CLINICAL STUDY

Molecular characteristics and clinical features of multifocal glioblastoma

Antonio Dono1,[2](http://orcid.org/0000-0002-8041-8399) · Emily Wang³ · Victor Lopez‑Rivera4 · Arvind V. Ramesh3 · Nitin Tandon1,5 · Leomar Y. Ballester1,2,5 · Yoshua Esquenazi1,5,[6](http://orcid.org/0000-0002-9757-1453)

Received: 17 April 2020 / Accepted: 14 May 2020 / Published online: 21 May 2020 © Springer Science+Business Media, LLC, part of Springer Nature 2020

Abstract

Introduction Glioblastomas (GBMs) usually occur as a solitary lesion; however, about 0.5–35% present with multiple lesions (M-GBM). The genetic landscape of GBMs have been thoroughly investigated; nevertheless, diferences between M-GBM and single-foci GBM (S-GBM) remains unclear. The present study aimed to determine diferences in clinical and molecular characteristics between M-GBM and S-GBM.

Methods A retrospective review of multifocal/multicentric infltrative gliomas (M-IG) from our institutional database was performed. Demographics, clinical, radiological, and genetic features were obtained and compared between M-GBM IDHwild type (IDH-WT) vs 193 S-GBM IDH-WT. Mutations were examined by a targeted next-generation sequencing assay interrogating 315 genes.

Results 33M-IG were identifed from which 94% were diagnosed as M-GBM IDH-WT, the remaining 6% were diagnosed as astrocytomas IDH-mutant. M-GBM and S-GBM comparison revealed that *EGFR* alterations were more frequent in M-GBM (65% vs 42% p=0.019). Furthermore, concomitant *EGFR*/*PTEN* alterations were more common in M-GBM vs. S-GBM (36% vs 19%) as well as compared to TCGA (21%). No statistically signifcant diferences in overall survival were observed between M-GBM and S-GBM; however, within the M-GBM cohort, patients harboring *KDR* alterations had a worse survival (*KDR-*altered 6.7 vs *KDR-*WT 16.6 months, *p*=0.038).

Conclusions The results of the present study demonstrate that M-GBM genetically resembles S-GBM, however, M-GBM harbor higher frequency of *EGFR* alterations and co-occurrence of *EGFR*/*PTEN* alterations, which may account for their highly malignant and invasive phenotype. Further study of genetic alterations including diferences between multifocal and multicentric GBMs are warranted, which may identify potential targets for this aggressive tumor.

Keywords Multifocal glioblastoma · Multicentric glioblastoma · *EGFR* · *KDR*

Electronic supplementary material The online version of this article [\(https://doi.org/10.1007/s11060-020-03539-z](https://doi.org/10.1007/s11060-020-03539-z)) contains supplementary material, which is available to authorized users.

 \boxtimes Leomar Y. Ballester Leomar.Y.Ballester@uth.tmc.edu

 \boxtimes Yoshua Esquenazi Yoshua.EsquenaziLevy@uth.tmc.edu

- Vivian L. Smith Department of Neurosurgery, The University of Texas Health Science Center at Houston, Houston, TX, USA
- ² Department of Pathology and Laboratory Medicine, The University of Texas Health Science Center at Houston, Houston, TX, USA
- Rice University, Houston, TX, USA
- ⁴ Department of Neurology, The University of Texas Health Science Center at Houston, Houston, TX, USA
- ⁵ Memorial Hermann Hospital-TMC, Houston, TX, USA
- ⁶ Center for Precision Health, School of Biomedical Informatics, The University of Texas Health Science Center at Houston, Houston, TX, USA

Introduction

Glioblastoma (GBM) is the most common primary malignant tumor of the central nervous system (CNS) and is associated with poor prognosis despite current therapies [[1](#page-7-0)–[3\]](#page-7-1). GBMs usually occur as a solitary lesion; however, about 0.5 to 35% of all GBMs present with multiple lesions (M-GBM) [\[4](#page-7-2)–[8\]](#page-7-3). Based on radiographic features, M-GBMs can be further divided into multifocal and multicentric GBMs. Multifocal GBMs have a clear contiguity pathway of spread between foci, which can be demonstrated by contiguous areas of T2-weighted signal on magnetic resonance imaging (MRI) of the brain. Meanwhile, multicentric GBMs are widely separated lesions that cannot be attributed to contiguous pathways [\[9](#page-7-4)]. The prognosis of multifocal and multicentric GBMs (M-GBM) have been historically reported to be more dismal than single foci GBM $(S-GBM)$ $[6-8]$ $[6-8]$ $[6-8]$.

The genetic landscape of GBMs has been thoroughly investigated revealing important genetic pathways that infuence its clinicopathologic characteristics and outcomes [[10,](#page-7-6) [11](#page-7-7)]. The exact mechanism of multifocality/ multicentricity is still not fully comprehended, as studies defning genomic alterations of these tumors are scarce. The molecular characteristics of M-GBM might explain its inherent ability to migrate and invade, resulting in the poor outcome observed in these patients. It has been described that M-GBM lacks *IDH1*, *ATRX*, and *PDGFRA* mutations. Nevertheless, given the small sample sizes of prior studies, the true genomic diferences between S-GBM and M-GBM remain unclear [[8](#page-7-3)].

The goal of the present study is to determine from our institutional experience the molecular characteristics and clinical features between M-GBM and S-GBM, to allow a better understanding of genetic alterations in M-GBM that could explain their biology and behavior.

Material and methods

Patients and tumor samples

We performed a retrospective review of patients diagnosed with infltrative glioma between January 2004 and December 2019. Patients were included if they had (1) histologic diagnosis of difuse glioma; (2) multifocal or multicentric tumor; (3) mutation analysis by next-generation sequencing. Patients with multifocal tumors were defned as those having at least two distinct tumor foci, separated by at least 1 cm. Multifocal and multicentric GBM (M-GBM) were analyzed together and individually for a subanalysis. A

cohort including 193 S-GBM, IDH-WT was used for comparison of demographic, clinical, and molecular characteristics (Online Resource 1). Patients diagnosed with difuse astrocytic gliomas, IDH-WT, with molecular features of GBM, WHO grade IV were considered as GBM, IDH-WT according to the 3rd cIMPACT-NOW update [[12](#page-7-8)].

Study data were collected from the electronic medical record of Memorial Hermann Hospital and managed using REDCap electronic data capture tools hosted at the University of Texas Health Science Center at Houston (UT Houston) [[13](#page-7-9), [14\]](#page-7-10). These included age, gender, Karnofsky performance status (KPS), histologic diagnosis, tumor location, radiographic extent of resection, treatment strategy, recurrence, and survival. Tumors were classifed by a Board-certifed neuropathologist following the 2016 WHO Classifcation of Tumors of the Central Nervous System [[15\]](#page-7-11). This study was approved by the Institutional Review Board of the UT Houston and Memorial Hermann Hospital, Houston, TX.

Targeted sequencing

Tumor samples were analyzed for genomic alterations by a targeted next-generation sequencing assay (NGS) interrogating 315 genes and 28 gene rearrangements (FoundationOne CDx®, Foundation Medicine, Inc., Cambridge, MA, USA). The FoundationOne® assay was performed in a clinical laboratory improvement amendments certifed laboratory, as previously described [[16–](#page-7-12)[18](#page-8-0)]. Telomerase reverse transcriptase promoter (*TERTp*) status was not available for four patients.

Co‑occurrence of most common mutations

To evaluate the co-occurrence of *EGFR*/*PTEN*, *EGFR*/*KDR*, and *EGFR*/*PTEN*/*TERTp* alterations, we compared M-GBM vs S-GBM. In addition, to further investigate these diferences we compared our results to the known incidence of these mutations in The Cancer Genome Atlas (TCGA) PanCancer Atlas for *EGFR*/*PTEN* and *EGFR*/*KDR*, and to a published study evaluating co-occurrence of *EGFR*/*PTEN*/*TERTp* alterations, as TCGA lacks *TERTp* information [[19–](#page-8-1)[22\]](#page-8-2).

Statistical analyses

Clinical, demographic, and frequency of genomic alterations were evaluated by Fisher's exact test or Mann–Whitney U test for categorical and continuous variables, respectively. Overall survival was calculated as the time in months from diagnosis to death or date of last available follow-up. Kaplan–Meier method was used to plot survival curves and statistical signifcance was examined by the univariable log-rank test. Cox proportional hazard regression models were utilized for univariable and multivariable analysis to calculate the hazard ratio (HR) estimates with 95% confdence interval (CI). Multivariable Cox proportional hazard regression model analysis was adjusted for the following set of covariates: age, male gender, KPS, Stupp protocol, salvage bevacizumab, multifocal vs multicentric, and *KDR* mutant gene, as these are factors well-known to affect survival [\[23](#page-8-3)], *KDR* was selected as it was found to be signifcant in univariable analysis. A *p-*value≤0.05 was considered statistically signifcant. Statistical analyses were performed in EZR (1.40) [\[24\]](#page-8-4) and Prism v.8.2.1 (GraphPad, La Jolla, California, USA).

Results

Clinico‑demographic characteristics of multifocal infltrative gliomas

A total of 564 patients with infiltrative gliomas (IG) were identified, from which 41 (7.3%) had a multifocal/ multicentric profile (M-IG). There were 39/395 (9.9%) M-GBM IDH-WT patients, 2/69 (2.9%) astrocytomas IDH-mutant (IDH-MT) presented multiple lesions at the time of initial diagnosis. From the 41 patients, 33 met the inclusion criteria and were further analyzed (Online Resource 1).

The 33M-IG patients had a median age of 60 years (range 26–77). There were 17 males and 16 females, 24.2% had KPS scores ≥ 80 , and 29/33 (88%) underwent resection, meanwhile, the remaining 12% underwent biopsy. Temozolomide was prescribed to 31 (93.9%) patients with concomitant radiotherapy to 30 (90.9%) according to the Stupp protocol [\[25\]](#page-8-5). Moreover, some patients were treated with 1st line bevacizumab (7/33, 21.2%), irinotecan (5/33, 15.2%), and tumor-treating felds (5/33, 15.2%). Histologically, 31 patients (94%) were diagnosed as GBM, IDH-WT, 1 (3%) GBM IDH-mutant (IDH-MT), and 1 (3%) difuse astrocytoma (DA), IDH-MT. Furthermore, 22/33 (66.7%) patients could be further classifed as multicentric and 30 (90.9%) patients had enhancing tumors (Fig. [1](#page-2-0))*.* The demographic and clinical characteristics are depicted in Table [1](#page-3-0)*.*

The comparison between S-GBM and M-GBM revealed that the latter was less likely to have gross total resection (GTR, 12.9% vs 33.5% , $p = 0.02$) but was treated more frequently with 1st line irinotecan (1.5% vs 12.9%, $p = 0.007$) and bevacizumab (9.3% vs 19.4%, $p = 0.11$). Re-resection was less likely to occur in M-GBM patients

Fig. 1 M-GBM MRI imaging. Patient 1, axial T1 post contrast (**a**) and axial T2 fair (**b**) demonstrating a multifocal GBM IDH-WT. Patient 27, sagittal T1 post contrast (**c**) and axial T2 Flair (**d**) demonstrating a multicentric GBM IDH-WT. Patient 20, axial T2 Flair (**e**),

and axial T1 post contrast (**f**) demonstrating a multicentric partially enhancing GBM IDH-WT. Patient 32, axial T1 post contrast (**g**) demonstrating a multifocal GBM IDH-MT. Patient 33, sagittal (**h**) MRI demonstrating a multicentric difuse astrocytoma IDH-MT

GBM glioblastoma, *WT* wild type, *MT* mutant, *IQR* interquartile range, *S-GBM* single foci glioblastoma, *M-GBM* multifocal/multicentric glioblastoma

^aNot all patients had a recurrence, for the salvage therapies a total of 154 GBM IDH-WT were taken into account [128 S-GBM and 26M-GBM (9 multifocal and 17 multicentric)]

^bTested in 221 patients as *TERTp* status was not available for 3 patients. Fisher's exact test or Mann–Whitney U test was performed for categorical and continuous variables respectively. *P-*values were two-sided and a *p*≤0.05 was considered as statistically signifcant and are represented in bold

Fig. 2 Mutations in cancer-related genes of 33 multicentric/multifocal infltrative glioma patients*. PFS* progression-free survival (months), *OS* overall survival (months), *WT* wild type, *Mut* mutant,

M male, *F* female. *Anaplastic astrocytoma IDH-WT with molecular features of GBM WHO grade IV according to cIMPACT-NOW Update 3

 $(39.8\% \text{ vs } 19.2\%, p=0.07)$. Furthermore, the analysis between multifocal $(n = 10)$ and multicentric $(n = 21)$ GBM IDH-WT showed that within this group, multifocal patients were more likely to undergo re-resection and salvage therapies at recurrence (Table [1\)](#page-3-0).

Genetic alterations

NGS analysis revealed that *TERTp* was mutated in 22/29 (75.9%) patients. *CDKN2A*/*B* loss was observed in 25/33 (75.7%), while *EGFR* alterations in 21/33 (63.6%) patients. *PTEN* was mutated in 18/33 (54.5%) patients. The other frequently altered genes were *TP53* in 8/33 (24.4%), *KDR*, *KIT*, and *PDGFRA*, each mutated in 5/33 (15.2%) patients (Fig. [2;](#page-4-0) Table [1](#page-3-0)).

The comparison between S-GBM and M-GBM revealed that *EGFR* alterations were more frequently present in M-GBM (65% vs 42% *p* = 0.019, Table [1](#page-3-0) and Online Resource 2). Moreover, we observed that concomitant mutations in *EGFR* and *PTEN* genes were more frequently observed in M-GBM (M-GBM 36% vs S-GBM 19%, $p = 0.05$). Also, the percentage of patients harboring the aforementioned mutation concomitantly with *TERTp* mutations was higher in M-GBM (M-GBM 25% vs S-GBM 18%, $p = 0.43$). Analysis of TCGA data revealed a co-occurrence of *EGFR*/*PTEN* alterations in 81/378 (21.4%) GBM IDH-WT cases. This was similar to the S-GBM group (19%) but lower than M-GBM patients in our study [[19,](#page-8-1) [20](#page-8-6), [22](#page-8-2)]. The co-occurrence of *EGFR*, *PTEN*, and *TERTp* also appeared to be higher in M-GBM $(25%)$ compared to prior reports $(5.6%)$ [[21](#page-8-7)]. Also, we evaluated the co-occurrence of *EGFR*/*KDR* alterations in the entire cohort ($n=224$) and M-GBM ($n=31$), in which there was not an association between these genes ($p=0.34$) and 0.12, respectively). These results were further validated by the lack of association in the TCGA database between *EGFR*/*KDR* alterations (*p* = 0.49) and *EGFR* amplification and *KDR* alterations $(p=0.17)$ [[19](#page-8-1), [20,](#page-8-6) [22](#page-8-2)].

Prognosis and survival

Univariable analysis of GBM IDH-WT patients $(n=224)$ demonstrated that patients with age<55 years, KPS 80 or higher, and 1st line Stupp protocol had improved OS. However, only age and 1st line Stupp protocol were independent predictors of poor survival after multivariable analysis. Importantly, we observed a trend towards shorter survival (13.0 vs 17.9 months, $p = 0.31$) in M-GBM patients (Fig. [3](#page-5-0)a, Online Resource 3).

No statistically significant differences were observed between multifocal and multicentric GBM. However, multifocal GBM showed a trend towards better outcomes (Fig. [3b](#page-5-0)). In addition, M-GBM patients with *KDR* alterations had a worse survival than *KDR*-WT patients in univariable analysis (6.7 vs 16.6 months, $p=0.038$) and multivariable analysis (HR 9.3 [1.17–73.8], *p*=0.035, Fig. [3](#page-5-0)c; Table [2\)](#page-5-1). The efect of *KDR* alterations in the outcome of S-GBM was observed in the univariable log-rank test, in which *KDR* altered S-GBM had a worse survival than their *KDR*-WT counterparts (11.4 vs 18.5 months, *p*=0.015). However, this difference was not observed after multivariable analysis (*KDR* altered HR 1.78 [0.74–4.31], *p*=0.20). Survival diferences

Fig. 3 Overall survival in GBM IDH-WT. **a** Overall survival of GBM IDH-WT $(n=224)$ by multifocality, in which there was not statistically signifcant diference between the survival of M-GBM (n=31) and S-GBM (n=193, 13.0 vs 17.9 months, *p*=0.31). **b** Overall survival within M-GBM $(n=31)$. There was not statistically significant difference between multifocal $(n=10)$ and multicentric $(n=21)$ patients (27.7 vs 12.5 months, $p=0.07$). **c** Overall survival of $M-GBM$ ($n=31$) by KDR gene status, in which KDR mutant patients

Table 2 Multivariable Cox proportional hazard regression model of overall survival in patients with M-GBM $(n=31)$

Variable	Multivariable HR [95% CI]	p -value
Age at diagnosis > 55 years	0.51 [-0.13 to 1.91]	0.315
Male	4.87 [0.90 to 26.3]	0.065
KPS > 80	1.32 [0.36 to 4.80]	0.677
Stupp protocol	0.09 [0.006 to 1.39]	0.086
Salvage bevacizumab	0.55 [0.14 to 2.10]	0.382
Multifocal	0.38 [0.08 to 1.77]	0.219
KDR MT	9.30 [1.17 to 73.8]	0.035

P-values were two-sided and a $p < 0.05$ was considered as statistically signifcant and are represented in bold

M-GBM multifocal/multicentric glioblastoma, *KPS* Karnofsky performance status, *MT* mutant, *HR* hazard ratios, *CI* confdence interval

in M-GBM patients were not observed between *EGFR*-altered (12.5 months) and *EGFR*-WT (16.5 months) tumors (Fig. [3](#page-5-0)d; Table [2](#page-5-1)).

 $(n=4)$ had worse survival compared to KDR WT patients $(n=27, 6.7)$ vs 16.6 months, $p = 0.038$). **d** Overall survival of M-GBM by EGFR gene status, in which there was not statistically signifcant diference between EGFR altered ($n=20$) and EGFR WT ($n=10$) patients (12.5) vs 16.5 months, *p*=0.54). *GBM* glioblastoma, *M-GBM* multifocal/ multicentric GBM, *S-GBM* single foci GBM, *WT* wild type. Kaplan– Meier curves were examined by the log-rank test, a $p \le 0.05$ was considered as statistically signifcant

Discussion

Several studies on M-GBM have reported decreased survival when compared to S-GBM; however, studies focusing on the molecular characteristics of M-GBM are scarce. It is still unclear whether M-GBM represents a distinct biologic variant of GBM or if multifocal progression is part of the natural history of the disease $[7, 8]$ $[7, 8]$ $[7, 8]$ $[7, 8]$. The current study represents, the largest M-GBM cohort with comprehensive genomic characterization, in which subtle molecular diferences were observed. Our data suggest that M-GBM genetically resembles typical GBM.

In the present cohort, we identifed that 9.9% of GBM IDH-WT had multiple lesions at diagnosis, which is comparable to recent studies that have used similar criteria to defne M-GBM [[5,](#page-7-14) [7,](#page-7-13) [8\]](#page-7-3). Interestingly, we identifed 2 cases (2.9%) of multifocal astrocytomas IDH-MT. A recent study reported that multifocality is an independent predictor for *IDH1*-WT status, as only 1/102 (1%) patients had an *IDH1* mutation $[26]$. The multifocal tumor, in this case, was an oligodendroglioma, IDH*-*mutant and 1p/19q co-deleted. A prior study by Liu et al. [\[8](#page-7-3)] reported all multifocal patients to be IDH-WT. In our study, both patients (32 and 33) were astrocytomas, IDH-mutant (1p/19q intact). Thus, to the best of our knowledge, these cases represent the frst reports of multifocal IDH-MT astrocytomas (Figs. [1g](#page-2-0), h, [2](#page-4-0)).

Previous studies have described the diferential expression of MAPK, PTEN, MGMT, and EGFR between S-GBM and M-GBM; however, these studies are limited by their small sample size and lack of comprehensive molecular characterization [[5](#page-7-14), [7](#page-7-13), [27](#page-8-9)]. A recent study on 18M-GBM patients with mutation analysis identifed the absence of *IDH1*, *ATRX*, or *PDGFRA* mutations in all patients [[8](#page-7-3)]. These results contrast with our fndings in which in addition to the observed IDH1 p.R132H mutation in two patients*,* we also observed mutations in *ATRX* and *PDGFRA* amplifcation in 2.9% and 15.2% of patients, respectively. *ATRX* mutations are widely distributed across the gene and are mostly truncating (including frame shift and nonsense variants) as seen in the current study (patient 32 with an *ATRX* N111fs*16 and patient 33 with an *ATRX* E1541*) [[28](#page-8-10)]. *ATRX* mutations occur in 78% and 63% of GBM, IDH-MT and astrocytoma IDH-MT, respectively, and only in 3% of GBM IDH-WT [[29\]](#page-8-11). Moreover, a Japanese study identifed loss of ATRX expression in 28.6% of 14 multicentric astrocytomas, IDH-WT [\[27](#page-8-9)]. Thus, it is not surprising that *ATRX* mutations were not observed in previous smaller cohorts. *PDGFRA* discrepancies between our study and previous studies could be also explained by sample size, as we observed that *PDGFRA* is amplifed in 15.2% of M-IG and 12.9% of M-GBM. This represents roughly the same known frequency of *PDGFRA* amplifcations in GBM, as 13.5% of S-GBM and 14.7% of GBMs in TCGA harbored *PDGFRA* amplifcation [[19\]](#page-8-1). Moreover, *PDGFRA* amplifcation has been reported in 16.7% of a small M-GBM study [\[30](#page-8-12)].

EGFR alterations are found in 30–50% of all GBMs and are associated with tumor development and progression [[19,](#page-8-1) [31](#page-8-13)]. Therefore, *EGFR* has been proposed as a potential therapeutic target for GBMs; however, clinical trials targeting this gene have been unsuccessful [\[32](#page-8-14), [33](#page-8-15)]. A previous study with six M-GBM found aberrations in *EGFR* in all patients [\[30\]](#page-8-12). In our study, we also identifed that *EGFR* alterations were more common in M-GBM than in S-GBM (65% vs $42\%, p=0.019$.

Importantly, *EGFR* alterations have been associated with an increased infltrative and invasive phenotype [[30](#page-8-12), [34](#page-8-16)], and therefore with a multifocal appearance as seen in our study. Notably, we observed that co-occurrence of *EGFR* and *PTEN* alterations, as well as, *EGFR*, *PTEN*, and *TERTp* alterations, was increased in M-GBM compared to both S-GBMs in our cohort and published literature [\[19–](#page-8-1)[22](#page-8-2)]. These fndings are in accordance with the results of a previous study that described 67% of M-GBM showed alterations in *EGFR*, *PTEN*, and *TERTp* [\[30](#page-8-12)]. Therefore M-GBM genetically resembles typical GBM with an increased incidence

of *EGFR* mutations and co-occurrence of *EGFR*, *PTEN*, and *TERTp* alterations.

A previous study has also identifed a higher expression of *CYB5R2* in M-GBM, which has been independently associated with worse OS and has been proposed as a potential prognostic and diagnostic marker [[8\]](#page-7-3). Unfortunately, we did not examine the expression of this gene in our study.

Overall survival and multifocality

Multifocality has been consistently correlated with poor survival in GBM [\[4](#page-7-2)[–8](#page-7-3), [35,](#page-8-17) [36\]](#page-8-18). In our study, M-GBM trended towards a worse survival compared to S-GBM; however, it did not reach statistical signifcance. This could be explained due to the limited sample size, the relatively small percentage of biopsies performed in these cases (12.2%) compared to prior reports $(28-100\%)$ [5-[7](#page-7-13), 35-[38\]](#page-8-19), and the more aggressive adjuvant upfront approach demonstrated by increased utilization of chemotherapeutic agents in these patients (bevacizumab and irinotecan) compared to S-GBM (Online Resource 4). Prior centers have also described an aggressive approach for these cases, suggesting multiple craniotomies for maximal cytoreduction that could potentially improve survival [\[9](#page-7-4)]. Interestingly, we identifed for the frst time that alterations in *KDR* predicted worse survival within the M-GBM group. Even though our results should be taken with caution and validated in a larger cohort, *KDR*, also known as *VEGFR2*, is a tyrosine kinase receptor that plays an important role in GBM angiogenesis, aggressiveness, and progression [[39](#page-8-20)[–42\]](#page-8-21). Previous studies have demonstrated that the *KDR* CAGT haplotype increases GBM aggressiveness and that concomitant high mRNA expression of *KDR*, *FLT1*, and *VEGFA* has been associated with shorter survival [\[39,](#page-8-20) [40](#page-8-22)]. Bevacizumab, a monoclonal antibody towards VEGF, is one of the few drugs currently approved for GBM treatment. A recent study has demonstrated that *EGFR*altered patients with recurrent GBM, have a signifcantly shorter time to progression when treated with bevacizumab [\[43\]](#page-8-23). Moreover, *KDR* activation through paracrine secretion of VEGF-C has been demonstrated to represent an escape mechanism employed by GBM to counteract bevacizumab therapy, which could potentially explain the worse survival seen in *KDR*-altered patients [\[44](#page-8-24)]. While *KDR* alterations have been demonstrated to predict bevacizumab response in other malignancies such as colon cancer [[45\]](#page-8-25), it is still unknown if alterations in *KDR* and its related genes *FLT1* and *VEGFA* could predict response to bevacizumab in GBM.

Multifocal vs multicentric GBM

Finally, we investigated the diferences between multifocal and multicentric GBM. While no demographic diferences were observed, our study revealed that multicentric GBM

was less likely to undergo re-resection, salvage therapies, and GTR. Prior studies have also reported worse outcomes in multicentric GBM compared to multifocal GBM (3 vs. 11 months) [\[6](#page-7-5)], which is in concordance with our results. Interestingly, various genetic diferences were observed in the univariate analysis between these groups demonstrating diferences in *CDKN2A*/*B*, *CDK4*, *PTEN*, and *TP53* genes that have not been previously described and may warrant further study (Table [1](#page-3-0)).

Despite several limitations, such as the retrospective nature, limited sample size, and lack of sequencing information from distinct tumor foci, the present study represents the largest cohort of multifocal/multicentric gliomas undergoing comprehensive genetic characterization and demonstrates the unique molecular features of this aggressive type of GBM.

Conclusions

The results of the present study demonstrate that M-GBM genetically resembles S-GBM, however, M-GBM have a higher frequency of *EGFR* alterations and co-occurrence of *EGFR*/*PTEN* alterations, which may account for their highly malignant and invasive phenotype.

Acknowledgements None.

Author contributions Study design: AD, LYB, and YE. Data recollection: AD, EW, and AR Data analysis: AD and VLR. Manuscript writing: AD, EW, and YE. Manuscript revision and editing: AD, NT, LYB, and YE. Study supervision: LYB and YE. Approved fnal manuscript: all authors.

Funding No funding to disclose.

Data availability The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.Code availability Not applicable.

Consent to participate Not applicable.

Consent for publication Not applicable.

Compliance with ethical standards

Conflict of interest The authors declared no confict of interest.

Ethical approval This retrospective study was approved by the Institutional Review Board of The University of Texas Health Science Center at Houston and Memorial Hermann Hospital, Houston, TX following the 1964 Helsinki declaration and its later amendments.

References

- 1. Ostrom QT, Gittleman H, Truitt G et al (2018) CBTRUS Statistical Report: primary brain and other central nervous system tumors diagnosed in the United States in 2011–2015. Neuro-oncology 20:iv1–iv86. <https://doi.org/10.1093/neuonc/noy131>
- 2. Zhu P, Du XL, Zhu J-J, Esquenazi Y (2019) Improved survival of glioblastoma patients treated at academic and high-volume facilities: a hospital-based study from the National Cancer Database. J Neurosurg.<https://doi.org/10.3171/2018.10.JNS182247>
- 3. Ferlay J, Colombet M, Soerjomataram I et al (2019) Estimating the global cancer incidence and mortality in 2018: GLOBOCAN sources and methods. Int J Cancer 144:1941-1953. [https://doi.](https://doi.org/10.1002/ijc.31937) [org/10.1002/ijc.31937](https://doi.org/10.1002/ijc.31937)
- Showalter TN, Andrel J, Andrews DW et al (2007) Multifocal glioblastoma multiforme: prognostic factors and patterns of progression. Int J Radiat Oncol Biol Phys 69:820–824. [https://doi.](https://doi.org/10.1016/j.ijrobp.2007.03.045) [org/10.1016/j.ijrobp.2007.03.045](https://doi.org/10.1016/j.ijrobp.2007.03.045)
- 5. Patil CG, Yi A, Elramsisy A et al (2012) Prognosis of patients with multifocal glioblastoma: a case–control study. J Neurosurg 117:705–711. <https://doi.org/10.3171/2012.7.JNS12147>
- 6. Thomas RP, Xu LW, Lober RM et al (2013) The incidence and signifcance of multiple lesions in glioblastoma. J Neurooncol 112:91–97.<https://doi.org/10.1007/s11060-012-1030-1>
- 7. Paulsson AK, Holmes JA, Peifer AM et al (2014) Comparison of clinical outcomes and genomic characteristics of single focus and multifocal glioblastoma. J Neurooncol 119:429–435. [https://](https://doi.org/10.1007/s11060-014-1515-1) doi.org/10.1007/s11060-014-1515-1
- 8. Liu Q, Liu Y, Li W et al (2015) Genetic, epigenetic, and molecular landscapes of multifocal and multicentric glioblastoma. Acta Neuropathol 130:587–597. [https://doi.org/10.1007/s0040](https://doi.org/10.1007/s00401-015-1470-8) [1-015-1470-8](https://doi.org/10.1007/s00401-015-1470-8)
- 9. Hassaneen W, Levine NB, Suki D et al (2011) Multiple craniotomies in the management of multifocal and multicentric glioblastoma. J Neurosurg 114:576–584. [https://doi.](https://doi.org/10.3171/2010.6.JNS091326) [org/10.3171/2010.6.JNS091326](https://doi.org/10.3171/2010.6.JNS091326)
- 10. Yan H, Parsons DW, Jin G et al (2009) Mutations in gliomas. N Engl J Med 360:765–773. [https://doi.org/10.1056/NEJMoa0808](https://doi.org/10.1056/NEJMoa0808710) [710](https://doi.org/10.1056/NEJMoa0808710)
- 11. Brennan CW, Verhaak RGW, McKenna A et al (2013) The somatic genomic landscape of glioblastoma. Cell 155:462. [https](https://doi.org/10.1016/j.cell.2013.09.034) [://doi.org/10.1016/j.cell.2013.09.034](https://doi.org/10.1016/j.cell.2013.09.034)
- 12. Brat DJ, Aldape K, Colman H et al (2018) cIMPACT-NOW update 3: recommended diagnostic criteria for "Difuse astrocytic glioma, IDH-wildtype, with molecular features of glioblastoma, WHO grade IV". Acta Neuropathol 136:805–810. [https://doi.](https://doi.org/10.1007/s00401-018-1913-0) [org/10.1007/s00401-018-1913-0](https://doi.org/10.1007/s00401-018-1913-0)
- 13. Harris PA, Taylor R, Thielke R et al (2009) Research electronic data capture (REDCap)—a metadata-driven methodology and workflow process for providing translational research informatics support. J Biomed Inform 42:377–381. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jbi.2008.08.010) [jbi.2008.08.010](https://doi.org/10.1016/j.jbi.2008.08.010)
- 14. Harris PA, Taylor R, Minor BL et al (2019) The REDCap Consortium: building an international community of software platform partners. J Biomed Inform 95:103208. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jbi.2019.103208) [jbi.2019.103208](https://doi.org/10.1016/j.jbi.2019.103208)
- 15. Louis DN, Perry A, Reifenberger G et al (2016) The 2016 World Health Organization Classifcation of Tumors of the Central Nervous System: a summary. Acta Neuropathol 131:803–820. [https://](https://doi.org/10.1007/s00401-016-1545-1) doi.org/10.1007/s00401-016-1545-1
- 16. Zorofchian S, El-Achi H, Yan Y, et al (2018) Characterization of genomic alterations in primary central nervous system lymphomas. J Neurooncol 140:509–517. [https://doi.org/10.1007/s1106](https://doi.org/10.1007/s11060-018-2990-6) [0-018-2990-6](https://doi.org/10.1007/s11060-018-2990-6)
- 17. Frampton GM, Fichtenholtz A, Otto GA et al (2013) Development and validation of a clinical cancer genomic profling test based on massively parallel DNA sequencing. Nat Biotechnol 31:1023–1031. <https://doi.org/10.1038/nbt.2696>
- 18. Schwaederle M, Krishnamurthy N, Daniels GA et al (2018) Telomerase reverse transcriptase promoter alterations across cancer types as detected by next-generation sequencing: a clinical and molecular analysis of 423 patients. Cancer 124:1288–1296. [https](https://doi.org/10.1002/cncr.31175) [://doi.org/10.1002/cncr.31175](https://doi.org/10.1002/cncr.31175)
- 19. McLendon R, Friedman A, Bigner D et al (2008) Comprehensive genomic characterization defnes human glioblastoma genes and core pathways. Nature 455:1061–1068. [https://doi.org/10.1038/](https://doi.org/10.1038/nature07385) [nature07385](https://doi.org/10.1038/nature07385)
- 20. Gao J, Aksoy BA, Dogrusoz U et al (2013) Integrative analysis of complex cancer genomics and clinical profles using the cBioPortal. Sci Signal 6:1–20. <https://doi.org/10.1126/scisignal.2004088>
- 21. Nonoguchi N, Ohta T, Eun J (2013) TERT promoter mutations in primary and secondary glioblastomas. 931–937. [https://doi.](https://doi.org/10.1007/s00401-013-1163-0) [org/10.1007/s00401-013-1163-0](https://doi.org/10.1007/s00401-013-1163-0)
- 22. Cerami E, Gao J, Dogrusoz U et al (2012) The cBio Cancer Genomics Portal: an open platform for exploring multidimensional cancer genomics data. Cancer Discov 2:401–404. [https://](https://doi.org/10.1158/2159-8290.CD-12-0095) doi.org/10.1158/2159-8290.CD-12-0095
- 23. Thakkar JP, Dolecek TA, Horbinski C et al (2014) Epidemiologic and molecular prognostic review of glioblastoma. Cancer Epidemiol Biomark Prev 23:1985–1996. [https://doi.org/10.1158/1055-](https://doi.org/10.1158/1055-9965.EPI-14-0275) [9965.EPI-14-0275](https://doi.org/10.1158/1055-9965.EPI-14-0275)
- 24. Kanda Y (2013) Investigation of the freely available easy-to-use software "EZR" for medical statistics. Bone Marrow Transplant 48:452–458.<https://doi.org/10.1038/bmt.2012.244>
- 25. Stupp R, Mason WP, van den Bent MJ et al (2005) Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. N Engl J Med 352:987–996. [https://doi.org/10.1056/NEJMoa0433](https://doi.org/10.1056/NEJMoa043330) 30
- 26. Park YW, Han K, Ahn SS et al (2018) Prediction of IDH1-mutation and 1p/19q-codeletion status using preoperative MR imaging phenotypes in lower grade gliomas. Am J Neuroradiol 39:37–42. <https://doi.org/10.3174/ajnr.A5421>
- 27. Karlowee V, Amatya VJ, Hirano H et al (2017) Multicentric glioma develops via a mutant IDH1-independent pathway: immunohistochemical study of multicentric glioma. Pathobiology 84:99–107.<https://doi.org/10.1159/000447951>
- 28. Heaphy CM, De Wilde RF, Jiao Y et al (2011) Altered telomeres in tumors with ATRX and DAXX mutations. Science (80-) 333:425.<https://doi.org/10.1126/science.1207313>
- 29. Pekmezci M, Rice T, Molinaro AM et al (2017) Adult infltrating gliomas with WHO 2016 integrated diagnosis: additional prognostic roles of ATRX and TERT. Acta Neuropathol 133:1001– 1016.<https://doi.org/10.1007/s00401-017-1690-1>
- 30. Abou-El-Ardat K, Seifert M, Becker K et al (2017) Comprehensive molecular characterization of multifocal glioblastoma proves its monoclonal origin and reveals novel insights into clonal evolution and heterogeneity of glioblastomas. Neuro-oncology 19:546– 557.<https://doi.org/10.1093/neuonc/now231>
- 31. Parsons DW, Jones S, Zhang X et al (2008) An integrated genomic analysis of human glioblastoma multiforme. Science (80-) 321:1807–1812.<https://doi.org/10.1126/science.1164382>
- 32. Lassman AB, Van Den Bent MJ, Gan HK et al (2019) Safety and efficacy of depatuxizumab mafodotin $+$ temozolomide in patients with EGFR-amplifed, recurrent glioblastoma: results

from an international phase I multicenter trial. Neuro-oncology 21:106–114.<https://doi.org/10.1093/neuonc/noy091>

- 33. Lassman AB, Pugh SL, Wang TJC, Aldape K, Gan HK, Preusser M, Vogelbaum MA, Sulman E, Won M, Zhang P, Moazami G, Macsai MS, Gilbert MR, Bain E, Blot V, Ansell PJ, Samanta S, Kundu MG, Seidel C, de Vos FY, Hsu S, Cardona AF, Lombardi G, Bentsion D, Peterson R, Gedye C, Lebrun-Frenay C, Wick A, Curran WJ, Mehta M (2019) Epidermal Growth Factor Receptor (EGFR) amplifed (amp) newly diagnosed glioblastoma (nGBM). In: Paper presented at the annual meeting of Society of Neuro-Oncology, Phoenix, AZ
- 34. Talasila KM, Soentgerath A, Euskirchen P, et al (2013) EGFR wild-type amplifcation and activation promote invasion and development of glioblastoma independent of angiogenesis. 683– 698.<https://doi.org/10.1007/s00401-013-1101-1>
- 35. Syed M, Liermann J, Verma V et al (2018) Survival and recurrence patterns of multifocal glioblastoma after radiation therapy. Cancer Manag Res 10:4229–4235. [https://doi.org/10.2147/](https://doi.org/10.2147/CMAR.S165956) [CMAR.S165956](https://doi.org/10.2147/CMAR.S165956)
- 36. Pérez-Beteta J, Molina-García D, Villena M et al (2019) Morphologic features on MR imaging classify multifocal glioblastomas in diferent prognostic groups. Am J Neuroradiol 40:634–640. [https](https://doi.org/10.3174/ajnr.A6019) [://doi.org/10.3174/ajnr.A6019](https://doi.org/10.3174/ajnr.A6019)
- 37. Singh G, Mehrotra A, Das K et al (2015) Multiple glioblastomas: are they diferent from their solitary counterparts? Asian J Neurosurg 10:266.<https://doi.org/10.4103/1793-5482.162685>
- 38. Burger MC, Breuer S, Cieplik HC et al (2017) Bevacizumab for patients with recurrent multifocal glioblastomas. Int J Mol Sci 18:1–11. <https://doi.org/10.3390/ijms18112469>
- 39. Vasconcelos VCA, Lourenço GJ, Brito ABC et al (2019) Associations of VEGFA and KDR single-nucleotide polymorphisms and increased risk and aggressiveness of high-grade gliomas. Tumor Biol 41:1–10.<https://doi.org/10.1177/1010428319872092>
- 40. Zhang SD, Leung KL, McCrudden CM, Kwok HF (2015) The prognostic signifcance of combining VEGFA, FLT1 and KDR mRNA expressions in brain tumors. J Cancer 6:812–818. [https://](https://doi.org/10.7150/jca.11975) doi.org/10.7150/jca.11975
- 41. Sjöström S, Wibom C, Andersson U et al (2011) Genetic variations in VEGF and VEGFR2 and glioblastoma outcome. J Neurooncol 104:523–527.<https://doi.org/10.1007/s11060-010-0504-2>
- 42. Wu HB, Yang S, Weng HY et al (2017) Autophagy-induced KDR/ VEGFR-2 activation promotes the formation of vasculogenic mimicry by glioma stem cells. Autophagy 13:1528–1542. [https](https://doi.org/10.1080/15548627.2017.1336277) [://doi.org/10.1080/15548627.2017.1336277](https://doi.org/10.1080/15548627.2017.1336277)
- 43. Hovinga KE, McCrea HJ, Brennan C et al (2019) EGFR amplifcation and classical subtype are associated with a poor response to bevacizumab in recurrent glioblastoma. J Neurooncol 142:337– 345.<https://doi.org/10.1007/s11060-019-03102-5>
- 44. Michaelsen SR, Staberg M, Pedersen H et al (2018) VEGF-C sustains VEGFR2 activation under bevacizumab therapy and promotes glioblastoma maintenance. Neuro-oncology 20:1462–1474. <https://doi.org/10.1093/neuonc/noy103>
- 45. Zhang SD, McCrudden CM, Meng C et al (2015) The signifcance of combining VEGFA, FLT1, and KDR expressions in colon cancer patient prognosis and predicting response to bevacizumab. Oncotargets Ther 8:835–843. [https://doi.org/10.2147/OTT.S8051](https://doi.org/10.2147/OTT.S80518) [8](https://doi.org/10.2147/OTT.S80518)

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional afliations.