REGULAR ARTICLE

Contribution of winter cover crop amendments on global warming potential in rice paddy soil during cultivation

Sang Yoon Kim · Chang Hoon Lee · Jessie Gutierrez · Pil Joo Kim

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Abstract

Background and aims Winter cover crop cultivation during the fallow season has been strongly recommended in mono-rice paddy soil to improve soil quality, but its impact in increasing the greenhouse gases (GHGs) emissions during rice cultivation when applied as green manure has not been extensively studied. In order to recommend a preferable cover crop which can increase soil productivity and suppress GHG emission impact in paddy soil, the effect of winter cover crop addition on rice yield and total global warming potential (GWP) was studied during rice cultivation.

Methods Two cover crops (Chinese milk vetch, *Astragalus sinicus* L., hereafter vetch, and rye, *Secale cerealis*) having different carbon/nitrogen (C/N) ratios were cultivated during the rice fallow season. The fresh above-ground biomasses of vetch [25 Mg fresh weight (FW) ha⁻¹, moisture content (MC) 86.9 %, C/N ratio 14.8] and rye (29 Mg rye FW ha⁻¹, MC 78.0 %, C/ N ratio 64.3) were incorporated as green manure 1 week before rice transplanting (NPK + vetch, and NPK + rye).

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P. J. Kim

Institute of Agriculture and Life Science, Gyeongsang National University, Jinju 660-701, South Korea The NPK treatment was installed for comparison as the control. During the rice cultivation, methane (CH₄) and nitrous oxide (N₂O) gases were collected simultaneously once a week using the closed-chamber method, and carbon dioxide (CO₂) flux was estimated using the soil C balance analysis. Total GWP impact was calculated as CO₂ equivalents by multiplying the seasonal CH₄, CO₂, and N₂O fluxes by 25, 1, and 298, respectively.

Results Methane mainly covered 79-81 % of the total GWP, followed by CO₂ (14-17 %), but the N₂O contribution was very small (2-5 %) regardless of the treatment. Seasonal CH₄ fluxes significantly increased to 61 and 122 % by vetch and rye additions, respectively, compared to that of the NPK treatment. Similarly, the estimated seasonal CO₂ fluxes increased at about 197 and 266 % in the vetch and rye treatments, respectively, compared with the NPK control plots. Based on these results, the total GWP increased to 163 and 221 % with vetch and rye applications, respectively, over the control treatment. Rice productivity was significantly increased with the application of green manure due to nutrient supply; however, vetch was more effective. Total GWP per grain yield was similar with the vetch (low C/N ratio) and NPK treatments, but significantly increased with the rye (high C/N ratio) application, mainly due to its higher CH₄ emission characteristic and lower rice productivity increase. Conclusions A low C/N ratio cover crop, such as vetch, may be a more desirable green manure to reduce total GWP per grain yield and to improve rice productivity.

Keywords Greenhouse gases · Green manure · Methane · Nitrous oxide · Global warming potential · Paddy soil · Rice

S. Y. Kim · C. H. Lee · J. Gutierrez · P. J. Kim (⊠) Division of Applied Life Science (BK 21 Program), Gyeongsang National University, Jinju 660-701, South Korea e-mail: pjkim@gnu.ac.kr

Introduction

In temperate zone countries like Korea, Japan and China, winter cover crop cultivation as green manure has been strongly recommended to improve soil quality. In particular, green manuring is considered an important management practice to increase the soil organic matter level and reduce the dependence on mineral fertilizers (Elfstrand et al. 2007). In rice paddy soil, there are two groups of winter cover crops, high biomass yielding non-leguminous plants and N-fixing leguminous plants. For example, leguminous vetch, like Chinese milk vetch (Astragalus sinicus) or hairy vetch (Vicia villosa), can increase the soil N content through symbiotic N fixation (Kim et al. 2007; Na et al. 2007), while non-leguminous cover crops, like rye (Secale cerealis) and barley (Hordeum vulgare), have comparatively higher biomass productivity (Zhang et al. 2007). These winter cover crops are usually sown in late autumn, harvested the next spring, and then incorporated as green manure before rice cultivation. Both cover crops have a strong resistance to a hard, cold winter climate.

The application of cover crop as green manure may increase GHGs emission like CH₄, CO₂, and N₂O, but this impact has not been well studied. In particular, rice cultivation in the flooded paddy soil is one of the major anthropogenic sources of CH₄, which has a 25 times higher GWP than carbon dioxide (CO2) over a 100-year time horizon (IPCC 2007). Since CH₄ is mainly produced from the decomposition of organic matter by the methanogenic archaea under extremely reduced conditions (Garcia et al. 2000), cover crop application may markedly increase CH₄ emission during rice cultivation. In comparison, the contribution of N₂O to global warming is 298 times higher than CO₂; however, its emission from rice paddy soil during cultivation may be negligible (Kreye et al. 2007). Since N₂O emission level generally depends on the amount of N available in the soil (Bouwman 1996; Brown et al. 2000; Maggiotto et al. 2000), high N containing cover crop like vetch may increase largely N₂O emission, but the effect of cover crop application is not clear.

There were numerous studies done about the GHG emission characteristics in rice paddy soils. However, such studies only focused on the individual GHG emission impact without the overall quantification of the total GWP from the combined emission contributions of these main GHGs like CH₄, CO₂, and N₂O (Rath et al. 1999; Naser et al. 2007). To properly

control the total global warming impact of these GHGs, it is necessary to simultaneously investigate their individual contributions during the cultivation season (Nishimura et al. 2011; Burney et al. 2010). Among the major cereals in the world, rice has comparatively higher GWP at 3.8 Mg CO₂ ha⁻¹ season⁻¹ than wheat (*Triticum aestivum*, 0.7 Mg ha⁻¹ season⁻¹) and maize (*Zea mays*, 1.4 Mg ha⁻¹ season⁻¹) suggesting the importance of mitigating the GWP for rice systems (Linguist et al. 2012; Godfray et al. 2011).

In order to select a preferable cover crop which can improve soil quality and reduce GHG emission impact, we compared the effects of two representative cover crops having different C/N ratio on rice productivity and total GWP by monitoring the seasonal CH_4 , CO_2 , and N_2O fluxes during rice cultivation.

Materials and methods

Winter cover cropping and rice cultivation

Winter cover cropping followed by rice cultivation was carried out in the Agronomy Field 1, Gyeongsang National University, Jinju (35°8'56.73"N, 128°5' 46.27"E), South Korea in 2010. The soil was classified as fine silty, mixed, mesic Typic Endoaquepts. The soil pH before the study was neutral and had low fertility (Table 1). The 10 m \times 10 m treatment plots were in a randomized block design and replicated three times. The recommended rates of vetch (50 kg ha^{-1}) and rye seeds (120 kg ha⁻¹) were sown after rice harvest in mid-October 2009, harvested in late May 2010, and their nutrients were determined after harvest by collecting the above parts of the cover crops, washed by tap water, oven-dried at 70 °C for 72 h, ground and then digested using an acid solution (H₂O:H₂SO₄:HClO₄, 5:1:9 volume/volume) (Table 1). The above-ground fresh biomass of vetch and rye were cut into small pieces and mechanically mixed into the soil surface (0-15 cm depth) 1 week before rice transplanting.

Mineral fertilizers (NPK) were applied to the cover crop treatments (NPK + vetch and NPK + rye) at rates of N:P₂O₅:K₂O=110:45:58 kg ha⁻¹, which is the Korean recommended fertilization level for rice cultivation (RDA 1999). The mineral fertilizer treatment alone (NPK) was the control. The basal mineral fertilizers applied 1 day before transplanting were 55 kg N ha⁻¹, 45 kg P₂O₅ ha⁻¹, and 40.6 kg K₂O ha⁻¹. Thirty-

Table 1 Chemical properties of initial soil, and cover crops used	Soil	Values	Cover crops	Rye	Vetch
for the field experiment	Bulk density (g cm^{-3})	1.2±0.2	Moisture contents $(\%, \text{ wt wt}^{-1})$	78.0±2.5	86.9±3.2
	pH (1:5 with H_2O)	H ₂ O) 6.6 ± 0.3 Total concentration $(g kg^{-1})^a$	Total concentration $(g kg^{-1})^a$		
	Total organic matter (g kg ⁻¹)	$15.5{\pm}0.4$	С	540±23	521 ± 18
	Available phosphorus (mg kg ⁻¹)	30.3 ± 6.2	Ν	$8.4{\pm}0.8$	35 ± 3.8
	Exchangeable cation $(\text{cmol}^+ \text{kg}^{-1})$		Р	$2.8 {\pm} 0.3$	17±1.5
	K	$0.2 {\pm} 0.1$	K	21.2 ± 3.4	34.6±4.6
	Ca	$6.1 {\pm} 0.3$	Ca	2.7±0.5	10.0 ± 1.7
^a All chemical properties are based on dried matter	Mg	$1.1{\pm}0.1$	C/N ratio	$64.3{\pm}8.6$	14.8 ± 1.1

day-old seedlings (3 plants per hill) of rice (cultivar,

Dongjinbyeo, Japonica) were transplanted by hand on June 8, 2010, at a spacing of 30 cm×15 cm. Tillering fertilizer (22 kg N ha⁻¹) was broadcast about 2 weeks after rice transplanting and panicle fertilizer (33 kg N ha^{-1} , 17.4 kg K₂O ha^{-1}) 6 weeks after transplanting. The rice was harvested on October 21, 2010, and its productivity was recorded following the RDA methods (RDA 1995). Throughout the cropping season, the water level was maintained at a depth of 5-7 cm above the soil surface by using an automatic water level controller.

Methane and N₂O gas sampling and analysis

Methane and N₂O emission characteristics were investigated during the rice cropping season using the closed-chamber method (Rolston 1986). In each plot, three transparent acryl chambers (width 62 cm, length 62 cm, and height 112 cm) were placed permanently into the soil after transplanting the rice seedlings. There were four holes in the bottom of each chamber to control the amount of floodwater. The chamber was equipped with a circulating fan to ensure complete gas mixing during the period of sampling. Eight rice plants were covered by each chamber.

Gas sampling was carried out 3 times a day (08:00, 12:00, and 16:00 hours) to determine the average daily CH₄ emission during the cropping season. In detail, gas samples in triplicates were collected once a week using 50-ml air-tight plastic syringes at 0-, 15-, and 30-min intervals after manually closing the chamber. The collected gas samples were transferred into 30-ml airevacuated glass vials sealed with a butyl rubber septum.

Methane concentrations were measured using a gas chromatograph (Shimadzu, GC-2010) with a stainless steel column packed with Porapak NQ column (Q 80-100 mesh) and a flame ionization detector (FID). The temperature of the column, injector and detector were adjusted to 80, 100, and 110 °C, respectively. Nitrous oxide concentrations were determined using a similar gas chromatograph (Shimadzu, GC-2010) with a stainless steel column packed with Porapak Q column (Q 80-100 mesh) but equipped with a ⁶³Ni electron capture detector (ECD). The temperature of the column, injector, and detector were adjusted at 70, 80, and 320 °C, respectively. Helium and H₂ gases were used as the carrier and burning gases, respectively.

Total global warming potential (GWP) calculation

Methane and N₂O emission rates were calculated from the increase in CH₄ and N₂O concentrations per unit surface area of the chamber during a specific time interval. The following closed-chamber equation was used to estimate the CH₄ and N₂O fluxes from each treatment (Rolston 1986):

$$F = \rho \times (V/A) \times (\Delta c/\Delta t) \times (273/T)$$

where F is the CH₄ flux (mg CH₄ m⁻² h⁻¹) or N₂O flux (μ g N₂O m⁻² h⁻¹), ρ is the gas density of CH₄ or N_2O under a standardized state (mg cm⁻³), V is the volume of the chamber (m^3) , A is the chamber area (m^2) , $\Delta c/\Delta t$ is the rate of CH₄ or N₂O gas accumulation in the chamber (mg m⁻³ h⁻¹ for CH₄, µg m⁻³ h⁻¹ for N_2O), and T is the absolute temperature (273+ mean temperature in the chamber, °C).

The seasonal CH₄ or N₂O flux for the entire cropping period was computed by the formula of Singh et al. (1999):

Seasonal CH₄or N₂O flux =
$$\sum_{i}^{n} (R_i \times D_i)$$

where $R_{\rm i}$ is the rate of CH_4 or N_2O flux (g m^{-2} day^{-1} for CH₄, mg m⁻² day⁻¹ for N₂O) in the *i*th sampling interval, D_i is the number of days in the *i*th sampling interval, and n is the number of sampling intervals.

However, it was not possible to quantify the CO₂ emission rate along with the CH₄ and N₂O emission rates with the use of the closed-chamber method, since the CO₂ might be absorbed by the plant during photosynthesis. In this study, we indirectly estimated the CO_2 flux using C budgeting (Lal 2002). The instantaneous

level of the pool was assessed by computing the balance between input and output.

$$(SOC)_{g} = antecedent pool + input - losses$$

$$(SOC)_{g} = C_{o} + (C_{r} + C_{b}) - (C_{e} + C_{l} + C_{m})$$

where $(SOC)_g$ is the gross SOC pool, C_o is the antecedent SOC pool, $C_{\rm r}$ is the addition of C in crop residue, and $C_{\rm b}$ is the addition as other biosolids. The losses of soil C may be due to accelerated erosion (C_e), leaching as DOC (C_l), and mineralization or oxidation (C_m) . The losses of soil C by mineralization or oxidation (C_m) included CH₄ and CO2 emission losses during rice cultivation.

The gross SOC pool and the antecedent SOC pool were evaluated by soil C stocks before and after rice cultivation, and the soil C stock was calculated using the soil C content and bulk density in the surface layer (0-15 cm) (Shang et al. 2010) (Table 2). Since rice straw is

Table 2 Soil properties, rice growth and yield characteristics at rice harvesting stage	Parameters	Treatments			LSD _{0.05}	
		NPK	NPK + rye	NPK + vetch		
	Soil properties					
	pH (1:5, H ₂ O)	6.5	6.6	6.4	ns	
	Mean Eh value (mV)	-171	-200	-186	13.6	
	Total C (g kg^{-1})	8.7	9.9	9.2	0.6	
	Total N (g kg ⁻¹)	0.49	0.50	0.55	0.03	
	Available P_2O_5 (mg kg ⁻¹)	63	62	101	14	
	Exchangeable cations $(\text{cmol}^+ \text{kg}^{-1})$					
	K^+	0.3	0.3	0.3	ns	
	Ca ²⁺	8.2	7.9	8.2	ns	
	Mg^{2+}	0.5	0.6	0.5	ns	
	Hot water extractable C (mg kg^{-1})	636	957	796	127	
	PLFA profile (umol kg ⁻¹)					
	Total phospholipid fatty acids	138	238	221	51	
	Bacteria	78.8	162.8	120.4	26.5	
	Actinomycetes	14.3	25.2	17.8	3.0	
	Fungi	15.9	27.7	29.3	12.5	
	Rice growth and yield characteristics					
	Plant height (cm)	93.7	95.3	105.6	3.2	
	Tiller number per hill	11.7	12.4	12.8	ns	
	Straw yield (Mg ha ⁻¹)	5.7	6.6	7.0	0.5	
	Total biomass (Mg ha ⁻¹)	11.2	13.6	14.3	0.6	
LSD _{0.05} least significant differ-	Root volume per hill (cm ³)	33.4	36.4	48.4	5.4	
ence at the 5 % level, <i>ns</i> not significant	Root weight per hill (g)	29.8	32.7	44.1	4.3	

LSD_{0.05} least significant difference at the 5 % level, ns not significant

generally removed in paddy soils for animal feeding, only the rice roots and applied cover crop biomass were considered as the added C from crop residue (C_r) and other biosolids (C_b), respectively.

The losses of soil C by leaching (C_l) and erosion (C_e) during the rice cultivation were not measured in this study but were estimated using the organic N loss data in general Korean paddy soils. Since soil organic C has a mean C/N ratio of 10 (McLean 1930; Kendall et al. 2001), 63.6 and 315.8 kg C ha⁻¹ of C leaching (C_l) and erosion loss (C_e) , respectively, were estimated from the 6.36 and 31.6 kg N ha⁻¹ of organic N leaching and erosion losses, respectively, during rice cultivation (Han et al. 1998; Cho et al. 2000).

With the information on the flux of GHGs over the rice growing season, the relative ability of gases, also called the global warming potential (GWP) of a production system, were expressed in terms of CO_2 equivalent (Robertson et al. 2000), and the GWP is 1 for CO_2 , 25 for CH_4 , and 298 for N₂O (IPCC 2007):

$$GWP(CO_2 \ equivalent) = CO_2 + 25(CH_4) + 298(N_2O)$$

Investigation of soil properties and rice yield characteristics

The Eh electrode was installed permanently at 0-5 cm soil depth throughout the rice cultivation period. The soil redox potential (Eh) was measured in each plot during gas sampling using an Eh meter (PRN-41, DKK-TOA).

The analysis of other soil chemical properties was performed using the soil samples after the rice harvest, collected at 0–15 cm soil depth from five selected points in each plot, air-dried, and sieved (<2 mm). The chemical analysis included soil pH (1:5, with H₂O), exchangeable cations (Ca²⁺, Mg²⁺, and K⁺ with 1 N ammonium acetate solution at pH 7.0), available phosphate (RDA 1988), hot-water extractable C (extracted at 80 °C; Ghani et al. 2000), and total C and N concentrations (CHNS-932 Analyzer; Leco, USA).

Fresh soil samples were collected at the harvesting stage and the fatty acids were extracted using the FAME (fatty acid methyl ester) methodology to evaluate the microbial activity and diversity. The total microbial biomass was evaluated using phospholipid fatty acid (PLFA) extraction according to the Bligh–Dyer method (Frostegard et al. 1996; Zelles and Bai 1993). The resulting lipid material was fractionated into neutral lipids, glycolipids, and phospholipids in a silica-bonded phase column (SPE-SI; Bond Elute, Analytical Chem International, USA) by elution with chloroform, acetone, and methanol, respectively. After mild alkaline hydrolysis, the resulting fatty acid methyl esters were separated using a capillary gas chromatograph and identified by the MIS automatic identification system (Agilent 6850, FID, TSBA50; MIDI, Sherlock, USA). Individual phospholipid fatty acids were designated by the total number of carbon atoms:number of double bonds. The position of the double bond is defined by the symbol ' ω ' followed by the number of carbons from the methyl end of the fatty acid molecule. The prefixes 'i' and 'a' indicate iso- and anteiso-branching, respectively, and 'cy' indicates a cyclopropane fatty acid. 'Me' refers to the position of the methyl group from the carboxyl-end of the chain. The sum of the PLFAs considered to be predominantly of bacterial origin (i15:0, a15:0, 15:0, i16:0, 16:1w7c, 16:1w5c, i17:0, a17:0, cy17:0, 17:0, $18:1\omega7c$, cy19:0) were chosen to represent the bacterial biomass (Frostegard et al. 1993). The fatty acids 10Me 16:0, 10Me 17:0, and 10Me 18:0 were used as an indicator of actinomycetes (Klose et al. 2006). The fatty acids $18:2\omega 6,9$ and $18:2\omega 6,9c$ were used to signify the fungal biomass (Frostegard et al. 1996; Olsson 1999).

Rice plant growth parameters such as plant height, straw yield, total biomass, root weight, and root volume were determined at the harvesting stage. The root volume was measured by the water displacement method (Kar and Ghildyal 1975).

Statistical analyses were conducted using SAS software (SAS Institute 2003). A one-way ANOVA was carried out to compare the means of the different treatments. Fisher's protected least significant difference (LSD) was calculated at the 0.05 probability level for making treatment mean comparisons.

Results

Changes of soil redox potential

The soil Eh values gradually decreased from +150 mV in the control (NPK) treatment with flooding and at 10 days after transplanting reached less than -200 mV. This extremely reduced soil condition is suitable for CH₄ production (Fig. 1). However, the soil Eh values had immediately dropped below -200 mV after flooding in the cover crop

Fig. 1 Soil Eh values, and CH₄ and N₂O emission rates in rice paddy soil during cultivation



applied treatments (NPK + vetch, and NPK + rye) and remained within -200 to -240 mV throughout the entire flooding period. This value rapidly increased after the final drainage 2 weeks before the harvesting. The cover crop amended treatments gave statistically lower mean Eh values compared with the control. Methane, nitrous oxide, and carbon dioxide fluxes

Methane emission patterns showed a near inverse relationship with the changes of the Eh values during rice cultivation (Fig. 1). In the NPK treatment, CH_4 emission was comparably lower at the initial rice

growth stage and increased with the development of soil reductive conditions and rice growth. In comparison, CH_4 emission rates increased from the early vegetative growth stage of the rice plants in the two cover crop applications plots. Net CH_4 emission rates almost dropped to near zero values at the grain maturation stage, irrespective of the treatment.

The seasonal CH₄ flux was 483 kg ha⁻¹ in the NPK treatment during rice cultivation, and the cover crop additions as green manure significantly increased (P< 0.05) the seasonal CH₄ flux to 61 and 122 % by vetch and rye applications, respectively (Fig. 2). However, vetch having a low C/N ratio had lower CH₄ emissions than rye with its high C/N ratio. Labile (hot water-extractable) and total C concentrations in the soil at the rice harvesting stage were significantly increased (P< 0.05) by the two cover crop additions (Table 2). These increases were more pronounced with the addition of rye than vetch, which may have resulted in the significant increase of CH₄ emissions during rice cultivation.

The N2O emission patterns and rates were different from those of CH₄ (Fig. 1). Nitrous oxide emission rates were negligible in the early rice growing season and became slightly higher in a short time period during the rice panicle formation stage, probably due to the N fertilization and the warm temperature. However, N₂O emission rates were generally very low during the whole rice cultivation, compared to the CH₄ emission rates. Comparing the N₂O emission rates in NPK, the vetch addition slightly increased the emission rates while they slightly decreased with the rye addition. The seasonal N₂O flux was just 2.5 kg ha⁻¹ in the NPK treatment and increased 20 % by the application of low C/N ratio vetch. In contrast, the high C/N ratio rye addition slightly decreased the seasonal N2O flux by 8 % compared with the NPK treatment (Fig. 1).

To estimate the seasonal CO₂ loss, we used the C budgeting analysis in this study. For example, the total C input was estimated to be 847 kg C ha⁻¹, which originated from the two main C sources (urea: 20 kg C ha⁻¹; rice root biomass: 827 kg C ha⁻¹) (Table 3). We also computed the soil C stock by using the soil organic C content and bulk density (Lee et al. 2009), and its difference before and after rice cultivation was approximated to be -475 kg C ha⁻¹. The difference between the total C input (847 kg C ha⁻¹) and soil C stock change (-475 kg C ha⁻¹), which included

the C mineralization loss (362 kg Cha^{-1} of CH_4 loss, and CO₂ loss) and the leaching and erosion losses $(379 \text{ kg C ha}^{-1})$ during rice cultivation. Since we did not investigate the C leaching and erosion losses, we used the average C lost by leaching (64 kg C ha^{-1}) and erosion (316 kg C ha⁻¹) of a general Korean rice paddy soil during rice cultivation (Han et al. 1998; Cho et al. 2000). Based on these results, around 151 kg C ha⁻¹ of CO₂ emission loss was estimated in the NPK treatment and significantly increased in the NPK + rye and NPK + vetch treatments due to higher amounts of C additions from the cover crop biomass (rye: 3,446 kg C ha⁻¹; vetch: 1,709 kg ha⁻¹). Between the two selected cover crops, rye addition more significantly increased the CO₂ flux, to 266 % of that of the NPK, than the vetch treatment (197 %) (Fig. 2). Based on this result, lower C/N ratio vetch was more effective on regulating CO₂ emission than rye, having a higher C/N ratio, during rice cultivation.

Global warming potentials

Irrespective of the fertilization, CH_4 was the most influential GHG in increasing the growth scale of the total GWP during rice cultivation (Fig. 2). The contribution of the seasonal CH_4 flux to the total GWP was 79–81 %, followed by the estimated CO_2 flux (14–17 %), but the N₂O flux contribution was very low (2–5 %).

Total GWP was approximately 14.9 Mg CO_2 ha⁻¹ in the NPK during the rice cultivation, but significantly increased in the cover crop applied treatments (Fig. 2). The high C/N ratio rye increased more significantly the total GWP (221 % increase to that of the NPK) than the low C/N ratio vetch (163 % increase). As a result, lower C/N ratio vetch could be more effective in reducing the total GWP impact than the high C/N ratio rye as a green manure in rice paddy soil.

Rice growth and yields and soil properties

The rice grain yield averaged 5.4 Mg ha⁻¹ in the NPK control plot, and significantly ($P \le 0.05$) increased with the application of green manure (Fig. 3). Vetch application increased the grain productivity to 35 % over the NPK treatment, while the effect of rye addition was lower than the vetch application (29 % increase). Vetch addition was more effective than the rye application in improving rice plant growth characteristics

Fig. 2 Seasonal CH₄, CO₂, and N₂O fluxes, and total GWP for the different fertilization treatments during rice cultivation. *NS* not significant



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Table 3 Comparison of soil C stock during rice cultivation

Parameters	Treatm	LSD _{0.05}			
	NPK	NPK + rye	NPK + vetch		
C input (kg C ha ⁻¹)					
Urea	20	20	20		
Rice root ^a	827	827	827		
Green manure	0	3,446	1,709	126.8	
Sum	847	4,293	2,556	126.7	
C output (kg C ha ⁻¹)					
CH ₄ emission	362	802	582	168	
CO ₂ emission	580	1,544	1,143	720	
Leaching and erosion ^b	379	379	379		
Sum	1,322	2,726	2,104	708	
Soil C stock changes (kg C ha ⁻¹) ^c	-475	1,567	452	685	

 a Rice root biomass yield was 2.2 Mg ha $^{-1}\,$ with 37.6 % (wt wt $^{-1}\,$) of total C content (Kim et al. 2009)

^b Leaching and erosion loss data are cited from Han et al. (1998) and Cho et al. (2000)

 $^{\rm c}$ Soil C stock changes were the difference of soil C stock between before and after rice cultivation in the surface layer

like plant height, straw yield, total biomass, root volume per hill, and root weight per hill than rye (Table 2).

The higher N supply with vetch application might have contributed more effectively to rice plant development and yield improvement than rye. For example, 25 Mg FW ha⁻¹ vetch addition could supply around 120 and 28 kg ha⁻¹ of N and P₂O₅, respectively, which were comparable with the 55 and 28 kg ha⁻¹ of N and P₂O₅ addition by the 29 Mg FW ha⁻¹ rye application. In addition, the two cover crop applications significantly improved soil chemical and microbial properties (Table 2). However, rye was more effective on increasing soil C accumulation and boosting microbial activity than vetch at the harvesting stage. Among the microorganisms identified, bacteria were predominant, followed by actinomycetes and fungi, which changed at similar trends to the microbial activity changes.

However, the increase of soil organic C content by cover crop addition may have adversely and significantly increased the GHGs emissions during rice cultivation. In particular, the hot water-extractable C and total C contents were significantly (P<0.05) and positively correlated with the seasonal CH₄ and CO ₂ fluxes, while the total N content was significantly (P<0.05) and positively correlated with the seasonal N₂O flux (Table 4). As a result, low C/N ratio vetch could be more effective in regulating the CH₄ and CO ₂ emissions than high C/N ratio rye. Although vetch addition largely increased the seasonal N₂O flux, its contribution to the total GWP increase was very minimal in a rice paddy soil condition.

Total GWP per grain yield

Total GWP per grain yield was approximately 2.8 Mg $CO_2 Mg^{-1}$ in the NPK treatment, which did not differ



Fig. 3 Grain yield and total GWP per grain yield for the different fertilization treatments during rice cultivation

Table 4 Correlation between the seasonal GHGs, and soil properties and plant growth characteristics at rice harvesting (n=9)

Parameters	Co-efficient (r)				
	CH ₄ flux	N ₂ O flux	CO ₂ flux		
Soil properties					
pН	0.208	-0.001	0.438		
Mean Eh value	-0.830**	0.011	-0.601		
Total C	0.930***	-0.316	0.591		
Total N	0.011	0.788**	0.324		
Available P ₂ O ₅	-0.020	0.379	0.076		
Exchangeable cations					
K^+	-0.056	-0.230	0.036		
Ca ²⁺	-0.242	0.151	-0.194		
Mg^{2+}	0.277	-0.513	0.281		
Hot water extractable C	0.965***	-0.174	0.743*		
PLFA profile					
Total phospholipid fatty acids	0.790**	-0.046	0.689*		
Total bacteria	0.896***	-0.222	0.743*		
Actinomycetes	0.883***	-0.171	0.707*		
Fungi	0.847**	-0.339	0.634*		
Plant growth & yield prope	erties				
Plant height	0.124	0.338	0.162		
Tiller number per hill	0.212	0.227	0.476		
Straw yield	0.638*	0.207	0.633*		
Grain yield	0.744*	0.311	0.731*		
Total biomass	0.700*	0.273	0.702*		
Root volume	0.168	0.570	0.304		
Root weight	0.266	0.552	0.164		

*, **, and *** denotes significance at the 5, 1, and 0.1 % levels, respectively

significantly from the 3.4 Mg $CO_2 Mg^{-1}$ in the NPK + vetch (Fig. 3). However, the rye addition significantly increased the total GWP per grain yield to 180 % compared with the NPK treatment, due to higher CH₄ and CO₂ emission and lower grain productivity than vetch addition.

Discussion

As shown in the NPK treatment, CH_4 was emitted at a comparably low rate during the initial rice growing stage, increased drastically with the development of soil reductive conditions and plant growth, and

recorded the peak emission rate at the reproductive stage (Neue and Roger 1993; Adhya et al. 1994; Chidthaisong et al. 1999). It is a well-known fact that CH₄ emitted from rice fields is transported mostly (60-90 %) through the aerenchyma of rice plants rather than by molecular diffusion across the waterair interfaces or the release of gas bubbles (Butterbach-Bahl et al. 1997; Aulakh et al. 2000). Since the apparent growth of rice plant are maximized at the reproductive stage, the well-developed aerenchyma might also provide an effective channel for CH₄ gas exchange between the atmosphere and the anaerobic soil (Nouchi et al. 1990; Butterbach-Bahl et al. 1997). In addition, the higher release of root exudates, which are good substrates for methanogenic archaea (Pusatjapong et al. 2003), increased CH₄ emissions at this stage (Aulakh et al. 2001). Methane emissions decreased after flowering because the rate of photosynthesis declined after the commencement of grain development and hence decreased the supply of available assimilates for CH₄ production (Sinha 1995).

However, the applications of the two cover crops drastically decreased the Eh values and increased CH₄ emission rates immediately after transplanting (Fig. 1). Within 40 days after transplanting, approximately 60 % of the total CH₄ was emitted in the vetch and rye applied plots, which was comparable with about 30 % in the NPK treatment plots. These results indicated high levels of CH₄ could be emitted by the direct diffusion-ebullition pathways from the initial rice growing stage (Kruger et al. 2002), irrespective of plant growth development. This was supported by the sudden reduction of soil Eh values (below -200 mV) right after the start of flooding. In our study, the amendment of organic matter like rye and vetch to a flooded rice field decreased the soil Eh more drastically compared with the NPK treatment (Fig. 1). The vetch and rye amendments provided C sources to methanogens, and thus increased the CH₄ production (Dubey 2005) which led to very high CH₄ emission rates.

In general, flooded rice paddies are not considered to be an important source of atmospheric N_2O (Granli and Bockman 1994). In this study, the N_2O emission was at very minimal values during the whole rice cultivation (Fig. 1). Nitrous oxide is formed by nitrifying and denitrifying bacteria in aerobic or upland soils and is enhanced by higher N availability (Gomes et al. 2009; Xiong et al. 2002), but under intensive anaerobic conditions like the paddy soil, N_2O is rapidly reduced to N₂ and then its emission becomes negligible (Granli and Bockman 1994). Comparatively high N₂O emission rates were observed at the panicle initiation stage, probably due to the effect of N side-dressing and soil temperature increase (Kurganova and Lopes de Gerenyu 2010). Incorporation of leguminous organic material creates a pool of readily available N and therefore stimulates N₂O emissions (Flessa and Beese 1995; Lemke et al. 1999). Slightly increased net N₂O emissions were observed in the vetch incorporated plots during rice cultivation, while the high C/N ratio of rye decreased the seasonal N₂O flux by 8 % compared with the NPK treatment (Figs. 1, 2). It is known that, with the addition of high C/N organic matter, microorganisms compete with the plants for available N in soil, which reduces N loss, similar to N leaching and volatilization (Kirschmann and Witter 1992). High C/N ratio cover crop cultivation and incorporation could be considered to reduce N₂O emissions from arable lands rather than low C/N ratio cover crops (Rosecrance et al. 2000).

The GWP can be used as an index to measure how much a given mass of a GHG contributes to global warming and as a basis to compare the effectiveness of each GHG to trap heat in the atmosphere relative to CO₂ (IPCC 2007; Zhang et al. 2008). To evaluate the effect of cover crop cultivation and incorporation as green manure to total GWP, GHGs emission should be characterized during a whole year including the period of cover crop and rice cultivations. Unfortunately, the GHGs emissions were not quantified during the cover crop cultivation in the fallow season in this study. Compared with the CH_4 , CO_2 , and N_2O emission levels during the flooded rice cultivation in the hot summer season under highly fertilized soil conditions, these GHGs emissions might be very small during the cover crop cultivation under upland conditions in a cold season. Cover crops like vetch and rye cultivations are not generally fertilized in Korea and are grown during colder seasons from November to April. Over 80 % of a whole year's GWP by CH₄ and N₂O emission was contributed during rice cultivation from a paddy field using a typical Japanese conventional water and fertilizer management system, in which the intermittent drainage system was introduced during rice cultivation and 6 Mg ha⁻¹ of rice straw was incorporated after rice harvest (Nishimura et al. 2004). In comparison, our field experiment was carried out in a Korean conventional rice cultivation system under continuous flooding without rice straw addition. This means that the contribution of our rice cropping season to the total GWP might be much higher than that of the Japanese study. Therefore, controlling GHGs emission during rice cultivation rather than the fallow season might be a key factor on regulating the total GWP impact in the rice paddy field.

Methane contributed very highly to the total GWP (79-81 %) during the entire rice cultivation period, followed by the CO_2 (14–17 %), while the contribution of N_2O was very low (2–5 %) (Fig. 2). Since the contribution of N₂O was very small irrespective of the treatment, the effective control of CH₄ and CO₂ emissions could be a useful countermeasure to reduce the impact of the total GHG emissions in rice paddy soil, suggesting that the cultivation and application of low C/N ratio cover crops, like vetch, could be more reasonable than high C/N ratio cover crops like rye, since low C/N ratio cover crops are more effective in minimizing CH₄ and CO₂ emissions. The vetch application increased the N2O emission during rice cultivation, but its contribution to the total GWP was very minimal.

Although both cover crop additions significantly increased rice grain yields and plant growth (Fig. 3; Table 2), leguminous vetch was more effective. This yield increase could be caused by the addition of available nutrients through biomass incorporation (Table 1) and soil fertility improvement (Table 2). The 25 Mg FW ha⁻¹ vetch application supplied around 120:28:92 kg ha⁻¹ of N:P₂O₅:K₂O, respectively, which was comparable with the 55:28:82 kg ha^{-1} by the rye application (29 Mg FW ha^{-1}). In Korea, 110:45:58 kg ha⁻¹ of N:P₂O₅:K₂O fertilization is recommended for rice cultivation (RDA 1995). Except for phosphate, the recycling of the two cover crop biomasses can satisfy the main nutrients fertilization requirements. However, vetch can supply more nutrients than rye, which could then lead to a higher increase in terms of rice productivity. Legumes green manure species such as vetch (Astragalus sinicus), sesbania (Sesbania rostrafa), and sola pith (Aeschynomene afraspero) supply large quantities of biologically fixed N to lowland rice cropping activities (Ladha et al. 1992) and improve soil productivity (Singh et al. 1991).

Soil chemical and biological properties were significantly improved by cover crop applications compared with those in the NPK treatment, but rye was more effective than vetch. Metabolic characteristics and diversity of soil microbial communities are known to be sensitive to management, and may also provide information on the status and activity of the microbial community (Marx et al. 2001). Total microbial activity, which was estimated by total PLFA concentration (Green and Scow 2000), was significantly improved with cover crop application, but rye was more effective than vetch (Table 2), probably due to higher soil organic C accumulation (Denef et al. 2009). Bacteria were the dominant microorganisms, followed by actinomycetes, and fungi (Bai et al. 2000), but the activities of these microorganisms changed similarly with the total microbial activity.

On the other hand, cover crop additions were effective in soil organic C accumulation, but adversely increased GHGs emissions. Since higher organic C accumulation can more significantly boost the methanogens activity and CH₄ production, the rye application may have stimulated higher CH₄ and CO₂ emissions than the vetch addition during rice cultivation. Methanogenic activity in paddy soil is determined mainly by the presence of exogenous substrates such as acetate or hydrogen supplied by hydrolytic and fermentative microbes in the ecosystem through the decomposition of organic matter (Zinder 1993; Conrad 1999). In particular, the concentration of hot water-extractable C, which is an important substrate for CH₄ production (Yavitt and Lang 1990), was significantly increased by the green manure amendments, but the increase was greater in the rye than the vetch treatment. The hot water-extractable C, being a component of the labile SOM (Ghani et al. 2003), is readily available to soil microbial biomass (Sparling et al. 1998), and rich in amorphous polysaccharides, which are similar to what originates from microbial and plant exudates (Feller et al. 1991) and is utilized by microbial communities for microbial processes like the methanogens. In comparison, total N concentration was significantly increased by the vetch addition, and consequently affected higher N₂O emissions in the vetch plots.

Total GWP per grain yield, which can be an indicator of the global warming impact from rice production (Li et al. 2006; Mosier et al. 2006; Qin et al. 2010), was similar in the NPK and NPK + vetch treatments, but it was increased more, to 172 %, in the NPK + rye-treated plots (Fig. 3). Our results indicated that the addition of a low C/N ratio leguminous green manure like vetch during rice cultivation could be more desirable than cover crops like rye, having a high C/N ratio, which have a greater potential to increase the total GWP per grain yield.

Conclusion

The addition of cover crops as green manure significantly increased the GHG emissions during rice cultivation. In particular, CH₄ emissions, which contributed 79-81 % to the total GWP, followed by the estimated CO_2 emissions (14–17 %), were markedly increased by the addition of high C/N ratio rye. Nitrous oxide emission was greatly increased by low C/N ratio vetch application, but its contribution to the total GWP was very small. The addition of green manure effectively improved rice productivity and soil quality. Vetch addition increased the total GWP to 163 % over the NPK control, but was increased more, to 221 %, with rye application. Total GWP per grain yield was similar between the NPK + vetch and NPK treatments, but increased to 171 % with rye addition. Conclusively, a low C/N ratio cover crop such as vetch may be a more recommendable green manure to minimize the total GWP per grain yield, and to improve rice productivity and soil quality in mono-rice cultivation systems.

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References

- Adhya TK, Rath AK, Gupta PK, Rao VR, Das SN, Parida KM, Parashar DC, Sethunathan S (1994) Methane emission from flooded rice fields under irrigated conditions. Biol Fertil Soils 18:245–248
- Aulakh MS, Khera TS, Doran JW (2000) Mineralization and denitrification in upland, nearly-saturated and flooded subtropical soil. II. Effect of organic varying in N contents and C:N ratio. Biol Fertil Soils 31:168–174
- Aulakh MS, Wassmann R, Bueno C, Kreuzwieser J, Rennenberg H (2001) Characterization of root exudates at different growth stages of ten rice (*Oryza sativa L.*) cultivars. Plant Biol 3:139–148

- Bai Q, Guttinger A, Zelles L (2000) Characterization of microbial consortium in paddy soil by phospholipid analysis. Microb Ecol 39:273–281
- Bouwman AF (1996) Direct emissions of nitrous oxide from agricultural soils. Nutr Cycle Agroecosyst 46:53–70
- Brown HA, Waggner-Riddle C, Thurtell GW (2000) Nitrous oxide flux from solid dry manure in storage as affected by water content and redox potential. J Environ Qual 29:630–638
- Burney JA, Davisc SJ, Lobell DB (2010) Greenhouse gas mitigation by agricultural intensification. Proc Natl Acad Sci USA 107:12052–12057
- Butterbach-Bahl K, Papen H, Rennenberg H (1997) Impact of gas transport through rice cultivars on methane emission from rice paddy fields. Plant Cell Environ 20:1175–1183
- Chidthaisong A, Obata H, Watanabe I (1999) Methane formation and substrate utilization in anaerobic rice soils as affected by fertilization. Soil Biol Biochem 31:135–143
- Cho JY, Han KW, Choi JK (2000) Balance of nitrogen and phosphorus in a paddy field of Central Korea. Soil Sci Plant Nutr 46(2):343–354
- Conrad R (1999) Contribution of hydrogen to methane production and control of hydrogen concetration in methanogenic soils and sediments. FEMS Microbiol Ecol 28:193–202
- Denef K, Roobroeck D, Mihiri CW, Lootens P, Boecks P (2009) Microbial community composition and rhizo depositcarbon assimilation in differently managed temperate grassland soils. Soil Biol Biochem 41:144–153
- RDA (Rural Development Administration, Korea) (1995) Standard Investigation Methods for Agriculture Experiment, Suwon
- Dubey SK (2005) Microbial ecology of methane emission in rice agroecosystem: a review. Appl Ecol Env Res 3(2):1–27
- Elfstrand S, Hedlund K, Martensson A (2007) Soil enzyme activities, microbial community composition and function after 47 years of continuous green manuring. Appl Soil Ecol 35:610–621
- Feller C, Francois C, Villemin G, Portal JM, Tontain F, Morel JL (1991) Nature des matieres organiques associees aux fractions argileuses d'un sol ferrallitique. CR Acad Sci Paris 312:1491–1497
- Flessa H, Beese F (1995) Effects of sugarbeet residues on soil redox potential and nitrous oxide emission. Soil Sci Soc Am J 59:1044–1051
- Frostegard A, Baath E, Tunlid A (1993) Shift in the structure of soil microbial communities in limed forests as revealed by phospholipid fatty acid analysis. Soil Biol Biochem 25:723–730
- Frostegard A, Tunlid A, Baath E (1996) Changes in microbial community structure during long-term incubation in two soils experimentally contaminated with metals. Soil Biol Biochem 28:55–63
- Garcia JL, Patel BCK, Ollivier B (2000) Taxonomic, Phylogenetic and ecological diversity of methanogenic archaeal. Anaerobe 6:205–226
- Ghani A, Dexter M, Sarathchandra U, Perrott KW, Singleon P (2000) Assessment of extractable hot-water carbon as an indicator of soil quality on soils under long-term pastoral cropping, market gardening and native vegetation. Proceedings of Australian and New Zealand Second Joint Soils Conference. Lincoln, New Zealand, pp 119–120

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- Ghani A, Dexter M, Perott KW (2003) Hot water-extractable carbon in soil: a sensitive measurement for determining impacts of fertilization, grazing, and cultivation. Soil Biol Biochem 35:1231–1243
- Godfray HCJ, Pretty J, Thomas SM, Warham EJ, Beddington JR (2011) Linking policy on climate and food. Science 331:1013–1014
- Gomes J, Bayer C, de Souza Costa F, de Cassia Piccolo M, Zanata JA, Vieira FCB, Six J (2009) Soil nitrous oxide emissions in a long-term cover crops-based rotations under subtropical climate. Soil Till Res 106:36–44
- Granli T, Bockman OC (1994) Nitrous oxide from agriculture. Norw J Agric Sci Suppl 12:7–128
- Green C, Scow K (2000) Analysis of phospholipid fatty acids (PLFA) to characterize microbial communities in aquifers. Hydrogeol J 8:126–141
- Han KW, Cho JY, Son JG (1998) Losses of chemical components by infiltration water during the rice cultivation at silt moam paddy soil. Korean J Environ Agric 17(3):268–273
- IPCC (2007) Technical summary. In: Solomon S, Qin D, Manning M, Chen Z, Marquis M, Averyt KB, Tignor M, Miller HL (eds) Climate change: the physical science basis. Contribution of Working Group 1 to the Forth Assessment report of the intergovernmental panel on climate change. Cambridge University Press, Cambridge
- Kar S, Ghildyal BP (1975) Rice root growth in relation to size, quantity and rigidity of pores. Plant Soil 43:627–637
- Kendall C, Silva S, Kelly V (2001) Carbon and nitrogen isotopic compositions of particulate organic matter in from large rive r systems across the United States. Hydrol Process 15:1301–1346
- Kim SY, Lee BJ, Kim JH, Oh SH, Hwang WH, Hwang DY, Ahn JW, Oh BJ, Ku YC (2007) The timing for incorporating Chinese milk vetch plant into soil for natural reseeding in the southern part of Korean peninsula. Korean J Crop Sci 52:127
- Kim GY, Ko BG, Jeong HC, Roh KA, Shim KM, Lee JT, Lee DB, Hong SY, Kwon SI (2009) Estimating carbon fixation of 14 crops in Korea. Korean J Environ Agric 42(6):460– 466 (*in Korean*)
- Kirschmann H, Witter E (1992) Composition of fresh aerobic and anaerobic from animal dung. Bioresour Technol 40:137–142
- Klose S, Acosta-Martinez V, Ajwa HA (2006) Microbial community composition and enzyme activities in a sandy loam soil after fumigation with methyl bromide or alternative biocides. Soil Biol Biochem 38:1243–1254
- Kreye C, Dittert K, Zheng X, Zhang X, Lin S, Tao H (2007) Fluxes of methane and nitrous oxide in water-saving rice production in north China. Nutr Cycl Agroecosyst 77:293–304
- Kruger M, Eller G, Conrad R, Frenzel P (2002) Seasonal variation in pathways of CH₄ production and CH₄ oxidation in rice fields determined by stable isotope and specific inhibitors. Glob Change Biol 8:265–280
- Kurganova I, Lopes de Gerenyu V (2010) Effect of the temperature and moisture on the N₂O emission from some arable soils. Eurasian Soil Sci 43(8):919–928
- Ladha J, Pareek RP, Beeker M (1992) Stem-nodulating legume *Rhizobium* symbiosis and its agronomic use in lowland rice. Adv Soil Sci 20:148–192
- Lal R (2002) Soil carbon dynamics in cropland and rangeland. Environ Pollut 116:353–362

- Lee JH, Hopmans JW, Rolston DE, Baer SG, Six J (2009) Determining soil carbon stock changes: Simple bulk density corrections fail. Agric Ecosyst Environ 134(3–4):251–256
- Lemke RL, Izaurralde RC, Nyborg M, Solberg ED (1999) Tillage and N source influence soil-emitted nitrous oxide in the Alberta Parkland region. Can J Soil Sci 79:15–24
- Li C, Salas W, DeAngelo B, Rose S (2006) Assessing alternative for mitigating net greenhouse gas emissions and increasing yields from rice production in China over the next twenty years. J Environ Qual 35:1554–1565
- Linguist B, van Groenigen KJ, Adviento-Borbe MA, Pittelkow C, van Kessel C (2012) An agronomic assessment of greenhouse gas emissions from major cereal crops. Glob Change Biol 18(1): 194-209
- Maggiotto SR, Webb JA, Waggner-Riddle C, Thurtell GW (2000) Nitrous and nitrogen oxide emissions from turfgrass receiving different forms of nitrogen fertilizer. J Environ Qual 29:621–630
- Marx MC, Wood M, Jarvis SC (2001) A microplate fluorimetric assay for the study of enzyme diversity in soils. Soil Biol Biochem 33:1633–1640
- McLean W (1930) The carbon-nitrogen ratio of soil organic matter. J Agric Sci 20:348–354
- Mosier AR, Halvorson AD, Reule CA, Liu XJ (2006) Net global warming potential and greenhouse gas intensity in irrigated cropping systems in Northeastern Colorado. J Environ Qual 35:1584–1598
- Na CS, Lee YH, Hong SH, Jang CS, Kang BH, Lee JK, Kim TH, Kim W (2007) Change of seed quality of Chinese milk vetch (*Astragalus sinicus* L.) during seed developmental stages. Korean J Crop Sci 52:363–369
- Naser HM, Nagata O, Tamura S (2007) Methane emissions from five paddy fields with different amounts of rice straw application in Central Hokkaido, Japan. Soil Sci Plant Nutr 53:95–101
- Neue HU, Roger PA (1993) Rice agriculture; factors affecting emissions. In: Khalil MAK (ed) Atmospheric methane: Sources sinks and role in global change. Springer, Berlin, pp 254–298
- Nishimura S, Sawamoto T, Akiyama H, Sudo S, Yagi K (2004) Methane and nitrous oxide emissions from a paddy field with Japanese conventional water management and fertilizer application. Glob Biogeochem Cy 18:GB2017
- Nishimura S, Akiyama H, Sudo S, Fumoto T, Cheng W, Yagi K (2011) Combined emission of CH_4 and N_2O from a paddy field was reduced by preceding upland crop cultivation. Soil Sci Plant Nutr 57:167–178
- Nouchi I, Mariko S, Aoki K (1990) Mechanism of methane transport from the rhizosphere to the atmosphere through rice plants. Plant Physiol 94:59–66
- Olsson PA (1999) Signature fatty acids provide tools for determination of the distribution and interactions of mycorrhizal fungi in soil. FEMS Microbiol Ecol 29:303–310
- Pusatjapong W, Kerdchoechuen O, Towprayoon S (2003) Glucose, fructose, and sucrose accumulation in root and root exudates of rice cv. Supandari 1. KMUTT Res Develop J 26(3):339–350
- Qin Y, Liu S, Guo Y, Liu Q, Zou J (2010) Methane and nitrous oxide emissions from organic and conventional rice cropping systems in Southeast China. Biol Fertil Soils 46:825– 834

- Rath AK, Swain B, Ramakrishnan B, Panda D, Adhya TK, Rao VR, Sethunathan N (1999) Influence of fertilizer management and water regime on methane emission from rice fields. Agric Ecosyst Environ 76:99–107
- RDA (Rural Development Administration, Korea) (1988) Methods of soil chemical analysis. National Institute of Agricultural Science and Technology, Suwon
- RDA (Rural Development Administration, Korea) (1999) Fertilization standard of crop plants. National Institute of Agricultural Science and Technology, Suwon
- Robertson GP, Paul E, Harwood R (2000) Greenhouse gases in intensive agriculture: Contributions of individual gases to the radiative forcing of the atmosphere. Science 289:1922– 1925
- Rolston DE (1986) Gas flux. In: Klute A (ed) Methods of soil analysis part 1. 2nd edn. Agron Monogr 9. ASA and SSSA, Madison, pp 1103–1119
- Rosecrance R, McCarty G, Shelton D, Teasdale J (2000) Denitrification and N mineralization from hairy vetch (*Vicia villosa* Roth) and rye (*Secale cereale* L.) cover crop monocultures and biocultures. Plant Soil 227:283–290
- SAS Institute (2003) System for Windows Release 9.1. SAS Institute, Cary
- Shang Q, Yang X, Gao C, Wu P, Liu J, Xu Y, Shen Q, Zou J, Guo S (2010) Net annual global warming potential and greenhouse gas intensity in Chinese double rice-cropping systems: a 3-year field measurement in long-term fertilizer experiments. Glob Change Biol 17(6):2196–2210
- Singh J, Khud CS, Singh B (1991) Efficient management of leguminous green manure in wetland rice. Adv Agron 45:135–189
- Singh S, Singh JS, Kashyap AK (1999) Methane flux from irrigated rice fields in relation to crop growth and Nfertilization. Soil Biol Biochem 31:1219–1228
- Sinha SK (1995) Global methane emission from rice paddies: excellent methodology but poor extrapolation. Curr Sci 68:643–646
- Sparling G, Vojvodic-Vukovic M, Schipper LA (1998) Hot water soluble C as a simple measure of labile organic matter: the relationship with microbial biomass C. Soil Biol Biochem 30:1469–1472
- Xiong Z, Zing G, Tsuruta H, Shen G, Shi S, Su L (2002) Field study on nitrous oxide emissions from upland cropping systems in China. Soil Sci Plant Nutr 48(4):539–546
- Yavitt JB, Lang GE (1990) Methane production in contrasting wetland sites: response to organo-chemical components of peat and to sulfate reduction. Geomicrobiol J 8:27–46

Zelles L, Bai QY (1993) Fractionation of fatty acids derived from soil lipids by solid phase extraction and their quantitative analysis by GC-MS. Soil Biol Biochem 25:495–507

- Zhang YS, Lee GJ, Joo JH, Lee JT, Ahn JH, Park CS (2007) Effect of winter rye cultivation to improve soil fertility and crop production in alpine upland in Korea. Korean J Environ Agric 26(4):300–305
- Zhang JE, Ouyang Y, Huang ZX (2008) Characterization of Nitrous Oxide Emission from Rice-Duck Farming System in South China. Arch Environ Contam Toxicol 54:167–172
- Zinder SH (1993) Physiological ecology of methanogens. In: Ferry JG (ed) Methanogens: Ecology, physiology, biochemistry and genetics. Chapman and Hall, New York, pp 128–206