

Genetic Divergence and Biogeographical Patterns in *Amentotaxus argotaenia* Species Complex

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Abstract The *Amentotaxus argotaenia* (Taxaceae) species complex is comprised of four relict and endangered gymnosperms. *Amentotaxus argotaenia* and *A. yunnanensis* are scattered throughout South and Central China, and *A. formosana*, and *A. poilanei* are restricted to Taiwan and Vietnam. In the present study, we examined genetic divergence and biogeographical patterns in the species complex based on chloroplast DNA *atpI-atpH* and *rpS16-trnK* intergenic spacers, the mitochondrial DNA *nadI* intron, and microsatellite markers. Genealogical analyses revealed

systematic inconsistencies between organellar DNA markers, with paraphyletic species inferred based on cpDNA, versus a monophyletic *A. formosana*, and clustering of *A. poilanei* within *A. yunnanensis* at mtDNA. AMOVA revealed that most organellar DNA variants resided among species and populations, a pattern also supported by clustering with STRUCTURE analyses on microsatellites. Higher levels of genetic diversity in the Yongxin population of *A. argotaenia* reflected features of a refugium. Most speciation events were rather recent, with divergence no earlier than 2.50 MYA, although many ancient lineages derived 4.12–13.84 MYA were maintained within species. Significant genetic structuring within *A. argotaenia* was detected among three geographical regions—a phylogeographical pattern likely attributable to past fragmentations. In contrast to paraphyletic *A. argotaenia* based on the maintenance of ancestral polymorphisms, interspecific hybridization likely contributed to polyphyly in *A. yunnanensis*. Furthermore, despite the low likelihood of recurrent gene flow due to geographical isolation, IM analyses revealed continued interspecific gene flow after species divergence, reflecting a parapatric speciation model.

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Introduction

Amentotaxus (Taxaceae) represents an ancient lineage of six relict species (Farjon 2010). Tertiary fossil evidence indicates this genus was once widespread in the Northern Hemisphere (Harris 1976). However, currently *Amentotaxus* is restricted to subtropical, southeast Asia, from Taiwan west across southern China to Assam in the eastern Himalaya, and south to Vietnam (Ferguson et al. 1978; Royer et al. 2003). Four species of

Amentotaxus, namely *A. argotaenia* Pilg., *A. formosana* H. L. Li, *A. yunnanensis* H. L. Li, and *A. poilanei* D. K. Ferguson are considered a species complex, which remains a taxonomic challenge due to the existence of intermediate morphotypes based on species diagnostic traits, i.e., stomatal band width and leaf shape (Cheng 1978; Zhou 2001). All four species are characteristic of subtropical forests, and exhibit an allopatric distribution with clear habitat differentiation. *A. yunnanensis* is confined to southwest China, growing in limestone habitats; *A. poilanei* is restricted to moist montane forests of Vietnam; *A. formosana* is an element of temperate broadleaf forests in Taiwan; and *A. argotaenia* is restricted to limestone mountains, forests, ravines, shady and damp stream banks across 11 provinces of South and Central China. Demographically, all populations remain in small, isolated fragments, primarily with fewer than 100 individuals; and only a single *A. formosana* population remains in southern Taiwan (Li and Keng 1994). According to IUCN categories (IUCN 2013), *A. argotaenia* and *A. poilanei* are under vulnerable conditions (VU), and *A. formosana* and *A. yunnanensis* are under serious threats (EN), all with small population numbers and sizes.

Geographical barriers, such as the Taiwan Strait and the Himalayan mountain ranges might have played an integral role in genetic differentiation and speciation of this complex. Nevertheless, species boundaries among gymnosperms remain poorly explored. Although allopolyploid and homoploid speciation has been found much more common than expected in conifers (Chiang et al. 2006; Wachowiak and Prus-Glowacki 2008; Wang et al. 2012; Zhou et al. 2010), gene flow levels among species remain poorly understood. Occurrence of natural hybrids does not necessarily reject allopatric speciation—a long-accepted model in which gene flow ceases immediately following speciation—as it has long been suggested hybrids can act as a reproductive barrier between progenitor species (Wang et al. 2000). In fact, the existence of natural hybrids is not equivalent to the occurrence of interspecific gene flow. In this study, we examined allopatric speciation in *Amentotaxus* by examining the extent of gene flow since species divergence.

Extant and historical distributions of *Amentotaxus* species show a demographic history attributable to fluctuating climates. Based on the palynoflora, Quaternary glaciers largely removed most temperate species, like *Pinus massoniana* and *P. hwangshanensis*, in the north regions of China (Zhou et al. 1994, 2010; Ge et al. 2012), resulting in range shifts that forced plants into southern refugia, and a rich south China flora (Hewitt 1999; Tong et al. 1992). In contrast to North Europe and North America, where many of the Tertiary plant species went extinct, refugia in central and southwestern China (below 31° N) supported many southward migrants during the last glacial maximum (LGM) (Ge et al. 2012; Harrison et al. 2001; Liu 1991; Ray and Adams 2001; Wang

et al. 2011; Xu et al. 2008). Furthermore, Del Tredici et al. (1992) suggested another refugium located in Eastern China. Geographically, primarily east–west oriented mountain ranges buffered climate change effects across a latitudinal gradient. Tertiary relicts, such as *Cathaya*, *Ginkgo*, *Metasequoia*, and *Taiwania*, which were once widespread across Europe and North America, survived in this region, with a scattered distribution and small population sizes (Traula 1968). However, subsequent postglacial recolonization did not substantially recover the species ranges, due largely to dry climate in northern China, where vegetation has since been dominated by steppe (Tong et al. 1999). Accordingly, postglacial colonization resulted in a range of phylogeographical patterns in species from different latitudes (Hewitt 1996; Chiang and Schaal 2006).

The *Amentotaxus argotaenia* complex, which spans the subtropical mainland, and a continental island provides an ideal model with which to examine genetic divergence among sister species, and genetic structure among populations across species geographical ranges, and to further elucidate speciation processes. Species evolving through glacial cycles are of particular interest to evolutionists, as climate changes during the ice ages had a dramatic influence on species, causing isolation, migration, and extinction, as well as accelerating speciation (Taberlet et al. 1998). Under conditions of small population sizes shaped by random genetic drift, rare species possess low levels of genetic variation, and display evident geographic structuring due to the stochastic loss of genetic polymorphisms from populations. Ge et al. (2005) reported significant genetic differentiation among species of the *A. argotaenia* complex, where all species populations grow under forest canopies, with a scattered and patch-like distribution.

In this study, we reconstructed phylogenies and population structure of the *Amentotaxus* species complex by investigating the genetic variation of organellar DNAs and microsatellite fingerprinting. The following questions were addressed: (1) Are the four species genetically distinct entities? (2) Did these taxa experienced interspecific gene flow since divergence? (3) *A. argotaenia* exhibits a much larger geographic area than the other three species; therefore, does it possess higher levels of genetic diversity? And (4) what is the phylogeographic pattern of relict *Amentotaxus* species in hypothetical refugia of China?

Materials and Methods

Population, Sampling and DNA Extraction

Seventeen populations throughout the whole geographical range of distribution of the *Amentotaxus argotaenia* complex were surveyed. In total, 98 individuals were sampled from 11

populations of *A. argotaenia*, 12 individuals from the single population of *A. formosana*, 38 individuals from four populations of *A. yunnanensis*, and 11 individuals from the single population of *A. poilanei*. Rare distributions of this complex pose a difficulty in sampling (Tran 2011) (Table 1). Within the distributional range of *A. argotaenia*, three regions can be discerned based on the geographical topography. The East region was separated from West and Central regions by the Mt. Nanling and the Mt. Wuyi; West and Central regions were separated from each other by the Yangtze River (Fig. 1). Identical materials were previously used for an ISSR fingerprinting analyses (Ge et al. 2005), except for Bamianshan (HU) population of *A. argotaenia* in China, and Lao Va Chai Municipality (VL) of *A. poilanei* in Vietnam. Young, healthy leaves were collected and dried in silica gel until DNA extraction. Total genomic DNA was extracted from leaves following a CTAB methodology (Doyle and Doyle 1987).

Organelle DNA Analyses

The cpDNA (chloroplast DNA) spacers and mtDNA (mitochondrial DNA) intron were amplified by the polymerase chain reaction (PCR) using primers *atpI* 5' TAT TTA CAA GCG GTA TTC AAG CT 3' and *atpH* 5' CCA ACC CAG

CAG CAA TAA C 3', *trnK* 5' TTA AAA GCC GAG TAC TCT ACC 3' and *rpS16* 5' AAA GTG GGT TTT TAT GAT CC 3', and *nad1/F* 5' GAT CGG CCATAA ATG TAC TCC 3' and *nad1/R* 5' CCC CAT ATA TTC CCG GAG C 3'. PCR products were electrophoresed on agarose gels and the DNA fragment was excised from the gel and purified. Eluted PCR products were sequenced directly in both directions on an ABI 377XL automated sequencer (Applied Biosystem, Foster City, CA).

Microsatellite Genotyping

Fifteen variable microsatellite loci were amplified and genotyped (Ho et al. 2012). PCR amplification was carried out in a 50 uL reaction, with forward primers for PCR labelled with fluorescent dye. PCR products were separated with an ABI 3100 automated sequencer, and fragment sizes were assessed using GENEMAPPER software version 3.7 (Applied Biosystems).

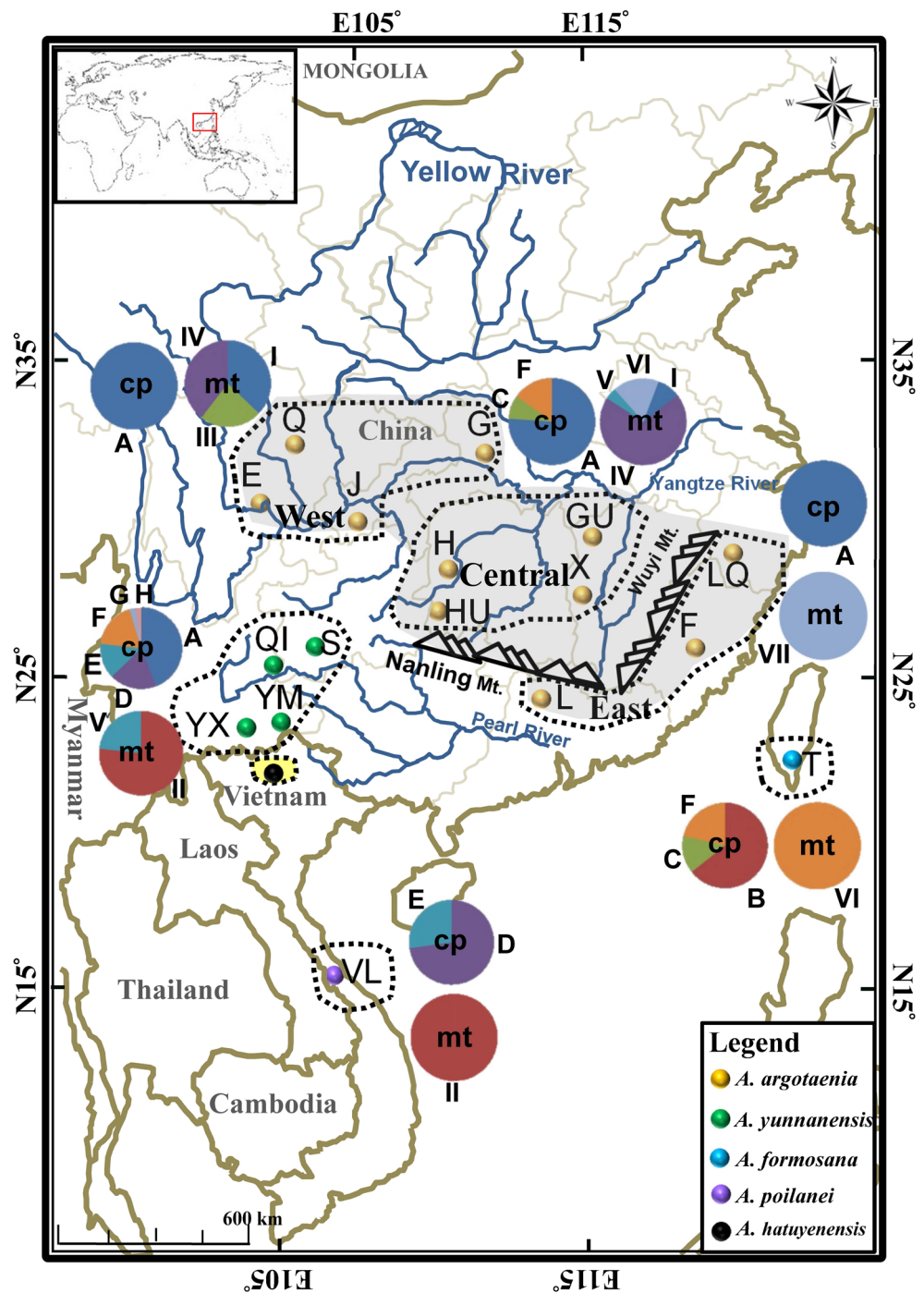
Phylogenetic and Phylogeographic Analyses

Nucleotide sequences were aligned with the program BlastAlign (Belshaw and Katzourakis 2005). Indels were coded as binary characters with SeqState version 1.4.1 (Müller 2005). Maximum-likelihood (ML) analyses were

Table 1 Localities and sample numbers of the 17 investigated populations of *Amentotaxus argotaenia* complex

Species	Location	Population symbol	Latitude (N)	Longitude (E)	Sample size (<i>n</i>)	
<i>A. argotaenia</i>	East region	Datian, Fujian	F	25°40'	117°48'	15
		Longquan, Zhejiang	LQ	28°13'	119°00'	11
		Lianshan, Guangdong	L	24°33'	112°01'	9
	Central region	Suining, Hunan	H	26°29'	110°10'	12
		Guanshan, Jiangxi	GU	28°25'	114°30'	14
		Yongxin, Jiangxi	X	26°56'	114°30'	10
		Bamianshan, Hunan	HU	28°02'	109°47'	10
	West region	Xingshan, Hubei	G	31°05'	110°55'	11
		Jinfoshan, Chongqing	J	29°00'	107°10'	10
		Emeishan, Sichuan	E	29°33'	103°20'	12
Dujiangyan, Sichuan		Q	31°2'	103°35'	10	
<i>A. formosana</i>	Chinshuiying, Taiwan	T	22°23'	120°46'	14	
<i>A. yunnanensis</i>	Xingyi, Guizhou	QI	24°50'	104°45'	9	
	Anlong, Guizhou	S	25°11'	105°30'	10	
	Malipo, Yunnan	YM	23°16'	104°58'	12	
	Xichou, Yunnan	YX	23°16'	104°13'	12	
<i>A. poilanei</i>	Lao Va Chai Municipality, Vietnam	VL	23°06'	105°04'	11	

Fig. 1 Map showing distributions and frequencies of clades of organellar DNA in the populations of *Amentotaxus* species. Clades for cpDNA: dark blue A, brown B, green C, purple D, blue E, orange F, light blue G, light orange H; clades for mtDNA: dark blue I, brown II, green III, purple IV, blue V, orange VI, light blue VII. For abbreviations of populations see Table 1. *A. hatuyenensis* was not included in this study



performed using PHYLIP v. 3.67 (Felsenstein 2007). Confidence of the lineages reconstructed was tested by bootstrapping (Felsenstein 1985) with 1,000 replicates using unweighted characters. Genetic distances between haplotypes were calculated based on the transition/transversion ratio using MEGA, version 5.1 (Tamura et al. 2011), and were used to construct a minimum spanning network with MINSPNET software (Excoffier and Smouse 1994).

Population Genetic Analyses

To investigate the genetic structure of *Amentotaxus* populations, several genetic analyses within and between populations were conducted. Genetic analyses within and between populations were conducted for cp- and mt-DNA sequences. The two cpDNA fragments were combined, and treated as a single marker. Statistics such as the number of segregating

sites (S), and genetic diversity (Hd) were determined (Nei 1987). The level of genetic diversity within populations was quantified by measures of nucleotide divergence, θ (Watterson 1975) and π (Tajima 1989), using DnaSP (Rozas et al. 2003). In order to make inferences about demographic dynamics of *Amentotaxus*, we employed both mismatch distributions (Slatkin and Hudson 1991) and statistical tests of neutrality. We calculated Tajima's D (Tajima 1989), Fu and Li's D* statistic (Fu and Li 1993), and Fu's Fs statistics (Fu 1997). Mismatch distributions have a unimodal pattern when the population expands from a single source or recovers from a bottleneck; mismatch distributions are multimodal in populations with a stable effect size (Rogers and Harpending 1992). A hierarchical analysis of molecular variance (AMOVA, Excoffier et al. 1992) was conducted to summarize the variation among and within populations with the computer program Arlequin (Excoffier et al. 2005). The Φ statistics were computed using haplotype frequencies alone, and the significance of departures from zero of Φ statistics and genetic variance components was tested using 10,000 permutations. Patterns of geographical subdivision were estimated hierarchically using DnaSP. Statistical significance of Φ_{st} estimates was assessed using software Arlequin with permutations. The differentiation parameter of G_{st} was also estimated for the total data set and for each species using PerMut 1.0 (Pons and Petit 1996).

The xmf2struct program (<http://www.xavierdidelot.xtremhost.com/clonalframe.htm>) was used to convert sequence data of cpDNA and mtDNA into genotype data for HP-RARE 1.0 (Kalinowski 2005) and FSTAT V2.9.3.2 (Goudet 1995). HP-RARE 1.0 was used to calculate standardized haplotype richness (H_t) and private allelic richness (H_p), equalizing to a sample size using rarefaction. The permutation tests of significance for allelic richness were implemented in FSTAT V2.9.3.2.

A Bayesian estimate of group delimitation and the ages of TMRCA of the *Amentotaxus* clades was obtained using *BEAST (Melo-Ferreira et al. 2012) and BEAST v. 1.6 (Drummond et al. 2006). For estimating divergence between populations or species, a well-documented evolutionary rate is needed. The fossil record of the Taxads extends from the Lower Jurassic to the present (Stewart and Rothwell 1993). Even so, it is difficult to assess the divergence times using the fossil record. In this study, evolutionary rates for cp DNA and mtDNA have been shown to reach $1.10\text{--}2.90 \times 10^{-9}$, and $2.00\text{--}3.00 \times 10^{-10}$ substitutions per site per year (Wolfe et al. 1987; Chiang et al. 2006). Posterior estimates of age of the TMRCA were obtained by BEAST v. 1.6, with samples drawn every 500 steps over a total of 1,000,000 steps. Adequate sampling and convergence to the stationary distribution were checked using TRACER v. 1.6 (Rambaut and Drummond 2004). Posterior estimates of parameters were all found to be

distinctly unimodal (although with wide 95 % highest posterior densities), and all parameters appeared to be identifiable, despite the relatively low information content in the sequences and the small age range of the sequences.

For microsatellite loci, the genetic diversity within species/populations was estimated by calculating allele number, observed, and expected heterozygosities of microsatellites with Arlequin (Excoffier et al. 2005). The program was also performed to calculate population differentiation, and molecular variance. The standardized allelic richness (A_r) and private allelic richness (A_p), with significance for allelic richness via permutation tests, and differentiation parameters (G_{st}) were assessed using programs HP-RARE 1.0, FSTAT V2.9.3.2, and PerMut 1.0. Program STRUCTURE version 2.1 (Pritchard and Wen 2003) applies a Bayesian method to infer the number of clusters (K) without using prior information of individual sampling locations. Program STRUCTURE v. 2.1 (Pritchard and Wen 2003) applies a Bayesian method to infer the number of clusters (K=1–17) without using prior information of individual sampling locations and was estimated by the Markov chain Monte Carlo (MCMC) method with 20 separate runs for each number of clusters to evaluate the consistency of the results. Each run was pursued for 1,000,000 MCMC interactions, with an initial burn-in of 100,000 and an ancestry model that allowed for admixture. The final posterior probability of K, $Pr(X|K)$, was computed using the runs with highest probability for each K (Pritchard et al. 2000). However, as indicated in the STRUCTURE documentation, $Pr(X|K)$ usually plateaus or increases slightly after the 'right' K is reached. Thus, following Evanno et al. (2005), ΔK , where the modal value of the distribution is located at the real K, was calculated by STRUCTURE HARVESTER v. 0.6.8 (Earl and vonHoldt 2012).

IM Estimation of Demographic Dynamics

To determine the demographic history that the species experienced, we used the simulation program IMA (Hey and Nielsen 2004) to investigate the isolation with migration model in microsatellites and organelle DNAs. Using coalescent simulations and Bayesian computation procedures, IM yields six demographic parameters, including the divergence time (t), bidirectional migration rates (m_1 and m_2), and the effective population sizes of the ancestral (θ_A) and descendent populations (θ_1 and θ_2). The stepwise mutation model (SMM) and Hasegawa-Kishino-Yano (HKY) models were applied to microsatellite and organelle sequence data, respectively. The posterior probability densities of these parameters are generated by MCMC simulations, and simulations were run with individual simulations being updated 100,000 times. Within each simulation, we used the procedure to swap among ten heated chains (Metropolis coupling) and observed sufficient swapping rate while the simulation was running. These

simulations were carried out using ten independent runs, with each chain started at a different starting point and initiated with a burn-in period of 100,000 updates. Convergence upon the stationary distribution was assessed by estimating the effective sample size (ESS) for the parameters, based on the autocorrelation of parameter values measured over the course of the run. The analysis was considered to have converged upon a stationary distribution if the independent runs generated similar posterior distributions with a minimum ESS of 100. Each simulation yielded a marginal density histogram for the six parameters of interest. The peaks of the resulting distributions were considered as the maximum likelihood estimates (MLE) of the parameter with credibility intervals equaling the 95 % highest posterior density (HPD) intervals. Demographic parameters were scaled using the neutral mutation rate (u). For loci of cpDNA and mtDNA, evolutionary rates have been shown to reach $1.10\text{--}2.90 \times 10^{-9}$, and $2.00\text{--}3.00 \times 10^{-10}$ substitutions per site per year (Wolfe et al. 1987; Chiang et al. 2006). Then we used the geometric mean ($u = 4.55 \times 10^{-7}$ substitutions/locus/year) based on the above rate for rescaling parameters. For microsatellites, 6.3×10^{-4} mutations per locus were applied (O'Connell and Ritland 2004). The effective population size for ancestral and present times were estimated by $N=(\theta/2u)$ and $N=(\theta/4u)$ for organelle and SSR data.

Results

Sequence Variation and Genetic Diversity

We examined organellar DNA genetic diversity in the *Amentotaxus* species complex (Tables 2–3). In total, 38 haplotypes (C1–C38) of the cpDNA were identified (GenBank accession numbers: HG424279–HG424291 for *atpI-atpH* spacer; HG424292–HG424316 for *rpS16-trnK* spacer), only five of which (C5, 9, 10, 23, and 24) were shared among species. Twenty-six mtDNA haplotypes were identified (M1–M26, GenBank accession numbers: EF418631–EF418656), and almost all were private, i.e., none were shared among species (Fig. 2b).

Population level genetic diversity (Hd) based on cpDNA was 0.78, and nucleotide diversity (θ) was 4.00×10^{-3} . *Amentotaxus yunnanensis* exhibited the highest nucleotide diversity ($\theta=3.21 \times 10^{-3}$), followed by *A. formosana* ($\theta=2.57 \times 10^{-3}$), *A. poilanei* ($\theta=1.86 \times 10^{-3}$), and *A. argotaenia* ($\theta=1.75 \times 10^{-3}$). The *A. argotaenia* population sampled from Yongxin had the highest nucleotide diversity; and *A. yunnanensis* collected from Xichou exhibited the highest nucleotide diversity level (Table 2). The standardized haplotype richness (Hr) and private haplotype richness (Hp) of populations ranged from 1.00 to 1.38 and 0.00 to 0.21,

respectively. Higher private allelic richness was found in *A. formosana*, and *A. yunnanensis* (Table 2). The mtDNA population genetic diversity (Hd) was 0.91, and nucleotide diversity (θ) was 7.86×10^{-3} . *Amentotaxus argotaenia* possessed the highest nucleotide diversity level ($\theta=6.03 \times 10^{-3}$), followed by *A. yunnanensis* ($\theta=2.10 \times 10^{-3}$), and *A. formosana* ($\theta=1.40 \times 10^{-3}$), while no variation was detected in *A. poilanei*. *Amentotaxus argotaenia* exhibited 16 of 22 haplotypes private to populations with the highest private allelic richness ($Hp=0.33$), while three haplotypes were restricted to a single *A. formosana* population ($Hp=0.17$). *Amentotaxus argotaenia* from Yongxin had the highest nucleotide diversity and haplotype richness, whereas the Anlong population of *A. argotaenia* was fixed at haplotype M26 (Table 3). No significant differences in the allelic richness were detected among species ($P=0.19$, cpDNA; $P=0.43$, mtDNA).

Gene Genealogy of CpDNA and MtDNA Haplotypes

A maximum likelihood tree was reconstructed based on cpDNA haplotypes, and eight major lineages (A–H) were identified. Haplotype data indicated paraphyly in all species in the complex. No distinct geographical subdivisions were detected among lineages. A dominant basal haplotype C5 (46.35 %) was resolved in Cluster A, which occurred exclusively in *A. argotaenia* and *A. yunnanensis* (Fig. 2a). A minimum spanning network was also reconstructed based on mutational changes. Eight major lineages (A–H) were identified, and lineages C, D, and G were nested as the interior node, independently linked to A, B, E, F, and H (Fig. 3a).

Seven major mtDNA lineages (I–VII) were identified (Fig. 2b), with I–V forming a cluster, and sister to VI+VII. Lineages II and VI occurred exclusively in *A. formosana*, and *A. yunnanensis* and *A. poilanei*, respectively; while the remainder occurred in *A. argotaenia*. *A. yunnanensis* and *A. poilanei* were nested within *A. argotaenia* (Fig. 3b). Furthermore, *A. argotaenia* and *A. yunnanensis* were respectively paraphyletic and polyphyletic in genetic composition; monophyly was observed in *A. formosana* and *A. poilanei*. *Amentotaxus argotaenia* was monophyletic in populations distributed in the East region, included in lineage VII; while paraphyletic in both Central and West regions. A minimum spanning network of mtDNA, comprised of two major lineages I+II+III+IV+V and VI+VII, corresponded to clusters of the maximum likelihood tree (Fig. 3b).

The *BEAST population tree based on the combined organelle makers in the species complex identified two major groups, i.e., *A. argotaenia* vs. other three species (Supplementary Fig. 1). Within the *A. argotaenia* group, geographical subdivision nevertheless was not supported. Admixture of haplotypes across geographical populations, plus a low posterior possibility (Pp=0.61) for the internal

Table 2 Genetic variability of cpDNA within *Amentotaxus* populations and within geographical regions/species. *Hr* haplotype richness, *Hp* private haplotype richness

Species, region: population (symbol)	h	Hr	Hp	Hd	$\pi (\times 10^{-3})$	$\Theta (\times 10^{-3})$	Fu & Li's D*	Tajima's D	Mismatch analyses
<i>A. argotaenia</i>	15	1.21	0.08	0.60	0.94	1.75	-0.11	-1.25	Unimodal
East region:	6	1.00	0.00	0.59	0.48	0.61	-0.54	-0.54	Equilibrium
Datian (F)	5	1.00	0.00	0.64	0.48	0.77	-0.60	-1.22	Unimodal
Longquan (LQ)	2	1.00	0.00	0.18	0.11	0.21	-1.29	-1.13	Equilibrium
Lianshan (L)	1	1.00	0.00	0.00	0.00	0.00	-	-	-
Central region:	12	1.26	0.05	0.80	1.73	2.01	-0.35	-0.43	Unimodal
Suining (H)	3	1.00	0.00	0.59	0.88	0.83	-0.46	0.18	Bimodal
Guanshan (GU)	3	1.05	0.02	0.28	0.18	0.40	-1.83	-1.48	Equilibrium
Yongxin (X)	5	1.00	0.00	0.76	0.73	1.11	-1.35	-1.39	Unimodal
Bamianshan (HU)	3	1.05	0.05	0.51	0.35	0.45	-0.28	-0.69	Equilibrium
West region:	4	1.05	0.02	0.26	0.17	0.44	-0.37	-1.31	Equilibrium
Xingshan (G)	2	1.03	0.03	0.44	0.27	0.21	0.78	0.67	Equilibrium
Jinfoshan (J)	2	1.00	0.00	0.20	0.13	0.22	-1.24	-1.11	Equilibrium
Emeishan (E)	3	1.00	0.00	0.32	0.21	0.42	-1.72	-1.45	Equilibrium
Duijiangyan (Q)	1	1.00	0.00	0.00	0.00	0.00	-	-	-
<i>A. formosana</i>									
Chinshuiying (T)	8	1.33	0.21	0.86	2.78	2.57	0.14	0.33	Bimodal
<i>A. yunnanensis</i>	16	1.35	0.18	0.88	2.16	3.21	-1.89	-1.08	Unimodal
Xingyi (QI)	1	1.00	0.00	0.00	0.00	0.00	-	-	-
Anlong (S)	1	1.00	0.00	0.00	0.00	0.00	-	-	-
Malipo (YM)	6	1.23	0.01	0.80	1.18	1.87	-1.62	-1.50	Unimodal
Xichou (YX)	10	1.38	0.18	0.96	2.18	3.12	-1.27	-1.29	Bimodal
<i>A. poilanei</i>									
Lao Va Chai Municipality (VL)	4	1.10	0.04	0.75	0.71	1.86	-0.40	-0.64	Unimodal
Overall	38	-	-	0.78	2.05	4.00	-0.88	-1.42	Unimodal

* $P < 0.05$

node of coalescence, indicated effects of ancestral lineage sorting or occurrence of gene flow.

Genetic Diversity at Microsatellite Loci

The species complex was polymorphic at all 15 microsatellite loci, and the number of alleles for each locus ranged from 1.93 to 5.47 (Table 4). *Amentotaxus argotaenia* exhibited the highest number of alleles per locus, while the lowest number was resolved in *A. poilanei*. The eastern geographic region had a higher number of alleles per locus than other regions within *A. argotaenia*. Observed (H_o) and expected (H_e) heterozygosity varied respectively from 0.05 to 0.29, and 0.24 to 0.61 (Table 4). The standardized allelic richness (Ar) and private allelic richness (Ap) of populations ranged from 1.91 to 4.90 and 0.00 to 0.65, respectively. Higher private allelic richness was found in *A. argotaenia*, and *A. formosana*. Overall, four loci deviated from Hardy-Weinberg expectations in *A. formosana*, *A. yunnanensis*, and *A. argotaenia* in all populations due to heterozygote deficiencies, the exception

being *A. poilanei*. At the population level, heterozygote deficiencies were also detected in most populations. Like organellar DNA, no significant differences of allelic richness among species was detected in microsatellites ($P=0.08$).

Subsequently, data were analyzed to examine genetic structure at a finer resolution using STRUCTURE (Pritchard et al. 2000). The best, second, and third fit numbers of groupings were inferred as 2, 3, and 6 based on ΔK , i.e. $\Delta K=8.46$ at $K=2$, $\Delta K=3.64$ at $K=3$, and $\Delta K=2.84$ at $K=6$, based on Bayesian clustering analyses on microsatellites (Fig. 4a). At $K=2$, the *A. argotaenia* species complex was divided into two differentiated groups; *A. formosana* and *A. argotaenia* were allied in one cluster, and *A. yunnanensis* and *A. poilanei* were grouped in the other cluster (Fig. 4b). Each individual possessed a high percentage of component 1 (green segment in Fig. 4b) or component 2 (blue segment in Fig. 4b), with the exception of 11 *A. argotaenia* individuals, which showed > 20 % of component 2, revealing genetic admixture composition. Similarly, at $K=3$, *A. formosana* and *A. argotaenia* were separated from an *A. yunnanensis* and *A. poilanei* cluster

Table 3 Genetic variability of mtDNA within *Amentotaxus* populations and within geographical regions/species

Species, region: population (symbol)	h	Hr	Hp	Hd	$\pi (\times 10^{-3})$	$\Theta (\times 10^{-3})$	Fu & Li's D*	Tajima's D	Mismatch analyses
<i>A. argotaenia</i>	19	1.64	0.33	0.91	9.65	6.03	1.41	1.57	Bimodal
East region:	3	1.06	0.03	0.51	0.87	0.74	0.83	0.39	Bimodal
Datian (F)	1	1.00	0.00	0.00	0.00	0.00	–	–	–
Longquan (LQ)	1	1.00	0.00	0.00	0.00	0.00	–	–	–
Lianshan (L)	2	1.03	0.03	0.39	0.55	0.52	0.84	0.16	Equilibrium
Central region:	10	1.56	0.06	0.73	7.81	6.23	1.57*	0.89	Bimodal
Suining (H)	4	1.07	0.03	0.78	1.49	1.04	1.06	1.50	Unimodal
Guanshan (GU)	1	1.00	0.00	0.00	0.00	0.00	–	–	–
Yongxin (X)	6	1.47	0.03	0.93	9.15	7.60	1.55*	1.05	Bimodal
Bamianshan (HU)	2	1.00	0.00	0.53	0.75	0.50	0.80	1.30	Unimodal
West region:	13	1.14	0.11	0.82	2.68	2.80	0.68	–0.12	Unimodal
Kingshan (G)	2	1.03	0.03	0.22	0.31	0.52	–1.190	–1.09	Equilibrium
Jinshoshan (J)	2	1.00	0.00	0.56	0.78	0.52	0.84	1.40	Unimodal
Emeishan (E)	3	1.06	0.07	0.64	1.02	1.04	–0.22	–0.06	Unimodal
Dujiangyan (Q)	2	1.03	0.03	0.54	0.76	0.54	0.89	1.17	Bimodal
<i>A. formosana</i>									
Chinshuiying (T)	3	1.10	0.17	0.53	1.94	1.40	1.11	1.27	Bimodal
<i>A. yunnanensis</i>	4	1.17	0.04	0.75	2.49	2.10	1.64*	0.12	Bimodal
Xingyi (QI)	2	1.00	0.00	0.56	0.78	0.52	0.84	1.40	Unimodal
Anlong (S)	1	1.00	0.00	0.00	0.00	0.00	–	–	–
Malipo (YM)	2	1.00	0.00	0.36	0.50	0.50	0.80	0.02	Equilibrium
Xichou (YX)	2	1.03	0.03	0.36	0.50	0.50	0.80	0.02	Bimodal
<i>A. poilanei</i>									
Lao Va Chai Municipality (VL)	1	1.00	0.00	0.00	0.00	0.00	–	–	–
Overall	26	–	–	0.91	10.52	7.86	1.65*	0.99	Bimodal

* $P < 0.05$

Fig. 2a,b Unrooted maximum likelihood tree of haplotypes in *Amentotaxus*. Numbers at nodes represent bootstrap values and divergence times between major lineages. Distribution of each clade is indicated in parentheses: ARG *A. argotaenia*, ARG_(C) Central region, ARG_(E) East region, ARG_(W) West region), FOR *A. formosana*, YUN *A. yunnanensis*, POI *A. poilanei*. **a** cpDNA, **b** mtDNA

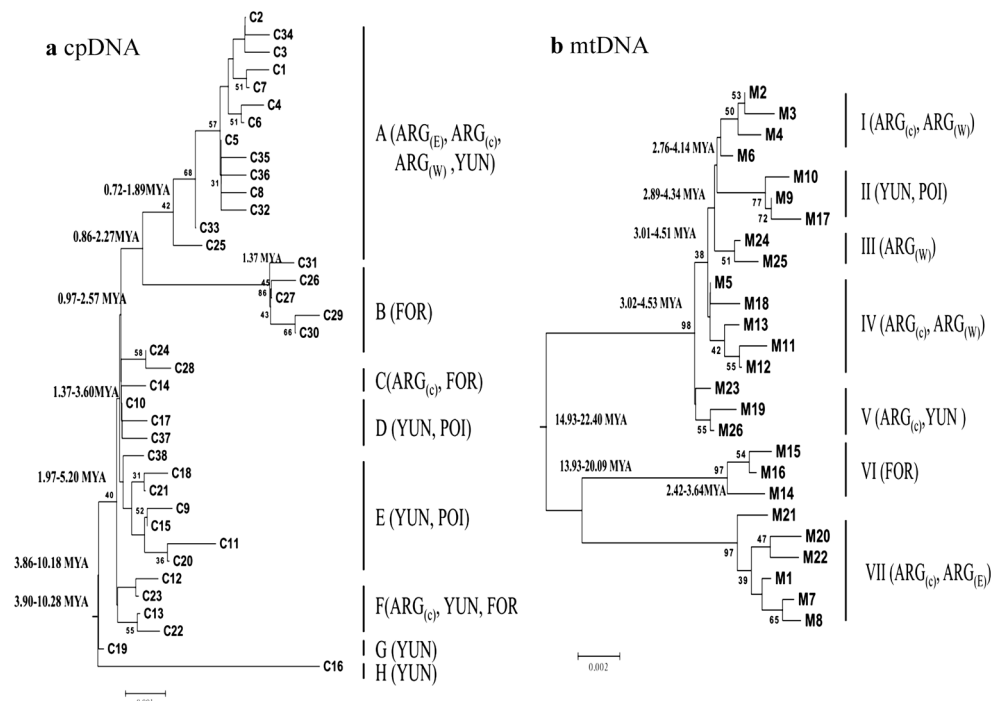
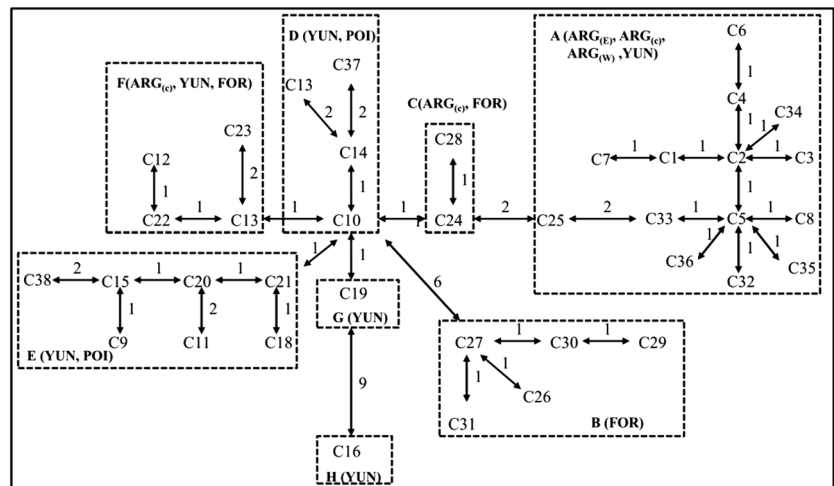
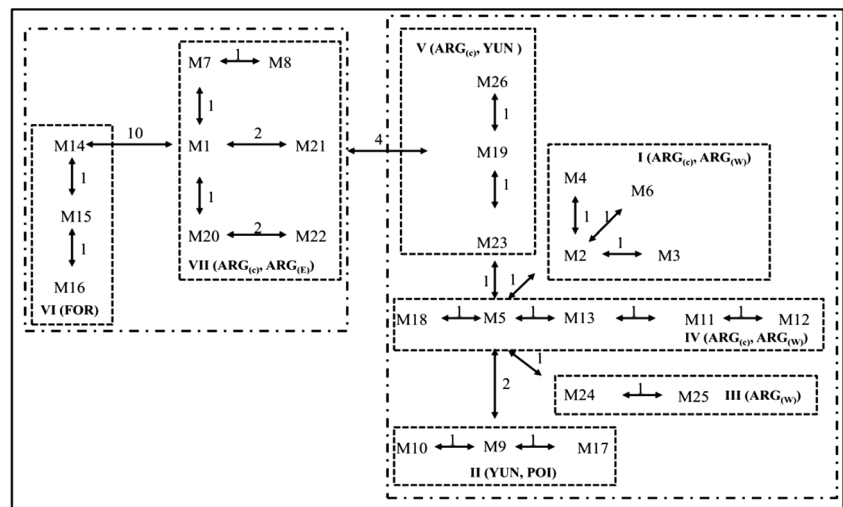


Fig. 3 Minimum spanning network of haplotypes in *Amentotaxus*. Geographical distribution of each haplotype and lineage is indicated. Distribution of each clade is indicated in parentheses: *ARG* *A. argotaenia*, *ARG_(C)* Central region, *ARG_(E)* East region, *ARG_(W)* West region, *FOR* *A. formosana*, *YUN* *A. yunnanensis*, *POI* *A. poilanei*. **a** cpDNA, **b** mtDNA

a cpDNA



b mtDNA



(Fig. 4b); at $K=6$, *A. argotaenia* was divided into four clusters, which did not correspond to geographic subdivisions (Fig. 4b).

Population Genetic Structure and Demographic Dynamics

Hierarchical and nonhierarchical AMOVAs were conducted to determine the spatial apportionment of genetic variants among *Amentotaxus* species and populations (Table 5). The AMOVA of cpDNA revealed only 13.48 % of the total variants within populations ($\Phi_{SC}=0.82$, $P<0.01$) and the largest level of variation (59.87 %) was explained by differences among populations ($\Phi_{ST}=0.87$, $P<0.01$). However, 26.65 % of the total variation was due to differences among species ($\Phi_{CT}=0.27$, $P<0.01$) (Table 5). A hierarchical AMOVA was conducted based on three geographic subdivisions of *A. argotaenia*, i.e., west, central, and east regions. Results

showed 82.54 % of the total variation was among regions, and 17.46 % within populations. Most of genetic differentiation within *A. argotaenia* and *A. yunnanensis* was partitioned among populations, with respective values of 82.54 % and 89.92 %. The mtDNA AMOVA results yielded similar patterns. Results indicated the highest level of variation was among populations (63.49 %, $\Phi_{ST}=0.92$, $P<0.01$), with only 8.22 % within population variation ($\Phi_{SC}=0.89$, $P<0.01$) (Table 5), and 28.30 % of the variance was explained by differences among species ($\Phi_{CT}=0.32$, $P<0.01$). In contrast to organellar DNA loci, AMOVA of microsatellite loci revealed that the largest level of variation (51.13 %) was due to differences within populations ($\Phi_{SC}=0.22$, $P<0.01$) (Table 5), and only 14.68 % of the total variation was among populations within species ($\Phi_{ST}=0.49$, $P<0.01$). Nonetheless, 34.19 % of the variance was explained by differences among species ($\Phi_{CT}=0.34$, $P<0.01$). The estimates

Table 4 Genetic variability of microsatellites within *Amentotaxus* populations and within geographical regions/species. *A* Allele number, *Ar* allelic richness, *Ap* private allelic richness, *Ho* observed heterozygosity, *He* expected heterozygosity, *F_{IS}* fixation index

Species, region: population (symbol)	A	Ar	Ap	Ho	He	F _{IS}
<i>A. argotaenia</i>	9.80	5.48	3.72	0.12	0.61	0.81*
East region:	8.14	5.37	1.11	0.12	0.69	0.82*
Datian (F)	4.00	3.70	0.65	0.08	0.52	0.84*
Longquan (LQ)	3.73	3.62	0.27	0.15	0.46	0.70*
Lianshan (L)	3.60	3.60	0.14	0.13	0.49	0.75*
Central region:	7.13	4.91	0.65	0.11	0.57	0.81*
Suining (H)	3.27	3.20	0.01	0.08	0.49	0.84*
Guanshan (GU)	5.47	4.90	0.04	0.11	0.55	0.82*
Yongxin (X)	2.87	2.83	0.20	0.10	0.38	0.75*
Bamianshan (HU)	3.53	3.49	0.00	0.15	0.52	0.73*
West region:	7.27	4.88	0.56	0.12	0.54	0.78*
Xingshan (G)	2.20	2.19	0.00	0.10	0.33	0.72*
Jinfoshan (J)	3.40	3.34	0.03	0.09	0.42	0.80*
Emeishan (E)	4.93	4.62	0.14	0.11	0.57	0.81*
Dujiangyan (Q)	4.13	4.04	0.00	0.20	0.50	0.61*
<i>A. formosana</i>						
Chinshuiying (T)	3.47	3.32	2.11	0.05	0.61	0.92*
<i>A. yunnanensis</i>	2.73	2.46	0.62	0.23	0.36	0.36*
Xingyi (QI)	2.00	2.00	0.00	0.25	0.30	0.18
Anlong (S)	2.13	2.10	0.09	0.23	0.34	0.335*
Malipo (YM)	2.20	2.18	0.00	0.21	0.31	0.34*
Xichou (YX)	1.93	1.91	0.00	0.24	0.24	0.01
<i>A. poilanei</i>						
Lao Va Chai Municipality (VL)	2.07	2.01	0.52	0.29	0.27	−0.06
Overall	13.00	–	–	0.15	0.71	0.79

* $P < 0.05$

of *G_{st}* for the total data set and for each species group were presented. High levels of *G_{st}* were detected in the total data set (cpDNA: 0.45; mtDNA: 0.64; microsatellites: 0.41), and similar patterns were detected in *A. argotaenia* (cpDNA: 0.40; mtDNA: 0.55; microsatellites: 0.24) and *A. yunnanensis* (cpDNA: 0.55; mtDNA: 0.67; microsatellites: 0.19).

Pairwise comparisons of genetic differentiation between species and between regions were conducted using DnaSP. Significant genetic differentiation was detected between species. *Amentotaxus yunnanensis* was closely related to *A. poilanei*, as indicated by the lowest *F_{st}* values for all loci. *Amentotaxus argotaenia* populations from the Central region were related most closely to populations from the West at all loci but cpDNA (Table 6).

We used mismatch distribution analyses to infer the long-term demographic history of populations. A bimodal mismatch distribution was displayed in populations H, T, and YX for the cpDNA, and populations X, Q, T, and YX for mtDNA. Unimodal distributions were detected in F, X, YM,

and VL for cpDNA, and in H, HU, J, E, and QI populations for mtDNA (Tables 2–3). All loci displayed a bimodal or unimodal mismatch in most regions at the geographical level. The data used to generate mismatch distributions are not independent (Groth and Barrowclough 1999), therefore Tajima's *D*, Fu and Li's *D**, and Fu's *F_s* statistics were applied to detect departures from population equilibrium. In this study, most of Tajima's *D*, Fu and Li's *D** values yielded nonsignificant statistics for the three markers (Tables 2–3). The Bayesian skyline plot was applied to infer past demographic patterns; all three loci revealed a long history of constant population sizes for each species, followed by subsequent demographic expansion (Fig. 5).

Effective Population Size and Gene Flow

The IM program was used to refine six demographic parameters in *Amentotaxus* based on organellar DNA and the microsatellite data matrix. All pairwise

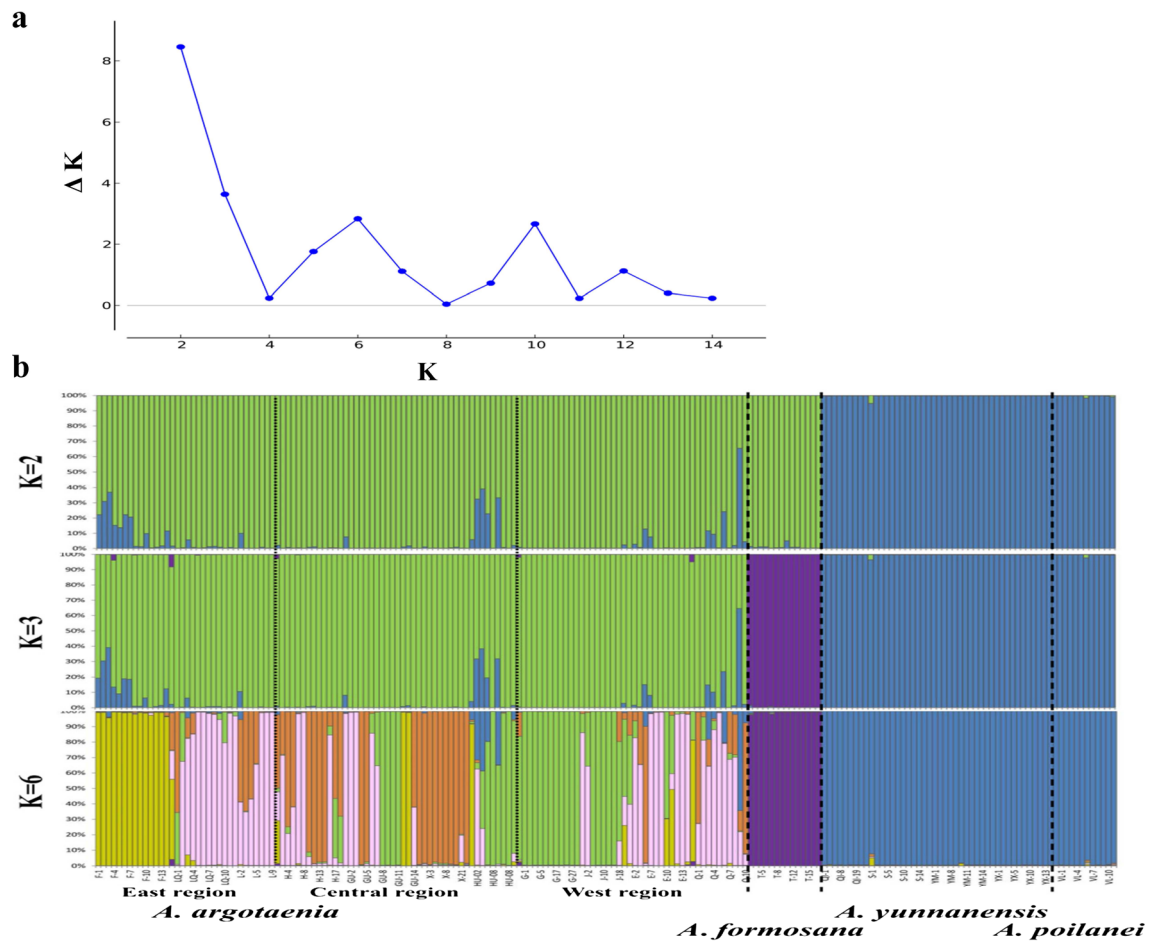


Fig. 4a,b Bayesian inference of the number of clusters (K) of *Amentotaxus* based on microsatellites. **a** The scatter plots of ΔK . The ΔK is based on the rate of change of $\ln P(X/K)$ between successive K values. **b** The two, three, and six clusters ($K=2, 3,$ and 6) detected from STRUCTURE analyses

comparisons indicated that ancestral effective population sizes were larger than the present, suggesting demographic shrinking (Supplementary Table S1). SSR fingerprints estimated an ancestral effective population size hundreds of times greater than the present size. The migration parameters further revealed a peak at the lower resolution limits in one direction, i.e., from *A. yunnanensis* to *A. poilanei*, from *A. argotaenia* to *A. poilanei*, and from *A. formosana* to *A. poilanei* in organellar DNAs. The population migration rates [$M_{12} = N_1 m_{12} = m_{12} * (\theta_1/4)$] estimated from *A. poilanei* to the three other species were substantially lower than vice versa. However, approximating estimates between ancestral and recent gene flow were obtained based on SSRs. Between inland and island species, the estimates of gene flow were not consistent among markers (Supplementary Table S1), revealing extremely low, recent gene flow based on SSRs. Altogether, at the species level, nonzero and asymmetric migrations were detected ever since species divergence.

Molecular Dating

Based on the maximum likelihood reconstructed tree, the node coalescence times were estimated using the BEAST program. For cpDNA, all haplotypes coalesced to approximately 3.90–10.28 MYA, while lineages A+B+C+D+E and F coalesced to 1.97–5.20 MYA. The coalescence of two primary mtDNA lineages (I+II+III+IV+V and VI+VII) in *Amentotaxus* was dated to 14.93–22.40 MYA. Similarly, *A. formosana* lineage VI and *A. argotaenia* lineage VII diverged 13.93–20.09 MYA; while *A. yunnanensis* lineage II and *A. argotaenia* lineage I did not coalesce until recently, approximately 2.76–4.14 MYA (Fig. 2).

Between-species divergence times estimated by organellar DNAs were calculated based on the isolation with migration model using the $T=t/u$ formula. Pairwise comparisons revealed divergence times between species ranging from 1.10 to 13.84 MYA, and divergence between island and inland species ranging from 2.40 to 13.84 MYA (Supplementary Table 1). Divergence times between inland species were

Table 5 AMOVA design and results of the variants in *Amentotaxus* species/populations

Grouping Source of variation	cpDNA			mtDNA			Microsatellites			
	d.f.	Sum of squares	Variance components	Percentage of variation	Sum of squares	Variance components	Percentage of variation	Sum of squares	Variance components	Percentage of variation
Among species	3	256.57	1.40	26.65	254.26	1.40	28.30	525.93	2.18	34.19
Among populations within species	13	464.95	3.15	59.87	458.17	3.13	63.49	316.11	0.94	14.68
Within populations	175	124.22	0.71	13.48	70.96	0.41	8.22	1,211.13	3.27	51.13
<i>A. argotaenia</i>										
Among populations	10	223.20	1.95	82.54	428.24	3.77	88.56	270.11	1.04	22.45
Within populations	113	46.56	0.41	17.46	54.95	0.49	11.44	852.45	3.60	77.55
<i>A. yunnanensis</i>										
Among populations	3	241.74	7.45	89.92	29.93	0.92	84.17	46.01	0.59	20.73
Within populations	39	32.58	0.84	10.08	6.72	0.17	15.83	192.31	2.27	79.27

Table 6 Genetic differentiation between geographical regions and between species. ARG: *A. argotaenia*, FOR: *A. formosana*, YUN: *A. yunnanensis*, POI: *A. poilanei*

	ARG				FOR	YUN	POI
		East	Central	West			
cpDNA\mtDNA							
ARG					0.69	0.39	0.51
East			0.78	0.94			
Central		0.14		0.15			
West		0.20	0.17				
FOR	0.61					0.91	0.97
YUN	0.32				0.39		0.20
POI	0.72				0.47	0.24	
Microsatellites							
ARG							
East							
Central		0.07					
West		0.09	0.03				
FOR	0.38						
YUN	0.37				0.50		
POI	0.40				0.50	0.23	

estimated from 1.10 to 2.31 MYA, with the longest divergence between *A. yunnanensis* and *A. poilanei*.

Discussion

Species Delimitation in *Amentotaxus*

Amentotaxus is an ancient genus with origins that date to the early Tertiary, and this genus diverged from *Torreya* ca. 55 MYA (Leslie et al. 2012). Members of the genus survived Pleistocene climatic oscillations, and must have experienced range contractions, species extinctions, and subsequent diversification. Molecular dating revealed a long period of isolation among *Amentotaxus* species. Monophyletic species are therefore expected. However, cp- and mtDNAs revealed para- and polyphyletic *Amentotaxus* species (Fig. 2). The absence of reciprocal monophyly might result from lineage sorting (Chiang and Schaal 2006; Chiang et al. 2009), or hybridization/introgression (Chiang et al. 2001). Stochastic effects of genetic drift might have also resulted in the systematic inconsistencies between gene trees at cp- and mtDNA markers (Chiang et al. 2001, 2009).

Hennigian cladists would recognize the four taxa as a single species, as reciprocal monophyly is the only criterion for grouping taxa (Dias et al. 2005; Potter and Freudenstein 2005). Nevertheless, Chiang and Schaal (2006) indicated that

paraphyly/polyphyly is a rule rather than an exception for the plant species complexes of East Asia, and is not even rare between/among well-defined species. In addition, gymnosperm species with East Asian geographic divisions revealed shared haplotypes due to ancient ancestral polymorphisms (Chiang et al. 2006, 2009). Various approaches to species delimitations have been developed. Wiens and Penkrot's (2002) methodology serves to identify species exclusivity instead of reciprocal monophyly, where the term "exclusive" was used to describe haplotype phylogenies. If the haplotypes were not exclusive within species, and instead paraphyletic or polyphyletic with respect to other species, the focal species would likely represent multiple species. In the present study, mtDNA genetic analyses, which showed stronger geographic structuring and deeper divergence in phylogenetic gene tree clustering than other loci, was adequate to assess species delimitations and phylogeographical patterns (Fig. 2b). Case study of the Pinaceae species in central and southern China also suggested that maternally inherited mtDNA markers are more prone to introgression than paternally inherited cpDNA markers, and the latter therefore provide better resolution when delimitating species (Zhou et al. 2010; Wang et al. 2011; Peng et al. 2012). The populations of *A. formosana* were relatively unique, with no genetic compositions shared

with other species (Figs. 2b and 4b). Lineage VI was detected in *A. yunnanensis* and *A. poilanei*; however the H13 haplotype nested in *A. argotaenia* (lineage III) indicated polyphyletic origins of *A. yunnanensis*. The assignment test of microsatellite fingerprints also indicated that genetic compositions of *A. yunnanensis* tended to be similar to *A. poilanei* (Fig. 4b). Following the Wiens-Penkrot methodology, *A. formosana* represents a well-defined species. In contrast, *A. argotaenia*, *A. yunnanensis*, and *A. poilanei* might possibly represent identical species. Significant genetic differentiation between species revealed by STRUCTURE analyses of microsatellite fingerprints (Fig. 4), and *BEAST population tree of organelle DNA (Supplementary Fig. 1), consistent with ISSR fingerprinting (Ge et al. 2005), further supported *A. argotaenia* and *A. yunnanensis* represent independent species; however, data support the recognition of *A. yunnanensis* and *A. poilanei* as one species.

Divergence times from 1.10 to 13.84 MYA among the four allopatric species (Supplementary Table S1) coincided with late Tertiary and Quaternary glaciations (Wang et al. 2013). The complex landscapes and fragmented habitats had significant effects on spatial and temporal patterns of genetic polymorphisms (Hewitt 1996), and speciation in *Amentotaxus*. In addition, divergence between *A. formosana* in Taiwan, and

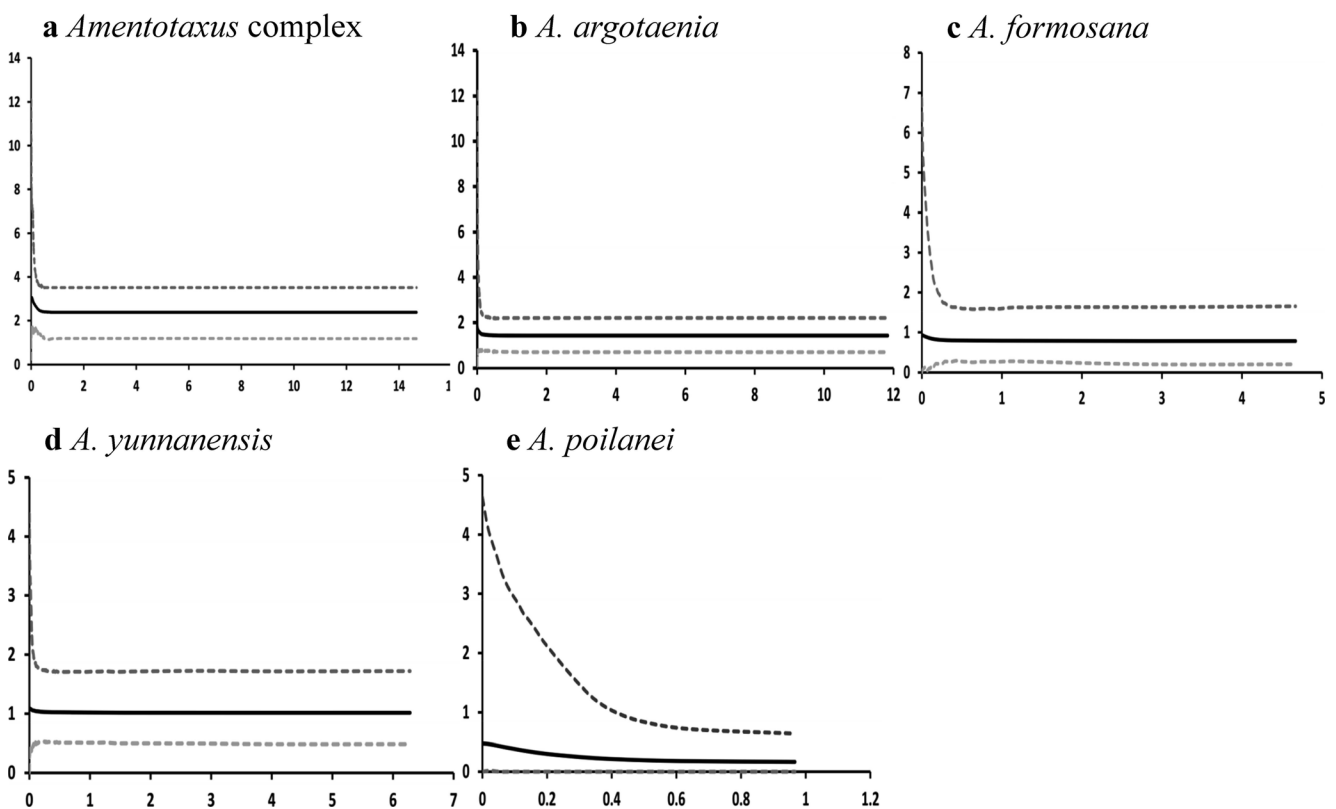


Fig. 5a–e Bayesian skyline plots combining organelle DNA datasets for the effective population size fluctuation through time. *Y*-axes Population size \times generation time; *x*-axes time [years before present (million years

ago)]. **a** *Amentotaxus* complex, **b** *A. argotaenia*, **c** *A. formosana*, **d** *A. yunnanensis*, **e** *A. poilanei*. Solid lines Median estimations; area between gray dashed lines 95 % confidence intervals

A. argotaenia can be traced back 2.40 MYA, a time period that approximates the coalescence of haplotypes resolved in *A. formosana*, suggesting that genetic divergence following colonization in the island may have occurred. The progenitor–derivative speciation scenario also indicated that island species *Picea morrisonicola* derived from continental species *P. wilsonii* within the more recent past (Zou et al. 2013). In this study, significant genetic differentiation was detected between *A. argotaenia* and *A. formosana* at all markers (Table 6), including ISSR fingerprints ($F_{st}=0.761 - 0.850$, Ge et al. 2005). The facts no mtDNA haplotypes were shared among species, and private cpDNA haplotypes were detected within species decreased the possibility of recurrent interspecific gene flow.

Apparently, cpDNA provided evidence for species paraphyly, and the maintenance of long-term ancestral polymorphisms within species. Such a phenomenon, regulated by random genetic drift across population fragments, often occurred in once widespread relict species in Pliocene forests, which subsequently became fragmented during the Quaternary. Vendramin et al. (2000) reported population-specific haplotypes as a general attribute in long diverged conifer species. Furthermore, recent population fragmentation accelerated genetic differentiation among populations, and fixation of alleles within populations/species.

In contrast to the inferred long divergence time between *A. argotaenia* and *A. formosana*, divergence between *A. argotaenia*, *A. yunnanensis*, and *A. poilanei* was more recent, between 1.10 to 2.31 MYA in the inland species. The three mainland species exhibit high morphological similarities; however different evolutionary processes can serve to explain the shared character traits. Only one haplotype (M9) was shared by *A. poilanei* and *A. yunnanensis*, indicating a descent-descendent relationship. A polyphyletic origin of *A. yunnanensis* suggests the divergence time was overestimated, with three haplotypes (M9, 10 and 17) representing lineage II and one haplotype (M26) in lineage V (Fig. 2b). Lineage II, including *A. yunnanensis* was sister to lineage I, which included *A. argotaenia* populations. Accordingly, the coalescence of Yunnan's haplotypes was even more recent. In contrast, the phylogenetically distant haplotype M26 likely represents immigrants from population X (M19 and 23) of *A. argotaenia* (Fig. 3b) (cf. Matos and Schaal 2000). All *A. yunnanensis* populations (including S) showed evidence of clustering based on ISSR (Ge et al. 2005) and microsatellite fingerprinting, plus mismatch analyses displaying a bimodal distribution, which supports a scenario of two lineages merging within *A. yunnanensis* due to obtaining the foreign M26 haplotype. This hybrid origin hypothesis, nevertheless, remains to be tested.

Recurrent gene flow via seeds and pollen, between *A. yunnanensis* and *A. argotaenia* is constrained by geographical barriers and ecological preferences, as indicated by

significant genetic differentiation based on all markers in this study, and ISSR fingerprinting at the species level (Ge et al. 2005). The strong genetic structure at the species level (Fig. 4), and extreme low levels of recent migration (Supplementary Table S1) inferred from microsatellites suggest again that historical gene flow was likely between the species. Providing a low possibility of recurrent genetic exchanges, coancestry instead accounts for the lower level of genetic differentiation. Accordingly, morphological similarities in *Amentotaxus* are likely attributable to high developmental constraints. Nevertheless, diversification and allopatric distribution in *Amentotaxus* do not guarantee a model of allopatric speciation. Nonzero and asymmetric gene flow among species inferred from the three organellar markers revealed that gene flow lasted for a certain period following species divergence, particularly given historical connectivity among species (Wang et al. 2000). This finding further confirms the recognition of paraphyletic species in a recently evolving species complex.

Low Genetic Diversity within Species versus High Genetic Differentiation between Species/Populations

Organellar DNA genetic variation in the *Amentotaxus* complex was low compared with many other species (Chiang et al. 2004, 2009; Ge et al. 2012). Similarly, for microsatellite loci, all populations/species in this complex exhibited lower observed heterozygosity than other conifers (Moriguchi et al. 2009; Parchman et al. 2011). Rare species tend to possess low levels of genetic diversity due to stochastic losses of genetic polymorphisms resulting from genetic drift. Low levels of genetic variability at all loci examined suggested demographic fragmentation caused the notable loss of genetic variation. Nevertheless, the loci did not show congruent variability. For example, *A. argotaenia* possessed the highest levels of mtDNA variation and microsatellite allele number, while having the lowest level of cpDNA diversity (Tables 2–4). In contrast, there were no significant differences among species for the haplotype richness for all loci ($P=0.19$, cpDNA; $P=0.43$, mtDNA; $P=0.08$, microsatellites). The larger numbers of private alleles led *A. argotaenia* to have higher levels of mtDNA and microsatellites variations than cpDNA (Tables 2–4). These results indicated stochastic effects of genetic drift.

Hierarchical AMOVA revealed significant genetic differentiation between species and among geographical regions/populations in this complex (Tables 5–6), a pattern congruent with the population structuring based on ISSR fingerprinting (Ge et al. 2005). The reconstructed phylogeographical patterns suggested colonization likely occurred at the time of Taiwan's formation via long distance dispersal in *A. formosana* (Lin 1966) from F or LQ populations of the East region that are geographically near the Taiwan

population, and characterized by fixation of the M1 haplotype, closely related to M14 (Figs. 2–3). Geological evidence shows Taiwan was connected to the Asian landmass by a land bridge during the LGM as sea levels dropped to 120 m lower than present (Liu 1988). The land bridge enhanced gene flow in many plants (Chiang and Schaal 2006). Nevertheless, in this study, results suggested gene flow between populations of the mainland and Taiwan during the Holocene were very restricted or unlikely to have occurred. The vicariant event was the formation of the Taiwan Strait, which we can conclude prevented gene flow between mainland and island populations. Subsequently, as no foreign alleles were introgressed into Taiwan species, many ancestral polymorphisms were stochastically lost and replaced by newly mutated haplotypes, resulting in monophyly within *A. formosana* and its divergence from *A. argotaenia* in mtDNA.

Spanning a large distributional range, *A. argotaenia* exhibited the highest genetic diversity levels in the species complex, evidenced by two major mtDNA lineages I+II+III+IV+V and VI+VII, which diverged about 22.4 MYA (Fig. 2b). Furthermore, geographical subdivision, i.e., mtDNA clades I+II+III+IV+V of West and Central regions, and VI+VII of the East region and population X (Central region), also contributed to high diversity levels. The divergence times between *A. argotaenia* and other three species ranging from 1.10 to 2.39 MYA indicated a large effective population size of widespread species based on IMA analysis (Supplementary Table 1). Nevertheless, most populations today remain small in the wild, and possess low levels of nucleotide diversity (Table 3). Data suggested habitat fragmentation triggered genetic differentiation among populations. Even so, only three populations of *A. argotaenia* were fixed at a single allele, while most populations were polymorphic, a pattern also occurring in *A. formosana* and *A. yunnanensis*. In contrast, *A. poilanei* was fixed at a single allele, likely resulting from stochastic losses. Furthermore, limited geographic distribution of low-frequency alleles represents footprints of recent demographic expansion (Chiang et al. 2006).

A general assumption inherent to most phylogeographic studies of postglacial colonization is that glacial refugia are expected to harbor more genetic diversity than recolonized areas (Comes and Kaderait 1998; Hewitt 1996, 1999, 2000; Petit et al. 2003). Rapid range expansions out of refugia involving serial population bottlenecks in founder events are expected to exhibit decreasing genetic diversity, particularly in allele or haplotype number, as sampling extends away from source populations (Hewitt 1996). Increased genetic diversity in refugia often reflects stable population dynamics and large population sizes (Hewitt 1996). Therefore, populations with high levels of genetic variation are generally recognized as refugia (DeChaine and Martin 2005). Here, population X with high ISSR fingerprinting diversity was also among the highest genetic levels based on all molecular markers (Tables 2–4) in

Amentotaxus populations (Ge et al. 2005), reflecting features of a refugium.

Nevertheless, a large population size alone would not explain sustained maintenance of long diverged lineages. Bimodal mismatch distributions for mtDNA (Table 3) suggested that population X was likely a melting pot that maintained genetic diversity. In addition, populations H and E might represent other Quaternary refugia based on mismatch distributions, and higher genetic diversity levels in most genetic markers (Table 2). Phylogeographical analyses of mtDNA variant spatial distributions in *Amentotaxus* identified refugia in southwest (Sichuan Province) and central (Hunan Province) China (Fig. 1), congruent with fossil evidence (Axelrod et al. 1996). However, we did not detect refugia in eastern China (Del Tredici et al. 1992).

Amentotaxus yunnanensis exhibited very low mtDNA diversity levels, consistent with small populations (Table 4). However, all individuals from population S possessed the M26 haplotype—the haplotype delimiting lineage V—and clustered within *A. argotaenia* lineages III and IV (Figs. 2–3). If population S was composed of foreign migrants, the level of genetic diversity across the remaining populations would be reduced dramatically in mtDNA ($\pi=1.12 \times 10^{-3}$ and $\theta=0.72 \times 10^{-3}$)—a level even lower than in *A. formosana* (Table 2). Consequently, colonization via small founders must have occurred; leading not only to low genetic variation, but also to a unique genetic composition in each population. Interestingly, although population S was fixed in organellar DNA, microsatellites and ISSR fingerprinting displayed levels of variation that exceeded other populations (Ge et al. 2005). A demographic bottleneck might have occurred in this population, driving the cpDNA and mtDNA loci towards fixation, followed by subsequent expansion that recovered genetic polymorphisms at nuclear loci. Lower haplotype numbers and levels of genetic diversity were detected in *A. poilanei* compared with other species, indicating effects of recent demographic bottlenecks. Dominant haplotype fixation within species for most loci contributed to the depletion of genetic diversity in *A. poilanei*, and the absence of genetic differentiation between *A. yunnanensis* and *A. poilanei*. However, increased levels of genetic diversity were detected in microsatellites, showing a similar demographic history in *A. yunnanensis* (Table 5). Sample size is one of the critical factors affecting the accuracy of the estimation of population genetic diversity. Here all markers suggested similar patterns. Nucleotide diversity of *A. poilanei* ($\theta=1.86 \times 10^{-3}$) in cpDNA was close to that of *A. argotaenia* ($\theta=1.75 \times 10^{-3}$), both with larger sample size, whereas lower than that of *A. yunnanensis* ($\theta=3.21 \times 10^{-3}$), altogether indicating that the lower genetic diversity of *A. poilanei* likely resulted from genetic drift rather than from small sample size. In summary, long diverged *Amentotaxus* species exhibited significant genetic differentiation across the species' complex geographic range, but low genetic diversity

within species/populations, likely due to past fragmentation and recent habitat degradation. In contrast to ecologically pioneering gymnosperms, e.g., *Pinus*, *Amentotaxus* species are subordinate in well-developed forests, where forest canopies prevent seed and pollen exchange among populations. Most gene flow therefore is restricted within populations. In addition, human overexploitation has accelerated isolation among population fragments, leading to threats of extinction for these relic species.

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