

# Role of insulin resistance in endothelial dysfunction

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**Abstract** Insulin resistance is frequently associated with endothelial dysfunction and has been proposed to play a major role in cardiovascular diseases. Insulin exerts pro- and anti-atherogenic actions on the vasculature. The balance between nitric oxide (NO)-dependent vasodilator actions and endothelin-1- dependent vasoconstrictor actions of insulin is regulated by phosphatidylinositol 3-kinase-dependent (PI3K) - and mitogen-activated protein kinase (MAPK)-dependent signaling in vascular endothelium, respectively. During insulin-resistant conditions, pathway-specific impairment in PI3K-dependent signaling may cause imbalance between production of NO and secretion of endothelin-1 and lead to endothelial dysfunction. Insulin sensitizers that target pathway-selective impairment in insulin signaling are known to improve endothelial dysfunction. In this review, we discuss the cellular mechanisms in the endothelium underlying vascular actions of insulin, the role of insulin resistance in mediating endothelial dysfunction, and the effect of insulin sensitizers in restoring the balance in pro- and anti-atherogenic actions of insulin.

**Keywords** Nitric oxide · Insulin resistance · Endothelial dysfunction · Metabolic syndrome

## 1 Introduction

Insulin resistance plays a patho-physiological role in type 2 diabetes and is frequently present in obesity, hypertension, coronary artery disease, dyslipidemias, and metabolic syndrome [1, 2]. Global epidemic of obesity is driving the increased incidence and prevalence of insulin resistance and its cardiovascular complications [3]. Insulin regulates glucose homeostasis not only by promoting glucose disposal in skeletal muscle and adipose tissue and inhibiting gluconeogenesis in liver [2], but also by regulating nutrient delivery to target tissues by actions on microvasculature [4]. Insulin-induced nitric oxide (NO) production from vascular endothelium leads to increased blood flow that further enhances glucose uptake in skeletal muscle [5].

At the cellular level, balance between phosphatidylinositol 3-kinase- (PI3K)-dependent insulin-signaling pathways that regulate endothelial NO production and mitogen activated protein kinase (MAPK)-dependent insulin-signaling pathways regulating the secretion of the vasoconstrictor endothelin-1 (ET-1) determines the vascular response to insulin. Insulin resistance is typically defined as decreased sensitivity or responsiveness to metabolic actions of insulin such as insulin-mediated glucose disposal. However, diminished sensitivity to the vascular actions of insulin also plays an important role in the pathophysiology of insulin-resistant states [6, 7]. Indeed, endothelial insulin resistance is typically accompanied by reduced PI3K-NO pathway and an intact or heightened MAPK-ET-1 pathway. Insulin resistant states are associated with metabolic abnormalities that include glucotoxicity, lipotoxicity, and inflammation that also lead to endothelial dysfunction. Indeed, pathway-specific impairment in PI3K-dependent insulin signaling contributes

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to reciprocal relationships between insulin resistance and endothelial dysfunction that foster the clustering of metabolic and cardiovascular diseases in insulin-resistant states [8]. Herein, we discuss the role of insulin resistance in endothelial dysfunction and therapeutic interventions that may simultaneously improve both metabolic and endothelial function in insulin-resistant conditions.

**Endothelial dysfunction** The term “endothelial dysfunction” refers to a maladapted endothelial phenotype characterized by reduced NO bioavailability, increased oxidative stress, elevated expression of pro-inflammatory and pro-thrombotic factors, and abnormal vasoreactivity [9]. Healthy endothelium plays a central role in homeostatic functions by actively secreting various molecules that act in a paracrine, autocrine, and endocrine fashion. These molecules affect vascular tone, endothelial and vascular smooth muscle (VSMC) growth and proliferation, endothelial-leukocyte interactions, platelet adhesion, coagulation, inflammation, and permeability. Vascular tone is modulated by endothelium-derived vasoactive substances that include vasodilators (e.g., NO, prostaglandins (PGI<sub>2</sub>), endothelium-derived hyperpolarization factor (EDHF), epoxyeicosatrienoic acids (EETs)) and vasoconstrictors (e.g., Angiotensin II (Ang II), ET-1, prostanoids, isoprostanes). In addition to affecting vascular tone, key vasodilators such as NO and prostacyclin are anti-proliferative and anti-inflammatory, while vasoconstrictors such as ET-1 and Ang II are mitogenic and pro-inflammatory. Both in the macro- and microvasculature (arterioles, capillaries and venules that are less than 150 μm in diameter), the balance in actions of vasodilators and vasoconstrictors determine vascular tone and endothelial function.

NO, an important determinant of endothelial function, is produced in vascular endothelium by activation of endothelial NO synthase (eNOS) [10]. Vasodilatory actions of NO are primarily mediated via reductions in VSMC intracellular calcium Ca<sup>2+</sup> concentrations secondary to NO-mediated guanylate cyclase activation and cGMP formation. Elevations in endothelial cytoplasmic Ca<sup>2+</sup> secondary to the activation of serpentine G protein-coupled receptors (e.g. acetylcholine receptor) promote binding of calmodulin to and subsequent activation of eNOS. In addition, phosphorylation of eNOS at Ser<sup>1177</sup> by serine kinases including Akt, AMPK, and PKA also stimulate production of NO in a Ca<sup>2+</sup>-independent manner. Availability of L-arginine (substrate for eNOS) and enzymatic cofactors (NADPH, flavin adenine dinucleotide [FAD], flavin mononucleotide [FMN], and tetrahydrobiopterin [BH<sub>4</sub>]) also play a role in regulating NO production by eNOS [11]. In addition to decreasing vascular tone, NO decreases expression of vascular cell adhesion molecules, attenuates production of pro-inflammatory cytokines, decreases leukocyte recruitment, inhibits VSMC proliferation, opposes

apoptosis, attenuates platelet aggregation, and reduces monocyte adhesion to the vascular wall [12]. Thus, reductions in NO bioavailability favor a dysfunctional vascular phenotype. Due to the systemic nature of the dysfunction, the manifestations of endothelial dysfunction vary depending on the nature of the vascular bed.

A dysfunctional endothelium is a harbinger of atherosclerosis and can also contribute to cardiovascular events. Importantly, endothelial dysfunction is independently associated with and predicts cardiac death, myocardial infarction, and stroke [13]. Interestingly, endothelial dysfunction is also linked to insulin-resistant states including diabetes, obesity, and the metabolic syndrome [14]. This increases the susceptibility of patients with these metabolic diseases to cardiovascular complications including accelerated atherosclerosis, coronary heart disease, and hypertension.

**Assessment of endothelial function** Studies designed to evaluate endothelial function in humans often assess NO-dependent vasodilation. Measurement of the changes in coronary artery diameter and blood flow in response to intra-coronary infusion of acetylcholine is regarded as the “gold standard” technique. Changes in limb blood flow (assessed by plethysmography) or conduit artery diameter (assessed by ultrasound) in response to intra-arterial infusion of agents that stimulate endothelium-dependent production of NO such as acetylcholine are used primarily in research settings to evaluate endothelial function [9]. Another less-invasive method involves shear stress-induced flow-mediated dilatation (FMD) of the brachial artery. High resolution doppler ultrasonography is used to measure changes in arterial diameter and blood flow in the brachial artery in response to shear stress induced by inflating and deflating a blood pressure cuff (reactive hyperemia). Using this same principle, peripheral endothelial function is measured by finger plethysmography. Increases in digital pulse amplitude after reactive hyperemia are recorded by peripheral arterial tonometry (EndoPAT) [15]. Endothelial function measured using this technique correlates well with coronary endothelial-mediated vasodilatory response to acetylcholine [16]. Currently, limited resolution of current imaging techniques precludes robust assessment of microvascular endothelial function in humans. Nevertheless, positron emission tomography (PET) [17], contrast-imaging with microbubbles [18], and laser-doppler perfusion [19] monitoring have been used to monitor changes in capillary blood flow in various vascular beds in response to agents/interventions known to augment endothelial-mediated blood flow.

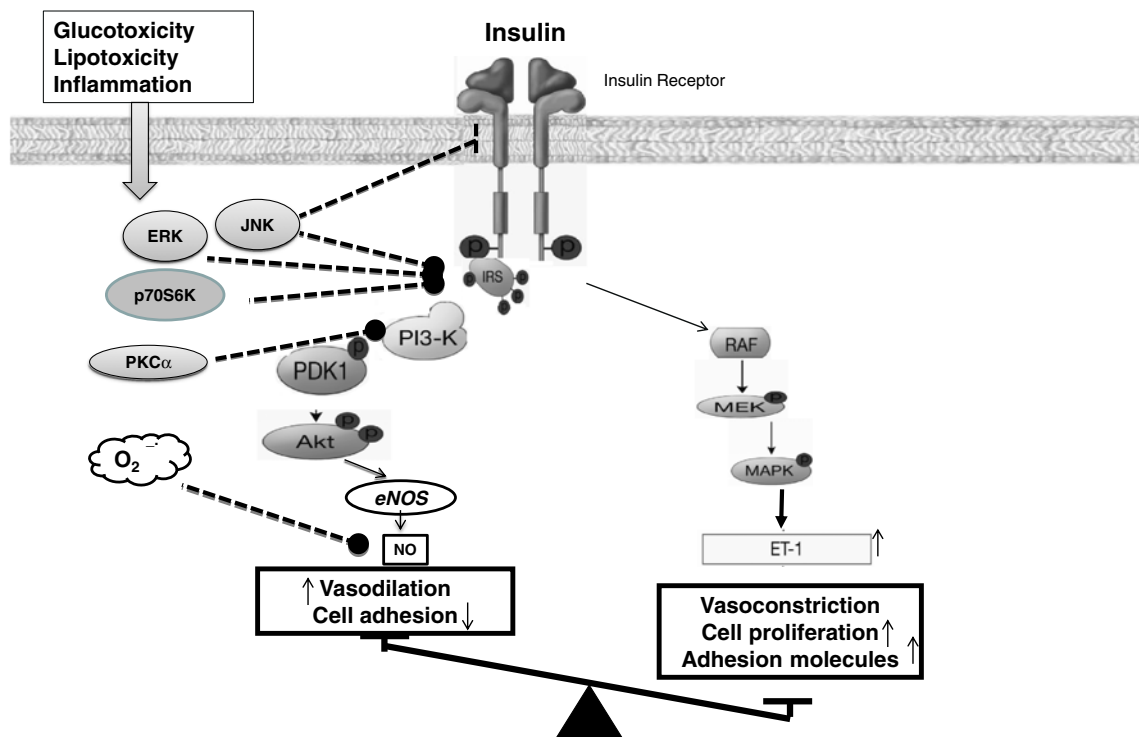
Elevated circulating plasma concentrations of biomarkers for inflammation, hemostasis, and oxidative stress are also used as indicators that accompany and promote endothelial dysfunction [9]. Among them circulating levels of P- and E-selectin, soluble inter-cellular adhesion molecules (sICAM-1), soluble

vascular cell adhesion molecule (sVCAM), oxidized low-density lipoprotein (oxLDL), plasminogen activator inhibitor-1 (PAI-1), and asymmetrical dimethylarginine (ADMA) have been used as a marker of endothelial dysfunction.

**Selective resistance to insulin actions in the endothelium** Insulin resistant states are characterized by attenuation of some cell-specific insulin responses, while other actions of insulin not only persist but may be exaggerated. This selective insulin resistance is observed in the liver where insulin-mediated suppression of glucose production is impaired but not lipogenesis [20]. Similarly, in the endothelium, insulin-stimulated NO, but not ET-1 production is impaired [21]. This selective insulin resistance is explained by the pathway-specific impairment in insulin signaling pathways (Fig. 1). Insulin-signaling pathways regulating endothelial NO and ET-1 production have been elucidated [4, 22]. Detailed description of these pathways is described elsewhere in this issue but briefly, insulin binding to its receptor results in phosphorylation of IRS-1 which then binds and activates PI3K. PI-3,4,5-triphosphate (PIP<sub>3</sub>), a product of PI3K activity, stimulates phosphorylation and activation of PDK-1 that in turn phosphorylates and activates Akt.

Akt directly phosphorylates eNOS at Ser<sup>1177</sup> resulting in increased eNOS activity and subsequent NO production. Insulin stimulates ET-1 production using MAPK-dependent (but not PI3K-dependent) signaling pathways [23]. Insulin stimulates increased expression of PAI-1, VCAM-1 and E-selectin on endothelium using MAPK-dependent pathway [24, 25]. Inhibition of PI3K or Akt increases insulin-induced PAI-1 and expression of adhesion molecules [24]. These findings suggest that insulin-stimulated PI3K/Akt pathways oppose atherothrombotic factors in endothelium by multiple mechanisms including production of beneficial molecules such as NO and inhibition of pathogenic molecules including PAI-1, ICAM-1, VCAM-1, and E-selectin. In the setting of selective impairment of PI3K activation and compensatory hyperinsulinemia, the pro-hypertensive, atherogenic, thrombogenic, and pro-coagulant actions of insulin dominates to lead to endothelial dysfunction.

**Selective insulin resistance in animal models** Selective insulin resistance has been observed in various rodent models of global insulin resistance [23, 26]. Using genetic approaches, various groups have examined the role of endothelium-specific insulin resistance in vascular dysfunction [27–30].



**Fig. 1** Pathway-specific impairment of insulin signaling pathway and endothelial dysfunction. PI 3-kinase branch of insulin signaling regulates NO production and vasodilation in vascular endothelium. MAP-kinase branch of insulin signaling controls secretion of endothelin-1 (ET-1) and adhesion molecule expression in vascular endothelium. Glucotoxicity, lipotoxicity, and various cytokines activate signaling molecules that

inhibit PI3K/Akt signaling. eNOS, endothelial nitric oxide synthase; IRS, insulin receptor substrate; MEK, MAPK kinase; PDK, phosphoinositide-dependent protein kinase; PKC, protein kinase C; ERK, extracellular signal-regulated kinase; JNK, C-Jun N-terminal kinase; p70S6K, p70 ribosomal S6 kinase; AP-1, activator protein-1; NO, nitric oxide; and ET-1, endothelin-1

Mice lacking insulin receptors specifically in vascular endothelium (VENIRKO) have normal glucose metabolism and blood pressure, but reduced expression of eNOS and ET-1 in endothelium [27]. However, when challenged with high-salt diet, VENIRKO mice develop insulin resistance and elevated blood pressure [27]. Similarly, mice expressing endothelium-specific expression of a dominant negative mutant human insulin receptor develop endothelial dysfunction. This was evident by impaired NO release to insulin, acetylcholine, or a calcium ionophore. Furthermore, there was an increase in the expression of NADPH oxidase that leads to oxidative stress and reduced NO bioavailability [28]. In the vasculature of heterozygous global insulin receptor knockout mice (IRKO) with metabolic insulin resistance, insulin-stimulated phosphorylation and activation of eNOS is impaired resulting in reduced basal and insulin-stimulated NO release [29]. Likewise, IRS-1 (IRS-1<sup>-/-</sup>) and IRS-2 (IRS-2<sup>-/-</sup>) deficient mice not only exhibit resistance to the metabolic actions of insulin, but also demonstrate diminished endothelial NO activity [30]. These findings suggest that complex interactions between insulin action, eNOS, ET-1, and oxidative stress affect the metabolic and vascular phenotype in these mice. In addition, in these models, the proximal nodes in insulin signaling (IR and IRS-1) are affected thereby affecting both the PI3K and MAPK pathways. Perhaps this explains the relatively mild endothelial dysfunction observed in these rodent models.

In contrast to previously mentioned rodent models, endothelial dysfunction is pronounced in rodents that exhibit selective-impairment in insulin resistance. Genetic ablation of Akt1 in high fat-fed ApoE<sup>-/-</sup> mice leads to increased atherogenesis and endothelial dysfunction [31]. Obese Zucker rats demonstrate pathway selective insulin resistance in PI3K-dependent signaling (with intact MAPK signaling) in the vasculature [26]. This results in impaired NO-mediated vasodilation and augmented ET-1-mediated vasoconstriction in response to insulin. In spontaneously hypertensive rats (SHRs) NO-dependent vasodilator response to insulin is significantly impaired. However, inhibiting MAPK-dependent pathways unmasks vasodilator actions of insulin [23]. Treatment of micro-vessels with ET-1 receptor antagonists, BQ788 and BQ123 also improves insulin-mediated vasodilation in these rats. These findings suggest that selective impairment in PI3K- pathways leads to decreased endothelial production of NO while increased insulin signaling through MAPK-dependent pathways leads to elevated secretion of ET-1. Thus, partial and selective defects in insulin signaling or NO activity are sufficient to cause endothelial dysfunction during nutritional stress, inflammation, and dysmetabolic states.

*Insights about selective insulin resistance in human studies* Studies in patients with extremely rare mutations in either insulin receptor or Akt2 genes support the concept

of selective insulin resistance in the liver [20]. Patients with IR mutations manifest a loss of suppression of hepatic glucose output without enhanced hepatic lipogenesis [20]. However, increased *de novo* lipogenesis and hepatic gluconeogenesis are observed in patients with Akt2 mutations. These results suggest that selective insulin resistance in the PI3K-Akt pathway uncouples gluconeogenesis and lipogenesis. Endothelial function has not been studied in these patients. Nevertheless, findings from other studies support the notion of pathway-selective insulin resistance in the human endothelium. Subjects with IRS-1 polymorphism (glycine to arginine at codon 972), are not only insulin-resistant, but also have endothelial dysfunction [32]. In endothelial cells from subjects carrying the G972R-IRS-1 variant, insulin-mediated PI3-K/Akt/eNOS activation is diminished [33]. Similarly, activation of Akt is impaired in internal mammary arteries obtained from patients with diabetes when compared with non-diabetics [34]. In addition, the absolute levels of phospho-eNOS (Ser1177) are also decreased in vascular tissue from diabetic patients [34]. These findings suggest that impaired endothelial insulin signaling and reduced NO activity contributes to endothelial dysfunction in insulin resistant states.

As previously discussed, pathway-specific insulin resistance results in enhanced effects of insulin to stimulate ET-1 production and promote increased vasoconstrictor tone. Coronary vessels with vulnerable and obstructive atherosclerosis are characterized by an increase in ET-1 activity and endothelial dysfunction [35]. The parallel increase in ET-1 activity and diminished NO bioactivity contributes to abnormal vascular function. Human studies in overweight [36], obese [37], hypertensive [38, 39] and diabetic [37, 40] subjects support this notion. Vascular ET<sub>A</sub>/ET<sub>B</sub> receptor blockade in the forearm significantly increases endothelium-dependent vasodilatation in overweight, insulin-resistant subjects but not in lean, healthy controls [36, 38]. Similarly, selective ET<sub>A</sub> receptor blockade in the forearm significantly increases forearm blood flow in patients with type 2 diabetes [40]. Hyperinsulinemia stimulates ET-1 secretion [41] and accentuated ET-1 activity may cause insulin resistance [42]. Thus, human studies support the idea that increased endogenous activity of ET-1 and reduced is a feature of endothelial dysfunction in insulin resistance, obesity, hypertension, and diabetes mellitus. This phenotype is a manifestation of pathway-selective insulin resistance in the endothelium.

*Relationship between insulin resistance and endothelial dysfunction: a meta-analysis* Many cross-sectional studies have examined the relationship between insulin resistance/sensitivity and endothelial function [7, 43–56]. In these studies, endothelial function was evaluated by high resolution brachial artery ultrasound, venous occlusion plethysmography, and laser doppler imaging techniques. Insulin

sensitivity was assessed by euglycemic hyperinsulinemic clamps and surrogate indices (HOMA-IR, frequently sampled intravenous glucose tolerance test). In an effort to evaluate the contribution of insulin resistance to endothelial dysfunction, we pooled univariate correlation coefficients from these studies using Schmidt-Hunter model. Figure 2 presents the pooled correlation coefficient for 12 studies ( $N=3,190$ ) was 0.14 ( $p<0.001$ , 95 % CI:  $-0.09$ – $-0.20$ ). These results suggest that the correlation between insulin resistance and endothelial function is rather weak. The test for heterogeneity in the included studies was significant ( $p<0.05$ ). Given the heterogeneity of the studies, the different techniques to assess insulin resistance and endothelial function, and the limited number of studies reviewed, it is possible that we are underestimating the strength of this relationship. Nevertheless, in many studies, upon multivariate analysis that included adjustment of other potential modulators of endothelial function, insulin resistance was no longer a significant predictor of endothelial dysfunction [46, 48–50].

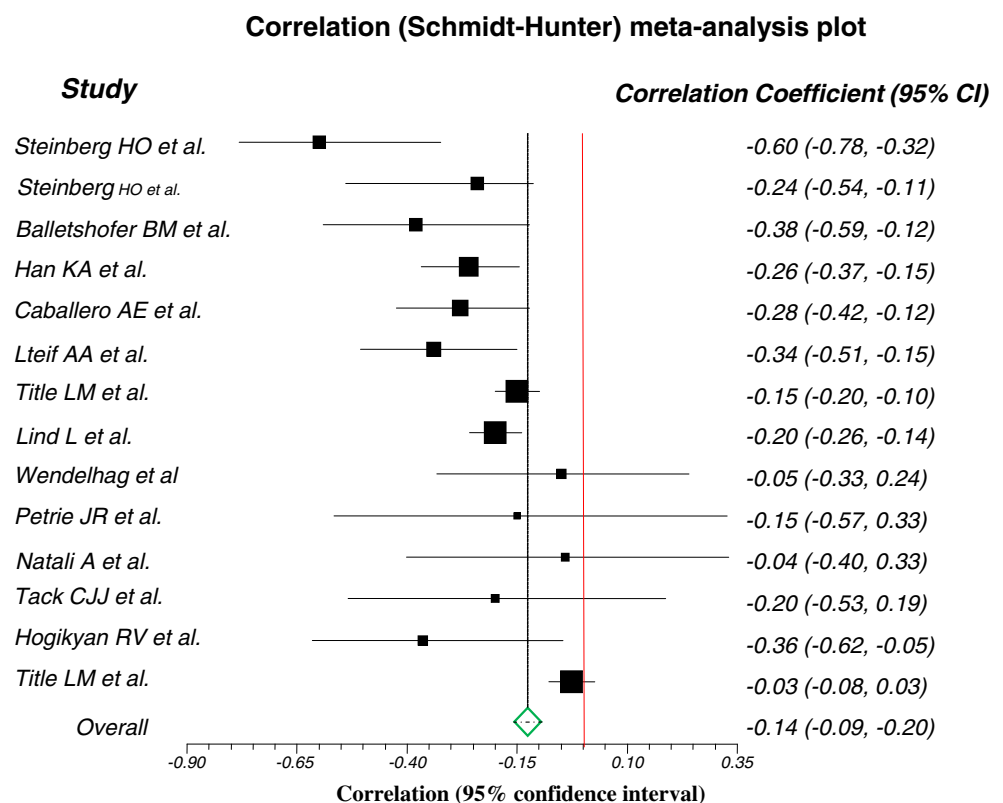
These findings together suggest that insulin resistance may only partly explain the impaired endothelial function. It is also possible that other manifestations of insulin resistance such as hyperglycemia, dyslipidemia, inflammation, and obesity may be intermediary mediators that act in concert with insulin resistance to mediate endothelial dysfunction. The role of glucotoxicity, lipotoxicity, and inflammation in endothelial dysfunction is reviewed in detail elsewhere in this issue.

However, hyperglycemia, hyperlipidemia, and various cytokines are known to selectively impair PI3K/Akt/eNOS pathway, increase oxidative stress, and enhance the release of ET-1 from the endothelium (Fig. 1) [4].

*Insulin sensitizers and endothelial dysfunction* Calorie restriction and physical exercise, interventions known to improve insulin sensitivity, also improves endothelial dysfunction [57–69]. Calorie restriction improves NO-dependent vasodilation and simultaneously reduces circulating ET-1 levels in insulin resistant individuals [70–72]. Regular exercise increases vascular eNOS protein expression and activity via PI3K/Akt-dependent phosphorylation in humans [73]. Thus the beneficial effects of these life style modifications may involve enhanced insulin signaling, increased eNOS activity, and reductions in oxidative and inflammatory stress that leads to the rebalancing the vasoconstrictor and vasodilator actions of insulin

Metformin, a frequently used insulin sensitizer stimulates adenylyl monophosphate kinase (AMP-kinase) that in turn phosphorylates and activates eNOS in endothelial cells in culture [74]. Consequently, administration of metformin not only improves endothelium-dependent vasodilation but simultaneously decreases circulating ET-1 levels in insulin-resistant individuals [75]. Similarly, administration of another insulin sensitizer, rosiglitazone also improves FMD and acetylcholine-mediated vasodilation in individuals with the metabolic syndrome [76]. It is unclear whether improvement in insulin sensitivity per se was the proximal mediator of

**Fig. 2** Correlation Meta-analysis. Univariate correlation coefficients between insulin sensitivity and endothelial function and corresponding 95 % confidence intervals



improved endothelial function in these studies. However, metformin has been shown to significantly reduce CV events in patients with insulin resistance [77]. Considering that prevention of insulin resistance is predicted to reduce myocardial infarctions by 42 % [78], therapies aimed at improving insulin sensitivity may offer a more rational choice in treating endothelial dysfunction in dysmetabolic states.

**Summary** Endothelium specific selective impairment of PI3K-dependent insulin signaling pathways favors the phenotype of endothelial dysfunction and insulin resistance. Targeting this selective impairment has the potential to simultaneously ameliorate endothelial dysfunction and insulin resistance. Future studies in patients with genetic mutations in insulin receptor, Akt, or PTEN, may offer additional insights aimed at delineating the role of insulin resistance in endothelial dysfunction. In the meantime however, insulin sensitization therapy over insulin provision should be considered in insulin-resistant subjects to improve endothelial health and reduce cardiovascular disease.

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