell Endocrinol*. 2015;418(Pt 2):108–19.
63. Monteleone AM, Piscitelli F, Dalle Grave R, El Ghoch M, Di Marzo V, Maj M, et al. Peripheral endocannabinoid responses to hedonic eating in binge-eating disorder. *Nutrients*. 2017;9.
64. Mathes WF, Brownley KA, Mo X, Bulik CM. The biology of binge eating. *Appetite*. 2009;52:545–53.
65. Avena NM. The study of food addiction using animal models of binge eating. *Appetite*. 2010;55:734–7.
66. Geiger BM, Haburcak M, Avena NM, Moyer MC, Hoebel BG, Pothos EN. Deficits of mesolimbic dopamine neurotransmission in rat dietary obesity. *Neuroscience*. 2009;159:1193–9.
67. Wang GJ, Volkow ND, Logan J, Pappas NR, Wong CT, Zhu W, et al. Brain dopamine and obesity. *Lancet*. 2001;357:354–7.
68. Johnson PM, Kenny PJ. Dopamine D2 receptors in addiction-like reward dysfunction and compulsive eating in obese rats. *Nat Neurosci*. 2010;13:635–41.
69. Bello NT, Hajnal A. Dopamine and binge eating behaviors. *Pharmacol Biochem Behav*. 2010;97:25–33.
70. Blundell JE. Serotonin manipulations and the structure of feeding behaviour. *Appetite*. 1986;7(Suppl):39–56.
71. Steiger H. Eating disorders and the serotonin connection: state, trait and developmental effects. *J Psychiatry Neurosci*. 2004;29:20–9.
72. Klump KL, Suisman JL, Burt SA, McGue M, Iacono WG. Genetic and environmental influences on disordered eating: an adoption study. *J Abnorm Psychol*. 2009;118:797–805.
73. Panaro BL, Tough IR, Engelstoft MS, Matthews RT, Digby GJ, Moller CL, et al. The melanocortin-4 receptor is expressed in enteroendocrine L cells and regulates the release of peptide YY and glucagon-like peptide 1 *in vivo*. *Cell Metab*. 2014;20:1018–29.
74. Zhang X, Qi Q, Zhang C, Smith SR, Hu FB, Sacks FM, et al. FTO genotype and 2-year change in body composition and fat distribution in response to weight-loss diets: the POUNDS LOST trial. *Diabetes*. 2012;61:3005–11.
75. Yilmaz Z, Hardaway JA, Bulik CM. Genetics and epigenetics of eating disorders. *Adv Genomics Genet*. 2015;5:131–50.
76. Hudson JI, Lalonde JK, Berry JM, Pindyck LJ, Bulik CM, Crow SJ, et al. Binge-eating disorder as a distinct familial phenotype in obese individuals. *Arch Gen Psychiatry*. 2006;63:313–9.
77. Mitchell KS, Neale MC, Bulik CM, Aggen SH, Kendler KS, Mazzeo SE. Binge eating disorder: a symptom-level investigation of genetic and environmental influences on liability. *Psychol Med*. 2010;40:1899–906.
78. Javaras KN, Laird NM, Reichborn-Kjennerud T, Bulik CM, Pope HG Jr, Hudson JI. Familiality and heritability of binge eating

- disorder: results of a case-control family study and a twin study. *Int J Eat Disord*. 2008;41(2):174–9.
79. Levitan RD, Masellis M, Basile VS, Lam RW, Kaplan AS, Davis C, et al. The dopamine-4 receptor gene associated with binge eating and weight gain in women with seasonal affective disorder: an evolutionary perspective. *Biol Psychiatry*. 2004;56:665–9.
 80. Lebrun B, Bariouhay B, Moyse E, Jean A. Brain-derived neurotrophic factor (BDNF) and food intake regulation: a minireview. *Auton Neurosci*. 2006;126-127:30–8.
 81. Helder SG, Collier DA. The genetics of eating disorders. *Curr Top Behav Neurosci*. 2011;6:157–75.
 82. Beckers S, Peeters A, Zegers D, Mertens I, Van Gaal L, Van Hul W. Association of the BDNF Val66Met variation with obesity in women. *Mol Genet Metab*. 2008;95:110–2.
 83. Kessler RM, Hutson PH, Herman BK, Potenza MN. The neurobiological basis of binge-eating disorder. *Neurosci Biobehav Rev*. 2016;63:223–38.
 84. Feng J. Role of Cyfip2 in binge eating. *Biol Psychiatry*. 2017;81:e77–8.
 85. Steiger H, Thaler L, Gauvin L, Joober R, Labbe A, Israel M, et al. Epistatic interactions involving DRD2, DRD4, and COMT polymorphisms and risk of substance abuse in women with binge-purge eating disturbances. *J Psychiatr Res*. 2016;77:8–14.
 86. Rozenblat V, Ong D, Fuller-Tyszkiewicz M, Akkermann K, Collier D, Engels R, et al. A systematic review and secondary data analysis of the interactions between the serotonin transporter 5-HTTLPR polymorphism and environmental and psychological factors in eating disorders. *J Psychiatr Res*. 2017;84:62–72.
 87. Monteleone P, Tortorella A, Castaldo E, Di Filippo C, Maj M. The Leu72Met polymorphism of the ghrelin gene is significantly associated with binge eating disorder. *Psychiatr Genet*. 2007;17:13–6.
 88. Qasim A, Mayhew AJ, Ehtesham S, Alyass A, Volckmar AL, Herpertz S, Hinney A, et al. Gain-of-function variants in the melanocortin 4 receptor gene confer susceptibility to binge eating disorder in subjects with obesity: a systematic review and meta-analysis. *Obes Rev*. 2019;20(1):13–21. <https://doi.org/10.1111/obr.12761>.
 89. Davis C, Levitan RD, Kaplan AS, Carter J, Reid C, Curtis C, et al. Reward sensitivity and the D2 dopamine receptor gene: a case-control study of binge eating disorder. *Prog Neuro-Psychopharmacol Biol Psychiatry*. 2008;32:620–8.
 90. Davis C, Levitan RD, Yilmaz Z, Kaplan AS, Carter JC, Kennedy JL. Binge eating disorder and the dopamine D2 receptor: genotypes and sub-phenotypes. *Prog Neuro-Psychopharmacol Biol Psychiatry*. 2012;38:328–35.
 91. Noble EP. D2 dopamine receptor gene in psychiatric and neurologic disorders and its phenotypes. *Am J Med Genet B Neuropsychiatr Genet*. 2003;116B:103–25.
 92. Jonsson EG, Nothen MM, Grunhage F, Farde L, Nakashima Y, Propping P, et al. Polymorphisms in the dopamine D2 receptor gene and their relationships to striatal dopamine receptor density of healthy volunteers. *Mol Psychiatry*. 1999;4:290–6.
 93. Shinohara M, Mizushima H, Hirano M, Shioe K, Nakazawa M, Hiejima Y, et al. Eating disorders with binge-eating behaviour are associated with the s allele of the 3'-UTR VNTR polymorphism of the dopamine transporter gene. *J Psychiatry Neurosci*. 2004;29:134–7.
 94. Kaye WH, Frank GK, Bailer UF, Henry SE, Meltzer CC, Price JC, et al. Serotonin alterations in anorexia and bulimia nervosa: new insights from imaging studies. *Physiol Behav*. 2005;85:73–81.
 95. Monteleone P, Zanardini R, Tortorella A, Gennarelli M, Castaldo E, Canestrelli B, et al. The 196G/a (val66met) polymorphism of the BDNF gene is significantly associated with binge eating behavior in women with bulimia nervosa or binge eating disorder. *Neurosci Lett*. 2006;406:133–7.
 96. Olszewski PK, Levine AS. Central opioids and consumption of sweet tastants: when reward outweighs homeostasis. *Physiol Behav*. 2007;91:506–12.
 97. Nakagawa T, Tsuchida A, Itakura Y, Nonomura T, Ono M, Hirota F, et al. Brain-derived neurotrophic factor regulates glucose metabolism by modulating energy balance in diabetic mice. *Diabetes*. 2000;49:436–44.
 98. Lilienfeld LR, Kaye WH, Greeno CG, Merikangas KR, Plotnicov K, Pollice C, et al. A controlled family study of anorexia nervosa and bulimia nervosa: psychiatric disorders in first-degree relatives and effects of proband comorbidity. *Arch Gen Psychiatry*. 1998;55:603–10.
 99. Strober M, Freeman R, Lampert C, Diamond J, Kaye W. Controlled family study of anorexia nervosa and bulimia nervosa: evidence of shared liability and transmission of partial syndromes. *Am J Psychiatry*. 2000;157:393–401.
 100. Monteleone P, Bifulco M, Di Filippo C, Gazzo P, Canestrelli B, Monteleone F, et al. Association of CNR1 and FAAH endocannabinoid gene polymorphisms with anorexia nervosa and bulimia nervosa: evidence for synergistic effects. *Genes Brain Behav*. 2009;8:728–32.
 101. Muller TD, Greene BH, Bellodi L, Cavallini MC, Cellini E, Di Bella D, et al. Fat mass and obesity-associated gene (FTO) in eating disorders: evidence for association of the rs9939609 obesity risk allele with bulimia nervosa and anorexia nervosa. *Obes Facts*. 2012;5:408–19.
 102. Thaler L, Groleau P, Badawi G, Sycz L, Zeramdini N, Too A, et al. Epistatic interactions implicating dopaminergic genes in bulimia nervosa (BN): relationships to eating- and personality-related psychopathology. *Prog Neuro-Psychopharmacol Biol Psychiatry*. 2012;39:120–8.
 103. Steiger H, Joober R, Israël M, Young SN, Ng Ying Kin NM, Gauvin L, Bruce KR, et al. The 5HTTLPR polymorphism, psychopathologic symptoms, and platelet [3H]-paroxetine binding in bulimic syndromes. *Int J Eat Disord*. 2005;37(1):57–60.
 104. Baker JH, Thornton LM, Bulik CM, Kendler KS, Lichtenstein P. Shared genetic effects between age at menarche and disordered eating. *J Adolesc Health*. 2012;51:491–6.
 105. Nilsson M, Naessén S, Dahlman I, Lindén Hirschberg A, Gustafsson JA, Dahlman-Wright K. Association of estrogen receptor beta gene polymorphisms with bulimic disease in women. *Mol Psychiatry*. 2004;9(1):28–34.
 106. Pagotto U, Marsicano G, Cota D, Lutz B, Pasquali R. The emerging role of the endocannabinoid system in endocrine regulation and energy balance. *Endocr Rev*. 2006;27:73–100.
 107. Steiger H, Richardson J, Joober R, Gauvin L, Israel M, Bruce KR, et al. The 5HTTLPR polymorphism, prior maltreatment and dramatic-erratic personality manifestations in women with bulimic syndromes. *J Psychiatry Neurosci*. 2007;32:354–62.
 108. Le Foll B, Gallo A, Le Strat Y, Lu L, Gorwood P. Genetics of dopamine receptors and drug addiction: a comprehensive review. *Behav Pharmacol*. 2009;20:1–17.
 109. Jimerson DC, Lesem MD, Kaye WH, Brewerton TD. Low serotonin and dopamine metabolite concentrations in cerebrospinal fluid from bulimic patients with frequent binge episodes. *Arch Gen Psychiatry*. 1992;49:132–8.
 110. Tauscher J, Pirker W, Willeit M, de Zwaan M, Bailer U, Neumeister A, et al. [123I] beta-CIT and single photon emission computed tomography reveal reduced brain serotonin transporter availability in bulimia nervosa. *Biol Psychiatry*. 2001;49:326–32.
 111. Kaplan AS, Levitan RD, Yilmaz Z, Davis C, Tharmalingam S, Kennedy JL. A DRD4/BDNF gene-gene interaction associated with maximum BMI in women with bulimia nervosa. *Int J Eat Disord*. 2008;41:22–8.
 112. Frayling TM, Timpson NJ, Weedon MN, Zeggini E, Freathy RM, Lindgren CM, Perry JR, et al. A common variant in the FTO gene

- is associated with body mass index and predisposes to childhood and adult obesity. *Science*. 2007;316(5826):889–94.
113. Scuteri A, Sanna S, Chen WM, Uda M, Albai G, Strait J, et al. Genome-wide association scan shows genetic variants in the FTO gene are associated with obesity-related traits. *PLoS Genet*. 2007;3:e115.
114. Dina C, Meyre D, Gallina S, Durand E, Komer A, Jacobson P, et al. Variation in FTO contributes to childhood obesity and severe adult obesity. *Nat Genet*. 2007;39:724–6.
115. Qasim A, Mayhew AJ, Ehtesham S, Alyass A, Volckmar AL, Herpertz S, Hinney A, et al. Gain-of-function variants in the melanocortin 4 receptor gene confer susceptibility to binge eating disorder in subjects with obesity: a systematic review and meta-analysis. *Obes Rev*. 2019;20(1):13–21. <https://doi.org/10.1111/obr.12761>.
116. Potoczna N, Branson R, Kral JG, Pic G, Steffen R, Ricklin T, Hoehe MR, et al. Gene variants and binge eating as predictors of comorbidity and outcome of treatment in severe obesity. *J Gastrointest Surg*. 2004;8(8):971–81.
117. Hirata T, Uemura T, Shinohara M, Hirano M. Association between Dopamine Transporter Gene (DAT1) Polymorphisms and Eating Disorders with Binge Eating Behavior. *Open Journal of Psychiatry*. 2017;7:329–343.
118. Nilsson M, Naessén S, Dahlman I, Lindén Hirschberg A, Gustafsson JA, Dahlman-Wright K. Association of estrogen receptor beta gene polymorphisms with bulimic disease in women. *Mol Psychiatry*. 2004 Jan;9(1):28–34.
119. Thaler L, Groleau P, Joober R, Bruce KR, Israel M, Badawi G, Sycz L, Steiger H. Epistatic interaction between 5HTTLPR and TPH2 polymorphisms predicts novelty seeking in women with bulimia nervosa spectrum disorders. *Psychiatry Res*. 2013;208(1):101–3. <https://doi.org/10.1016/j.psychres.2012.11.028>

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