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Unsaturated Analogues of the Neurotransmitter GABA: trans-4-Aminocrotonic, cis-4-Aminocrotonic and 4-Aminotetrolic Acids

Graham A. R. Johnston¹

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Abstract Analogues of the neurotransmitter GABA containing unsaturated bonds are restricted in the conformations they can attain. This review traces three such analogues from their synthesis to their use as neurochemicals. trans-4-Aminocrotonic acid was the first conformationally restricted analogue to be extensively studied. It acts like GABA across a range of macromolecules from receptors to transporters. It acts similarly to GABA on ionotropic receptors. cis-4-Aminocrotonic acid selectively bicuculline-insensitive GABA_C activates receptors. 4-Aminotetrolic acid, containing a triple bond, activates bicuculline-sensitive GABA_A receptors. These findings indicate that GABA activates GABAA receptors in extended conformations and GABA_C receptors in folded conformations. These and related analogues are important for the molecular modelling of ionotropic GABA receptors and to the development of new agents acting selectively on these receptors.

Keywords GABA \cdot *trans*-4-Aminocrotonic acid \cdot *cis*-4-Aminocrotonic acid \cdot 4-Aminotetrolic acid \cdot GABA_A receptors \cdot GABA_C receptors

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Graham A. R. Johnston grahamj@mail.usyd.edu.au

¹ Pharmacology, Faculty of Medicine, School of Medical Sciences, The University of Sydney, Sydney, NSW 2006, Australia

Introduction

The neurotransmitter, GABA, is a highly flexible molecule so it is possible that different conformations bind to its receptive macromolecules such as transporters, receptors or catabolising enzymes. Structural analogues of the neurotransmitter GABA that contain double or triple bonds are limited in the shapes they can attain in contrast to GABA being a highly flexible molecule. Such conformationally restricted analogues are useful neurochemicals with which to probe the different shapes that GABA may adopt in its interactions with receptive macromolecules. The simple unsaturated analogues-trans-4-aminocrotonic acid and 4-aminotetrolic acid-were the subject of Philip Beart's PhD thesis in 1972 (The Neurochemistry of Amino Acid Transmitters, Australian National University). Subsequently 4-aminotetrolic acid was converted into cis-4aminocrotonic acid and this led to the discovery of a new subtype of ionotropic GABA receptors (Fig. 1).

trans-4-Aminocrotonic Acid

trans-4-Aminocrotonic acid (TACA, *E*-4-aminobut-2-enoic acid) was the first conformationally restricted analogue of GABA to be studied extensively. The constraints of the double bond held the four carbon atoms in a plane, thus preventing free rotation around the C2-C3 bond in GABA.

TACA had been prepared in 1954 by dehydration of the readily available 4-amino-3-hydroxybutyric acid [1], and is now available commercially. Crystal structure analysis showed its similarity to GABA [2, 3] as did pKa measurements—GABA 4.04, 10.71; TACA 3.55, 9.46 [4, 5] with TACA being a somewhat stronger acid than GABA due to the electron withdrawing double bond.

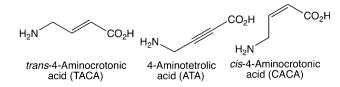


Fig. 1 Structures of simple unsaturated analogues of GABA

The action of TACA on nervous system function was first studied in 1960 by Purpura who described a GABA-like depressant action on neuronal firing [6]. Then Krnjevic and Phillis in 1963 described TACA as having an inhibitory activity comparable with that of GABA on neuronal firing in the mammalian cerebral cortex [7].

With the advent of agents to distinguish different subtypes of GABA postsynaptic receptors, TACA was found to have a more potent action than that of GABA on the ionotropic GABA_A and GABA_C receptors [8] and a much less potent action on the metabotropic GABA_B receptors [9].

TACA was also found to be a potent competitive inhibitor of GABA uptake into rat brain slices [5] and a weak substrate for GABA transaminase activity in rat brain extracts [10].

In an electroconvulsive threshold test in mice, TACA (at 15 and 25 mg/kg i.p.) significantly decreased the threshold indicating proconvulsant properties on systemic administration [11]. Such an effect was completely opposite to the authors' primary assumption and expectance. The authors suggested that TACA-induced stimulation of GABA_C receptors, highly expressed in superior colliculus, may lead to the proconvulsant action. Further experiments are needed to clarify this finding.

Numerous analogues of TACA have been prepared. Perhaps the most interesting are analogues substituted in the 2-position. The 2-fluoro analogue, *trans*-4-amino-2-fluorobut-2-enoic acid, was found to be a potent agonist (K_D =2.43 μ M) at human ρ 1 GABA_C receptors expressed in oocytes. In contrast, the 2-methyl analogue, *trans*-4-amino-2-methylbut-2-enoic acid, was found to be a moderately potent antagonist (IC₅₀=31.0 μ M and K_B=45.5 - μ M). These observations highlight the possibility that subtle structural substitutions may change an agonist into an antagonist when interacting with GABA_C receptors [12] (Fig. 2).

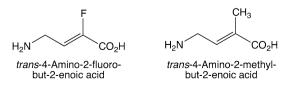


Fig. 2 Structures of some 2-substituted derivatives of 4-aminocrotonic acid

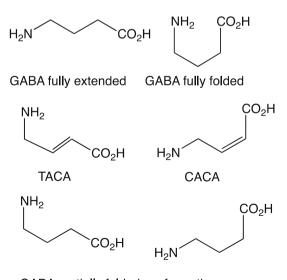
4-Aminotetrolic Acid

4-Aminotetrolic acid (ATA, 4-aminobut-2-ynoic acid) was first prepared by Philip Beart as part of his PhD studies in a hazardous multi-step synthesis [13]. The triple bond holds the four carbons atoms in a line thus representing a unique fully extended conformation of GABA with no possible folding. It is a much stronger acid than GABA with pKa values of 1.80 and 8.34 [14]. An X-ray crystal structure and molecular orbital calculations support the concept of ATA as a GABA analogue [15, 16]. It is not available commercially (Fig. 3).

Reduction of the triple bond in ATA and its derivatives with tritium has provided a convenient route to the preparation of GABA labelled to high specific activity due to the incorporation of four tritium atoms [17–20]. An improved synthesis of ATA has been reported [21].

ATA has an inhibitory action on the firing of a spinal interneuron in anesthetized cats 20–50 % as strong as that of GABA [13]. The inhibitory action of ATA, like that of GABA, could be reversibly antagonized by bicuculline whereas the glycine antagonist strychnine was ineffective as an antagonist. It has a similar action on bicuculline sensitive GABA receptors in the isolated superior cervical ganglion of the rat [22]. ATA has no activity on GABA_B receptors [13]. Like TACA, ATA was found to be a competitive inhibitor of GABA uptake into rat brain slices [5]. ATA is also an inhibitor of GABA transaminase activity without influence on glutamate decarboxylase [23].

We wrote at the time "The successful interaction of 4-aminotetrolic acid with bicuculline-sensitive postsynaptic receptors supports the view that GABA acts on these receptors in an extended rather than a folded conformation,



GABA partially folded conformations

Fig. 3 Some conformations of GABA, TACA and CACA

and further supports our original suggestion regarding the structural similarities of bicuculline, GABA and muscimol" [13]. The actions of bicuculline and muscimol on GABA receptors have been recently reviewed [24, 25].

cis-4-Aminocrotonic Acid

cis-4-Aminocrotonic acid (CACA, *Z*-4-aminobut-2-enoic acid) was first reported in 1975 and initially synthesised by partial hydrogenation of ATA by Philip Beart before undertaking postdoctoral research with Leslie Iversen at the University of Cambridge [8]. It was his last synthesis as a medicinal chemist before becoming a fully fledged neurochemist. Subsequently, an improved synthesis from a derivative of ATA was reported [17]. It has been available commercially since 1992. Radioactive CACA has been prepared by reduction of an acetylenic intermediate with tritium gas [18]. The resultant high specific activity preparation of [³H]CACA was found to be toxic producing burning sensations to the face, eyes and hands of personnel handling the preparation, thus limiting the extent of the binding studies that can be carried out.

CACA was particularly important as an analogue of GABA in a partially folded conformation and this led to the discovery of a new subtype of ionotropic GABA receptor. CACA is close to TACA in pKa values (3.93 and 9.84) but has very different actions on GABA systems [8].

Light slowly converts CACA into the more thermodynamically stable TACA. Contamination of some samples of CACA with up to 0.1 % TACA as detected by nuclear magnetic spectroscopy can complicate interpretation of the effects of high concentrations of CACA, given the relatively high potency of TACA in many test systems compared to that of pure CACA [26].

CACA is a weak substrate for a transporter that transports GABA, β -alanine and nipecotic acid in glial cells isolated from guinea-pig retina [27]. It stimulates the release of GABA and β -alanine from slices of rat spinal cord and cerebellum [28]. These studies are consistent with the idea that CACA, β -alanine, nipecotic acid and GABA are substrates for a common transporter, that may be related to the GAT-3 transport protein cloned from rat CNS (Fig. 4).

In terms of shape, CACA represents more folded conformations of GABA while TACA represents more extended conformations. Importantly GABA and TACA can adopt similar partially folded conformations, but CACA cannot match the more extended conformations of GABA.

Although CACA has an inhibitory action of the firing of spinal neurones, this action is not blocked by the $GABA_A$ antagonist bicuculline under conditions whereby TACA had a potent bicuculline-sensitive action [8]. The inhibitory action of CACA is not blocked by the glycine antagonist

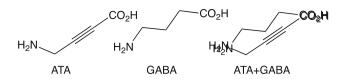


Fig. 4 Overlapping structures of ATA and GABA

strychnine thus ruling out an interaction with glycine receptors. Unlike the $GABA_B$ agonist baclofen, CACA is equally potent as a inhibitor of the firing of Renshaw cells and spinal interneurones. These results indicated that CACA is not an agonist for $GABA_A$ or $GABA_B$ receptors on spinal neurones [8].

CACA was described as a 'disturber of the peace' by Krogsgaard-Larsen and colleagues [29]. The concept of a third major class of GABA receptors arose in 1984 from studies on the lack of effect of CACA on the binding of radioactive baclofen to rat cerebellar membranes: "folded analogs of GABA may interact with a class of binding site (GABA_C?) insensitive to (-)-baclofen and bicuculline" [30].

Peace was restored in 1993 when, following the release of CACA commercially, three groups described the action of CACA on bicuculline-insensitive, baclofen-insensitive GABA receptors in the retina and receptors cloned from the retina [31–33]. These became known as GABA_C receptors made up of ρ -subunits [26, 34], although he nomenclature of these ionotropic GABA receptors remains unsettled. This has been discussed in a comprehensive review of ionotropic GABA receptors [35]. It is important to distinguish between GABA_A and GABA_C receptors, as these receptors appear to have opposing roles in nervous system function [36, 37].

CACA is a partial agonist at human recombinant $\rho 1$ and $\rho 2$ GABA_C receptors [26, 34]. It is much more selective as a GABA_C receptor ligand than the more potent TACA, which interacts strongly with a variety of macromolecules that recognize GABA. Unlike TACA, CACA is, at best, a very weak GABA_A receptor agonist and is neither a substrate for, nor an inhibitor of, GABA transaminase in extracts of rat brain mitochondria. In addition, it does not influence the activity of glutamate decarboxylase in rat brain extracts [8]. The *cis*-double bond in CACA appears to restrict the conformations available to CACA to those that interact with GABA_C receptors and GAT3 transporters. CACA is tenfold weaker as a substrate for the transporter than as a partial agonist for GABA_C receptors (Fig. 5).

Used in conjunction with TACA and the GABA_C receptor antagonist TPMPA [38, 39], CACA has been used to aid in the characterisation of GABA_C receptors in the retina [31, 33], hippocampus [40], colliculus [41, 42], pituitary [43], pelvic ganglia [44], and the gut [45–47]. CACA also has a peripheral anti-nociceptive effect in a paw pressure test [48] that may involve GABA_C receptors.

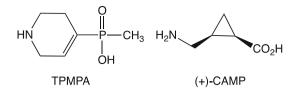


Fig. 5 Structures of the selective GABA_c antagonist TPMPA and the selective GABA_c agonist (+)-CAMP

There is evidence for CACA activating receptors other than purely ρ -containing GABA_C receptors. It activates $\alpha 6$ subunit-containing GABA_A receptors but not GABA_C receptors in cerebellar granule cells in a bicuculline-sensitive, TPMPA-insensitive manner [49]. In cerebellar Purkinje cells, CACA has a mixed action activating both bicuculline- and TPMPA-sensitive receptors [50]. CACA has been used to provide evidence for the co-assembly of GABA_A and GABA_C receptor subunits in the brain stem and hippocampus [51, 52].

Conclusion

The conformationally restricted unsaturated analogues of GABA introduced by Philip Beart and his colleagues continue to provide valuable information on the function of ionotropic GABA receptors. ATA shows selectivity for GABA_A over GABA_C receptors while CACA shows selectivity for GABA_C over GABA_A receptors. TACA activates both classes of ionotropic receptor. The structures of ATA, CACA and TACA are important for the molecular modelling studies of GABAA and GABAC receptors that aid in the discovery of drugs interacting with ionotropic GABA receptors [53, 54]. Unfortunately, the synthesis of pure CACA uncontaminated by TACA has remained difficult, limiting access to N-substituted analogues of CACA. Additionally, the unsaturated bond in these compounds alters the acidity in comparison to GABA. This can be avoided by using ring structures to restrict conformational freedom. Indeed the cyclopropane analogues of GABA have significant actions of GABA_C receptors with cis-2-(aminomethyl)cyclopropanecarboxylic acid ((+)-CAMP) being a potent and selective full agonist at GABA_C receptors [55, 56]. Neurochemicals such as these are important to the development of new agents acting selectively on ionotropic GABA receptors.

Acknowledgments It is a pleasure to contribute to this special issue honouring Philip Beart who was my first PhD student. We have maintained our collaboration over many years most recently on aspects of the history of the International Society for Neurochemistry. I am grateful to Philip Beart, Jane Hanrahan and Ken Mewett for their helpful comments on the manuscript.

References

- Musashi A (1954) The synthesis of γ-aminocrotonic acid and γaminobutyric acid. Hoppe-Seyler's Z Physiol Chem 297:71–73
- Tomita KI (1971) Crystal data and some structural features of γaminobutyric acid, 3-aminopropane sulfonic acid and their derivatives. Tetrahedron Lett 12:2587–2588
- Jones GP, Pauling PJ (1975) Crystal and molecular structure of trans-4-aminocrotonic acid. J Chem Soc Perkin Trans 2:1059–1062
- King EJ (1954) The thermodynamics of ionization of amino acids.1. The ionization constants of gamma-aminobutyric acid. J Am Chem Soc 76:1006–1008
- Beart PM, Johnston GAR, Uhr ML (1972) Competitive inhibition of GABA uptake in rat brain slices by some GABA analogues of restricted conformation. J Neurochem 19:1855–1861
- Purpura DP (1960) Pharmacological actions of ω-amino acid drugs on different cortical synaptic organizations. In: Inhibition nervous system gamma-Amino-butyric Acid, Proc Intern Symp, Duarte, Calif vol 1959, pp. 495–514
- Krnjevic K, Phillis JW (1963) Iontophoretic studies of neurons in the mammalian cerebral cortex. J Physiol 165:274–304
- Johnston GAR, Curtis DR, Beart PM, Game CJA, McCulloch RM, Twitchin B (1975) *cis-* and *trans*-4-Aminocrotonic acid as GABA analogues of restricted conformation. J Neurochem 24:157–160
- Falch E, Hedegaard A, Nielsen L, Jensen BR, Hjeds H, Krogsgaard-Larsen P (1986) Comparative stereostructure-activity studies on GABA-A and GABA-B receptor-sites and GABA uptake using rat-brain membrane preparations. J Neurochem 47:898–903
- Beart PM, Johnston GAR (1973) Transamination of analogues of γ-aminobutyric acid by extracts of rat brain mitochondria. Brain Res 49:459–462
- Borowicz KK, Zadrozniak M, Czuczwar SJ (2005) GABA_A and GABA_C receptors, shows a proconvulsant action in the electroconvulsive threshold test in mice. Pharmacol Rep PR 57:121–123
- Chebib M, Vandenberg RJ, Johnston GAR (1997) Analogues of γ-aminobutyric acid (GABA) and *trans*-4-aminocrotonic acid (TACA) substituted in the 2 position as GABA_C receptor antagonists. Br J Pharmacol 122:1551–1560
- Beart PM, Curtis DR, Johnston GAR (1971) 4-Aminotetrolic acid: new conformational-restricted analogue of γ-aminobutyric acid. Nature 234:80–81
- Beart PM, Johnston GAR (1972) Acetylenic analogues of γaminobutyric acid. Aust J Chem 25:1359–1361
- Jones GP, Pauling PJ (1976) Crystal and molecular structure of 4-aminotetrolic acid (4-amino-but-2-ynoic acid). J Chem Soc Perkin Trans 2:32–33
- Warner D, Borthwick PW, Stewart EG (1975) Complete neglect of differential overlap/2 molecular orbital studies on the electronic structure and conformational modes of 4-aminotetrolic acid. J Mol Struct 25:397–402
- Allan RD, Johnston GAR (1985) Synthesis of analogues of GABA. XIII An alternative route to (Z)-4-aminocrotonic acid. Aust J Chem 38:1647–1650
- Duke RK, Allan RD, Drew CA, Johnston GAR, Mewett KN, Long MA, Than C (1993) The preparation of tritiated E-4-aminobut-2-enoic and Z-4-aminobut-2-enoic acids, conformationally restricted analogues of the inhibitory neurotransmitter 4-aminobutanoic acid (GABA). J Label Compd Radiopharm 33:527–540
- Ahern DG, Laseter AG, Filer CN (2003) The synthesis and characterization of [2,3-3H] γ-aminobutyric acid via 4-aminotetrolic acid at high specific activity. Appl Radiat Isot 58:477–479

- Filer CN (2010) GABAergic radioligands labelled with tritium. J Label Compd Radiopharm 53:120–129
- Ahern DG, Laseter AG, Filer CN (2002) An improved synthesis of 4-aminotetrolic acid. Synth Commun 32:665–667
- 22. Bowery NG, Jones GP (1976) A comparison of γ-aminobutyric acid and the semi-rigid analogues 4-aminotetrolic acid, 4-aminocrotonic acid and imidazole-4-acetic acid on the isolated superior cervical ganglion of the rat. Br J Pharmacol 56:323–330
- Beart PM, Johnston GAR (1972) Bicuculline and GABAmetabolising enzymes. Brain Res 38:226–227
- 24. Johnston GAR (2013) Advantages of an antagonist: bicuculline and other GABA antagonists. Br J Pharmacol 169:328–336
- Johnston GAR (2014) Muscimol as an ionotropic GABA receptor agonist. Neurochem Res 39:148–159
- Johnston GAR (1996) GABA_C receptors: relatively simple transmitter-gated ion channels? Trends Pharmacol Sci 17:319–323
- Biedermann B, Eberhardt W, Reichelt W (1994) GABA uptake into isolated retinal Muller glial cells of the guinea-pig detected electrophysiologically. NeuroReport 5:438–440
- Chebib M, Johnston GAR (1997) Stimulation of [3H] GABA and beta-[3H] alanine release from rat brain slices by *cis*-4aminocrotonic acid. J Neurochem 68:786–794
- Krogsgaard-Larsen P, Frølund B, Ebert B (1997) GABA-A receptor agonists, partial agonists, and antagonists. In: Enna SJ, Bowery NG (eds) The GABA receptors, 2nd edn. Humana, Totowa, pp 37–81
- Drew CA, Johnston GAR, Weatherby RP (1984) Bicucullineinsensitive GABA receptors: studies on the binding of (-)-baclofen to rat cerebellar membranes. Neurosci Lett 52:317–321
- Qian H, Dowling JE (1993) Novel GABA responses from roddriven retinal horizontal cells. Nature 361:162–164
- Kusama T, Spivak CE, Whiting P, Dawson VL, Schaeffer JC, Uhl GR (1993) Pharmacology of GABA ρ1 and GABA a/b receptors expressed in *Xenopus* oocytes and COS cells. Br J Pharmacol 109:200–206
- Feigenspan A, Wassle H, Bormann J (1993) Pharmacology of GABA receptor Cl⁻ channels in rat retinal bipolar cells. Nature 361:159–162
- Bormann J, Feigenspan A (1995) GABA_C receptors. Trends Neurosci 18:515–519
- Chebib M, Johnston GAR (2000) GABA-activated ligand gated ion channels: medicinal chemistry and molecular biology. J Med Chem 43:1427–1447
- 36. Pasternack M, Boller M, Pau B, Schmidt M (1999) GABA_A and GABA_C receptors have contrasting effects on excitability in superior colliculus. J Neurophysiol 82:2020–2023
- Gibbs ME, Johnston GAR (2005) Opposing roles for GABA_A and GABA_C receptors in short-term memory formation in young chicks. Neuroscience 131:567–576
- 38. Johnston GAR, Chebib M, Hanrahan JR, Mewett KN (2010) Neurochemicals for the investigation of $GABA_C$ receptors. Neurochem Res 35:1970–1977
- Murata Y, Woodward RM, Miledi R, Overman LE (1996) The first selective antagonist for a GABA_C receptor. Bioorg Med Chem Lett 6:2073–2076
- Alakuijala A, Alakuijala J, Pasternack M (2006) Evidence for a functional role of GABA receptors in the rat mature hippocampus. Eur J Neurosci 23:514–520

- Edwards MD, White AM, Platt B (2002) Characterisation of rat superficial superior colliculus neurones: firing properties and sensitivity to GABA. Neuroscience 110:93–104
- Kirischuk S, Akyeli J, Iosub R, Grantyn R (2003) Pre- and postsynaptic contribution of GABA_C receptors to GABAergic synaptic transmission in rat collicular slices and cultures. Eur J Neurosci 18:752–758
- Gamel-Didelon K, Kunz L, Fohr KJ, Gratzl M, Mayerhofer A (2003) Molecular and physiological evidence for functional γaminobutyric acid GABA_C receptors in growth hormone-secreting cells. J Biol Chem 278:20192–20195
- Munakata Y, Tsurusaki M, Akasu T (1998) GABA_C receptors mediate slow membrane potentials in neurons of the rat major pelvic ganglia. Kurume Med J 45:295–299
- 45. Jansen A, Hoepfner M, Herzig KH, Riecken EO, Scherubl H (2000) GABA_C receptors in neuroendocrine gut cells: a new GABA-binding site in the gut. Pflugers Archiv 441:294–300
- 46. Zizzo MG, Mule F, Serio R (2007) Functional evidence for GABA as modulator of the contractility of the longitudinal muscle in mouse duodenum: role of $GABA_A$ and $GABA_C$ receptors. Neuropharmacology 52:1685–1690
- Reis HJ, Vanden Berghe P, Romano-Silva MA, Smith TK (2006) GABA-induced calcium signaling in cultured enteric neurons is reinforced by activation of cholinergic pathways. Neuroscience 139:485–494
- Reis GM, Duarte ID (2007) Involvement of chloride channel coupled GABA_C receptors in the peripheral antinociceptive effect induced by GABA_C receptor agonist *cis*-4-aminocrotonic acid. Life Sci 80:1268–1273
- Wall MJ (2001) *cis*-4-Amino-crotonic acid activates alpha 6 subunit-containing GABA_A but not GABA_C receptors in granule cells of adult rat cerebellar slices. Neurosci Lett 316:37–40
- Harvey VL, Duguid IC, Krasel C, Stephens GJ (2006) Evidence that GABA ρ subunits contribute to functional ionotropic GABA receptors in mouse cerebellar Purkinje cells. J Physiol 577:127–139
- Milligan CJ, Buckley NJ, Garret M, Deuchars J, Deuchars SA (2004) Evidence for inhibition mediated by coassembly of GABA_C and GABA_C receptor subunits in native central neurons. J Neurosci 24:7241–7250
- Hartmann K, Stief F, Draguhn A, Frahm C (2004) Ionotropic GABA receptors with mixed pharmacological properties of GABA_A and GABA_C receptors. Eur J Pharmacol 497:139–146
- Bergmann R, Kongsbak K, Sorensen PL, Sander T, Balle T (2013) A unified model of the GABA_A receptor comprising agonist and benzodiazepine binding sites. PLoS One [Electronic Resource] 8:e52323
- 54. Abdel-Halim H, Hanrahan JR, Hibbs DE, Johnston GAR, Chebib M (2008) A molecular basis for agonist and antagonist actions at GABA_C receptors. Chem Biol Drug Des 71:306–327
- 55. Duke RK, Chebib M, Balcar VJ, Allan RD, Mewett KN, Johnston GAR (2000) (+)- and (-)-*cis*-2-Aminomethylcyclopropanecarboxy acids show opposite pharmacology at recombinant $\rho(1)$ and $\rho(2)$ GABA_C receptors. J Neurochem 75:2602–2610
- 56. Ng CK, Kim HL, Gavande N, Yamamoto I, Kumar RJ, Mewett KN, Johnston GAR, Hanrahan JR, Chebib M (2011) Medicinal chemistry of ρ GABA_C receptors. Fut Med Chem 3:197–209