



BCL2 expression is associated with a poor prognosis independent of cellular origin in primary central nervous system diffuse large B-cell lymphoma

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Abstract

Purpose Primary central nervous system diffuse large B-cell lymphoma (CNS-DLBCL) is a distinct clinicopathological entity with a poor prognosis. Concurrent MYC and BCL2 overexpression predicts inferior prognosis in systemic DLBCL, although their prognostic significance remains unclear in primary CNS-DLBCL.

Methods Pretreatment diagnostic biopsy samples were retrospectively evaluated for 79 patients with primary CNS-DLBCL who were treated between January 2001 and December 2017. Histological and immunohistochemical testing were performed to evaluate the patients' statuses for various markers, which were also evaluated for associations with survival outcomes.

Results According to the Hans criteria, 26 patients (32.9%) had the germinal center B-cell subtype and 53 patients (67.1%) had the activated B-cell subtype. Forty-one cases (51.9%) were positive for MYC (expression of $\geq 40\%$), 33 cases (41.8%) were positive for BCL2 (expression of $\geq 70\%$), 22 patients (27.8%) were positive for both MYC and BCL2, and 27 patients (34.2%) were negative for both MYC and BCL2. There were no significant differences in survival between the germinal center and activated B-cell subtypes. Furthermore, MYC positivity was not associated with overall survival ($p=0.369$) or progression-free survival (PFS) ($p=0.253$). However, BCL2 positivity was significantly associated with poor overall survival ($p=0.039$) and PFS ($p=0.036$). Co-expression of MYC and BCL2 was not associated with survival.

Conclusion Our data suggest that evaluating BCL2 expression may help predict the prognosis in cases of primary CNS-DLBCL.

Keywords Primary central nervous system lymphoma · MYC · BCL2 · Immunohistochemistry · Cell of origin

Introduction

Primary central nervous system diffuse large B-cell lymphoma (CNS-DLBCL) is an aggressive non-Hodgkin's lymphoma that affects the brain or spinal cord without evidence of systemic involvement at the diagnosis. However, primary CNS-DLBCL is rare and only accounts for <1% of all non-Hodgkin's lymphomas and approximately 3% of all primary brain tumors [1]. Compared to systemic DLBCL,

CNS-DLBCL has a poorer prognosis and it remains unclear whether this is related to the tumor's location, the microenvironment, an inherently aggressive biology, or a combination of those factors. The cellular origin of CNS-DLBCL is thought to be a late germinal center B-cell (GCB), based on results from immunophenotypic analysis, gene expression profiling, and molecular characterization of the immunoglobulin genes [2, 3]. In addition, the GCB-like subtype of DLBCL is thought to have a better prognosis than the activated B-cell (ABC)-like subtype [4, 5]. However, most primary CNS-DLBCLs involve the ABC subtype, which may explain the poor prognosis of primary CNS-DLBCL [6].

The MYC gene has recently been shown to be translocated in 5–15% of systemic DLBCLs. This genetic alteration worsens the response to chemotherapy and prognosis, especially when it occurs with concomitant BCL2 and/or BCL6 translocations (double-hit and triple-hit lymphomas)

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[7–10]. Furthermore, recent studies have demonstrated that immunohistochemical detection of high MYC and BCL2 protein expressions (double expressing lymphomas) is associated with an aggressive behavior and inferior overall survival (OS), regardless of any MYC and BCL2 rearrangements [11–15]. However, these proteins' relevance remains unclear in primary central nervous system lymphoma (PCNSL) without systemic involvement. Moreover, one large multi-center analysis revealed that the prognostic value of the cell of origin (COO) was lost after adjusting for double expressing lymphomas, which suggests that immunohistochemical detection of MYC and BCL2 co-expression was a more reliable prognostic factor among patients with DLBCL [12]. The present study used immunohistochemistry to evaluate the expressions of the MYC and BCL2 proteins, as well as their prognostic value relative to the COO concept, in CNS-DLBCL.

Materials and methods

Patients

Pretreatment diagnostic biopsy samples were retrospectively retrieved for 79 immunocompetent patients with newly diagnosed PCNSL who were treated between January 2001 and December 2017. The treatment involved induction therapy using a cycle of high-dose methotrexate (MTX) at 3.5 g/m² on days 1, 22, and 43, which was followed by radiotherapy as previously described [16]. All patients had provided written informed consent for their treatment, and this study's retrospective protocol was approved by our institutional ethics review board.

Histological and immunohistochemical testing

Tissue samples were fixed in formaldehyde and embedded in paraffin using standard methods. Sections were stained using hematoxylin and eosin before the immunohistochemical analysis. Routine deparaffinization, rehydration, and blocking of endogenous peroxidase activity were performed before the slide-mounted sections were immersed in 0.01 M sodium citrate buffer (pH 6.0) and heated at maximum power for 15 min in a 700-W microwave oven.

Immunohistochemical staining was performed for CD20 (L26; 1:200), CD10 (56C6; 1:25), and BCL6 (LN22; 1:100) using antibodies from Leica Biosystems (Wetzler, Germany). In addition, staining was performed for MUM-1 (MUM1p; 1:50), BCL2 (124; 1:200), and Ki-67 (MIB1; 1:50) using antibodies from DAKO (CA, USA). Staining for c-MYC (Y69; 1:200) was performed using antibodies from Abcam (Cambridge, UK). The staining pattern for MYC was distinctly nuclear and the staining pattern for

BCL2 was clearly cytoplasmic. The cutoff values for positivity were defined as $\geq 40\%$ for MYC and $\geq 70\%$ for BCL2, as previously described [11]. The cutoff values for positivity were defined as $\geq 30\%$ for CD10, BCL6, and MUM1. Cases were assigned to the GCB group based on positive results for CD10 or to the ABC group (non-GCB) based on negative results for CD10 and BCL6. If the results were negative for CD10 and positive for BCL6, the case was assigned to the GCB group if there was a negative result for MUM1 and to the ABC group if there was a positive result for MUM1 [17].

Statistical analysis

The survival intervals were calculated from the diagnosis to death or the last follow-up for OS and from the diagnosis to the first instance of disease progression, relapse, or death for progression-free survival (PFS). Survival curves were created using the Kaplan–Meier method and analyzed using the log-rank test and GraphPad Prism software (GraphPad Software Inc., San Diego, USA). The analyzed variables included patient age and sex, Karnofsky performance status (KPS) at admission, the number of lesions, and the expression statuses for MYC and/or BCL2. The Memorial Sloan Kettering Cancer Center prognostic score (MSKCC) was also calculated based on age and KPS [18]. All other statistical analyses were performed using StatView software (version 5; Abacus Concepts Inc., Berkeley, USA) and p-values of < 0.05 were considered statistically significant.

Results

The clinical features of the 79 patients with PCNSL are listed in Table 1. The median age was 68 years (range 41–86 years), the median KPS at admission was 40% (range 20–100%), and the median follow-up was 30.3 months (range 0.4–159.3 months). All patients were treated using high-dose MTX-based chemotherapy and achieved median OS and PFS values of 42.4 and 6.5 months, respectively. However, no significant differences in survival were observed according to the various clinical features, such as age, sex, KPS, number of lesions, and Memorial Sloan Kettering Cancer Center class (Table 2).

All cases had diffuse large cell morphology and were immunohistochemically positive for CD20, which indicated that all cases involved diffuse large B-cell lymphomas. The patients were subsequently grouped according to their immunohistochemical expressions of CD10, BCL6, and MUM1, with 26 patients (32.9%) having the GCB subtype and 53 patients (67.1%) having the ABC subtype (Table 1). No significant differences in OS and PFS were observed between the two subtypes (Table 2).

Table 1 Clinical characteristics of patients

Characteristic	Number (%)
Age (years)	
Median (range)	68 (41–86)
≥ 60	61 (77.2)
< 60	18 (22.8)
Sex	
Male	43 (54.4)
Female	36 (45.6)
KPS at admission (%)	
Median (range)	40 (20–100)
≥ 50	32 (40.5)
< 50	47 (59.5)
Number of lesions	
Multiple	47 (59.5)
Single	32 (40.5)
MSKCC class	
Class 1	9 (11.4)
Class 2	13 (16.4)
Class 3	57 (72.2)
Cell of origin	
GCB	26 (32.9)
ABC	53 (67.1)

KPS Karnofsky performance scale, MSKCC Memorial Sloan Kettering Cancer Center, GCB germinal center B-cell, ABC activated B-cell

Immunohistochemical testing for the MYC and BCL2 proteins revealed that 41 patients (51.9%) were positive for MYC and 33 patients (41.8%) were positive for BCL2 (Table 2). Twenty-two patients (27.8%) were positive for both MYC and BCL2, while 27 patients (34.2%) were negative for both MYC and BCL2. Figure 1 shows the immunohistochemical findings from a representative case involving the GCB subtype. Figure 2 shows that univariate analyses of BCL2 expression revealed significant associations with poor OS (31.8 vs. 78.7 months; $p=0.039$) and poor PFS (5.2 vs. 6.5 months; $p=0.036$). However, MYC expression with or without BCL2 co-expression did not significantly affect OS ($p=0.369$ and $p=0.136$, respectively) or PFS ($p=0.253$ and $p=0.056$, respectively) (Table 2). The significant factors in the univariate analyses were included in the multivariate analyses of OS and PFS (Table 3), which revealed that BCL2 positivity independently affected OS (hazard ratio [HR]: 3.137, $p=0.016$) but not PFS (HR: 2.134, $p=0.071$).

Discussion

The present study evaluated the patients' immunochemical statuses for MYC and BCL2, as well as the COO concept, to better predict the prognosis of primary CNS-DLBCL. Our

Table 2 Treatment results and univariate analysis of clinical and immunohistochemical features

	Overall survival		Progression free survival	
	Months	P value	Months	P value
Total (n=79)	42.4		6.5	
Sex				
Male (n=43)	43.0	0.943	5.2	0.530
Female (n=36)	42.4		6.5	
Age				
≥ 60 (n=61)	32.2	0.149	5.1	0.538
< 60 (n=18)	78.7		11.7	
KPS				
≥ 50 (n=32)	56.6	0.109	18.7	0.130
< 50 (n=47)	31.8		4.9	
Number of lesions				
Multiple (n=47)	31.8	0.340	5.1	0.952
Single (n=32)	43.0		9.4	
MSKCC class				
1 (n=9)	78.7	0.223	7.6	0.497
2 (n=13)	87.9		17.9	
3 (n=57)	32.2		5.1	
MYC				
Positive (n=41)	34.1	0.369	4.9	0.253
Negative (n=38)	56.6		12.9	
BCL2				
Positive (n=33)	31.8	0.039	5.2	0.036
Negative (n=46)	78.7		6.5	
Double				
Positive (n=22)	31.3	0.136	5.1	0.056
Negative (n=57)	51.1		7.6	
Cell of origin				
GCB (n=26)	31.2	0.358	11.7	0.349
ABC (n=53)	56.6		5.2	

KPS Karnofsky performance scale, MSKCC Memorial Sloan Kettering Cancer Center, GCB germinal center B-cell, ABC activated B-cell

results indicate that BCL2 expression had significant value for predicting both OS and PFS. However, MYC expression, co-expression of MYC and BCL2, the COO concept, and traditional prognostic factors (e.g., age and KPS) did not predict survival among our patients. Patient age and KPS have been suggested as prognostic factors and strongly influence the therapeutic decisions for PCNSL. For example, Abrey et al. [18] proposed the MSKCC model and indicated that patients who were < 50 years old had improved OS and PFS. However, we did not identify significant associations of these prognostic factors and the MSKCC score with our patients' outcomes. Our previous study evaluated treatment results in patients with PCNSL and examined various prognostic factors including the MSKCC score [16]. That study revealed

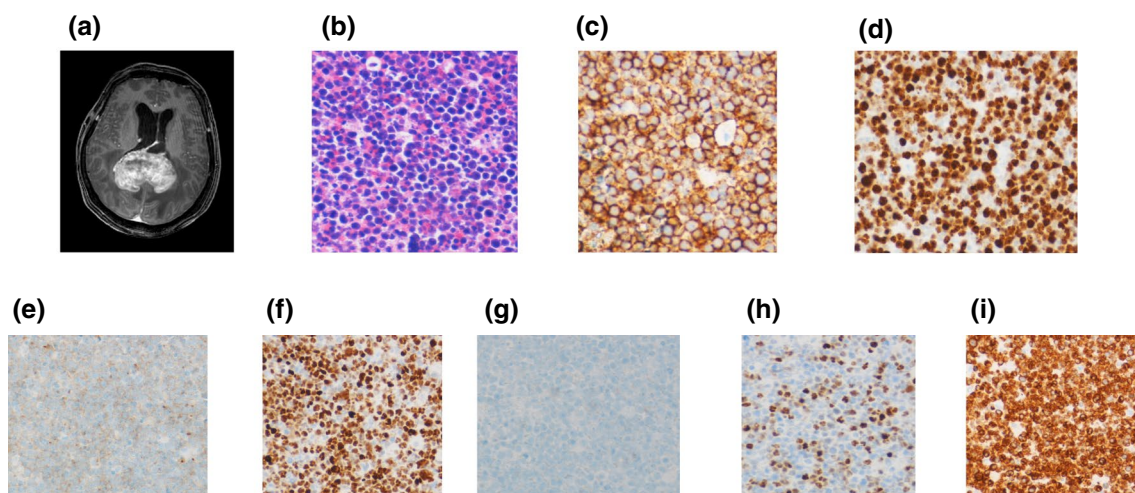


Fig. 1 A representative case of the GCB subtype. Homogenous enhancement of the lesion in the splenium of the corpus callosum during magnetic resonance imaging with gadolinium-DTPA (**a**). Diffusely infiltrating large neoplastic lymphocytes (**b**). Positive staining for CD20 (**c**). A high proliferative rate was detected via staining

for MIB-1 (**d**). Negative staining for CD10 (**e**). Positive staining for BCL6 (**f**). Negative staining for MUM1 (**g**). Positive staining for MYC (**h**). Positive staining for BCL2 (**i**). Panels (**b–i**) are shown at $\times 200$ magnification

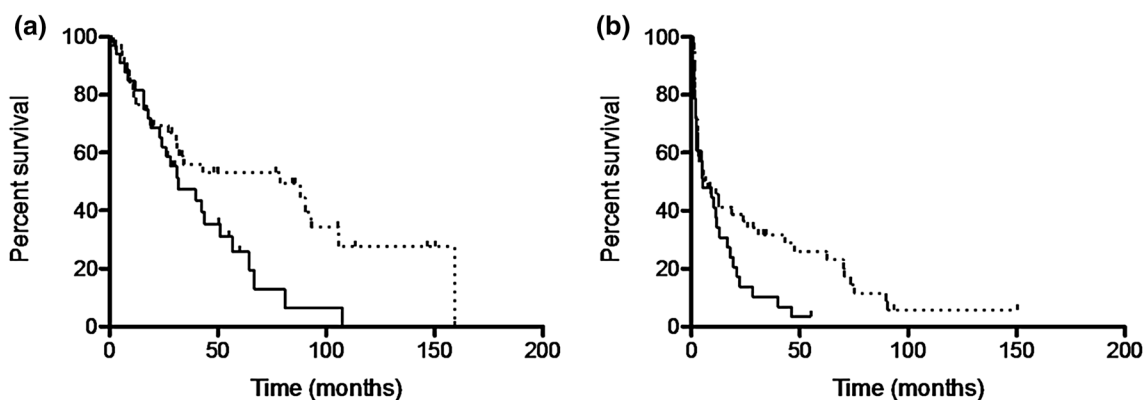


Fig. 2 Kaplan–Meier curves for overall survival (**a**) and progression-free survival (**b**) among patients with primary CNS-DLBCL that was positive for BCL2 ($n=33$; solid line) or negative for BCL2 ($n=46$; dashed line)

Table 3 Multivariate analysis of clinical and immunohistochemical features

	Overall survival			Progression free survival		
	HR	95% CI	P value	HR	95% CI	P value
Sex (male)	2.000	0.982–4.073	0.056	1.370	0.744–2.522	0.312
Age (≥ 60)	1.903	0.512–7.073	0.334	1.411	0.551–3.615	0.473
KPS (≥ 50)	0.639	0.291–1.404	0.265	0.724	0.378–1.385	0.329
Number of lesions (multiple)	1.326	0.704–2.495	0.382	1.008	0.595–1.708	0.977
MSKCC class 1	1.246	0.250–6.216	0.789	1.715	0.532–5.527	0.367
Class 2	0.776	0.233–2.585	0.680	0.843	0.353–2.017	0.702
MYC (positive)	1.81	0.736–4.452	0.196	1.591	0.736–3.439	0.237
BCL2 (positive)	3.137	1.234–7.974	0.016	2.134	0.937–4.859	0.071
Double (positive)	0.567	0.163–1.972	0.372	0.745	0.245–2.266	0.605
COO (ABC)	0.522	0.266–1.023	0.058	1.070	0.602–1.902	0.818

KPS Karnofsky performance scale, MSKCC Memorial Sloan Kettering Cancer Center, COO cell of origin, ABC activated B-cell

that completion of three cycles of high-dose MTX chemotherapy, rather than the traditional prognostic factors (e.g., age and KPS), independently predicted better OS relative to patients who did not complete three cycles of chemotherapy (56.4 vs. 24.0 months, $p=0.013$). In this context, *BCL2* is a known antiapoptotic molecule and its expression might be related to the response to MTX chemotherapy. The present study included 50 patients who completed three cycles of chemotherapy and experienced marginally better OS than patients who did not complete the chemotherapy (51.1 vs. 26.1 months, $p=0.071$). Interestingly, the completion rates were similar among patients with and without *BCL2* expression (63.6 and 63.1%, respectively). Thus, although we did not identify a significant relationship between *BCL2* expression and response to high-dose MTX, we believe that further studies are needed to investigate this potential relationship.

The COO concept is based on gene expression profiles and is considered a strong prognostic factor for systemic DLBCL, with the COO subtypes having different responses to various therapeutic agents [19, 20]. In addition, several reports have indicated that the COO concept might predict prognosis among patients with DLBCL who underwent chemotherapy [17, 21, 22]. However, a few recent studies have indicated that COO defined using immunochemistry could not predict the prognosis of DLBCL [23, 24], which may be related to the immunohistochemical detection of COO producing inconsistent results among patients with primary CNS-DLBCL, similar to among patients with systemic DLBCL. Most primary CNS-DLBCLs are the non-GCB subtype, with Lin et al. reporting that 40 of 51 primary CNS-DLBCL cases (78%) involved the non-GCB subtype, while only 11 cases (22%) involved the GCB subtype [25]. Similarly, Hattab et al. found that 26 of 31 patients (84%) with primary CNS-DLBCL had the non-GCB subtype [26], while Camilleri-Broët et al. reported that 80 of 83 patients (94.3%) with primary CNS-DLBCL had the non-GCB subtype [6]. The present study revealed that 26 patients had the GCB subtype and that 53 patients (67.1%) had the ABC/non-GCB subtype, which is consistent with the findings of the previous reports. Interestingly, Camilleri-Broët et al. suggested that the poor prognosis of primary CNS-DLBCL might be partly related to its ABC subtype [6], which may indicate that the GCB subtype is associated with a better prognosis. However, several reports have described no significant difference in survival when the two subtypes were compared among patients with primary CNS-DLBCL [26–28]. Similarly, we failed to detect significant differences in OS and PFS when we compared the two subtype groups.

Double-hit lymphomas are rare (3–5% of DLBCL cases) and have chromosomal breaks involving *MYC* and *BCL2* [12, 29]. In addition, DLBCL with co-rearrangement of *MYC* and *BCL2* has poor OS and PFS outcomes [12, 29], while co-expression of *MYC* and *BCL2* (double expressing

lymphomas) is associated with a poor prognosis in systemic DLBCL, which is independent of the COO subtype [12, 13, 30]. Several studies have also examined co-expression and co-rearrangement of *MYC* and *BCL2* in primary CNS-DLBCL. Although *MYC* expression in primary CNS-DLBCL significantly exceeds that in systemic DLBCL, *MYC* rearrangement is similar for primary CNS-DLBCL and systemic DLBCL [31]. Therefore, the rarity of *MYC* rearrangement and the high frequency of *MYC* overexpression is an interesting phenomenon that may be attributed to other genetic aberrations, such as *MYC* mutations or altered regulation of expression.

The reported rates of *MYC* expression are 70–90% for CNS-DLBCL [27, 31, 32], which are higher than the reported rates for systemic DLBCL [12, 13]. Although high *MYC* expression reportedly predicts poor OS and/or PFS for CNS-DLBCL [32–34], our results revealed no significant relationship between *MYC* expression and survival (OS: $p=0.369$, PFS: $p=0.253$) and similar results have also been reported [27]. Other reports have indicated that the rates of *BCL2* expression are 59–73% for PCNSL [28, 32–34], with Shi et al. and Kim et al. reporting that high *BCL2* expression was significantly associated with poor OS [32, 33], although other studies failed to detect an association with prognosis [28, 34]. The present study revealed a slightly lower *BCL2* expression rate (41.8%), although we found that high *BCL2* expression was significantly associated with poor OS and PFS.

Interestingly, co-expression of *MYC* and *BCL2* in primary CNS-DLBCL did not have prognostic value, which conflicts with Guo et al.'s report that *MYC* expression and co-expression of *MYC* and *BCL2* were significantly associated with outcomes among patients with primary CNS-DLBCL [35]. Moreover, Shi et al. investigated 77 patients with primary CNS-DLBCL and reported that co-expression of *MYC* and *BCL2* strongly predicted the prognosis for primary CNS-DLBCL, independent of the COO concept [32]. Kim et al. have also reported that patients with primary CNS-DLBCL co-expressing *MYC* and *BCL2* experienced poor PFS outcomes in the MTX-treated group [33]. In contrast, Gill et al. reported that expression of *MYC* and/or *BCL2* was not associated with survival outcomes [27]. The present study revealed co-expression of *MYC* and *BCL2* in 27.8% of cases, although this factor was not significantly associated with OS or PFS. These discrepancies may be related to the use of different antibodies and immunohistochemical cutoff values. For example, Kim et al. used antibodies against *MYC* (EP121, Cell Marque) and *BCL2* (124, DAKO) with cutoff values of $\geq 40\%$ for *MYC* and $\geq 30\%$ for *BCL2* [33], which generated positivity rates of 18.1% for *MYC* and 75% for *BCL2* that predicted survival outcomes. In contrast, Gill et al. used different antibodies against *MYC* (9E10, Epitomics Inc.) and *BCL2* (bcl-2/100

D5, Novocastera) with cutoff values of $\geq 40\%$ for MYC and $\geq 70\%$ for BCL2 [27], which generated positivity rates of 73% for MYC and 71% for BCL2 that did not predict survival outcomes. Moreover, Guo et al. and Shi et al. used different antibodies against MYC (Y69, Abcam) and BCL2 (124, DAKO) with cut-off values of 33.3 and 78% for MYC and 81.8 and 61% for BCL2 [32, 35]. Both studies used a $\geq 40\%$ cutoff value for MYC, which agreed with previous studies, although the cutoff values for BCL2 that were 30 or 70% higher than in the other studies. Interestingly, Guo et al. only detected significant prognostic value for MYC expression, while Shi et al. detected significant prognostic value for both MYC and BCL2 expression. The present study used the same antibodies (Y69 for MYC and 124 for BCL2) with cutoff values of $\geq 40\%$ for MYC and $\geq 70\%$ for BCL2. The positivity rates were 51.9% for MYC and 41.8% for BCL2, but only BCL2 had significant prognostic value. Therefore, it is important to be aware of the specific antibodies and cutoff values that are being used to examine the prognostic values of these markers.

In conclusion, the present study revealed that BCL2 expression had good prognostic value, independent of the COO concept, among patients with primary CNS-DLBCL. The prognostic value of BCL2 is especially important in the era of targeted therapy, as this expression may predict a good response to anti-BCL2 targeted drugs.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflicts of interest.

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