# ORIGINAL PAPER

# Effect of Hydrogen Sulfide on Cyclic AMP Production in Isolated Bovine and Porcine Neural Retinae

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Abstract Hydrogen sulfide (H<sub>2</sub>S) has been reported to exert pharmacological effects on neural and non-neural tissues from several mammalian species. In the present study, we examined the role of the intracellular messenger, cyclic AMP in retinal response to H<sub>2</sub>S donors, sodium hydrosulfide (NaHS) and sodium sulfide (Na<sub>2</sub>S) in cows and pigs. Isolated bovine and porcine neural retinae were incubated in oxygenated Krebs buffer solution prior to exposure to varying concentrations of NaHS, Na<sub>2</sub>S or the diterpene activator of adenylate cyclase, forskolin. After incubation at different time intervals, tissue homogenates were prepared for cyclic AMP assay using a well established methodology. In isolated bovine and porcine retinae, the combination of both phosphodiesterase inhibitor, IBMX (2 mM) and forskolin (10 µM) produced a synergistic increase (P < 0.001) in cyclic AMP concentrations over basal levels. NaHS (10 nM-100 µM) produced a

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time-dependent increase in cyclic AMP concentrations over basal levels which reached a maximum at 20 min in both bovine and porcine retinae. At this time point, both NaHS and Na<sub>2</sub>S (10 nM-100 µM) caused a significant (P < 0.05) dose-dependent increase in cyclic AMP levels in bovine and porcine retinae. For instance, NaHS (100 nM) elicited a four-fold and three-fold increase in cyclic AMP concentrations in bovine and porcine retinae respectively whilst higher concentrations of Na2S  $(100 \ \mu M)$  produced a much lesser effect in both species. In bovine and porcine retinae, the effects caused by forskolin (10 µM) on cyclic AMP production were not potentiated by addition of low or high concentrations of both NaHS and Na<sub>2</sub>S. We conclude that H<sub>2</sub>S donors can increase cyclic AMP production in isolated neural retinae from cows and pigs. Bovine retina appears to be more sensitive to the stimulatory effect of H<sub>2</sub>S donors on cyclic nucleotide production than its porcine counterpart indicating that species differences exist in the magnitude of this response. Furthermore, effects produced by forskolin on cyclic AMP formation were not additive with those elicited by H<sub>2</sub>S donors suggesting that these agents may share a common mechanism in their action on the adenylyl cyclase pathway.

**Keywords** Hydrogen sulfide · Cyclic AMP · Retina · Sodium sulfide · Sodium hydrosulfide

#### Abbreviations

$H_2S$	Hydrogen sulfide
Na <sub>2</sub> S	Sodium sulfide
NaHS	Sodium hydrosulfide
Cyclic AMP	Adenosine 3',5' cyclic monophosphate
PDE	Phosphodiesterase
COX	Cyclooxygenase

#### Introduction

In the past decade, interest in the non-toxic actions of hydrogen sulfide (H<sub>2</sub>S) has led to several studies aimed at elucidating its potential physiological and pathological effects in mammalian tissues. H<sub>2</sub>S, a colorless gas characterized by its pungent odor (commonly described as the smell of rotten eggs) has been known for decades only as an environmental pollutant with a broad toxicity spectrum [1, 2]. Recently, there is evidence that this "toxic" gas can serve as an endogenous neurotransmitter, a smooth muscle relaxant and a regulator of immune reactions [1-4]. Endogenous H<sub>2</sub>S is generated in mammalian tissues from L-cysteine, a reaction catalyzed by two endogenous pyridoxal-5'-phosphate dependent-enzymes, cystathionine- $\beta$ -synthase (CBS) and cystathionine- $\gamma$ -lyase (CSE) [5–8], and is a component of foodstuffs, human feces [9, 10] and a product of bacterial and helminth metabolism [11, 12].

In the eye, the presence of the enzymes (CBS and CSE) responsible for H<sub>2</sub>S biosynthesis has recently been reported in ocular tissues [13, 14]. Both CBS and CSE were shown to be highly localized in the retina indicating the presence of a functional trans-sulfuration pathway and thus suggesting a potential role of H<sub>2</sub>S as a gaseous neuromodulator in this tissue. In the eye, toxicity associated with exposure to lethal concentrations of H<sub>2</sub>S is mostly at the mucus membrane level leading to keratoconjunctivitis [15]. Furthermore, studies have reported that deficiency of CBS is often associated with many eye disorders including retinal degeneration, retinal detachment, optical atrophy and glaucoma [16, 17]. Evidence from our laboratory demonstrate that H<sub>2</sub>S (using sodium hydrosulfide, NaHS and sodium sulfide, Na<sub>2</sub>S as donors) can induce pharmacological effects in mammalian ocular tissues [18–20]. We found that both NaHS and Na<sub>2</sub>S relaxed pre-contracted isolated porcine irides [18] and inhibited sympathetic and glutamatergic neurotransmission from isolated porcine and bovine anterior uvea and retina, an effect that was shown to be dependent on intramural biosynthesis of this gas [19, 20].

In the cardiovascular system, the pharmacological effects of  $H_2S$  has been reported to be mediated largely by ATP-sensitive potassium channels ( $K_{ATP}$ ) [21, 22]. However, in neuronal tissues  $H_2S$  has been shown to stimulate the production of adenosine 3',5' cyclic monophosphate (cyclic AMP) and thus activate cyclic AMP-dependent processes [23]. Cyclic AMP is an intracellular second messenger that plays an essential role in numerous neuronal functions such as cell survival, axon regeneration and modulation of axonal guidance [24, 25]. Taken together, it appears that both  $K_{ATP}$  channels and cyclic AMP may mediate the pharmacological actions of  $H_2S$  in mammalian tissues. In the present study, we examined the effect of  $H_2S$ 

(using NaHS and Na<sub>2</sub>S as donors) on cyclic AMP production in isolated bovine and porcine neural retinae. Parts of the data reported in this paper have been communicated in an abstract form [26].

# Methods

#### Chemicals

NaHS, Na<sub>2</sub>S, Ethylenediamine tetra-acetic acid solution (EDTA), Isobutylmethylxanthine (IBMX) and Forskolin were all purchased from Sigma–Aldrich, St. Louis, MO. Flurbiprofen was procured from Cayman Chemicals, Ann Arbor, MI. Stock solutions of IBMX, flurbiprofen and forskolin were dissolved in Dimethylsulfoxide (DMSO) whilst solutions of NaHS and Na<sub>2</sub>S were prepared with distilled water. At pH 7.4 the concentration of H<sub>2</sub>S solution is relatively stable [22, 27, 28]. All test agents were freshly prepared immediately before use on the day of the study.

### Preparation of Retinal Tissues

Bovine and Porcine eyes were obtained from a local slaughterhouse (Fisher Ham & Meat Co., Houston, Texas) and transported to the laboratory on ice following decapitation of animals. The eyeballs were enucleated, and the anterior chambers were carefully removed. The resulting eye cups were inverted and placed in fresh oxygenated Krebs solution containing the following composition (mM): potassium chloride, 4.8; sodium chloride, 118; calcium chloride, 2.3, potassium dihydrogen phosphate, 1.2; sodium bicarbonate, 25; magnesium sulfate, 2.0; and dextrose, 10. The neural retinae were detached by gently movement and incubated immediately in freshly prepared oxygenated Krebs solution (pH 7.4) containing the cyclooxygenase (COX) inhibitor, flurbiprofen (3 µM), with or without the presence of the cyclic nucleotide phosphodiesterase (PDE) inhibitor, IBMX (2 mM) for 30 min. Time elapsed between animal sacrifice and retina preparation was less than 24 h.

#### Cyclic AMP Assay

The methodology employed for cyclic AMP assay was essentially the same as reported by [29] with some modifications. Immediately following the 30 min incubation, the isolated bovine or porcine neural retinae were transferred to 2 ml of freshly prepared Krebs solution (pH 7.4) containing flurbiprofen (3  $\mu$ M) with or without IBMX (2 mM). In studies, were we examined the role of PDE on metabolism of cyclic AMP, isolated neural retinae without prior incubation with IBMX, were exposed

to either IBMX (2 mM), forskolin (10 uM) or a combination of both at the desired time of 20 min (this time scale was chosen based on results from our preliminary experiments that showed that maximal effect of agonists on cyclic AMP was achieved at this time). All subsequent experiments on the effect of H<sub>2</sub>S donors and forskolin were performed in the presence of IBMX (2 mM). Tissues were treated with varying concentrations of, H<sub>2</sub>S donors (NaHS, Na<sub>2</sub>S) or the intracellular cyclic AMP-elevating agent forskolin (10 µM) and exposed for 20 min. Control tissues were exposed to an appropriate volume of vehicle (0.9% saline) for the same time period. In experiments were we examined the combined effect of forskolin and H2S donors on cyclic AMP production, isolated neural retinae were exposed to forskolin (10 µM) for 20 min before the addition of the H<sub>2</sub>S donors. After an additional 10 min the reaction was terminated by addition of 4 mM ice-cold EDTA. The tissue homogenates were boiled for 20 min and then centrifuged at 3,000 rpm for 10 min. Pellets obtained were dissolved in 1 N NaOH at 60°C for protein determination by method of Bradford and aliquots of the supernatant were employed for measurement of cyclic AMP content using a cyclic AMP enzyme immunoassay kit purchased from Cayman Chemicals, Ann Arbor, Michigan.

#### Data Analysis

Results are expressed as pmol/µg of protein. Values given are arithmetic means  $\pm$  SEM. The means were determined from two separate experiments performed in triplicates. Significance of differences between control and treatment groups were evaluated using one-way analysis of variance (ANOVA) followed by Newman–Keuls comparison test. Drug treatment, time and interaction between drug treatment and time were assessed by two-way ANOVA followed by post-hoc Bonferroni test (Graph Pad Prism Software, San Diego, CA). A *P* value of <0.05 was considered as statistically significant.

### Results

# Effects of Activation of Adenylyl Cyclase and Inhibition of Phosphodiesterase on Cyclic AMP Levels in Neural Retina

Isobutylmethylxanthine is a phosphodiesterase (PDE) inhibitor that prevents the breakdown of accumulated intracellular cyclic AMP. In this study, we considered the possibility that PDE inhibition may contribute to the responses observed with forskolin (diterpene activator of

adenylyl cyclase) in both bovine and porcine isolated neural retinae. Furthermore, PDE inhibition could also allow optimization of the yield of cyclic AMP in response to the H<sub>2</sub>S donors. The PDE inhibitor, IBMX (2 mM) caused a two-three fold increase in basal cyclic AMP concentrations in bovine (Fig. 1a) and porcine (Fig. 1b) retinae respectively. Tissues stimulated with forskolin (10  $\mu$ M) alone did not have any significant effect on cyclic AMP concentrations when compared to basal levels in both species (Fig. 1a, b). In the presence of IBMX, responses elicited by forskolin were enhanced significantly (*P* < 0.001) in both bovine (Fig. 1a) and porcine (Fig. 1b) retinae.



Fig. 1 Effects of PDE inhibition on cyclic AMP levels in isolated bovine (a) and porcine (b) retina: control, in the presence of IBMX (2 mM) and Forskolin (10  $\mu$ M). *Vertical bars* represent means  $\pm$  SEM; n = 6. \*\*P < 0.001, \*P < 0.01 significantly different from controls; <sup>@</sup>P < 0.01 significantly different from IBMX-Forskolin treated groups

# Effects of NaHS on Cyclic AMP Levels in Neural Retina

Since there is evidence that exogenous H<sub>2</sub>S increases the production of cyclic AMP in neurons [30], we investigated the effects of the H<sub>2</sub>S donor, NaHS on cyclic AMP formation in bovine and porcine neural retinae. Based on data obtained in studies described in Fig. 1, all subsequent experiments on the effect of H<sub>2</sub>S donors were performed in the presence of IBMX (2 mM). As illustrated in Fig. 2, concentrations of 10 nM, 1 µM or 100 µM of NaHS produced a time-dependent significant (P < 0.05) increase in cyclic AMP levels over basal concentrations in both bovine (Fig. 2a) and porcine (Fig. 2b) retinae which reached a maximum at 20 min. In bovine retina, NaHS (100 uM) caused a five-fold and six-fold increase in cyclic AMP concentrations over basal levels at 5- and 20-min time intervals, respectively (Fig. 2a). In contrast, in porcine retina, NaHS (100 µM) elicited a two-fold and three-fold increase in cyclic AMP concentrations over basal levels at 5- and 20 min time intervals, respectively (Fig. 2b).

In subsequent experiments, we examined the effect of different concentrations of NaHS on cyclic AMP production using a 20 min incubation time for both bovine and porcine neural retinae. NaHS (10 nM–100  $\mu$ M) produced a concentration-dependent significant (P < 0.05) increase in cyclic AMP levels reaching a maximal effect at 100 nM (Fig. 3a). At this concentration, NaHS (100 nM) caused a four-fold increase in cyclic AMP concentrations above basal levels in bovine retina. NaHS (10 nM–100  $\mu$ M) also caused concentration-dependent significant (P < 0.05) increases in cyclic AMP levels in porcine retina with a maximal effect observed at 1  $\mu$ M (Fig. 3b). In porcine retina, NaHS (1  $\mu$ M) produced a three-fold increase in cyclic AMP concentrations above basal levels. The diterpene activator of adenylate cyclase, forskolin (10  $\mu$ M) also



Fig. 2 Time-dependent effect of NaHS on cyclic AMP in isolated bovine (a) and porcine (b) retina: control and in the presence of NaHS (10 nM–100  $\mu$ M). *Vertical bars* represent means  $\pm$  SEM; n = 6. \*\*P < 0.001, \*P < 0.01 significantly different from controls; <sup>†</sup>P < 0.01 significantly different among drug-treated groups (10 nM). Two-way ANOVA: In bovine retina (a) time, P < 0.004;

produced a significant (P < 0.001) increase in cyclic AMP concentrations above basal levels (Fig. 3a, b).

# Effects of Na<sub>2</sub>S on Cyclic AMP Levels in Neural Retina

We next examined the effect of another H<sub>2</sub>S donor, Na<sub>2</sub>S on cyclic nucleotide production in bovine and porcine neural retinae. At 20 min of incubation, Na<sub>2</sub>S (10 nM–100  $\mu$ M) elicited a concentration-dependent increase in cyclic AMP levels for both bovine and porcine neural retinae (Fig. 4a, b). At a concentration of 100  $\mu$ M, Na<sub>2</sub>S caused a two-one halffold increase in cyclic AMP concentration over basal levels in both bovine (Fig. 4a) and porcine (Fig. 4b) retinae. Upon stimulation with the positive control, forskolin (10  $\mu$ M), both bovine and porcine retinae responded with a significant (P < 0.001) increase in cyclic AMP concentrations over basal levels (Fig. 4a, b).

# Effects of H<sub>2</sub>S Donors on Activation of Adenylyl Cyclase by Forskolin

We examined the combined effect of forskolin and  $H_2S$  donors on cyclic AMP production in both bovine and porcine retinae. In bovine retina, submaximal (10 nM) and maximal (100 nM) concentrations of NaHS were examined on forskolin (10  $\mu$ M) stimulated cyclic AMP production (Fig. 5a). The response observed to forskolin was not enhanced by the presence of both low and high concentrations of NaHS (Fig. 5a). Likewise, in porcine retina, the effects caused by forskolin (10  $\mu$ M) on cyclic AMP formation were not enhanced by addition of submaximal (10 nM) and maximal (1  $\mu$ M) concentrations of NaHS (Fig. 5b).

We next investigated the combined effect of forskolin and Na<sub>2</sub>S on cyclic AMP formation in both bovine and



drug treatment, P < 0.002. For both groups, treatment and interaction (treatment × time of study) = NS; P < 0.280. In porcine retina (b) time, P < 0.001; drug treatment, P < 0.001. For both groups, treatment and interaction (treatment × time of study) = significant; P < 0.01



**Fig. 3** Concentration-dependent effect of NaHS on cyclic AMP in isolated bovine (**a**) and porcine (**b**) retina: control, in the presence of NaHS (10 nM–100  $\mu$ M) and Forskolin (10  $\mu$ M). *Vertical bars* represent means  $\pm$  SEM; n = 6. \*\*P < 0.001, \*P < 0.01 significantly different from controls; <sup>@</sup>P < 0.01 significantly different from Forskolin treated groups; <sup>†</sup>P < 0.01 significantly different among drug-treated groups (10 nM)

porcine retinae. In bovine retina, the effects elicited by forskolin (10  $\mu$ M) on cyclic AMP production were not potentiated by addition of submaximal (10 nM) and maximal (1  $\mu$ M) concentrations of Na<sub>2</sub>S (Fig. 6a). Similarly, in porcine retina, the effects produced by forskolin (10  $\mu$ M) on cyclic AMP formation were not enhanced by submaximal (1  $\mu$ M) and maximal (100  $\mu$ M) concentrations of Na<sub>2</sub>S (Fig. 6b).

### Discussion

In the last two decades, there has been a surge of interest in the biological effects of  $H_2S$ , a gas that is now deemed to serve as a gaseous transmitter along with nitric oxide and



**Fig. 4** Concentration-dependent effect of Na<sub>2</sub>S on cyclic AMP in isolated bovine (**a**) and porcine (**b**) retina: control, in the presence of Na<sub>2</sub>S (10 nM–100  $\mu$ M) and Forskolin (10  $\mu$ M). *Vertical bars* represent means  $\pm$  SEM; n = 6. \*\*P < 0.001, \*P < 0.01 significantly different from controls; <sup>@</sup>P < 0.01 significantly different from Forskolin treated groups; <sup>†</sup>P < 0.01 significantly different among drug-treated groups (100  $\mu$ M)

carbon monoxide [1]. Indeed the existence of the  $H_2S$ biosynthetic enzymes, CBS and CSE, in the retina suggests a potential physiological role for  $H_2S$  in this tissue [13, 14]. In a previous study, we found that the H<sub>2</sub>S donors, NaHS and Na<sub>2</sub>S, can inhibit excitatory amino acid transmission from isolated bovine and porcine retinae. Furthermore, this effect was determined to be dependent, at least in part, on intramural biosynthesis of this gas [20]. Based on the known actions of this gas on the vasculature and brain, numerous investigators have reported that possible mechanistic effects may involve potassium-sensitive ATP (K<sub>ATP</sub>) channels, reactive oxygen species (ROS), intracellular calcium and mitogen-activated protein (MAP) kinases [1, 2, 31]. However, the mechanism by which H<sub>2</sub>S elicits its physiological effects in the retina has not been clearly elucidated.





Fig. 5 Effects of NaHS on Forskolin (10  $\mu$ M)-stimulated cyclic AMP production in isolated bovine (a) and porcine (b) retina: control, in the presence of NaHS (10 nM–1  $\mu$ M) and Forskolin (10  $\mu$ M). *Vertical bars* represent means  $\pm$  SEM; n = 6. \*\*P < 0.001, \*P < 0.01 significantly different from controls; <sup>@</sup>P < 0.01 significantly different from groups treated with forskolin alone

The cyclic nucleotide, adenosine 3',5' cyclic monophosphate (cyclic AMP) is a ubiquitous cellular second messenger that is important in many biological processes. It is used for intracellular signal transduction and has been reported to be involved in numerous neuronal functions including cell survival, axon regeneration, and modulation of axonal guidance [24, 25]. In the present study, we examined the role of PDE in the metabolism of cyclic AMP in isolated bovine and porcine neural retinae. Inhibition of PDE with IBMX enhanced the responses elicited by the diterpene activator of adenylyl cyclase, forskolin suggesting that catabolism of cyclic AMP does occur in these tissues. Indeed, there is evidence that PDE is present in several retinal cells such as the ganglion, bipolar,

**Fig. 6** Effects of Na<sub>2</sub>S on Forskolin (10  $\mu$ M)-stimulated cyclic AMP production in isolated bovine (**a**) and porcine (**b**) retina: control, in the presence of Na<sub>2</sub>S (10 nM–100  $\mu$ M) and Forskolin (10  $\mu$ M). *Vertical bars* represent means  $\pm$  SEM; n = 6. \*\*P < 0.001, \*P < 0.01 significantly different from controls; <sup>@</sup>P < 0.001 significantly different form groups treated with forskolin alone

horizontal, amacrine and rod photoreceptors [32]. We included IBMX in all subsequent studies on the effects of  $H_2S$  donors on cyclic AMP production in the retina. Furthermore, we employed forskolin as a positive control in all assays performed in the present study.

Studies have reported that intracellularly,  $H_2S$  enhances *N*-methyl-D-aspartate (NMDA) receptor-mediated response via cyclic AMP production and that exogenous  $H_2S$ increases production of cyclic AMP in primary cultures of rat cerebral and cerebellar neurons, or in some neuronal and glial cell lines [30]. In the present study, we investigated the effect of  $H_2S$  donors on cyclic AMP production in isolated mammalian retinae. We report that the  $H_2S$ donor, NaHS produced a time-dependent increase in cyclic

AMP concentrations over basal levels in both isolated bovine and porcine retinae. Data from these experiments enabled us to determine an optimum time (20 min) for interaction between H<sub>2</sub>S donors and the adenylate cyclase pathway. Both NaHS and Na<sub>2</sub>S significantly increased cyclic AMP levels in bovine and porcine retinae in a concentration-related manner indicating that this nucleotide serves as a mediator of effects caused by H<sub>2</sub>S in these tissues. Data from these studies also show that NaHS was more potent than Na<sub>2</sub>S in stimulating cyclic AMP production in both bovine and porcine retinae. A similar change in sensitivity of tissues to NaHS and Na<sub>2</sub>S has been reported by our laboratory in studies of sympathetic neurotransmission in porcine iris-ciliary body [19], amino acid transmission in bovine retina [20] and in the relaxation of isolated porcine irides to both  $H_2S$  donors [18]. In the present study, we also observed that the ability of H<sub>2</sub>S donors to increase cyclic AMP concentrations was greater in bovine than porcine retina, suggesting that species differences exist in the response of this tissue to H<sub>2</sub>S donors. Opere et al. [20] also reported that species differences exist in the effects caused by H<sub>2</sub>S donors on glutamatergic transmission.

So far, data obtained from the present study shows that both  $H_2S$  donors and forskolin can increase the production of cyclic AMP in bovine and porcine retinae. To establish whether  $H_2S$  donors and forskolin utilize the same pathways for increasing cyclic AMP formation, we designed experiments that exposed retina to both agents simultaneously. We observed that in both bovine and porcine retinae, responses elicited by forskolin on cyclic formation was not additive with those produced by NaHS and Na<sub>2</sub>S indicating that a common pathway may mediate the observed responses. It is, however, unclear whether the effects of  $H_2S$  donors on the adenylyl cyclase pathway are due to a direct and/or indirect action on this enzyme.

In conclusion,  $H_2S$  donors can cause a time- and dosedependent increase in cyclic AMP production in isolated neural retina from cows and pigs. Bovine retina appears to be more sensitive to the stimulatory effect of  $H_2S$  donors on cyclic nucleotide production than its porcine counterpart indicating that species differences exist in the magnitude of this response. Effects produced by forskolin on cyclic AMP formation were not additive with those elicited by  $H_2S$ donors suggesting that these agents may share a common mechanism in their action on the adenylyl cyclase pathway. Taken together, our findings suggest that  $H_2S$  may play a regulatory role in signal transduction processes in mammalian retina.

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#### References

- 1. Lowicka E, Beltowski J (2007) Hydrogen sulfide (H<sub>2</sub>S)—the third gas of interest for pharmacologists. Pharmacol Rep 59:4–24
- Qu K, Lee SW, Bian JS et al (2008) Hydrogen sulfide: neurochemistry and neurobiology. Neurochem Int 52:155–165
- Abe K, Kimura H (1996) The possible role of hydrogen sulfide as an endogenous neuromodulator. J Neurosci 16:1066–1071
- Kimura H, Nagai Y, Umemura K et al (2005) Physiological roles of hydrogen sulfide: synaptic modulation, neuroprotection, and smooth muscle relaxation. Antioxid Redox Signal 7:795–803
- 5. Bukovska G, Kery V, Kraus JP (1994) Expression of human cystathionine beta-synthase in *Escherichia coli*: purification and characterization. Protein Expr Purif 5:442–448
- Erickson PF, Maxwell IH, Su LJ et al (1990) Sequence of cDNA for rat cystathionine gamma-lyase and comparison of deduced amino acid sequence with related *Escherichia coli* enzymes. Biochem J 269:335–340
- Stipanuk MH, Beck PW (1982) Characterization of the enzymic capacity for cysteine desulphhydration in liver and kidney of the rat. Biochem J 206:267–277
- Swaroop M, Bradley K, Ohura T et al (1992) Rat cystathionine beta-synthase. Gene organization and alternative splicing. J Biol Chem 267:11455–11461
- Florin TH (1991) Hydrogen sulphide and total acid-volatile sulphide in faeces, determined with a direct spectrophotometric method. Clin Chim Acta 196:127–134
- Kraft AA, Brant AW, Ayres JC (1956) Detection of hydrogen sulphide in packaged meals and in broken-out shell eggs. Food Technol 10:443–444
- Goffredi SK, Childress JJ, Desaulniers NT et al (1997) Sulfide acquisition by the vent worm Riftia pachyptila appears to be via uptake of HS-, rather than H<sub>2</sub>S. J Exp Biol 200:2609–2616
- Pitcher MC, Beatty ER, Cummings JH (2000) The contribution of sulphate reducing bacteria and 5-aminosalicylic acid to faecal sulphide in patients with ulcerative colitis. Gut 46:64–72
- Persa C, Osmotherly K, Chao-Wei CK et al (2006) The distribution of cystathionine beta-synthase (CBS) in the eye: implication of the presence of a trans-sulfuration pathway for oxidative stress defense. Exp Eye Res 83:817–823
- Pong WW, Stouracova R, Frank N et al (2007) Comparative localization of cystathionine beta-synthase and cystathionine gamma-lyase in retina: differences between amphibians and mammals. J Comp Neurol 505:158–165
- Beauchamp RO Jr, Bus JS, Popp JA et al (1984) A critical review of the literature on hydrogen sulfide toxicity. Crit Rev Toxicol 13:25–97
- 16. Kraus JP, Kozich V (2001) Cystathionine  $\beta$ -synthase and its deficiency. In: Carmel R, Jacobsen DW (eds) Homocysteine in health and disease. Cambridge University Press, Cambridge, pp 223–243
- Mudd SH, Levy HL, Kraus JP (2001) Disorders of trans-sulfuration. In: Scriver CR, Beaudet AL, Sly WS, Valle D, Childs B, Kinzler K, Vogelstein B (eds) The metabolic and molecular bases of inherited disease. McGraw-Hill, New York, pp 2007–2056
- Monjok EM, Kulkarni KH, Kouamou G et al (2008) Inhibitory action of hydrogen sulfide on muscarinic receptor-induced contraction of isolated porcine irides. Exp Eye Res 87:612–616
- Kulkarni KH, Monjok EM, Zeyssig R et al (2009) Effect of hydrogen sulfide on sympathetic neurotransmission and catecholamine levels in isolated porcine iris-ciliary body. Neurochem Res 34:400–406
- Opere CA, Monjok EM, Kulkarni KH et al (2009) Regulation of [(3)h] d: -aspartate release from mammalian isolated retinae by hydrogen sulfide. Neurochem Res 34:1962–1968

- Cheng Y, Ndisang JF, Tang G et al (2004) Hydrogen sulfideinduced relaxation of resistance mesenteric artery beds of rats. Am J Physiol Heart Circ Physiol 287:H2316–H2323
- 22. Zhao W, Zhang J, Lu Y et al (2001) The vasorelaxant effect of H(2)S as a novel endogenous gaseous K(ATP) channel opener. EMBO J 20:6008–6016
- 23. Kimura H (2002) Hydrogen sulfide as a neuromodulator. Mol Neurobiol 26:13–19
- 24. Hu Y, Cui Q, Harvey AR (2007) Interactive effects of C3, cyclic AMP and ciliary neurotrophic factor on adult retinal ganglion cell survival and axonal regeneration. Mol Cell Neurosci 34:88–98
- Teng FY, Tang BL (2006) Axonal regeneration in adult CNS neurons—signaling molecules and pathways. J Neurochem 96:1501–1508
- 26. Njie Y, Bongmba OYN, Opere CA et al (2008) Regulation of mammalian retinal neurotransmitter function by hydrogen sulfide: role of cyclic AMP. Invest Ophthalmol Vis Sci E-Abstract 2005

- Hosoki R, Matsuki N, Kimura H (1997) The possible role of hydrogen sulfide as an endogenous smooth muscle relaxant in synergy with nitric oxide. Biochem Biophys Res Commun 237:527–531
- Zhao W, Wang R (2002) H(2)S-induced vasorelaxation and underlying cellular and molecular mechanisms. Am J Physiol Heart Circ Physiol 283:H474–H480
- 29. Ohia SE, Opere C, Tang L et al (1995) Role of cyclic AMP in prostaglandin mediated responses in the neural retina. J Ocul Pharmacol Ther 11:73–81
- Kimura H (2000) Hydrogen sulfide induces cyclic AMP and modulates the NMDA receptor. Biochem Biophys Res Commun 267:129–133
- Chen CQ, Xin H, Zhu YZ (2007) Hydrogen sulfide: third gaseous transmitter, but with great pharmacological potential. Acta Pharmacol Sin 28:1709–1716
- Whitaker CM, Cooper NG (2009) The novel distribution of phosphodiesterase-4 subtypes within the rat retina. Neuroscience 163:1277–1291