

pK_a of ubiquinone, menaquinone, phylloquinone, plastoquinone, and rholoquinone in aqueous solution

Ryo Hasegawa¹ · Keisuke Saito^{1,2} · Tomohiro Takaoka¹ · Hiroshi Ishikita^{1,2} 

Received: 13 December 2016 / Accepted: 3 April 2017 / Published online: 12 April 2017
© The Author(s) 2017. This article is an open access publication

Abstract Quinones can accept two electrons and two protons, and are involved in electron transfer and proton transfer reactions in photosynthetic reaction centers. To date, the pK_a of these quinones in aqueous solution have not been reported. We calculated the pK_a of the initial protonation (Q^- to QH) and the second protonation (QH^- to QH_2) of 1,4-quinones using a quantum chemical approach. The calculated energy differences of the protonation reactions Q^- to QH and QH^- to QH_2 in the aqueous phase for nine 1,4-quinones were highly correlated with the experimentally measured $pK_a(Q^-/QH)$ and $pK_a(QH^-/QH_2)$, respectively. In the present study, we report the $pK_a(Q^-/QH)$ and $pK_a(QH^-/QH_2)$ of ubiquinone, menaquinone, phylloquinone, plastoquinone, and rholoquinone in aqueous solution.

Keywords Photosystem II · Bacterial photosynthetic reaction centers · *Rhodobacter sphaeroides* · *Blastochloris viridis* · Green non-sulfur bacteria · Evolutionary transition

Introduction

Quinones can accept two electrons and two protons via the initial protonation of semiquinone, Q^- to QH , and second protonation of hydroquinone, QH^- to QH_2 . Ubiquinone serves as an electron acceptor at the Q_A and Q_B binding sites in reaction centers of purple bacteria (PbRC) from *Rhodobacter sphaeroides* (Fig. 1). Similarly, menaquinone (vitamin K_2) is the acceptor at the Q_A site in PbRC from *Blastochloris viridis*, while phylloquinone (vitamin K_1) is the active center at the A_{1A} and A_{1B} sites in photosystem I (PSI). In reaction centers of green non-sulfur bacteria from *Chloroflexus aurantiacus*, menaquinones are also located at both the Q_A and Q_B sites (Hale et al. 1983). It should be noted that phylloquinone and menaquinone have the same head-group structure (Fig. 2). Plastoquinone serves as an electron acceptor at the Q_A and Q_B sites in photosystem II (PSII) (Fig. 1) (Robinson and Crofts 1984; Rutherford et al. 1984; Okamura et al. 2000; Brettel and Leibl 2001; Wraight 2004). Rholoquinone is a required cofactor for anaerobic respiration in *Rhodospirillum rubrum* (Okayama et al. 1968). Because rholoquinone is assumed to have a higher $pK_a(Q^-/QH)$ than ubiquinone, rholoquinone-substituted PbRC has been used to investigate the mechanism of proton transfer to Q_B (e.g., Graige et al. 1999; Maroti et al. 2015).

In PbRC and PSII, Q_A/Q_A^- acts as a one-electron redox couple and donates an electron to the second quinone Q_B , without undergoing protonation itself. In contrast, Q_B reduction involves two consecutive one-electron reduction reactions with a series of associated proton uptake reactions (reviewed in Diner and Rappaport 2002; Renger and Renger 2008; Holzwarth 2008; Cardona et al. 2012; Muh et al. 2012; Petrouleas and Crofts 2005). Both Q_A and Q_B are located near the non-heme Fe^{2+} and the

Electronic supplementary material The online version of this article (doi:10.1007/s11120-017-0382-y) contains supplementary material, which is available to authorized users.

✉ Hiroshi Ishikita
hiro@appchem.t.u-tokyo.ac.jp

¹ Department of Applied Chemistry, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8654, Japan

² Research Center for Advanced Science and Technology, The University of Tokyo, 4-6-1 Komaba, Meguro-ku, Tokyo 153-8904, Japan

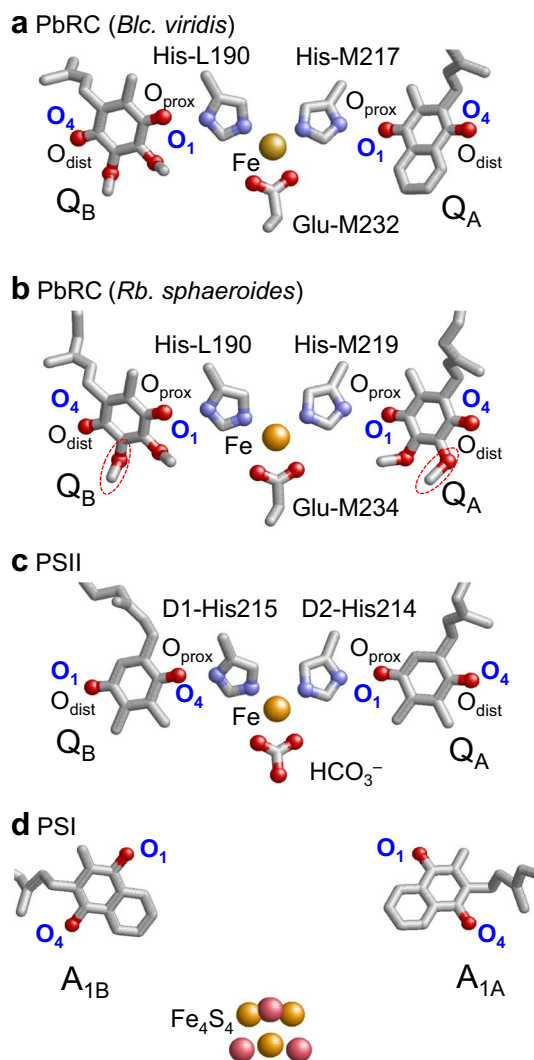


Fig. 1 Quinones in photosynthetic reaction centers: **a** menaquinone as Q_A and ubiquinone as Q_B in bacterial photosynthetic reaction centers from *Blastochloris viridis* (*Blc. viridis*, PDB ID: 2I5N) (Li et al. 2006); **b** ubiquinone as Q_A and Q_B in bacterial photosynthetic reaction centers from *Rhodobacter sphaeroides* (*Rb. sphaeroides*, PDB ID: 3I4D), where the 2-methoxy group of each ubiquinone is red circled; **c** plastoquinone as Q_A and Q_B in PSII (PDB ID: 3ARC) (Umena et al. 2011); and **d** phylloquinone as A_{1A} and A_{1B} in PSI (PDB ID: 1JB0) (Jordan et al. 2001). Red and blue balls indicate O and N atoms, respectively. In PbRC and PSII, O_{prox} and O_{dist} stand for O atoms of the quinones at the proximal and distal positions with respect to the non-heme Fe complex, respectively. Note that except for Q_B in PSII, O_{prox} is O1 and O_{dist} is O4 in PbRC and PSII

ligands to the Fe²⁺ (i.e., His-L190 and His-M217 (or M219) in PbRC and D1-His215 and D2-His214 in PSII) donate an H-bond to the carbonyl O atoms of quinones that are nearer to the Fe complex (O_{prox}) (Fig. 1a–c). The carbonyl O atoms of quinones at the distal position (O_{dist}) also form H-bonds with the proteins. On the other hand, in PSI, the non-heme Fe²⁺ is absent, but the

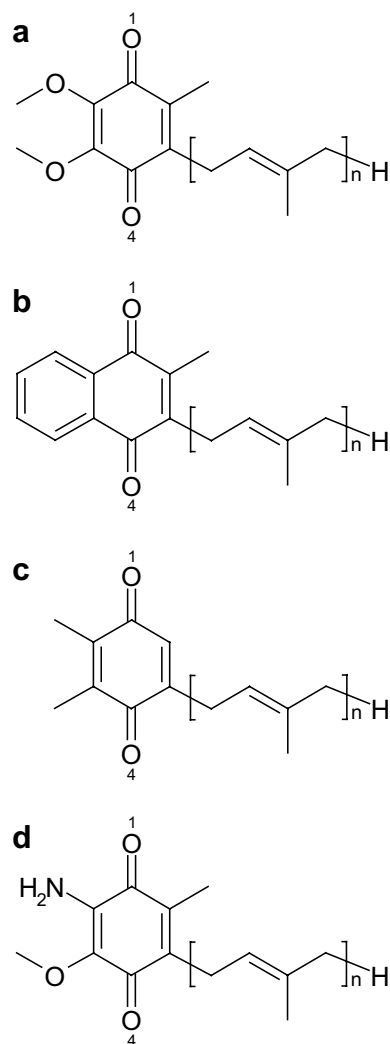


Fig. 2 Molecular structures of **a** ubiquinone ($n=10$), **b** menaquinone/phyloquinone ($n=3-9$), **c** plastoquinone ($n=6-9$), and **d** rholoquinone ($n=10$), where n is the number of isoprene units

Fe₄S₄ cluster F_X is located near the two A₁ binding sites (Fig. 1d). For many 1,4-quinones, experimentally measured pK_a(Q⁻/QH[•]) in aqueous solution were summarized by Swallow (1982) and experimentally measured pK_a(QH⁻/QH₂) in aqueous solution were reported by Bishop and Tong (1965).

As far as we are aware, experimentally measured pK_a(Q⁻/QH[•]) and pK_a(QH⁻/QH₂) for ubiquinone, menaquinone, plastoquinone, and rholoquinone in aqueous solution have not been reported, because of their insoluble hydrophobic isoprene side-chains (Fig. 2). The pK_a(Q⁻/QH[•]) for ubiquinone was roughly estimated to be 4.9 in aqueous solution (Swallow 1982) or measured to be 6.5 in methanol (Land and Swallow 1970; Swallow 1982). In theoretical studies by Cape et al. (2006), pK_a(Q⁻/QH[•]) for ubiquinone, plastoquinone, and rholoquinone were calculated

to be 5.35, 4.86, and 5.09, respectively (Cape et al. 2006), although $pK_a(Q^-/QH)$ of rhodoquinone is assumed to be higher than $pK_a(Q^-/QH)$ of ubiquinone (Graige et al. 1999). The $pK_a(QH^-/QH_2)$ for ubiquinone were measured to be 13.3 in 80% ethanol (Morrison et al. 1982). On the other hand, to calculate $pK_a(QH^-/QH_2)$ of Q_B in PbRC from *R. sphaeroides*, Zhu and Gunner used a $pK_a(QH^-/QH_2)$ of 10.7 for ubiquinone in aqueous solution in their theoretical studies (Zhu and Gunner 2005). Reliable $pK_a(Q^-/QH)$ and $pK_a(QH^-/QH_2)$ values for ubiquinone, menaquinone, and plastoquinone in aqueous solution are required for understanding the mechanisms of reactions involving quinones in PbRC, PSI, and PSII. Here, we report the $pK_a(Q^-/QH)$ and $pK_a(QH^-/QH_2)$ for ubiquinone, menaquinone, phyloquinone, and plastoquinone in aqueous solution, obtained by adopting a quantum chemical approach.

Computational procedures

In the deprotonation reaction of the protonated state (AH) to deprotonated state (A^-) in aqueous solution, pK_a is defined as

$$pK_a = \frac{\Delta G_{aq}}{2.303RT}, \quad (1)$$

where ΔG_{aq} is the free energy difference between (AH) and ($A^- + H^+$) (i.e., $\Delta G_{aq} = G_{aq}(A^-) - G_{aq}(AH) + G_{aq}(H^+)$), R is the gas constant, and T is the temperature. ΔG_{aq} can also be approximated as

$$\Delta G_{aq} = k\Delta E_{QM/PCM} + C, \quad (2)$$

where k is the scaling factor, $\Delta E_{QM/PCM}$ is the energy difference between AH and A^- in aqueous phase (i.e., $\Delta E_{QM/PCM} = E_{QM/PCM}(A^-) - E_{QM/PCM}(AH)$), which can be calculated using a quantum chemical (QM) approach with the polarizable continuum model (PCM) method, and C is the constant [“simple pK_a estimation with energy of the optimized geometry scheme” (Matsui et al. 2012)]. If pK_a of molecules are obtained at the same temperature, Eq. 1 can be written into Eq. 3 using Eq. 2,

$$pK_a = k'\Delta E_{QM/PCM} + C', \quad (3)$$

where k' is the scaling factor and C' is constant. To determine k' and C' , we calculated $\Delta E_{QM/PCM}$ for nine 1,4-quinones whose experimentally measured $pK_a(Q^-/QH)$ (Swallow 1982) and $pK_a(QH^-/QH_2)$ (Bishop and Tong 1965) are reported. We employed the unrestricted DFT method with the B3LYP functional and 6-31g++** basis sets, using the Gaussian (Frisch et al. 2004) program code with the PCM method, i.e., water molecules were considered implicitly. This treatment is more appropriate to describe H-bonds between quinones and bulk water molecules, in which the

H-bond patterns are not unique. We evaluated all possible conformations of each protonated quinone (i.e., QH and QH_2) regarding the $-OH$ group orientation, and we took the energetically lowest conformation as the relevant structure. In the resulting structures, the $-OH$ group was essentially in the plane of the quinone ring (SI Dataset 1 for atomic coordinates). Because the isoprene units are not composed of conjugated double bonds, the isoprene side-chain length n (Fig. 2) was set to 1 for the calculations of ubiquinone, menaquinone, phyloquinone, and plastoquinone. This could also reduce the number of the possible conformations. In fact, the length of the ubiquinone does not practically affect the energetics of ubiquinones, as demonstrated by the experimentally measured redox potential values of ubiquinone-1 and -10 in dimethylformamide being -611 and -602 mV versus saturated calomel reference electrode, respectively, essentially no difference (Prince et al. 1983).

Results and discussion

Correlation of calculated energies with pK_a for 1,4-quinones

The calculated $\Delta E_{QM/PCM}$ for deprotonation of QH to Q^- for nine 1,4-quinones were highly associated with the experimentally measured $pK_a(Q^-/QH)$ ranging from 4.0 to 5.1 (summarized in Swallow 1982), which was best fitted in the following equations (Fig. 3):

$$pK_a(Q^-/QH) = \frac{1}{4.20} (\Delta E_{QM/PCM} - 268.48 [\text{kcal/mol}]). \quad (4)$$

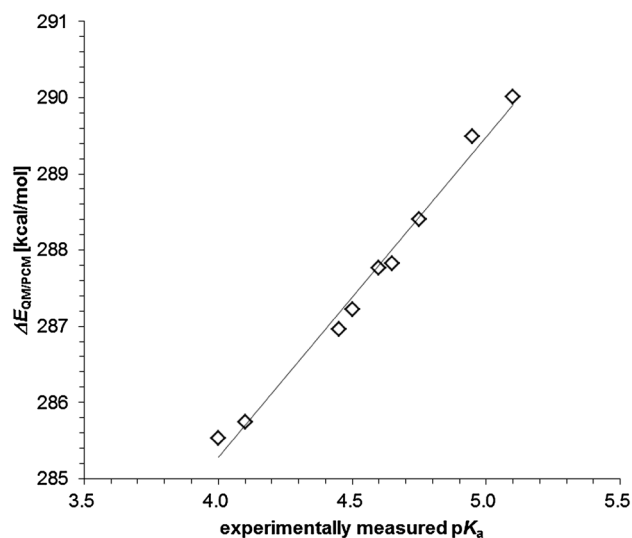


Fig. 3 Correlation between experimentally measured $pK_a(Q^-/QH)$ and calculated $\Delta E_{QM/PCM}$ (coefficient of determination $R^2=0.99$). $\Delta E_{QM/PCM}$ can be calculated using a quantum chemical approach with the PCM method. The solid line was drawn according to Eq. 4

The term of -268.48 kcal/mol corresponds to the proton solvation energy, which typically ranges from -252.6 to -271.7 kcal/mol [see Schmidt am Busch and Knapp (2004) and references therein]. Using Eq. 4, the calculated $pK_a(Q^-/QH)$ for nine 1,4-quinones are listed in Table 1, which confirms that Eq. 4 can reproduce the experimentally measured $pK_a(Q^-/QH)$. It should be noted that the accuracy of the experimentally measured pK_a values is generally considered to be within 0.2 units (Swallow 1982).

Symmetrically shaped quinones (e.g., benzoquinone and 2,3-dimethylbenzoquinone) had a single pK_a value, whereas asymmetrically shaped quinones (e.g., methylbenzoquinone and 2,6-dimethylbenzoquinone) had two distinguishable pK_a values (Table 1). The difference in pK_a values is caused by the difference in the chemical environment of the two O sites (O1 and O4). For example, the calculated $pK_a(Q^-/QH)$ of the O4 site in 2,6-dimethylbenzoquinone was larger than the calculated $pK_a(Q^-/QH)$ of the O1 site by 0.46 (Table 1), because the protonation at O1 (i.e., the formation of $-OH$ at O1) increases the steric hindrance with the two methyl groups, which results in a decrease in pK_a at O1. Since protonation of Q^- to QH occurs predominantly at one of the two O sites with a higher $pK_a(Q^-/QH)$,

the higher $pK_a(Q^-/QH)$ can be considered to be relevant for asymmetrically shaped quinones and are used to obtain Eq. 4.

The calculated $\Delta E_{QM/PCM}$ for deprotonation of QH_2 to QH^- for nine 1,4-quinones were also highly correlated with the experimentally measured $pK_a(QH^-/QH_2)$ ranging from 9.85 to 11.25 (Bishop and Tong 1965), which was best fitted in the following equations (Fig. 4):

$$pK_a(QH^-/QH_2) = \frac{1}{2.40} (\Delta E_{QM/PCM} - 280.00 [\text{kcal/mol}]) \quad (5)$$

The $pK_a(QH^-/QH_2)$ calculated for nine quinones using Eq. 5 are listed in Table 2, which confirms that Eq. 5 can reproduce the experimentally measured $pK_a(QH^-/QH_2)$ when $\Delta E_{QM/PCM}$ can be calculated appropriately.

$pK_a(Q^-/QH)$ and $pK_a(QH^-/QH_2)$ for ubiquinone, menaquinone, phyloquinone, plastoquinone, and rhodoquinone in aqueous solution

Using Eqs. 4 and 5, we also calculated $pK_a(Q^-/QH)$ and $pK_a(QH^-/QH_2)$ for ubiquinone,

Table 1 Calculated $pK_a(Q^-/QH)$ for the first protonation process Q^- to QH in water, using Eq. 4

	$pK_a(Q^-/QH)$		
	Measured (in water)	Calculated (in water)	(Lower)
Benzoquinone	4 ^{a,b,c}	4.06	n.d
Methylbenzoquinone	4.45 ^b	4.40	(4.24)
2,3-Dimethylbenzoquinone	4.65 ^b	4.60	n.d
2,5-Dimethylbenzoquinone	4.6 ^{b,d,e}	4.59	n.d
2,6-Dimethylbenzoquinone	4.75 ^b	4.74	(4.28)
Trimethylbenzoquinone	4.95 ^b	5.00	(4.73)
Duroquinone	5.1 ^{b,d,e}	5.13	n.d
1,4-Naphthoquinone	4.1 ^{b,d,e}	4.11	n.d
2-Methyl-1,4-naphthoquinone	4.5 ^{b,e}	4.46	(4.15)
Ubiquinone	n.d	5.31 at O4	(5.30 at O1)
Menaquinone/phyloquinone	n.d	4.92 at O4	(4.52 at O1)
Plastoquinone	n.d	5.11 at O4	(5.01 at O1)
Rhodoquinone	n.d	5.78 at O4	(4.23 at O1)

Lower $pK_a(QH/Q^-)$ values, if available (i.e., asymmetrically shaped quinones), are listed in the bracket. Since protonation of Q^- to QH occurs predominantly at one of the two O sites, O1 and O4, with a higher $pK_a(Q^-/QH)$, the higher $pK_a(Q^-/QH)$ can be considered to be experimentally measurable. Experimentally measured $pK_a(QH/Q^-)$ are summarized in Swallow (1982). The error for nine quinones between experimentally measured and calculated $pK_a(Q^-/QH)$ was 0.04 in pK_a unit. See Figs. 1 and 2 for the location of the O1 and O4 sites. n.d. = not determined

^aAdams and Michael (1967)

^bPatel and Willson (1973)

^cSteenken and O'Neill (1977)

^dRao and Hayon (1973)

^eWillson (1971)

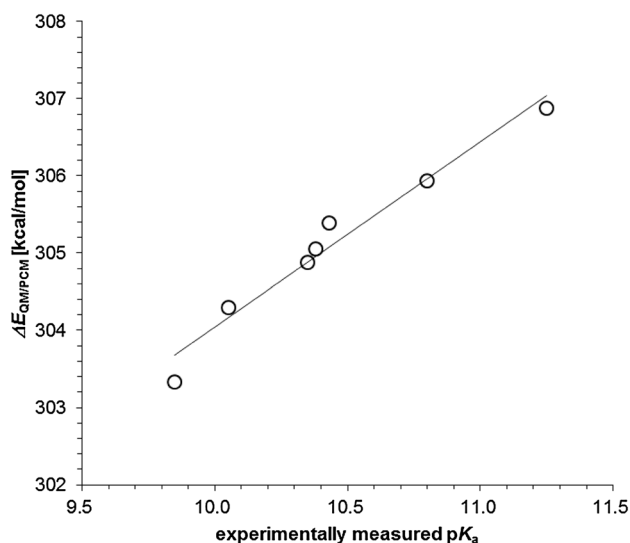


Fig. 4 Correlation between experimentally measured $pK_a(QH^-/QH_2)$ and calculated $\Delta E_{QM/PCM}$ (coefficient of determination $R^2=0.96$). $\Delta E_{QM/PCM}$ can be calculated using a quantum chemical approach with the PCM method. The solid line is drawn according to Eq. 5

Table 2 Calculated $pK_a(QH^-/QH_2)$ for the second protonation process QH^- to QH_2 in water, using Eq. 5

	$pK_a(QH^-/QH_2)$	
	Measured (in water)	Calculated (in water)
Benzoquinone	9.85 ^a	9.70
Methylbenzoquinone	10.05 ^a	10.10
2,3-Dimethylbenzoquinone	10.43 ^a	10.56
2,5-Dimethylbenzoquinone	10.38 ^a	10.42
2,6-Dimethylbenzoquinone	10.35 ^a	10.35
Trimethylbenzoquinone	10.8 ^a	10.79
Duroquinone	11.25 ^a	11.18
Ubiquinone	n.d	10.86
Menaquinone/phyloquinone	n.d	9.16
Plastoquinone	n.d	10.74
Rhodoquinone	n.d	9.81

The error for seven quinones between experimentally measured and calculated $pK_a(Q^-/QH)$ was 0.09 in pK_a unit. *n.d.* not determined

^aBishop and Tong (1965)

menaquinone, phyloquinone, and plastoquinone in aqueous solution, respectively. The calculated $pK_a(Q^-/QH)$ were 5.31 for ubiquinone, 4.92 for menaquinone/phyloquinone, and 5.11 for plastoquinone in aqueous solution (Table 1). $pK_a(Q^-/QH)$ did not significantly differ between O1 and O4 in plastoquinone and ubiquinone (Table 1). This result suggests that which of the two O sites, O1 and O4, serves as the initial (i.e., Q^- to QH) and second (i.e., QH^- to QH_2) protonation sites of Q_B in

PbRC and PSII, is predominantly determined by the protein environments.

The calculated $pK_a(Q^-/QH)$ for ubiquinone was closer to the value of 4.9 roughly estimated for aqueous solution (Swallow 1982) than the value of 6.5 measured in methanol (Land and Swallow 1970; Swallow 1982). Because methanol (dielectric constant=33) is less polar than water (dielectric constant=80), the negatively charged Q^- state is less stable in methanol than in water; this could explain the high $pK_a(Q^-/QH)$ in methanol with respect to water.

In theoretical studies by Cape et al., the calculated $pK_a(Q^-/QH)$ of 5.09 for rhodoquinone was lower than their calculated $pK_a(Q^-/QH)$ of 5.35 for ubiquinone (Cape et al. 2006). In contrast, in the present study, the calculated $pK_a(Q^-/QH)$ of rhodoquinone was significantly high, 5.78 (Table 1). It should be noted that $pK_a(Q^-/QH)$ of rhodoquinone is assumed to be higher than $pK_a(Q^-/QH)$ of ubiquinone (e.g., Graige et al. 1999), which is consistent with the present result.

The calculated $pK_a(QH^-/QH_2)$ of 10.86 for ubiquinone (Table 2) is very close to $pK_a(QH^-/QH_2)$ of 10.8 measured for trimethylbenzoquinone (Bishop and Tong 1965). Zhu and Gunner considered $pK_a(QH^-/QH_2)$ for trimethylbenzoquinone being more relevant to the $pK_a(QH^-/QH_2)$ for ubiquinone in aqueous solution (Zhu and Gunner 2005) than $pK_a(QH^-/QH_2)=13.3$ in 80% ethanol (Morrison et al. 1982). The present study supports their conclusion. The calculated $pK_a(QH^-/QH_2)$ of 10.86 for ubiquinone is also close to the calculated $pK_a(QH^-/QH_2)$ of 10.74 for plastoquinone (Table 2), which might be associated with the similarity in the Q_B protonation events for PbRC and PSII (Robinson and Crofts 1984; Rutherford et al. 1984; Okamura et al. 2000; Wraight 2004; Ishikita and Knapp 2005).

Influence of the 2-methoxy group orientation in $pK_a(Q^-/QH)$ for ubiquinone

It was proposed that difference in the 2-methoxy orientation of ubiquinone (Fig. 1b) was responsible for (i) the E_m difference of more than 160 mV between Q_A and Q_B in PbRC (Taguchi et al. 2013) or (ii) the quantum chemically obtained electron affinity difference of more than 170 meV between Q_A and Q_B in PbRC (de Almeida et al. 2014). On the other hand, FTIR studies suggested that the Q_A binding to PbRC (i.e., interaction between Q_A and the PbRC protein environment) was affected by the H-bond interaction with the protein environment but not by the methoxy orientation (Remy et al. 2003). In the present study, using the crystal structure (PDB: 3I4D; Fig. 1b) whose different 2-methoxy orientations between Q_A and Q_B were highlighted in ref. (de Almeida et al. 2014), $pK_a(Q^-/QH)$ were calculated. The calculated $pK_a(Q^-/QH)$ were the same, 5.30 at the O4 site in the Q_A conformation and 5.31 at the O4 site in the

Q_B conformation in water (Table S1), irrespective of the different 2-methoxy orientations in the quantum chemically optimized quinone structures (SI Dataset 2 for atomic coordinates). In the protonated QH^+ state, the energetically lowest conformations showed that the $-OH$ group was oriented toward the 2-methoxy O atom ($-O-H\cdots O_{\text{methoxy}}$ angle = 116° and $-O\cdots O_{\text{methoxy}}$ distance = 2.1 \AA in both Q_A and Q_B conformations). When the $-OH$ group was oriented away from the 2-methoxy O atom, the calculated $pK_a(Q^-/QH)$ were lowered by ~ 0.6 to $1 pK_a$ unit from the $pK_a(Q^-/QH)$ value of the energetically lowest conformation (Table S1). In the quantum chemically optimized Q_A and Q_B conformations, the 2-methoxy group was out of the quinone ring, as identified in the crystal structure (PDB: 3I4D). These results suggest that $pK_a(Q^-/QH)$ is more affected by the difference in the $-OH$ orientation than the difference in the 2-methoxy orientation. The influence of the 2-methoxy orientation might be more pronounced when calculated in vacuum (i.e., in the absence of the protein environment), where H-bond partners of ubiquinone (e.g., bulk water molecules and protein environments) are absent. In vacuum, alteration in the molecular configuration is the only way to alter the stability of the Q^- and QH^+ states. It should be noted that the influence of $-OH$ orientation can be ignored when considering $E_m(Q/Q^-)$, because $-OH$ is absent in both the Q and Q^- states.

Evolutionary transition of quinones in photosynthetic reaction centers

Among all of the 1,4-quinones investigated here, plastoquinone and ubiquinone employed at Q_B in PbRC and PSII have the second and third largest $pK_a(Q^-/QH)$ (Table 1). This might be a reason why nature uses these quinones exclusively as the terminal electron acceptor Q_B in PbRC and PSII. The large $pK_a(Q^-/QH)$ would result in a larger population of QH . This will then enable it to accept the second electron, leading to the fully protonated Q_BH_2 state. Thus, the large $pK_a(Q^-/QH)$ is advantageous in fixing the two transferred electrons using protonation in the form of Q_BH_2 . Remarkably, Q_A in PbRC from *B. viridis* and A_{1A} and A_{1B} in PSI, which serve as electron donors (to Q_B and the Fe_4S_4 cluster F_X , respectively, Fig. 1) without altering the protonation states, are menaquinone/phyloquinone with lower $pK_a(Q^-/QH) = 4.9$ (Table 1).

Menaquinone is assumed to represent the ancestral type of quinones in bioenergetics systems, whereas ubiquinone and plastoquinone are assumed to represent “more recent” quinones (Schoepp-Cothenet et al. 2009). Intriguingly, $E_m(Q/Q^-)$ of the “more recent” quinones are higher [by $\sim 100 \text{ mV}$ (Prince et al. 1983)] than $E_m(Q/Q^-)$ of the “ancestral” quinone. Reduced menaquinone becomes rapidly oxidized in the presence of O_2 , whereas ubiquinone

and plastoquinone are less sensitive toward oxygen, minimizing loss of the unproductive reduced power (Schoepp-Cothenet et al. 2009); it seems plausible that the difference in $E_m(Q/Q^-)$ is associated with rising levels of dioxygen 2.5 billion years ago. In most PbRC, the tightly bound cofactor at the Q_A site can be either menaquinone or ubiquinone, whereas the cofactor at the terminal electron acceptor Q_B site is ubiquinone [except for, e.g., PbRC from *Halorhodospira halophila*, which has menaquinone at both the Q_A and Q_B site (Schoepp-Cothenet et al. 2009)]. The higher $E_m(Q/Q^-)$ of ubiquinone at the Q_B site would contribute to the driving force of the electron transfer from Q_A to Q_B when menaquinone is the cofactor at the Q_A site (e.g., *B. viridis*). The present results show that the “more recent” quinones, ubiquinone and plastoquinone, have larger $pK_a(Q^-/QH)$ and $pK_a(QH^-/QH_2)$ values than the “ancestral” quinone, menaquinone/phyloquinone (Tables 1, 2), which is also consistent with a role of Q_B , fixation of two electrons and two protons in the form of Q_BH_2 .

In photosynthetic reaction centers of the green non-sulfur bacteria (e.g., *Chloroflexus aurantiacus*), which is assumed to have evolved first as the type-II reaction center (Gupta 2003), menaquinone occupies at both the Q_A and Q_B sites (Hale et al. 1983). One of the simplest ways to generate the driving force for electron transfer between the identical menaquinone cofactors would be to alter the solvent accessibility between the Q_A and Q_B sites. As the quinone-binding site is more exposed to the bulk water region, it can stabilize the Q^- state with respect to Q state, which results in an increase in $E_m(Q)$, i.e., the case for $E_m(Q_B)$, even if the amino acid sequences near the Q_A and Q_B binding sites were similar. In addition, the exposure of the quinone-binding site makes (P)bRC easier to hire external quinones. The effort to make the solvent accessibility of the quinone-binding sites different between the two quinone-binding sites in the homodimeric type-I reaction center would lead to formation of the heterodimeric type-II reaction center with $E_m(Q_B) > E_m(Q_A)$ and might also explain why the cofactor can be ubiquinone at the Q_B site in most PbRC.

Conclusions

Experimentally measured $pK_a(Q^-/QH)$ (Swallow 1982) and $pK_a(QH^-/QH_2)$ (Bishop and Tong 1965) values of nine 1,4-quinones in aqueous solution were highly correlated with the quantum chemically calculated energy differences ($\Delta E_{QM/PCM}$) between the protonated and deprotonated states (Figs. 3, 4). They can be best fitted to Eqs. 4 and 5, respectively. The calculated $pK_a(Q^-/QH)$ values were 5.31 for ubiquinone, 4.92 for menaquinone/phyloquinone, 5.11 for plastoquinone, and

5.78 for rholoquinone in aqueous solution (Table 1). $pK_a(Q^-/QH^-)$ for plastoquinone and ubiquinone in aqueous solution are the largest among all of the 1,4-quinones but rholoquinone listed in Table 1, partially explaining why nature employs these two 1,4-quinones specifically as the terminal electron acceptors Q_B in PbRC and PSII.

In PbRC and PSII, the initial protonation of Q^- to QH^- predominantly occurs at the distal carbonyl site O_{dist} of Q_B (Fig. 1) (Okamura et al. 2000; Wraight 2004; Saito et al. 2013). The $pK_a(Q^-/QH^-)$ at the O1 and O4 sites did not differ significantly for each quinone (Table 1), suggesting that the protein environments predominantly determine the initial protonation O site of Q_B .

The $pK_a(QH^-/QH_2)$ values were calculated to be 10.86 for ubiquinone, 9.16 for menaquinone/phyloquinone, 10.74 for plastoquinone, and 9.81 for rholoquinone in aqueous solution using Eq. 5 (Table 2). The calculated $pK_a(QH^-/QH_2)$ for ubiquinone was closer to the experimentally measured value of 10.8 for trimethylbenzoquinone (Bishop and Tong 1965) than the experimentally measured $pK_a(QH^-/QH_2)$ of 13.3 for ubiquinone in 80% ethanol (Morrison et al. 1982), as already pointed out (Zhu and Gunner 2005).

The $pK_a(Q^-/QH^-)$ and $pK_a(QH^-/QH_2)$ for ubiquinone, menaquinone/phyloquinone, and plastoquinone in aqueous solution, which were determined in the present study, will help understand the mechanisms of quinone-mediated reactions in photosynthetic reaction centers, such as formation of Q_BH_2 in PbRC and PSII (Robinson and Crofts 1984; Rutherford et al. 1984; Okamura et al. 2000; Wraight 2004; Saito et al. 2013), formation of Q_AH_2 in PSII (van Mieghem et al. 1995; Noguchi 2002), and evolutionary relationship between the type-I and type-II photosynthetic reaction centers (Gupta 2003; Rutherford and Faller 2003; Schoepp-Cothenet et al. 2009).

Acknowledgements This research was supported by JST CREST (JPMJCR1656 to H.I.), JSPS KAKENHI (JP26800224 to K.S., JP15H00864, JP16H06560, JP26105012, and JP26711008 to H.I., and JP22740276 to K.S. and H.I.), Materials Integration for engineering polymers of Cross-ministerial Strategic Innovation Promotion Program (SIP, H.I.), and Interdisciplinary Computational Science Program in CCS, University of Tsukuba. Theoretical calculations were partly performed using Research Center for Computational Science, Okazaki, Japan.

Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

References

- Adams GE, Michael BD (1967) Pulse radiolysis of benzoquinone and hydroquinone. Semiquinone formation by water elimination from trihydroxy-cyclohexadienyl radicals. *Trans Faraday Soc* 63(0):1171–1180. doi:10.1039/tf9676301171
- Bishop CA, Tong LKJ (1965) Equilibria of substituted semiquinones at high pH. *J Am Chem Soc* 87:501–505
- Brettel K, Leibl W (2001) Electron transfer in photosystem I. *Biochim Biophys Acta* 1507:100–114
- Cape JL, Bowman MK, Kramer DM (2006) Computation of the redox and protonation properties of quinones: towards the prediction of redox cycling natural products. *Phytochemistry* 67(16):1781–1788. doi:10.1016/j.phytochem.2006.06.015
- Cardona T, Sedoud A, Cox N, Rutherford AW (2012) Charge separation in photosystem II: a comparative and evolutionary overview. *Biochim Biophys Acta* 1817(1):26–43
- de Almeida WB, Taguchi AT, Dikanov SA, Wraight CA, O'Malley PJ (2014) The 2-methoxy group orientation regulates the redox potential difference between the primary (Q_A) and secondary (Q_B) quinones of Type II bacterial photosynthetic reaction centers. *J Phys Chem Lett* 5(15):2506–2509. doi:10.1021/jz500967d
- Diner BA, Rappaport F (2002) Structure dynamics, and energetics of the primary photochemistry of photosystem II of oxygenic photosynthesis. *Annu Rev Plant Biol* 53:551–580
- Frisch MJ, Trucks GW, Schlegel HB, Scuseria GE, Robb MA, Cheeseman JR, Montgomery JA Jr., Vreven T, Kudin KN, Burant JC, Millam JM, Iyengar SS, Tomasi J, Barone V, Mennucci B, Cossi M, Scalmani G, Rega N, Petersson GA, Nakatsuji H, Hada M, Ehara M, Toyota K, Fukuda R, Hasegawa J, Ishida M, Nakajima T, Honda Y, Kitao O, Nakai H, Klene M, Li X, Knox JE, Hratchian HP, Cross JB, Bakken V, Adamo C, Jaramillo J, Gomperts R, Stratmann RE, Yazyev O, Austin AJ, Cammi R, Pomelli C, Ochterski JW, Ayala PY, Morokuma K, Voth GA, Salvador P, Dannenberg JJ, Zakrzewski VG, Dapprich S, Daniels AD, Strain MC, Farkas O, Malick DK, Rabuck AD, Raghavachari K, Foresman JB, Ortiz JV, Cui Q, Baboul AG, Clifford S, Cioslowski J, Stefanov BB, Liu G, Liashenko A, Piskorz P, Komaromi I, Martin RL, Fox DJ, Keith T, Al-Laham MA, Peng CY, Nanayakkara A, Challacombe M, Gill PMW, Johnson B, Chen W, Wong MW, Gonzalez C, Pople JA (2004). Gaussian 03. Gaussian, Inc., Wallingford
- Graige MS, Paddock ML, Feher G, Okamura MY (1999) Observation of the protonated semiquinone intermediate in isolated reaction centers from *Rhodobacter sphaeroides*: implications for the mechanism of electron and proton transfer in proteins. *Biochemistry* 38(35):11465–11473. doi:10.1021/bi990708u
- Gupta RS (2003) Evolutionary relationships among photosynthetic bacteria. *Photosynth Res* 76(1–3):173–183. doi:10.1023/A:1024999314839
- Hale MB, Blankenship RE, Fuller RC (1983) Menaquinone is the sole quinone in the facultatively aerobic green photosynthetic bacterium *Chloroflexus aurantiacus*. *Biochimica et Biophysica Acta* 723(3):376–382. doi:10.1016/0005-2728(83)90044-0
- Holzwarth AR (2008) Ultrafast primary reactions in the photosystems of oxygen-evolving organisms. In: Braun M, Gilch P, Zinth W (eds) Ultrashort laser pulses in biology and medicine. Springer, Dordrecht, pp 141–164
- Ishikita H, Knapp E-W (2005) Control of quinone redox potentials in photosystem II: electron transfer and photoprotection. *J Am Chem Soc* 127:14714–14720
- Jordan P, Fromme P, Witt HT, Klukas O, Saenger W, Krauß N (2001) Three-dimensional structure of cyanobacterial photosystem I at 2.5 Å resolution. *Nature* 411:909–917

- Land EJ, Swallow AJ (1970) One-electron reactions in biochemical systems as studied by pulse radiolysis. 3. Ubiquinone. *J Biol Chem* 245(8):1890–1894
- Li L, Mustafi D, Fu Q, Tereshko V, Chen DL, Tice JD, Ismagilov RF (2006) Nanoliter microfluidic hybrid method for simultaneous screening and optimization validated with crystallization of membrane proteins. *Proc Natl Acad Sci USA* 103(51):19243–19248. doi:10.1073/pnas.0607502103
- Maroti A, Wraight CA, Maroti P (2015) Protonated rhodosemiquinone at the Q_B binding site of the M265IT mutant reaction center of photosynthetic bacterium *Rhodobacter sphaeroides*. *Biochemistry* 54(12):2095–2103. doi:10.1021/bi501553t
- Matsui T, Baba T, Kamiya K, Shigeta Y (2012) An accurate density functional theory based estimation of pK_a values of polar residues combined with experimental data: from amino acids to minimal proteins. *Phys Chem Chem Phys* 14(12):4181–4187
- Morrison LE, Schelhorn JE, Cotton TE, Bering CL, Loach PA (1982) Electrochemical and spectral properties of ubiquinone and synthetic analogs: relevance to bacterial photosynthesis. In: Trumpower BL (ed) *Function of quinones in energy conserving systems*. Academic Press, New York, pp 35–58
- Muh F, Glockner C, Hellmich J, Zouni A (2012) Light-induced quinone reduction in photosystem II. *Biochim Biophys Acta* 1817(1):44–65
- Noguchi T (2002) Dual role of triplet localization on the accessory chlorophyll in the photosystem II reaction center: photoprotection and photodamage of the D1 protein. *Plant Cell Physiol* 43:1112–1116
- Okamura MY, Paddock ML, Graige MS, Feher G (2000) Proton and electron transfer in bacterial reaction centers. *Biochim Biophys Acta* 1458:148–163
- Okayama S, Yamamoto N, Nishikawa K, Horio T (1968) Roles of ubiquinone-10 and rhodoquinone in photosynthetic formation of adenosine triphosphate by chromatophores from *Rhodospirillum rubrum*. *J Biol Chem* 243(11):2995–2999
- Patel KB, Willson RL (1973) Semiquinone free radicals and oxygen. Pulse radiolysis study of one electron transfer equilibria. *J Chem Soc Faraday Trans 1* 69(0):814–825. doi:10.1039/f19736900814
- Petrouleas V, Crofts AR (2005) The quinone iron acceptor complex. In: Wydrzynski T, Satoh K (eds) *Photosystem II: the water/plastoquinone oxido-reductase in photosynthesis*. Springer, Dordrecht, pp 177–206
- Prince RC, Dutton PL, Bruce JM (1983) Electrochemistry of ubiquinones: menaquinones and plastoquinones in aprotic solvents. *FEBS Lett* 160:273–276
- Rao PS, Hayon E (1973) Ionization constants and spectral characteristics of some semiquinone radicals in aqueous solution. *J Phys Chem* 77:2274–2276
- Remy A, Boers RB, Egorova-Zachernyuk T, Gast P, Lugtenburg J, Gerwert K (2003) Does different orientation of the methoxy groups of ubiquinone-10 in the reaction centre of *Rhodobacter sphaeroides* cause different binding at Q_A and Q_B ? *Eur J Biochem* 270(17):3603–3609
- Renger G, Renger T (2008) Photosystem II: the machinery of photosynthetic water splitting. *Photosynth Res* 98(1–3):53–80
- Robinson HH, Crofts AR (1984) Kinetics of proton uptake and the oxidation-reduction reactions of the quinone acceptor complex of photosystem II from pea chloroplasts. In: Sybesma C (ed) *Advances in photosynthesis research*, vol 1. Martinus Nijhoff/Dr. W. Junk Publishers, The Hague, pp 477–480
- Rutherford AW, Faller P (2003) Photosystem II: evolutionary perspectives. *Phil Trans R Soc Lond B* 358:245–253
- Rutherford AW, Renger G, Koike H, Inoue Y (1984) Thermoluminescence as a probe of photosystem II. The redox and protonation states of the secondary acceptor quinone and the O_2 -evolving enzyme. *Biochim Biophys Acta* 767(3):548–556
- Saito K, Rutherford AW, Ishikita H (2013) Mechanism of proton-coupled quinone reduction in Photosystem II. *Proc Natl Acad Sci USA* 110(3):954–959
- Schmidt am Busch M, Knapp E-W (2004) Accurate pK_a determination for a heterogeneous group of organic molecules. *ChemPhysChem* 5:1513–1522
- Schoepp-Cothenet B, Lieutaud C, Baymann F, Vermeglio A, Friedrich T, Kramer DM, Nitschke W (2009) Menaquinone as pool quinone in a purple bacterium. *Proc Natl Acad Sci USA* 106(21):8549–8554. doi:10.1073/pnas.0813173106
- Steenken S, O'Neill P (1977) Oxidative demethoxylation of methoxylated phenols and hydroxybenzoic acids by the hydroxyl radical. An in situ electron spin resonance, conductometric pulse radiolysis and product analysis study. *J Phys Chem* 81:505–508
- Swallow AJ (1982) Physical chemistry of semiquinones. Trumpower BL (ed) *Function of quinones in energy conserving systems*. Academic Press, New York, pp 59–72
- Taguchi AT, Mattis AJ, O'Malley PJ, Dikanov SA, Wraight CA (2013) Tuning cofactor redox potentials: the 2-methoxy dihedral angle generates a redox potential difference of >160 mV between the primary (Q_A) and secondary (Q_B) quinones of the bacterial photosynthetic reaction center. *Biochemistry* 52(41):7164–7166. doi:10.1021/bi4011896
- Umena Y, Kawakami K, Shen J-R, Kamiya N (2011) Crystal structure of oxygen-evolving photosystem II at a resolution of 1.9 Å. *Nature* 473:55–60
- van Mieghem F, Brettel K, Hillman B, Kamlowski A, Rutherford AW, Schlodder E (1995) Charge recombination reactions in photosystem II. 1. Yields, recombination pathways, and kinetics of the primary pair. *Biochemistry* 34:4798–4813
- Willson RL (1971) Semiquinone free radicals: determination of acid dissociation constants by pulse radiolysis. *J Chem Soc D* 20:1249–1250. doi:10.1039/c29710001249
- Wraight CA (2004) Proton and electron transfer in the acceptor quinone complex of photosynthetic reaction centers from *Rhodobacter sphaeroides*. *Front Biosci* 9:309–337
- Zhu Z, Gunner MR (2005) Energetics of quinone-dependent electron and proton transfers in *Rhodobacter sphaeroides* photosynthetic reaction centers. *Biochemistry* 44:82–96