REGULAR ARTICLE

Carbon allocation in *Larrea tridentata* plant-soil systems as affected by elevated soil moisture and N availability

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Abstract

Background and Aims Global change will likely express itself in southwestern United States arid lands through changes in amounts and timing of precipitation in response to elevated CO_2 concentrations. In addition, increased nitrogen (N) deposition may occur due to increased urban development. This study addressed the effects of water and N availability on C allocation in arid land soil-plant systems.

Methods Columns filled with Mojave Desert topsoil containing *Larrea tridentata* seedlings with two treatment levels each of N and soil moisture were labeled by exposure to 13 C-enriched CO₂.

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Department of Natural Resources and Environmental Science, University of Nevada, Reno, 1664 North Virginia Street, Reno, NV 89557, USA *Results* Increased soil moisture increased plant biomass, total ¹³C uptake, ¹³C levels in leaves, soil organic matter, and soil respiration, decreased relative C allocation to stems but increased allocation to soil organic matter. Increased soil N availability increased N uptake but decreased C allocation to soil respiration presumably due to decreased substrate supply for microbes. There was no detectable label in carbonate C, suggesting that this pool does not significantly contribute to ecosystem C fluxes.

Conclusions Our study indicates that increased water availability causes increased C uptake with increased C allocation to soil organic matter in *Larrea tridentata*-dominated communities while increased N deposition will have a minimal impact on C sequestration.

Keywords Larrea tridentata \cdot Mojave Desert \cdot Carbon allocation \cdot Soil moisture \cdot Nitrogen deposition \cdot Climate change

Introduction

Rising atmospheric CO₂ concentrations are predicted to cause an increase in atmospheric temperature by 4–5 °C in the next century in the arid lands of the southwestern U.S. while mean precipitation (PPT) is expected to decrease, with an increase in severity and frequency of droughts (Seager et al. 2007; Cayan et al. 2010; Fawcett et al. 2011; Notaro et al. 2012). In northern U.S. drylands, PPT is expected to change from a winter to a fall/spring-dominated regime (Solomon et al. 2007), thereby shifting the timing of available moisture. In addition, high-intensity storms likely will become more common (Archer and Predick 2008). Climate change predictions are however highly uncertain for U.S. drylands both in terms of magnitude and direction of PPT as the processes responsible for climate forcing are poorly understood in this region (Friggens et al. 2012). Since biological processes in drylands typically are constrained by moisture availability, any changes in PPT amount and distribution likely will have important implications for C cycling in these ecosystems (e.g., Weltzin et al. 2003; Austin et al. 2004; Huxman et al. 2004). In addition to changes in PPT, ecosystem processes in the western U.S. may be impacted by rapid development of urban areas causing large increases in nitrogen (N) deposition in this region (e.g., Noy-Meir 1973; Smith et al. 1997), with deposition estimates ranging from 29 to 45 kg ha⁻¹ yr⁻¹ downwind from major cities (Fenn et al. 2003).

Despite arid regions covering a substantial portion of the earth's surface (± 40 %; Reynolds 2001), relatively little is known about the role of these regions in the global C cycle and how their role might change in response to environmental change. Recently, several studies have suggested that arid ecosystems may represent an important sink for C (Jasoni et al. 2005; Wohlfahrt et al. 2008; Xie et al. 2009; Stone 2008). Potential C sinks that could contribute to C uptake include net primary productivity (e.g., Miller and Huenneke 1996; Rundel and Gibson 1996), biological crusts (Su et al. 2012; Li et al. 2012), and inorganic C (soil carbonates; Emmerich 2003; Xie et al. 2009); but none of these uptake mechanisms either by themselves or collectively appears to be large enough to explain uptake quantities obtained by ecosystem flux measurements (Schlesinger et al. 2009).

Generally, increases in resource availability appear to favor allocation of C from roots to leaves (e.g., Hunt et al. 1996; Poorter and Nagel 2000). Consequently, as resource limitations change, belowground C allocation may change as well which could affect soil C cycling through potential changes in root litter production and priming of soil organic matter decomposition (e.g., Dijkstra and Cheng 2007). Several studies show that plant photosynthesis as well as water and nutrient uptake in arid ecosystems increase in response to increased water and/or N availability (e.g., BassiriRad et al. 1999; Hamerlynck et al. 2000; Gebauer and Ehleringer 2002; Schwinning et al. 2002, 2005a, b; Snyder et al. 2004; Barker et al. 2006), but it is unclear how these changes affect total C uptake and/or C allocation.

Barker et al. (2006) showed increased photosynthetic activity following increases in summer rainfall and N deposition in a Mojave Desert ecosystem. This increased C uptake, however, was not accompanied by significant changes in aboveground biomass (Newingham et al. 2012) or fine root production (Verburg et al. 2013). Similarly, Clark et al. (2010) observed that fine root production and root respiration did not increase under elevated CO₂ concentrations despite measuring increased photosynthetic activity (Smith et al. 2000; Housman et al. 2006). They speculated that the extra C taken up may be released as root exudates since C-limited microbial processes were stimulated under elevated CO₂ (Billings et al. 2004; Jin and Evans 2007). These results suggest that the increased C uptake may not necessarily result in increased C sequestration in biomass as C may be respired at higher rates as well (e.g., Huxman et al. 2004; Cable et al. 2011). None of these studies assessed potential changes in C allocation to soil organic C pools in response to changes in resource availability.

Although responses of arid lands to changes in water and N availability appear modest, timing of resource availability can be critical. For instance, several studies show that plant production in the arid lands increased with increased spring moisture availability as most above- and belowground biomass production occurs during this period (e.g., Reynolds et al. 1999; Schwinning et al. 2005b; Phillips et al. 2006; Verburg et al. 2013). Consequently, for these ecosystems, changes in timing of resource availability in response to climate change may have a significant impact on C sequestration.

One mechanism that could potentially be important in arid lands that has received recent attention is sequestration of C in secondary carbonates (e.g., Stone 2008). In many arid soils, carbonate C is the largest C reservoir (Eswaran et al. 2000; Lal and Kimble 2000). Several field and laboratory studies have indicated a potential contribution of carbonate C to ecosystem C fluxes (e.g., Xie et al. 2009; Kuzyakov et al. 2006; Emmerich 2003; Stevenson and Verburg 2006). In contrast, Serna-Perez et al. (2006) found little evidence of C originating from carbonates in soil respiration measured in calcareous soils in New Mexico. Lapenis et al. (2008) proposed that carbonate dissolution and subsequent leaching could account for CO₂ uptake of 2.1–7.4 g C m⁻² yr⁻¹, while Marion et al. (2008) suggested that sequestration rates of carbonate-C would be as low as $0.2 \text{ g m}^{-2} \text{ yr}^{-1}$

indicating a minor contribution to net ecosystem C exchange.

The main objective of our study was to quantify the effects of changes in water and N availability on C allocation in native desert shrubs under controlled chamber conditions using isotopic tracers. We exposed plants to ¹³C-enriched CO₂ which allowed us to trace recently assimilated C through plant and soils and allowed us to detect relatively small changes in C flows in response to experimental treatments (e.g., Gorissen et al. 1995; Sangster et al. 2010). This approach is more sensitive than measuring total C in plants and soils since small changes in total C content are often difficult to detect against large background amounts of C in plants and soils especially when plants are slow-growing. We specifically hypothesized that (1) total net C uptake increases under increased soil moisture conditions; (2) relatively less C is being allocated belowground with increasing moisture levels, but absolute amounts may increase depending on the total increase in biomass; (3) increased N availability does not affect C allocation unless water availability increases as well; and (4) inorganic C can contribute to (belowground) C fluxes.

Materials and methods

Experimental setup

We used Larrea tridentata (DC.) Coville plants obtained from a nursery in the fall of 2006. The plants were approximately 20 cm tall. During the spring of 2007, 40 seedlings were transplanted into PVC columns (diameter 152 mm; height 250 mm) containing a mixture of 50 % Mojave Desert topsoil (0-20 cm) and 50 % Hydroton[®] extruded clay pebbles by volume. The soil was collected from the Mojave Global Change Facility (MGCF) located on the Nevada Test Site in the northern Mojave Desert, Nye County, Nevada (36°49'N, 115°55' W). Surface soils at MGCF are coarse-textured; gravel content is 40 % of the whole soil mass while the <2 mm fraction is dominated by sand (90 %). Soils are alkaline derived from calcareous alluvium (pH=8.2), and low in organic matter (organic C=1.0 %; inorganic C=2.5 %; Stevenson and Verburg 2006; Marion et al. 2008). The soils were sieved at 2 mm and homogenized. Given that size of the gravel varied widely we added Hydroton® to account for gravel content and to have a more even distribution of gravel material between the soil-plant columns. Soil crust material was mixed within the soil and not left intact.

To assess the effect of moisture on C allocation, gravimetric moisture content in 20 columns was kept at 7 % (w/w), while moisture content in the remaining 20 columns was kept at 15 % (w/w). The amount of moisture was adjusted weekly by weighing the columns. We used de-ionized water for our watering treatments. While soil moisture content is highly variable in the field, we chose to keep moisture constant throughout the experiment but used levels similar to those observed in the field following average and wet winters (Holmes 2009). Consequently, these treatments would reflect conditions of both increased spring and summer moisture. Ten low and ten high moisture columns received 36.4 mg N, equivalent to 40 kg ha^{-1} yr⁻¹ to simulate increased N deposition. This amount was chosen because it is similar to the amount of N added in a companion field study at the MGCF (Barker et al. 2006; Verburg et al. 2013) and reflects current N deposition rates in the vicinity of Los Angeles, California (Bytnerowicz and Fenn 1996). The N was added as 38-0-0 slow-release fertilizer (NITROFORM®). The remaining 20 columns acted as ambient N controls. The columns were randomly assigned to the water and N treatments. In May 2007, all columns were covered with a PVC lid. The lids were sealed with silicone rubber (Q3-3481; Dow Chemical) at the base of the plants to separate the shoot from the root and soil compartment. Each lid had a PVC tube (diameter 2 cm, length 10 cm) containing two layers of soda lime separated by a cotton plug allowing for free exchange of oxygen and trapping CO₂ evolved from the soil surface (soil respiration). Approximately 8 g of soda lime were used in the lower layer to capture soil respiration. The upper layer, containing 6 g soda lime, prevented entrance of CO_2 from the surrounding air (Verburg et al. 1998). A second tube (diameter and length 1 cm) acted as an access port to allow for watering the plants. This tube was sealed with a rubber stopper in between watering.

At the end of May 2007, the columns were divided over two growth chambers with each chamber containing columns subjected to all water and N treatment combinations. The CO₂ concentrations inside the chambers were kept at 380 μ l l⁻¹ CO₂ and temperatures were set at 25°C during the day and 15°C during the night. Temperature settings were triggered by ambient light levels. Maximum photon flux density was 1,866 μ mol photons m⁻² s⁻¹ during the middle of the day. To avoid chamber effects, plants were rotated between growth chambers on a weekly basis. Once per week, 20 plants from one chamber were put inside a UV-transparent acrylic chamber. To allow for tracking uptake and allocation of recently assimilated CO₂, we injected ten cc of pure ¹³C-CO₂ into the chamber using a syringe through a rubber septum, and the CO₂ was allowed to assimilate for 2 h. The air inside the chamber was circulated using a small fan. The same plants were labeled each week. The 20 remaining plants were used as unlabeled controls.

To quantify the amount of C released from the autotrophic (root) and heterotrophic (microbial) respiration, we measured CO_2 respiration bi-weekly throughout the study; and the amount of respired CO_2 was calculated based on the weight increase in soda lime (e.g., Verburg et al. 1998; Serna-Perez et al. 2006). Samples were pooled to have three measurement periods during the experiment. Isotopic values of the respired CO_2 were determined by dissolving the soda lime in concentrated phosphoric acid in an air-tight 250 ml glass jar and sampling the headspace of the jar after all soda lime was dissolved. We used fresh soda lime to account for any CO_2 that may have been absorbed prior to adding the soda lime to the traps.

After being in the growth chambers for 6 months, plants were harvested and separated into leaves, shoots, and roots. All plant materials were weighed, and plant materials and soils were analyzed for C, N (plants only) and δ^{13} C values to calculate C allocation in the different plant and soil compartments. Isotopic composition of soil organic C was measured after removal of carbonates using 1 M phosphoric acid. Total C, N (plants only) and isotopic values of vegetation material, respired CO₂, and organic C were measured at the Colorado Plateau Stable Isotope Facility at Northern Arizona University (Flagstaff, AZ). To estimate the amount of C incorporated in the inorganic C pool isotopic composition of inorganic C was measured at the University of Wyoming Stable Isotope Facility (Laramie, WY). Isotopic composition was reported as δ^{13} C, and total ¹³C uptake of the label was calculated by using isotope ratio and total C data correcting for the amounts of ¹³C present in the unlabeled plant-soil systems (Verburg et al. 1998, 2004).

Data analysis

Our study included four experimental treatments: (1) low moisture and low N (-W-N); (2) low moisture

and high N, (-W+N); (3) high moisture and low N, (+W-N); and (4) high moisture and high N (+W+N) with each treatment initially having five replicates within each labeling treatment. During the experiment a five out of 40 plants died, but each treatment had a minimum of four replicates at the end of the study. We used analysis of variance (ANOVA) to assess effects of water and N treatments on measured parameters for the labeled plants. To test whether or not the isotope label had been incorporated in plants and soils, we used t-tests between control and labeled plant-soil columns. Patterns in respired CO₂ as well as their isotopic composition were analyzed separately for control and labeled chambers using a repeated-measures ANOVA. Statistical analyses were carried out using DataDesk version 6.0.

Results

Plant biomass and total soil respiration

Soil moisture did not affect stem or root biomass, but average leaf biomass increased by 140 % and total biomass increased by 36 % at higher soil moisture levels (Fig. 1). In both cases the increases were marginally significant (p=0.053 for leaf biomass and p=0.073 for total biomass; Table 1). Average stem biomass increased by 31 % while average total biomass increased by 21 % at higher levels of N availability (Fig. 1) but neither of the increases were significant (Table 1). Average shoot/ root ratios increased by 47 % with higher N availability in the low soil moisture treatment (p=0.092; Table 1).

Leaf N content significantly increased by 127 % while total plant N content tended to increase by 32 % (p=0.091) at higher moisture availability (Fig. 1). Stem N content significantly increased by 52 % and total plant N content tended to increase by 31 % (p=0.099) with increasing N availability.

Total soil respiration rates significantly decreased through time (Fig. 2). The N treatment did not significantly affect total soil respiration (Table 1), but the water treatments tended to affect respiration through time as indicated by a marginally significant interaction (p=0.062) between "H₂O" and "time" (not shown) with soil respiration being 60 % higher during the second and 78 % higher during the last time period in the high water treatments (Fig. 2).



Isotopic values of plants, soils and soil respiration

The labeling significantly increased δ^{13} C values of leaves by 254‰ (p<0.001), stems by 92‰ (p<0.001), roots by 77‰ (p<0.001), and soil organic C by 2‰

(p<0.001); but inorganic δ^{13} C values were not significantly affected by the labeling (p=0.701; Figs. 3 and 4). None of the treatments affected isotopic values of the stems and leaves, but the δ^{13} C values of the roots tended to be higher (p=0.090)

Table 1 Analysis of variance results for biomass and isotopic data using water (H2O) and N as main factors

	Leaf	Stem	Roots	Total	S/R	OC	IC	Resp
Biomass ^a								
H_2O	0.053	0.174	0.222	0.073	0.114	-	_	0.131
Ν	0.922	0.149	0.395	0.347	0.458	_	_	0.951
H ₂ O*N	0.998	0.555	0.397	0.896	0.092	_	_	0.183
N content ^b								
H_2O	0.013	0.396	0.423	0.091	0.111	_	_	_
Ν	0.380	0.007	0.281	0.099	0.125	_	_	_
H ₂ O*N	0.627	0.123	0.959	0.842	0.378	_	_	_
$\delta^{13}C^c$								
H ₂ O	0.940	0.297	0.090	_	-	0.005	0.202	0.013
Ν	0.261	0.392	0.358	_	_	0.467	0.694	0.892
H ₂ O*N	0.678	0.463	0.666	_	_	0.443	0.840	0.508
Net ¹³ C uptake	d							
H_2O	0.039	0.139	0.101	0.009	0.935	0.002	_	0.017
Ν	0.415	0.128	0.190	0.309	0.617	0.332	_	0.839
H ₂ O*N	0.518	0.781	0.462	0.531	0.354	0.762	_	0.462
¹³ C allocation ^e								
H_2O	0.346	0.019	0.318	_	_	0.003	_	0.172
Ν	0.491	0.413	0.523	_	_	0.180	_	0.029
H ₂ O*N	0.678	0.518	0.605	-	-	0.258	-	0.165

Significant effects (p < 0.05) are given in bold. Trends (0.1 > p > 0.05) are shown in italics

^a Leaf, stem and root biomass in total grams, soil respiration in mgC/day

^b Leaf, stem and root N content in mg/plant

 $^{c}\,\delta^{13}\,C$ in ‰

^d Net ¹³ C uptake in mg ¹³ C

e13 C allocation in %



Fig. 2 Soil-respired CO₂ from soil-plant systems captured by soda lime in response to water and N addition treatments. $^{-}W-$ N'=low water, low N, $^{-}W+$ N'=low water, high N, $^{+}W-$ N'= high water, low N, and $^{+}W+$ N'=high water and high N

in high water treatments (66.6‰) compared to the low water treatment (39.9‰).

The δ^{13} C values of soil organic C significantly increased with increasing soil moisture from -21.6 to -19.8‰ in the labeled plants, but the N treatments had no effect on the isotopic value of soil organic C. None of the treatments affected the δ^{13} C of inorganic C. Isotopic values of the respired CO₂ significantly increased from -35.4 to -31.0‰ through time in the unlabeled plants with respiration being 0.4‰ higher in the low moisture treatment at the end of the study. In the labeled treatments, isotopic values of respired CO₂ significantly increased through time from -29.0 to -10.2‰, with values being 10‰ higher in the high water treatments at the end of the study (Fig. 5).

Net ¹³C uptake and ¹³C allocation

Total net ¹³C uptake was about twice as high for the high moisture treatment with the increase mainly due to larger amounts being allocated to leaves, soil organic C, and soil respiration (Fig. 6). The ¹³C shoot/root ratio was not affected by the any of the treatments, however. Relative C allocation to stems decreased from 27 to 17 %, while allocation to soil organic C increased from 10 to 25 % under higher soil moisture conditions (Fig. 6). In addition, relative C allocation to soil respiration decreased from 32 to 20 % in the high N treatment. Specific uptake, defined as total ¹³C uptake per unit leaf biomass, increased from 3.34 to 6.41 µg g⁻¹ leaf biomass in the high moisture treatments but was not affected by the N treatment (Fig. 7). Since none of the ¹³C label was incorporated into the inorganic C pool, we



Fig. 3 Isotopic composition of *larrea tridentata* leaves (a), stems (b) and roots (c) in unlabeled and ¹³C-labeled plant-soil systems in response to water and N additions. -W-N'=low water, low N, (-W+N'=low water, high N, (+W-N'=high water, low N, and (+W+N'=high water and high N

did not include inorganic C in the 13 C uptake and allocation calculations.

Discussion

Effects of water availability on biomass and C allocation

We initially hypothesized that (1) total net C uptake would increase under increased soil moisture



Fig. 4 Isotopic composition of organic (a) and inorganic (b) soil C in unlabeled and ¹³C-labeled plant-soil systems in response to water and N additions. '-W-N'=low water, low N, '-W+N'=low water, high N, '+W-N'=high water, low N, and '+W+N'=high water and high N

conditions, and that (2) relatively less C would be allocated belowground with increasing moisture levels. The results from our study showed that leaf and total biomass, N content and ¹³C content as well as total ¹³C uptake either significantly increased or tended to increase with increasing soil moisture in *Larrea tridentata* (Figs. 1 and 6; Table 1) supporting our first hypothesis. In addition, soil respiration was higher in the high water treatment toward the end of the study (Fig. 2). Biomass and ¹³C shoot/root ratios were however not affected by the moisture treatments in contrast to our second hypothesis.

While the overall responses in biomass and soil respiration to changed water availability were not significant, the isotopic data showed clear effects of the water treatments. The absence of a significant effect on biomass may have been due to the relatively short growing period of about 5 months while seedlings were already 2 years old at the start of the study. In addition, the low level of replication may have limited our ability to detect significant effects of moisture even though the absolute increases in leaf and total biomass were quite large (140 % for leaves and 36 % for total biomass). The increase in net C uptake was a direct function of total leaf mass but also leaf photosynthetic activity as indicated by the increased specific ¹³C uptake (Fig. 7), agreeing with Barker et al. (2006) who observed increased photosynthetic activity in response to summer irrigation in the Mojave Desert.

Increased moisture availability resulted in more ¹³C being allocated to soil organic matter and soil respiration (Fig. 6a). Relative C allocation (i.e., fraction of total ¹³C taken up allocated to specific plant-soil components) deviated from this pattern because a larger relative amount of C was allocated to soil organic matter at the expense of C allocated to stems (Fig. 6). Thus, increased moisture availability increased absolute belowground C allocation and proportionately more so to soil organic



Fig. 5 Isotopic composition of soil-respired CO_2 in unlabeled (a) and ¹³C-labeled (b) plant-soil systems in response to water and N additions. '-W-N'=low water, low N, '-W+N'=low water, high N, '+W-N'=high water, low N, and '+W+N'=high water and high N



Fig. 6 Net ¹³C uptake (a) and ¹³C allocation (b) in plant-soil systems exposed to ¹³C-labeled CO_2 in response to water and N additions. '-W-N'=low water, low N, '-W+N'=low water, high

matter. When calculating the ratio of aboveground to belowground allocation as (total ${}^{13}C_{shoots}+total$ ${}^{13}C_{stems}$)/(total ${}^{13}C_{roots}+total$ ${}^{13}C_{soil}$ organic matter+total ${}^{13}C_{soil}$ respiration), however, no effects of the treatments were found showing that—while total C uptake increased under higher moisture availability—the ratio of above-to-belowground allocation remained unchanged contrasting our second hypothesis.

Hunt et al. (1996) and Poorter and Nagel (2000) suggested that C allocation to roots decreases as resource availability increases. Several studies in arid environments have shown increased root proliferation in water and nutrient-rich patches, however (Eissenstat and Caldwell 1988; Jackson and Caldwell 1989; Caldwell et al. 1991). Reynolds et al. (1999) observed that root production in *Larrea tridentata* decreased with reduced summer moisture in a Chihuahuan Desert site in New Mexico. While no aboveground data were presented in these studies, they appear to contrast the idea that



Fig. 7 Specific ${}^{13}C$ uptake per unit leaf biomass in plant-soil systems exposed to ${}^{13}C$ -labeled CO₂ in response to water and N additions. '-W-N'=low water, low N, '-W+N'=low water, high N, '+W-N'=high water, low N, and '+W+N'=high water and high N

N, '+W-N'=high water, low N, and '+W+N'=high water and high N. Error bars represent standard errors of each individual soil-plant compartment

resource limitations increase belowground C allocation. The data from our study suggest that, while increased moisture favors C uptake, no changes in C allocation occurred when considering biomass and ¹³C shoot/root ratios as well as total ¹³C allocation data in contrast to our second hypothesis. More C may, however, be allocated to soil organic matter under higher moisture availability suggesting that production rates of organic matter exceeded potential moisture-induced increases in decomposition rates.

Effects of N availability on biomass and C allocation

We hypothesized that increased N availability would only affect C allocation when moisture limitation were alleviated. Increased N availability did not impact biomass, net ¹³C uptake, specific ¹³C uptake, or C allocation even at higher soil moisture levels rejecting our initial hypothesis. Leaf and total biomass N content increased indicating that N availability affected N uptake but this did not result in a significant increase in biomass production even when moisture levels were higher. Increased N availability resulted in an increase biomass shoot/root ratios at low moisture levels but this pattern was not apparent in the ¹³C data. The lack of responses to increasing N availability was somewhat surprising but consistent with results from a metaanalysis conducted by LeBauer and Treseder (2008) who found no impact of increased N availability on net primary activity (NPP) in three desert ecosystems. In addition, previous studies found no effects of N additions on fine root dynamics (Verburg et al. 2013) or aboveground plant production (Newingham et al.

2012). Barker et al. (2006) observed a significant increase in net photosynthesis in *Larrea tridentata* in response to N additions in combination with increased water availability during the summer at the site from which the soils were collected but this pattern was reversed during the spring. In our study, even at higher moisture availability we did not find an increase in net ¹³C uptake or specific ¹³C uptake suggesting that the patterns we found were more representative of increased spring moisture conditions.

The only effect of N availability observed in this study was a decrease in relative C allocation to soil respiration. Results from previous studies have shown reductions (e.g., Burton et al. 2004; Janssens et al. 2010) and increases (e.g., Deng et al. 2010) in soil respiration in response to N additions in forested environments. Burton et al. (2004) and Janssens et al. (2010) ascribed reductions in soil respiration to decreased belowground C allocation most likely resulting in reduced substrate supply to microbes, thereby reducing heterotrophic respiration. Deng et al. (2010) indicated that increased soil respiration occurring in the first year following N additions may have been caused by increased root biomass in response to N fertilization in a study using tree seedlings. This increased root biomass subsequently could result in increased belowground root litter production that could then stimulate heterotrophic respiration. Our study supports the suggestions from Burton et al. (2004) and Janssens et al. (2010) since previous studies in the Mojave Desert have shown no effects of water, N, and elevated CO₂ concentrations on root biomass (Verburg et al 2013; Ferguson and Nowak 2011; Newingham et al. 2012, 2013) or root respiration (Clark et al. 2010).

Contribution of inorganic C pools to C sequestration

Our last goal was to assess if carbonate C could play a role in ecosystem C sequestration in arid soils. The labeling did not result in a significant change in the δ^{13} C value of the carbonate C suggesting that no new C was incorporated into the carbonate C pool. One potential reason for this lack of a change in the δ^{13} C value may have been that the soils had a relatively large amount of inorganic C (2.5 %), so no clear change could be detected given the large background. Still, the amount of inorganic C; and the δ^{13} C data showed that a significant amount of new C was incorporated into the carbonate shows that a significant amount of new C was incorporated into the solution of the carbonate c was incorporated into the carbonate c

organic C pool especially under high soil moisture availability. In addition, the δ^{13} C data of the respired CO₂ in the labeled chamber suggested a significant amount of new C being present in soil CO₂ which potentially could have been incorporated in the inorganic C pool. Our results agree with Marion et al. (2008) suggesting very little contribution of carbonate C to ecosystem C sequestration. Still, Xie et al. (2009) found large C uptake in carbonate C; but no clear explanation has been given in terms of the mechanisms that could explain this large uptake. In addition, Kuzyakov et al. (2006) observed increases in C sequestration in soil carbonate pools in a loess soil planted with maize. In that study, soil CO₂ concentrations were much higher than those found in most arid soils potentially facilitating more rapid exchange of C between the atmosphere and carbonate pool.

Implications for C sequestration in arid lands

Our study showed that increased soil moisture had a larger impact on biomass production and C uptake and allocation than increased N availability with more C being taken up under wetter conditions supporting our initial hypotheses. The main pools that showed a net new C sequestration were the leaf and soil organic C pools, but the relative amounts of C allocated above- vs. belowground remained unchanged in contrast with our initial hypothesis. A substantial amount of the newly fixed C left the system as soil respiration but the amount of C sequestered in the soil-plant systems was still larger when moisture availability was high. While we observed active C uptake, particularly under wetter conditions, we cannot assess if the sink strength inferred from flux-based measurements (Jasoni et al. 2005; Wohlfahrt et al. 2008) can be explained by accumulations of C in biomass and/or organic matter as suggested by our study. First, we could not quantify the absolute amounts of new C that were incorporated into the various ecosystem pools because we conducted weekly labelings (i.e., a sequence of ¹³C pulses instead of exposure to a continuous ¹³C label; Sangster et al. 2010). Second, we did not measure C release through aboveground plant respiration which is included in flux-based net ecosystem measurements. Third, the plants used in our study were relatively young and most likely showed different C accumulation rates than mature shrubs. Finally, our study did not include intact biotic soil crusts which could potentially play an important role in C cycling in arid ecosystems (e.g., Su et al. 2012; Li et al. 2012). Still, the isotopic data from our study show that these relatively slow-growing plants species actively accumulate C not only in biomass but also in soil organic C pools which could help explain net C uptake observed in previous studies.

The responses of soils and vegetation to moisture availability found in our study were larger than those observed in field studies carried out in the Mojave Desert ecosystems since Newingham et al. (2012) and Verburg et al. (2013) found little change in above- and belowground biomass production after 5-6 years of summer water additions in these systems. Above- and belowground biomass also did not respond to 10 years of exposure to elevated CO₂ concentrations at a site in close proximity to the previously mentioned studies (Newingham et al. 2013; Ferguson and Nowak 2011). The lack of responses to N additions was however similar in our study and related field studies. One potential explanation for the discrepancy between results from our study and previous field studies may be that in our study moisture content was held constant throughout our experiment. As a result, the effects of increased moisture availability are likely to be magnified in our study since under field conditions changes in soil moisture may be more short-lived (episodic). Also, our study was conducted with 2 year old seedlings rather than mature shrubs. Still, Verburg et al. (2013) showed significant effects of winter precipitation on fine root dynamics while summer precipitation did not impact fine roots showing that timing of moisture availability is critical for these ecosystems. Given that we started our study in early spring, our water treatments were more representative of changes in spring water availability when most above- and belowground biomass production occurs. Consequently, if climate change results in differences in winter moisture, C sequestration may be impacted. Comparing results from our study with those from field studies suggest that our N treatments are likely to be more representative of field conditions where N deposition in wet and/or dry forms occurs throughout the year.

Overall, the results from our study suggest that changes in soil moisture may have an impact on C uptake by plants that could result in changing biomass and soil organic C levels in arid systems. These changes could contribute to increased C sequestration under wetter conditions. In contrast, increased N availability in response to atmospheric N deposition is not likely to have a major impact on C uptake in these ecosystems. The data from our study further suggested that inorganic C may not contribute significantly to C cycling in these systems compared to plants and soil organic C.

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References

- Archer SR, Predick KI (2008) Climate change and ecosystems of the Southwestern USA. Rangelands 30:23–28
- Austin AT, Yahdjian L, Stark JM et al (2004) Water pulses and biogeochemical cycles in arid and semiarid ecosystems. Oecologia 141:221–235
- Barker DH, Vanier C, Naumburg E, Charlet TN, Nielsen KM, Newingham BA, Smith SD (2006) Enhanced monsoon precipitation and nitrogen deposition affect leaf traits and photosynthesis differently in spring and summer in the desert shrub *Larrea tridentata*. New Phytol 169:799–808
- BassiriRad H, Tremmel DC, Virginia RA, Reynolds JF, de Soyza AG, Brunell MH (1999) Short-term patterns in water and nitrogen acquisistion by two desert shrubs following a simulated summer rain. Plant Ecol 145:27–36
- Billings SA, Schaeffer SM, Evans RD (2004) Soil microbial activity and N availability with elevated CO₂ in Mojave Desert soils. Glob Biogeochem Cycles 18, GB1011. doi:10. 1029/2003GB002137
- Burton AJ, Pregitzer KS, Crawford JN, Zogg GP, Zak DR (2004) Simulated chronic NO₃⁻ deposition reduces soil respiration in northern hardwood forests. Glob Chang Biol 10:1080– 1091
- Bytnerowicz A, Fenn ME (1996) Nitrogen deposition in California forests: a review. Environ Pollut 92:127–146
- Cable JM, Ogle K, Lucas RW et al (2011) The temperature responses of soil respiration in deserts: a seven desert synthesis. Biogeochemistry 103:71–90
- Caldwell MM, Manwaring JH, Jackson RB (1991) Exploitation of phosphate from fertile soil microsites by three Great Basin perennials when in competition. Funct Ecol 5:757–764
- Cayan DR, Das T, Pierce DW, Barnett TP, Tyree M, Gershunova A (2010) Future dryness in the southwest US and the hydrology of the early 21st century drought. Proc Natl Acad Sci U S A 107:21271–21276
- Clark NM, Apple ME, Nowak RS (2010) The effects of elevated CO₂ on root respiration rates of two Mojave Desert shrubs. Glob Chang Biol 16:1566–1575
- Deng Q, Zhou G, Liu J, Liu S, Duan H, Zhang D (2010) Responses of soil respiration to elevated carbon dioxide and nitrogen addition in young subtropical forest ecosystems in China. Biogeosciences 7:315–328

- Dijkstra FA, Cheng W (2007) Interactions between soil and tree roots accelerate long-term soil carbon decomposition. Ecol Lett 10:1046–1053
- Eissenstat DM, Caldwell MM (1988) Seasonal timing of root growth in favorable microsites. Ecology 69:692–695
- Emmerich WE (2003) Carbon dioxide fluxes in a semiarid environment with high carbonate soils. Agric For Meteorol 116:91–102
- Eswaran H, Reich PF, Kimble JM, Beinroth FH, Padmanabhan E, Moncharoen P (2000) Global carbon sinks. In: Lal R, Kimble JM, Eswaran H, Stewart BA (eds) Global climate change and pedogenic carbonates. CRC Press, Boca Raton, pp 15–26
- Fawcett PJ, Werne JP, Anderson RS et al (2011) Extended megadroughts in the southwestern United States during Pleistocene interglacials. Nature 470:518–521
- Fenn ME, Haeuber R, Tonnesen GS et al (2003) Nitrogen emissions, deposition, and monitoring in the western United States. Bioscience 53:391–403
- Ferguson SD, Nowak RS (2011) Transitory effects of elevated atmospheric CO₂ on fine root dynamics in an arid ecosystem do not increase long-term soil carbon input from fine root litter. New Phytol 190:953–967
- Friggens MM, Warwell MV, Chambers JC, Kitche SG (2012) Modeling and predicting vegetation response of western U.S.A. grasslands, shrublands, and deserts to climate change. In: Finch D (ed) Climate change in grasslands, shrublands, and deserts of the interior American West: A review and needs assessment. Gen. Tech. Rep. RMRS-GTR-285. U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station, Fort Collins, pp 1–20
- Gebauer RLE, Ehleringer JR (2002) Interspecific competition and resource pulse utilization in a cold desert community. Ecology 83:2602–2616
- Gorissen A, Kuikman PJ, van de Beek H (1995) Carbon allocation and water use in juvenile Douglas fir under elevated CO₂. New Phytol 129:275–282
- Hamerlynck EP, McAuliffe JR, Smith SD (2000) Effects of surface and sub-surface soil horizons on the seasonal performance of *Larrea tridentata* (creosotebush). Funct Ecol 14: 596–606
- Holmes C (2009) Effects of three global climate change factors on soil water and sap flow of *Larrea tridentata* in the Mojave Desert. University of Nevada, Reno, Dissertation
- Housman DC, Naumburg E, Huxman TE, Charlet TN, Nowak RS, Smith SD (2006) Increases in desert shrub productivity under elevated carbon dioxide vary with water availability. Ecosystems 9:374–385
- Hunt HW, Elliott ET, Detling JK, Morgan JA, Chen DX (1996) Responses of a C_3 and C_4 perennial grass to elevated CO_2 and climate change. Glob Chang Biol 2:35–47
- Huxman TE, Snyder KA, Tissue D et al (2004) Precipitation pulses and carbon fluxes in semiarid and arid ecosystems. Oecologia 141:254–268
- Jackson RB, Caldwell MM (1989) The timing and degree of root proliferation in fertile-soil microsites for three cold-desert perennials. Oecologia 81:149–153
- Janssens I, Dieleman W, Luyssaert S et al (2010) Reduction of forest soil respiration in response to nitrogen deposition. Nat Geosci 3:315–322
- Jasoni RL, Smith SD, Arnone JA III (2005) Net ecosystem CO₂ exchange in Mojave Desert shrublands during the eighth year of exposure to elevated CO₂. Glob Chang Biol 11:749–756

- Jin VL, Evans RD (2007) Elevated CO₂ increases microbial carbon substrate use and N cycling in Mojave Desert soils. Glob Chang Biol 13:452–465
- Kuzyakov Y, Shevtzova E, Pustovoytov K (2006) Carbonate recrystallization in soil revealed by ¹⁴C labeling: Experiment, model and significance for paleo-environmental reconstructions. Geoderma 131:45–58
- Lal R, Kimble JM (2000) Inorganic carbon and the global C cycle: Research and development priorities. In: Lal R, Kimble JM, Eswaran H, Stewart BA (eds) Global climate change and pedogenic carbonates. CRC Press, Boca Raton, pp 291–302
- Lapenis AG, Lawrence GB, Bailey SW, Aparin BF, Shiklomanov AI, Speranskaya NA (2008) Climatically driven loss of calcium in steppe soil as a sink for atmospheric carbon. Glob Biogeochem Cycles 22, GB2010
- LeBauer DS, Treseder KK (2008) Nitrogen limitation of net primary productivity in terrestrial ecosystems is globally distributed. Ecology 89:371–379
- Li X, Zhang P, Su YG, Jia RL (2012) Carbon fixation by biological soil crusts following revegetation of sand dunes in arid desert regions of China: A four-year field study. Catena 97: 119–126
- Marion GM, Verburg PSJ, McDonald EV, Arnone JA III (2008) Modeling salt movement through a Mojave Desert soil. J Arid Environ 72:1009–1030
- Miller RE, Huenneke LF (1996) Size decline in *Larrea tridentata* (creosotebush). Southwest Nat 41:248–250
- Newingham BA, Vanier CH, Charlet TN, Smith SD (2012) Effects of enhanced summer monsoons, nitrogen deposition and soil disturbance on *Larrea tridentata* productivity and resulting herbivory in the Mojave Desert. J Arid Environ 87:19–28
- Newingham BA, Vanier CH, Charlet TN, Ogle K, Smith SD, Nowak RS (2013) No cumulative effect of ten years of elevated [CO2] on perennial plant biomass components in the Mojave Desert. Glob Chang Biol. doi:10.1111/gcb. 12177
- Notaro M, Holman K, Zarrin A, Fluck E, Vavrus S, Bennington V (2012) Influence of the Laurentian Great Lakes on Regional Climate. J Clim. doi:10.1175/JCLI-D-12-00140.1
- Noy-Meir I (1973) Desert ecosystems: environment and producers. Annu Rev Ecol Syst 4:23–51
- Phillips DL, Johnson MG, Tingey DT, Catricala CE, Hoyman TL, Nowak RS (2006) Effects of elevated CO₂ on fine root dynamics in a Mojave Desert community: a FACE study. Glob Chang Biol 12:61–73
- Poorter H, Nagel O (2000) The role of biomass allocation in the growth response of plants to different levels of light, CO₂, nutrients and water: a quantitative review. Aust J Plant Physiol 27:595–607
- Reynolds JF (2001) Desertification. In: Levin SA (ed) Encyclopedia of biodiversity. Academic Press, San Diego, pp 61–78
- Reynolds JF, Virginia RA, Kemp PR, De Soyza AG, Tremmel DC (1999) Impact of drought on desert shrubs: Effects of seasonality and degree of resource island development. Ecol Monogr 69:69–106
- Rundel PW, Gibson AC (1996) Ecological Communities and Processes in a Mojave Desert Ecosystem: Rock Valley. Cambridge University Press, Cambridge, UK, Nevada
- Sangster A, Knight D, Farrell R, Bedard-Haughn A (2010) Repeat-pulse ¹³CO₂ labeling of canola and field pea:

implications for soil organic matter studies. Rapid Commun Mass Spectrom 24:2791–2798

- Schlesinger WH, Belnap J, Marion GM (2009) On carbon sequestration in desert ecosystems. Glob Chang Biol 15:1488–1490
- Schwinning S, Davis K, Richardson L, Ehleringer JR (2002) Deuterium-enriched irrigation indicates different forms of rain use in shrub/grass species of the Colorado Plateau. Oecologia 130:345–355
- Schwinning S, Starr BI, Ehleringer JR (2005a) Summer and winter drought in a cold desert ecosystem (Colorado Plateau) part I: effects on soil water and plant water uptake. J Arid Environ 60:547–566
- Schwinning S, Starr BI, Ehleringer JR (2005b) Summer and winter drought in a cold desert ecosystem (Colorado Plateau) part II: effects on plant carbon assimilation and growth. J Arid Environ 61:61–78
- Seager R, Ting M, Held I et al (2007) Model projections of an imminent transition to a more arid climate in southwestern North America. Science 316:1181–1184
- Serna-Perez A, Monger HC, Herrick JE, Murray L (2006) Carbon dioxide emissions from exhumed petrocalcic horizons. Soil Sci Soc Am J 70:795–805
- Smith SD, Monson RK, Anderson JE (1997) Physiological ecology of North American desert plants. Springer, Berlin
- Smith SD, Huxman TE, Zitzer SF et al (2000) Elevated CO₂ increases productivity and invasive species success in an arid ecosystem. Nature 408:79–82
- Snyder KA, Donovan LA, James JJ, Tiller RL, Richards JJ (2004) Summer water pulses do not necessarily lead to canopy growth of Great Basin and Northern Mojave Desert shrubs. Oecologia 141:325–334
- Solomon S, Qin D, Manning M et al (2007) Technical summary. In: Solomon S, Qin D, Manning M, Chen Z, Marquis M,

Averyt KB, Tignor M, Miller HL (eds) Climate change 2007: the physical science basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge University Press, Cambridge, pp 19–92

- Stevenson B, Verburg PSJ (2006) Effluxed CO₂-¹³C from sterilized and unsterilized treatments of a calcareous soil. Soil Biol Biochem 38:1727–1733
- Stone R (2008) Have desert researchers discovered a hidden loop in the Carbon cycle? Science 320:1409–1410
- Su YZ, Wang XF, Yang R, Lee J (2012) Characteristics of carbon flux in two biologically crusted soils in the Gurbantunggut Desert, Northwestern China. Catena 96:41–48
- Verburg PSJ, Gorissen A, Arp WJ (1998) Carbon allocation and decomposition of root-derived organic matter in a plant-soil system of *Calluna vulgaris* as affected by elevated CO₂. Soil Biol Biochem 30:1251–1258
- Verburg PSJ, Cheng W, Johnson DW, Schorran DE (2004) Nonsymbiotic nitrogen fixation in three-year-old Jeffrey pines and the role of elevated [CO₂]. Can J For Res 34:1979–1984
- Verburg PSJ, Young AC, Stevenson BA, Glanzmann I, Arnone JA III, Marion GM, Holmes C, Nowak RS (2013) Do increased summer precipitation and N deposition alter fine root dynamics in a Mojave Desert ecosystem? Glob Chang Biol 19:948–956
- Weltzin JF, Loik ME, Schwinning S et al (2003) Assessing the response of terrestrial ecosystems to potential changes in precipitation. Bioscience 53:941–952
- Wohlfahrt G, Fenstermaker LF, Arnone JA III (2008) Large annual net ecosystem CO₂ uptake of a Mojave Desert ecosystem. Glob Chang Biol 14:1475–1487
- Xie J, Li Y, Zhai C, Li C, Lan Z (2009) CO_2 absorption by alkaline soils and its implication to the global carbon cycle. Environ Geol 56:953–961