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Effect of different light intensity on physiology, antioxidant capacity and photosynthetic characteristics on wheat seedlings under high CO₂ concentration in a closed artificial ecosystem

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Abstract

The growth of plants under high carbon dioxide (CO₂) concentrations (≥ 1000 ppm) is explored for the climate change and the bioregenerative life support system (BLSS) environment of long-duration space missions. Wheat (*Triticum aestivum* L.) is a grass cultivated for cereal grain—a global staple food including astronauts. Light and CO₂ are both indispensable conditions for wheat seedlings. This study provides insights on the physiology, antioxidant capacity and photosynthetic characteristics of wheat seedlings under a range of photosynthetic photon flux densities in a closed system simulating BLSS with high CO₂ concentration. We found that the F_v/F_m , F_v/F_0 , chlorophyll content, intrinsic water use efficiencies (WUE_i), membrane stability index (MSI), and malondialdehyde (MDA) of wheat seedlings grown under an intermediate light intensity of 600 µmol m⁻² s⁻¹ environment were all largest. Interestingly, the high light intensity of 1200 mol m⁻² s⁻¹ treatment group exhibits the highest net photosynthetic rate but the lowest MDA content. The stomatal conductance and F_0 of high light intensity of 1000 µmol m⁻² s⁻¹ treatment group were both significantly higher than that of other groups. Our study provides basic knowledge on the wheat growth in different environments, especially in a closed ecosystem with artificial lights.

Keywords Wheat \cdot High CO₂ concentration \cdot Light intensity \cdot Photosynthetic characteristics \cdot Physiology \cdot Antioxidant capacity

Abbreviations

HL	High light
IL	Intermediate light
LL	Low light
PPFD	Photosynthetic photon flux density
LED	Light-emitting diodes

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LP1	Lunar palace 1
BLSS	Bioregenerative life support system
MSI	Membrane stability index
RWC	Relative water content
POD	Peroxidase
MDA	Malondialdehyde
Α	Net photosynthetic rate
gs	Stomatal conductance
WUE _i	Intrinsic water use efficiencies
Chl a	Chlorophyll a
Chl b	Chlorophyll b
Chl $a + b$	Total chlorophyll content
PS II	Photosystem II
F_0	Minimum fluorescence
$F_{\rm m}$	Maximum fluorescence
$F_{\rm v}$	Variable fluorescence
$F_{\rm v}/F_{\rm m}$	Maximum quantum yield of PSII efficiency
$F_{\rm v}/F_0$	Potential activity of PS II
H_2O_2	Hydrogen peroxide

Introduction

Wheat (*Triticum aestivum* L.) is a globally important grain crop cultivated as a staple food, accounting for 20% of the calories consumed by human beings (Brenchley et al. 2012). It is also an important crop in bioregenerative life support system (BLSS) to provide food, oxygen, and necessary psychological well-being for spacecraft crews (Poughon et al. 2009; Wheeler 2017; Ushakova et al. 2018; Larrabee 2018).

Growth of wheat crops is affected by carbon dioxide (CO₂) level (Prasad et al. 2002). Increasing CO₂ concentration can improve photosynthesis in C₃ plants such as wheat via reduction of photorespiration which can increase wheat yield (Eshghizadeh et al. 2018). Due to the global warming effect, the atmospheric CO₂ concentrations have risen from about 280 to 370 ppm since 1800 (Prasad et al. 2002; Griggs and Noguer 2002) and may reach 500-900 ppm by the end of the century (Joos et al. 1999; Bloom et al. 2002). The CO_2 levels in closed systems such as growth chambers and greenhouses can readily exceed 1000 ppm (Wang et al. 2015). Research has shown that CO_2 concentration which was controlled between 500 and 5500 ppm in a ground-based comprehensive experimental facility of BLSS called Chinese Lunar Palace 1 (LP1) could reach to a relatively steady-state level of approximately 1000 ppm (Fu et al. 2016). Although terrestrial CO₂ concentrations are not expected to exceed 1000 ppm in near future, it is necessary to explore the growth of plants under high CO₂ concentration to meet the future climates and the needs of BLSS environments. Controlled environmental studies showed that when wheat was cultivated under low CO₂ concentration (210 ppm), increasing CO₂ concentration was favorable to wheat growth (Dong et al. 2014a). In particular, 1000 ppm CO₂ was more favorable for growth and development of wheat (Bender et al. 1999; Taub et al. 2008; Dong et al. 2016a, b). High CO₂ concentration (1500 ppm) in closed BLSS can make carbon cycle time (residence in atmosphere) much shorter than natural (350 ppm) (Nelson et al., 2003). Therefore, it is essential for us to understand and control (Leong et al. 2020) the growth of wheat seedlings in high CO₂ concentration environments although which are not ideal on earth.

Light source is also an indispensable condition for plant (Schneider et al. 2006). The change of light intensity has a significant impact on physiology of wheat, such as the time to ear emergence and tillering of wheat, the adaptability to changing environmental conditions and the yield (Evtushenko and Chekurov 2004; Page and Feller 2016). Artificial lightings such as light-emitting diodes (LED) facilitate energy conservation compared to traditional light sources, which is also used to increase plant productivity per unit of cultivation area (Dong et al. 2014a; Loconsole et al. 2019). In high CO_2 , higher growth rates may be obtainable as light intensity increases. If adequate energy inputs are available, the size of the food production component can be greatly reduced in a controlled environment life support system (Bugbee and Salisbury 1988).

In this study, we investigated the physiology, antioxidant capacity and photosynthetic characteristics of wheat (*Triticum aestivum* L.) seedlings under a range of photosynthetic photon flux densities (PPFD) in the controlled artificial closed ecosystem of Lunar Palace 1 (LP1), simulating BLSS. Spefically, we measured the plant growth and leaf water status by Membrane Stability Index (MSI) and relative water contents (RWC), respectively. We also understand the antioxidant capacity by measuring the contents of peroxidase, malondialdehyde, and proline in the plants. This study aims to reveal potential mechanisms for the physiological and developmental responses to different light intensities on wheat seedlings in a high CO_2 concentration environment.

Materials and methods

Plant material and cultivation conditions

Chinese Lunar Palace 1 (LP1) located at the Institute of Environmental Biology and Life Support Technology, Beihang University, Beijing, China, was used for plant cultivation in this study. It has one comprehensive cabin and two enclosed plant cabins (Fig. S1), with a total area of 160 m^2 and a volume of 500 m³. The comprehensive cabin hosts four bed cubicles, a bathroom, an animal-raising room, a common room, a storage room, and a waste-treatment room, which is the main activity area of spacecraft crew members. Each plant cabin has two divisions in which environmental conditions could be independently control, such as oxygen levels, water contents, and food production. Spring wheat plants (Triticum aestivum L. cv. 'Dwarf') were cultivated in vermiculite using a porous tube nutrient delivery system (PTNDS) with water supply on demand in plant cabin of LP1 (Wang et al. 2016; Dong et al. 2018). For this study, the wheat was planted at a density of 1200 seeds m⁻² (Fig. S1) in a plant cabin with continuous lighting (24/0 h light/ dark) and 1000 ppm CO₂. Temperature and relative humidity of the plant cabin were maintained at 21 ± 1.3 °C and $70 \pm 4.6\%$, respectively. Growth period of the wheat seedling was 20 days. The basic culture medium was the modified Hoagland nutrient solution (Hoagland and Arnon 1950) (Table S1).

Light intensity treatments

LEDs used for this study have uniform spectra of white (350-750 nm) and red (peak at 658 nm) light (W:R = 1:1) measured by a spectroradiometer (Avaspec-2048-UA, Avantes B.V., Netherlands) (Fig. S1). The LED light source was set at 70 cm on top of the wheat planting surface to ensure the unity of light intensity in the initial canopy of wheat. Wheat plants were exposed to 200, 400, 600, 800, 1000 and 1200 µmol m⁻² s⁻¹ PPFD, measured by a quantum sensor (Li-250A, Li-Cor, USA) (20 cm under the light). They were referred as low light (LL-200, 200 μ mol m⁻² s⁻¹; LL-400, 400 μ mol m⁻² s⁻¹), intermediate light (IL-600, 600 μ mol m⁻² s⁻¹; IL-800, 800 μ mol m⁻² s⁻¹) and high light (HL-1000, 1000 μ mol m⁻² s⁻¹; HL-1200, 1200 μ mol m⁻² s⁻¹) treatment groups, respectively. PPFD levels were measured every two days at the top of the plant canopy with the quantum sensor. Five samples of those wheat seedlings were randomly chosen for each measurement.

Straw height analyses

The height of wheat seedlings was measured every two days using a straight scale and vernier caliper. The straw height was estimated from the vermiculite surface to the tip of the last leaf. Five samples were randomly chosen for each plant height measurement.

Membrane Stability Index (MSI) measurements

The leaves' MSI of wheat seedlings was measured with a conductivity meter (HI8733, Hanna instruments, Italy). Samples were excised from the leaves of 10 wheat seedlings at the first leaf at the terminal bud for each treatment. The samples were divided into two commensurate parts (about 100 mg) and soaked in 10 ml double-distilled water. The first part was then heated at 40 °C for 30 min and conductivity C_1 was determined. The second part was heated at 100 °C for 10 min and conductivity C_2 was determined. The MSI was calculated as:

$$MSI = \left(1 - \frac{C_1}{C_2}\right) \times 100\%$$

Relative water content (RWC) measurements

To determine RWC, samples were excised from the leaves of 10 wheat seedlings at the first leaf at the terminal bud for each treatment during the wheat seedling period. About 0.5 g (m_1 : initial weight of the leaves) of fresh leaves were

soaked in double-distilled water at room temperature for 4 h and immediately weighed again as m_2 . The leaves were then dried at 65 °C for 48 h and weighed again as m_3 . RWC was calculated using the following equation:

$$RWC = \frac{m_1 - m_3}{m_2 - m_3} \times 100\%$$

Peroxidase (POD) activity determination

POD activity of wheat seedlings was analyzed following the change of absorbance at 470 nm due to guaiacol oxidation (Díaz et al. 2001). The POD activity was assayed for 3 min in a reaction solution composed of 0.15 ml of 4% (v/v) guaiacol, 0.15 ml of 1% (v/v) H_2O_2 , 2.66 ml of 0.1 M phosphate buffer (pH 7.0) and 40 µl of enzyme extract. Blank sample was made of the same mixture without enzyme extract.

Malondialdehyde (MDA) estimation

Based on the thiobarbituric acid reactive substances (TBARS), the accumulation of lipid peroxide in tissues was determined by estimating the content of MDA (Stewart and Bewley 1980). The supernatant was mixed with 10 ml of 0.1% trichloroacetic acid (TCA). Pestled homogenate was used to centrifuge at 4000 rpm for 10 min and 2 ml supernatant was added to 4 ml 5% thiobarbituric acid (TBA) in 20% TCA. The mixture was heated at 95 °C for 30 min, rapidly cooled on ice and centrifuged at 3000 rpm for 10 min. The absorbance of the supernatant was recorded at 532 nm. The value for non-specific absorption at 600 nm was subtracted. The MDA content was calculated with its extinction coefficient of 155 mM⁻¹ cm⁻¹.

Proline content extraction

Samples were excised from the leaves of ten wheat seedlings at the first leaf at the terminal bud for each treatment. Seedling leaves were rapidly frozen in liquid nitrogen, freezedried and stored until lab use. Proline content was extracted and the content analyzed spectrophotometrically according to the method of Bates et al. (1973).

Chlorophyll analysis

The contents of chlorophyll a (Chl a) and chlorophyll b (Chl b) were analyzed with an ultraviolet spectrophotometer (SP-75, Shanghai Spectrum Instruments Co., Ltd, Shanghai, China). Seedling leaves were frozen in liquid nitrogen and stored at -80 °C until measured.

Photosynthetic efficiency measurement

Photosynthetic characteristics were measured using a portable photosynthesis instrument (LI-6400XT, Li-Cor, USA). Leaf gas-exchange parameters including net photosynthetic rate (*A*) and stomatal conductance (gs) were determined from the first leaf in the seedling's terminal bud under saturating quantum flux densities (> 800 µmol m⁻² s⁻¹) at ambient temperature and relative humidity. Leaf temperatures were 24 ± 1.3 °C during the measurements. The intrinsic water use efficiencies (WUE_i) were calculated by dividing A by gs (Meloni et al. 2003).

Determination of chlorophyll fluorescence

Chlorophyll fluorescence was measured simultaneously with CO_2 gas exchange on the first leaves using a portable photosynthesis system (LI-6400XT, Li-Cor, USA). All measurements were carried out using the standard fluorescence leaf chamber (2 cm²). The seedling leaves were dark-adapted for 20 min prior to measurements. The minimum fluorescence (F_0) and maximum fluorescence (F_m) were obtained by the method described by Fu et al. (2012). The maximum quantum yield of PS II efficiency (F_v/F_m) and potential activity of PS II sss F_v/F_0) was calculated based on methods reported earlier (Maxwell and Johnson, 2000; Fu et al. 2012).

Statistics

All measurements were performed in triplicate and randomly arranged in each block. Data were subjected to oneway analysis of variance (ANOVA) using Statistica Software SPSS 20.0 (SPSS Inc., Chicago, Illinois). A significant level of 0.05 was used for all statistical tests by Tukey's multiple range test.

Results

The impact on plant height, light intensity on the canopy, RWC and MSI

Figure 1a shows the average plant height under different light intensities (i.e. 200, 400, 600, 800, 1000 and 1200 μ mol m⁻² s⁻¹) as a function of the light exposure time. The plant height increases with exposure time, for all light intensities under test. Although the plant treated by 800 μ mol m⁻² s⁻¹ is generally taller than others for the first 18 days and the light intensity on canopy greatly changes with the treatment time (Fig. 1b), all plants turn out to be of similar height after 23 days of growth regardless of the light intensities exposed to. Furthermore, the RWC and MSI contents of the plants do not vary much as the light intensity increases (Fig. 1c, d). Within the small variations, minimum values were both found in LL-400, while maximum values were found in LL-200 and IL-600, respectively.

Morphological characteristics antioxidant system

Figure 2a and b shows the plants' POD activity and MDA content as a function of light intensity. Among all groups, the POD activity of LL-400 scores the highest, which is nearly 74% and 36% larger than the LL-200 and HL-1200 treatment groups, respectively. The POD activity also fluctuates as the light intensity increases. For example, the POD activity of HL-1000 is higher than that of IL-800 and HL-1200. On the other hand, the MDA content exhibits a Gaussian distribution with the light intensity. In Fig. 2b, the MDA content initially increases with the light intensity and declines for light intensity larger than 600 μ mol m⁻² s⁻¹, with the lowest content (3.06 ± 0.03) observed in the HL-1200 group.

Photosynthetic characteristics

Responses of net photosynthetic rate (A), stomatal conductance (gs), intrinsic water use efficiency (WUE), Chl a, Chl b, and Chl (a + b) to different light intensities are plotted in Fig. 3. Compared to LL-200, the net photosynthetic rate (A) of LL-400, IL-600, IL-800, HL-1000 and HL-1200 is larger by 1.82, 3.19, 3.52, 3.48 and 4.12 times, respectively (Fig. 3a). The stomatal conductance (gs) of HL-1000 treatment group is 0.11, which is 18.1% and 15.5% higher than IL-800 and HL-1200 groups, respectively, and much larger than LL-200, LL-400 and IL-600 groups (Fig. 3b). Similarly, the WUE; exhibits large variations across different light intensities treatments. The WUE_i increases initially from LL-200 to IL-600, with a maximum value of $68.91 \pm 1.36 \ \mu mol \ CO_2 \ mol^{-1} \ H_2O^{-1}$ at IL-600, and then decreases with light intensity (Fig. 3c). It rises again at HL-1200 under high CO₂ concentration.

Chlorophyll fluorescence

The increase of minimal fluorescence (F_0) level is clearly visible in the HL-1000 group, which is 70.3 ± 4.8 and significantly higher than that of LL-200 and HL-1200 (Table 1). The stability of F_0 found among LL-400, IL-600 and IL-800 treatments indicates that IL caused negligible changes in the reaction centers. HL-1200 treatment does not impede the PS II reaction center under high CO₂ concentration owing to the reduced F_0 of HL-1200 (Table 1), which can be related to the inhibition of photorespiration. The trends of fluorescence (F_v) and maximum fluorescence (F_m) are similar to the F_0 (Table 1). When the light intensity was too high, both F_v and F_m reduces.



Fig. 1 The response of straw height (**a**) and light intensity (**b**) of wheat seedlings to different treatments; the values are means \pm SE (*n* = 10). The response of relative water content (RWC) (**c**) and membrane stability index (MSI) (**d**) of wheat seedlings to different treatmet.



ments; the values are means \pm SE (*n*=3). Different lowercase letters indicate significant differences among different light intensity treatments (*p* < 0.05)

Figure 4a shows that F_v/F_m of LL-400, HL-1000 and HL-1200 treatment groups is significantly lower than the IL-600 treatment group. Figure 4b clearly shows that the F_v/F_0 of the LL-400, HL-1000 and HL-1200 groups is much lower than the IL-600 group, by about 8.5%, 10.7% and 6.5%, respectively. The largest F_v/F_0 value is 5.029 in the IL-600 group. Meanwhile, the highest content of chlorophyll (Chl a, Chl b, Chl a + b) is in the IL-600 group (Fig. 3d–f). The decrease of high light intensity (HL-1000 and HL-1200) is observed in Fig. 4b, which has a similar condition to the change of chlorophyll content. Meanwhile, LL-400 has not enough light and chlorophyll content to sufficiently photosynthesize. Considering the chlorophyll fluorescence analysis performed in real growing conditions (Fig. 4c-f), the heat dissipative quantum ratio (F_0/F_m) and steady-state fluorescence yield (F_s) are invariant to the different growing

conditions. The maximum fluorescence yield (F'_m) and electron transport rate (ETR) exhibit similar trend, where the minimum values were both recorded in the LL-200 group. Meanwhile, the ETR is significantly higher in LL-1200.

Discussion

As a major scavenger of hydrogen peroxide (H_2O_2) , peroxidase (POD) catalyzes the dismutation of superoxide to H_2O_2 and oxygen (Fu et al. 2012). The level of POD reflects the status of physiological activity of plants and has the function of oxidase IAA. The growth of overground of wheat, especially for the elongation growth, could be promoted in a low level of POD (Dong et al. 2014c). Our findings show that the light intensity has a substantial impact in tuning



Fig. 2 The response of POD (**a**) and MDA (**b**) of wheat seedlings to different treatments. The values are means \pm SE (n=3). Different lowercase letters indicate significant differences among different light intensity treatments (p < 0.05)

the POD activity in wheat, similar to previous reports in romaine lettuce (Lactuca sativa L.), Phalaenopsis orchids, and Lithocarpus litseifolius (Hance) Chun. (Ali et al. 2005; Fu et al. 2012; Li et al. 2016). The POD activity of our lowlight intensity group (LL-400) scores the highest. This indicates that excess energy light could damage photosynthetic organism and reactive oxygen species (ROS), although the photosynthetic capacity is stronger in high light intensity samples (Li et al. 2016). With the increase of light intensity, the activity of POD does not vary much, which is mainly affected by the genetic characteristics of the variety itself (Ashraf and Harris 2004). The increment of ROS causes membrane lipid peroxidation, leading to the damage of cell membranes (Li et al. 2016) and the production of MDA. The MDA content measures degree of damage and stress in plant (Abdollahi et al. 2004; Dong et al. 2014a), and which formation is associated with the POD in leaves (Dong et al. 2014a, b). Previous work demonstrated that diverse responses of antioxidant enzymes to light-stressed plantlets, implying that oxidative stress might be one of the major component of light stress on *Phalaenopsis* (Ali et al. 2005). In our study, we speculate that the photorespiration was restrained by high CO₂ concentration from LL-600 to HL-1000, so that plants can efficiently utilize light energy for photosynthesis and ETR. Meanwhile, the damage caused by ROS reduces significantly. As a result of the reduction in POD activity from LL-600 to HL-1000 groups, more and more ROS was accumulated, damaging the chloroplast envelope and increasing the MDA content (Li et al. 2016).

Our net photosynthetic rate (A) results agree well with previous findings that the stronger the light intensity, the higher the net photosynthetic rate (Dong et al. 2014a, b, c). The A of the HL-1200 is significantly greater than others, indicating possibly that the former has a wide and long wheat leaf area (Jiang et al. 2003). Unlike the previous work, the differences observed between IL-800 and HL-1000 treatment groups are not significant (Fig. 3a), which could be due to the fact that Chl a, Chl b, and Chl (a+b) are not statistically significant between IL-800 and HL-100 (Fig. 3d-f). To a certain extent, Chlorophyll content determines the photosynthetic rate, and the change of chlorophyll content is closely related to the attenuation of A (Kura-Hotta et al. 1987; Spano et al. 2003). It has already been proposed that gs increases with light intensity (Wang et al. 2017; Xiong et al., 2018). In this study, the max gs was found in HL-1000 level because plants reduce their stomatal conductance to resist strong light (Dong et al. 2014a). However, the gs of IL and HL groups is significantly larger than that of LL. Lowlight intensity may decrease the gs to CO₂, thus inhibiting photosynthesis (Loconsole et al. 2019). Zhang et al. (2017) and Li et al. (2015) also reported that gs varies linearly with A. The intrinsic water use efficiency (WUE_i), an index of potential resource use efficiency was used to facilitate the comparisons among leaves under similar conditions (Dong et al. 2016a). Our results prove that the WUE; of different light intensities treatments has significant differences. This is consistent with earlier reports (Zhang et al. 2016). However, significant changes are not observed between IL-800 and HL-1200 treatment groups, indicating that the light intensity is excessive for the plant needs (Loconsole et al. 2019), due to the change in gs. At the same time, it can also be seen that water-use efficiency does not solely depend on the amount of water intake of plants, but is a comprehensive evaluation index of resource utilization efficiency that represents CO₂ concentration, humidity, and temperature or light in the environment (Dong et al. 2014a, 2016a; Medrano et al.



Fig. 3 The response of net photosynthetic rate (**a**), stomatal conductance (**b**) and intrinsic water efficiency (**c**) of wheat seedlings to different treatments; the values are means \pm SE (*n*=10). The response of chlorophyll a (**d**) and chlorophyll b (**e**) and chlorophyll a + b (**f**) of

wheat seedlings to different light intensity treatments; the values are means \pm SE (*n*=6). Different lowercase letters indicate significant differences among different light intensity treatments (*p* < 0.05)

Light intensity F_0 $F_{\rm m}$ F_{v} $(\mu mol m^{-2} s^{-1})$ $48.8 \pm 2.9a$ 200 $280.9 \pm 16.1a$ $232.1 \pm 13.3a$ 400 $57.4 \pm 4.8 ab$ 318.3 ± 21.8ab 260.9±17.1ab 600 63.1±1.0ab $380.3 \pm 5.7b$ $317.1 \pm 5.2b$ 800 $58.9 \pm 3.2ab$ 340.9 ± 16.3 ab 282.0±13.2ab 1000 $70.3 \pm 4.8b$ $386.8 \pm 26.7b$ $316.5 \pm 22.2b$ 1200 $53.8 \pm 3.4a$ $333.9 \pm 9.2a$ $242.3 \pm 17.1a$

Table 1 The response of potential activity of F_0 , F_m and F_v of wheat seedlings to different treatments

The values are means \pm SE (*n*=9). Different lowercase letters indicate significant differences among different light intensity treatments (*p* < 0.05)

2015). Lighting system with different light intensities gives different responses to chlorophyll synthesis and chlorophyll degradation in wheat seedling leaves (Dong et al. 2014a). The chlorophyll content of wheat leaves was found to be increased quickly in high CO₂ concentrations (700 ppm) (Gutiérrez et al. 2013). Chlorophyll concentrations increase when the light intensity rises from 210 to 420 μ mol m⁻² s⁻¹ during seedling stage (Dong et al. 2014a), similar to our observations. The decline of Chlorophyll concentrations under high light intensity observed in our study can be attributed to three reasons. First, the response of chlorophyll to light intensity is species-dependent. Second, the light intensity and CO₂ concentration both have impacts on the chlorophyll content, and their function was mutual and indivisible. Third, high CO₂ concentrations induce sugar accumulation and decrease nitrogen content, causing senescence and the loss of chlorophyll in leaves (Lotfiomran et al. 2016; Agüera and Haba 2018).

The change of fluorescence (F_0) level observed in this work is supported by a previous study (Li et al. 2015). Increased F_0 represents an impaired energy trapping efficiency in PS II reaction centers because of the stronger light intensity (Paunov et al. 2018). Decreased efficiency of the energy transfer from the antenna to the reaction center was demonstrated by Somersalo and Krause (1990). However, we found that HL-1200 treatment did not cause the destruction of the PS II reaction center under high CO₂ concentration. This may be related to the reduced F_0 of HL-1200 and the inhibition of photorespiration. To a certain extent, the high light intensity can cause them to rise, which means the reduction of electron accepting plastoquinone (Q_A) is reduced under low light intensity and super high light intensity. In brief, the increase in light intensity can increase the urgency of Q_A (Van Eerden et al. 2017). F_v/F_m reflects the original light energy efficiency of PS II (Li et al. 2015). Under non-environmental stress conditions, this parameter rarely changes and it is not affected by species and growth conditions. It is always used as a good indicator to reflect the degree of light inhibition (Wang et al. 2015; Mathur et al. 2016). According to Björkman and Demmig (1987), the optimal value of F_v/F_m was around 0.83 for healthy plants, and if F_v/F_m was higher than 0.8, the potential efficiency of PS II was not affected. In the present study, the largest $F_{\rm v}/F_{\rm m}$ value was 0.834 in IL-600 treatment, which indicates that the plant is healthy and not suffering from photosynthetic stress (Li et al. 2015). The $F_{\rm v}/F_{\rm m}$ value decreased with increasing light intensity. Wang et al. (2015) also reported that F_v/F_m also decreased significantly with super-elevated CO₂ concentration in soybean and maize leaves. The results showed that F_v/F_m of HL treatment groups were significantly lower than ILtreatment group in the range (0.817–0.821, higher than 0.8), which might indicate that the photosynthetic stress was prevented under high CO₂ concentration. $F_{\rm v}/F_0$ indicates the potential activity or the size and number of PS II in the photoreaction center in the chloroplast, which is always used to represent photosynthetic strength of the plant (Li et al. 2015). The change F_y/F_0 was similar to F_v/F_m in our study. The results strongly suggest that the original light energy efficiency of PS II and potential activity of PS II in the photoreaction center were not inhibited under IL-600, but they were significantly inhibited under LL-400, HL-1000 and HL-1200, relatively. However, the ETR was significantly higher in LL-1200, further indicating that higher light intensity caused a loss in the performance of the PSII under high CO₂ concentration. Similar results were found in different light intensities under normal atmospheric (Loconsole et al. 2019).

Conclusion

In this study, wheat seedlings were cultivated under different light intensities (200–1200 μ mol m⁻² s⁻¹) in high carbon dioxide concentration, and characterized by appropriate morphological characteristics, antioxidant capacity, photosynthesis parameters and chlorophyll fluorescence parameters. Our results show that wheats planted under 600 μ mol m⁻² s⁻¹ exhibit the best characteristics in terms of F_v/F_m , F_v/F_0 , chlorophyll content, WUE_i and MSI. Hence, 600 μ mol m⁻² s⁻¹ is the most suitable light intensity for wheat seedlings growth under high carbon dioxide concentration. Interestingly, the net photosynthetic rate always increased, however, the content of MDA increased first and then decreased gradually with increasing light intensity. The stomatal conductance and F_0 of HL-1000 treatment group were both significantly higher than that of other groups, and the WUE; also showed a trend of increasing first and then decreasing with the increasing light intensity. Our study provides insights on the role of light intensity on wheat growth and serves as a foundation to understand and predict the



Fig. 4 The chlorophyll fluorescence indexes measured on leaves of wheat seedlings to different light intensity treatments. The maximum quantum efficiency of PS II (F_v/F_m) (**a**), the potential activity of PS II (F_v/F_0) (**b**), the heat dissipative quantum ratio (F_0/F_m) (**c**), the steady-

state fluorescence yield (F_s) (**d**), the maximum fluorescence (F'_m) (**e**), and the electron transport rate (ETR) (**f**). The values are means ±SE (n=9). Different lowercase letters indicate significant differences among different light intensity treatments (p < 0.05)

yield of wheat growths in different environments, especially in a closed ecosystem.

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Compliance with Ethical Standards

Conflict of interest The authors declare that they have no conflict of interest.

References

- Abdollahi M, Ranjbar A, Shadnia S et al (2004) Pesticides and oxidative stress: a review. Medical Sci Monit 10(6):R141–R147
- Agüera E, De la Haba P (2018) Leaf senescence in response to elevated atmospheric CO_2 concentration and low nitrogen supply. Biol Plant 62(3):401–408
- Ali MB, Hahn EJ, Paek KY (2005) Effects of light intensities on antioxidant enzymes and malondialdehyde content during short-term acclimatization on micropropagated Phalaenopsis plantlet. Environ Exp Bot 54(2):109–120
- Ashraf M, Harris PJC (2004) Potential biochemical indicators of salinity tolerance in plants. Plant Sci 166(1):3–16
- Baresel JP, Rischbeck P, Hu Y et al (2017) Use of a digital camera as alternative method for non-destructive detection of the leaf chlorophyll content and the nitrogen nutrition status in wheat. Comput Electron Agric 140:25–33
- Barna M, Bosela M (2015) Tree species diversity change in natural regeneration of a beech forest under different management. For Ecol Manag 342:93–102
- Bates LS, Waldren RP, Teare ID (1973) Rapid determination of free proline for water-stress studies. Plant Soil 39(1):205–207
- Bender J, Hertstein U, Black CR (1999) Growth and yield responses of spring wheat to increasing carbon dioxide, ozone and physiological stresses: a statistical analysis of 'ESPACE-wheat' results. Eur J Agron 10(3–4):185–195
- Biederman LA, Harpole WS (2013) Biochar and its effects on plant productivity and nutrient cycling: a meta-analysis. GCB Bioenergy 5(2):202–214
- Björkman O, Demmig B (1987) Photon yield of O_2 evolution and chlorophyll fluorescence characteristics at 77 K among vascular plants of diverse origins. Planta 170(4):489–504
- Bloom AJ, Smart DR, Nguyen DT et al (2002) Nitrogen assimilation and growth of wheat under elevated carbon dioxide. Proc Natl Acad Sci USA 99(3):1730–1735
- Blum A, Ebercon A (1981) Cell membrane stability as a measure of drought and heat tolerance in wheat 1. Crop Sci 21(1):43–47
- Brenchley R, Spannagl M, Pfeifer M et al (2012) Analysis of the bread wheat genome using whole-genome shotgun sequencing. Nature 491(7426):705–710
- Bugbee BG, Salisbury FB (1988) Exploring the limits of crop productivity: I. Photosynthetic efficiency of wheat in high irradiance environments. Plant Physiol 88(3):869–878
- Croft H, Chen JM, Luo X et al (2017) Leaf chlorophyll content as a proxy for leaf photosynthetic capacity. Glob Change Biol 23(9):3513–3524
- Díaz J, Bernal A, Pomar F et al (2001) Induction of shikimate dehydrogenase and peroxidase in pepper (*Capsicum annuum* L.)

seedlings in response to copper stress and its relation to lignification. Plant Sci 161(1):179–188

- Dong C, Fu Y, Liu G, Liu H (2014a) Low light intensity effects on the growth, photosynthetic characteristics, antioxidant capacity, yield and quality of wheat (*Triticum aestivum* L.) at different growth stages in BLSS. Adv Space Res 53(11):1557–1566
- Dong C, Hu D, Fu Y et al (2014b) Analysis and optimization of the effect of light and nutrient solution on wheat growth and development using an inverse system model strategy. Comput Electron Agric 109:221–231
- Dong C, Fu Y, Liu G et al (2014c) Growth, photosynthetic characteristics, antioxidant capacity and biomass yield and quality of wheat (*Triticum aestivum* L.) exposed to LED light sources with different spectra combinations. J Agron Crop Sci 200(3):219–230
- Dong C, Shao L, Wang M, Liu G, Liu H, Xie B, Li B, Fu Y, Liu H (2016a) Wheat carbon dioxide responses in space simulations conducted at the Chinese Lunar Palace-1. Agron J 108(1):32–38
- Dong C, Liu G, Fu Y et al (2016b) Twin studies in Chinese closed controlled ecosystem with humans: The effect of elevated CO_2 disturbance on gas exchange characteristics. Ecol Eng 91:126–130
- Dong C, Chu Z, Wang M et al (2018) Influence of nitrogen source and concentrations on wheat growth and production inside "Lunar Palace-1". Acta Astronaut 144:371–379
- Dutta SS, Tyagi W, Rai M (2017) Physiological and molecular response to low light intensity in rice: a review. Agric Rev 38(3):209–215
- Embiale A, Hussein M, Husen A et al (2016) Differential sensitivity of *Pisum sativum* L. cultivars to water-deficit stress: changes in growth, water status, chlorophyll fluorescence and gas exchange attributes. J Agron 15(2):45–57
- Eshghizadeh HR, Zahedi M, Mohammadi S (2018) Differential growth responses of wheat seedlings to elevated CO₂. Not Sci Biol 10(3):400–409
- Evtushenko EV, Chekurov VM (2004) Inheritance of the light intensity response in spring cultivars of common wheat. Hereditas 141(3):288–292
- Fan XX, Xu ZG, Liu XY et al (2013) Effects of light intensity on the growth and leaf development of young tomato plants grown under a combination of red and blue light. Sci Hortic 153:50–55
- Freeman KW, Girma K, Arnall DB et al (2007) By-plant prediction of corn forage biomass and nitrogen uptake at various growth stages using remote sensing and plant height. Agron J 99(2):530–536
- Fu W, Li P, Wu Y (2012) Effects of different light intensities on chlorophyll fluorescence characteristics and yield in lettuce. Sci Hortic 135(135):45–51
- Fu Y, Li L, Xie B et al (2016) How to establish a bioregenerative life support system for long-term crewed missions to the moon or Mars. Astrobiology 16(12):925–936
- Griggs DJ, Noguer M (2002) Climate change 2001: the scientific basis. Contribution of working group I to the third assessment report of the intergovernmental panel on climate change. Weather 57(8):267–269
- Gutiérrez D, Morcuende R, Del Pozo A et al (2013) Involvement of nitrogen and cytokinins in photosynthetic acclimation to elevated CO₂ of spring wheat. J Plant Physiol 170(15):1337–1343
- Hoagland DR, Arnon DI (1950) The water-culture method for growing plants without soil. Calif Agric Exp Stn Circ 347(5406):357–359
- Jiang GM, Sun JZ, Liu HQ et al (2003) Changes in the rate of photosynthesis accompanying the yield increase in wheat cultivars released in the past 50 years. J Plant Res 116(5):347–354
- Joos F, Plattner GK, Stocker TF et al (1999) Global warming and marine carbon cycle feedbacks on future atmospheric CO₂. Science 284(5413):464–467
- Kreslavski V, Tatarinzev N, Shabnova N et al (2008) Characterization of the nature of photosynthetic recovery of wheat seedlings from short-term dark heat exposures and analysis of the

mode of acclimation to different light intensities. J Plant Physiol 165(15):1592–1600

- Kura-Hotta M, Satoh K, Katoh S (1987) Relationship between photosynthesis and chlorophyll content during leaf senescence of rice seedlings. Plant Cell Physiol 28(7):1321–1329
- Larrabee DA (2018) Climate change and conflicting future visions. Zygon 53(2):515–544
- Leong WH, Teh SY, Hossain MM et al (2020) Application, monitoring and adverse effects in pesticide use: the importance of reinforcement of good agricultural practices (GAPs). J Environ Manag 260:109987
- Li S, Yang W, Yang T et al (2015) Effects of cadmium stress on leaf chlorophyll fluorescence and photosynthesis of *Elsholtzia argyi* a cadmium accumulating plant. Int J Phytoremediat 17(1):85–92
- Li A, Li S, Wu X et al (2016) Effect of light intensity on leaf photosynthetic characteristics and accumulation of flavonoids in Lithocarpus litseifolius (Hance) Chun. (Fagaceae). Open J For 6:445–459
- Liu Q, Xiu WU, Chen B et al (2014) Effects of low light on agronomic and physiological characteristics of rice including grain yield and quality. Rice Sci 21(5):243–251
- Liu G, Bollier D, Gübeli C et al (2018) Simulated microgravity and the antagonistic influence of strigolactone on plant nutrient uptake in low nutrient conditions. npj Microgravity 4(1):20
- Loconsole D, Cocetta G, Santoro P et al (2019) Optimization of LED lighting and quality evaluation of Romaine lettuce grown in an innovative indoor cultivation system. Sustainability 11(3):841
- Lotfiomran N, Köhl M, Fromm J (2016) Interaction effect between elevated CO₂ and fertilization on biomass, gas exchange and C/N ratio of European beech (*Fagus sylvatica* L.). Plants 5(3):38
- Mackinney G (1941) Absorption of light by chlorophyll solutions. J Biol Chem 140(2):315–322
- Mathur S, Kalaji HM, Jajoo A (2016) Investigation of deleterious effects of chromium phytotoxicity and photosynthesis in wheat plant. Photosynthetica 54(2):185–192
- Maxwell K, Johnson GN (2000) Chlorophyll fluorescence—a practical guide. J Exp Bot 51(345):659–668
- Medrano H, Tomás M, Martorell S et al (2015) From leaf to wholeplant water use efficiency (WUE) in complex canopies: limitations of leaf WUE as a selection target. Crop J 3(3):220–228
- Meloni DA, Oliva MA, Martinez CA et al (2003) Photosynthesis and activity of superoxide dismutase, peroxidase and glutathione reductase in cotton under salt stress. Environ Exp Bot 49(1):69–76
- Monostori I, Heilmann M, Kocsy G et al (2018) LED lightingmodification of growth, metabolism, yield and flour composition in wheat by spectral quality and intensity. Front Plant Sci 9(605):1–16
- Monson RK, Rawsthorne S (2000) CO2 assimilation in C3–C4 intermediate plants. In: Leegood R, Sharkey TD, Caemmerer SC (eds) Photosynthesis. Springer, Dordrecht, pp 533–550
- Muneer S, Kim E, Park J et al (2014) Influence of green, red and blue light emitting diodes on multiprotein complex proteins and photosynthetic activity under different light intensities in lettuce leaves (*Lactuca sativa* L.). Int J Mol Sci 15(3):4657–4670
- Nam MH, Heo EJ, Kim JY et al (2003) Proteome analysis of the responses of Panax ginseng CA Meyer leaves to high light: use of electrospray ionization quadrupole-time of flight mass spectrometry and expressed sequence tag data. Proteomics 3(12):2351–2367
- Nelson M, Allen J, Ailing A et al (2003) Earth applications of closed ecological systems: relevance to the development of sustainability in our global biosphere. Adv Space Res 31(7):1649–1655
- Ommen OE, Donnelly A, Vanhoutvin S et al (1999) Chlorophyll content of spring wheat flag leaves grown under elevated CO₂ concentrations and other environmental stresses within the 'ESPACEwheat' project. Eur J Agron 10(3–4):197–203
- Page V, Feller U (2016) Light intensity selectively influences the distribution and further redistribution of macro-and micronutrients

in hydroponically grown wheat (*Triticum aestivum* L.). J Plant Nutr 39(3):428–437

- Pan JQ, Guo BL (2016) Effects of light intensity on the growth, photosynthetic characteristics, and flavonoid content of epimedium pseudowushanense B.L. Guo. Molecules 21(11):1475
- Paunov M, Koleva L, Vassilev A et al (2018) Effects of different metals on photosynthesis: Cadmium and zinc affect chlorophyll fluorescence in Durum Wheat. Int J Mol Sci 19(3):787
- Poorter H, Nagel O (2000) The role of biomass allocation in the growth response of plants to different levels of light, CO₂, nutrients and water: a quantitative review. Aust J Plant Physiol 27(6):595–607
- Poughon L, Farges B, Dussap CG et al (2009) Simulation of the MELiSSA closed loop system as a tool to define its integration strategy. Adv Space Res 44(12):1392–1403
- Prasad PVV, Boote KJ, Allen LH Jr et al (2002) Effects of elevated temperature and carbon dioxide on seed-set and yield of kidney bean (*Phaseolus vulgaris* L.). Glob Change Biol 8(8):710–721
- Sairam RK, Srivastava GC (2010) Water stress tolerance of wheat (*Triticum aestivum* L.): variations in hydrogen peroxide accumulation and antioxidant activity in tolerant and susceptible genotypes. J Agron Crop Sci 186(1):63–70
- Schneider S, Ziegler C, Melzer A (2006) Growth towards light as an adaptation to high light conditions in Chara branches. New Phytol 172(1):83–91
- Sinha RK (2004) Photosynthesis. In: Sinha RK (ed) Modern Plant Physiology. CRC Press, Boca Raton, pp 176–255
- Somersalo S, Krause GH (1990) Photoinhibition at chilling temperatures and effects of freezing stress on cold acclimated spinach leaves in the field: a fluorescence study. Physiol Plant 79(4):617–622
- Spano G, Di Fonzo N, Perrotta C et al (2003) Physiological characterization of 'stay green'mutants in durum wheat. J Exp Bot 54(386):1415–1420
- Stewart RRC, Bewley JD (1980) Lipid peroxidation associated with accelerated aging of soybean axes. Plant Physiol 65(2):245–248
- Stutte GW, Mackowiak CL, Yorio NC et al (1999) Theoretical and practical considerations of staggered crop production in a BLSS. Life Support Biosph Sci 6(4):287–291
- Taub DR, Miller B, Allen H (2008) Effects of elevated CO_2 on the protein concentration of food crops: a meta-analysis. Glob Change Biol 14(3):565–575
- Tikhomirov AA, Ushakova SA, Gribovskaya IA et al (2003) Light intensity and production parameters of phytocenoses cultivated on soil-like substrate under controled environment conditions. Adv Space Res 31(7):1775–1780
- Tilly N, Hoffmeister D, Cao Q et al (2014) Multitemporal crop surface models: accurate plant height measurement and biomass estimation with terrestrial laser scanning in paddy rice. J Appl Remote Sens 8(1):083671
- Ushakova S, Tikhomirova N, Velichko V et al (2018) Analysis of the gas exchange and water balance in a closed experimental model of the artificial ecosystem intended for an estimated portion of a human. Acta Astronaut 152:105–111
- Van Eerden FJ, Melo MN, Frederix PWJM et al (2017) Exchange pathways of plastoquinone and plastoquinol in the photosystem II complex. Nat Commun 8:15214
- Walker BJ, Drewry DT, Slattery RA et al (2018) Chlorophyll can be reduced in crop canopies with little penalty to photosynthesis. Plant Physiol 176(2):1215–1232
- Wang M, Xie B, Fu Y et al (2015) Effects of different elevated CO_2 concentrations on chlorophyll contents, gas exchange, water use efficiency, and PSII activity on C₃ and C₄ cereal crops in a closed artificial ecosystem. Photosynth Res 126(2–3):351–362
- Wang M, Liu H, Dong C et al (2016) Elevated CO₂ enhances photosynthetic efficiency, ion uptake and antioxidant activity of *Gynura*

bicolor DC. Grown in a porous tube nutrient delivery system under simulated microgravity. Plant Biol 18(3):391–399

- Wang N, Gao J, Zhang S (2017) Overcompensation or limitation to photosynthesis and root hydraulic conductance altered by rehydration in seedlings of sorghum and maize. Crop J 5(4):337–344
- Ware MA, Belgio E, Ruban AV (2015) Photoprotective capacity of non-photochemical quenching in plants acclimated to different light intensities. Photosynth Res 126(2–3):261–274
- Weatherley PE (2006) Studies in the water relations of the cotton plant. New Phytol 49(1):36–51
- Weiguo F, Pingping L, Yanyou W et al (2012) Effects of different light intensities on anti-oxidative enzyme activity, quality and biomass in lettuce. Hortic Sci 39(3):129–134
- Wheeler RM (2017) Agriculture for space: people and places paving the way. Open Agric 2(1):14–32
- Xiong D, Douthe C, Flexas J (2018) Differential coordination of stomatal conductance, mesophyll conductance, and leaf hydraulic conductance in response to changing light across species. Plant Cell Environ 41(2):436–450

- Yi Z, Liu H, Fu Y, et al (2018) Study on the application effect of cold tolerant biocontrol agent *Pseudochrobactrum kiredjianiae* A4 in BLSS. In: Proceedings of the 42nd COSPAR Scientific Assembly, p 42
- Zhang M, Cao T, Ni L et al (2010) Carbon, nitrogen and antioxidant enzyme responses of Potamogeton crispus to both low light and high nutrient stresses. Environ Exp Bot 68(1):44–50
- Zhang J, Liu J, Yang C et al (2016) Photosynthetic performance of soybean plants to water deficit under high and low light intensity. S Afr J Bot 105:279–287
- Zhang YJ, Sack L, Cao KF et al (2017) Speed versus endurance tradeoff in plants: leaves with higher photosynthetic rates show stronger seasonal declines. Sci Rep 7(42085):1–9

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