#### **ORIGINAL ARTICLE**



# **Static magnetic feld treatment enhanced photosynthetic performance in soybean under supplemental ultraviolet‑B radiation**

**Sunita Kataria1 · Meeta Jain<sup>1</sup> · Anshu Rastogi2,3 · Marian Brestic4,5**

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#### **Abstract**

The study was performed to analyze the impact of seed pretreatment by static magnetic feld (SMF) of 200 mT for 1 h on photosynthetic performance of soybean (*Glycine max*) seedlings under ambient (aUV-B) and supplemental ultraviolet-B  $_{a+\text{UV-B}}$ ) stress. Ambient and supplemental UV-B were found to decrease the plant growth, chlorophyll concentration, PSII efficiency, selected JIP-test parameters such as  $F_v/F_m$ ,  $\varphi$ Eo,  $\Delta V(I-P)$ ,  $PI_{\text{ABS}}$ ,  $PI_{\text{total}}$ , and rate of photosynthesis in the leaves of soybean seedlings emerged from untreated (UT) seeds.  $_{a}$ UV-B and  $_{a+s}$ UV-B were observed to increase the synthesis of UV-B-absorbing substances (UAS), reactive oxygen species (ROS) like superoxide radical  $(O_2^-)$  and hydrogen peroxide  $(H<sub>2</sub>O<sub>2</sub>)$ , antioxidants like ascorbic acid and  $\alpha$ -tocopherol and decrease the nitrate reductase (NR) activity; subsequently, it results in a decreased rate of photosynthesis, biomass accumulation, and yield. However, our results provided evidence that SMF pretreatment increased the tolerance of soybean seedlings to UV-B radiation by increased NO content and NR activity; higher efficiency of PSII, higher values of  $\varphi$ Eo,  $\Delta V(I-P)$ , PI<sub>ABS</sub>, and PI<sub>total</sub>, decreased intercellular CO<sub>2</sub> concentration, lower amount of UAS, ROS, and antioxidants that consequently improve the yield of soybean plants under <sub>a</sub>UV-B as well as  $_{a+s}$ UV-B stress. Thus, our results suggested that SMF pretreatment mitigates the adverse effects of UV-B stress by the enhancement in photosynthetic performance along with higher NO content which may be able to protect the plants from the deleterious efects of oxidative stress caused by UV-B irradiation.

Keywords Growth · Photosynthesis · PSII efficiency · Chl fluorescence · Nitric oxide · UV-B

#### **Abbreviations**



 $\boxtimes$  Sunita Kataria sunita\_kataria@yahoo.com

- <sup>1</sup> School of Biochemistry, Devi Ahilya University, Khandwa Road, Indore, M.P. 452001, India
- Laboratory of Bioclimatology, Department of Ecology and Environmental Protection, Poznan University of Life Sciences, Piątkowska 94, 60-649 Poznan, Poland
- <sup>3</sup> Faculty of Geo-Information Science and Earth Observation (ITC), University of Twente, 7500 AE Enschede, The Netherlands
- <sup>4</sup> Department of Plant Physiology, Slovak University of Agriculture, A. Hlinku 2, 94976 Nitra, Slovak Republic
- <sup>5</sup> Department of Botany and Plant Physiology, Faculty of Agrobiology, Food and Natural Resources, Czech University of Life Sciences Prague, 16500 Prague, Czech Republic



## **Introduction**

Plants growing in natural environments are exposed to different environmental signals that regulate responses at the plant level. Of those environmental signals, sunlight is of utmost importance as a source of energy for plants. In sunlight, wavelengths in the UV region are divided into UV-A (315–400 nm), UV-B (280–315 nm), and UV-C (100–280 nm) radiations (Björn [2015\)](#page-12-0). However, wavelengths below 290 nm are absorbed by the ozone layer and atmospheric oxygen, therefore are not detectable at the Earth's surface (Rowland [2006\)](#page-14-0). As a consequence of human activities worldwide, the chemical composition of the atmosphere is changing. A reduction in the stratospheric ozone layer is dangerous for most of the Earth's population as it leads to a higher level of UV-B radiation on Earth's surface (Bais et al. [2019](#page-12-1); Bornman et al. [2019](#page-12-2)). Due to the high sustainability of chloro-fuorocarbons (an industrial gas), the ozone layer is under the threat, and even if all the nations execute the Montreal Protocol, it is not possible to reduce the UV-B level reaching on the Earth surface to its pre-industrialization era by 2050 (Mohammed and Tarpley [2010](#page-13-0)). Though in the solar spectrum UV-B radiations consist of only a small part, it's high energy can degrade vital molecules like lipids, proteins, nucleic acids, and phytohormones (Jansen et al. [1998](#page-13-1); Singh et al. [2012](#page-14-1); Kataria et al. [2014a](#page-13-2), [b](#page-13-3); Vanhaelewyn et al. [2016](#page-15-0)). The enhanced UV-B radiation drastically hampered the physiological, morphological, and biochemical development of numerous plant species (Kakani et al. [2003a,](#page-13-4) [b](#page-13-5); Caldwell et al. [2007](#page-12-3); Kataria et al. [2014a](#page-13-2), [b\)](#page-13-3) and eventually decrease the crop yield (Liu et al. [2013](#page-13-6); Kataria et al. [2014a](#page-13-2), [b\)](#page-13-3). The morphological alterations by UV-B, such as decrease in plant height, leaf area and leaf length, thicker leaves, reduced internode length, curling of cotyledons/leaves, bronzing/glazing of leaves, chlorosis and necrotic spots in leaves, delayed seedling emergence and fowering have been reported in numerous crop plants (Caldwell et al. [1995](#page-12-4), [2007;](#page-12-3) Robson et al. [2015](#page-14-2); Suchar and Robberecht [2015\)](#page-15-1).

One of the most sensitive physiological processes in plants afected by UV-B exposure is photosynthesis which is closely related to biomass accumulation and crop yield. A reduction in plant growth parameters due to UV-B exposure was observed in several plant species (Kakani et al. [2003a,](#page-13-4) [b;](#page-13-5) Kataria et al. [2013\)](#page-13-7) which eventually reduced the crop productivity (Searles et al. [2001;](#page-14-3) Zuk-Golaszewska et al. [2003](#page-15-2)). UV-B radiations caused a reduction in photosynthetic activity due to the damage to carotenoids and chlorophyll, destruction of PSII proteins, decreased activity of Rubisco and sedoheptulose 1,7-biphosphatase (Allen et al. [1998](#page-12-5); Kataria et al. [2013](#page-13-7)), and damage to

PSII efficiency (Nogues and Baker [1995;](#page-14-4) Allen et al. [1998](#page-12-5); Yu et al. [2013\)](#page-15-3). In the last few decades, chlorophyll fluorescence observation has established itself as a reliable technique for the detection of photosynthetic processes (Kalaji et al. [2014](#page-13-8), [2018\)](#page-13-9). The 'JIP' test is a chlorophyll-*a* fuorescence-based method used to evaluate the status of photosynthetic apparatus under diferent abiotic stresses (Rastogi et al. [2019a](#page-14-5), [b,](#page-14-6) [2020;](#page-14-7) Akhter et al. [2021](#page-12-6)).

Due to their survival nature, plants evolved with time to reduce UV-induced damages. Some of the protection mechanisms include higher production and accumulation of phenolic compounds which shield diferent organelles, whereas DNA photolyase protects damages to DNA (Jenkins [2009\)](#page-13-10). Exposure to UV leads to the generation of reactive oxygen species (ROS) like superoxide anion radical  $(O_2^-)$ , hydrogen peroxide  $(H_2O_2)$ , and hydroxyl radical (HO−) (Rastogi and Pospisil [2013](#page-14-8)). Plants have developed several enzymatic and non-enzymatic mechanisms for the purpose to protect themselves from diferent ROS molecules (Jain et al. [2004](#page-13-11); Hasanuzzaman et al. [2012;](#page-13-12) Rastogi et al. [2014\)](#page-14-9). The non-enzymatic molecules include ascorbic acid (AsA), glutathione, α-tocopherol, and carotenoids (Munné-Bosch and Alegre [2002;](#page-14-10) Jain et al. [2004\)](#page-13-11), whereas enzymatic scavenger include Halliwell/Asada pathway enzymes, superoxide dismutase, and catalase (Jain et al. [2004\)](#page-13-11). A number of studies have suggested the participation of nitric oxide (NO) in response to UV-B radiation (An et al. [2005;](#page-12-7) Zhang et al. [2011\)](#page-15-4).

Considering the increasing infuence of UV-B radiation, it is required to look into diferent techniques for the purpose to improve crop yield efficiently. Among different available techniques, seed priming is one of the most commonly used processes for enhancing diferent stress tolerance (Sytar et al. [2019](#page-15-5); Prajapati et al. [2020;](#page-14-11) Shah et al. [2020\)](#page-14-12). The SMF priming is one of the techniques which is less explored for its interaction with UV-B radiation. Some of the earlier studies have indicated that SMF pretreatment alleviates the response of diferent abiotic stress factors on number of plant speciecs (Anand et al. [2012](#page-12-8); Thomas et al. [2013;](#page-15-6) Kataria et al. [2017,](#page-13-13) [2019](#page-13-14), [2020;](#page-13-15) Baghel et al. [2018](#page-12-9)).

The impact of ambient and enhanced UV-B radiation on soybean yield has been previously studied by several researchers (Liu et al. [2013;](#page-13-6) Baroniya et al. [2011,](#page-12-10) [2014](#page-12-11)). Some of the studies have indicated that the pretreatment of seeds by SMF enhances productivity under ambient UV-B stress conditions (Kataria et al. [2017](#page-13-13), [2020\)](#page-13-15). However, the mitigation of the adverse efects of enhanced or supplemental UV-B radiation on growth, the efficiency of PSII, JIP-test parameters, photosynthesis, and yield of soybean through magnetopriming have not been investigated yet. Therefore, the present study aims to evaluate the impact of SMF pretreatment on photosynthetic performance and yield of soybean under enhanced UV-B stress.

# **Materials and methods**

Soybean (*Glycine max* (L.) variety JS-335) seeds were obtained from the ICAR-Indian Institute of Soybean Research in Indore, India. The experiments were conducted in October to January'2018 under ambient conditions at the University campus (latitude 22°43′N) in Indore, India.

#### **Magnetic feld generation and magnetic treatment**

The magnetic feld was generated by a fabricated electromagnetic feld generator ("AETec" Academy of Embedded Technology, Delhi, India) as described in Kataria et al. [\(2020](#page-13-15)). The SMF of 200 mT was generated and provided to soybean seeds for an hour in a sample holder (transparent plastic) of capacity 42 cm<sup>3</sup> (2.7  $L \times 2.6 B \times 7.3 H$ ) at  $25 \pm 5$  °C. The untreated (UT) seeds were kept far away from the infuence of the electromagnetic feld generator. The local geomagnetic field was observed to be  $< 10$  mT.

#### **Supplementary UV‑B treatments**

The SMF-treated (MT) and untreated (UT) seeds were sown in nursery bags of  $34 \times 34$  cm, filled with a 5 kg mixture of homogeneous black soil, sand, and farm-yard manure (2:2:1 by volume). Before sowing, the recommended fungicides Bevistin (Ankur Agro company, Etawah, India) and Diathane M-45 (Sagan Agro Industries, Meerut, India) (2 g  $kg^{-1}$  seeds) were applied to all the seeds. These seeds were also properly mixed with *Rhizobium* culture (National Fertilizer limited, New Delhi, India) at 3 g kg<sup>-1</sup> seeds. Six seeds of similar size were packed in plastic nursery bags and after germination three plants were maintained in each bag. The black soil used was affluent in lime, iron, alumina, and magnesia. The average temperature ranged from 27 to 30 °C, relative humidity ranged from 55 to 75% during the experimental period.

From the time of germination, soybean seedlings emerged from UT and MT seeds were exposed to ambient  $_{a}$ UV-B  $(0.136 \text{ mWcm}^{-2} \text{ s}^{-1})$  and enhanced UV-B (<sub>a+s</sub>UV-B, 0.335) mWcm−2 s −1) radiation *via* UV-B lamps (a UV-B fuorescent tube,TL40W/12, Philips, Eindhoven, The Netherlands) which exhibited its emission > 280 nm to a maximum at 312 nm and ftted on steel frames at a distance of 45 cm directly above the plant canopy of nursery bags  $(34 \times 34 \text{ cm})$ that was maintained constant throughout the experimental period.

The experimental site was organized in randomised way and divided into three blocks containing three biological replications,  $n = 3$ . This split-plot design allowed us to test the efects of ambient UV-B and for enhanced UV-B (ambient + supplemental,  $_{a+s}$ UV-B; 280–315 nm) radiation on soybean plants.The intensity of UV-B radiation was measured by a radiometer, solar light Co. Inc. (PMA 2100), Glenside, PA, USA.

# **Measurement of growth, chlorophyll, PSII efficiency, and gas exchange parameters**

The sampling was done from 45-day-old soybean plants for the growth and biochemical parameters.The plant height and leaf area were measured. A portable laser leaf area meter, CI-202 (CID Inc., Camas, WA USA), was used for the measurement of the area of third trifoliate leaves. The specifc leaf weight was measured according to Hunt ([1982](#page-13-16)). The chlorophyll content was measured via the dimethyl sulfoxide (DMSO) method (Hiscox and Israelstam [1979\)](#page-13-17). The total chlorophyll concentration was calculated by equation of Wellburn's (Wellburn and Lichtenthaler [1984](#page-15-7)) and expressed as mg g−1 leaf fresh weight. For chlorophyll *a* (Chl*a*) fuorescence analysis, the handy PEA fuorimeter (Plant Efficiency Analyzer, Hansatech Instruments, Norfolk, UK) was used with the standard protocol of 30 min dark adaptation (Strasser et al. [2000;](#page-15-8) Kalaji et al. [2016](#page-13-18), [2018](#page-13-9)). The transients were induced by red light (peak at 650 nm) of 600 Wm<sup>2</sup> (3200 µE m<sup>-2</sup> s<sup>-1</sup>) supply through a range of six light-emitting diodes, fxed on the leaf surface in the clips on a spot of 4 mm diameter to provide homogenous illumination over the exposed area of the sample. Data were recorded for 1 s with 12-bit resolution; the data acquisition was done at every 10 µs for the frst 2 ms and every 1 ms thereafter (Strasser et al. [2000](#page-15-8)). The JIP-test parameters such as  $F_v/F_m$ , the maximum quantum yield (efficiency) of PSII photochemistry, *φ*Eo, the quantum yield of electron transport, Δ*V*(*I*–*P* phase), the amplitude of the relative contribution of the *I*-to-*P* rise to the OJIP transient,  $PI_{ABS}$ , performance index at absorption basis, and  $PI_{total}$  total performance index were calculated according to the equations reviewed by Bussotti et al. [\(2020\)](#page-12-12) and Banks ([2017\)](#page-12-13). The Biolyzer HP3 software (Bioenergetics Laboratory) was used for the calculation of the photosynthetic parameters.

The LI-6200 photosynthetic system (LICOR Inc., USA), was used to measure the rate of photosynthesis (*Pn*  $\mu$ mol CO<sub>2</sub> m<sup>-2</sup> s<sup>-1</sup>), stomatal conductance (gs, mmol H<sub>2</sub>O  $m^{-2} s^{-1}$ ), and internal CO<sub>2</sub> concentration (µmol CO<sub>2</sub> mol<sup>-1</sup>) in third trifoliate leaves of 45-day-old soybean plants. The measurement was conducted around noon according to the protocol as described previously by Fatima et al. [\(2020](#page-12-14)).

#### **Biochemical analyses**

All the biochemical analyses were made in third trifoliate leaves of 45-day-old soybean plants from untreated and SMF-treated seeds grown under  $_{a}$ UV-B as well as  $_{a+s}$ UV-B stress.

# **UV‑B absorbing substances (UAS)**

UAS accumulation in leaves of soybean was determined spectrophotometrically (Shimadzu-UV 1601) from acidifed methanol extract by the method of Mazza et al. ([1999](#page-13-19)) from the leaf disk of 0.50 cm diameter and expressed as units mg<sup>-1</sup> fresh weight of leaves.

# Hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>)

 $H<sub>2</sub>O<sub>2</sub>$  was estimated by the formation of titanium-hydroperoxide complex (Mukherjee and Choudhuri [1983](#page-14-13)) from500 mg of soybean leaves and expressed as  $\mu$ mol  $H_2O_2$  g<sup>-1</sup> fresh weight of leaves.

# **Superoxide anion radical (O<sub>2</sub><sup>−</sup>)**

The standard nitroblue tetrazolium chloride (NBT) reduction method was used to quantify  $O_2$ <sup>-</sup> in soybean leaves (100 mg) following the method of Chaitanya and Naithani ([1994\)](#page-12-15) and expressed as  $\mu$ mol  $g^{-1}$  fresh weight of leaves.

#### **Estimation of ascorbic acid (ASA)**

Total ASA was estimated following the protocol explained in Arakawa et al. ([1981](#page-12-16)). A total of 200 mg leaf tissue was used to determine the reduction of dehydroascorbic acid (DHA) to ASA by dithiothreitol and expressed as nmol  $g^{-1}$ leaf fresh weight.

# **Estimation of α‑tocopherol**

α-Tocopherol was estimated from 500 mg of leaf tissue following the slightly modifed procedure from Pearson et al. ([1970](#page-14-14)) and described before in Kataria et al. ([2019\)](#page-13-14) and represented as mg  $g^{-1}$  leaf fresh weight.

#### **Nitric oxide (NO) determination**

The procedure from Zhou et al. ([2005\)](#page-15-9) was used to estimate NO content from 500 mg of soybean leaves. The NO content was expressed in nmol  $g^{-1}$  fresh weight of leaves.

# **Nitrate reductase (NR) activity**

The procedure of Jaworski ([1971\)](#page-13-20) was followed to estimate the enzymatic activity of NR from 250 mg of soybean leaf tissue and expressed in µmol NO<sub>2</sub> g<sup>-1</sup> leaf fresh weight h<sup>-1</sup>.

#### **Yield**

After the crop harvest at 120 DAE, the number of pods and seeds, pod weight, and seed yield per plant were measured.

#### **Statistical analysis**

Statistical analysis was performed on Microsoft Excel and Prism 4 (GrafPad Software, La Jolla, California) software where mean and standard errors were calculated, and the analysis of variance (ANOVA) followed by post hoc Newman–Keuls Multiple Comparison Test were performed.  $^{#H#}p < 0.001$ ;  $^{#}p < 0.01$ ;  $^{#}p < 0.05$  indicate the significant diference among the seedlings emerged from untreated (UT) seeds grown-up in ambient UV-B with ambient  $+$  supplemental UV-B conditions;  $**^*p < 0.001$ ;  $**^*p < 0.01$ ;  $*p$ <0.05 indicate significant difference among the seedlings emerged from SMF pretreated (MT) seeds with the seedlings of untreated (UT) seeds grown-up in ambient and ambient+supplemental UV-B conditions.

# **Results**

The effects of UV-B radiation on morphological parameters of SMF pretreated seeds in 45-day-old soybean plants are presented in Fig. [1](#page-4-0). Enhanced UV-B  $_{(a+*s*)}$  significantly reduced the plant height (15%), leaf area (27%), total biomass (22%), and specifc leaf weight (27%) in plants obtained from untreated soybean seeds as compared to the plants from untreated seeds grown in ambient UV-B conditions; while all of these parameters were signifcantly increased by SMF pretreatment (Fig. [1b](#page-4-0)–d). The plant height maximally enhanced by 34% and 67% (Fig. [1](#page-4-0)a); leaf area enhanced by 60% and 115% (Fig. [1](#page-4-0)b), and specifc leaf weight was also increased by 60% and 96% (Fig. [1](#page-4-0)d), respectively, under  $_{\rm a}$ UV-B as well as  $_{\rm a+s}$ UV-B exposure by the SMF treatment as compared to their respective untreated ones.

Total chlorophyll content of leaves was decreased by 25% in enhanced UV-B irradiation plants in comparison to untreated seeds grown under only ambient UV-B, while it was signifcantly increased (21%) by SMF treatment even after the enhanced UV-B irradiation (Fig. [2a](#page-5-0)).

UV-B absorbing compounds were found to be increased in the leaves of soybean plant under the  $_{a}$ UV-B and  $_{a+<sub>s</sub>}$ UV-B (Fig. [2](#page-5-0)b). UAS contents in the leaves of plants from untreated seeds showed a signifcant increase of 28% under  $_{a+s}$ UV-B, whereas a significant reduction of 32% and 30% were recorded respectively under  $_{a}$ UV-B and  $_{a+s}$ UV-B as compared to their untreated ones (Fig. [2b](#page-5-0)).

ChlF data indicate that maximum fluorescence  $(F_m)$ decreased in  $_{a+s}$ UV-B treated plants when compared with



<span id="page-4-0"></span>**Fig. 1** Efect of SMF-priming (200 mT for 1 h) on plant height (**a**), leaf area (**b**), total biomass (**c**), and specifc leaf weight (**d**) of soybean plants under ambient and supplemental UV-B conditions. MT=seedlings from SMF pretreated and UT=seedlings from untreated seeds

ChlF from plants of untreated seeds grown under ambient UV-B. The transient fuorescence curve (OJIP) in plants exposed to ambient or  $_{a+s}$ UV-B after SMF pretreatment is shown in Fig. [3.](#page-6-0) The OJIP curve showed an increase in the *I*–*P* phase due to SMF treatment as compared to the leaves of plants that emerged from UT seeds under ambient as well as  $_{a+s}$ UV-B conditions (Fig. [3\)](#page-6-0).

The photosynthetic parameters such as  $F_v/F_m$ ,  $\varphi$ Eo=ETo/ ABS,  $\Delta V (I-P)$ ,  $PI_{ABS}$  and  $PI_{total}$  were decreased in plants from untreated seeds grown under  $_{a}$ UV-B and  $_{a+s}$ UV-B exposure (Fig. [4](#page-7-0)a–e). SMF treatment caused a slight increase in  $F_v/F_m$  as compared to their UT ones under the presence of UV-B stress (Fig. [4](#page-7-0)a). However, SMF pretreatment signifcantly increased the value of *φ*Eo by 50% and 113% and  $\Delta V (I-P)$  by 12.9% and 13%, respectively, under <sub>a</sub>UV-B and  $a_{+s}$ UV-B in comparison to their UT ones (Fig. [4c](#page-7-0)). Values of PI<sub>abs</sub> and PI<sub>total</sub> parameter derived from ChlF records (Fig. [4](#page-7-0)d, e) confrmed much higher responsiveness compared to  $F_v/F_m$ . A tremendous increase was found in  $PI_{ABS}$ after SMF treatment under  $_{a}$ UV-B (145%) and  $_{a+s}$ UV-B (254%) (Fig. [4](#page-7-0)d). SMF pretreatment also caused a signifcant increase of 43% and 68% in  $PI_{total}$ , respectively under  $_{a}$ UV-B and  $_{a+s}$ UV-B as compared to their UT ones (Fig. [4e](#page-7-0)).

Signifcant inhibition of the net rate of photosynthesis (23%) and stomatal conductance (42%) was observed under the enhanced UV-B, whereas internal  $CO<sub>2</sub>$  concentration was observed to be increased by 15% in the leaves of soybean plants emerged from untreated seeds as compared to the plants grown in ambient UV-B condition (Fig. [5a](#page-8-0)–c). On the other hand, *Pn* was increased by 36% and 46%, respectively, under  $_{a}$ UV-B and  $_{a+1}$ UV-B in the plants from SMF-treated seeds in comparison to the plants obtained from untreated seeds (Fig. [5a](#page-8-0)). SMF pretreatment caused 38% and 64% enhancement in stomatal conductance, respectively, under  $_{a}UV-B$  and  $_{a+s}UV-B$  in comparison to their untreated seedlings (Fig. [5b](#page-8-0)). However, SMF pretreatment caused a reduction of 56% and 52% in internal  $CO_2$  concentration, respectively, for <sub>a</sub>UV-B and  $_{a+e}$ UV-B conditions (Fig. [5c](#page-8-0)).

Superoxide anion radical  $(O_2^-)$  and hydrogen peroxide  $(H<sub>2</sub>O<sub>2</sub>)$  content was observed to be higher in the leaves of plants that emerged from UT seeds under  $_{a}$ UV-B and  $_{a+s}$ UV-B conditions when compared with plants from SMF pretreated seeds (Fig. [6](#page-9-0)a, b). Most of the antioxidant activities were found to be increased in presence of  $_{a+s}$ UV-B depicting a better defensive response of the soybean plants under  $_{a+s}$ UV-B exposure. Among non-enzymatic antioxidants, ascorbic acid showed a signifcant increase of 24% and  $\alpha$ -tocopherol showed an increase of 52% by  $_{3+\text{s}}$ UV-B as compared to their untreated plants grown under  $_{a}$ UV-B



<span id="page-5-0"></span>**Fig. 2** Efect of SMF-priming (200 mT for 1 h) on Total Chlorophyll (**a**), and UAS (**b**) in third trifoliate leaves of soybean plants under ambient and supplemental UV-B conditions. MT=seedlings from SMF pretreated and  $UT =$  seedlings from untreated seeds

condition (Fig. [6](#page-9-0)c, d), while SMF pretreatment caused a signifcant decrease in ASA and α-tocopherol content under  $_{a}$ UV-B and  $_{a+s}$ UV-B (Fig. [6c](#page-9-0), d).

Nitric oxide (NO) content was remarkably increased by 63% under  $_{a+s}$ UV-B in plants from untreated seeds as compared to their plants grown under ambient UV-B stress (Fig. [7a](#page-10-0)). SMF treatment increased the NO content by 58% and 23%, respectively, in  $_{a}$ UV-B and  $_{a+s}$ UV-B conditions as compared to their UT ones (Fig. [7](#page-10-0)a). NR activity was decreased by  $_{a+s}$ UV-B, while SMF-treatment significantly enhanced the NR activity in  $_{a}UV-B$  and  $_{a+s}$ UV-B conditions as compared to their respective UT ones (Fig. [7](#page-10-0)b).

A significant difference in yield parameters such as the number of pods/seeds and the weight of pods/seeds per plant was observed at crop maturity (Fig. [8](#page-11-0)a–d). We observed a negative effect of  $_{a+s}$ UV-B stress on the yield of soybean, but the plants that emerged from SMF-treated seeds showed a significant increase of 35% in the number of pods, 39% in pod weight, 45% in the number of seeds, and 54% in seed weight per plant in comparison to their respective UT ones under  $_{a+s}$ UV-B stress conditions (Fig. [8](#page-11-0)a–d).

# **Discussion**

In this study plant growth, the efficiency of PSII, performance indices, photosynthesis, and yield was negatively impacted by enhanced UV-B, whereas SMF pretreatment was observed to mitigate the UV-B-induced oxidative stress in soybean plants. A signifcant decline in several morphological and physiological characteristics (plant height, dry mass, leaf area, chlorophyll content, the efficiency of PSII, and photosynthesis) was observed in soybean after its exposure to  $_{a}$ UV-B together with  $_{s}$ UV-B. A reason behind the morphological and physiological changes in the plant can be related to an enhanced level of ROS ( $O_2$ <sup>--</sup> and H<sub>2</sub>O<sub>2</sub>) observed in this study and it can be related to previous studies (Reddy et al. [2004](#page-14-15); Shine and Guruprasad [2012](#page-14-16); Rastogi and Pospisil [2013;](#page-14-8) Kataria et al. [2020](#page-13-15)). The decline in dry mass and leaf area due to enhanced UV-B could be due to a reduction in the cytokinin content, the extent of cell division, and elongation as described in previous studies (Hopkins et al. [2002;](#page-13-21) Kataria et al. [2014a](#page-13-2), [b](#page-13-3); Singh et al. [2014;](#page-14-17) Kataria and Guruprasad [2018\)](#page-13-22). Reduction in plant height by enhanced UV-B could have been due to photo-oxidative damage of the phytohormone indole acetic acid followed by lower cell wall extensibility, as established in sunfower seedlings (Ros and Tevini [1995\)](#page-14-18). On the other hand, our results revealed that SMF pretreatment signifcantly enhanced the plant height, leaf area, specifc leaf weight, and total biomass accumulation even under  $_{a+s}$ UV-B. Some previous studies demonstrated that SMF pretreatment could ameliorate the inhibition of growth, photosynthesis, and yield caused by the ambient UV-B stress in soybean and maize (Shine and Guruprasad [2012](#page-14-16); Kataria et al. [2017,](#page-13-13) [2020;](#page-13-15) Baghel et al. [2015\)](#page-12-17). A signifcant increase in SLW after SMF pretreatment infuenced the plant's higher biomass and an increase in leaf thickness. This is the frst study showing that SMF  $(200$  mT for 1 h) pretreatment considerably increased efficiency of PSII, the quantum yield of electron transport, performance indices ( $PI<sub>abs</sub>$  and  $PI<sub>total</sub>$ ), rate of photosynthesis, NR activity, and NO content as compared to the plants that emerged from UT seeds grown under enhanced UV-B stress  $(a + sUV-B)$  conditions.

A decrease in total chlorophyll concentration under enhanced UV-B stress observed is in agreement with studies under elevated UV-B radiation (Zhao et al. [2003](#page-15-10); Reddy et al. [2004](#page-14-15)). UV-B might have caused the <span id="page-6-0"></span>**Fig. 3** Efect of SMF-priming (200 mT for 1 h) on chlorophyll fuorescence emission transients in third trifoliate leaves of the soybean seedlings grown under ambient and supplemental UV-B conditions. MT=seedlings from SMF pretreated and UT=seedlings from untreated seeds.  $MT<sub>a</sub>UV-B=$  plants emerged from SMF-treated seeds grown under ambient  $UV-B$ ;  $UT<sub>a</sub>UV-B$  = plants emerged from UT seeds grown under ambient UV-B;  $MT_{a+s}UV-B=$  plants emerged from SMF-treated seeds grown under ambient plus supplemental UV-B; and  $UT_{a+s}UV-B=$  plants emerged from UT seeds grown under ambient plus supplemental UV-B stress.The curve was normalized for  $F_{o}$  (fluorescence at time 0) and  $F_t/F_o$  (where  $F_t$  is fuorescence at time *t*) against the time



destruction of chloroplast structure, interferes in chlorophyll synthesis through destruction of enzymes, and may have enhanced chlorophyll degradation which may have resulted in a decrease in total chlorophyll concentration observed in this study and supported by previous studies (Sakaki et al. [1983;](#page-14-19) Kataria et al. [2013](#page-13-7), [2014a](#page-13-2), [b](#page-13-3)). The exposure of the plant to  $_{a+s}$ UV-B significantly increased the UV-B absorbing compounds that emerged from untreated seeds in the current study as reported previously (Kakani et al. [2004](#page-13-23); Reddy et al. [2004\)](#page-14-15). The increase in favonoids had been recognized as a common and quick response to UV-B stress (Tiitto et al. [2015\)](#page-15-11). In a study, the total phenolic and favonoid content was observed to increase in lettuce plants exposed to direct sunlight (UV exposed) in contrast to greenhouse conditions (Low UV) by the fuorescence excitation ratio method (Zivcak et al. [2017\)](#page-15-12). The favonoids have efective free radical-scavenging capabilities and could contribute directly to enhanced photoprotection against UV-B radiation (Mosadegh et al. [2018\)](#page-13-24). Thus, favonoids could protect photosynthetic pigments and may help in sustaining the photosynthetic activity (Day and Neale [2002\)](#page-12-18). In this study, the favonoids concentrations were observed to be decreased while total Chl concentration was increased in combined treatment of SMF with UV-B radiation.

It has been previously reported that the supplemental UV-B irradiance harms the physiological processes of plants including the reduction of photosynthetic efficiency, decrease of leaf stomatal conductance, and transpiration rate (Krupa and Kickert [1989;](#page-13-25) Chen and Zhang  $2007$ ; Kataria et al.  $2014a$ , [b](#page-13-3)). The photosynthetic efficiency of soybean seedlings was determined in terms of chlorophyll fuorescence (JIP-test parameters) (Figs. [3,](#page-6-0) [4](#page-7-0)) and gas exchange parameters (Fig. [5](#page-8-0)). It showed that  $_{a+s}$ UV-B reduced the *I–P* phase of the OJIP curve in the leaves of plants that emerge from UT seeds, while plants that emerged from SMF-treated seeds showed a signifcant increase in the  $I-P$  phase even under  $_{a+s}$ UV-B stress. The *I–P* phase is related to the electron transfer through PS I (Schansker et al. [2006\)](#page-14-20). The *I*–*P* phase may decline under several kinds of stresses such as nitrogen deficiency (Nikiforou and Manetas [2011\)](#page-14-21), salinity (Oukarroum et al. [2015](#page-14-22)), heavy metal (Bernardini et al. [2015\)](#page-12-20), drought (Pollastrini et al. [2014\)](#page-14-23), and ozone pollution (Bussotti et al. [2011](#page-12-21)). The increase in chlorophyll fuorescence specifcally in the *I*–*P* phase of the OJIP curve is characteristically recognized to the reduction of electron transporters (ferredoxin, intermediary acceptors, and NADP) of thePSI acceptor side (Kalaji et al. [2016](#page-13-18)). A positive correlation between Δ*V*(*I-P*) and net photosynthesis (Pn) have been observed in plants (Cascio et al. [2010](#page-12-22); Santos et al. [2019\)](#page-14-24). Δ*V*(*I-P*) is measured as a proxy of the concentration of PSI reaction centers (Schansker et al. [2005](#page-14-25)), which is implied in the production of NADPH<sup>+</sup> for  $CO<sub>2</sub>$  fixation. The analysis of



<span id="page-7-0"></span>**Fig. 4** Efect of SMF-priming (200 mT for 1 h) on maximum photochemical efficiency of PS II,  $F_v/F_m$  (a), quantum yield of electron transport, *φ*Eo (**b**), relative amplitude of the *I*–*P* phase of Chl*a* fuorescence,  $\Delta V (I-P)$  (c), performance index on absorbtion basis,  $PI_{ABS}$ ,

 $(d)$ , and performance index total,  $PI_{total}$   $(e)$  in third trifoliate leaves of soybean plants under ambient and supplemental UV-B conditions. MT=seedlings from SMF pretreated and UT=seedlings from untreated seeds

fast ChlF transient was applied in numerous studies in crop plants, to study the environmental efects such as salinity, drought, high/low temperature (Stirbet et al. [2018\)](#page-14-26), and light stress (Kalaji et al. [2018\)](#page-13-9). Signifcant suppression in Chl fuorescence induction curve was observed in saltsensitive genotypes, whereas for salt-resistant sorghum



<span id="page-8-0"></span>**Fig. 5** Efect of SMF-priming (200 mT for 1 h) on rate of photosynthesis (a), stomatal conductance  $(b)$ , and internal  $CO<sub>2</sub>$  concentration (**c**) in third trifoliate leaves of soybean plants under ambient and supplemental UV-B conditions. MT=seedlings from SMF pretreated and UT=seedlings from untreated seeds

genotypes the photosynthetic machinery was working even at a salt concentration of 200 mM (Rastogi et al. [2020\)](#page-14-7).

Our results showed that  $_{a+s}$ UV-B exposure reduced  $F_v/F_m$  and  $\Delta V(I-P)$  values, the quantum yield of electron transport,  $PI_{ABS}$  and  $PI_{total}$  in plants from untreated seeds (Fig. [4](#page-7-0)), indicating the destruction of PSII under UV-B stress (Albert et al. [2005;](#page-12-23) Kataria et al. [2013,](#page-13-7) [2014a](#page-13-2), [b](#page-13-3)). Similarly, Rastogi et al. ([2020](#page-14-7)) have found lower values of  $F_v/F_m$  and PI<sub>ABS</sub> in sorghum under salt stress. Kalaji et al. [\(2018](#page-13-9)) also observed that after 7 days of exposure to different abiotic stress factors  $PI_{\rm ABS}$  was most sensitive to the fuctuations in diferent conditions in barley landraces, as it characterizes the overall activity of PSII. Several performance indices (PIs) have been defned, which are suggested to provide information on the structure and function of PSII,

and the efficiencies of specific electron transport reactions in the thylakoid membrane (Stirbet et al. [2018\)](#page-14-26). Further, these PIs have been expected to quantify the plant tolerance to various abiotic stresses like drought, high light, high/ low temperature, or N-deficiency (Živčák et al. [2014a,](#page-15-13) [b](#page-15-14); Stirbet et al. [2018](#page-14-26); Kalaji et al. [2018\)](#page-13-9). In the present study, we have observed that  $\varphi$ Eo=ETo/ABS, PI<sub>ABS,</sub> and PI<sub>total</sub> were signifcantly increased by SMF pretreatment than commonly used parameter  $F_v/F_m$  under <sub>a</sub>UV-B as well  $_{a+s}$ UV-B conditions and it was shown to be well associated with photosynthetic capacity measured as  $CO<sub>2</sub>$  assimilation (Ripley et al. [2004\)](#page-14-27). Similarly, Živčák et al. ([2014b](#page-15-14)) indicated that the normally used parameter  $F_v/F_m$  was more or less insensitive to nitrogen treatment but the  $PI<sub>ABS</sub>$  and  $PI<sub>total</sub>$ were much more responsive and signifcant diferences were found among the plants of diferent nitrogen treatments.The authors also observed that the  $PI_{total}$  is sensitive in comparison to  $PI<sub>ABS</sub>$ , especially under field conditions, as  $PI<sub>total</sub>$  also considers the quantum efficiency of reduction in PSI ends acceptor (Redillas et al. [2011](#page-14-28); Živčák et al. [2014b;](#page-15-14) Banks [2017\)](#page-12-13). Redillas et al. [\(2011\)](#page-14-28) also observed a decrease in  $PI_{total}$  during N-depletion as a result of the decrease in the reduction of end electron acceptors on PSI. In this study, we have observed signifcant diferences between treatments for  $PI<sub>ABS</sub>$  and  $PI<sub>total</sub>$ , but the values of  $PI<sub>ARS</sub>$  showed a higher difference between the treatments, which indicated  $PI<sub>ARS</sub>$ are sufficiently sensitive for UV-B-related study in a plant. It has also been reported that reduction in quantum yield of electron transport (*φ*Eo) and electron fuxes per crosssection of the leaves in  $C_3$  and  $C_4$  plant species under ambient UV-B stress caused the signifcant decline in uptake of carbon (Kataria et al. [2013\)](#page-13-7). Thus our results suggested that higher  $\varphi$ Eo=ETo/ABS,  $\Delta V(I-P)$ , PI<sub>ABS,</sub> and PI<sub>total</sub> in the soybean plants obtained from SMF-primed seeds contributes to the higher efficiency of light-harvesting and as a result increase the biomass accumulation and uptake of  $CO<sub>2</sub>$  even in the presence  $_{a+s}$ UV-B stress.

Moreover, in the present study, a signifcant reduction in photosynthetic  $CO<sub>2</sub>$  uptake was observed with a decrease in stomatal conductance and increase in internal  $CO<sub>2</sub>$  concentration in plants from untreated seeds grown under  $_{a+s}$ UV-B (Fig. [5\)](#page-8-0), suggesting that the decline in photosynthesis may be due to the UV-B-induced ROS, destroyed chlorophyll, and decreased efficiency of PSII and Rubisco activity (Kataria et al. [2013;](#page-13-7) Sztatelman et al. [2015](#page-15-15)). Similarly, previous studies have reported that enhanced UV-B causes reduction in leaf thickness (Inostroza-Blancheteau et al. [2014](#page-13-26)), stomatal conductance (Martinez-Luscher et al. [2013](#page-13-27)), photosynthetic  $CO<sub>2</sub>$  assimilation (Basahi et al. [2014](#page-12-24)), and photosynthetic efficiency (related with photosystem II) (Jordan et al. [2016](#page-13-28)), as a result, it decrease the crop growth and productivity (Kakani et al. [2003a,](#page-13-4) [b;](#page-13-5) Kataria et al. [2013,](#page-13-7) [2014a,](#page-13-2) [b](#page-13-3)). Zivcak et al. ([2017\)](#page-15-12) found that in corresponding





<span id="page-9-0"></span>**Fig. 6** Efect of SMF-priming (200 mT for 1 h) on superoxide radical (**a**), hydrogen peroxide (**b**), ascorbic acid (**c**), and  $\alpha$ -tocopherol (**d**) content in third trifoliate leaves of soybean plants under ambient

and supplemental UV-B conditions. MT=seedlings from SMF pretreated and UT=seedlings from untreated seeds

with photochemical parameters, the assimilation of  $CO<sub>2</sub>$  was also robustly decreased in low light as compared to high light and they suggested this inhibition was not due to stomatal closure. Further, they found  $CO<sub>2</sub>$  assimilation was deficient by the PSI electron transport supply at low to moderate light intensities, but at high light, carboxylation rate seemed to be decreased due to the downregulated activity of Calvin cycle enzymes, which is fne-tuned by the redox signaling at the PSI acceptor side. The response of photosynthetic performance under  $_{a}$ UV-B and  $_{a+s}$ UV-B in soybean plants that emerged from SMF pretreated seeds in this study was in accordance with numerous previous studies on soybean (Shine et al. [2011](#page-14-29), [2012](#page-14-30); Kataria et al. [2020](#page-13-15)).

The production of ROS is the most common cause for the reduction in photochemical efficiency and photosynthetic rate (Shine and Guruprasad [2012;](#page-14-16) Kataria et al. [2014a,](#page-13-2) [b](#page-13-3)). PSII reaction center is the most sensitive component of photosynthetic apparatus to UV-B exposure (Kataria et al. [2014a,](#page-13-2) [b\)](#page-13-3). An increase in the  $H_2O_2$  content in the leaves of UV-B treated plants was observed by several researchers (Rybus-Zając [2005;](#page-14-31) Kubiś and Rybus-Zając [2008](#page-13-29)). In our study, we have also observed a higher amount of  $O_2^{\text{-}}$  and H<sub>2</sub>O<sub>2</sub> and antioxidants (ASA and α-tocopherol) in the plants from untreated seeds exposed to  $_{a+s}$ UV-B. However, SMFtreated plants showed a lower amount of ROS and antioxidants even after exposure to  $_{a+s}$ UV-B; this may support the soybean plants to utilize their metabolic energy to improve crop growth and productivity. Earlier, it has been reported that the reduction in the net photosynthetic rate of crops may be closely related to the inhibition of PSII efficiency, photosynthetic enzymes, and ultrastructural changes in chloroplasts due to higher ROS after exposure to UV-B (Jordan et al. [1992;](#page-13-30) He et al. [1994](#page-13-31); Greenberg et al. [1997](#page-12-25); Kataria et al. [2014a,](#page-13-2) [b\)](#page-13-3). Several reports have shown the involvement of ROS molecules in UV-B induced damages to the plants (Snyrychova et al. [2007](#page-14-32)). Thus, an increase in antioxidant defense such as higher ascorbic acid and α-tocopherol and oxidative membrane damage products were observed



<span id="page-10-0"></span>Fig. 7 Effect of SMF-priming (200 mT for 1 h) on nitric oxide content (**a**), and nitrate reductase activity (**b**) in third trifoliate leaves of soybean plant under ambient and supplemental UV-B conditions. MT=seedlings from SMF pretreated and UT=seedlings from untreated seeds

in plants exposed to UV-B stress (Rao and Ormrod [1995](#page-14-33); Malanga et al. [1997;](#page-13-32) Jain et al. [2004;](#page-13-11) Kataria et al. [2007](#page-13-33); Baroniya et al. [2013](#page-12-26); Dwivedi et al. [2015\)](#page-12-27).

Nitric oxide (NO) is an important signaling molecule involved in the amelioration of growth and development of plants under various biotic and abiotic stresses (Ahmad et al. [2016;](#page-11-1) Ahanger et al. [2020](#page-12-28)). In recent years NO was observed to play an important role defense mechanism in response to UV-B radiation (Kataria et al. [2020\)](#page-13-15). NR is an important enzyme in N metabolism, that is used to converts nitrate  $(NO<sub>3</sub><sup>-</sup>)$  into nitrite  $(NO<sub>2</sub><sup>-</sup>)$  and required for the formation of amino acids (Canovas et al. [2007\)](#page-12-29). In the present study, enhanced UV-B caused a decrease in NR activity in leaves of plants that emerged from untreated seeds as compared to plants grown in ambient UV-B. While SMF treatment enhanced the NO content and NR activity under ambient and enhanced UV-B stress as compared to their unprimed ones. NO has been found as the second messenger related to growth and development under UV-B irradiation (Zhang et al. [2003](#page-15-16)). Zhang et al. ([2011\)](#page-15-4) found a connection between UV-B-induced favonoid production and NR mediated NO production in *Betula pendula*. Nitrate reductase is involved in 24-epibrassinolide-induced NO synthesis to improve

tolerance toward iron defciency in strawberry and cadmium toxicity in pepper plants (Kaya et al. [2020a](#page-13-34), [b\)](#page-13-35). The response of soybean plants to  $_{a+s}$ UV-B and SMF pretreatment in this study was in accordance with several earlier studies on soybean (Baroniya et al. [2013](#page-12-26), [2014;](#page-12-11) Kataria et al. [2017](#page-13-13)). Related studies reported that an enhanced or supplemental UV-B radiation could signifcantly reduce the NR activity in soybean (Rockel [2002\)](#page-14-34), maize (Quaggiotti et al. [2004\)](#page-14-35), *Vigna* (Balakumar et al. [1999\)](#page-12-30), and barley (Ghisi et al. [2002\)](#page-12-31). The NR reducing activity also infuences the nitrogen metabolism by infuencing photosynthetic machinery (Baroniya et al. [2014\)](#page-12-11).

As the stratospheric ozone layer is depleting the farmers and scientists are concern about the decreasing crop yield. A decrease in biomass is generally observed with an increase in secondary metabolites (Zhang and Björn [2009](#page-15-17)), which indicates a less anticipated economic loss. In our study the fnal yield or biomass may have infuenced by various parameters that are afected by UV-B radiation; such as decreases in chlorophyll concentration, the efficiency of PSII, rate of photosynthesis, leaf area, and increase in ROS. The present study confrmed previous reports (Qiang et al. [2004;](#page-14-36) Liu et al. [2013\)](#page-13-6) that UV-B radiation on the canopy changed the soybean agronomic traits and decrease the yield per plant. Chen et al. ([2004\)](#page-12-32) reported that the seed weight of 20 soybean cultivars showed diferent sensitivity to UV-B radiation and found that the seed weight of the 15 soybean cultivars decreased quite signifcantly. However, SMF treatment caused a signifcant decrease in UV-B absorbing substances, ROS, ASA, and  $\alpha$ -tocopherol content while caused a remarkable increase in efficiency of PSII, and JIP-test parameters such as  $\varphi$ Eo,  $\Delta V(I-P)$ , PI<sub>ABS,</sub> and PI<sub>total</sub> along with a higher rate of photosynthesis, higher NO and NR activity; consequently, the SMF-treatment remarkably improve all the yield parameters studied under ambient and supplemental UV-B stress. Thus, SMF-priming of dry seeds of soybean was effective in mitigating the adverse effects of supplemental UV-B and improved the crop yield.

# **Conclusions**

In conclusion, enhanced/supplemental UV-B radiation had an adverse efect on plant height, leaf area, specifc leaf weight, biomass, efficiency of PSII, photosynthesis, and grain yield of soybean crop under feld conditions. The impact of UV-B on biological systems mainly on plants is found to be severe because of the sessile nature of the plants. The adverse efect of  $_{a+s}$ UV-B can be alleviated by magnetopriming with the SMF of 200 mT for 1 h in soybean plants. The strong defense strategy of SMF-treated plants was to reduce the damaging efects of ROS induced by enhanced UV-B stress especially on the photosynthetic performance of soybean plants. The



<span id="page-11-0"></span>**Fig. 8** Efect of SMF-priming (200 mT for 1 h) on yield parameters such as number of seeds (**a**), weight of pods (**b**), number of seeds (**c**), and seed weight (**d**) per plant of soybean under ambient and supple-



mental UV-B conditions. MT=seedlings from SMF pretreated and UT=seedlings from untreated seeds

enhanced photosynthetic efficiency, higher  $\varphi$ Eo,  $\Delta V(I-P)$ ,  $PI<sub>ABS</sub>$ ,  $PI<sub>total</sub>$ , and NO/NR activity subsequently caused improvement in yield of soybean plants emerged from SMFprimed seeds under UV-B stress; it is an integration of stimulation of tolerance towards enhanced or supplemental UV-B stress as compared to the plants emerged from untreated seeds. Future substantiation of the results under feld conditions can help in promoting magnetopriming as a feasible option for alleviating the adverse efect of enhanced UV-B stress.

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# **Declarations**

**Conflict of interest** The authors declare that they have no confict of interest.

# **References**

<span id="page-11-1"></span>Ahmad P, Abdel Latef AA, Hashem A, Abd\_Allah EF, Gucel S, Tran SP (2016) Nitric oxide mitigates salt stress by regulating levels

of osmolytes and antioxidant enzymes in Chickpea. Front Plant Sci 7:347. <https://doi.org/10.3389/fpls.2016.00347>

- <span id="page-12-28"></span>Ahanger MA, Aziz U, Alsahli AA, Alyemeni MN, Ahmad P (2020) Influence of exogenous salicylic acid and nitric oxide on growth, photosynthesis, and ascorbate-glutathione cycle in salt stressed *Vigna angularis*. Biomolecules 10(1):42. [https://](https://doi.org/10.3390/biom10010042) [doi.org/10.3390/biom10010042](https://doi.org/10.3390/biom10010042)
- <span id="page-12-23"></span>Albert KR, Mikkelsen TN, Ro-Poulsen H (2005) Efects of ambient versus reduced UV-B radiation on high arctic *Salix arctica* assessed by measurements and calculations of chlorophyll a fuorescence parameters from fuorescence transients. Physiol Plant 124:208–226. [https://doi.org/10.1111/j.1399-3054.2005.](https://doi.org/10.1111/j.1399-3054.2005.00502.x) [00502.x](https://doi.org/10.1111/j.1399-3054.2005.00502.x)
- <span id="page-12-5"></span>Allen DJ, Nougúes S, Baker NR (1998) Ozone depletion, and increased UV-B radiation: is there a real threat to photosynthesis? J Exp Bot 49:1775–1778. <https://doi.org/10.1093/jxb/49.328.1775>
- <span id="page-12-7"></span>An LZ, Liu YH, Zhang MX (2005) Efect of nitric oxide on growth of maize seedling leaves in the presence or absence of ultraviolet-B radiation. J Plant Physiol 162:317–326. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jplph.2004.07.004) [jplph.2004.07.004](https://doi.org/10.1016/j.jplph.2004.07.004)
- <span id="page-12-6"></span>Akhter MS, Noreen S, Mahmood S, Athar HR, Ashraf M, Abdullah AA, Ahmad P (2021) Infuence of salinity stress on PSII in barley (*Hordeum vulgare* L.) genotypes, probed by chlorophyll-a fuorescence. J King Saud Univ Sci 33:101239. [https://doi.org/](https://doi.org/10.1016/j.jksus.2020.101239) [10.1016/j.jksus.2020.101239](https://doi.org/10.1016/j.jksus.2020.101239)
- <span id="page-12-8"></span>Anand A, Nagarajan S, Verma A, Joshi D, Patha P, Bhardwaj J (2012) pretreatment of seeds with static magnetic feld ameliorates soil water stress in seedlings of maize (*Zea mays* L.). Indian J Biochem Biophys 49:63–70
- <span id="page-12-16"></span>Arakawa N, Tsutsumi K, Sanceda NG, Kurata T, Inagaki C (1981) A rapid and sensitive method for determination of ascorbic acid using 4,7-Diphenyl-l, 10-bathophenanthroline. Agric Biol Chem 45:1289–1290
- <span id="page-12-13"></span>Banks JM (2017) Continuous excitation chlorophyll fuorescence parameters: a review for practitioners.Tree Physiol 37:1128– 1136. <https://doi.org/10.1093/treephys/tpx059>
- <span id="page-12-17"></span>Baghel L, Kataria S, Guruprasad KN (2015) Impact of pre-sowing exposure of seeds to stationary magnetic feld on nitrogen and carbon metabolism in maize and soybean. Int J Trop Agric 33:977–983
- <span id="page-12-9"></span>Baghel L, Kataria S, Guruprasad KN (2018) Efect of SMF pretreatment on growth, photosynthetic performance and yield of soybean under water stress. Photosynthetica 56:718–730
- <span id="page-12-1"></span>Bais AF, Bernhard G, McKenzie RL, Aucamp PJ, Young PJ, Ilyas M, Jockel P, Deushi M (2019) Ozone–climate interactions and efects on solar ultraviolet radiation. Photochem Photobiol Sci 18:602–640
- <span id="page-12-30"></span>Balakumar T, Sevakumar V, Sathiameena K, Ilanchezhian CM, Paliwal K (1999) UV-B radiation mediated alterations in the nitrate assimilation pathway of cropplants-1. Kinetic characteristics of nitrate reductase. Photosynthetica 37:459–467
- <span id="page-12-10"></span>Baroniya SS, Kataria S, Pandey GP, Guruprasad KN (2011) Intraspecifc variationin sensitivity to ambient ultraviolet-B radiation in growth and yield characteris-tics of eight soybean cultivars grown under feld conditions. Braz J Plant Physiol 23:197–202
- <span id="page-12-26"></span>Baroniya SS, Kataria S, Pandey GP, Guruprasad KN (2013) Intraspecifc variations in antioxidant defense responses and sensitivity of soybean varieties to ambient UV radiation. Acta Physiol Plant 35:1521–1530
- <span id="page-12-11"></span>Baroniya SS, Kataria S, Pandey GP, Guruprasad KN (2014) Growth, photosynthesis and nitrogen metabolism in soybean varieties after exclusion of the UV-B and UV-A/B components of solar radiation. Crop J 2:388–397
- <span id="page-12-24"></span>Basahi JM, Ismail IM, Hassan IA (2014) Efects of enhanced UV-B radiation and drought stress on photosynthetic performance of

lettuce (*Lactuca sativa* L. Romaine) plants. Annu Res Rev Biol 4:1739

<span id="page-12-20"></span>Bernardini A, Salvatori E, Guerrini V, Fusaro L, Canepari S, Manes F (2015) Efects of high Zn and Pb concentrations on *Phragmites australis* (Cav.) Trin. Ex. Steudel: photosynthetic performance and metal accumulation capacity under controlled conditions. Int J Phytoremediat 18:16–24. [https://doi.org/10.1080/15226514.](https://doi.org/10.1080/15226514.2015.1058327) [2015.1058327](https://doi.org/10.1080/15226514.2015.1058327)

<span id="page-12-0"></span>Björn LO (2015) Ultraviolet-A, B and C. UV4 Plants Bull 1:17–18

- <span id="page-12-2"></span>Bornman JF, Barnes PW, Robson TM, Robinson SA, Jansen MAK, Ballaré CL, Flint SD (2019) Linkages between stratospheric ozone, UV radiation and climate change and their implications for terrestrial ecosystems. Photochem Photobiol Sci 18:681–716. <https://doi.org/10.1039/C8PP90061B>
- <span id="page-12-21"></span>Bussotti F, Desotgiu R, Cascio C, Pollastrini M, Gravano E, Gerosa G, Marzuoli C, Lorenzini G, Salvatori E, Manes F, Schaub M, Strasser RJ (2011) Ozone stress in woody plants assessed with chlorophyll a fuorescence. A critical reassessment of existing data. Environ Exp Bot 73:19–30. [https://doi.org/10.1016/j.envex](https://doi.org/10.1016/j.envexpbot.2010.10.022) [pbot.2010.10.022](https://doi.org/10.1016/j.envexpbot.2010.10.022)
- <span id="page-12-12"></span>Bussotti F, Gerosa G, Digrado A, Po llastrini M (2020) Selection of chlorophyll fuorescence parameters as indicators of photosynthetic efficiency in large scale plant ecological studies. Ecol Indic 108:105686. <https://doi.org/10.1016/j.ecolind.2019.105686>
- <span id="page-12-4"></span>Caldwell MM, Teramura AH, Tevini M, Bornman JF, Bjorn LO, Kulandaivellu G (1995) Efects of increased solar ultraviolet radiation on terrestrial plants. Ambio 24:166–173
- <span id="page-12-3"></span>Caldwell MM, Bornman JF, Ballaré CL, Flint SD, Kulandaivelu G (2007) Terrestrial ecosystems, increased solar ultraviolet radiation, and interactions with other climate change factors. Photochem Photobiol Sci 6:252–266
- <span id="page-12-29"></span>Canovas FM, Avila C, Canton FR, Canas RA, de la Torre F (2007) Ammonium assimilation and amino acid metabolism in conifers. J Exp Bot 58:2307–2318
- <span id="page-12-22"></span>Cascio C, Schaub M, Novak K, Desotgiu R, Bussotti F, Strasser RJ (2010) Foliar responses to ozone of *Fagus sylvatica* L. seedlings grown in shaded and in full sunlight conditions. Environ Exp Bot 68:188–197. <https://doi.org/10.1016/j.envexpbot.2009.10.003>
- <span id="page-12-15"></span>Chaitanya KS, Naithani SC (1994) Role of superoxide, lipid peroxidation and superoxide dismutase in membrane perturbation during loss of viability in seeds of *Shorea robusta* Gaertn.f. New Phytol 126:623–627
- <span id="page-12-19"></span>Chen L, Zhang S (2007) Efects of enhanced ultraviolet-B radiation on water use efficiency, stomatal conductance, leaf nitrogen content, and morphological characteristics of *Spiraea pubescens* in a warm-temperate deciduous broad-leaved forest. Front For China 2:401.<https://doi.org/10.1007/s11461-007-0064-6>
- <span id="page-12-32"></span>Chen JJ, Zu YQ, Chen HY, Li Y (2004) Infuence of enhanced UV-B radiation on growth and biomass allocation of twenty soybean cultivars. J Agro-Environ Sci 23:29–33
- <span id="page-12-18"></span>Day TA, Neale PJ (2002) Effects of UV-B radiation on terrestrial and aquatic primary producers. Annu Rev Ecol Syst 33:371–396
- <span id="page-12-27"></span>Dwivedi R, Singh VP, Kumar J, Prasad SM (2015) Diferential physiological and biochemical responses of two Vigna species under enhanced UV-B radiation. J Radiat Res Appl Sci 8:173–181
- <span id="page-12-14"></span>Fatima A, Kataria S, Prajapati R, Jain M, Agrawal AK, Singh B, Kashyap Y, Tripathi DK, Singh VP, Gadre R (2020) Magnetopriming efects on arsenic stress-induced morphological and physiological variations in soybean involving synchrotron imaging. Physiol Plant.<https://doi.org/10.1111/ppl.13211>
- <span id="page-12-31"></span>Ghisi R, Trentin AR, Masi A, Ferretti M (2002) Carbon and nitrogen metabolism in barley plants exposed to UV-B radiation. Physiol Plant 116:200–205
- <span id="page-12-25"></span>Greenberg BM, Wilson MI, Huang XD, Duxbury CL, Gerhaddt KE, Gensemer RW (1997) The effects of ultraviolet- B radiation on

higher plants. In: Wang W, Goursuch J, Hughes JS (eds) Plants for environmental studies. CRC Press, Boca Raton, pp 1–35

- <span id="page-13-12"></span>Hasanuzzaman M, Hossain MA, da Silva J, Fujita M (2012) Plant response and tolerance to abiotic oxidative stress, antioxidant defenseis a key factor. In: Venkateswarlu B, Shanker AK, Shanker C, Maheswari M (eds) Crop stress and its management, perspectives and strategies. Springer, Rijeka, pp 261–315
- <span id="page-13-31"></span>He J, Huang LK, Chow WS, Whitecross MI, Anderson JM (1994) Responses of rice and pea plants to hardening with low doses of ultraviolet-B radiation. Aust J Plant Physiol 21:563–574
- <span id="page-13-17"></span>Hiscox J, Israelstam G (1979) A method for the extraction of chlorophyll from leaf tissue without maceration. Can J Bot 57:1332–1334
- <span id="page-13-21"></span>Hopkins L, Bond MA, Tobin AK (2002) Ultraviolet-B radiation reduces the rates of cell division and elongation in the primary leaf of wheat (*Triticum aestivum* L. cv Maris Huntsman). Plant Cell Environ 25:617–624

<span id="page-13-16"></span>Hunt R (1982) Plant growth analysis. University Press, Baltimore, USA

- <span id="page-13-26"></span>Inostroza-Blancheteau C, Reyes-Díaz M, Arellano A, Latsague M, Acevedo P, Loyola R, Arce-Johnson P, Alberdi M (2014) Efects of UV-B radiation on anatomical characteristics, phenolic compounds and gene expression of the phenylpropanoid pathway in highbush blueberry leaves. Plant Physiol Biochem 85:85–95
- <span id="page-13-11"></span>Jain K, Kataria S, Guruprasad KN (2004) Oxyradicals under UV-B stress and their quenching by antioxidants. Indian J Exp Biol 42:884–892
- <span id="page-13-1"></span>Jansen AK, Gaba V, Greenberg BM (1998) Higher plants and UV-B radiation: balancing damage, repair and acclimation. Trends Plant Sci 3:131–135
- <span id="page-13-20"></span>Jaworski EG (1971) Nitrate reductase assay in intact plant tissue. Biochem Biophys Res Commun 43:1274–1279
- <span id="page-13-10"></span>Jenkins GI (2009) Signal transduction in response to UV-B radiation. Annu Rev Plant Biol 60:407–431. [https://doi.org/10.1146/annur](https://doi.org/10.1146/annurev.arplant.59.032607.092953) [ev.arplant.59.032607.092953](https://doi.org/10.1146/annurev.arplant.59.032607.092953)
- <span id="page-13-30"></span>Jordan BR, He J, Chow WS, Anderson JM (1992) Changes in mRNA levels and polypeptide subunits of ribulose-1,5-bisphosphate carboxylase in response to supplemental UV-B radiation. Plant Cell Environ 15:91–98
- <span id="page-13-28"></span>Jordan BR, Strid Å, Wargent JJ (2016) What role does UV-B play in determining photosynthesis? In: Pessarakli M (ed) Handbook of photosynthesis. CRC Press, Boca Raton, pp 275–286
- <span id="page-13-8"></span>Kalaji HM, Schansker G, Ladle RJ et al (2014) Frequently asked questions about in vivo chlorophyll fluorescence: practical issues. Photosynth Res 122:121–158. [https://doi.org/10.1007/](https://doi.org/10.1007/s11120-014-0024-6) [s11120-014-0024-6](https://doi.org/10.1007/s11120-014-0024-6)
- <span id="page-13-18"></span>Kalaji MH, Jajoo A, Oukarroum A, Brestic M, Zivcak M, Samborska AI, Cetner DM, Lukasik I, Goltsev V, Ladle JR (2016) Chlorophyll a fuorescence as a tool to monitor physiological status of plants under abiotic stress conditions. Acta Physiol Plant 38:102
- <span id="page-13-9"></span>Kalaji HM, Rastogi A, Živčák M et al (2018) Prompt chlorophyll fuorescence as a tool for crop phenotyping: an example of barley landraces exposed to various abiotic stress factors. Photosynthetica 56:953–961.<https://doi.org/10.1007/s11099-018-0766-z>
- <span id="page-13-4"></span>Kakani VG, Reddy KR, Zhao D, Mohammed AR (2003a) Efects of ultraviolet-B radiation on cotton (*Gossypium hirsutum* L.) morphology and anatomy. Ann Bot 91:817–826
- <span id="page-13-5"></span>Kakani VG, Reddy KR, Zhao D, Sailaja K (2003b) Field crop responses to ultraviolet-B radiation: a review. Agric For Meteorol 120:191–218.<https://doi.org/10.1016/j.agrformet.2003.08.015>
- <span id="page-13-23"></span>Kakani VG, Reddy KR, Zhao D, Goa W (2004) Senescence and hypospectral refectance of cotton leaves exposed to ultraviolet radiation and carbon dioxide. Physiol Plant 221:250–257
- <span id="page-13-22"></span>Kataria S, Guruprasad KN (2018) Interaction of cytokinins with UV-B (280–315 nm) on the expansion growth of cucumber cotyledons. Hortic Int J 2(2):45–53
- <span id="page-13-33"></span>Kataria S, Jain K, Guruprasad KN (2007) UV-B induced changes in antioxidant enzymes and their isoforms in cucumber (*Cucumis sativus* L.) .cotyledons. Ind J Biochem Biophys 44:31–37
- <span id="page-13-7"></span>Kataria S, Guruprasad KN, Ahuja S, Singh B (2013) Enhancement of growth, photosynthetic performance and yield by the exclusion of ambient UV components in  $C_3$  and  $C_4$  plants. Photochem Photobiol B Biol 127:140–152
- <span id="page-13-2"></span>Kataria S, Jajoo A, Guruprasad KN (2014a) Impact of increasing ultraviolet-B radiation on photosynthetic processes. J Photochem Photobiol B 137:55–66
- <span id="page-13-3"></span>Kataria S, Baroniya S, Baghel L, Kanungo M (2014b) Efect of exclusion of solar UV radiation on plants. Plant Sci Today 1:224–232. <https://doi.org/10.14719/pst.2014.1.4.61>
- <span id="page-13-13"></span>Kataria S, Baghel L, Guruprasad KN (2017) Alleviation of adverse efects of ambient UV stress on growth and some potential physiological attributes in soybean (*Glycine max*) by seed pretreatment with static magnetic feld. J Plant Growth Regul 36:550–565
- <span id="page-13-14"></span>Kataria S, Baghel L, Jain M, Guruprasad KN (2019) Magnetopriming regulates antioxidant defense system in soybean against salt stress. Biocatal Agric Biotechnol 18:101090
- <span id="page-13-15"></span>Kataria S, Rastogi A, Bele A, Jain M (2020) Role of nitric oxide and reactive oxygen species in static magnetic feld pretreatment induced tolerance to ambient UV-B stress in soybean. Physiol Mol Biol Plants 26:939–945. [https://doi.org/10.1007/](https://doi.org/10.1007/s12298-020-00802-5) [s12298-020-00802-5](https://doi.org/10.1007/s12298-020-00802-5)
- <span id="page-13-34"></span>Kaya C, Ashraf M, Alyemeni MN, Ahmad P (2020a) Nitrate reductase rather than nitric oxide synthase activity is involved in 24-epibrassinolide-induced nitric oxide synthesis to improve tolerance to iron defciency in strawberry (*Fragaria* × *annassa*) by up-regulating the ascorbate-glutathione cycle. Plant Physiol Biochem 151:486–499
- <span id="page-13-35"></span>Kaya C, Ashraf M, Alyemeni MN, Ahmad P (2020b) The role of nitrate reductase in brassinosteroid-induced endogenous nitric oxide generation to improve cadmium stress tolerance of pepper plants by up-regulating the ascorbate-glutathione cycle. Ecotoxicol Environ Saf 196:110483. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.ecoenv.2020.110483) [ecoenv.2020.110483](https://doi.org/10.1016/j.ecoenv.2020.110483)
- <span id="page-13-25"></span>Krupa SV, Kickert RN (1989) The greenhouse efect: impacts of ultraviolet (UV)-B radiation, carbon dioxide  $(CO<sub>2</sub>)$  and ozone  $(O<sub>3</sub>)$ on vegetation. Environ Pollut 61:263–392
- <span id="page-13-29"></span>Kubiś J, Rybus-Zając M (2008) Drought and excess UV-B irradiation diferentially alter the antioxidant system in cucumber leaves. Acta Biol Crac Ser Bot 50:35–41
- <span id="page-13-6"></span>Liu B, Liu X-B, Yan-Sheng L, Herbert SJ (2013) Efects of enhanced UV-B radiation on seed growth characteristicsand yield components in soybean. Field Crop Res 154:158–163
- <span id="page-13-32"></span>Malanga G, Calmanovici G, Puntarulo S (1997) Oxidative damage to chloroplast from *Chlorella vulgaris* exposed to ultraviolet-B radiation. Physiol Plant 101:455–462
- <span id="page-13-27"></span>Martinez-Luscher J, Morales F, Delrot S, Sanchez-Diaz M, Gomes E, Aguirreolea J, Pascual I (2013) Short and long-term physiological responses of grapevine leaves to UV-B radiation. Plant Sci 213:114–122
- <span id="page-13-19"></span>Mazza CA, Batista D, Zima AM, Szwarcberg-Bracchitta M, Giordano CV, Acevedo A, Scopel AL, Ballare CL (1999) The efects of solar UV-B radiation on the growth and yield of barley are accompanied by increased DNA damage and antioxidant responses. Plant Cell Environ 22:61–70
- <span id="page-13-0"></span>Mohammed AR, Tarpley L (2010) Diferential response of Southern US rice (*Oryza sativa* L.) cultivars to Ultraviolet-B radiation. J Agron Crop Sci 196:286–295
- <span id="page-13-24"></span>Mosadegh H, Trivellini A, Ferrante A, Lucchesini M, Vernieri P, Mensuali A (2018) Applications of UV-B lighting to enhance phenolic accumulation of sweet basil. Sci Hortic 229:107–116
- <span id="page-14-13"></span>Mukherjee SP, Choudhuri MA (1983) Implications of water stressinduced changes in the levels of endogenous ascorbic acid and hydrogen peroxide in Vigna seedlings. Physiol Plant 58:166–170
- <span id="page-14-10"></span>Munné-Bosch S, Alegre L (2002) The function of tocopherols and tocotrienols in plants. Crit Rev Plant Sci 21:31–57
- <span id="page-14-21"></span>Nikiforou C, Manetas Y (2011) Inherent nitrogen defciency in *Pistacia lentiscus* preferentially affects photosystem I: a seasonal field study. Funct Plant Biol 38:848–855
- <span id="page-14-4"></span>Nogues S, Baker NR (1995) Evaluation of the role of damage to photosystem II in the inhibition of  $CO<sub>2</sub>$  assimilation in pea leaves on exposure to UV-B radiation. Plant Cell Environ 18:781–787
- <span id="page-14-22"></span>Oukarroum A, Bussotti F, Goltsev V, Kalaji HM (2015) Correlation between reactive oxygen species production and photochemistry of photosystems I and II in *Lemna gibba* L. plants under salt stress. Environ Exp Bot 109:80–88. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.envexpbot.2014.08.005) [envexpbot.2014.08.005](https://doi.org/10.1016/j.envexpbot.2014.08.005)
- <span id="page-14-11"></span>Prajapati R, Kataria S, Jain M (2020) Seed priming for alleviation of heavy metal toxicity in plants: an overview. Plant Sci Today 7:16. <https://doi.org/10.14719/pst.2020.7.2.751>
- <span id="page-14-14"></span>Pearson CK, Davies RR, Barnes MM (1970) Separation of alphatocotrienol from alpha-tocopherol by polyehtylenecelite column chromatography. Chem Ind 8:275–276
- <span id="page-14-23"></span>Pollastrini M, Desotgiu R, Camin F, Ziller L, Gerosa G, Marzuoli R, Bussotti F (2014) Severe drought events increase the sensitivity to ozone on poplar clone. Environ Exp Bot 100:94–104. [https://](https://doi.org/10.1016/j.envexpbot.2013.12.016) [doi.org/10.1016/j.envexpbot.2013.12.016](https://doi.org/10.1016/j.envexpbot.2013.12.016)
- <span id="page-14-36"></span>Qiang WY, Yang H, Chen T, An LZ, Wang XL (2004) Efect of the combination of cadmium and UV-B radiation on soybean growth. Chin J Appl Ecol 15:697–700
- <span id="page-14-35"></span>Quaggiotti S, Trentin AR, Dalla Vecchia F, Ghisi R (2004) Response of maize nitrate reductase to UV-B radiation. Plant Sci 167:107–116
- <span id="page-14-33"></span>Rao MV, Ormrod DP (1995) Impact of UV-B and  $O_3$  on the free radical scavenging in *Arabidopsis thaliana* genotypes difering in favonoid biosynthesis. Photochem Photobiol 62:719–726
- <span id="page-14-8"></span>Rastogi A, Pospisil P (2013) Ultra-weak photon emission as a noninvasive tool for the measurement of oxidative stress induced by UVA radiation in *Arabidopsis thaliana*. J Photochem Photobiol B Biol 123:59–64
- <span id="page-14-9"></span>Rastogi A, Yadav D, Szymańska R et al (2014) Singlet oxygen scavenging activity of tocopherol and plastochromanol in *Arabidopsis thaliana*: relevance to photooxidative stress. Plant Cell Environ 37(2):392–401
- <span id="page-14-5"></span>Rastogi A, Stróżecki M, Kalaji HM, Łuców D, Lamentowicz M, Juszczak R (2019a) Impact of warming and reduced precipitation on photosynthetic and remote sensing properties of peatland vegetation. Environ Exp Bot 160:71–80
- <span id="page-14-6"></span>Rastogi A, Zivcak M, Tripathi DK, Yadav S, Kalaji HM, Brestic M (2019b) Phytotoxic efect of silver nanoparticles in *Triticum aestivum*: improper regulation of photosystem I activity as the reason for oxidative damage in the chloroplast. Photosynthetica 57(1):209–216
- <span id="page-14-7"></span>Rastogi A, Kovar M, He X, Zivcak M, Kataria S, Kalaji HM, Skalicky M, Ibrahimova UF, Hussain S, Mbarki S, Brestic M (2020) Special issue in honour of Prof. Reto J Strasser—JIP-test as a tool to identify salinity tolerance in sweet sorghum genotypes. Photosynthetica 58:518–528
- <span id="page-14-15"></span>Reddy KR, Kakanl VG, Zhao D, Kotl S, Gao W (2004) Interactive effects of ultraviolet-B radiation and temperature on cotton physiology, growth, development and hyperspectral refectance. Photochem Photobiol 79:416–427
- <span id="page-14-28"></span>Redillas MC, Jeong JS, Strasser RJ, Kim YS, Kim JK (2011) JIP analysis on rice (*Oryza sativa* cv. Nipponbare) grown under limited nitrogen conditions. J Korean Soc Appl Biol Chem 54:827–832
- <span id="page-14-27"></span>Ripley BS, Redfern SP, Dames JF (2004) Quantifcation of the photosynthetic performance of phosphorus-deficient Sorghum

by means of chlorophyll-a fuorescence kinetics. S Afr J Sci 100:615–618

- <span id="page-14-2"></span>Robson M, Klema K, Urban O, Jansen M (2015) Re-interpreting plant morphological responses to UV-B radiation. Plant Cell Environ 38:856–866. <https://doi.org/10.1111/pce.12374>
- <span id="page-14-34"></span>Rockel P (2002) Regulation of nitric oxide (NO) production by plant nitrate reductase in vivo and in vitro. J Exp Bot 53:103–110. <https://doi.org/10.1093/jexbot/53.366.103>
- <span id="page-14-18"></span>Ros J, Tevini M (1995) Interaction of UV-radiation and IAA during growth of seedlings and hypocotyls segments of sunfower. J Plant Physiol 146:295–302
- <span id="page-14-0"></span>Rowland F (2006) Stratospheric ozone depletion. Philos Trans Biol Sci 361(1469):769–790
- <span id="page-14-31"></span>Rybus-Zając M (2005) Oxidative stress generation in *Taxus baccata*  leaves afected by *Pestalotiopsis funerea* Desm. under diferent light conditions. Dendrobiology 54:51–56
- <span id="page-14-19"></span>SakakiT KN, Sugahara K (1983) Breakdown of photosynthetic pigments and lipids in spinach leaves with ozone fumigation: role of activated oxygen. Physiol Plant 9:28–34
- <span id="page-14-24"></span>Santos VA, Nelson BW, Rodrigues JV, Garcia MN, Ceron VB, Ferreira MJ (2019) Fluorescence parameters among leaf photosynthesisrelated traits are the best proxies for  $CO<sub>2</sub>$  assimilation in Central Amazon trees. Braz J Bot 42:239–247. [https://doi.org/10.1007/](https://doi.org/10.1007/s40415-019-00533-2) [s40415-019-00533-2](https://doi.org/10.1007/s40415-019-00533-2)
- <span id="page-14-20"></span>Schansker G, Toth SZ, Strasser RJ (2006) Dark-recovery of the Chl a fuorescence transient (OJIP) after light adaptation: the qTcomponent of non-photochemical quenching is related to an activated photosystem I acceptor side. Biochim Biophys Acta 1757:787–797
- <span id="page-14-25"></span>Schansker G, Tóth SZ, Strasser RJ (2005) Methylviologen and dibromothymoquinone treatments of pea leaves reveal the role of photosystem I in the Chl a fuorescence rise OJIP. Biochimica et Biophysica Acta 1706:250–261
- <span id="page-14-3"></span>Searles PS, Flint SD, Caldwell MM (2001) A meta-analysis of plant feld studies simulating stratospheric ozone depletion. Oecologia 127:1–10
- <span id="page-14-12"></span>Shah T, Latif S, Saeed F, Ali I, Ullah S, Abdullah A, Jan S, Ahmad P (2020) Seed priming with titanium dioxide nanoparticles enhances seed vigor, leaf water status, and antioxidant enzyme activities in maize (*Zea mays* L.) under salinity stress. J King Saud Univ Sci.<https://doi.org/10.1016/j.jksus.2020.10.004>
- <span id="page-14-16"></span>Shine MB, Guruprasad KN (2012) Oxyradicals and PS II activity in maize leaves in the absence of UV components of the solar spectrum. J Biosci 37:703–712
- <span id="page-14-29"></span>Shine MB, Guruprasad KN, Anand A (2011) Enhancement of germination, growth, and photosynthesis in soybean by pretreatment of seeds with a magnetic feld. Bioelectromagnetics 32:474–484
- <span id="page-14-30"></span>Shine MB, Guruprasad KN, Anand A (2012) Effect of stationary magnetic feld strengths of 150 and 200 mT on reactive oxygen species production in soybean. Bioelectromagnetics 33:428–437
- <span id="page-14-1"></span>Singh S, Kumari R, Agrawal M, Agrawal SB (2012) Differential response of radish plants to supplemental ultraviolet-B radiation under varying NPK levels: chlorophyll fuorescence, gas exchange and antioxidants. Physiol Plant 145:474–484. [https://](https://doi.org/10.1111/j.1399-3054.2012.01589.x) [doi.org/10.1111/j.1399-3054.2012.01589.x](https://doi.org/10.1111/j.1399-3054.2012.01589.x)
- <span id="page-14-17"></span>Singh S, Agrawal M, Agrawal SB (2014) Impact of ultraviolet-B radiation on photosynthetic capacity, antioxidative potential and metabolites in *Solanum tuberosum* L. under varying levels of soil NPK. Acta Physiol Plant 36:1441–1453. [https://doi.org/10.](https://doi.org/10.1007/s11738-014-1522-z) [1007/s11738-014-1522-z](https://doi.org/10.1007/s11738-014-1522-z)
- <span id="page-14-32"></span>Snyrychova I, Kos PB, Hideg E (2007) Hydroxyl radicals are not the protagonists of UV-B induced damage in isolated thylakoid membranes. Funct Plant Biol 34:1112–1121
- <span id="page-14-26"></span>Stirbet A, Lazár D, Kromdijk J, Govindjee G (2018) Chlorophyll a fuorescence induction: can just a one-second measurement be used

to quantify abiotic stress responses? Photosynthetica 56:86–104. <https://doi.org/10.1007/s11099-018-0770-3>

- <span id="page-15-8"></span>Strasser RJ, Tsimilli-Micheal M, Srivastava A (2000) The fuorescence transient as a tool to characterize and screen photosynthetic samples. In: Yunus M, Pathre U, Mohnanty P (eds) Probing photosynthesis: mechanisms regulation and adaptation. Taylor and Francis, London, UK, pp 445–483
- <span id="page-15-1"></span>Suchar VA, Robberecht R (2015) Integration and scaling of UV-B radiation efects on plants: from DNA to leaf. Ecol Evol 5:2544–2555
- <span id="page-15-5"></span>Sytar O, Kumari P, Yadav S et al (2019) Phytohormone priming: regulator for heavy metal stress in plants. J Plant Growth Regul 38:739–752. <https://doi.org/10.1007/s00344-018-9886-8>
- <span id="page-15-15"></span>Sztatelman O, Grzy J, Gabrys H, Banas AK (2015) The effect of UV-B on Arabidopsis leaves depends on light conditions after treatment. BMC Plant Biol 15:1–16
- <span id="page-15-11"></span>Tiitto R, Nenadis N, Neugart S, Robson M, Agati G, Vepsäläinen J, Zipoli G, Nybakken L, Winkler B, Jansen AKM (2015) Assessing the response of plant favonoids to UV radiation: an overview of appropriate techniques. Phytochem Rev 14:273–297. [https://](https://doi.org/10.1007/s11101-014-9362-4) [doi.org/10.1007/s11101-014-9362-4](https://doi.org/10.1007/s11101-014-9362-4)
- <span id="page-15-6"></span>Thomas S, Anand A, Chinnusamy V, Dahuja A, Basu S (2013) Magnetopriming circumvents the efect of salinity stress on germination in chickpea seeds. Acta Physiol Plant 35:3401–3411. [https://doi.](https://doi.org/10.1007/s11738-013-1375-x) [org/10.1007/s11738-013-1375-x](https://doi.org/10.1007/s11738-013-1375-x)
- <span id="page-15-0"></span>Vanhaelewyn L, Prinsen E, Van der Straeten D, Vandenbussche F (2016) Hormone, controlled UV-B responses in plants. J Exp Bot 67(15):4469–4482
- <span id="page-15-7"></span>Wellburn A, Lichtenthaler H (1984) Formulae and program to determine total carotenoids and chlorophylls a and b of leaf extracts in diferent solvents. In: Advances in photosynthesis research. Springer, pp 9–12
- <span id="page-15-3"></span>Yu GH, Li W, Yuan ZY, Cui HY, Lv CG, Gao ZP, Han B, Gong YZ, Chen GX (2013) The effects of enhanced UV-B radiation on photosynthetic and biochemical activities in super high-yield hybrid rice Liangyoupeijiu at the reproductive stage. Photosynthetica 51:33–44
- <span id="page-15-17"></span>Zhang WJ, Björn LO (2009) The efect of ultraviolet radiation on the accumulation of medicinal compounds in plants. Fitoterapia 80:207–218. [https://doi.org/10.1016/j.ftote.2009.02.006](https://doi.org/10.1016/j.fitote.2009.02.006)
- <span id="page-15-4"></span>Zhang M, Dong JF, Jin HH, Sun LN, Xu MJ (2011) Ultraviolet-Binduced favonoid accumulation in *Betula pendula* leaves is

dependent upon nitrate reductase-mediated nitric oxide signaling. Tree Physiol 31:798–807. [https://doi.org/10.1093/treephys/](https://doi.org/10.1093/treephys/tpr070) [tpr070](https://doi.org/10.1093/treephys/tpr070)

- <span id="page-15-16"></span>Zhang M, An L, Feng P, Chen T, Chen K, Liu Y, Tang H, Chang J, Wang X (2003) The cascade mechanisms of nitric oxideas a second messenger of ultraviolet-B ininhibiting mesocotyl elongations. Photochem Photobiol 77:219–225
- <span id="page-15-10"></span>Zhao D, Reddy KR, Kakani VG, Reed J, Sullivan J (2003) Growth and physiological responses of cotton (*Gossypium hirsutum* L.) to elevated carbon dioxide and ultraviolet-B radiation under controlled environment conditions. Plant Cell Environ 26:771–782
- <span id="page-15-9"></span>Zhou B, Guo Z, Xing J, Huang B (2005) Nitric oxide is involved in abscisic acid-induced antioxidant activities in *Stylosanthes guianensis*. J Exp Bot 56:3223–3228
- <span id="page-15-13"></span>Živčák M, Olšovská K, Slamka P et al (2014a) Measurements of chlorophyll fuorescence in diferent leaf positions may detectnitrogen defciency in wheat. Zemdirb Agric 101:437–443
- <span id="page-15-14"></span>Živčák M, Olšovská K, Slamka P et al (2014b) Application of chlorophyll fuorescence performance indices to assess the wheat photosynthetic functions influenced by nitrogen deficiency. Plant Soil Environ 60:210–215
- Živčák M, Brestic M, Kunderlikova K, Sytar O, Allakhverdiev SI (2015) Repetitive light pulse-induced photoinhibition of photosystem I severely affects  $CO<sub>2</sub>$  assimilation and photoprotection in wheat leaves. Photosynth Res. [https://doi.org/10.1007/](https://doi.org/10.1007/s11120-015-0121-1) [s11120-015-0121-1](https://doi.org/10.1007/s11120-015-0121-1)
- <span id="page-15-12"></span>Zivcak M, Bruckova K, Sytar O, Brestic M, Olsovska K, Allakhverdiev SI (2017) Lettuce favonoids screening and phenotyping by chlorophyll fuorescence excitation ratio. Planta 245(6):1215–1229. <https://doi.org/10.1007/s00425-017-2676-x>
- <span id="page-15-2"></span>Zuk-Golaszewska K, Upadhyaya MK, Golaszewski J (2003) The efect of UV-B radiation on plant growth and development. Plant Soil Environ 49:135–140

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