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Effects of forest thinning on soil-plant carbon and nitrogen dynamics

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Received: 18 August 2016 / Accepted: 5 September 2016 / Published online: 10 September 2016 © Springer International Publishing Switzerland 2016

Abstract

Aims Corymbia spp. (previously included in the genus *Eucalyptus*) are common species in sub/tropical Australia and produce high quality timber and round logs. Thinning of native forests helps to preserve native tree species and is more sustainable than replacing native forest stands with mono-species plantations to produce timber. This study aimed to explore the effects of native

Responsible Editor: Simon Jeffery.

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forest thinning on soil-plant carbon (C) and nitrogen (N) dynamics in two experimental sites, Esk (5 years post-thinning) and Herberton (7 years post-thinning), situated in Queensland, Australia.

Methods The two sites had different thinning regimes. The final stocking rates varied between 75 and 200 stems ha^{-1} at Esk and between 250 and 400 stems ha^{-1} at Herberton. The thinned plots were compared to un-thinned plots. Soil samples were collected to measure labile C and N. Leaf samples were collected from *C. variegata* and *C. citriodora* in Esk and Herberton respectively.

Results Thinning did not change soil total C, total N, δ^{15} N and inorganic N at either Esk or Herberton. However, at Esk, intensive thinning resulted in decreases in water soluble total N (WSTN). Foliar δ^{13} C did not vary with respect to thinning whereas foliar δ^{15} N values were more enriched in thinned areas than those of un-thinned plots. The stepwise linear regression indicated that both foliar total N and δ^{15} N were explained mainly by soil TN and WSTN.

Conclusions Thinning did not change soil C and N most likely due to the retention of thinned materials on site and their incorporation into soil. Foliar δ^{13} C was not thinning-dependent due to homeostatic maintenance of the ratio of intercellular to ambient CO₂ concentrations during photosynthesis. In our study, soil N was not a limiting factor for foliar N, however, foliar N was mainly driven by WSTN which may foreshadow a possible N limitation in severely thinned plots in the long term. We conclude that forest thinning does not decrease soil C

and N availability in native *Corymbia* forests for several years post-thinning if the thinned materials are retained on site.

Keywords Stable isotopes · Inorganic nitrogen · *Corymbia* spp. · *Eucalyptus* spp. · Farm forestry · Water soluble total N and C

Introduction

The protection of forest biodiversity maintains ecosystem function and productivity (Erskine et al. 2006; Piotto 2008). Privately-owned native forest management, or 'farm forestry', is practiced to bring over-stocked and genetically impoverished forests back to productivity and is far more sustainable than replacing native forest stands with mono-species plantations to produce timber (Paul et al. 2013). Increasing spacing through thinning in plantations or forests has been shown to increase individual plant growth (Briggs et al. 2000; Henskens et al. 2001; Bai et al. 2013). Improved plant growth postthinning is a direct response to increased nutrient availability including nitrogen (N), phosphorus and potassium as well as decreased competition for available resources (Briggs et al. 2000; Thibodeau et al. 2000; Zhang et al. 2006). Improved tree growth has been reported for periods over three decades post-thinning (Zhang et al. 2006). Despite improved tree growth, it is uncertain how thinning affects carbon (C) and N cycling post-thinning and how this will be in turn translated in foliar C and N contents.

Nitrogen dynamics have been closely coupled with C cycling (Bai et al. 2015a, 2015b), and different studies suggest that enhanced N availability results in increased C sequestration in both plant biomass and soil (Högberg 2007; Xu et al. 2009). Soil and foliar C and N isotope compositions (δ^{13} C and δ^{15} N) have been used to investigate long-term C and N cycling in different soil-plant systems including plantations, forests, revegetated areas and agricultural lands (Xu et al. 2000; Bai et al. 2013; Nottingham et al. 2015; Reverchon et al. 2015). The study of δ^{13} C and δ^{15} N provides valuable long-term information on environmental conditions, water and nutrient availability, and management practices in different ecosystems (Ogaya and Peñuelas 2008; Cernusak et al. 2009; Mullaney et al. 2015; Xu et al. 2015). Foliar δ^{13} C reflects long-term plant water-use efficiency because water regulates stomatal closure which influences $^{13}CO_2$ discrimination of photosynthesis (Farquhar et al. 1989). Photosynthesis is additionally controlled by N and light availability (Meir et al. 2007; Osunkoya et al. 2010; Ma et al. 2015). Enriched soil 15 N signals may indicate N losses or acceleration in N transformations as a result of increased microbial activities (Högberg 1997; Kleinebecker et al. 2013). Foliar δ^{15} N values in turn reflect soil N dynamics as non-legume species assimilate soil available N (Kähmen et al. 2008).

Australian members of the Myrtaceae family, such as Eucalyptus and Corymbia, produce an exceptional timber and constitute a high proportion of plantation trees in sub/tropical areas worldwide due to their high adaptability to harsh conditions and to their fast growth (Santos et al. 2004). In sub-tropical Australia, forests types are usually tall open forests dominated by Eucalyptus spp. and Corymbia spp. and are managed to preserve these native species to sustain timber production (Lee 2007; Johnson et al. 2009). Tree growth response to spacing in plantations worldwide has been well documented (Henskens et al. 2001; Alcorn et al. 2007; Vande Walle et al. 2007). However studies on tree response to different thinning regimes in managed native forests are scant. In a complementary study, we investigated the effects of different thinning regimes on plant growth at two sites located in sub-tropical Australia, 1 and 3 years postthinning (Bai et al. 2013). Our results indicated that the thinning of native forests to stocking rates ranging from 50 to 200 stems ha^{-1} improved tree growth significantly compared to un-thinned plots (Bai et al. 2013).

Many forest thinning studies have investigated soil respiration as well as C and N dynamics post thinning and revealed a range of responses (Briggs et al. 2000; Kaye et al. 2005; Tang et al. 2005; Moore et al. 2006; Cheng et al. 2014, 2015). However, studies investigating C and N dynamics in both soils and plants post-thinning are relatively rare. Hence, the extent to which forest thinning alters C and N cycling in both soil and plant remains uncertain. Our study aimed to investigate mechanisms involved in C and N dynamics of both soils and plants at two *Corymbia* spp. dominated forests located in subtropical and tropical Australia, 5 and 7 years post-thinning.

Methods

Site description

The two experimental sites were situated at Esk ($27^{\circ} 14'$ S 152° 23' E) and Herberton ($17^{\circ} 27'$ S 45° 25' E), located in south-east and north-east Queensland, Australia. Esk is located approximately 200 km south of Gympie. The Esk site was categorised as open forest dominated by *Corymbia variegata* (F. Muell.) K.D. Hill & L.A.S. Johnson (spotted gum), *C. intermedia* (R.T. Baker) K.D. Hill & L.A.S. Johnson (red bloodwood) and other *Eucalyptus* spp. with an understorey of *Acacia* spp. and whipstick *Lophostemon confertus* (R. Br.) Peter G. Wilson & J.T. Waterh. The annual precipitation at Esk varied between 600 mm and 1400 mm from 2008 to 2014 (Fig. 1a). Throughout the sample collection period at Esk for the current study, the monthly rainfall varied between 7 mm and 190 mm (Fig. 1b).

The Herberton site is located in the Atherton tablelands approximately 100 km south-west of Cairns. The vegetation class was open forest with remnant vegetation dominated by *C. citriodora* K.D. Hill & L.A.S. Johnson (lemon-scented spotted gum) followed by *Eucalyptus argillacea* W. Fitzg. (Northern grey box) and other *Eucalyptus* spp. Herberton received annual rainfall within a range of 800 mm to 1800 mm in the years following the thinning (from 2007 to 2014) (Fig. 1a). At Herberton site during the sample collection in 2014, the monthly rainfall varied between 0 mm and 350 mm (Fig. 1b).

Experimental design and treatments

At Esk, a randomised complete block design with three replications was established in 2008. The plot areas were 70 m \times 70 m (0.49 ha). The sample collections were undertaken in the centre of the plots. The treatments included thinning to a final stocking rate of 75 stems ha⁻¹ (11.5 m \times 11.5 m of spacing); 120 stems ha⁻¹ (9 m \times 9 m of spacing); and 200 stems ha⁻¹ (7.5 m \times 7.5 m of spacing). The un-thinned plots contained a stocking rate of approximately 1150 stems ha⁻¹. The thinned materials were not removed from the thinned plots and were left on site to be decomposed.

At the Herberton site, the thinning commenced in 2007. We established a randomised complete block design with three replications for thinned areas. However, the un-thinned plots were established with six



Fig. 1 Mean annual rainfall and mean annual maximum daily temperature at the experimental sites between 2007 and 2014, years following the thinning (**a**), mean monthly rainfall at the experimental sites throughout the sample collection (**b**), data extracted from Bureau of Methodology on-line database. Black column; rainfall (R) in Herberton, white column; rainfall in Esk, close circles; temperature (T) in Herberton and open circles; temperature (T) in Esk

replications to undertake more intensive sample collection due to high variability in tree density in the unthinned areas. The plots were chosen in a circle with a surface area of 0.49 ha. Despite the fact that the plots were square at Esk and circular in Herberton, the plots had the same surface area and the sample collections were undertaken in the centre of the plots. The treatments included thinning to a final stocking rate of: 250 stems ha⁻¹; and 400 stems ha⁻¹. The un-thinned plots contained a stocking rate of up to 700 stems ha⁻¹. The thinned materials were not removed from the thinned plots and were left on site to be decomposed.

The thinned materials were not removed from thinned plots at either experimental sites because no viable commercial market existed in Australia for such products at the time. The thinned stems were not appropriate for fencing as they were not durable enough and/or they had poor form (e.g. bent or small). The thinned species were also not suitable for paper pulp. Nonetheless, the thinned materials could be used as bio-fuel, which is also a limited use due to the cost of transport and the distance between forests and the bio-fuel premises.

Sample collections

The sample collection strategies were similar at both sites. All the sample collections within one site were done on the same time. At the Esk site, both soil and leaf sample collections were undertaken in November 2013 and March 2014 (60 and 64 months, over 5 years post-thinning – before and after the wet season, respectively). At Herberton, the soil and leaf samples were collected in February and November 2014 (84 and 91 months, over 7 years post-thinning –after and before the wet season, respectively). Sampling dates were chosen based on the rainfall pattern in Esk and Herberton to be undertaken at the beginning and end of wet season.

Soil sample collections and analyses

Soil samples were collected at five points chosen randomly within the plots to a depth of 5 cm. The soil samples were collected using an auger with a 64-mm internal diameter. Soil samples collected from the same plots were bulked and mixed to provide one homogenised sample at each plot. Soil samples were then sieved (2 mm) when field moist in the laboratory and air dried before analysis.

A sub-sample of each soil sample was oven dried (50 °C) to a constant weight and ground to a fine powder (RocklabsTM ring grinder). Approximately 20 mg were transferred into 8 mm × 5 mm tin capsules for total C (TC), total N (TN), δ^{13} C and δ^{15} N analysis using an isotope ratio mass spectrometer (GV Isoprime, Manchester, UK).

To measure water soluble organic C (WSOC) and water soluble total N (WSTN), 5 g soil were added to 35 ml deionised water, then shaken by an end-over-end shaker for 5 min followed by centrifuging at 10,000 rpm for 10 min. The suspension was filtered through a Whatman 42 filter paper followed by filtering through a 33 mm Millex syringe-driven 0.45 μ m filter. The concentration of filtered solution was measured using a Shimadzu TOC-VCSH/CSN TOC/N analyser.

Soil inorganic N content (NH_4^+ -N and NO_3^- -N) was measured after shaking 5 g soil with 40 ml 2 *M* KCl by an end-over-end shaker for 60 min followed by a 10 min centrifugation at 4000 rpm and filtration through a Whatman 42 filter paper. The NH_4^+ -N and NO_3^- -N concentrations were then determined using a SmartChem 200, Discrete Chemistry Analyser (DCA).

Foliar sample collections and analyses

Leaf samples were collected from *C. variegata* and *C. citriodora* in Esk and Herberton respectively. The species chosen for this study were the dominant species in the corresponding experimental sites. The outside middle canopy branches were targeted using a Big Shot slingshot. Five fully expanded leaves were collected from fallen branches. The leaf samples were oven dried at 50 °C to a constant weight and ground to a fine powder by a RocklabsTM ring grinder. Approximately 4 mg of homogenised powder were transferred into 8 mm × 5 mm tin capsules to analyse foliar TC, TN, δ^{13} C and δ^{15} N using an isotope ratio mass spectrometer (GV Isoprime, Manchester, UK).

Statistical analysis

Thinning regimes differed at the two experimental sites, hence, we performed a repeated measures ANOVA at each site to detect significant difference among thinning treatments, sampling time and their interaction followed by Tukey test where significant differences were detected to compare treatments and sampling times of soil and foliar C and N contents. All data were tested for normality using Shapiro Wilk normality test and for homogeneity of variance using Levene's test. A stepwise regression using a linear model was then performed to determine which soil variables best explained foliar C and N contents. SPSS 21 software was used for all above statistical analyses. A linear regression was performed between soil WSTN and δ^{15} N as well as soil WSTN and foliar δ^{15} N pooling all data from both sites to find a general trend. Linear regression was performed in XLSTAT 15.1.

Results

Soil C and N pools

Esk

Total C and TN did not differ significantly among thinning treatments and there was no interaction between thinning and sampling time (Table 1). TC varied between 1.93 % and 3.18 % at this site and TN varied between 0.079 % and 0.160 %. Soil δ^{15} N did not vary either among treatments but it was significantly

influenced by the sampling time, being significantly higher at the beginning of the wet season than at the end in the 100 stems ha^{-1} (Table 1).

No changes were detected in WSOC with respect to the thinning treatment whereas WSTN was significantly influenced by thinning (Table 1). In month 60, WSTN was significantly lower in 75 stems ha⁻¹ compared to 100 stems ha⁻¹ whereas in month 64, WSTN in 75 stems ha⁻¹ was significantly lower than that of other treatments including the un-thinned areas (Table 2). Both WSOC and WSTN were significantly responsive to the sampling time (Table 1). WSOC only increased in un-thinned plots from month 60 to month 64 whereas WSTN increased significantly in 75, 200 stems ha⁻¹ and un-thinned plots from month 60 to month 64 postthinning (Table 2).

Neither NH_4^+ -N nor NO_3^- -N were influenced by thinning treatments, however, these variables changed significantly over the sampling time (Table 1). In general, NH_4^+ -N increased from month 60 to 64 but the increased NH_4^+ -N was significant only in 200 stems ha⁻¹ and un-thinned (Table 2). In contrast, soil NO_3^- -N decreased from month 60 to month 64 but the decrease was significant only in 75 stems ha⁻¹, 100 stems ha⁻¹ and un-thinned plots (Table 2).

Herberton

At Herberton, soil TC and TN did not differ at different thinning treatments at either sampling point (Tables 1 and 2). There was also no significant change for soil TC and TN between sampling times (Tables 1 and 2). Soil TC varied between 1.71 % and 5.04 % regardless of thinning treatment and sampling time (Table 2). Soil TN also varied between 0.146 % and 0.290 % regardless of thinning treatment and sampling time (Table 2). No significant differences in soil δ^{15} N were observed with respect to both thinning treatments and sampling time (Table 1).

Neither WSOC nor WSTN showed significant differences among thinning treatments regardless of sampling times (Tables 1 and 2). However, both WSOC and WSTN changed significantly from month 84 to 91 postthinning (Tables 1 and 2). WSOC significantly decreased in 400 stems ha⁻¹ and un-thinned plots from month 84 to month 91 post-thinning (Table 2). WSTN also decreased only in 250 stems ha⁻¹ and un-thinned plots from month 84 to month 91 post-thinning (Table 3). Neither NH₄⁺-N nor NO₃⁻-N showed any significant change regardless of both thinning treatment and sampling time (Tables 1 and 2).

Foliar C and N content

Esk

Foliar TC and TN showed no significant differences regardless of the thinning (Table 3). However, there were significant changes for foliar TC and TN between the sampling times. Foliar TC decreased from month 60 to month 64 post- thinning in all plots including the unthinned plots (Table 4). A decrease in foliar TN was also observed in thinned plots but not in un-thinned plots from month 60 to month 64 post-thinning (Table 4).

No significant differences of foliar δ^{13} C with respect to thinning and sampling time were detected (Tables 3 and 4). Foliar δ^{15} N was significantly higher in 100 stems ha⁻¹ compared to that of un-thinned plots and no differences in foliar δ^{15} N were observed among 75 stems ha⁻¹, 200 stems ha⁻¹ and un-thinned plots at

Table 1 Probability of a two-way ANOVA for soil properties in a farm forestry practice located at Esk and Herberton

Soil	TC (%)	TN (%)	δ ¹⁵ N (‰)	WSOC ($\mu g \ g^{-1}$)	WSTN ($\mu g g^{-1}$)	$N{H_4}^+\!N\;(\mu g\;g^{-1})$	$NO_3^{-}-N (\mu g g^{-1})$
Esk site							
Sampling time	0.784	0.516	0.037	0.001	P < 0.001	0.004	0.023
Thinning	0.527	0.444	0.357	0.166	0.028	0.470	0.871
Sampling time × Thinning	0.976	0.797	0.741	0.877	0.285	0.617	0.752
Herberton site							
Sampling time	0.078	0.736	0.419	P < 0.001	P < 0.001	0.052	0.389
Thinning	0.357	0.176	0.306	0.277	0.300	0.808	0.097
Sampling time \times Thinning	0.971	0.838	0.288	0.675	0.218	0.906	0.815

Table 2 Soil total C (TC), total N (TN), C:N ratio, N isotope composition (δ^{15} N), water soluble organic C (WSOC), water soluble total N (WSTN), NH₄⁺N and NO₃⁻-N at Esk and Herberton measured on two sampling times. Means followed with

different letters show a significant difference at P < 0.05 at each sampling time. Values in bold cases represent significant differences between sampling time (P < 0.05). Mean standard errors are presented in the brackets

	TC (%)		TN (%)		δ ¹⁵ N (‰)			
Soil – Esk site	Post-thinning							
Stocking rates	Month 60	Month 64	Month 60	Month 64	Month 60	Month 64		
75 (stems ha^{-1})	2.29 (0.5)	1.93 (0.5)	0.113 (0.03)	0.079 (0.01)	4.00 (0.7)	2.97 (0.3)		
$100 (\text{stems ha}^{-1})$	3.18 (0.9)	2.97 (0.8)	0.160 (0.03)	0.124 (0.02)	3.50 (0.7)	1.73 (0.7)		
$200 \text{ (stems ha}^{-1}\text{)}$	2.75 (0.4)	2.56 (0.3)	0.116 (0.01)	0.120 (0.01)	3.36 (0.4)	3.05 (0.9)		
Un-thinned	2.77 (0.7)	2.99 (0.8)	0.130 (0.03)	0.138 (0.04)	3.00 (0.5)	1.75 (0.9)		
	$WSOC(\mu gg^{-1})$		$WSTN(\mu gg^{-1})$		$N{H_4}^+\!N(\mu gg^{-1})$		$NO_3^{-}N(\mu g g^{-1})$	
	Month 60	Month 64	Month 60	Month 64	Month 60	Month 64	Month 60	Month 64
75 (stems ha^{-1})	181 (28)	299 (27)	9.09 (1.6)b	15.3 (1.1)b	6.99 (1.8)	11.4 (0.5)	11.5 (2.1)	8.43 (0.1)
$100 (\text{stems ha}^{-1})$	263 (35)	346 (61)	15.3 (1.3)a	20.6 (0.7)a	9.29 (2.5)	14.6 (2.2)	12.4 (2.2)	8.63 (0.3)
$200 \text{ (stems ha}^{-1}\text{)}$	196 (4.8)	298 (24)	10.8 (1.6)ab	21.2 (2.4)a	6.18 (0.4)	18.7 (5.0)	9.65 (0.4)	8.89 (0.3)
Un-thinned	243 (35)	390 (65)	11.3 (2.1)ab	22.6 (2.4)a	9.31 (2.1)	20.0 (6.0)	12.2 (3.2)	8.42 (0.1)
Soil – Herberton	site							
	TC (%)		TN (%)		δ ¹⁵ N (‰)			
	Month 84	Month 91	Month 84	Month 91	Month 84	Month 91		
$250 \text{ (stems ha}^{-1}\text{)}$	3.46 (1.3)	1.71 (0.8)	0.206 (0.06)	0.165 (0.6)	3.23 (0.2)	3.13 (0.7)		
$400 \text{ (stems ha}^{-1}\text{)}$	5.04 (1.5)	3.64 (0.8)	0.263 (0.07)	0.290 (0.6)	2.97 (0.3)	2.90 (0.7)		
Un-thinned	4.75 (1.1)	2.29 (0.6)	0.181 (0.05)	0.146 (0.04)	2.01 (0.2)	3.05 (0.4)		
	$WSOC(\mu gg^{-1})$		$WSTN~(\mu g~g^{-1})$		$N{H_4}^+\!N(\mu gg^{-1})$		$NO_3^{-}N (\mu g g^{-1})$	
	Month 84	Month 91	Month 84	Month 91	Month 84	Month 91	Month 84	Month 91
$250 \text{ (stems ha}^{-1}\text{)}$	443 (113)	208 (41)	28.5 (2.7)	9.84 (3)	9.80 (1.5)	5.98 (2)	9.48 (0.5)	9.78 (0.9)
$400 \text{ (stems ha}^{-1}\text{)}$	674 (130)	272 (41)	21.2 (3.1)	13.7 (3)	10.0 (1.7)	7.86 (2)	9.49 (0.6)	10.5 (0.9)
Un-thinned	523 (101)	189 (29)	20.0 (2.4)	9.50 (2)	10.1 (1.3)	7.37 (1)	8.41 (0.5)	8.61 (0.6)

month 64 post-thinning (Fig. 2a). Foliar δ^{15} N significantly increased in all plots from month 60 to month 64 post-thinning (Fig. 2a).

Herberton

Foliar TC was significantly higher in all treatments at month 84 than in month 91 (Table 4). Foliar TN significantly varied with respect to thinning and sampling time and there was a significant interaction between thinning and sampling time (Table 3). At month 84, both 250 stems ha⁻¹ and 400 stems ha⁻¹ had significantly higher TN than un-thinned plots (Table 4). At month 93, however, no significant difference in foliar TN was found among all treatments (Table 4). Foliar TN in 250 stems ha⁻¹ and 400 stems ha⁻¹ significantly decreased between month 84 and month 91 post-thinning (Table 4). No significant difference in foliar δ^{13} C among treatments was observed in either sampling time (Table 4). However, foliar δ^{13} C increased in all plots from month 84 to 91 post-thinning (Table 4). Foliar δ^{15} N was significantly higher in 250 stems ha⁻¹ and 400 stems ha⁻¹ compared to un-thinned plots at either sampling time (Fig. 2b). Foliar δ^{15} N values decreased significantly in all plots from month 84 to month 91 post-thinning (Fig. 2b).

Relationship between soil and foliar C and N content

Relationship between foliar TC, TN, δ^{13} C and δ^{15} N with soil C and N were examined using a stepwise regression. At Esk, soil WSTN and TN explained 71 % of variation in foliar TC (Table 5). WSTN was the sole significant factor explaining foliar TN and δ^{15} N (57 % and 36 % respectively; Table 5). At Herberton,

	TC (%)	TN (%)	δ ¹³ C (‰)	δ ¹⁵ N (‰)
Leaf – Esk site				
Sampling time	P < 0.0001	P < 0.0001	0.235	P < 0.0001
Thinning	0.816	0.975	0.365	0.016
Sampling time × Thinning	0.741	0.177	0.066	0.837
Leaf – Herberton site				
Sampling time	P < 0.0001	P < 0.0001	P < 0.0001	P < 0.0001
Thinning	0.409	P < 0.0001	0.907	P < 0.0001
Sampling time × Thinning	0.918	P < 0.0001	0.365	0.200

Table 3 Probability of a two-way ANOVA for foliar properties of Corymbia variegata at Esk and C. citriodora at Herberton

both WSTN and NO₃⁻-N explained 66 % of variation in foliar TC (Table 5). At Herberton, whilst both soil WSTN and TN explained 55 % of variation in foliar TN, WSTN was the only variable explaining 47 % of variation in foliar δ^{15} N (Table 5). In general, WSTN explained variations of both soil and foliar δ^{15} N (*P* < 0.05; 13 % and 42 %, respectively) (Fig. 3 a, b).

Discussion

In general, thinning did not affect soil TC, TN, NH₄⁺-N and NO₃⁻-N, at neither Esk (5 years post-thinning) nor Herberton (7 years post-thinning). Both increased and decreased N availability have been reported after thinning in different thinning regimes (Grady and Hart 2006; Baena et al. 2013; Overby and Hart 2016). For example, a decreased rate of N mineralisation was

Table 4 Foliar total C (TC), total N (TN) and C isotope composition (δ^{13} C) of *Corymbia variegata* at Esk; and *C. citriodora* at Herberton at different stocking measured on two sampling times. Means followed with different letters show a significant difference

observed up to 15 years following thinning in a ponderosa pine forest in Southwestern America due to decreased organic matter inputs (Grady and Hart 2006) whereas Kaye et al. (2005) reported an increased N availability after thinning in the same ponderosa pine forest. An increase in soil mineral C and in N mineralisation was also observed by Overby and Hart (2016) 6 month post-thinning, most likely due to incorporation of organic matter into the soil after thinning. However, both mineral C and mineral N returned to their initial levels one year following thinning (Overby and Hart 2016). Recently, increased nutrient availability due to thinning has been reported to be lessened in the longer term (Ganzlin et al. 2016). The mechanisms through which this attenuation of N dynamics could take place have been attributed to the decrease of mineralisation-immobilisation flushes that occur immediately after thinning, to N losses through leaching,

at P < 0.05 at each sampling time. Values in bold cases represent significant differences between sampling time (P < 0.05). Mean standard errors are presented in the brackets

		5	-			
Leaf – Esk site	TC (%)		TN (%)		δ ¹³ C (‰)	
	Post-thinning					
Stocking rates	Month 60	Month 64	Month 60	Month 64	Month 60	Month 64
75 (stems ha^{-1})	58.0(0.6)	48.1(0.6)	1.44(0.05)	1.18(0.05)	-28.6(0.25)	-28.9(0.25)
100 (stems ha ⁻¹)	58.0(0.6)	48.3(0.5)	1.46(0.05)	1.20(0.05)	-29.1(0.25)	-29.1(0.23)
200 (stems ha ⁻¹)	58.7(0.6)	48.6(0.6)	1.43(0.06)	1.20(0.05)	-29.2(0.26)	-29.1(0.25)
Un-thinned	58.6(0.6)	47.7(0.6)	1.34(0.05)	1.29(0.05)	-29.5(0.25)	-28.5(0.25)
Leaf - Herberton site	TC (%)		TN (%)		δ ¹³ C (‰)	
	Month 84	Month 91	Month 84	Month 91	Month 84	Month 91
250 (stems ha ⁻¹)	44.0(0.4)	32.1(0.4)	2.43(0.10)a	1.36(0.10)	-29.8(0.20)	-28.5(0.20)
400 (stems ha ⁻¹)	43.6(0.4)	32.1(0.4)	2.13(0.10)b	1.32(0.10)	-29.5(0.20)	-28.7(0.20)
Un-thinned	44.3(0.3)	32.5(0.3)	1.41(0.08)c	1.22(0.08)	-29.5(0.16)	-28.7(0.17)



Fig. 2 Foliar N isotope composition $(\delta^{15}N)$ of *Corymbia* variegata at Esk – close symbols (**a**); and *C. citriodora* at Herberton – open symbols (**b**) at different stocking measured on two sampling times. Stocking rates of 75 (stems ha⁻¹) – close rectangular, 100 (stems ha⁻¹) – close triangular, 200 (stems ha⁻¹) –

erosion, or denitrification, or to N adsorption by phenolic compounds (Hättenschwiler and Vitousek 2000; Dannenmann et al. 2006; Ganzlin et al. 2016).

At Herberton, the higher tree density in thinned areas than the one mentioned in Overby and Hart (2016) (116 stems ha⁻¹), as well as the lack of thinned material removal from the thinned plots, could explain the lack of thinning effects on soil C and N. At Esk, a severe thinning had been performed (75 stems ha⁻¹), however, lack of thinned material removal from the thinned plots may explain the absence of significant decreases in TC and TN in thinned areas compared to un-thinned areas. No effects of thinning on soil C and N were reported even three decades post-thinning which was partly associated

Table 5 Stepwise regression to identify driving factors explaining foliar C and N using soil variables, including soil TN, WSTN, NH_4^+N and NO_3^--N as independent parameters

Dependent	Independent	\mathbb{R}^2	Probability	
Esk site				
Foliar TC	WSTN TN	0.71	P < 0.0001	
Foliar TN	WSTN	0.57	0.004	
Foliar $\delta^{15}N$	WSTN	0.36	0.002	
Herberton site				
Foliar TC	WSTN	0.66	P < 0.0001	
Foliar $\delta^{13}C$	WSTN	0.41	0.001	
Foliar TN	WSTN TN	0.55	P < 0.0001	
Foliar $\delta^{15}N$	WSTN	0.47	P < 0.0001	



close diamond, un-thinned – close circle at Esk; and 250 (stems $ha^{-1})$ – open rectangular, 400 (stems $ha^{-1})$ – open triangular and un-thinned – open circle at Herberton. Astricts indicate significances at P < 0.05

to leaving thinned materials on site (Nilsen and Strand 2008). However, those authors found a decrease in above ground C storage by increased thinning intensities which may suggest thinning may require over 30 years to affect soil C and N storage (Nilsen and Strand 2008).

Thinning has been shown to change plant cover composition with increased herbaceous species (Moore et al. 2006). Nitrogen mineralisation is a microbially mediated process which can be influenced by alteration of vegetation composition, time since thinning and other environmental factors, which may explain the contradictory results reported in different studies undertaken in the same experimental site (Kaye et al. 2005; Grady and Hart 2006; Overby and Hart 2016). Thinning may change soil organic layer C and N, due to the decrease in organic matter input but without affecting the mineral layer of soil C and N (Inagaki et al. 2011) which may additionally explain those contrasting findings.

Intensive thinning may limit N availability in long term which was supported by the lowest WSTN observed in the intensively thinned plots at the Esk. The soil WSTN mainly originates from organic matter inputs and is an important indicator of soil N availability (Huang and Schoenau 1998). However, soluble forms of soil N are also highly susceptible to leaching and lighter N (¹⁴N) is more prone to be leached compared to heavier N (¹⁵N), which leads to soil δ^{15} N enrichment (Högberg 1997). The negative relationship between soil WSTN and soil δ^{15} N observed in our data (Fig. 3a) further indicates a possible acceleration of leaching in intensively thinned plots.

The sample collection in this study was undertaken before and after the wet season. WSOC and WSTN were higher after the wet season compared to before



Fig. 3 Relationship between soil water soluble total nitrogen (WSTN) and soil N isotope composition (δ^{15} N) (**a**) and between WSTN and foliar δ^{15} N (**b**)

wet season mainly in less intensively thinned areas and un-thinned plots. Soil organic C and N as well as microbial biomass C and N were investigated under thinning in both summer and autumn in two Mediterranean forests (Baena et al. 2013). There was a significant relationship between soil labile C and N and soil moisture, as soil labile C was higher in autumn than summer when temperature was also lower (Baena et al. 2013). Soil moisture is one of the main driving factors for microbial C and N transformations (Merilä et al. 2002). Increases and decreases of WSOC and WSTN at both sites in the current study concurred with the precipitation patterns. Despite the fact that WSOC and WSTN were responsive to precipitation patterns, soil TC and TN remained unaffected. We think that less intensively thinned and un-thinned plots may have resulted in increased organic inputs to temporally increase WSOC and WSTN. However, the inputs were not sufficiently high to change soil TC and TN.

In this study, foliar δ^{13} C was not tree density-dependent. Foliar δ^{13} C can change with water, light and resource availability. For example, foliar δ^{13} C is expected to increase due to increased water limitation (Qiu et al. 2013; Mullaney et al. 2015) as a consequence of increased stomatal closure leading to greater ¹³C fixation in the carboxylation site (Farquhar et al. 1989). Lack of foliar δ^{13} C response to tree density has also been observed in other studies, suggesting that increased light intensity, water and N availability post-thinning may not be translated into an overall change in foliar δ^{13} C (Geßler et al. 2001; Fotelli et al. 2003; Skov et al. 2004; Inagaki et al. 2011). Our results are contradictory

with a study undertaken in overstory and understory trees of *E. marginata* post-thinning where increases in water availability and decreases in N availability led to decreased foliar δ^{13} C of *E. marginata* juveniles (Qiu et al. 2013). Those authors suggested that N limitation was affecting foliar δ^{13} C in *E. marginata* juveniles post-thinning, but also attributed this decrease in N availability to an extremely dry climate combined with their thinning experiment (Qiu et al. 2013).

The lack of foliar δ^{13} C response to tree density observed in our study may be explained through different mechanisms. For example, it has been shown that increased light interception and N availability may have a counteracting effect to increased water availability, because they generally tend to increase δ^{13} C by increasing the drawdown of CO_2 from the atmosphere to the sites of carboxylation (Cernusak et al. 2013). This acts in opposition to increased water availability, which tends to decrease the drawdown of CO₂ from the atmosphere to the carboxylation sites by allowing stomata to be more open. Additionally, homeostatic regulation of the ratio of intercellular CO2 to ambient CO2 concentrations during photosynthesis may also associate with a lack of foliar δ^{13} C response to tree density because homeostatic maintenance of gas exchange in plants adjusts carbon fixation at the carboxylation site for a given stomatal conductance (Wong et al. 1979; Skov et al. 2004; McDowell et al. 2006). Homeostatic maintenance is evident in anisohydric plants giving them an advantage over isohydric plants to cope with water limitation (McDowell and Allen 2015). Some Eucalyptus spp. have been shown to be anisohydric, maintaining higher

stomatal conductance under a given foliar water potential compared to isohydric species, which makes these species more tolerant to water limitation (Franks et al. 2007; McDowell et al. 2008). Hence, we think that although plants at Esk were experiencing a very dry year with 609 mm rainfall in 2014, the foliar δ^{13} C did not vary due to the fact that there might have been maintenance of a relatively constant ratio of intercellular to ambient CO₂ concentrations for photosynthesis, even while trees were experiencing water scarcity.

Foliar δ^{15} N tended to be more enriched in thinned plots compared to un-thinned plots. Foliar $\delta^{15}N$ indicates the condition of N at the moment of plant growth and is therefore linked to soil $\delta^{15}N$ (Ibell et al. 2013). Various mechanisms have been shown to influence leaf ¹⁵N signatures. Firstly, when canopy density reduces after thinning, a higher N uptake would lead to higher ¹⁵N fixation in leaves (Qiu et al. 2013). Secondly, an increase in foliar $\delta^{15}N$ indicates an acceleration of N transformation due to an increase in soil microbial activity and/or increased leaching (Garten 1993; Ibell et al. 2010; Reverchon et al. 2014). Soil microbial N transformation changes the $^{15}\mathrm{N}$ signal of both $\mathrm{NH_4^+N}$ and NO₃⁻-N (Natelhoffer and Fry 1988). An enhanced leaching and N turnover in thinned areas are likely to occur in the long term due to the release of added biomass residues to the soil from thinned materials (Qiu et al. 2012). A positive relationship between soil N availability and enriched foliar δ^{15} N has also been observed after N fertilisation and understorey removal which was associated with enhanced mineralization and nitrification as well as increased leaching (Matsushima et al. 2012). This is consistent with the results from our regression analysis where WSTN, which is an indicator of fast N availability, explained 42 % of variation in foliar δ^{15} N (Fig. 3b). Thirdly, many tree species including Eucalyptus spp. have a strong mutualistic relationship with mycorrhizal fungi which additionally affect their N uptake (Horton et al. 2013). Isotopic fractionation occurs when N is transferred from mycorrhizal fungi to plants and mycorrhizal ¹⁵N signals are more enriched than plants (Hobbie et al. 2000; Spriggs et al. 2003). It is possible that trees in un-thinned plots are more reliant upon mycorrhizal colonisation to overcome the competition leading to less enriched foliar ¹⁵N signals in un-thinned plots compared to thinned plots. In addition, thinning has been shown to alter myorrhizal fungal communities (Buée et al. 2005; Teste et al. 2012). We did not measure ¹⁵N signal of soil inorganic N nor mycorrhizal ¹⁵N in this study, hence, the magnitude of their influence on foliar ¹⁵N signals remains uncertain.

In our previous study, we found that there were no significant differences in tree growths in thinned plots at the Esk site (Bai et al. 2013). Therefore, decreased WSTN in intensively thinned plots (75 stems ha^{-1}) at Esk did not compromise plant growth probably due to lower competition for available N in severely thinned plots compared to other thinning regimes. However, it may imply a decrease in organic matter input in this site. Nitrogen utilisation efficiency has been reported to remain unaffected by thinning in a hinoki cypress plantation in Japan (Inagaki et al. 2011) despite a decrease in organic soil N content. However, those authors suggested that in the long-term, plant N uptake might be limited (Inagaki et al. 2011). While severe thinning may be the only practical and sustainable solution to improve the genetics of a native forest that has been progressively harvested of the best timber for generations, it also has implications for the soil organic matter. Thinning promotes understory regeneration due to improved light and resource availability which compensates for part of the decreased organic matter inputs after thinning (Grady and Hart 2006; Moore et al. 2006). The current study also suggests leaving thinned organic matter in thinned plots further ensures preserving soil total C and N if high intensity thinning is required to be applied.

Conclusion

Soil and plant interactions following thinning have been studied and shown to improve tree growth due to increased nutrient and light availability as well as decreased competition (Giuggiola et al. 2015; Chase et al. 2016; Zhang et al. 2016). Farm forestry has also been revealed to be economically viable in areas receiving annual rainfall over 600 mm and if not replacing agricultural lands (Paul et al. 2013). However, the present study showed that soil TC and TN were not influenced by thinning most likely due to the fact that thinned materials had not been removed from the thinned plots and/or due to stimulation of understorey growth. Lack of significant differences in soil C and N with respect to thinning was also consistent with a lack of response of foliar TC and TN. Possible soil N limitation in the long term in intensively thinned plots was supported by decreased WSTN and increased foliar δ^{15} N which was an indication of increased leaching and N turnover. Lack of foliar δ^{13} C response to tree density may be attributed to offsetting effects of increased light interception and N availability acting in opposition to effects of increased water availability, as well as homeostatic regulation of the ratio of intercellular CO₂ to ambient CO₂ concentrations during photosynthesis. Considering that tree growth was not influenced in intensively thinned plots compared to less intensively thinned plots (Bai et al. 2013), less intensive thinning may ensure longer term sustainability in the system compared to intensive thinning. If intensive thinning is required to be undertaken, incorporating thinning residues in soil helps to retain soil C and N.

Acknowledgment This study was funded by James Cook University and Griffith University Collaborative Scheme. The authors would like to acknowledge the owner of the Herberton site (Mr. Olivier Lambert) and the owner of the Esk site for providing the access to the experimental sites. The authors would like to acknowledge Private Forestry Service Queensland (PFSQ) for providing the long term growth data and access to the Esk site. We thank Mr. Geoffrey Lambert for his assistance in both laboratory and sample collection, Mr. Yan Zhao for his assistance in sample collection and Mr. Rene Diocares for stable isotope analyses. SHB was supported by Collaborative Research Network – University of the Sunshine Coast Research Futures project.

References

- Alcorn PJ, Pyttel P, Bauhus J, Smith RGB, Thomas D, James R, Nicotra A (2007) Effects of initial planting density on branch development in 4-year-old plantation grown *Eucalyptus pilularis* and *Eucalyptus cloeziana* trees. For Ecol Manag 252:41–51
- Baena CW, Andrés-Abellán M, Lucas-Borja ME, Martínez-García E, García-Morote FA, Rubio E, López-Serrano FR (2013) Thinning and recovery effects on soil properties in two sites of a Mediterranean forest, in Cuenca Mountain (south-eastern of Spain. For Ecol Manag 308:223–230
- Bai, S. H., Blumfield, T. J., Reverchon, F., Ryan, R., 2013. Profitable practice management in native forests to sustain timber production: an opportunity for the private sector. *In* International Union of Forest Research Organizations (IUFRO) 2013 Fukuoka, Japan
- Bai SH, Blumfield TJ, Reverchon F, Amini S (2015a) Do young trees contribute to soil labile carbon and nitrogen recovery? J Soils Sediments 15:503–509
- Bai SH, Xu Z, Blumfield TJ, Reverchon F (2015b) Human footprints in urban forests: implication of nitrogen deposition for nitrogen and carbon storage. J Soils Sediments 15:1927– 1936
- Briggs RD, Hornbeck JW, Smith CT, Lemin RC, McCormack ML (2000) Long-term effects of forest management on nutrient cycling in spruce-fir forests. For Ecol Manag 138:285–299

- Buée M, Vairelles D, Garbaye J (2005) Year-round monitoring of diversity and potential metabolic activity of the ectomycorrhizal community in a beech (*Fagus silvatica*) forest subjected to two thinning regimes. Mycorrhiza 15: 235–245
- Cernusak LA, Winter K, Turner BL (2009) Physiological and isotopic (δ^{13} C and δ^{18} O) responses of three tropical tree species to water and nutrient availability. Plant Cell Environ 32:1441–1455
- Cernusak LA, Ubierna N, Winter K, Holtum JAM, Marshall JD, Farquhar GD (2013) Environmental and physiological determinants of carbon isotope discrimination in terrestrial plants. New Phytol 200:950–965
- Chase CW, Kimsey MJ, Shaw TM, Coleman MD (2016) The response of light, water, and nutrient availability to precommercial thinning in dry inland Douglas-fir forests. For Ecol Manag 363:98–109
- Cheng X, Han H, Kang F, Liu K, Song Y, Zhou B, Li Y (2014) Short-term effects of thinning on soil respiration in a pine (*Pinus tabulaeformis*) plantation. Biol Fertil Soils 50:357– 367
- Cheng X, Kang F, Han H, Liu H, Zhang Y (2015) Effect of thinning on partitioned soil respiration in a young *Pinus* tabulaeformis plantation during growing season. Agric For Meteorol 214–215:473–482
- Dannenmann M, Gasche R, Ledebuhr A, Papen H (2006) Effects of forest management on soil N cycling in beech forests stocking on calcareous soils. Plant Soil 287:279–300
- Erskine PD, Lamb D, Bristow M (2006) Tree species diversity and ecosystem function: can tropical multi-species plantations generate greater productivity? For Ecol Manag 233:205–210
- Farquhar GD, Ehleringer JR, Hubick KT (1989) Carbon isotope discrimination and photosynthesis. Annu Rev Plant Biol 40: 503–537
- Fotelli MN, Rennenberg H, Holst T, Mayer H, Geßler A (2003) Carbon isotope composition of various tissues of beech (*Fagus sylvatica*) regeneration is indicative of recent environmental conditions within the forest understorey. New Phytol 159:229–244
- Franks PJ, Drake PL, Froend RH (2007) Anisohydric but isohydrodynamic: seasonally constant plant water potential gradient explained by a stomatal control mechanism incorporating variable plant hydraulic conductance. Plant Cell Environ 30:19–30
- Ganzlin PW, Gundale MJ, Becknell RE, Cleveland CC (2016) Forest restoration treatments have subtle long-term effects on soil C and N cycling in mixed conifer forests. Ecol Appl 26: 1503–1516
- Garten CT (1993) Variation in foliar ¹⁵N abundance and the availability of soil nitrogen on Walker branch watershed. Ecology 74:2098–2113
- Geßler A, Schrempp S, Matzarakis A, Mayer H, Rennenberg H, Adams MA (2001) Carbon isotope composition of phloem sap, wood and foliage of beech (*Fagus sylvatica* L.): effects of water availability and radiation during the growing season. New Phytol 150:653–664
- Giuggiola A, Ogée J, Rigling A, Gessler A, Bugmann H, Treydte K (2015) Improvement of water and light availability after thinning at a xeric site: which matters more? A dual isotope approach, New Phytologist

- Grady KC, Hart SC (2006) Influences of thinning, prescribed burning, and wildfire on soil processes and properties in southwestern ponderosa pine forests: a retrospective study. For Ecol Manag 234:123–135
- Hättenschwiler S, Vitousek PM (2000) The role of polyphenols in terrestrial ecosystem nutrient cycling. Trends Ecol Evol 15: 238–243
- Henskens FL, Battaglia M, Cherry ML, Beadle CL (2001) Physiological basis of spacing effects on tree growth and form in *Eucalyptus globulus*. Trees 15:365–377
- Hobbie EA, Macko SA, Williams M (2000) Correlations between foliar δ^{15} N and nitrogen concentrations may indicate plant-mycorrhizal interactions. Oecologia 122:273–283
- Högberg P (1997) Tansley review no. 95¹⁵N natural abundance in soil-plant systems. New Phytol 137:179–203
- Högberg P (2007) Environmental science: nitrogen impacts on forest carbon. Nature 447:781–782
- Horton BM, Glen M, Davidson NJ, Ratkowsky D, Close DC, Wardlaw TJ, Mohammed C (2013) Temperate eucalypt forest decline is linked to altered ectomycorrhizal communities mediated by soil chemistry. For Ecol Manag 302:329–337
- Huang WZ, Schoenau JJ (1998) Fluxes of water-soluble nitrogen and phosphorus in the forest floor and surface mineral soil of a boreal aspen stand. Geoderma 81:251–264
- Ibell PT, Xu Z, Blumfield TJ (2010) Effects of weed control and fertilization on soil carbon and nutrient pools in an exotic pine plantation of subtropical Australia. J Soils Sediments 10: 1027–1038
- Ibell PT, Xu Z, Blumfield TJ (2013) The influence of weed control on foliar δ^{15} N, δ^{13} C and tree growth in an 8 year-old exotic pine plantation of subtropical Australia. Plant Soil 369:199– 217
- Inagaki Y, Nakanishi A, Fukata H (2011) Soil properties and nitrogen utilization of hinoki cypress as affected by strong thinning under different climatic conditions in the Shikoku and Kinki districts in Japan. J For Res 16:405–413
- Johnson I, Carnegie A, Henson M (2009) Growth, form and Quambalaria shoot blight tolerance of spotted gum in northeastern new South Wales, Australia. Silvae Genetica 58:180
- Kähmen A, Wanek W, Buchmann N (2008) Foliar δ¹⁵N values characterize soil N cycling and reflect nitrate or ammonium preference of plants along a temperate grassland gradient. Oecologia 156:861–870
- Kaye JP, Hart SC, Fulé PZ, Covington WW, Moore MM, Kaye MW (2005) Initial carbon, nitrogen, and phosphorus fluxes following ponderosa pine restoration treatments. Ecol Appl 15:1581–1593
- Kleinebecker T, Poelen MD, Smolders AJ, Lamers LP, Hölzel N (2013) Fast and inexpensive detection of total and extractable element concentrations in aquatic sediments using nearinfrared reflectance spectroscopy (NIRS. PLoS One 8: e70517
- Lee DJ (2007) Achievements in forest tree genetic improvement in Australia and New Zealand 2: development of Corymbia species and hybrids for plantations in eastern Australia. Aust For 70:11–16
- Ma L, Rao X, Lu P, Bai SH, Xu Z, Chen X, Blumfield TJ, Xie J (2015) Ecophysiological and foliar nitrogen concentration responses of understorey Acacia spp. and Eucalyptus sp. to prescribed burning. Environ Sci Pollut Res 22:10254–10262

- Matsushima M, Choi WJ, Chang SX (2012) White spruce foliar δ^{13} C and δ^{15} N indicate changed soil N availability by understory removal and N fertilization in a 13-year-old boreal plantation. Plant Soil 361:375–384
- McDowell NG, Allen CD (2015) Darcy's law predicts widespread forest mortality under climate warming. Nat Clim Chang 5: 669–672
- McDowell NG, Adams HD, Bailey JD, Hess M, Kolb TE (2006) Homeostatic maintenance of ponderosa pine gas exchange in response to stand density changes. Ecol Appl 16:1164–1182
- McDowell N, Pockman WT, Allen CD, Breshears DD, Cobb N, Kolb T, Plaut J, Sperry J, West A, Williams DG, Yepez EA (2008) Mechanisms of plant survival and mortality during drought: why do some plants survive while others succumb to drought? New Phytol 178:719–739
- Meir P, Levy PE, Grace J, Jarvis PG (2007) Photosynthetic parameters from two contrasting woody vegetation types in West Africa. Plant Ecol 192:277–287
- Merilä P, Smolander A, Strömmer R (2002) Soil nitrogen transformations along a primary succession transect on the landuplift coast in western Finland. Soil Biol Biochem 34:373– 385
- Moore MM, Casey CA, Bakker JD, Springer JD, Fule PZ, Covington WW, Laughlin DC (2006) Herbaceous vegetation responses (1992–2004) to restoration treatments in a ponderosa pine forest. Rangel Ecol Manag 59:135–144
- Mullaney J, Trueman SJ, Lucke T, Bai SH (2015) The effect of permeable pavements with an underlying base layer on the ecophysiological status of urban trees. Urban For Urban Green 14:686–693
- Natelhoffer KJ, Fry B (1988) Controls on natural nitrogen-15 and carbon-13 abundances in forest soil organic matter. Soil Sci Soc Am J 52:1633–1640
- Nilsen P, Strand LT (2008) Thinning intensity effects on carbon and nitrogen stores and fluxes in a Norway spruce (*Picea abies* (L.) karst.) stand after 33 years. For Ecol Manag 256: 201–208
- Nottingham AT, Turner BL, Stott AW, Tanner EV (2015) Nitrogen and phosphorus constrain labile and stable carbon turnover in lowland tropical forest soils. Soil Biol Biochem 80:26–33
- Ogaya R, Peñuelas J (2008) Changes in leaf δ^{13} C and δ^{15} N for three Mediterranean tree species in relation to soil water availability. Acta Oecol 34:331–338
- Osunkoya OO, Bayliss D, Panetta FD, Vivian-Smith G (2010) Variation in ecophysiology and carbon economy of invasive and native woody vines of riparian zones in South-Eastern Queensland. Austral Ecology 35:636–649
- Overby ST, Hart SC (2016) Short-term belowground responses to thinning and burning treatments in Southwestern ponderosa pine forests of the USA. Forests 7:45
- Paul KI, Reeson A, Polglase P, Crossman N, Freudenberger D, Hawkins C (2013) Economic and employment implications of a carbon market for integrated farm forestry and biodiverse environmental plantings. Land Use Policy 30:496–506
- Piotto D (2008) A meta-analysis comparing tree growth in monocultures and mixed plantations. For Ecol Manag 255:781–786
- Qiu S, Bell RW, Hobbs RJ, McComb AJ (2012) Estimating nutrient budgets for prescribed thinning in a regrowth eucalyptus forest in south-West Australia. Forestry 85:51–61
- Qiu S, Bell RW, Hobbs RJ, McComb AJ (2013) Overstorey and juvenile response to thinning and drought in a jarrah

(Eucalyptus marginata Donn ex Sm.) forest of southwestern Australia. Plant Soil 365:291–305

- Reverchon F, Flicker RC, Yang H, Yan G, Xu Z, Chen C, Bai SH, Zhang D (2014) Changes in δ^{15} N in a soil–plant system under different biochar feedstocks and application rates. Biol Fertil Soils 50:275–283
- Reverchon F, Bai SH, Liu X, Blumfield TJ (2015) Tree plantation systems influence nitrogen retention and the abundance of nitrogen functional genes in the Solomon Islands. Front Microbiol 6:1439. doi:10.3389/fmicb.2015.01439
- Santos PETD, Geraldi IO, Garcia JN (2004) Estimates of genetic parameters of wood traits for sawn timber production in *Eucalyptus grandis*. Genet Mol Biol 27:567–573
- Skov KR, Kolb TE, Wallin KF (2004) Tree size and drought affect ponderosa pine physiological response to thinning and burning treatments. For Sci 50:81–91
- Spriggs AC, Stock WD, Dakora FD (2003) Influence of mycorrhizal associations on foliar $\delta^{15}N$ values of legume and nonlegume shrubs and trees in the fynbos of South Africa: implications for estimating N₂ fixation using the ¹⁵N natural abundance method. Plant Soil 255:495–502
- Tang J, Qi Y, Xu M, Misson L, Goldstein AH (2005) Forest thinning and soil respiration in a ponderosa pine plantation in the sierra Nevada. Tree Physiol 25:57–66
- Teste FP, Lieffers VJ, Strelkov SE (2012) Ectomycorrhizal community responses to intensive forest management: thinning alters impacts of fertilization. Plant Soil 360:333–347
- Thibodeau L, Raymond P, Camiré C, Munson AD (2000) Impact of precommercial thinning in balsam fir stands on soil

nitrogen dynamics, microbial biomass, decomposition, and foliar nutrition. Can J For Res 30:229-238

- Walle IV, Van Camp N, Van de Casteele L, Verheyen K, Lemeur R (2007) Short-rotation forestry of birch, maple, poplar and willow in Flanders (Belgium) I—biomass production after 4 years of tree growth. Biomass Bioenergy 31:267–275
- Wong S-C, Cowan IR, Farquhar GD (1979) Stomatal conductance correlates with photosynthetic capacity. Nature 282:424–426
- Xu ZH, Saffigna PG, Farquhar GD, Simpson JA, Haines RJ, Walker S, Osborne DO, Guinto D (2000) Carbon isotope discrimination and oxygen isotope composition in clones of the F-1 hybrid between slash pine and Caribbean pine in relation to tree growth, water-use efficiency and foliar nutrient concentration. Tree Physiol 20:1209–1217
- Xu Z, Chen C, He J, Liu J (2009) Trends and challenges in soil research 2009: linking global climate change to local longterm forest productivity. J Soils Sediments 9:83–88
- Xu CY, Bai SH, Hao Y, Rachaputi RC, Xu Z, Wallace HM (2015) Peanut shell biochar improves soil properties and peanut kernel quality on a red Ferrosol. J Soils Sediments 15: 2220–2231
- Zhang SY, Chauret G, Swift DE, Duchesne I (2006) Effects of precommercial thinning on tree growth and lumber quality in a jack pine stand in New Brunswick, Canada. Can J For Res 36:945–952
- Zhang J, Webster J, Young DH, Fiddler GO (2016) Effect of thinning and soil treatments on *Pinus ponderosa* plantations: 15-year results. For Ecol Manag 368:123–132