9,10-ANTHRAQUINONE DITHIOCARBAMATES AS POTENTIAL PHARMACEUTICAL SUBSTANCES WITH PLEIOTROPIC ACTIONS: COMPUTERIZED PREDICTION OF BIOLOGICAL ACTIVITY AND EXPERIMENTAL VALIDATION

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The pleiotropic pharmacological actions of 9,10-anthraquinone dithiocarbamates were studied. The *Way2Drug* web portal was used to establish that cytotoxic, antiviral, and anticonvulsant actions were predicted for the study molecules, along with moderate toxicity. *In vitro* experimental studies showed that all study compounds had cytotoxic effects on tumor cell lines Hep-2, CHO, HeLa, Raji, BL, and B95-8. The high levels of cytotoxicity against cell lines Raji and B95-8 seen with derivatives **II**, **IV**, **VI**, and **X** was evidence of direct actions on Epstein-Barr virus (EBV). Along with the cytotoxic action, dithiocarbamates **V**, **VIII**, and **IX** showed antiviral effects against herpes virus HSV-2. Compounds **VI** and **X**, which had cytotoxic and antiviral actions, produced anticonvulsant effects 3 and 6 h after administration, while dithiocarbamate **III**, which had cytotoxic activity, retained its antiviral effect 24 h after administration. Dithiocarbamates with pleiotropic actions were moderately toxic $(LD_{50} > 1000 \text{ mg/ml}, \text{ mice}, \text{p.o.})$. Interactions between the structures and the three types of biological activity studied experimentally were assessed.

Keywords: 9,10-anthraquinone dithiocarbamates, computer prediction, *Way2Drug*, cytotoxic activity, antiviral activity, anticonvulsant activity.

Malignant neoplasms are the second most common cause of death throughout the world. Chemotherapy is widely used in the treatment of cancer and for various types of cancer, including leukemia, it remains the only effective method. Viral infections in patients with oncological disease lead to serious complications [1]. Furthermore, a number of oncoviruses, such as Epstein-Barr virus and herpes simplex virus (HSV), are associated with the occurrence and development of cancers $[2 - 4]$. Many patients with gliomas, meningiomas, and systemic oncological diseases develop convulsions, which generates the need for anticonvulsant therapy [5]. The combined prescription of multiple drugs (antitumor, antibacterial, antifungal, antiviral, antiemetic, anti-inflammatory, anticonvulsant, immunomodulator, etc.) with different mechanisms of action is widely used in the treatment of patients with malignant neoplasms. However, this treatment produces, along with positive effects, increases in the risk of unwanted interactions between drugs, resulting in increases in the number of side effects $[6 - 8]$ and/or therapeutic resistance $[9, 10]$. Use of pharmacological agents with pleiotropic actions prevents the development and decreases the consequences of infectious complications in the chemotherapy of malignant neoplasms $[11 - 13]$. As a result, the development of low-toxicity substances with pleiotropic actions, combining cytotoxic, antiviral, antibacterial, antifungal, and anticonvulsant effects in a single molecule, represents a potential direction in the search for and creation of novel chemotherapeutic drugs.

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Derivatives of 9,10-anthraquinone display various pharmacological effects – antitumor, antimicrobial, antiviral, antioxidant, antiprotozoal, antidiabetic, etc. $[14 - 17]$. The search for novel pharmaceutical substances based on compounds of this class is ongoing $[18 - 20]$. A number of successfully functionalized derivatives of 9,10-anthraquinone are currently in various stages of preclinical and clinical trials [21]. We have previously reported the synthesis of 9,10-anthraquinone dithiocarbamates and studies of their antioxidant and antiplatelet activities [22, 23]. We have also studied the cytotoxic actions of dithiocarbamates in human lung (A549), prostate (PC3), large intestine (HT29), and breast (MCF7) tumor cell lines using the SRB method [24].

We report here the results of computer prediction of the biological activities of 9,10-anthraquinone dithiocarbamates and experimental testing of the cytotoxic, antiviral, and anticonvulsant activities, as well as the acute toxicity of these compounds, and evidence was obtained of the potential of the search for pleiotropic pharmacological substances among compounds of this chemical class.

EXPERIMENTAL SECTION

Predictions of biological activity were made using the web services *PASS Online, CLC-Pred*, and *Acute Rat Toxicity* in the *Way2Drug* web portal [25 – 28]. Input data for obtaining prediction results were the structural formulas of substances presented as MOL files. MOL files were prepared using the program MarvinSketch (www.chemaxon.com). Prediction results were lists of probable types of biological activity with assessments of two probabilities - the presence of activity Pa and the absence of activity Pi.

Cytotoxic activity was studied using cell cultures included in the catalog of the European Collection of Cell Cultures (ECACC): Hep-2 (human throat epidermoid carcinoma cells), HeLa (human cervical cancer cells), Raji (human type B undifferentiated lymphoblasts from a Burkitt's lymphoma, which contain EBV), CHO cells (Chinese hamster ovary cells), BL (an experimental B-cell line), and B95-8 (EBV-transformed marmoset leukocytes which chronically produce EBV). Epithelial cell line were grown in sterile flasks (Sigma, USA) in medium containing 45% DMEM (Sigma, USA), 45% RPMI 1640 (Sigma, USA), and 10% heat-inactivated (56°C) fetal calf serum (Sigma, USA), and antibiotics – penicillin/streptomycin (0.1%). Cells were reseeded after forming monolayers. Cell dilutions were determined using a Goryaev counting chamber and a Lomo inverted microscope (Russia) with magnification $\times 70$. Cells were removed from flask surfaces with 0.02% Versene solution, suspended in nutrient medium, and their concentrations in suspension were then adjusted to 3×10^5 cells/ml. Lymphoblast cell lines were cultured in medium containing 90% RPMI 1640 (Sigma, USA), 10% heat-inactivated (56°C) embryonic calf serum (Sigma, USA), and 0.1% penicillin/streptomycin. Plates with cell cultures were incubated at 37° C in a 5% CO₂ atmosphere. Starting solutions of test substances $I - X$ were prepared in DMSO. Substance dilutions were prepared in growth medium. Solutions were filtered through syringe membrane filters (Sarstedt, Germany) with pore diameter 0.45 μ m and stored for use at +4 $\rm{°C}$. Studies used the MTT method [29]. Cells were grown in 96-well plates in medium. After incubation for 24 h, culture medium was replaced with medium containing dissolved substances at different concentrations $(200 \mu l \text{ per well})$. Two-fold serial dilutions of compounds were made in nutrient medium without fetal calf serum. Old medium was replaced with fresh in control samples (no substances). Platelets with cells were kept in an incubator at 37° C in a 5% CO₂ atmosphere for 72 h. Cell monolayers were monitored using a Lomo inverted microscope (Russia) (ocular $\times 7$, objective \times 10). MTT(sigma, USA) was dissolved in sterile phosphate buffer pH 7.2 at room temperature to a concentration of 5 mg/ml; 20 μ l of MTT solution was placed in each well of the 96-well plate and incubated for $2 - 4$ h at 37°C. Medium was removed after incubation and 150μ of 96% ethanol was added to dissolve formazan crystals. The optical densities of solution were determined spectrophotometrically using a Multiscan FC universal microplate reader (Thermo Scientific, USA) at 540 nm. Concentrations of compounds producing 50% inhibition of cell viability (CC_{50}) were compared with controls. Analysis of substance CC_{50} concentrations was by linear regression in Microsoft Excel. The cytotoxic actions of study compounds were compared with that of reference agent mitoxantrone.

Antiviral activity was studied in strains of human serotype 5 adenovirus (Ad h5), US herpes simplex virus type 1 (HSV-1/US), BH herpes simplex type 2 virus (HSV-2/BH). Ad h5 was obtained from the collection of the Institute of Microbiology, Budapest Medical University. HSV-1/US was obtained from the collection of the Institute of Antiviral Chemotherapy, Center for Clinical and Theoretical Medicine (Germany). HSV-2/BH was obtained from the collection of the Institute of Epidemiology and Infectious Diseases, Ukrainian Academy of Medical Sciences. Antiviral actions of substances were evaluated in terms of inhibition of the cytopathic effects of viruses using the spectrophotometric MTT method. Cells were infected with 40 µl of a previously determined dose of virus at 24 h of cell growth and formation of monolayers in 96-well plates. Virus was adsorbed at room temperature for $2 h$ in a 5% CO₂ atmosphere. Cells were washed to remove unadsorbed virus with Hank's balanced salt solution and $200 \mu l$ of two-fold dilutions of test substances was added to each well. Platelets with cells were incubated in an incubator at 37° C in a 5% CO₂ atmosphere, 20 μ l of MTT solution (5 mg/ml) were added to each well, and incubation was continued in a 5% CO₂ atmosphere at 37°C for 4 h. Medium was then removed from each well and $150 \mu l$ of DMSO was added to wash stain from cells. plates were read using a Multiscan FC automatic plate reader (Thermo Scientific, USA) at 540 nm. The optical density of each well was compared with the optical density of solvent as control. Percentage inhibition of virus multiplication was determined as

⁹sinhibition =
$$
\frac{(OD_{\text{es}}) - (OD_{\text{vc}})}{(OD_{\text{cs}}) - (OD_{\text{vc}})}
$$
·100%, (1)

where OD_{cs} is the mean optical density of control samples, *OD*_{vc} is the mean optical density of the virus control, and OD_{es} is the mean optical density of the experimental samples.

The extent of the antiviral activity of compounds was assessed in terms of EC_{50} – the concentration of compound producing 50% inhibition of virus replication in cultures of infected cells. The index of selectivity (IS) was used to identify compounds for further studies and was calculated using Equation (2). Compounds with $IS \geq 4$ were regarded as active, with potential for further *in vivo* study.

$$
SI = \frac{CC_{50}}{EC_{50}}.\t(2)
$$

The antiviral activities of test substances were compared with those of reference agents acyclovir (for HSV-1/US $EC_{50} = 7.7 \pm 0.9$ µg/ml, HSV-2/BH $EC_{50} = 1.75 \pm 0.33$) and ganciclovir (for Ad h5 $EC_{50} = 60 \pm 0.85$ µg/ml).

The anticonvulsant activities of compounds $I - X$ were studied in white male mice $(18 - 22 \text{ g})$. All animals were kept in conditions of a 12-h light regime and in a standard location for animals, with free access to water and food in compliance with the European Convention for the Protection of Vertebrates Used for Experimental and Other Specific Purposes (Strasbourg, 1986) and the Guidelines of the European Council for Studies on Biological Materials of Human Origin (2006). Animals were obtained from Odessa National Medical University (ONMU); the studies were approved by the ONMU Ethics Committee. The anticonvulsant activities of compounds were studied at a dose of 100 mg/kg and were compared with reference drug – valproic acid (VPA), used at a dose of 200 mg/kg. Mice received compounds p.o. in Tween 80/water emulsion and controls consisted of Tween 80/water emulsion. The anticonvulsant activities of compounds were determined using a corasol model, which includes determination of the minimum effective dose (MED) of corasol inducing clonic-tonic convulsions (CTC) and tonic extension (TE) in the experimental animals on i.v. infusion of 1% aqueous corasol solution into the tail vein. The corasol doses for induction of clonic-tonic convulsions (DCTC) and tonic extension (DTE) were calculated in relation to controls. The anticonvulsant effects of compounds were evaluated at specified time points after administration (3, 6, and 24 h) as compared with the control group. MED (%) was calculated as:

$$
MED(\%) = \frac{V}{m} \cdot 10^{4}, \tag{3}
$$

where MED is the minimum effective dose of corasol inducing DCTC or DTE, *V* is the volume of corasol solution, and *m* is the weight of the animal, g.

The acute toxicity of compounds was studied using a method analogous to that described in [30]. Acute toxicity (LD_{50}) in mice was determined by p.o. administration of compounds **I–X**. Mice were divided into groups of 10 animals in each and anthraquinone derivatives were given as single doses of 100, 250, 500, and 1000 mg/kg in Tween-80/water emulsion, while the control group received only Tween-80 in water. Observations for possible toxic symptoms in the animals were continued for 24 h after substance administration. The toxicity class for anthraquinone derivatives was determined using the OECD classification [31].

Data from all biological experiments were processed statistically using Student's *t* test; results were regarded as significant at $p < 0.05$.

RESULTS AND DISCUSSION

Prediction of Biological Activity

A rational approach to the search for novel biologically active substances with the necessary properties is based on computer prediction of biological activity on the basis of their structural formulas $[32 - 34]$. The computer program *PASS Online* [35] has now been used to predict more than 4000 types of biological activity [26]. The effectiveness of using *PASS Online* has been confirmed by many years of employment by 23,000 researchers from more than 100 countries around the world. Several hundred predictions of biological activity using this web service have been confirmed experimentally [36, 37]. The potential for computer prediction of biological activity suggested by this web service has significantly widened in recent years as a result of use of the *Way2Drug* platform [25]. The present study used in silico prediction to identify the most likely of the previously unknown types of biological activity of derivatives for the 9,10-anthraquinone dithiocarbamates **I–X** studied here (Fig. 1) [22].

Results obtained by computer evaluation of biological activity using *PASS Online* are given in Table 1. As shown from the data in this Table, all compounds are predicted to be neurotransmitter reuptake inhibitors, pointing to the likely presence of anticonvulsant activity, and most compounds were predicted also to have antiviral activity.

Cytotoxicity in relation to cancer cell lines was predicted using *PASS CLC-Pred* (Cell Line Cytotoxicity Predictor) [27]. Analysis of the data showed (Table 2) that the most likely were interactions between the study compounds with lung, lymphoid, breast gland tissue cell lines and for some

Fig. 1. Structural formulae of substances for which computer prediction and experimental testing of biological activities were performed.

compounds also uterine cervix, brain, and large intestine cells.

Relatively low probabilities Pa in the predictions (Tables 1 and 2) may be evidence for significant novelty in the study compounds as compared with substances for which information is present in the PASS training set.

Acute toxicity LD_{50} was predicted for rats via four different routes of administration $-$ i.p., i.v., p.o., and s.c. $-$ using the online resource *Acute Rat Toxicity* [28]. The results (Table 3) showed that the study dithiocarbamates could be classified as intermediate-toxicity, low-toxicity, or nontoxic compounds on the classification described in [31].

Thus, computer results point to the potential for studying the antitumor, antiviral, and anticonvulsant activities of dithiocarbamate derivatives of 9,10-anthraquinones **I–X**. Experimental confirmation of prediction results would identify pleiotropic compounds with sets of pharmacological actions.

Biological Activities of CompoundsI–X

Studies of the cytotoxic actions of compounds $I - X$ used available cell cultures – Hep-2, CHO, HeLa, Raji, BL, and B95-8, which provided for assessment of the cytotoxicity of dithiocarbamates $I - X$ in relation to epidermoid carcinoma, ovary, uterine cervix, and lymphoblastoid cells, effects on which were predicted by *PASS CLC-Pred.* This resulted in determination of the concentrations of study compounds (Table 4) suppressing cell viability by 50% (CC_{50}). Analysis of the data in Table 4 shows that the cytotoxic effects of dithiocarbamates $I - X$ on the test cell lines Hep-2, CHO, A549, HeLa, Raji, BL, and B85-8 confirmed the prediction data in all cases.

In particular, the study compounds displayed cytotoxicity against cultures of human throat carcinoma Hep-2 cells in the range $CC_{50} = 0.7 - 1 \mu M$; the most active was compound **IV**. Study dithiocarbamates **I–X** showed cytotoxic effects in cultures of CHO Chinese hamster ovary cells with $CC_{50} = 0.4 - 19.7 \mu M$; CHO cells were most sensitive to dithiocarbamate V. Cytotoxicity for compounds **I–X** in HeLa uterine cervix cell cultures gave $CC_{50} = 0.7 - 2.8 \mu M$, and compound V produced a cytotoxic effect at a lower concentration than the other test substances. Studies of the cytotoxicity of compounds **I–X** against cultures of undifferentiated human B-type lymphoblastoid cells from Burkitt's lymphoma (Raji cells) showed that CC_{50} values of dithiocarbamates were in the range CC_{50} < 0.001 to 0.2 μ M, while cultures of experimental cell line BL obtained from a patient with type B lymphoma had cytotoxicity from <0.001 to 0.5 µM. Compounds **IV** and **X** showed high cytotoxicity to Raji and BL cells with CC_{50} concentrations of <0.001 μ M, while dithiocarbamate **VI** had an analogous level of cytotoxic actions only in relation to the Raji cell population.

The effects of substituents and their positions in the anthraquinone ring on the level of cytotoxic action were also studied. In the case of Hep-2 cells, changes in the location of the substituent from position 1 (compounds $I - III$) to position 2 (derivatives $IV - VI$) of the anthraquinone ring led to an increase in the cytotoxic effect. Introduction of a chlorine atom at position 3 (compounds VII and VIII) increased CC_{50} as compared with dithiocarbamates **IV** and V; the only case in which there was a decrease in the concentration eliciting 50% inhibition of this cell population compared with dithiocarbamate **VI** was derivative **X**. The following patterns were identified in the actions of study compounds on CHO cells. Movement of the diethyl fragment to position 2 led to an increase in CC_{50} , while additional introduction of a chlorine atom had no significant effect on activity. The presence of a morpholine ring in position 2 of 9,10-anthraquinone increased the cytotoxic effect, though introduction of a chlorine atom in position 3 led to a sharp decrease. Substitution of the morpholine fragment (compound **VIII**) by a piperidine

Fig. 2. Anticonvulsant activity of anthraquinone derivatives 3 h after p.o. administration at a dose of 100 mg/kg (only compounds with positive results are shown). Values are given as mean \pm SEM, $n = 5$ mice. For all compounds, ** $p < 0.01$ compared with controls.

Fig. 3. Anticonvulsant activity of anthraquinone derivatives 6 h after p.o. administration at a dose of 100 mg/kg (only compounds with positive results are shown). Values are given as mean \pm SEM, $n = 5$ mice. For all compounds, ** $p < 0.01$ compared with controls.

fragment (compound IX) increased CC_{50} . Movement of the pyrrolidine ring to position 2 (compound **VI**) and introduction of an additional halogen atom (compound **X**) led to a significant decrease the parameter expressing cytotoxicity and, thus, an increase in cytotoxic activity. In the case of the actions of dithiocarbamates **I** – **VI** on HeLa cells, substituent positions had no significant effects on activity except for derivative V, where there was a reduction in CC_{50} . The presence of a chlorine atom in the molecules of compounds **VII** and **VIII** significantly decreased the cytotoxic action. Results obtained with Raji cells showed the following. Compound **IV**, which contained a diethyl residue in position 2, was more active in terms of cytotoxicity than compounds **I** and **VII**. The presence of a pyrrolidine fragment in position 2 and introduction of a halogen into the molecule led to signifi-

Fig. 4. Anticonvulsant activity of anthraquinone derivatives 24 h after p.o. administration at a dose of 100 mg/kg (only compounds with positive results are shown). Values are given as mean \pm SEM, $n = 5$ mice. For all compounds, ** $p < 0.01$ compared with controls.

cant decreases in CC_{50} . Analysis of data for BL cells showed that the presence of the diethyl fragment in positions 1 and 2 in compounds **I** and **IV** had no effect on the cytotoxic effect. The presence of a morpholine ring in compounds **V** and **VIII** led to increases in CC_{50} , while its replacement with a piperidine residue had virtually no effect on the cytotoxic action. Of interest were results obtained in B95-8 cells with compound **II**, which led to elimination of 50% of the cell population from a concentration as low as $0.01 \mu M$; this may be evidence of a direct antiviral action against Epstein-Barr virus (EBV).

Studies of the **antiviral activity** of dithiocarbamates **I–X** against strains of human adenovirus 5 serotype Ad h5 and herpes simplex types 1 (HSV-1/US) and 2 (HSV-2/BH) showed that study compounds had no antiadenovirus activ-

TABLE 1. Types of Activity Predicted for Compounds **I–X** by the PASS Program with Pa > 20%

	Predicted biological activity					
Compound	Antiviral activity	Anticonvulsant activity				
I	Antiviral (Herpes) 0.207 Antiviral (Adenovirus) 0.245	Neurotransmitter uptake inhibitor 0.567				
\mathbf{I}	Antiviral (Influenza) 0.236 Antiviral (Picornavirus) 0.305	Neurotransmitter uptake inhibitor 0.526				
Ш	Antiviral (Adenovirus) 0.231 Antiviral (Picornavirus) 0.290	Neurotransmitter uptake inhibitor 0.639				
IV	Antiviral (Adenovirus) 0.245 Antiviral (Herpes) 0.207 Antiviral $(Influenza)$ 0.258	Neurotransmitter uptake inhibitor 0.567				
V	Antiviral (Influenza) 0.306 Antiviral (Picornavirus) 0.305	Neurotransmitter uptake inhibitor 0.523				
VI	Antiviral (Adenovirus) 0.231 Antiviral (Influenza) 0.249 Antiviral (Picornavirus) 0.290	Neurotransmitter uptake inhibitor 0.639				
VII	NP	Neurotransmitter uptake inhibitor 0.495				
VIII	NP	Neurotransmitter uptake inhibitor 0.456				
IX	NP	Neurotransmitter uptake inhibitor 0.589				
X	NP	Neurotransmitter uptake inhibitor 0.568				

* NP (not predicted) - stated activity absent in prediction at Pas $>$ 20%.

ity. Experiments on antiherpes activity against herpesviruses HSV-1 showed that derivative **IX** had minor antiviral activity. Antiviral activity against type 2 herpesvirus HSV-2 was seen with compounds V, **VIII**, and **IX**, which had actions close to that of reference agent ganciclovir (EC_{50}) values were 75 ± 0.9 , 70 ± 1.2 , and 60 ± 0.8 ug/ml respectively). Compound **VIII** had the greatest potential: the index of selectivity was $IS = 7$. Studies were also run in Raji cells, which contain the Epstein-Barr virus genome in episomal form and, due to synthesis of certain oncolytic proteins, maintains the culture in the immortal state, providing a convenient model for studies of the latent form of EBV infection. Analysis of the values obtained identified compounds **IV**, **VI**, and **X**, which at extremely low concentrations $(< 0.5 \mu g/ml$) led to elimination of virus-associated cells, such that they can be regarded as having potential for further studies for antitumor therapy. It is important to note the fact that for compounds **IV** and **X**,

the effects seen here were confirmed in cell cultures obtained from a patient with B lymphoma.

As noted above, antiviral activity was predicted with low probability, which may be evidence that these compounds have significant novelty as compared with substances included in the PASS training set. Thus, comparison of prediction results with experimental data leads to the conclusion that prediction is supported in relation to antiviral activity in the study compounds without any differentiation of their actions on particular viruses (an antiadenovirus effect was predicted, but not supported on testing with the available adenovirus strains; compounds **V**, **VIII**, and **IX** showed activity against herpesvirus type 2, while substances **IV**, **VI**, and **X** showed activity against EBV, which was not predicted).

The anticonvulsant effects of anthraquinone derivatives **I–X** were studied using the corasol test, which is a widely used research model of generalized myoclonic convulsions

TABLE 2. Predicted Cytotoxicity (Pa Values) for Compounds **I–X** Against Various Cancer Cell Lines

		Cell line/tissue/organ							
Com- pound	Lung carcinoma/ Lung	Adult immunoblastic lymphoma/ Haematopoietic and lymphoid tissue	Breast carcinoma/ Breast	Cervical adenocarcinoma adenocarcinoma /Cervix	Lung μ_{Lung}	Non-small cell lung carci- noma/Lung	Colon carci- noma/Colon	Breast adenocarcinoma /Breast	Oligodendrogli- oma/Brain
$\mathbf I$	0.641 DMS-114	0.587 SR	0.375 T47D			0.233 NCI-H522 0.204 $HOP-18$			
\mathbf{I}				0.383 HeLa	0.329 $PC-9$	0.206 $HOP-18$	0.334 RKO	0.241 MDA-MB-361	0.281 Hs 683
Ш				0.392 HeLa	0.220 $PC-9$	0.243 $HOP-18$	0.316 RKO	0.309 MDA-MB-361	0.316 Hs 683
IV	0.641 DMS-114	0.627 SR	0.447 T47D	$\overline{}$	$\overline{}$	0.334 NCI-H522 0.204 $HOP-18$			
\bf{V}				0.493 HeLa	0.329 $PC-9$	0.209 NCI-H522 0.206 $HOP-18$		0.241 MDA-MB-361	0.281 Hs 683
VI			0.207 T47D	0.510 HeLa		0.243 $HOP-18$			0.316 Hs 683
VII	0.562 DMS-114	0.604 SR	0.383 T47D	$\overline{}$	$\overline{}$	0.327 NCI-H522			0.267 Hs 683
VIII	0.221 DMS-114	$\overline{}$	$\qquad \qquad -$	0.448 HeLa	0.307 $PC-9$		0.281 RKO	0.247 MDA-MB-361	0.288 Hs 683
IX				0.446 HeLa	$\overline{}$	0.227 $HOP-18$			0.295 Hs 683
\mathbf{X}				0.460 HeLa					0.324 Hs 683

"-" indicates absence of predicted activity at Pa $>$ 30%.

in humans [38]. The dynamics of this action were studied for a dose of 100 mg/kg at early (3 and 6 h) and late (24 h) time points after single p.o. doses. These results showed that only five compounds (**I**, **III**, **VI**, **IX**, and **X**) had anticonvulsant effects throughout the *in vivo* test period. As shown in Figs. 2 and 3, studies of the anticonvulsant actions of compounds gave mean values of DCTC and DTE of 145% and 144% at 3 h post-administration and 168% and 162% at 6 h post-administration as compared with animals of the control group (100%). Thus, anthraquinone derivatives **I**, **III**, **VI**, **IX**, and **X** showed minor antiviral effects at the early time points. It should be noted that the antiviral action of compound **III** at 3 and 6 h after single p.o. doses were analogous to that of the reference agent valproic acid (200 mg/kg). The data indicate that administration of study compounds 24 h before i.v. corasol produced anticonvulsant activity, with mean DCTC and DTE of 145% and 143% respectively, which is evidence of a minor prolongation of the effect in these derivatives (Fig. 4). The effects of all compounds were less than that of valproic acid, which had DCTC and DTE values of 238% and 236% respectively. Analysis of the structure-activity interaction showed that substitution of the diethyldithiocarbamate fragment by a pyrrolidinedithiocarbamate fragment in position 1 of 9,10-anthraquinone led to an increase in anticonvulsant activity. At the same time, movement of the latter to position 2 (compound **VI**) and the presence of a chlorine atom in position 3 (compound **X**) decreased the anticonvulsant effect. Substitution of the pyrrolidine residue by a piperidine residue had no effect on the anticonvulsant properties.

Comparison of results on prediction of anticonvulsant activity with the experimental results showed that anticonvulsant activity was confirmed experimentally for compounds **I**, **III**, **VI**, **IX**, and **X**, which had the highest Pa values.

Studies of **acute toxicity** showed that no deaths or clear clinical signs occurred in animals of the study groups receiving compounds $I - X$. The LD_{50} values of all anthraquinone derivatives $I - X$ were greater than 1000 mg/kg, which, in terms of the standard classification, allows all compounds to be classified as toxicity class **III**, i.e., moderately toxic. It should be noted that the experimental data on acute p.o. toxicity correlated completely with results obtained by prediction using the online resource *Acute Rat Toxicity* (Table 3).

The results obtained from the present studies using in silico, *in vitro*, and *in vivo* methods identified that the moderately toxic 9,10-anthraquinone dithiocarbamate derivatives **III**, **IV**, **VI**, **IX**, and X are potential pharmacological substances with pleiotropic actions whose selectivities in relation to particular cell lines and different viruses are due to the features of their chemical structures, determining differences in the mechanisms of the pleiotropic actions. These studies established that the presence of cytotoxic, antiviral, and anticonvulsant actions and acute toxicity predicted using *PASS Online*, *CLC-Pred*, and *Acute Rat Toxicity* (Tables 1 – 3) on the *Way2Drug* platform were confirmed by *in vitro* and *in vivo* experimental results. A search based on structural similarity in the information system containing data on more than half a million actively studied pharmaceutical substances, *Clarivate Analytics Integrity* [39], showed that even using a threshold for the Tanimoto coefficient of similarity $TC > 70\%$ did not identify any analogs of the test compounds. As previous studies [40] have shown that at $TC > 85\%$ there is only a 30% probability that structural analogs will have the same biological activity, this result supports the novelty of the 9,10-anthraquinone dithiocarbamates studied here as noted above.

The test compounds were found to have marked cytotoxic activity against tumor cell lines Hep-2, HeLa, Raji, CHO, BL, and B95-8, while compounds **II**, **IV**, **VI**, and **X** included particularly promising compounds, as they displayed

TABLE 3. Assessment of Acute Toxicity in Rats (LD₅₀, mg) using the Program *Acute Rat Toxicity* for Compounds $I - X (LD₅₀)$ and Toxicity Class)

	LD_{50} , mg/kg / toxicity class					
Compound	i.p.	i.v.	p.o.	S.C.		
I	480/4	111/4	1495/4	976/4		
\mathbf{I}	928/5	194/4	1177/4	1343/5		
Ш	717/5	146/4	755/4	661/4		
IV	538/5	113/4	1375/4	1230/4		
V	1165/5	175/4	1056/4	1366/5		
VI	867/5	120/4	644/4	1222/5		
VII	653/5	76/4	890/4	1247/5		
VIII	1120/5	156/4	1060/4	2593/nontoxic		
IX	766/5	88/4	855/4	612/4		
X	808/5	106/4	651/4	720/4		

JU.	$\tilde{}$	\mathbf{r}					
	Concentration inducing 50% inhibition of cell viability, μM						
Compound	Hep-2	CHO	HeLa	Raji	BL	$B95 - 8$	
I	0.9	0.5	1.0	0.1	< 0.001	0.08	
П	0.9	0.7	1.2	0.2	0.3	0.01	
Ш	1.2	1.2	0.9	0.1	0.1	0.04	
IV	0.7	1.2	1.1	< 0.001	${}< 0.001$	0.09	
V	0.8	0.4	0.7	0.4	0.4	0.1	
VI	1.0	1.0	1.0	${}< 0.001$	0.2	0.09	
VII	0.9	1.0	0.9	0.1	0.03	0.1	
VIII	1.2	19.7	2.8	0.04	0.5	0.08	
IX	0.9	0.9	1.4	0.1	0.5	0.1	
X	0.8	0.5	2.6	< 0.001	${}< 0.001$	0.08	
Mitoxantrone	0.8	ND	10	83	0.9	12.3	

TABLE 4. CC_{50} Values for Study Compounds in Cell Lines

* ND (not determined) – absence of effects on cell lines;

** $p < 0.01$ for all compounds compared with controls.

cytotoxic effects against EBV-containing tumor cells Raji and B95-8. Dithiocarbamate **VIII** had activity against herpesvirus type 2 HSV-2 with $EC_{50} = 70 \text{ µg/ml with } IS = 7$. Studies of anticonvulsant activity using the corasol test showed that dithiocarbamate **III** had the greatest anticonvulsant activity in the early (3 and 6 h) and late (24 h) time points as compared with compounds **I**, **VI**, **IX**, and **X**. Assessment of acute toxicity in mice given p.o. substances showed that test compounds were moderately toxic $(LD₅₀ > 1000 \text{ mg/kg})$. The effects of different substituents on the types of biological activity studied here were investigated, indicating the potential for further targeted optimization of the structure and properties of substances of this class.

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REFERENCES

- 1. V. T. DeVita, T. S. Lawrence, and S. A. Rosenberg, *Cancer: Principles and Practice of Oncology*, Wolters Kluwer Lippincott Williams & Wilkins, Philadelphia (2011).
- 2. P. De Paoli and A. Carbone, *Int. J. Cancer*, **133**(7), 1517 1529 (2013).
- 3. J. L. Kutok and F. Wang, *Annu. Rev Pathol.*, No. 1, 375 404 (2006).
- 4. J. Jalouli, M. M. Jalouli, D. Sapkota, et al., *Anticancer Res.*, **32**(2), 571 – 580 (2012).
- 5. C. P. Benit and CJ. Vecht, *Neuro-Oncology Practice*, **3**, No. 4, $245 - 260(2016)$.
- 6. H. Brötz-Oesterhelt and N. A. Brunner, *Curr. Opin. Pharmacol.*, **8**(5), 564 – 573 (2008).
- 7. A. L. Hopkins, G. R. Bickerton, I. M. Carruthers, et al., *Cur. Top. Med. Chem.*, **11**(10), 1292 – 1300 (2011).
- 8. J. U. Peters, *J. Med. Chem.*, **56**(22), 8955 8971 (2013).
- 9. G. Housman, S. Byler, S. Heerboth, et al., *Cancers,* **6**, 3, 1769 – 1792 (2014).
- 10. H. Zahreddine and K. L. Borden, *Front. Pharmacol.*, **4**, 28 (2013).
- 11. B. Deslouches and Y. P. Di, *Oncotarget*, **8**(28), 46635 46651 (2017).
- 12. V. W. C. Soo, B. W. Kwan, H. Quezada, et al. *Curr. Top. Med. Chem.*, **17**(10), 1157 – 1176 (2017).
- 13. M. Micozzi, *Fundamentals of Complementary, Alternative, and Integrative Medicine*, Elsevier, Riverport Lane (2018).
- 14. A. Q. Acton, *Anthraquinones Advances in Research and Application*, Scholarly Editions, Atlanta (2013).
- 15. V. I. Zvarich, M. V. Stasevich, O. V. Stan'ko, et al., *Pharm. Chem. J.*, **48**(9), 584 – 588 (2014).
- 16. M. Stasevych, V. Zvarych, V. Lunin, et al., *Indian J. Pharm. Sci.*, **77**(5), 634 – 637 (2015).
- 17. M. Stasevych, V. Zvarych, V. Lunin, et al., *Monatsh. Chem.*, **149**(6), 1111 – 1119 (2018).
- 18. E. M. Malik and C. E. Müller, *Med. Res. Rev.*, **36**(4), 705 748 (2016).
- 19. L. M. L. Nollet and J. A. Gutierrez-Uribe, *Phenolic Compounds in Food. Characterization and Analysis*, CRC Press, Boca Raton (2018).
- 20. A. S. Tikhomirov, A. A. Shtil, and A. E. Shchekotikhin, *Recent Pat. Anti-Cancer Drug Discov.*, **13**, 159 – 183 (2018).
- 21. Clarivate Analytics Integrity, 2018; http: // integrity. Thomson-Pharma.com (accessed June 23, 2018).
- 22. V. Zvarych, M. Stasevych, V. Lunin, et al., *Monatsh. Chem.*, **147**(12), 2093 – 2101 (2016).
- 23. T. I. Halenova, I. V. Nikolaeva, M. V. Stasevych, et al., *Res. J. Pharm. Biol. Chem. Sci.*, **8**, 1626 – 1632 (2017).
- 24. M. Stasevych, V. Zvarych, V. Lunin., et al., *SAR QSAR Environ. Res*., **28**(5), 355 – 366 (2017).
- 25. D. S. Druzhilovskiy, A. V. Rudik, D. A. Filimonov, et al., *Rus. Chem. Bull.,* **66**(10), 1832 – 1841 (2017).

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- 26. D. A. Filimonov, D. S. Druzhilovskiy, A. A. Lagunin, et al., *Biomed. Chem. Res. Meth.*, **1**, e00004 (2018).
- 27. A. A. Lagunin, V. I. Dubovskaja, A. V. Rudik, et al., *PLOS One*, **13**, e0191838 (2018).
- 28. A. Lagunin, A. Zakharov, D. Filimonov, et al., *Mol. Inf.,* $30(2-3)$, $241-250$ (2011).
- 29. T. Mosmann, *Immunol. J. Meth.*, **65**(1 2), 55 63 (1983).
- 30. M. Ahmed, *Pharmacol. Pharmacy*, **6**, 185 189 (2015).
- 31. J. Švarc-Gajić, *General Toxicology*, Nova Science Publishers Inc., New York (2010).
- 32. S. M. Ivanov, A. A. Lagunin D. A. Filimonov, et al., *Pharm. Chem. J.*, **52**(8), 758 – 762 (2018).
- 33. R. K. P. Tripathi and S. R. Ayyannan, *Med. Chem. Res.*, **27**(3), $709 - 725$ (2018).
- 34. V. V. Knyazev, V. S. Rogovskii, E. D. Sveshnikova, et al., *Pharm. Chem. J.*, **52**(3), 205 – 208 (2018).
- 35. A. Lagunin, A. Stepanchikova, D. Filimonov, et al., *Bioinformatics*, **16**(8), 747 – 748 (2000).
- 36. *PASS Online*, 2019; http: // www.way2drug.com / PASSOnline / index.php (accessed July 15, 2019).
- 37. K. Anusevicius, V. Mickevicius, M. Stasevych, et al., *Res. Chem. Intermed.,* **41**(10), 7517 – 7540 (2015).
- 38. M. V. Nesterkina and I. A. Kravchenko, *Chem. Nat. Compd.,* **52**(2), 237 – 239 (2016).
- 39. *Clarivate Analytics Integrity* (2019); https: // integrity.clarivate.com /(accessed July 15, 2019).
- 40. Y. C. Martin, J. L. Kofron, and L. M. Traphagen, *J. Med. Chem.*, **45**(19), 4350 – 4358 (2002).