REVIEW

Ischemia Reperfusion Injury Induced Blood Brain Barrier Dysfunction and the Involved Molecular Mechanism

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Abstract

Stroke is characterized by the abrupt failure of blood flow to a specific brain region, resulting in insufficient supply of oxygen and glucose to the ischemic tissues. Timely reperfusion of blood fow can rescue dying tissue but can also lead to secondary damage to both the infarcted tissues and the blood–brain barrier, known as ischemia/reperfusion injury. Both primary and secondary damage result in biphasic opening of the blood–brain barrier, leading to blood–brain barrier dysfunction and vasogenic edema. Importantly, blood–brain barrier dysfunction, infammation, and microglial activation are critical factors that worsen stroke outcomes. Activated microglia secrete numerous cytokines, chemokines, and infammatory factors during neuroinfammation, contributing to the second opening of the blood–brain barrier and worsening the outcome of ischemic stroke. TNF-α, IL-1β, IL-6, and other microglia-derived molecules have been shown to be involved in the breakdown of blood–brain barrier. Additionally, other non-microglia-derived molecules such as RNA, HSPs, and transporter proteins also participate in the blood–brain barrier breakdown process after ischemic stroke, either in the primary damage stage directly infuencing tight junction proteins and endothelial cells, or in the secondary damage stage participating in the following neuroinfammation. This review summarizes the cellular and molecular components of the blood–brain barrier and concludes the association of microglia-derived and non-microglia-derived molecules with blood–brain barrier dysfunction and its underlying mechanisms.

Keywords Ischemic stroke · Blood–brain barrier · Microglia · Cytokines · Transporters · Ion channels

Introduction

According to the diferent etiology, there are two types of stroke: hemorrhagic stroke and ischemic stroke. Hemorrhagic stroke is characterized by the rupture of blood vessels in the brain, leading to bleeding into the brain. However, the pathophysiology of ischemic stroke features an abrupt failure

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of blood fow supply to a specifc brain region, which leads to insufficient oxygen and glucose in the ischemic brain tissue [[1\]](#page-10-0). Notably, ischemic stroke brings about magnifcent changes in the morphology of cells in the brain. After the organelles' swell and the cell contents' leak into the extracellular space, necrosis occurs, eventually resulting in the loss of neuronal function. Additionally, lactic acid production increases due to the transition to anaerobic metabolism, leading to tissue acidosis. Consequently, excess tissue acidosis and insufficient ATP production cause membrane transporter dysfunction and also compromise cellular Ca^{2+} buffering, which then initiates the cascade of phosphorylation and neuronal damage [\[2](#page-10-1), [3\]](#page-10-2). Importantly, blood–brain barrier dysfunction, infammation, and activation of glial cells, among other pathophysiologies accompanying ischemic stroke, are critical factors that worsen the stroke outcome.

The blood–brain barrier (BBB) is a hallmark of the mature central nervous system (CNS), which functions as a difusive barrier selectively ruling out most blood-borne substances from getting into the brain. The BBB is formed by an endothelial layer wrapped with pericytes (PCs), which share the same basement membrane with endothelial cells (ECs), and the astrocyte (AC) end-feet continuously surround this layer. Likewise, neurons form innervation on the surface with the AC end-feet [[4\]](#page-10-3). Simultaneously, these cells are components of the neurovascular unit (NVU) as well. As an integral part of BBB, tight junctions (TJs) present between the cerebral ECs and contribute to the difusive function of the BBB. In the setting of ischemic stroke, insufficient oxygen and glucose signifcantly infuence BBB function, altering its TJs expression and distribution, the morphology of its cellular parts, transporter functions, and so on. Most of the TJ proteins get degraded while the expression of transporters may increase or decrease depending on their location in the infarcted tissues. This BBB breakdown can lead to edema, ionic homeostasis disruption, and immune infltration [[5\]](#page-10-4). Importantly, the neuroinfammation following ischemic stroke contributes to the biphasic opening of the BBB, during which cytokines and infammatory factors secreted by glial cells and neurons play a key role. In the present review, we will discuss the cellular and molecular dysfunction of the BBB after ischemia and its underlying mechanisms. Furthermore, we will discuss the implications of these mechanisms which give insight into clinical diseases and treatment.

Basic Components of BBB and Their Dysfunction Following Ischemia/ Reperfusion Injury

The components of the BBB involve unique cells, such as AC end-feet, PCs, ECs, and neuronal innervation. ECs contact with PCs by "peg-and-socket" junctions in a shared basal lamina with AC end-feet that form the ensheathment of brain capillaries [[6](#page-10-5)], and the main components of BBB are shown in Fig. [1.](#page-1-0) Additionally, interneurons and perivascular microglia contact with ECs, PCs, and ACs to form the construction of the NVU [[7\]](#page-10-6). Rather than looking at each cell type separately, these cells interact intimately and reciprocally. Furthermore, TJs seal adjacent endothelial cells along with ACs and PCs to form a selective barrier that regulates solute movement, maintaining normal BBB function.

Ischemia can rapidly elicit cerebral edema, developing stepwise from cytotoxic edema to vasogenic edema [[8\]](#page-10-7). This is partly due to the BBB's increased permeability, resulting from cellular components' dysfunction and disruption of TJ proteins. And the degradation of TJ proteins is the hallmark of BBB dysfunction. Here we concluded the changes in diferent components of the BBB caused by ischemia/ reperfusion(I/R) injury and their fnal infuence on BBB.

Fig. 1 The components of BBB. The constitution of the BBB involves astrocytes end-feet, pericytes, and endothelial cells. These cells interact intimately and reciprocally with each other and together maintain the integrity of the BBB

Endothelial Cell and Tight Junction Proteins

Endothelial cells (ECs), composing the BBB, are equipped with specialized TJ structures, shaping a selective barrier that effluxes potential toxins and delivers specific nutrients into the brain [[9\]](#page-10-8). Subtle changes can result in the signifcant alteration in BBB permeability. Although more attention has been paid to TJ protein degradation during BBB dysfunction induced by ischemic stroke, an increasing focus has shifted to the dysfunction of the cerebral endothelium directly [\[10](#page-10-9)]. Consequently, ischemic stroke impairs normal endothelial morphology and function. As investigated, the vessels in the ischemic penumbra showed signs of swelling and edema of the endothelium $[11]$ $[11]$ $[11]$, and vasodilation and vasoconstriction function gets compromised $[12, 13]$ $[12, 13]$ $[12, 13]$, which ruins the restoration of cerebral blood fow. Furthermore, I/R injury also elicits endothelial apoptosis and necroptosis [\[14,](#page-10-13) [15](#page-10-14)], leading to the impaired function of the BBB, thereby limiting substrates to pass the selective barrier.

In the CNS, TJs are part of the ECs, which confer ECs to their CNS phenotype. TJs are critical to sustaining functional BBB integrity, enabling the BBB to restrict the free flow of water and solutes. Three integral proteins form TJs on the cell membrane–claudin, occludin, and JAM, and many cytoplasmic accessory proteins including ZO-1,

Fig. 2 The organization of tight junction proteins (TJs). The primary constituents of an integral TJ, including proteins on the membrane—claudin, occludin, and junction adhesion molecules (JAM), and accessory proteins in the cytoplasm— ZO-1, ZO-2, ZO-3, cingulin, etc.

ZO-2, ZO-3, cingulin, and others [\[16\]](#page-10-15), and the main components of TJs and their arrangement are shown in Fig. [2.](#page-2-0) The zonula occludens proteins (ZO-1, ZO-2, and ZO-3) are members of the MAGUK family [\[17](#page-10-16)]. Adherens junctions (AJs) recruit ZO-1 and ZO-2 to nascent adhesion sites to accomplish their formation [[18\]](#page-10-17). Additionally, the assembly of TJ proteins induces claudin polymerization and conjugation of an incessant TJ belt [\[19](#page-10-18)]. Claudin proteins are the most abundant transmembrane proteins of the TJs and play a determining role in paracellular ion conductance [\[20](#page-10-19)]. By regulating the degradation and distribution of Claudin 5, autophagy could mitigate the hypoxic injury to the BBB induced by ischemic stroke [\[21\]](#page-10-20). Claudin proteins are also substrates of diverse kinases, including protein kinase C (PKC), protein kinase A (PKA), and mitogen-activated protein kinase (MAPK), so they serve as a modulator to regulate TJs [[22\]](#page-10-21). Moreover, Occludin interacts with many other TJassociated proteins. Notably, ZO-1 and ZO-2 were initially found to directly interplay with occluding [\[23](#page-10-22)]. Likewise, VE-cadherin is one of the adherens junction proteins regulating adhesion between ECs. In the CNS, VE-cadherin is exclusive to ECs, assisting them to interact with other same type cells. Therefore, VE-cadherin plays an important role in the maintenance of the BBB [[24\]](#page-10-23). Evidence demonstrates that recycling endocytosed VE-cadherin increased the

integrity of the vascular system following BBB dysfunction [\[25](#page-10-24)]. Ischemic injury arouses various forms of pathological dysfunction in the brain and its insult in the BBB is mainly due to the degradation, disintegration, and redistribution of the TJ proteins, such as VE-cadherin, occludin and ZO-1 [\[26–](#page-10-25)[28](#page-10-26)]. TJs breaking down compromises BBB integrity, and its permselectivity becomes damaged as a result of this.

Pericytes

Pericytes (PCs) are the NVU's perivascular cells inserted into the basement membrane (BM), surrounding ECs and acting as a roundabout target for small molecules to regulate BBB function [[29\]](#page-10-27). Its contractile ability contributes to the normal function of the cerebral vasculature. Pericytes enhance angiogenesis in leptomeninges and infarct brain regions, contributing to the recovery of cerebral blood fow in the affected areas thus improving tissue repair $[30]$ $[30]$. By trapping blood cells, pericytes could inhibit the occluded vessels from reperfusion after ischemic stroke, thus functioning as a potential therapeutic target [\[31](#page-11-1)]. Previous studies have shown that pericytes are vulnerable to ischemic injury, and they could die a few minutes after insufficient blood flow and therefore constrict the microcirculation [\[32](#page-11-2)]. In the setting of chronic cerebral hypoperfusion, neurotoxic molecules pass through the BBB via endothelial transcytosis, which leads to the loss of pericytes and ultimately, the dysfunction of the BBB [[33\]](#page-11-3).

Astrocyte

Astrocytes (ACs) exist around cerebral microvessels and mediate BBB function via AC-derived factors and astrocytic terminal processes (end-feet) [[34](#page-11-4)]. Under ischemic conditions, the number of activated astrocytes increased, most of which are not perivascular astrocytes, indicating that the activated astrocytes mainly got involved in neurological infammation instead of restoring and regulating the impaired vasculature. Additionally, ischemic insult activates perivascular astrocytes, develops swollen end-feet, and retracts processes from blood vessels, leading to the opening of gap junctions and disassembly of TJ proteins [\[35\]](#page-11-5). Further studies showed that under OGD/R condition (oxygen glucose deprivation/reperfusion, the I/R condition for cultured cells), the level of apoptosis of astrocytes increases and neurological infammation is elicited [\[36](#page-11-6)]. Notably, the crosstalk between ECs and perivascular ACs is critical to maintaining the function of the selective barrier and its completeness [\[37\]](#page-11-7). Furthermore, the changes in reactivated perivascular astrocytes insulted by ischemic injury ruined the astrocyteendothelium interaction, leading to morphological alterations in BBB and its dysfunction [[38\]](#page-11-8).

Microglia

Microglia are the resident immune cells in the CNS, whose role in maintaining and regulating the structural and functional completeness of the BBB is yet to be elucidated [\[39](#page-11-9)]. In the setting of ischemic stroke, microglia are activated, which initiates following neuroinfammation. As a part of the NVU, microglia can secrete numerous infammatory factors, such as TNF- α , IL-1 and IL-6, to affect the permeability of the BBB [\[40](#page-11-10)]. Microglia could also phagocytize cell debris to suppress the infammatory response, consequently assisting recovery of the BBB. The dual function of microglia relates to its diferent phenotypes which are M1 (proinfammatory type) and M2 (anti-infammatory type) [\[41](#page-11-11)]. Notably, activated microglia contribute to BBB leakage via the downregulation of endothelial junctional proteins such as claudin-5 and occluding [[42](#page-11-12)]. Furthermore, M1 microglia secrete TNF- α to elicit necroptosis in endothelial cells, resulting in compromised BBB integrity and function during the pathological condition of I/R [[43\]](#page-11-13). Simultaneously, microglia also polarizes into the M2 phenotype in acute ischemic stroke, limiting poststroke infammation progression in the early phase of ischemic stroke [[44\]](#page-11-14).

Neuron

As a part of the neurovascular unit, neurons interact with ECs [\[45](#page-11-15)] and their activities impact cerebral blood flow [[46](#page-11-16)]. Studies show that experience-dependent neuronal activity alterations could infuence endothelial function [\[47](#page-11-17)]. Additionally, neuronal activity mediates brain endothelial gene expression and is inversely correlated with the expression of BBB ABC (ATP-binding cassette) efflux transporters in the BBB [[46](#page-11-16)]. Following I/R injury, transient autophagy activation in neurons occurs, leading to lysosomal dysfunction during the later phase and contributing to the compromised synaptic plasticity in the reperfusion phase [\[48\]](#page-11-18). Moreso dysfunction of the BBB exacerbates neuronal injury as CLEC14A defciency increases leakage in the endothelial barrier, which leads to larger infarct areas of afected brain tissue and severe neurological defects following I/R injury [[49\]](#page-11-19) (The changes in cells following I/R injury are summerized in Fig. [3](#page-4-0)).

Mechanisms Underlying BBB Dysfunction Following I/R Injury

Microglia‑Derived Molecules Involved in BBB Breakdown

During ischemia stroke, high levels of cytokines and chemokines produced by microglia, such as TNF-α, IL-1β,

Fig. 3 I/R insult induces alterations in the cellular composition of BBB and the involved molecules. They ultimately cause TJs degradation and breakdown of the endothelial belt

and IL-6, upregulate EC adhesion molecules and speed up the infltration of leukocytes, contributing to the impairment of the BBB [\[50](#page-11-20)]. Importantly, diferent infammatory factors and chemokines secreted by microglia exert diferent function phenotypes. Here, we concluded common cytokines and chemokines that function on TJ proteins and ECs, ultimately afecting the integrity and function of the BBB.

Cytokines

Tumor necrosis factor (TNF)- α , an inflammatory factor released from M1 phenotype-activated microglia, is a pleiotropic molecule that regulates cell proliferation, death, immunity, and metabolism [\[51](#page-11-21)]. Upon injury, TNF- α increases as early as 6–24 h following permanent middle cerebral artery occlusion (pMCAO) [[52](#page-11-22)]. A. Q. Chen et al. found that M1 microglia-derived TNF- α could bond to its receptors on EC to initiate the necroptosis process, leading to endothelial necroptosis and destroying the completeness of the endothelial barrier [\[14\]](#page-10-13). Notably, ZO-1 is a downstream target of TNF- α signaling, and its level drops in cerebral I/R injury [[53\]](#page-11-23). However, the intercellular ZO-1 distribution and its expression got improved when inhibiting the TNF- α receptor with its antagonist, R-7050. Simultaneously, R-7050 also increases TEER (trans-endothelial electrical resistance) and lowers the permeability of the BBB [\[54](#page-11-24)].

IL-1 is mainly produced by resident microglia in the brain [\[55\]](#page-11-25), and its concentration increases up to 40 to 60- fold during the first 24 h after the MCAO condition [[56](#page-11-26)]. Importantly, systemically infusing IL-1β antibody ameliorates parenchymal brain injury and attenuates apoptosis and caspase-3 activity in non-neurons [[57\]](#page-11-27). R. Wong et al. found that deleting cerebral ECs IL-1R1 greatly decreased both ischemic lesions and IgG leakage. However, there's no sig-nificant impact on neurons [[58](#page-11-28)]. Another isoform of IL-1, IL-1α, is widely known as a direct regulator of angiogenesis after ischemic injury [\[59\]](#page-11-29). Furthermore, it exerts an identical function as IL-1β via IL-1RI [[60\]](#page-11-30). However, further explorations are necessary to determine the exact mechanisms underlying the functions.

In the acute phase of stroke, activated microglia would secrete IL-6 to contribute to post-stroke inflammation [\[61](#page-11-31)]. Previous studies showed that the expression and bioactivity of IL-6 were induced in the infarcted hemisphere in the pMCAO rat model [[62\]](#page-11-32). Additionally, after 24 h exposure to IL-6, BBB permeability was compromised [[63\]](#page-11-33). Likewise, the TJ protein expression decreased after treatment with IL-6, IL-17, or TNF-α. However, their study showed that IL-6 reduced ischemic damage by activating the STAT3 transcription factor singling pathway [[64](#page-12-0), [65\]](#page-12-1). Notably, oncostatin M (OSM) is the most powerful molecule among the IL-6 family to impair BBB function [[66\]](#page-12-2). Its expression peaked at 12 h post-stroke and remained stable until 72 h [\[67](#page-12-3)]. Recent evidence demonstrates its close relationship with the integrity of BBB. OSM was found to downregulate TJs thus impairing the endothelial integrity of BBB under both normal and infammatory conditions via prolonged activation of the JAK/STAT3 signaling pathway

[\[68\]](#page-12-4). Additionally, OSM also stimulates brain cells to produce prostaglandins and cytokines, thus decreasing TEER of in vitro BBB model [\[69](#page-12-5)]. The above fndings illustrate that IL-6 family closely associates with BBB function and may be a vital regulator to maintain an intact BBB.

Chemokines

Chemokines are a big group of small proteins secreted by resident microglia in post-stroke infammation and signal via G protein-coupled heptahelical chemokine receptors on the cell surface, attracting and activating leukocytes further [\[70](#page-12-6)]. Chemokine (C–C motif) ligand 2 (CCL2) participates in the infammatory response and modulates monocytes migrating into tissues and the subsequent diferentiation of monocytes into macrophages [[71](#page-12-7)]. CCL2/CCR2 increased its expression on brain ECs and ACs in the ipsilateral hemisphere, and the highest expression of CCL2 was observed 24 h after stroke. Additionally, CCL2 regulates BBB permeability via the interaction with CCR2 on brain endothelial cells [[72](#page-12-8)]. When mesenchymal stromal cells overexpressing CCR2 were transplanted into animal and cell models, ischemic lesions, BBB permeability, and neurological function improved. Furthermore, MSCCCR2 (mesenchymal stromal cells expressing CCR2 on the cell surface) partially restored BBB integrity in a PRDX4-mediated antioxidant manner after stroke [\[73](#page-12-9)]. Overall, CCL2 is one chemokine that links tightly to ECs tightly and is therefore relevant to the integrity of the BBB. Notwithstanding, more chemokines may take part in the mediation of BBB during physiological and pathophysiological conditions, which requires more research to illustrate.

Matrix Metalloproteinases (MMPs)

Matrix metalloproteinases (MMPs) are members of the endopeptidase's family. They function to degrade and remodel the extracellular matrix (ECM) proteins [\[74](#page-12-10)]. Studies demonstrated that MMP-2/-9 released from activated microglia could induce TJs degradation BBB dysfunction [\[75\]](#page-12-11). Besides microglia, astrocytes and some cortical and cerebellum neurons also express MMPs in the brain [\[76,](#page-12-12) [77](#page-12-13)]. MMP-9 (gelatinase B), a member of MMPs, are well known for its regulation of I/R-induced BBB breakdown via the degradation of TJ proteins and the basal lamina of capillaries and infltration of immune cells [[78](#page-12-14)]. Furthermore, the activation of endothelial MMP-9 was observed when endothelial cells were exposed to astrocytes, indicating a relationship between endothelial MMP-9 activity and astrocytic infuence. At the same time, ACs could release soluble factors to mediate BBB function via the activation of endothelial MMP-9 [\[79\]](#page-12-15). Recent fndings showed that ICS II, which is derived from Herbal Epimediiand, could bind to and inhibit MMP2/9 by attenuating the MMP/ TIMP1 balance to improve BBB dysfunction and suppress the death of hipponeurons [[80\]](#page-12-16). Besides, pretreatment with CB2R agonist JWH-133 prevented activating MMP9 and upregulated perivascular expression of STLR4/MMP9 after cerebral I/R injury [[81\]](#page-12-17). However, MMPs are emerging in research and demonstrate regulatory functions on the BBB, and more research is required to elucidate their underlying mechanisms.

Non‑Microglia Derived Molecules Mediate BBB Permeability

Micro‑RNAs

Micro-RNAs(miRNAs) that are small non-coding RNAs have emerged as a kind of gene expression regulator by inhibiting or degrading target mRNAs [[82](#page-12-18)]. During the last decade, accumulating research has emerged to prove the therapeutic function of miRNAs on experimental I/R murine models. Many miRNAs are abundant in endothelial cells, which resist the I/R insult by targeting TJ proteins or cellular components of ECs. The miRNA function in the I/R condition is listed in the following Table [1.](#page-6-0)

Circular RNAs

Circular RNAs (circRNAs) are a particular kind of RNA with a circular structure rather than a linear structure formed by a back-splicing mechanism that joins 30 and 50 methyl guanosine caps together [\[112](#page-13-0)]. They can specifcally combine with miRNAs to regulate the expression of relevant genes by acting as miRNA sponges [[113\]](#page-13-1). Circ-Foxo3 is highly expressed in normal tissues and its expression is usually downregulated in some pathological conditions [[114\]](#page-13-2). Recently, studies proved that circ-Foxo3 expression was elevated following tMCAO with autophagy activation. Moreover, short-FoxO3 exacerbated the TJ deformation, which can be attenuated by circ-FoxO3 overexpression, suggesting circ-FoxO3 is needed for the integrity of the BBB following ischemic stroke in an autophagy-dependent man-ner [[115](#page-13-3)]. With more focus on gene modifications via noncoding RNAs, miRNA and circRNAs may be a potential therapy for I/R injury.

Long Non‑coding RNAs

Long non-coding RNAs (LncRNAs) are non-coding RNAs that are longer than 200 nucleotides (nt), they can function in a regulatory role at post-transcriptional, epigenetic, translational, and post-translational levels. Unlike miR-NAs, lncRNAs are highly cell-specific [[116](#page-13-4)]. Of the many lncRNAs expressed in ECs, some are exclusively related to

miRNA		Locations Changes in I/R condi- tion	Treatment in experiments Function on I/R		Reference
miRNA-98	Xp11.22	↓	Overexpression	Improve neurological outcomes; Attenuate post-stroke inflammation; Reduce BBB disruption	[83]
m iRNA-126	9q34.3	↓	Knockdown Overexpression	Impair angiogenesis and vessel integrity; Suppress inflammation; Reduce infarct and edema volume; Elevate BBB leakage;	[84, 85]
miRNA-30a	6q13	↓	Inhibition	Reduce the permeability; Block the loss of TJ proteins Decrease zinc accumulation and the loss of ZnT4 in the microvessels of the ischemic brain	[86]
m _{RNA} -182	7q32.2	↑	Knockdown Inhibition	Target endothelial cells Reduce infarction volume; Preserve BBB integrity preserved; Attenuate degradation of TJ proteins	[87, 88]
miRNA-130a	11q12.1	↑	Inhibition	Regulate occludin via HoxA5; Reduce the size of cerebral infarct; Suppressed OGD-induced increase of paracellular perme- ability	[89]
miRNA-15a and $13q14.2$ m RN A-16-1	13q14.2	↑	Deletion	Upregulate Claudin-5; Inhibited M1-type microglia/macrophage polarization in the peri-infarct area; Enhanced angiogenesis; Improved long-term functional recovery	[90]
miRNA-132/212 17p13.3	11; 11B5	↑	Overexpression	Attenuates cerebral injury via repressing MMP; Target TJ proteins including Cldn1, Jam3, and Tjap1; Attenuate the decline in TEER values after OGD	[91, 92]
			Downregulation	Aggravate ischemic brain injury; Aggravate neuronal damage via RBFox-1; Suppress vascular bed; Destabilize tight junction protein expression	
miRNA-34a	1p36.22	↑	Knockout	Participate in the opening of BBB during the early phase of [93] ischemic reperfusion; Ameliorate stroke infarction; Reduce BBB permeability; Alleviates disruption of tight junctions; Improves stroke outcomes	
miRNA-92b	1q22	↓	Overexpression	Maintain the BBB integrity; Raise the viability and lessen the permeability of OGD- induced BMECs;	[94]
m iRNA-668	14q32.31		Inhibition	Attenuate infarction volume and BBB leakage; Attenuate neurological functions; Attenuate BBB permeability; Attenuate increase in cytokine levels; Attenuate neuronal cell apoptosis	[95]
miRNA-149-5	2q37.3	↓	Overexpression	Regulate pericyte migration, N-cadherin expression; Regulate BBB permeability via targeting S1PR2; Improved the outcome of transient middle cerebral artery occlusion (tMCAO) rats	[96, 97]
miRNA-29b	7q32.3	↓	Overexpression	Attenuate BBB dysfunction; Inhibit Aquaporin 4; Reduce infarct volume and edema; Decrease apoptosis of neurons and hCMEC/D3 cells	[98, 99]
m iRNA-150	19q13.33 \downarrow		Overexpression	Increase BBB permeability in vivo and in vitro; Decrease BMEC survival; Decrease claudin-5 expression	$[100]$

Table 1 miRNAs tightly associated with BBB damage in I/R conditions

Table 1 (continued)

miRNA	Locations	Changes in $\textsf{I/R}$ condi- tion	Treatment in experiments Function on I/R		Reference
m iRNA-122	18q21.31 \downarrow		Overexpression	Promote proliferation; Inhibit apoptosis and autophagy; Decrease the infarct area	[101, 102]
$miRNA-210$	11p15.5		Overexpression	Decrease infarct volume and hemispheric swelling	[103, 104]
m iRN-21	17q23.1		Inhibition	Downregulate MMP-9; Improve BBB permeation	[105, 106]
m _{RNA} -539	14q32.31		Overexpression	Inhibit OGD/R-induced BBB disruption; Suppress the expression of MMP-9 mRNA and protein	$[107]$
m _{RNA} -155	21q21.3		Deletion	Protect I/R-induced brain injury and neurological deficits;	[108]
m iRNA-149-5p	2q37.3		Overexpression	Attenuate pericytes migration; Increase N-cadherin in pericytes; Increase BBB integrity; Promote neurological recovery after stroke	[97]
miRNA-1	20q13.33		Inhibition	Reduce infarct volume; Reduce brain edema: Decrease BBB permeability	[109]
m iRNA-503	Xq26.3	↑	Inhibition	Reduce infarct volume; Improve BBB dysfunction; Increase cerebral blood flow (CBF); Decrease apoptosis and the production of ROS and NO in ECs Increase permeability of HBMECs	$[110]$
m iRNA-141-3p	$12p13.31 \quad \downarrow$		Inhibition	Higher cell apoptosis rate Increase ROS level in PC 12 cells	[111]

BBB function via infuencing the TJs dynamics. LncRNA HOTAIR also modifes the levels of occludin, claudin-5 and ZO-1 by interacting with upstream stimulatory factor 1 (USF1) [[117](#page-13-10)]. Some lncRNAs interact with miRNAs to mediate TJs, primarily functioning as miRNA sponges or competing endogenous RNAs (ceRNAs) to suspend the target genes of miRNAs. As reported, lnc00462717 could bind to polypyrimidine tract binding protein (PTBP1) to downregulate miR-186-5p, thus regulating occludin levels in an in vitro blood-tumor barrier model [[33\]](#page-11-3). Additionally, Linc00174 and FOS like 2 (FOSL2) bind to miR-138-5p and miR150-5p to exert TJs regulator function [[51](#page-11-21)]. Moreover, lncRNA MIAT competes with miR-140-3p to adjust TJs expression [[95](#page-12-31)]. Under the I/R condition, XIST was downregulated, leading to the decreased expression of ZO-1 and claudin-5 [[118](#page-13-11)]. Besides, inhibiting lncRNA H19 neuron exosomes signifcantly reduced the permeability of the EC monolayer in in vitro OGD/R conditions [[119\]](#page-13-12). Reported as the most upregulated lncRNA, the upregulation of Malat1 was found to contribute to the survival of brain microvascular endothelial cells (BMECs) by promoting the autophagy of BMECs [[120\]](#page-13-13). Meanwhile, You at el. found that MEG3 expression of BMECs increased in OGD/R condition and inhibiting its expression could relieve the hyperpermeability of the BBB [[121](#page-13-14)]. Several signaling pathways are known to

be involved, including the Wnt/β-catenin signaling pathway and Notch pathway, as well as some molecules like STAT3, HIF-1α, P53, NOX4 and VEGF [[121,](#page-13-14) [122](#page-13-15)].

Heat Shock Proteins

Heat shock proteins (HSPs) are a group of proteins binding to DNA to regulate gene expression [[123](#page-13-16)]. An early study reported that the expression of HSPB8 was upregulated to attenuate cell death induced by OGD/R injury [\[124](#page-13-17)]. Likewise, Li et al. found that HSPB8 signifcantly alleviated brain injury during I/R. However, they also observed aggravated mitophagy and TJs degradation following HSPB8 overexpression, which boosted autophagic fux and weakened its protective efect on BBB after MCAO/R [[124,](#page-13-17) [125](#page-13-18)]. Additionally, HSP70 overexpression facilitated neuronal repair and improved recovery of neurological function after stroke as well. Moreover, Jiang found that Pla-Exo inherited HSP70, and it reduced I/R injury by reducing ROS generation and maintaining the integral mitochondria. Notably, it also protects BBB from dysfunction by elevating the integrity of TJ proteins [\[126](#page-13-19)]. Furthermore, recent studies show that endothelial HSP70 induction by GGA could inhibit infammation BBB leakage following stroke [[127\]](#page-13-20).

Transporters and Ion Channels Participate in BBB Breakdown

Ion disturbance occurs early during I/R injury, contributing to hypoxic injury. Several ion channels and transporters are suggested to be involved in the regulation of ionic homeostasis during I/R, which is essential for BBB function, such as $Na⁺/H⁺$ exchanger isoform 1 (NHE1), $Na⁺/Ca²⁺$ exchanger (NCX), Na^+/Ca^{2+} cotransporters (NBCs), voltage-gated Na^+ channel (Navs), ATP-sensitive K^+ channel (K_{ATP} channel), $Na⁺-K⁺-ATP$ pump, and so on [[128](#page-13-32)[–131\]](#page-14-0). Here we try to give insights into the updates of these channel's roles in the setting of I/R injury.

NHE1

NHE1 mainly functions in astrocytes as a pathway for H^+ efflux in exchange for Na^+ influx [[132\]](#page-14-1). Recently, evidence demonstrated that infarction and neurological function both are improved in *Nhe1* null mice. Simultaneously, microvessel damage and BBB integrity were improved in *Nhe1* null mice in the setting of ischemic stroke [[133](#page-14-2)]. Moreover, in the astrocyte OGD/R model, NHE1 was activated, leading to Na⁺ overloading and swelling at the end feet [\[134](#page-14-3)]. Knockout of *Nhe1* in astrocytes resulted in reduced paracellular permeability and enhanced cerebral perfusion and angiogenesis. Additionally, the endothelial Wnt/β-catenin signaling pathways play a vital role in this process [[135\]](#page-14-4). Furthermore, NHE1 also exists at the abluminal side of endothelial cells in the brain capillaries, which regulates $Li⁺$ influx in brain endothelium and is critical to maintaining $Li⁺-Na⁺$ homeostasis [\[136\]](#page-14-5). Thus, the close association between NHE1 and astrocytes and its role in ionic homeostasis contributes to BBB regulation during I/R condition.

NCX

To sustain normal Ca^{2+} -Na⁺ homeostasis, NCX plays an indispensable part. NCX1-3 is selectively expressed in the brain tissues, such as pyramidal neurons in the cortex and dentate gyrus within the hippocampus [[137](#page-14-6), [138](#page-14-7)]. During ischemic stroke, NCX is downregulated in the core but upregulated in the penumbra [[139](#page-14-8)]. Moreso, recent evidence shows that the expression of NCX1 and NCX3 are reduced in the hippocampus of neonatal mice subjected to hypoxic-ischemic. In contrast, the activation of NCX significantly reduced hippocampal injury and improved short-term memory and motor performance [[140](#page-14-9)]. BBB dysfunction is closely related to intracellular calcium levels and calciummediated signaling, such as C3a/C3aR signaling, which functions as a second messenger to mediate endothelial pMLC activity and endothelial VE-cadherin homeostasis [\[141\]](#page-14-10).

NBCs

NBCs are widely expressed in the neurons and astrocytes [[142](#page-14-11), [143](#page-14-12)], and play a significant part in intracellular pH regulation and ionic homeostasis. It was found to increase within 12 h after I/R and it began to decrease 2 days after I/R and reached the lowest level at 2 days following I/R. Notably, neuronal death was improved with the application of an inorganic anion exchanger blocker DIDs [[144\]](#page-14-13). Another study showed that NBCn and NBCe1 were upregulated both in RNA and protein levels in experimental I/R conditions, which induced astrocytic death. However, NBC inhibition with S0859 aggravated astrocytic death [[145](#page-14-14)]. Conversely, S0859 attenuates neuronal death in the gerbil hippocampus [[130\]](#page-14-15). Considering its significant impact on astrocytes and neurons, two crucial components of the BBB, there's much potential for NBCs to play a part in BBB regulation following I/R insult. Furthermore, more studies are needed to elucidate its role in that function.

Other Molecules Involved in Post‑stroke BBB Permeability Modulation

Peroxiredoxin 4 (Prx4), belonging to the antioxidant enzyme family ($Prx1-6$), has been identified as an efficient scavenger for H_2O_2 [[146\]](#page-14-16). It likely also plays a direct role in protecting ECs from ROS and functions as a peroxidase that is associated with the membrane in ECs [[147\]](#page-14-17). Therefore, Prx4 may prevent endothelial dysfunction and alleviate BBB breakdown. However, more evidence is needed to illustrate the role Prx4 plays in BBB integrity and its underlying mechanism.

The Hippo/YAP kinase cascade induces the phosphorylation of YAP and TAZ which are crucial downstream efectors and co-activators of transcription. Activation of YAP or TAZ protected the brain tissue from cerebral edema elicited by I/R insult. Additionally, YAP could reduce the infarction volume and maintain TJ protein integrity, consequently, attenuating BBB dysfunction [[148\]](#page-14-18). However, some evidence shows that inhibition of YAP ameliorated I/R-induced damage and preserved the BBB integrity [\[149](#page-14-19)]. Thus, the contradictory efects of YAP on BBB function require added investigations for clarity.

Insights for Treatment

In the setting of I/R, many molecules are involved in the pathological process exacerbating or attenuating the injury, which provides targets for adjuvant treatment for ischemic stroke accompanying thrombolytic therapy with tPA. Microglia mainly participate in post-stroke infammation, and the strategy to suppress infammation can remarkably reduce the secretion of microglia-derived molecules, thus attenuating I/R injury. TPCD NPs are bioactive nanoparticle-derived multifunctional nanoparticles whose efectiveness in treating ischemic stroke has been tested in vivo and in vitro. Notably, studies show that TPCD NPs could signifcantly reduce the production of intracellular ROS, alleviate oxidative stress, and suppress infammation in OGD conditions. Simultaneously, TPCD NPs also mitigated neuronal death that was induced by activated microglia [\[150](#page-14-20)]. Furthermore, cottonseed oil can also suppress microglial and astrocytic activation via inhibiting TLR4/NF-κB, thus reducing brain edema induced by ischemic injury [\[151](#page-14-21)]. Moreover, a neutralized antibody targeting MMP-9 could attenuate BBB dysfunction. Furthermore, it even could neutralize MMP-9 in stroke patients' serum and brain tissue samples, providing strong evidence to support its clinical potential for acute stroke treatment [[152](#page-14-22)]. Liposomes are widely used to enhance the efficiency of treatment for cerebral vascular disease because of their ability to cross the BBB. Ginkgolide B (GB) is one ingredient of G. biloba leaf extract, which possesses the strongest efect in cerebral vascular disease treatment. Recent studies showed that a GB-DHA complex, which can be integrated into the lipid bilayer, thus crossing the BBB was beneficial to I/R injury $[153]$ $[153]$. Evidence has shown the efficiency of Baicalin (BA), an extract from Radix Scodelario, for I/R injury treatment. And Yu Long et al. found that the complex of Borneo and BA was highly lipid-soluble and could enhance its efficiency and could reduce BBB hyperpermeability induced by I/R injury[[154\]](#page-14-24). However, most of the efficiency of these treatments is still at the basic experimental stage. Therefore, more evidence and trials are needed to explore whether they are qualifed for clinical use.

Conclusion

The normal function of BBB is maintained by its essential cellular components, TJ proteins, and extracellular matrix (ECM). However, insufficient oxygen and glucose condition-the core of stroke pathology induces BBB dysfunction, resulting in its dysfunction. Simultaneously, the primary damage also causes neuroinfammation, which worsens BBB dysfunction, leading to the second opening of BBB. Microglia are activated following I/R injury, which is the hallmark of neuroinfammation. Once microglia are activated, they polarize into either the M1 phenotype to promote neuroinfammation, or the M2 phenotype to suppress neuroinfammation. M1 phenotype-activated microglia occur in the early phase of stroke and secretes infammatory factors to exert its proinfammatory function, which would deteriorate the prognosis of ischemic stroke. However, M2 phenotype-activated microglia emerge in the relatively later phase of ischemic stroke, suppressing the infammation and counteracting the tissue damage induced by M1 phenotype microglia. Notably, TNF-α, IL-1β, and IL-6, which are secreted by M1 phenotype microglia and increased upon I/R injury, can destroy TJs or infuence the function of ECs via receptors on their membrane, aggravating BBB dysfunction. Importantly, suppressing TNF- α and Il-1 β expression are reported to attenuate BBB damage induced by I/R injury. However, IL-6 is proven to play a protective role in I/R damage. Activated microglia are one of the MMPs resources which also play a part in BBB breakdown following ischemia/reperfusion injury. For instance, MMP-2 and MMP-9 have been proven to infuence either TJs, ACs, or ECs. Besides, non-microgliaderived molecules also contribute to BBB breakdown following I/R either in the primary damage stage or the secondary damage stage. miRNAs and circ-FoxO3 have been discovered to maintain BBB integrity during stroke. Additionally, HSPs, peroxiredoxin 4, and YAP participate in the disease progression of stroke, and they are involved in the autophagy process or/and infammatory process. The same goes for the ion channels and transporters responsible for the regulation of BBB function, which play a large part in maintaining the ion balance and homeostasis. Perturbations in the ion balance may lead to cascades that trigger microglial activation, infammation, autophagy, or necroptosis in cells, which eventually causes EC dysfunction, TJs degradation, and BBB breakdown. However, the precise mechanism of how these molecules protect BBB and brain tissues against stroke damage or aggravate its outcome is yet to be illustrated, which would give more clinical insight into treating ischemic stroke and improving its prognosis.

Numerous studies have highlighted the importance of microglia during ischemic stroke, as well as the role of microglia-derived and non-microglia-derived molecules in the progression of I/R injury. Microglia-derived molecules are mainly involved in the secondary damage to the central nervous system (CNS) and BBB dysfunction, whereas nonmicroglia-derived molecules can contribute to either primary or secondary damage, resulting in biphasic opening of the BBB. However, there are still several unanswered questions: How do these molecules contribute to primary damage? Do they only function in a neuroinfammatory manner? Do non-microglia-derived and microglia-derived molecules interact with each other to worsen disease progression, given that they may both engage in infammatory and non-infammatory processes? If so, how do they interact? To shed light on the role of microglia and its secreted molecules, as well as non-microglia-derived molecules, in I/R injury, particularly BBB breakdown, more studies are needed.

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content of the review. MJ and QW checked the grammar of the review. JW made critical revisions and approved the fnal version of the manuscript. All authors have read and agreed to the published version of the manuscript.

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Declarations

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References

- 1. Kuriakose D, Xiao Z (2020) Pathophysiology and treatment of stroke: present status and future perspectives. Int J Mol Sci. <https://doi.org/10.3390/ijms21207609>
- 2. Siesjö BK, Katsura K, Kristian T et al (1996) Molecular mechanisms of acidosis-mediated damage. Mechanisms of secondary brain damage in cerebral ischemia and trauma. Springer, Vienna
- 3. Liu L, Kearns KN, Eli I et al (2021) Microglial calcium waves during the hyperacute phase of ischemic stroke. Stroke 52(1):274–283. [https://doi.org/10.1161/STROKEAHA.120.](https://doi.org/10.1161/STROKEAHA.120.032766) [032766](https://doi.org/10.1161/STROKEAHA.120.032766)
- 4. Menaceur C, Gosselet F, Fenart L et al (2021) The blood-brain barrier, an evolving concept based on technological advances and cell-cell communications. Cells 11(1):133. [https://doi.org/](https://doi.org/10.3390/cells11010133) [10.3390/cells11010133](https://doi.org/10.3390/cells11010133)
- 5. Wang Y, Zhu Y, Wang J et al (2023) Purinergic signaling: a gatekeeper of blood-brain barrier permeation. Front Pharmacol. <https://doi.org/10.3389/fphar.2023.1112758>
- 6. Liebner S, Dijkhuizen RM, Reiss Y et al (2018) Functional morphology of the blood–brain barrier in health and disease. Acta Neuropathol 135:311–336. [https://doi.org/10.1007/](https://doi.org/10.1007/s00401-018-1815-1) [s00401-018-1815-1](https://doi.org/10.1007/s00401-018-1815-1)
- 7. Schaefer S, Iadecola C (2021) Revisiting the neurovascular unit. Nat Neurosci 24(9):1198–1209. [https://doi.org/10.1038/](https://doi.org/10.1038/s41593-021-00904-7) [s41593-021-00904-7](https://doi.org/10.1038/s41593-021-00904-7)
- 8. Chen S, Shao L, Ma L (2021) Cerebral edema formation after stroke: emphasis on blood–brain barrier and the lymphatic drainage system of the brain. Front Cell Neurosci 15:716825. [https://](https://doi.org/10.3389/fncel.2021.716825) doi.org/10.3389/fncel.2021.716825
- 9. Alajangi HK, Kaur M, Sharma A et al (2022) Blood–brain barrier: emerging trends on transport models and new-age strategies for therapeutics intervention against neurological disorders. Mol Brain 15(1):1–28.<https://doi.org/10.1186/s13041-022-00937-4>
- 10. Sashindranath M, Nandurkar HH (2021) Endothelial dysfunction in the brain: setting the stage for stroke and other cerebrovascular complications of COVID-19. Stroke 52(5):1895–1904. [https://](https://doi.org/10.1161/STROKEAHA.120.032711) doi.org/10.1161/STROKEAHA.120.032711
- 11. Krueger M, Mages B, Hobusch C et al (2019) Endothelial edema precedes blood-brain barrier breakdown in early time points after experimental focal cerebral ischemia. Acta Neuropathol Commun 7(1):1–17.<https://doi.org/10.1186/s40478-019-0671-0>
- 12. Ahnstedt H, Sweet J, Cruden P et al (2016) Effects of early postischemic reperfusion and tPA on cerebrovascular function and nitrosative stress in female rats. Transl Stroke Res 7(3):228–238. <https://doi.org/10.1007/s12975-016-0468-4>
- 13. Krueger M, Härtig W, Frydrychowicz C et al (2017) Strokeinduced blood–brain barrier breakdown along the vascular tree–no preferential afection of arteries in diferent animal models and in humans. J Cereb Blood Flow Metab 37(7):2539– 2554. <https://doi.org/10.1177/0271678X16670922>
- 14. Chen AQ, Fang Z, Chen XL et al (2019) Microglia-derived TNF-alpha mediates endothelial necroptosis aggravating blood brain-barrier disruption after ischemic stroke. Cell Death Dis 10(7):487.<https://doi.org/10.1038/s41419-019-1716-9>
- 15. Shi W, Wei X, Wang Z et al (2016) HDAC 9 exacerbates endothelial injury in cerebral ischaemia/reperfusion injury. J Cell Mol Med 20(6):1139–1149. [https://doi.org/10.1111/jcmm.](https://doi.org/10.1111/jcmm.12803) [12803](https://doi.org/10.1111/jcmm.12803)
- 16. Heinemann U, Schuetz A (2019) Structural features of tightjunction proteins. Int J Mol Sci 20(23):6020. [https://doi.org/10.](https://doi.org/10.3390/ijms20236020) [3390/ijms20236020](https://doi.org/10.3390/ijms20236020)
- 17. Funke L, Dakoji S, Bredt DS (2005) Membrane-associated guanylate kinases regulate adhesion and plasticity at cell junctions. Annu Rev Biochem 74:219–245. [https://doi.org/10.1146/annur](https://doi.org/10.1146/annurev.biochem.74.082803.133339) [ev.biochem.74.082803.133339](https://doi.org/10.1146/annurev.biochem.74.082803.133339)
- 18. Campbell HK, Maiers JL, DeMali KA (2017) Interplay between tight junctions & adherens junctions. Exp Cell Res 358(1):39–44. <https://doi.org/10.1016/j.yexcr.2017.03.061>
- 19. Beutel O, Maraspini R, Pombo-Garcia K et al (2019) Phase separation of zonula occludens proteins drives formation of tight junctions. Cell 179(4):923–936. [https://doi.org/10.1016/j.cell.](https://doi.org/10.1016/j.cell.2019.10.011) [2019.10.011](https://doi.org/10.1016/j.cell.2019.10.011)
- 20. Tsukita S, Tanaka H, Tamura A (2019) The claudins: from tight junctions to biological systems. Trends Biochem Sci 44(2):141– 152. <https://doi.org/10.1016/j.tibs.2018.09.008>
- 21. Yang Z, Lin P, Chen B et al (2021) Autophagy alleviates hypoxiainduced blood-brain barrier injury via regulation of CLDN5 (claudin 5). Autophagy 17(10):3048–3067. [https://doi.org/10.](https://doi.org/10.1080/15548627.2020.1851897) [1080/15548627.2020.1851897](https://doi.org/10.1080/15548627.2020.1851897)
- 22. Paradis T, Bègue H, Basmaciyan L et al (2021) Tight junctions as a key for pathogens invasion in intestinal epithelial cells. Int J Mol Sci 22(5):2506.<https://doi.org/10.3390/ijms22052506>
- 23. Itoh M, Morita K, Tsukita S (1999) Characterization of ZO-2 as a MAGUK family member associated with tight as well as adherens junctions with a binding affinity to occludin and alpha catenin. J Biol Chem 274(9):5981–5986. [https://doi.org/10.1074/](https://doi.org/10.1074/jbc.274.9.5981) [jbc.274.9.5981](https://doi.org/10.1074/jbc.274.9.5981)
- 24. Li W, Chen Z, Chin I et al (2018) The role of VE-cadherin in blood-brain barrier integrity under central nervous system pathological conditions. Curr Neuropharmacol 16(9):1375–1384. <https://doi.org/10.2174/1570159x16666180222164809>
- 25. Anquetil T, Solinhac R, Jafre A et al (2021) PI3KC2β inactivation stabilizes VE-cadherin junctions and preserves vascular integrity. EMBO Rep.<https://doi.org/10.15252/embr.202051299>
- 26. Weinl C, Castaneda Vega S, Riehle H et al (2015) Endothelial depletion of murine SRF/MRTF provokes intracerebral hemorrhagic stroke. Proc Natl Acad Sci 112(32):9914–9919. [https://](https://doi.org/10.1073/pnas.1509047112) doi.org/10.1073/pnas.1509047112
- 27. Zhang S, An Q, Wang T et al (2018) Autophagy-and MMP-2/9-mediated reduction and redistribution of ZO-1 contribute to hyperglycemia-increased blood–brain barrier permeability during early reperfusion in stroke. Neuroscience 377:126–137. <https://doi.org/10.1016/j.neuroscience.2018.02.035>
- 28. Kim K-A, Kim D, Kim J-H et al (2020) Autophagy-mediated occludin degradation contributes to blood–brain barrier disruption during ischemia in bEnd.3 brain endothelial cells and rat ischemic stroke models. Fluids Barriers CNS. [https://doi.org/10.](https://doi.org/10.1186/s12987-020-00182-8) [1186/s12987-020-00182-8](https://doi.org/10.1186/s12987-020-00182-8)
- 29. Sato Y, Falcone-Juengert J, Tominaga T et al (2022) Remodeling of the neurovascular unit following cerebral ischemia and hemorrhage. Cells 11(18):2823.<https://doi.org/10.3390/cells11182823>
- 30. Tachibana M, Ago T, Wakisaka Y et al (2017) Early reperfusion after brain ischemia has beneficial effects beyond rescuing neurons. Stroke 48(8):2222–2230. [https://doi.org/10.1161/](https://doi.org/10.1161/STROKEAHA.117.016689) [STROKEAHA.117.016689](https://doi.org/10.1161/STROKEAHA.117.016689)
- 31. Zhou S-Y, Guo Z-N, Zhang D-H et al (2022) The role of pericytes in ischemic stroke: fom cellular functions to therapeutic targets. Front Mol Neurosci. [https://doi.org/10.3389/fnmol.](https://doi.org/10.3389/fnmol.2022.866700) [2022.866700](https://doi.org/10.3389/fnmol.2022.866700)
- 32. Dalkara T, Alarcon-Martinez L, Yemisci M (2019) Pericytes in ischemic stroke. Springer, Cham
- 33. Sun Z, Gao C, Gao D et al (2021) Reduction in pericyte coverage leads to blood-brain barrier dysfunction via endothelial transcytosis following chronic cerebral hypoperfusion. Fluids Barriers CNS 18(1):21.<https://doi.org/10.1186/s12987-021-00255-2>
- 34. Michinaga S, Koyama Y (2019) Dual roles of astrocyte-derived factors in regulation of blood-brain barrier function after brain damage. Int J Mol Sci 20(3):571. [https://doi.org/10.3390/ijms2](https://doi.org/10.3390/ijms20030571) [0030571](https://doi.org/10.3390/ijms20030571)
- 35. Haley MJ, Lawrence CB (2017) The blood–brain barrier after stroke: structural studies and the role of transcytotic vesicles. J Cereb Blood Flow Metab 37(2):456–470. [https://doi.org/10.](https://doi.org/10.1177/0271678X16629976) [1177/0271678X16629976](https://doi.org/10.1177/0271678X16629976)
- 36. Xu D, Kong T, Shao Z et al (1867) (2021) Orexin-A alleviates astrocytic apoptosis and infammation via inhibiting OX1Rmediated NF-κB and MAPK signaling pathways in cerebral ischemia/reperfusion injury. Biochimica et Biophysica Acta (BBA) 11:166230.<https://doi.org/10.1016/j.bbadis.2021.166230>
- 37. Daneman R, Engelhardt B (2017) Brain barriers in health and disease. Neurobiol Dis 107:1–3. [https://doi.org/10.1016/j.nbd.](https://doi.org/10.1016/j.nbd.2017.05.008) [2017.05.008](https://doi.org/10.1016/j.nbd.2017.05.008)
- 38. Eilam R, Segal M, Malach R et al (2018) A strocyte disruption of neurovascular communication is linked to cortical damage in an animal model of multiple sclerosis. Glia 66(5):1098–1117. <https://doi.org/10.1002/glia.23304>
- 39. Ronaldson PT, Davis TP (2020) Regulation of blood-brain barrier integrity by microglia in health and disease: a therapeutic opportunity. J Cereb Blood Flow Metab 40(1):S6–S24. [https://](https://doi.org/10.1177/0271678X20951995) doi.org/10.1177/0271678X20951995
- 40. Varatharaj A, Galea I (2017) The blood-brain barrier in systemic infammation. Brain Behav Immun 60:1–12. [https://doi.org/10.](https://doi.org/10.1016/j.bbi.2016.03.010) [1016/j.bbi.2016.03.010](https://doi.org/10.1016/j.bbi.2016.03.010)
- 41. Jiang X, Andjelkovic AV, Zhu L et al (2018) Blood-brain barrier dysfunction and recovery after ischemic stroke. Prog Neurobiol 163–164:144–171. [https://doi.org/10.1016/j.pneurobio.2017.10.](https://doi.org/10.1016/j.pneurobio.2017.10.001) [001](https://doi.org/10.1016/j.pneurobio.2017.10.001)
- 42. Keaney J, Campbell M (2015) The dynamic blood–brain barrier. FEBS J 282(21):4067–4079.<https://doi.org/10.1111/febs.13412>
- 43. Chen A-Q, Fang Z, Chen X-L et al (2019) Microglia-derived TNF-α mediates endothelial necroptosis aggravating blood brain–barrier disruption after ischemic stroke. Cell Death Dis 10(7):1–18.<https://doi.org/10.1038/s41419-019-1716-9>
- 44. Dong R, Huang R, Wang J et al (2021) Efects of microglial activation and polarization on brain injury after stroke. Front Neurol 12:620948.<https://doi.org/10.3389/fneur.2021.620948>
- 45. Li S, Kumar TP, Joshee S et al (2018) Endothelial cell-derived GABA signaling modulates neuronal migration and postnatal behavior. Cell Res 28(2):221–248. [https://doi.org/10.1038/cr.](https://doi.org/10.1038/cr.2017.135) [2017.135](https://doi.org/10.1038/cr.2017.135)
- 46. Pulido RS, Munji RN, Chan TC et al (2020) Neuronal activity regulates blood-brain barrier efflux transport through endothelial circadian genes. Neuron 108(5):937–952. [https://doi.org/10.](https://doi.org/10.1016/j.neuron.2020.09.002) [1016/j.neuron.2020.09.002](https://doi.org/10.1016/j.neuron.2020.09.002)
- 47. Hrvatin S, Hochbaum DR, Nagy MA et al (2018) Single-cell analysis of experience-dependent transcriptomic states in the mouse visual cortex. Nat Neurosci 21(1):120–129. [https://doi.](https://doi.org/10.1038/s41593-017-0029-5) [org/10.1038/s41593-017-0029-5](https://doi.org/10.1038/s41593-017-0029-5)
- 48. Zhang X, Wei M, Fan J et al (2021) Ischemia-induced upregulation of autophagy preludes dysfunctional lysosomal storage and associated synaptic impairments in neurons. Autophagy 17(6):1519–1542. [https://doi.org/10.1080/15548627.2020.18407](https://doi.org/10.1080/15548627.2020.1840796) [96](https://doi.org/10.1080/15548627.2020.1840796)
- 49. Kim Y, Lee S, Zhang H et al (2020) CLEC14A defciency exacerbates neuronal loss by increasing blood-brain barrier permeability and infammation. J Neuroinfammation 17(1):1–14. [https://](https://doi.org/10.1186/s12974-020-1727-6) doi.org/10.1186/s12974-020-1727-6
- 50. Shao Z, Tu S, Shao A (2019) Pathophysiological mechanisms and potential therapeutic targets in intracerebral hemorrhage. Front Pharmacol 10:1079. [https://doi.org/10.3389/fphar.2019.](https://doi.org/10.3389/fphar.2019.01079) [01079](https://doi.org/10.3389/fphar.2019.01079)
- 51. Bu L, Cao X, Zhang Z et al (2020) Decreased secretion of tumor necrosis factor-alpha attenuates macrophages-induced insulin resistance in skeletal muscle. Life Sci 244:117304. [https://doi.](https://doi.org/10.1016/j.lfs.2020.117304) [org/10.1016/j.lfs.2020.117304](https://doi.org/10.1016/j.lfs.2020.117304)
- 52. Yang T, Feng C, Wang D et al (2020) Neuroprotective and antiinfammatory efect of tangeretin against cerebral ischemia-reperfusion injury in rats. Infammation 43(6):2332–2343. [https://](https://doi.org/10.1007/s10753-020-01303-z) doi.org/10.1007/s10753-020-01303-z
- 53. Fang M, Zhong WH, Song WL et al (2018) Ulinastatin ameliorates pulmonary capillary endothelial permeability induced by sepsis through protection of tight junctions via inhibition of TNF-alpha and related pathways. Front Pharmacol 9:823. [https://](https://doi.org/10.3389/fphar.2018.00823) doi.org/10.3389/fphar.2018.00823
- 54. Lin SY, Wang YY, Chang CY et al (2021) TNF-alpha receptor inhibitor alleviates metabolic and infammatory changes in a rat model of ischemic stroke. Antioxidants (Basel). [https://doi.org/](https://doi.org/10.3390/antiox10060851) [10.3390/antiox10060851](https://doi.org/10.3390/antiox10060851)
- 55. Liu X, Quan N (2018) Microglia and CNS interleukin-1: beyond immunological concepts. Front Neurol 9:8. [https://doi.org/10.](https://doi.org/10.3389/fneur.2018.00008) [3389/fneur.2018.00008](https://doi.org/10.3389/fneur.2018.00008)
- 56. Lambertsen KL, Biber K, Finsen B (2012) Inflammatory cytokines in experimental and human stroke. J Cereb Blood Flow Metab 32(9):1677–1698.<https://doi.org/10.1038/jcbfm.2012.88>
- 57. Chen X, Hovanesian V, Naqvi S et al (2018) Systemic infusions of anti-interleukin-1beta neutralizing antibodies reduce shortterm brain injury after cerebral ischemia in the ovine fetus. Brain Behav Immun 67:24–35. [https://doi.org/10.1016/j.bbi.2017.08.](https://doi.org/10.1016/j.bbi.2017.08.002) [002](https://doi.org/10.1016/j.bbi.2017.08.002)
- 58. Wong R, Lenart N, Hill L et al (2019) Interleukin-1 mediates ischaemic brain injury via distinct actions on endothelial cells and cholinergic neurons. Brain Behav Immun 76:126–138. <https://doi.org/10.1016/j.bbi.2018.11.012>
- 59. Salmeron KE, Maniskas ME, Edwards DN et al (2019) Interleukin 1 alpha administration is neuroprotective and neuro-restorative following experimental ischemic stroke. J Neuroinfammation 16:1–14.<https://doi.org/10.1186/s12974-019-1599-9>
- 60. Fettelschoss A, Kistowska M, LeibundGut-Landmann S et al (2011) Infammasome activation and IL-1β target IL-1α for secretion as opposed to surface expression. Proc Natl Acad Sci 108(44):18055–18060.<https://doi.org/10.1073/pnas.1109176108>
- 61. Zhang W, Tian T, Gong S-X et al (2021) Microglia-associated neuroinfammation is a potential therapeutic target for ischemic stroke. Neural Regen Res 16(1):6. [https://doi.org/10.4103/1673-](https://doi.org/10.4103/1673-5374.286954) [5374.286954](https://doi.org/10.4103/1673-5374.286954)
- 62. Wang X, Yue TL, Young PR et al (1995) Expression of interleukin-6, c-fos, and zif268 mRNAs in rat ischemic cortex. J Cereb Blood Flow Metab 15(1):166–171. [https://doi.org/10.1038/](https://doi.org/10.1038/jcbfm.1995.18) [jcbfm.1995.18](https://doi.org/10.1038/jcbfm.1995.18)
- 63. Voirin AC, Perek N, Roche F (2020) Infammatory stress induced by a combination of cytokines (IL-6, IL-17, TNF- α) leads to a loss of integrity on bEnd.3 endothelial cells in vitro BBB model. Brain Res 1730:146647. [https://doi.org/10.1016/j.brainres.2020.](https://doi.org/10.1016/j.brainres.2020.146647) [146647](https://doi.org/10.1016/j.brainres.2020.146647)
- 64. Jung JE, Kim GS, Chan PH (2011) Neuroprotection by interleukin-6 is mediated by signal transducer and activator of transcription 3 and antioxidative signaling in ischemic stroke. Stroke 42(12):3574–3579. [https://doi.org/10.1161/strokeaha.](https://doi.org/10.1161/strokeaha.111.626648) [111.626648](https://doi.org/10.1161/strokeaha.111.626648)
- 65. Mahdiani S, Omidkhoda N, Rezaee R et al (2022) Induction of JAK2/STAT3 pathway contributes to protective efects of diferent therapeutics against myocardial ischemia/reperfusion. Biomed Pharmacother 155:113751. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.biopha.2022.113751) [biopha.2022.113751](https://doi.org/10.1016/j.biopha.2022.113751)
- 66. Takata F, Dohgu S, Matsumoto J et al (2018) Oncostatin M– induced blood-brain barrier impairment is due to prolonged activation of STAT3 signaling in vitro. J Cell Biochem 119(11):9055–9063.<https://doi.org/10.1002/jcb.27162>
- 67. Han J, Feng Z, Xie Y et al (2019) Oncostatin M-induced upregulation of SDF-1 improves bone marrow stromal cell migration in a rat middle cerebral artery occlusion stroke model. Exp Neurol 313:49–59
- 68. Hermans D, Houben E, Baeten P et al (2022) Oncostatin M triggers brain infammation by compromising blood–brain barrier integrity. Acta Neuropathol 144(2):259–281. [https://doi.](https://doi.org/10.1007/s00401-022-02445-0) [org/10.1007/s00401-022-02445-0](https://doi.org/10.1007/s00401-022-02445-0)
- 69. Repovic P, Mi K, Benveniste EN (2003) Oncostatin M enhances the expression of prostaglandin E2 and cyclooxygenase-2 in astrocytes: synergy with interleukin-1β, tumor necrosis factor-α, and bacterial lipopolysaccharide. Glia 42(4):433– 446.<https://doi.org/10.1002/glia.10182>
- 70. Hughes CE, Nibbs RJ (2018) A guide to chemokines and their receptors. FEBS J 285(16):2944–2971. [https://doi.org/10.1111/](https://doi.org/10.1111/febs.14466) [febs.14466](https://doi.org/10.1111/febs.14466)
- 71. Hao Q, Vadgama JV, Wang P (2020) CCL2/CCR2 signaling in cancer pathogenesis. Cell Commun Signal 18:1–13. [https://](https://doi.org/10.1186/s12964-020-00589-8) doi.org/10.1186/s12964-020-00589-8
- 72. Dimitrijevic OB, Stamatovic SM, Keep RF et al (2006) Efects of the chemokine CCL2 on blood-brain barrier permeability during ischemia-reperfusion injury. J Cereb Blood Flow Metab 26(6):797–810. <https://doi.org/10.1038/sj.jcbfm.9600229>
- 73. Huang Y, Wang J, Cai J et al (2018) Targeted homing of CCR2-overexpressing mesenchymal stromal cells to ischemic brain enhances post-stroke recovery partially through PRDX4 mediated blood-brain barrier preservation. Theranostics 8(21):5929–5944.<https://doi.org/10.7150/thno.28029>
- 74. Kapoor C, Vaidya S, Wadhwan V et al (2016) Seesaw of matrix metalloproteinases (MMPs). J Cancer Res Ther 12(1):28. <https://doi.org/10.4103/0973-1482.157337>
- 75. Ruan Z, Zhang D, Huang R et al (2022) Microglial activation damages dopaminergic neurons through MMP-2/-9-mediated increase of blood-brain barrier permeability in a Parkinson's disease mouse model. Int J Mol Sci 23(5):2793. [https://doi.org/](https://doi.org/10.3390/ijms23052793) [10.3390/ijms23052793](https://doi.org/10.3390/ijms23052793)
- 76. Lee E-J, Park J-S, Lee Y-Y et al (2018) Anti-infammatory and anti-oxidant mechanisms of an MMP-8 inhibitor in lipoteichoic acid-stimulated rat primary astrocytes: involvement of NF-κB, Nrf2, and PPAR-γ signaling pathways. J Neuroinfammation. <https://doi.org/10.1186/s12974-018-1363-6>
- 77. Beroun A, Mitra S, Michaluk P et al (2019) MMPs in learning and memory and neuropsychiatric disorders. Cell Mol Life Sci 76(16):3207–3228. [https://doi.org/10.1007/](https://doi.org/10.1007/s00018-019-03180-8) [s00018-019-03180-8](https://doi.org/10.1007/s00018-019-03180-8)
- 78. Asahi M, Wang X, Mori T et al (2001) Efects of matrix metalloproteinase-9 gene knock-out on the proteolysis of blood-brain barrier and white matter components after cerebral ischemia. J Neurosci 21(19):7724–7732. [https://doi.org/10.1523/jneurosci.](https://doi.org/10.1523/jneurosci.21-19-07724.2001) [21-19-07724.2001](https://doi.org/10.1523/jneurosci.21-19-07724.2001)
- 79. Spampinato SF, Merlo S, Sano Y et al (2017) Astrocytes contribute to Aβ-induced blood-brain barrier damage through activation

 $\circled{2}$ Springer

of endothelial MMP9. J Neurochem 142(3):464–477. [https://doi.](https://doi.org/10.1111/jnc.14068) [org/10.1111/jnc.14068](https://doi.org/10.1111/jnc.14068)

- 80. Liu MB, Wang W, Gao JM et al (2020) Icariside II attenuates cerebral ischemia/reperfusion-induced blood-brain barrier dysfunction in rats via regulating the balance of MMP9/TIMP1. Acta Pharmacol Sin 41(12):1547–1556. [https://doi.org/10.1038/](https://doi.org/10.1038/s41401-020-0409-3) [s41401-020-0409-3](https://doi.org/10.1038/s41401-020-0409-3)
- 81. Jing N, Fang B, Li Z et al (2020) Exogenous activation of cannabinoid-2 receptor modulates TLR4/MMP9 expression in a spinal cord ischemia reperfusion rat model. J Neuroinfamm 17(1):1–14.<https://doi.org/10.1186/s12974-020-01784-7>
- 82. Fish JE, Santoro MM, Morton SU et al (2008) miR-126 regulates angiogenic signaling and vascular integrity. Dev Cell 15(2):272– 284. <https://doi.org/10.1016/j.devcel.2008.07.008>
- 83. Bernstein DL, Zuluaga-Ramirez V, Gajghate S et al (2020) miR-98 reduces endothelial dysfunction by protecting blood-brain barrier (BBB) and improves neurological outcomes in mouse ischemia/reperfusion stroke model. J Cereb Blood Flow Metab 40(10):1953–1965. <https://doi.org/10.1177/0271678x19882264>
- 84. Harris TA, Yamakuchi M, Ferlito M et al (2008) MicroRNA-126 regulates endothelial expression of vascular cell adhesion molecule 1. Proc Natl Acad Sci USA 105(5):1516–1521. [https://doi.](https://doi.org/10.1073/pnas.0707493105) [org/10.1073/pnas.0707493105](https://doi.org/10.1073/pnas.0707493105)
- 85. Pan J, Qu M, Li Y et al (2020) MicroRNA-126-3p/-5p overexpression attenuates blood-brain barrier disruption in a mouse model of middle cerebral artery occlusion. Stroke 51(2):619– 627. <https://doi.org/10.1161/strokeaha.119.027531>
- 86. Wang P, Pan R, Weaver J et al (2021) MicroRNA-30a regulates acute cerebral ischemia-induced blood-brain barrier damage through ZnT4/zinc pathway. J Cereb Blood Flow Metab 41(3):641–655.<https://doi.org/10.1177/0271678x20926787>
- 87. Li Y, Zhang D, Wang X et al (2015) Hypoxia-inducible miR-182 enhances HIF1α signaling via targeting PHD2 and FIH1 in prostate cancer. Sci Rep 5:12495–12495. [https://doi.org/10.1038/](https://doi.org/10.1038/srep12495) [srep12495](https://doi.org/10.1038/srep12495)
- 88. Yi H, Huang Y, Yang F et al (2017) MicroRNA-182 aggravates cerebral ischemia injury by targeting inhibitory member of the ASPP family (iASPP). Arch Biochem Biophys 620:52–58. <https://doi.org/10.1016/j.abb.2016.05.002>
- 89. Wang Y, Wang MD, Xia YP et al (2018) MicroRNA-130a regulates cerebral ischemia-induced blood-brain barrier permeability by targeting Homeobox A5. Faseb J 32(2):935–944. [https://doi.](https://doi.org/10.1096/fj.201700139RRR) [org/10.1096/f.201700139RRR](https://doi.org/10.1096/fj.201700139RRR)
- 90. Sun P, Zhang K, Hassan SH et al (2020) Endothelium-targeted deletion of microRNA-15a/16-1 promotes poststroke angiogenesis and improves long-term neurological recovery. Circ Res 126(8):1040–1057. [https://doi.org/10.1161/CIRCRESAHA.119.](https://doi.org/10.1161/CIRCRESAHA.119.315886) [315886](https://doi.org/10.1161/CIRCRESAHA.119.315886)
- 91. Burek M, König A, Lang M et al (2019) Hypoxia-induced MicroRNA-212/132 alter blood-brain barrier integrity through inhibition of tight junction-associated proteins in human and mouse brain microvascular endothelial cells. Transl Stroke Res 10(6):672–683.<https://doi.org/10.1007/s12975-018-0683-2>
- 92. Yan H, Kanki H, Matsumura S et al (2021) MiRNA-132/212 regulates tight junction stabilization in blood-brain barrier after stroke. Cell Death Discov 7(1):380. [https://doi.org/10.1038/](https://doi.org/10.1038/s41420-021-00773-w) [s41420-021-00773-w](https://doi.org/10.1038/s41420-021-00773-w)
- 93. Hu H, Hone EA, Provencher EAP et al (2020) MiR-34a interacts with cytochrome c and shapes stroke outcomes. Sci Rep 10(1):3233.<https://doi.org/10.1038/s41598-020-59997-y>
- 94. Shen J, Li G, Zhu Y et al (2021) Foxo1-induced miR-92b downregulation promotes blood-brain barrier damage after ischaemic stroke by targeting NOX4. J Cell Mol Med 25(11):5269–5282. <https://doi.org/10.1111/jcmm.16537>
- 95. He J, Zhang X (2020) miR-668 inhibitor attenuates mitochondrial membrane potential and protects against neuronal

apoptosis in cerebral ischemic stroke. Folia Neuropathol 58(1):22–29. <https://doi.org/10.5114/fn.2020.94003>

- 96. Sayed ASM, Xia K, Li F et al (2015) The diagnostic value of circulating microRNAs for middle-aged (40–60-year-old) coronary artery disease patients. Clinics (Sao Paulo) 70(4):257– 263. [https://doi.org/10.6061/clinics/2015\(04\)07](https://doi.org/10.6061/clinics/2015(04)07)
- 97. Wan Y, Jin HJ, Zhu YY et al (2018) MicroRNA-149-5p regulates blood-brain barrier permeability after transient middle cerebral artery occlusion in rats by targeting S1PR2 of pericytes. Faseb J 32(6):3133–3148. [https://doi.org/10.1096/f.](https://doi.org/10.1096/fj.201701121R) [201701121R](https://doi.org/10.1096/fj.201701121R)
- 98. Ma X, Yun HJ, Elkin K et al (2022) MicroRNA-29b suppresses inflammation and protects blood-brain barrier integrity in ischemic stroke. Mediators Infamm 2022:1–11. [https://doi.org/](https://doi.org/10.1155/2022/1755416) [10.1155/2022/1755416](https://doi.org/10.1155/2022/1755416)
- 99. Wang Y, Huang J, Ma Y et al (2015) MicroRNA-29b is a therapeutic target in cerebral ischemia associated with aquaporin 4. J Cereb Blood Flow Metab 35(12):1977–1984. [https://doi.org/10.](https://doi.org/10.1038/jcbfm.2015.156) [1038/jcbfm.2015.156](https://doi.org/10.1038/jcbfm.2015.156)
- 100. Fang Z, He QW, Li Q et al (2016) MicroRNA-150 regulates blood-brain barrier permeability via Tie-2 after permanent middle cerebral artery occlusion in rats. FASEB J 30(6):2097–2107. [https://doi.org/10.1096/f.201500126](https://doi.org/10.1096/fj.201500126)
- 101. Wu Y, Gao Z, Zhang J (2020) <p>Transcription Factor E2F1 aggravates neurological injury in ischemic stroke via micro-RNA-122-targeted sprouty2</p>. Neuropsychiatr Dis Treat 16:2633–2647.<https://doi.org/10.2147/ndt.s271320>
- 102. Wang M, Liu X, Wu Y et al (2021) ΜicroRNA-122 protects against ischemic stroke by targeting Maf1. Exp Ther Med. <https://doi.org/10.3892/etm.2021.10048>
- 103. Pfeifer S, Tomašcová A, Mamrak U et al (2021) AMPK-regulated miRNA-210-3p is activated during ischaemic neuronal injury and modulates PI3K-p70S6K signalling. J Neurochem 159(4):710–728.<https://doi.org/10.1111/jnc.15347>
- 104. Liang C, Ni G-X, Shi X-L et al (2020) Astragaloside IV regulates the HIF/VEGF/Notch signaling pathway through miRNA-210 to promote angiogenesis after ischemic stroke. Restor Neurol Neurosci 38(3):271–282. <https://doi.org/10.3233/RNN-201001>
- 105. Deng X, Zhong Y, Gu L et al (2013) MiR-21 involve in ERKmediated upregulation of MMP9 in the rat hippocampus following cerebral ischemia. Brain Res Bull 94:56–62. [https://doi.org/](https://doi.org/10.1016/j.brainresbull.2013.02.007) [10.1016/j.brainresbull.2013.02.007](https://doi.org/10.1016/j.brainresbull.2013.02.007)
- 106. Zhai K, Duan H, Wang W et al (2021) Ginsenoside Rg1 ameliorates blood–brain barrier disruption and traumatic brain injury via attenuating macrophages derived exosomes miR-21 release. Acta Pharmaceutica Sinica B 11(11):3493–3507. [https://doi.org/](https://doi.org/10.1016/j.apsb.2021.03.032) [10.1016/j.apsb.2021.03.032](https://doi.org/10.1016/j.apsb.2021.03.032)
- 107. Fan F, Yang J, Xu Y et al (2018) MiR-539 targets MMP-9 to regulate the permeability of blood-brain barrier in ischemia/ reperfusion injury of brain. Neurochem Res 43(12):2260–2267. <https://doi.org/10.1007/s11064-018-2646-0>
- 108. Suofu Y, Wang X, He Y et al (2020) Mir-155 knockout protects against ischemia/reperfusion-induced brain injury and hemorrhagic transformation. NeuroReport 31(3):235–239. [https://doi.](https://doi.org/10.1007/s12975-020-00794-0) [org/10.1007/s12975-020-00794-0](https://doi.org/10.1007/s12975-020-00794-0)
- 109. Talebi A, Rahnema M, Bigdeli MR (2019) Efect of intravenous injection of antagomiR-1 on brain ischemia. Mol Biol Rep 46(1):1149–1155.<https://doi.org/10.1007/s11033-018-04580-y>
- 110. Zhang H, Pan Q, Xie Z et al (2020) Implication of Micro-RNA503 in brain endothelial cell function and ischemic stroke. Transl Stroke Res 11(5):1148–1164. [https://doi.org/10.1007/](https://doi.org/10.1007/s12975-020-00794-0) [s12975-020-00794-0](https://doi.org/10.1007/s12975-020-00794-0)
- 111. Zhou B, Liu H-Y, Zhu B-L et al (2019) MicroRNA-141 protects PC12 cells against hypoxia/reoxygenation-induced injury via regulating Keap1-Nrf2 signaling pathway. J Bioenerg Biomembr 51(4):291–300. <https://doi.org/10.1007/s10863-019-09804-9>
- 112. Guerra BS, Lima J, Araujo B et al (2021) Biogenesis of circular RNAs and their role in cellular and molecular phenotypes of neurological disorders. Semin Cell Dev Biol 114:1–10. [https://](https://doi.org/10.1016/j.semcdb.2020.08.003) doi.org/10.1016/j.semcdb.2020.08.003
- 113. Panda AC (2018) Circular RNAs act as miRNA sponges. Circular RNAs: biogenesis and functions. Springer, Singapore, pp 67–79
- 114. Yang T, Li Y, Zhao F et al (2021) Circular RNA Foxo3: a promising cancer-associated biomarker. Front Genet 12:652995. [https://](https://doi.org/10.3389/fgene.2021.652995) doi.org/10.3389/fgene.2021.652995
- 115. Yang Z, Huang C, Wen X et al (2021) Circular RNA circ-FoxO3 attenuates blood-brain barrier damage by inducing autophagy during ischemia/reperfusion. Mol Ther. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.ymthe.2021.11.004) [ymthe.2021.11.004](https://doi.org/10.1016/j.ymthe.2021.11.004)
- 116. Quinn JJ, Chang HY (2016) Unique features of long non-coding RNA biogenesis and function. Nat Rev Genet 17(1):47–62. <https://doi.org/10.1038/nrg.2015.10>
- 117. Sa L, Li Y, Zhao L et al (2017) The role of HOTAIR/miR-148b-3p/USF1 on regulating the permeability of BTB. Front Mol Neurosci 10:194.<https://doi.org/10.3389/fnmol.2017.00194>
- 118. Wang C, Dong J, Sun J et al (2021) Silencing of lncRNA XIST impairs angiogenesis and exacerbates cerebral vascular injury after ischemic stroke. Molecular Therapy-Nucleic Acids 26:148– 160. <https://doi.org/10.1016/j.omtn.2021.06.025>
- 119. Wang J, Cao B, Sun R et al (2022) Exosome-transported long non-coding ribonucleic acid H19 induces blood–brain barrier disruption in cerebral ischemic stroke Via the H19/micro ribonucleic acid-18a/vascular endothelial growth factor axis. Neuroscience 500:41–51. [https://doi.org/10.1016/j.neuroscience.2022.07.](https://doi.org/10.1016/j.neuroscience.2022.07.028) [028](https://doi.org/10.1016/j.neuroscience.2022.07.028)
- 120. Li Z, Li J, Tang N (2017) Long noncoding RNA Malat1 is a potent autophagy inducer protecting brain microvascular endothelial cells against oxygen-glucose deprivation/reoxygenation-induced injury by sponging miR-26b and upregulating ULK2 expression. Neuroscience 354:1–10. [https://doi.org/10.](https://doi.org/10.1016/j.neuroscience.2017.04.017) [1016/j.neuroscience.2017.04.017](https://doi.org/10.1016/j.neuroscience.2017.04.017)
- 121. You D, You H (2019) Repression of long non-coding RNA MEG3 restores nerve growth and alleviates neurological impairment after cerebral ischemia-reperfusion injury in a rat model. Biomed Pharmacother 111:1447–1457. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.biopha.2018.12.067) [biopha.2018.12.067](https://doi.org/10.1016/j.biopha.2018.12.067)
- 122. Wang S, Liang K, Hu Q et al (2017) JAK2-binding long noncoding RNA promotes breast cancer brain metastasis. J Clin Investig 127(12):4498–4515.<https://doi.org/10.1016/j.dib.2020.106260>
- 123. Zhang X, Li Y, Sun Y et al (2020) Regulatory efect of heat shock transcription factor-1 gene on heat shock proteins and its transcriptional regulation analysis in small abalone Haliotis diversicolor. BMC Mol Cell Biol 21:1-12. [https://doi.org/10.1186/](https://doi.org/10.1186/s12860-020-00323-9) [s12860-020-00323-9](https://doi.org/10.1186/s12860-020-00323-9)
- 124. Li F, Yang B, Li T et al (2019) HSPB8 over-expression prevents disruption of blood-brain barrier by promoting autophagic fux after cerebral ischemia/reperfusion injury. J Neurochem 148(1):97–113.<https://doi.org/10.1111/jnc.14626>
- 125. Li F, Tan J, Zhou F et al (2018) Heat shock protein B8 (HSPB8) reduces oxygen-glucose deprivation/reperfusion injury via the induction of mitophagy. Cell Physiol Biochem 48(4):1492–1504. <https://doi.org/10.1159/000492259>
- 126. Jiang Y, He R, Shi Y et al (2020) Plasma exosomes protect against cerebral ischemia/reperfusion injury via exosomal HSP70 mediated suppression of ROS. Life Sci 256:117987. [https://doi.](https://doi.org/10.1016/j.lfs.2020.117987) [org/10.1016/j.lfs.2020.117987](https://doi.org/10.1016/j.lfs.2020.117987)
- 127. Li F, Gong X, Yang B (2021) Geranylgeranylacetone ameliorated ischemia/reperfusion induced-blood brain barrier breakdown through HSP70-dependent anti-apoptosis efect. Am J Transl Res 13(1):102–114
- 128. Yang D, Ma L, Wang P et al (2019) Normobaric oxygen inhibits AQP4 and NHE1 expression in experimental focal ischemic

stroke. Int J Mol Med 43(3):1193–1202. [https://doi.org/10.3892/](https://doi.org/10.3892/ijmm.2018.4037) [ijmm.2018.4037](https://doi.org/10.3892/ijmm.2018.4037)

- 129. Pignataro G, Tortiglione A, Scorziello A et al (2004) Evidence for a protective role played by the Na+/Ca2+ exchanger in cerebral ischemia induced by middle cerebral artery occlusion in male rats. Neuropharmacology 46(3):439–448. [https://doi.org/](https://doi.org/10.1016/j.neuropharm.2003.09.015) [10.1016/j.neuropharm.2003.09.015](https://doi.org/10.1016/j.neuropharm.2003.09.015)
- 130. Jia M, Zhang Q, Guo X et al (2022) Na+/HCO3-co-transporters inhibitor S0859 attenuates global cerebral ischemia-reperfusion injury of the CA1 neurons in the Gerbil's hippocampus. CNS Neurol Disord. [https://doi.org/10.1016/j.neuropharm.2003.09.](https://doi.org/10.1016/j.neuropharm.2003.09.015) [015](https://doi.org/10.1016/j.neuropharm.2003.09.015)
- 131. Xue J, Zhou D, Yao H et al (2008) Role of transporters and ion channels in neuronal injury under hypoxia. Am J Physiol Regul Integr Comp Physiol 294(2):R451–R457. [https://doi.org/](https://doi.org/10.1152/ajpregu.00528.2007) [10.1152/ajpregu.00528.2007](https://doi.org/10.1152/ajpregu.00528.2007)
- 132. Rose CR, Karus C (2013) Two sides of the same coin: sodium homeostasis and signaling in astrocytes under physiological and pathophysiological conditions. Glia 61(8):1191–1205. [https://](https://doi.org/10.1002/glia.22492) doi.org/10.1002/glia.22492
- 133. Begum G, Song S, Wang S et al (2018) Selective knockout of astrocytic Na+/H+ exchanger isoform 1 reduces astrogliosis, BBB damage, infarction, and improves neurological function after ischemic stroke. Glia 66(1):126–144. [https://doi.org/10.](https://doi.org/10.1002/glia.23232) [1002/glia.23232](https://doi.org/10.1002/glia.23232)
- 134. Luo J, Chen H, Kintner DB et al (2005) Decreased neuronal death in Na+/H+ exchanger isoform 1-null mice after in vitro and in vivo ischemia. J Neurosci 25(49):11256–11268. [https://](https://doi.org/10.1523/JNEUROSCI.3271-05.2005) doi.org/10.1523/JNEUROSCI.3271-05.2005
- 135. Song S, Huang H, Guan X et al (2021) Activation of endothelial Wnt/β-catenin signaling by protective astrocytes repairs BBB damage in ischemic stroke. Prog Neurobiol 199:101963. [https://](https://doi.org/10.1016/j.pneurobio.2020.101963) doi.org/10.1016/j.pneurobio.2020.101963
- 136. Luo H, Gauthier M, Tan X et al (2018) Sodium transporters are involved in lithium infux in brain endothelial cells. Mol Pharm 15(7):2528–2538. [https://doi.org/10.1021/acs.molpharmaceut.](https://doi.org/10.1021/acs.molpharmaceut.8b00018) [8b00018](https://doi.org/10.1021/acs.molpharmaceut.8b00018)
- 137. Quednau BD, Nicoll D, Philipson KD (1997) Tissue specifcity and alternative splicing of the Na^+/Ca^{2+} exchanger isoforms NCX1, NCX2, and NCX3 in rat. Am J Physiol Cell Physiol 272(4):C1250–C1261. [https://doi.org/10.1152/ajpcell.1997.](https://doi.org/10.1152/ajpcell.1997.272.4.C1250) [272.4.C1250](https://doi.org/10.1152/ajpcell.1997.272.4.C1250)
- 138. Canitano A, Papa M, Boscia F et al (2002) Brain distribution of the Na^{+}/Ca^{2+} exchanger-encoding genes NCX1, NCX2, and NCX3 and their related proteins in the central nervous system. Ann NY Acad Sci 976(1):394–404. [https://doi.org/10.1111/j.](https://doi.org/10.1111/j.1749-6632.2002.tb04766.x) [1749-6632.2002.tb04766.x](https://doi.org/10.1111/j.1749-6632.2002.tb04766.x)
- 139. Boscia F, Gala R, Pignataro G et al (2006) Permanent focal brain ischemia induces isoform-dependent changes in the pattern of Na+/Ca2+ exchanger gene expression in the ischemic core, periinfarct area, and intact brain regions. J Cereb Blood Flow Metab 26(4):502–517. <https://doi.org/10.1038/sj.jcbfm.9600207>
- 140. Cerullo P, Brancaccio P, Anzilotti S et al (2018) Acute and longterm NCX activation reduces brain injury and restores behavioral functions in mice subjected to neonatal brain ischemia. Neuropharmacology 135:180–191. [https://doi.org/10.1016/j.neuro](https://doi.org/10.1016/j.neuropharm.2018.03.017) [pharm.2018.03.017](https://doi.org/10.1016/j.neuropharm.2018.03.017)
- 141. Propson NE, Roy ER, Litvinchuk A et al (2021) Endothelial C3a receptor mediates vascular infammation and blood-brain barrier permeability during aging. J Clin Invest. [https://doi.org/10.1172/](https://doi.org/10.1172/JCI140966) [JCI140966](https://doi.org/10.1172/JCI140966)
- 142. Theparambil SM, Hosford PS, Ruminot I et al (2020) Astrocytes regulate brain extracellular pH via a neuronal activity-dependent

bicarbonate shuttle. Nat Commun 11(1):5073. [https://doi.org/10.](https://doi.org/10.1038/s41467-020-18756-3) [1038/s41467-020-18756-3](https://doi.org/10.1038/s41467-020-18756-3)

- 143. Du L, Zahra A, Jia M et al (2021) Understanding the functional expression of Na+-Coupled SLC4 transporters in the renal and nervous systems: a review. Brain Sci. [https://doi.org/10.3390/](https://doi.org/10.3390/brainsci11101276) [brainsci11101276](https://doi.org/10.3390/brainsci11101276)
- 144. Sohn Y, Yoo K-Y, Park OK et al (2011) Na+/HCO3− cotransporter immunoreactivity changes in neurons and expresses in astrocytes in the gerbil hippocampal CA1 region after ischemia/reperfusion. Neurochem Res 36(12):2459–2469. [https://doi.org/10.1007/](https://doi.org/10.1007/s11064-011-0572-5) [s11064-011-0572-5](https://doi.org/10.1007/s11064-011-0572-5)
- 145. Yao H, Azad P, Zhao HW et al (2016) The Na⁺/HCO^{3−} co-transporter is protective during ischemia in astrocytes. Neuroscience 339:329–337. [https://doi.org/10.1016/j.neuroscience.2016.09.](https://doi.org/10.1016/j.neuroscience.2016.09.050) [050](https://doi.org/10.1016/j.neuroscience.2016.09.050)
- 146. Tavender TJ, Bulleid NJ (2010) Peroxiredoxin IV protects cells from oxidative stress by removing $H2O₂$ produced during disulphide formation. J Cell Sci 123(Pt 15):2672–2679. [https://doi.](https://doi.org/10.1242/jcs.067843) [org/10.1242/jcs.067843](https://doi.org/10.1242/jcs.067843)
- 147. Okado-Matsumoto A, Matsumoto A, Fujii J et al (2000) Peroxiredoxin IV is a secretable protein with heparin-binding properties under reduced conditions1. J Biochem 127(3):493–501. [https://](https://doi.org/10.1093/oxfordjournals.jbchem.a022632) doi.org/10.1093/oxfordjournals.jbchem.a022632
- 148. Ouyang T, Meng W, Li M et al (2020) Recent advances of the Hippo/YAP signaling pathway in brain development and glioma. Cell Mol Neurobiol 40(4):495–510. [https://doi.org/10.1007/](https://doi.org/10.1007/s10571-019-00762-9) [s10571-019-00762-9](https://doi.org/10.1007/s10571-019-00762-9)
- 149. Gong S, Ma H, Zheng F et al (2021) Inhibiting YAP in endothelial cells from entering the nucleus attenuates blood-brain barrier damage during ischemia-reperfusion injury. Front Pharmacol 12:777680. <https://doi.org/10.3389/fphar.2021.777680>
- 150. Yuan J, Li L, Yang Q et al (2021) Targeted treatment of ischemic stroke by bioactive nanoparticle-derived reactive oxygen species responsive and infammation-resolving nanotherapies. ACS Nano 15(10):16076–16094.<https://doi.org/10.1021/acsnano.1c04753>
- 151. Liu M, Xu Z, Wang L et al (2020) Cottonseed oil alleviates ischemic stroke injury by inhibiting the infammatory activation of microglia and astrocyte. J Neuroinfamm 17(1):1–15. [https://](https://doi.org/10.1186/s12974-020-01946-7) doi.org/10.1186/s12974-020-01946-7
- 152. Ji Y, Gao Q, Ma Y et al (2023) An MMP-9 exclusive neutralizing antibody attenuates blood-brain barrier breakdown in mice with stroke and stroke patient-derived MMP-9 activity. Pharmacol Res. <https://doi.org/10.1016/j.phrs.2023.106720>
- 153. Li Y, Zhang M, Li S et al (2023) Selective ischemic-hemisphere targeting Ginkgolide B liposomes with improved solubility and therapeutic efficacy for cerebral ischemia-reperfusion injury. Asian J Pharmaceut Sci. [https://doi.org/10.1016/j.ajps.2023.](https://doi.org/10.1016/j.ajps.2023.100783) [100783](https://doi.org/10.1016/j.ajps.2023.100783)
- 154. Long Y, Liu S, Wan J et al (2023) Brain targeted borneol-baicalin liposome improves blood-brain barrier integrity after cerebral ischemia-reperfusion injury via inhibiting HIF-1α/VEGF/eNOS/ NO signal pathway. Biomed Pharmacother 160:114240. [https://](https://doi.org/10.1016/j.biopha.2023.114240) doi.org/10.1016/j.biopha.2023.114240

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