V. N. Mukhin, K. I. Pavlov, and V. M. Klimenko

Translated from Rossiiskii Fiziologicheskii Zhurnal imeni I. M. Sechenova, Vol. 102, No. 2, pp. 113–129, February, 2016. Original article submitted October 23, 2015. Revised version received November 24, 2015.

One of the main elements in the pathogenesis of Alzheimer's disease consists of larger decreases in the numbers of neurons in various parts of the brain than seen in normal ageing. The relevance of studying the pathogenesis of this process arises from the fact that neuron loss starts at the early, preclinical stage of Alzheimer's disease, when amyloid plaques and neurofibrillary tangles (the main morphological signs of the disease) have still not formed; neuron loss correlates with the extent of clinical signs of the disease. Data have now accumulated on the likely pathogenetic mechanisms of neuron loss. The aim of the present literature review was to summarize these data.

Keywords: Alzheimer's disease, neurodegeneration, apoptosis, β-amyloid, postnatal neurogenesis.

 Alzheimer's disease is a chronic neurodegenerative disease apparent as impairments to memory, thinking, spatial orientation, and speech, along with disorders of the emotional domain. The disease generally starts at ages of more than 60 years, has an irreversible, progressive course, and in its late stages is characterized by amnesia, agnosia, apraxia, aphasia, complete degradation of personality, and loss of self-care ability; it ends with the patient's death from loss of regulation of vital functions or infectious complications. In Russia, Alzheimer's disease affects more than 1.2 million people -4.5% of those aged more than 60 years [2, 4].

 One of the basic elements in the pathogenesis of Alzheimer's disease consists of more intense decreases in the numbers of neurons in various areas of the brain than seen in normal aging [30, 40]. Neuron loss starts at the early, preclinical stage of Alzheimer's disease, when amyloid plaques and neurofibrillary tangles (the main morphological manifestations of the disease) have not yet formed [45, 103]. During this time, there are decreases in the numbers of neurons in hippocampal field CA1, the dentate fascia, the subiculum, and layer 2 of the entorhinal cortex [16, 45, 101, 113]. The process then spreads to the temporal (superior temporal gyrus), frontal, and parietal (supramarginal gyrus) lobes of the cortex [30]. At the late stages of the disease, the process of neuron loss affects the whole brain, including the olfactory bulbs [120], the amygdala [66, 124], the basal nucleus of Meynert [13, 131, 132], the substantia nigra [137], the locus ceruleus [21, 128], and the dorsal raphe nucleus [85].

 Decreases in the number of neurons, particularly in hippocampal field CA1 and the entorhinal cortex, correlate with the severity of memory impairments [44]. The likely reason for this is loss of the capacity for rearrangement of the neuronal organization of cerebral structures and addition of new neurons to them, which are required for normal learning processes to take place [9, 32, 47, 112, 134].

 These points make studies of the pathogenetic mechanisms of neuron loss in Alzheimer's disease very relevant. Further studies require summaries of the data thus far accumulated. This is the aim of the present review.

The Normal Dynamics of Neuron Numbers in the Brain during Ontogeny

In Alzheimer's disease, the decrease in neuron numbers evidently occurs as a result of impairments to the normal processes maintaining them: postnatal neurogenesis and natural neuron death. We will describe these here.

 Neurogenesis is the process whereby new neurons appear in the brain as a result of neural stem cell division and subsequent proliferation and differentiation of daughter

Physiology Department, Institute of Experimental Medicine, St. Petersburg, Russia; e-mail: Valery.Mukhin@gmail.com.

cells and their functional integration into defined brain structures [5]. This process only occurs at a high level during prenatal development. By birth, neurogenesis terminates in most brain structures and neuron counts reach a stable level, which persists to middle age [96, 99]. During this time, the number of neurons in the adult human brain is, according to different sources, 85–200 billion [15, 93, 133]. Of these, 12–20 billion neurons are in the telencephalon (most in the neocortex, about 20–30 million in the hippocampus, 4.5–7 million in the subiculum, and 10 million in the amygdala), 70–130 billion in the cerebellum, and fewer than one billion in the brainstem and spinal cord [12, 15, 113, 124, 133].

 In contrast to other brain structures, the appearance of new neurons in the ventromedial zone of the prefrontal cortex, the dentate gyrus, and the striatum continues after birth in humans (postnatal neurogenesis). The daughters of neural stem cells in the subventricular zone migrate to the prefrontal cortex and striatum, where they become interneurons [107]. We note that another hypothesis holds that new striatal neurons are derived from astrocytes in this brain structure [36, 77]. In the dentate gyrus, new granular neurons appear as a result of the division of stem cells in the subgranular zone [65]. The incorporation of new neurons into the olfactory bulbs in adult humans is dubious (in contrast to other animal species) [18].

 In those brain structures in which postnatal neurogenesis is observed, neuron numbers are determined by the dynamic equilibrium between the appearance of new neurons and their natural death. For example, the intensity of neuron replacement in the human dentate gyrus is 0.004% of the population per day, while the proportion of neurons replaced over the whole life is 35% [117].

 With age, the intensity of postnatal neurogenesis in the dentate gyrus and subventricular zones in humans (and other mammals) decreases; quickly in the first months of life and then gradually [36, 46, 67, 117]. The inclusion of new neurons into the prefrontal cortex ceases during the first year of life [107]. In the dentate gyrus, postnatal neurogenesis, gradually weakening, continues to old age [67].

Another process influencing the numbers of neurons in the brain is naturally occurring neuron death [76, 91]. Neurons in various stages of maturity undergo naturally occurring death. Some 50% of immature neurons appearing as a result of prenatal and postnatal neurogenesis are not integrated into brain structures and undergo natural termination of their existence [118]. Naturally occurring death is not characteristic of mature neurons, as these cells have well developed mechanisms suppressing it [68].

 The main mechanism of naturally occurring neuronal death is apoptosis. Autophagy, pyroptosis, and oncosis are significantly rarer [91, 118, 138]. The intensity of naturally occurring neuronal death can increase with age. Thus, the mouse hypothalamus shows age-related increases in the proportion of apoptotic neurosecretory cells [1].

 Naturally occurring neuronal death leads to age-related decreases in the numbers of neurons and their density per unit tissue volume in the hippocampal formation, neocortex, thalamus, and cerebellum [30, 103].

 The process of age-related decreases in neuron numbers and densities occurs most intensely in the hippocampal formation, affecting the subiculum and field CA1. By the end of life, neuron death in these structures amounts to 50% of the initial number of cells and the rate of death reaches 3.6% for each 10 years of life [86, 113, 130]. Published data suggest that the natural decrease in the number of neurons in the subiculum and field CA1 terminates after 60 years. In studies demonstrating this reduction, the lower limit of the age range was significantly below 60 years, while studies on the over-60 contingent did not see this pattern [101].

 The number of neurons in the neocortex decreases in old age by 10% of the initial number (by an average of one neuron per second), mainly because of the postcentral gyrus, the frontal associative cortex, the anterior zone of the cingulate gyrus, the temporal associative cortex, the inferior parietal gyrus, and the occipital lobe [30, 95, 96]. However, the decrease in the number of neurons in the neocortex was small and some authors have not detected it at all [88, 121].

Mechanisms of Decreases in Neurogenesis and Maturation of Neurons in Alzheimer's Disease

 Decreases in neuron numbers in Alzheimer's disease arise from impairment to both of the processes maintaining it: postnatal neurogenesis and naturally occurring neuronal death.

Impairments to postnatal neurogenesis. Changes in postnatal neurogenesis are different in different parts of the brain. The subventricular zone shows decreases, while the subgranular zone of the dentate gyrus shows increases. In addition, new neurons appear where this does not usually $occur - in field CA1$. These conclusions are based on data showing changes in the intensity of the expression of the molecular markers of neural precursor cells [6] in the brains of dead patients [61, 142]. However, some investigators have suggested that increased neurogenesis in the subgranular zone is more apparent than real, as increased expression of the molecular markers of neurogenesis is linked with increases in glial and vascular rather than neuronal proliferation [20]. Modeling of Alzheimer's disease in animals has demonstrated: increased neurogenesis in the subgranular zone was seen only at the late stages of the disease; at the earlier stages, weakening of neurogenesis occurs in both progenitor cell division zones and at all steps in cell division, including the step forming mature neurons [27, 105].

 The literature contains only fragmentary data on the mechanisms weakening neurogenesis in Alzheimer's disease.

 Decreases in allopregnanolone have been suggested to be significant, this substance being able to simulate neuronal proliferation via GABA receptors [26, 57, 126]. A decrease in the allopregnanolone level have been seen in the frontal cortex of patients [80]. Studies in a transgenic model of Alzheimer's disease in mice showed that an increased allopregnanolone level restores impaired cognitive functions [127]. The likely cause of the decrease in allopregnanolone in patients is deranged cholesterol metabolism.

 Another suggested cause for the weakening of neurogenesis is cholinergic denervation of the hippocampus and neocortex. Dysfunction of the basal cholinergic system of the brain is one of the main elements in the pathogenesis of Alzheimer's disease. Accumulation of soluble β-amyloid oligomers in brain tissue leads to decreased secretion and increased reuptake of acetylcholine by neurons of the basal cholinergic system, along with degeneration of the axons of these neurons [7]. Experimental weakening of the activity of the basal cholinergic system leads to decreases in the level of neurogenesis in the olfactory bulbs and dentate gyrus of these animals [31].

 At the cellular-molecular level, the mechanism of the decrease in neurogenesis in the presence of β-amyloid includes dysregulation of calcium homeostasis and activation of calpains and caspases [51].

Impairment to neuron maturation. The maturation of neurons formed during neurogenesis in the dentate gyrus is impaired in Alzheimer's disease [75]. Neuron processes are unable to develop into axons (neuronal polarization), to lengthen, or to undergo functional integration. These processes are prevented by high levels of glycogen synthase kinase 3β (GSK3β) activity, which are seen in neurons in Alzheimer's disease and are due to increases in extracellular levels of soluble β-amyloid oligomers [7, 74, 98].

 Neuron polarization can only occur at low levels of GSK3β in cells [42, 60, 111, 136]. High GSK3β activity impairs the stability and remodeling of neuron microtubules, thus preventing neuronal polarization [55, 115]. The pathogenesis of impaired neuron maturation in Alzheimer's disease may involve three mechanisms of GSK3β action on microtubule remodeling.

 Firstly, GSK3β phosphorylates tau protein [59, 70]. Hyperphosphorylation of tau protein is one of the main elements in the pathogenesis of Alzheimer's disease, leading to the formation and accumulation of neurofibrillary tangles in patients' cerebral neurons. Hyperphosphorylated tau protein in neurofibrillary tangles [104] is believed to be unable to interact with the tubulin in microtubules. These undergo depolymerization and their stability decreases, and this leads to impairments to neuron maturation [62].

 Secondly, increased GSK3β activity corresponds to inactivation (phosphorylation) of collapsin response mediator protein 2 (CRMP-2) [29, 60, 136]. Inactivation of CRMP-2 decreases its binding to tubulin in microtubules, impairing their polymerization and, thus, axon growth and branching [41, 136].

 Thirdly, GSK3β phosphorylates adenomatous polyposis coli protein (APC). This has the result that the interaction of APC with microtubules decreases and their stability is reduced [143].

510 Mukhin, Pavlov, and Klimenko

Increases in the probability of neuron death in Alzheimer's disease. The cellular-molecular mechanism of neuron death in the brain in Alzheimer's disease has not been firmly established. Results from studies of post mortem neuron structure showed that the main mechanism of death was apoptosis. This conclusion was based on the morphological and biochemical changes seen in these neurons. Patients' brains show increases in the number of neurons with signs of DNA fragmentation and forming autophagic vacuoles [114, 135]. However, several researchers have shown that the canonical signal pathways for apoptosis are not the main ones operating. Apoptosis in its canonical version is a quite rapid process, and if it were the main mechanism of death, then neurons showing signs of apoptosis would be significantly less frequent than is in fact the case [141]. On this basis it can be suggested that neuron death in Alzheimer's disease involves nonstandard signal pathways, inducing the same morphological changes in neurons as apoptosis. In addition, the role of programmed forms of necrosis (necroptosis, aponecrosis) have been studied [37].

Pathways increasing the calcium concentration in the neuron cytoplasm. The central component initiating the cellular-molecular mechanisms of neuron death in Alzheimer's disease is an increase in the cytoplasmic calcium concentration [8, 119]. The sources of calcium are the extracellular space and the endoplasmic reticulum (ER).

Increases in the calcium flux from the extracellular space can occur via several pathways. One is the formation of ion channels in neuron membranes by β-amyloid molecules, with passage of calcium ions through these into the neuron cytoplasm (see Fig. 1, *A*) [14, 35, 102]. Another pathway is via the action of soluble β-amyloid oligomers on $α7$ N-cholinoreceptors [52]. This interaction, direct or indirect, evokes functional activity of α 7 N-cholinoreceptors and a calcium ion current through their channels [34, 71, 84]. The mechanism of excitotoxicity may also take part in the increases in neuronal calcium ion concentration. Animal experiments have shown that increased extracellular soluble β-amyloid oligomer levels lead to hyperpolarization of astrocyte membranes, with decreased glutamate reuptake and intracellular accumulation by these cells. An excess extracellular glutamate level leads to extreme excitation of neuron glutamate receptors, increased entry of calcium ions through them, and triggering of pathological intracellular mechanisms [50].

 Release of calcium ions from the endoplasmic reticulum results from stress (tension) on the endoplasmic reticulum (see Fig. 1, *B*) [3, 19]. The cause of stress is the accumulation of protein molecules with impaired secondary and tertiary structure (misfolded proteins) in the ER, i.e., β-amyloid and hyperphosphorylated tau protein. The source of intracellular β-amyloid in cerebral neurons remains to be firmly established. Two sources for β -amyloid accumulation have been considered: production within the neuron itself (see Fig. 1, *C*) and internalization of β-amyloid secreted into the intracellular space by other cells (see Fig. 1, *D*).

Fig. 1. Hypothetical pathways for neuron death in Alzheimer's disease. $A\beta - \beta$ -amyloid; Ca²⁺ – calcium ions; ApoE – apolipoprotein E; LRP1 – low density lipoprotein; ER – endoplasmic reticulum; RyR – ryanodine receptors; IP3-R – inositol triphosphate receptors; ROS – reactive oxygen species; CitC – cytochrome c; AIF – apoptosis-inducing factor; Apaf-1 – apoptosis protease-activating factor; p20 – active fragments of BAP31; BAP31 – B-cell receptor-associated protein; BCL-2 – B-cell lymphoma protein; PKB – protein kinase B; GSK3β – glycogen synthase kinase 3β; UPS – unfolded protein response system; mPTP – mitochondrial permeability transition pores. Neuron death in Alzheimer's disease is due to accumulation of calcium ions in the cytoplasm. Sources of calcium accumulation are the intercellular space (*A*) and endoplasmic reticulum (*B).* Calcium release from the endoplasmic reticulum is induced by stress of the endoplasmic reticulum due to accumulation of β-amyloid molecules with unfolded structure within it. These molecules are produced by the endoplasmic reticulum itself (*C*) or enter the neuron from the extracellular space (*D*). Accumulation of calcium ions in the neuron cytoplasm can activate the classical pathway of apoptosis (*E*). Uptake of calcium ions by mitochondria (*F*) induces mitochondrial dysfunction. Mitochondrial dysfunction can induce apoptosis via several pathways. Cytochrome c released by mitochondria triggers the classical apoptosis pathway via apoptosome formation with Apaf-1 protein and procaspase-9 (*G*). AIF released from mitochondria initiates the caspase-independent apoptosis pathway (*H*). Reactive oxygen species released by mitochondria also initiate apoptosis (*I*). Stress on the neuronal endoplasmic reticulum leads to activation of one of the signal pathways of apoptosis, not associated with calcium accumulation in the cytoplasm (*J*). Mitochondrial dysfunction can be induced by accumulation within them not only of calcium, but also β-amyloid (*K*).

 Intracellular β-amyloid accumulation as a result of its production by the dead neurons themselves is a natural phenomenon, as β-amyloid is formed in neuronal endosomes as a result of proteolysis of amyloid precursor protein (APP) [94]. This occurs when APP molecules are not cleaved by α -secretase on the cell surface [33, 97, 125]. β-Amyloid molecules can then be degraded in lysosomes (autophagy), secreted by axons [28, 63] or dendrites [129], or transferred to other organelles (Golgi complex, endoplasmic reticulum, mitochondria). Inadequate degradation of β-amyloid in neurons is due to weakening of the mechanisms of autophagy (a cellular dysfunction currently regarded as a candidate for the key component in the pathogenesis of Alzheimer's disease) [92].

 Another source for β-amyloid accumulation in the ER is its internalization (uptake from the extracellular space) (see Fig. 1, *D*) [87]. The dominant view is that internalization of

β-amyloid occurs via clathrin-mediated endocytosis on interaction of molecules with LRP1 (low density lipoprotein-related protein 1) receptor (see [87] for review). Interaction of β-amyloid molecules with LRP1 occurs indirectly, via a coreceptor or in complex with apolipoprotein E, for which the LRP1 receptor is also specific. Other internalization hypotheses involve lipid rafts containing ganglioside GM1, α7 N-cholinoreceptors, or receptors for advanced glycation end products (RAGE) [106]. After internalization, β-amyloid is located in neuronal endosomes and lysosomes, where molecules are degraded by proteases or form protease-resistant aggregates which are transferred to the ER and mitochondria.

 In normal conditions, protein molecules with abnormal structure present in the ER and mitochondria are destroyed by a special molecular-cellular system, i.e., the UPR (unfolded protein response). In overtension (stress) of the ER, when the UPR system is ineffective, various signal pathways leading to cell death are activated [19]. Data confirming mobilization of this mechanism in Alzheimer's disease include the presence of incorrectly folded β-amyloid molecules (soluble oligomers of this substance) in the ER, hyperactivation of the neuronal UPR system, and activation of ER stress markers in the brains of dead patients [38, 48, 53]. ER stress leads to activation of several signal pathways for neuronal death. Some of these are initiated by release of calcium ions from the ER via ryanodine and inositol triphosphate receptors [64, 82].

 The β-amyloid precursor protein intracellular domain (AICD) is able to potentiate the release of calcium from the ER [69, 89]. Molecules of this substance are released within the neuron in response to γ -secretase at the second stage of cleavage of the β-amyloid precursor protein (APP) molecule. The activation of the amyloidogenic pathway of APP cleavage in Alzheimer's disease unavoidably leads to an increase in AICD production. AICD molecules functioning as a transcription factor activate the expression of the genes for proteins involved in controlling cellular calcium balance. The inositol phosphate signal pathway activated by this process leads to activation of IP3 receptors in the endoplasmic reticulum and increased release of calcium ions through these receptors to the cytoplasm [73].

Pathways of apoptosis initiated by cytoplasmic calcium. Calcium ions activate proteases of the calpain and caspase-12 families, which trigger the classical caspase cascade of the apoptosis reaction (see Fig. 1, *E*) [19]. Involvement of this mechanism in the pathogenesis of Alzheimer's disease has been confirmed by experimental data. β-Amyloid induces release of calcium from the ER [83]. Apoptosis induced by β-amyloid is mediated by caspase-12 [90].

 Calcium accumulation in the cytoplasm leads to its uptake by mitochondria (see Fig. 1, *F*). Overload of mitochondria with calcium activates mitochondrial permeability transition pores (mPTP), increasing the permeability of the mitochondrial membrane and the release of proapoptotic factors from mitochondria into the cytoplasm: cytochrome c, apoptosis-inducing factor (AIF), and reactive oxygen species [54].

 Cytochrome c interacts with Apaf-1 protein (apoptotic protease-activating factor-1) and procaspase-9 to form molecular complexes, i.e., apoptosomes. The appearance of apoptosomes in the cytoplasm leads to activation of caspase-3 molecules and triggering of the classical apoptosis pathway reaction cascade (see Fig. 1, *G*) [23].

 The suggestion that caspases are involved in neuron death in Alzheimer's disease is supported by data showing increases in their expression and activity in those cerebral neurons in patients' brains displaying morphological changes typical of apoptosis. The process of β-amyloid accumulation and activation of the apoptosis signal pathways occur synchronously and are colocated in the brain. This particularly applies to caspases-2, -3, -6, -7, and -8 [11, 43, 81, 100] or caspase-2 (independently of caspase-3) [110, 123].

512 Mukhin, Pavlov, and Klimenko

 The interaction of β-amyloid and caspases may form a vicious circle (such situations underlie many pathological states): β-amyloid activates caspases and caspases can increase β-amyloid synthesis by activating β-secretase [122].

 An increasingly widely held view is that activation of caspases in Alzheimer's disease is not an element of the process of cell death, but is only associated with functional impairments in neurons [56, 116, 139]. Within the framework of this theory, concepts of the role of the caspase-independent apoptosis pathway in Alzheimer's disease, associated with activation of another product of mitochondrial dysfunction – AIF – has been developed [24, 72]. AIF is a mitochondrial flavoprotein which functions to protect cells against reactive oxygen species [39, 109]. Molecules of the mature form of this substance are produced from a precursor and are located on the internal mitochondrial membrane. Proapoptotic factors, particularly β-amyloid, produce proteolysis (involving calpains) of AIF molecules to form the soluble form of this substance, which enters the neuronal cytoplasm and then the nucleus, where it triggers the process of DNA fragmentation and cell death (see Fig. 1, *H*) [49].

 Reactive oxygen species are produced by mitochondrial dysfunction – they induce oxidative stress in neurons (one of the main signs of Alzheimer's disease). It is another proposed cause of the activation of apoptosis signal pathways (Fig. 1, *I*) [10, 22]. In addition, oxidative stress-induced lipid peroxidation of cell membranes is accompanied by release of phosphatidylserine into the cytoplasm, which facilities the association between β-amyloid and the membrane and formation of β-amyloid ionic channels and the entry of calcium into the cell through these channels [8]. Thus, the formation of β-amyloid channels leading through a chain of events to oxidative stress and oxidative stress promoting the formation of β-amyloid cannels operates as a vicious cycle which can theoretically amplify the pathological process in the neuron.

Signal pathways of apoptosis not linked with calcium accumulation in the cytoplasm. The literature contains descriptions of various apoptosis signal pathways able to lead to neuron death in Alzheimer's disease but not linked with calcium accumulation in the cytoplasm.

 Application of stress to the endoplasmic reticulum initiates apoptosis not only by release of calcium into the cytoplasm, but also by formation of a complex of integral proteins in the ER membrane: BAP31 (or BCAP31 – B-cell receptor-associated protein 31), BCL2 (C-cell lymphoma protein), and caspase-8 (see Fig. 1, *J*). BAP31 undergoes cleavage in this complex, with release of its p20 fragment. p20, acting on mitochondria, increases their membrane permeability and cytochrome c release [23]. In the cytoplasm, cytochrome c can activate the apoptosis signal pathway described above, forming apoptosomes and caspase-3.

 Mitochondrial dysfunction is an obligatory component of neuronal death signal pathways in Alzheimer's disease [17, 140]. It has been suggested that mitochondrial dysfunction in Alzheimer's disease can be elicited not only by

mitochondrial calcium overload, but also the accumulation of structurally damaged β-amyloid molecules within them, coming from the endoplasmic reticulum (see Fig. 1, *K*) [19]. Transport of β-amyloid from the ER into mitochondria probably does not occur over a distance, but within the framework of mitochondrial membrane-associated endoplasmic reticulum [108]. In mitochondria, β-amyloid molecules bind alcohol dehydrogenase molecules, producing impairments to oxidative phosphorylation and ATP synthesis and decreasing membrane potential [25, 79]. The increase in mitochondrial membrane permeability induced by these processes leads to release of cytochrome c, AIF, and reactive oxygen species from mitochondria into the cytoplasm, these being factors triggering the mechanisms of apoptosis described above.

 The proapoptotic actions of intracellular β-amyloid may be mediated by glycogen synthase kinase 3β (GSK3β) [78]. If this is so, β-amyloid monomers will inactivate protein kinase B (PKB) via caspase-3; the active form of PKB inactivates GSK3β. The resulting increase in GSK3β activity may lead to mobilization of apoptosis signal pathways [58].

Conclusions

 Summarizing published data leads to the conclusion that one element in the pathogenesis of Alzheimer's disease is neuron loss in the brain. In those brain areas in which neuron death is seen in normal aging, this process is more intense in Alzheimer's disease (the precentral and postcentral gyri, parietal and temporal associative areas of the cortex, subiculum, and hippocampal field CA1). In addition, the number of neurons starts to decrease in those brain areas where there is essentially no change in normal aging (the dentate gyrus, entorhinal cortex, olfactory bulbs, basal nucleus of Meynert, amygdala, substantia nigra, locus ceruleus, raphe nuclei). Neuron death starts at the early stages of disease development and is one of the causes of the impairments of brain function seen in this disease. The cause of the decrease in the number of neurons is an increase in the probability that they will die, with suppression of the maturation and functional integration of new neurons in the dentate gyrus, along with weakening of postnatal neurogenesis in the subventricular zone and, according to some data, in the dentate gyrus. The cellular-molecular mechanism of neuron death in Alzheimer's disease has not been established. Neurons show elements of apoptosis, though there are data excluding the operation of its canonical mechanisms. The possibility that other, poorly studied mechanisms of cell death operate is under study. The most likely initiator of neuron death is the intracellular accumulation of β-amyloid induced by dysfunction of the endoplasmic reticulum and mitochondria and leading to activation of intracellular caspase-independent apoptosis pathways.

REFERENCES

1. E. D. Bazhanova, V. N. Molodtsov, and K. I. Pavlov, "Changes in the expression of apoptosis-associated molecules in neurosecretory cells of the hypothalamus in mice during aging," *Morfologiya*, **130**, No. 6, 35–39 (2006).

- Yu. B. Belousov, S. K. Zyryanov, D. Yu. Belousov, and A. S. Beketov, "Clinical-economical aspects of the treatment of Alzheimer's disease in Russia," *Kachestv. Klin. Prakt.*, S1, 3–28 (2009).
- 3. I. B. Besprozvannyi, "The calcium signaling system in neurodegeneration," *Acta Nature (Russian version)*, **2**, No. 1, 80–88 (2010).
- 4. S. I. Gavrilov and Ya. B. Kalyn, "Socially mediated factors and the state of mental health in the elderly population," *Vestn. Ros. Akad. Med. Nauk.*, No. 9, 15–20 (2002).
- 5. D. E. Korzhevskii, "Neurogenesis and neural stem cells," *Med. Akad. Zh.*, **10**, No. 4, 175–182 (2010).
- 6. D. E. Korzhevskii, O. V. Kirik, and E. G. Gilerovich, "Postnatal neurogenesis: cell identification and terminology," *Morfologiya*, 144, No. 4, 88–92 (2013).
- 7. V. N. Mukhin, "Pathogenetic mechanisms of dysfunction of the cholinergic system in Alzheimer's disease," *Ros. Fiziol. Zh.*, **99**, No. 7, 793–804 (2013).
- 8. E. A. Popugaeva, O. L. Vlasova, and I. B. Besprozvannyi, "The role of intracellular calcium in the development of the pathogenesis of Alzheimer's disease," *Nauchno-Tekhn. Ved. St. Peterburg. Univ. Fiz. Mat. Nauki*, **189**, No. 1, 79–90 (2014).
- 9. J. B. Aimone, J. Wiles, and F. H. Gage, "Potential role for adult neurogenesis in the encoding of time in new memories," *Nature Neurosci.*, **9**, No. 6, 723–727 (2006).
- 10. A. D. Buttelfield, A. Castegna, C. M. Lauderback, and J. Drake, "Evidence that amyloid beta-peptide-induced lipid peroxidation and its sequelae in Alzheimer's disease brain contribute to neuronal death," *Neurobiol. Aging*, **23**, No. 5, 655–664 (2002).
- 11. J. W. Allen, B. A. Eldadah, X. Huang, et al., "Multiple caspases are involved in β-amyloid-induced neuronal apoptosis," *J. Neurosci. Res.*, **65**, No. 1, 45–53 (2001).
- 12. B. B. Andersen, L. Korbo, and B. Pakkenberg, "A quantitative study of the human cerebellum with unbiased stereological techniques," *J. Comp. Neurol.*, **326**, No. 4, 549–560 (1992).
- 13. T. Arendt, V. Bigl, A. Arendt, and A. Tennstedt, "Loss of neurons in the nucleus basalis of Meynert in Alzheimer' s disease, paralysis agitans and Korsakoff's disease," *Acta Neuropathol.*, **61**, No. 2, 101– 108 (1983).
- 14. N. Arispe, H. B. Pollard, and E. Rojas, "β-Amyloid Ca2+-channel hypothesis for neuronal death in Alzheimer disease," *Mol. Cell*. *Biochem.*, **140**, No. 2, 119–125 (1994).
- 15. F. A. C. Azevedo, L. R. B. Carvalho, L. T. Grinberg, et al., "Equal numbers of neuronal and nonneuronal cells make the human brain an isometrically scaled-up primate brain," *J. Comp. Neurol.*, **513**, No. 5, 532–541 (2009).
- 16. M. J. Bali, "Neuronal loss, neurofibrillary tangles and granulovacuolar degeneration in the hippocampus with ageing and dementia," *Acta Neuropathol.*, **37**, No. 2, 111–118 (1977).
- 17. M. F. Beal, "Mitochondria take center stage in aging and neurodegeneration," *Ann. Neurol.*, **58**, No. 4, 495–505 (2005).
- 18. O. Bergmann, J. Liebl, S. Bernard, et al., "The age of olfactory bulb neurons in Humans," *Neuron*, **74**, No. 4, 634–639 (2012).
- 19. S. Bernales, M. A. Morales Soto, and E. McCullagh, "Unfolded protein stress in the endoplasmic reticulum and mitochondria: a role in neurodegeneration," *Front. Aging Neurosci.*, **4**, 5 (2012).
- 20. K. Boekhoorn, M. Joels, and P. J. Lucassen, "Increased proliferation reflects glial and vascular-associated changes, but not neurogenesis in the presenile Alzheimer hippocampus," *Neurobiol. Dis.*, **24**, No. 1, 1–14 (2006).
- 21. W. Bondareff, C. Q. Mountjoy, and M. Roth, "Loss of neurons of origin of the adrenergic projection to cerebral cortex (nucleus locus ceruleus) in senile dementia," *Neurology*, **32**, No. 2, 164–168 (1982).
- 22. M. A. Bradley-Whitman and M. A. Lovell, "Biomarkers of lipid peroxidation in Alzheimer disease (AD), an update," *Arch. Toxicol.*, **89**, No. 7, 1035–1044 (2015).
- 23. D. E. Bredesen, R. V. Rao, and P. Mehlen, "Cell death in the nervous system," *Nature*, **443**, No. 7113, 796–802 (2006).
- 24. A. Camins, M. Pallas, and J. S. Silvestre, "Apoptotic mechanisms involved in neurodegenerative diseases: Experimental and therapeutic approaches," *Methods Find. Exp. Clin. Pharmacol.*, **30**, No. 1, 43 (2008).
- 25. C. Caspersen, N. Wang, J. Yao, et al., "Mitochondrial Abeta: a potential focal point for neuronal metabolic dysfunction in Alzheimer's disease," *FASEB J.*, **19**, No. 14, 2040–2041 (2005).
- 26. S. Chen, J. M. Wang, R. W. Irwin, et al., "Allopregnanolone promotes regeneration and reduces β-amyloid burden in a preclinical model of Alzheimer's disease," *PLoS One*, **6**, No. 8, e24293 (2011).
- 27, T. T. Chuang, "Neurogenesis in mouse models of Alzheimer's disease," *Biochem. Biophys. Acta Mol. Basis Dis.*, **1802**, No. 10, 872–880 (2010).
- 28. J. R. Cirrito, K. A. Yamada, M. B. Finn, et al., "Synaptic activity regulates interstitial β-amyloid-beta levels in vivo," *Neuron*, **48**, No. 6, 913–922 (2005).
- 29. A. R. Cole, A. Knebel, N. A. Morrice, et al., "GSK-3 phosphorylation of the alzheimer epitope within collapsin response mediator proteins regulates axon elongation in primary neurons," *J. Biol. Chem.*, **279**, No. 48, 50 176–50 180 (2004).
- 30. P. D. Coleman and D. G., Flood, "Neuron numbers and dendritic extent in normal aging and Alzheimer's disease," *Neurobiol. Aging*, **8**, No. 6, 521–545 (1987).
- 31. C. M. Cooper-Kuhn, J. Winkler, and H. G. Kuhn, "Decreased neurogenesis after cholinergic forebrain lesion in the adult rat," *J. Neurosci. Res.*, **77**, No. 2, 155–165 (2004).
- 32. W. Deng, M. D. Saxe, I. S. Gallina, and F. H. Gage, "Adult-born hippocampal dentate granule cells undergoing maturation modulate learning and memory in the brain," *J. Neurosci.*, **29**, No. 43, 13532–13542 (2009).
- 33. B. De Strooper, R. Vassar, and T. Golde, "The secretases: enzymes with therapeutic potential in Alzheimer disease," *Nat. Rev. Neurol.*, **6**, No. 2, 99–107 (2010).
- 34. K. T. Dineley, M. Westerman, D. Bui, et al., "β-Amyloid activates the mitogen-activated protein kinase cascade via hippocampal α7 nicotinic acetylcholine receptors: in vitro and in vivo mechanisms related to Alzheimer's disease," *J. Neurosci.*, **21**, No. 12, 4125–4133 (2001).
- 35. C. Di Scala, J-D. Troadec, C. Lelievre, et al., "Mechanism of cholesterol-assisted oligomeric channel formation by a short Alzheimer β-amyloid peptide," *J. Neurochem.*, **128**, No. 1, 186–195 (2014).
- 36. A. Ernst, K. Alkass, S. Bernard, et al., "Neurogenesis in the Striatum of the adult human brain," *Cell*, **156**, No. 5, 1072–1083 (2014).
- 37. S. M. Fayaz, V. S. Suvanish Kumar, and G. K. Rajanikant, "Necroptosis: Who knew there were so many interesting ways to die?" *CNS Neurol. Disord. Drug Targets*, **13**, No. 1, 42–51 (2014).
- 38. P. Fernández-Vizarra, A. P. Fernández, S. Castro-Blanco, et al., "Intra and extracellular Abeta and PHF in clinically evaluated cases of Alzheimer's disease," *Histol. Histopathol.*, **19**, No. 3, 823–844 (2004).
- 39. P. Ferreira, R. Villanueva, L. A. Cabon, et al., "The oxido-reductase activity of the apoptosis inducing factor: A promising pharmacological tool?" *Curr. Pharm. Des.*, **19**, No. 14, 2628–2636 (2013).
- 40. A. M. Fjell and K. B. Walhovd, "Structural brain changes in aging: courses, causes and cognitive consequences," *Rev. Neurosci.*, **21**, No. 3, 187–222 (2011).
- 41. Y. Fukata, T. J. Itoh, T. Kimura, et al., "CRMP-2 binds to tubulin heterodimers to promote microtubule assembly," *Nature Cell Biol.*, **4**, No. 8, 583–591 (2002).
- 42. J. J. Garrido, D. Simón, O. Varea, and F. Wandosell, "GSK3 alpha and GSK3 beta are necessary for axon formation," *FEBS Lett.*, **581**, No. 8, 1579–1586 (2007).
- 43. M. C. Gastard, J. C. Troncoso, and V. E. Koliatsos, "Caspase activation in the limbic cortex of subjects with early Alzheimer's disease," *Ann. Neurol.*, **54**, No. 3, 393–398 (2003).
- 44. P. Giannakopoulos, F. R. Herrmann, T. Bussiere, et al., "Tangle and neuron numbers, but not amyloid load, predict cognitive status in Alzheimer's disease," *Neurology*, **60**, No. 9, 1495–1500 (2003).
- 45. T. Gómez-Isla, J. L. Price, D. W. McKeel, Jr., et al., "Profound loss of layer II entorhinal cortex neurons occurs in very mild Alzheimer's disease," *J. Neurosci.*, **16**, No. 14, 4491–4500 (1996).
- 46. C. Göritz and J. Frisén, "Neural stem cells and neurogenesis in the adult," *Cell Stem Cell*, **10**, No. 6, 657–659 (2012).
- 47. E. Gould, A. Beylin, P. Tanapat, et al., "Learning enhances adult neurogenesis in the hippocampal formation," *Nature Neurosci.*, **2**, No. 3, 260–265 (1999).
- 48. G. K. Gouras, J. Tsai, J. Naslund, et al., "Intraneuronal Af342 accumulation in human brain," *Am. J. Pathol.*, **156**, No. 1, 15–20 (2000).
- 49. E. Hangen, K. Blomgren, P. Bénit, et al., "Life with or without AIF," *Trends Biochem. Sci.*, **35**, No. 5, 278–287 (2010).
- 50. T. Harkany, I. Ábraham, W. Timmerman, et al., "β-Amyloid neurotoxicity is mediated by a glutamate-triggered excitotoxic cascade in rat nucleus basalis," *Eur. J. Neurosci.*, **12**, No. 8, 2735–2745 (2000).
- 51. N. J. Haughey, A. Nath, S. L. Chan, et al., "Disruption of neurogenesis by amyloid β-peptide, and perturbed neural progenitor cell homeostasis, in models of Alzheimer's disease," *J. Neurochem.*, **83**, No. 6, 1509–1524 (2002).
- 52. C. M. Hernandez, and K. T. Dineley, "α7 Nicotinic acetylcholine receptors in Alzheimer's disease: neuroprotective, neurotrophic or both?" *Curr. Drug Targets*, **13**, No. 5, 613–622 (2012).
- 53. J. J. M. Hoozemans, R. Veerhuis, E. S. V. Haastert, et al., "The unfolded protein response is activated in Alzheimer' s disease," *Acta Neuropathol.*, **110**, No. 2, 165–172 (2005).
- 54. J. Hroudova, N. Singh, and Z. Figar, "Mitochondrial dysfunctions in neurodegenerative diseases: Relevance to Alzheimer's disease," *BioMed Res. Int.*, 2014, e175062 (2014).
- 55. E.-M. Hur and F.-Q. Zhou, "GSK3 signalling in neural development," *Nat. Rev. Neurosci.*, **11**, No. 8, 539–551 (2010).
- 56. B. T. Hynan, "Caspase activation without apoptosis: insight into Aβ initiation of neurodegeneration," $Nat. Neurosci., 14, No. 1, 5–6 (2011).$
- 57. R. W. Irwin, J. M. Wang, S. Chen, and R. D. Brinton, "Neuroregenerative mechanisms of allopregnanolone in Alzheimer's disease," *Front. Endocrinol. (Lausanne)*, **12**, No. 2, 117 (2012).
- 58. K. M. Jacobs, S. R. Bhave, D. J. Ferraro, et al., "GSK-3β, a bifunctional role in cell death pathways," *Int. J. Cell Biol.*, **2012**, 2012, e930710 (2012).
- 59. T. Jaworski, S. Kügler, and F. Van Leuven, "Modeling of tau-mediated synaptic and neuronal degeneration in Alzheimer's disease," *Int. J. Alzheimer's Dis.*, **2010**: 1–10 (2010).
- 60. H. Jiang, W. Guo, X. Liang, and Y. Rao, "Both the establishment and the maintenance of neuronal polarity require active mechanisms: Critical roles of GSK-3β and its upstream regulators," *Cell*, **120**, No. 1, 123–135 (2005).
- 61. K. Jin, A. L. Peel, X. O. Mao, et al., "Increased hippocampal neurogenesis in Alzheimer's disease," *Proc. Natl. Acad. Sci. USA*, **101**, No. 1, 343–347 (2004).
- 62. G. V. Johnson and W. H. Stoothoff, "Tau phosphorylation in neuronal cell function and dysfunction," *J. Cell Sci.*, **117**, No. 24, 5721–5729 (2004) .
- 63. F. Kamenetz, T. Tomita, H. Hsieh, et al., "APP processing and synaptic function," *Neuron*, **37**, No. 6, 925–937 (2003).
- 64. M. Kelliher, J. Fastbom, R. F. Cowburn, et al., "Alterations in the ryanodine receptor calcium release channel correlate with Alzheimer's disease neurofibrillary and β-amyloid pathologies," *Neuroscience*, **92**, No. 2, 499–513 (1999).
- 65. G. Kempermann and F. H., Gage, "New nerve cells for the adult brain," *Sci. Am.*, **280**, No. 5, 48–53 (1999).
- 66. S. Knafo, "Amygdala in Alzheimer's disease," in: *The Amygdala – a Discrete Multitasking Manager*, B. Ferry (ed.), InTech (2012), pp. 375–384.

514 Mukhin, Pavlov, and Klimenko

- 67. R. Knoth, L. Singec, M. Ditter, et al., "Murine features of neurogenesis in the human hippocampus across the lifespan from 0 to 100 years," *PLoS One*, 5 (1), e8809 (2010).
- 68. A. J. Kole, R. P. Annis, and M. Deshmukh, "Mature neurons: equipped for survival," *Cell Death Dis.*, **4** No. 6, e689 (2013).
- 69. U. Konietzko, "AICD nuclear signaling and its possible contribution to Alzheimer's diseasem" *Curr. Alzheimer Res.*, **9**, No. 2, 200–216 (2012).
- 70. A. Kremer, J. V. Louis, T. Jaworski, and F. Van Leuven, "GSK3 and Alzheimer's disease: facts and fiction," *Front. Mol. Neurosci.*, No. 4 (2011).
- 71. P. Kurup, Y. Zhang, J. Xu, et al., "Aβ-Mediated NMDA receptor endocytosis in Alzheimer's disease involves ubiquitination of the tyrosine phosphatase STEP61," *J. Neurosci.*, **30**, No. 17, 5948–5957 (2010).
- 72. J.-H. Lee, Y.-H. Cheon, R.-S. Woo, et al., "Evidence of early involvement of apoptosis inducing factor-induced neuronal death in Alzheimer brain," *Anatomy Cell Biol.*, **45**, No. 1, 26 (2012).
- 73. M. A. Leissring, M. P. Murphy, T. R. Mead, et al., "A physiologic signaling role for the y-secretase-derived intracellular fragment of APP," *Proc. Natl. Acad. Sci. USA*, **99**, No. 7, 4697–4702 (2002).
- 74. K. Leroy, Z. Yilmaz, and J.-P. Brion, "Increased level of active GSK-3β in Alzheimer's disease and accumulation in argyrophilic grains and in neurones at different stages of neurofibrillary degeneration," *Neuropathol. Appl. Neurobiol.*, **33**, No. 1, 43–55 (2007).
- 75. B. Li, H. Yamamori, Y. Tatebayashi, et al., "Failure of neuronal maturation in Alzheimer disease dentate gyrus," *J. Neuropathol. Exp. Neurol.*, **67**, No. 1, 78–84 (2008).
- 76. L. Lossi and A. Merighi, "In vivo cellular and molecular mechanisms of neuronal apoptosis in the mammalian CNS," *Progr. Neurobiol.*, **69**, No. 5, 287–312 (2003).
- 77. J. P. Magnusson, C. Göritz, J. Tatarishvili, et al., "A latent neurogenic program in astrocytes regulated by Notch signaling in the mouse," *Science*, **346**, No. 6206, 237–241 (2014).
- 78. J. Magrané, K. M. Rosen, R. C. Smith, et al., "Intraneuronal β-amyloid expression downregulates the Akt survival pathway and blunts the stress response," *J. Neurosci.*, **25**, No. 47, 10 960–10 969 (2005).
- 79. M. Manczak, T. S. Anekonda, E. Henson, et al., "Mitochondria are a direct site of Af3 accumulation in Alzheimer's disease neurons: implications for free radical generation and oxidative damage in disease progression," *Hum. Mol. Genet.*, **15**, No. 9, 1437–1449 (2006).
- 80. C. E. Marx, W. T. Trost, L. J. Shampine, et al., "The neurosteroid allopregnanolone is reduced in prefrontal cortex in Alzheimer's disease," *Biol. Psychiatry*, **60**, No. 12, 1287–1294 (2006).
- 81. T. Matsui, K. Ramasamy, M. Ingelsson, et al., "Coordinated expression of caspase 8, 3 and 7 mRNA in temporal cortex of Alzheimer disease: Relationship to formic acid extractable Aβ42 levels," *J. Neuropathol. Exp. Neurol.*, **65**, No. 5, 508–515 (2006).
- 82. M. P. Mattson, "Pathways towards and away from Alzheimer's disease," *Nature*, **430**, No. 7000, 631–639 (2004).
- 83. M. P. Mattson, B. Cheng, D. Davis, et al., "Beta-amyloid peptides destabilize calcium homeostasis and render human cortical neurons vulnerable to excitotoxicity," *J. Neurosci.*, **12**, No. 2, 376–389 (1992).
- 84. T. K. Mehta, J. J. Dougherty, J. Wu, et al., "Defining pre-synaptic nicotinic receptors regulated by beta amyloid in mouse cortex and hippocampus with receptor null mutants," *J. Neurochem.*, **109**, No. 5, 1452–1458 (2009).
- 85 C. C. Meltzer, G. Smith, S. T. DeKosky, et al., "Serotonin in aging, late-life depression, and Alzheimer's disease: The emerging role of functional imaging," *Neuropsychopharmacology*, **18**, No. 6, 407–430 (1998).
- 86. A. K. H. Miller, R. L. Alston, C. Q. Mountjoy, and J. A. N. Corsellis, "Automated differential cell counting on a sector of the normal human hippocampus: The influence of age," Neuropathol. Appl. Neu*robiol.*, **10**, No. 2, 123–141 (1984).
- 87. A. Mohamed and F. Posse de Chaves, "Aβ internalization by neurons and glia," *Int. J. Alzheimer's Disease*, **2011**, 1–17 (2011).
- 88. J. H. Morrison and P. R. Hof, "Life and death of neurons in the aging brain," *Science*, **278**, No. 5337, 412–419 (1997).
- 89. T. Müller, H. E. Meyer, R. Egensperger, and K. Marcus, "The amyloid precursor protein intracellular domain (AICD) as modulator of gene expression, apoptosis, and cytoskeletal dynamics – Relevance for Alzheimer's disease," *Progr. Neurobiol.*, **85**, No. 4, 393–406 (2008).
- 90. T. Nakagawa, H. Zhu, N. Morishima, et al., "Caspase-12 mediates endoplasmic-reticulum-specific apoptosis and cytotoxicity by amyloid-β," *Nature*, **403**, No. 6765, 98–103 (2000).
- 91. L. Lossi and A. Merighi (eds.), *Neuronal Cell Death: An Overview of Its Different Forms in Central and Peripheral Neurons*, Springer, New York (2015).
- 92. R. A. Nixon and D.-S. Yang, "Autophagy failure in Alzheimer's disease-locating the primary defect," *Neurobiol. Dis.*, **43**, No. 1, 38–45 (2011).
- 93. S. C. Noctor, V. Martinez-Cerdeño, and A. R. Kriegstein, "Contribution of intermediate pronitor cells to cortical histogenesis," *Arch. Neurol.*, **64**, No. 5, 639–642 (2007).
- 94. R. J. O'Brien and P. C. Wong, "Amyloid precursor protein processing and Alzheimer's disease," *Annu. Rev. Neurosci.*, **34**, 185–204 (2011).
- 95. B. Pakkenberg and H. J. G. Gundersen, "Neocortical neuron number in humans: Effect of sex and age," *J. Comp. Neurol.*, **384**, No. 2, 312–320 (1997).
- 96. B. Pakkenberg, D. Pelvig, L. Marner, et al., "Aging and the human neocortex," *Exp. Gerontology*, **38**, No. 1–2, 95–99 (2003).
- 97. M. S. Parihar and G. J. Brewer, "Amyloid beta as a modulator of synaptic plasticity," *J. Alzheimers Dis.*, **22**, No. 3, 741–763 (2010).
- 98. J. J. Pei, E. Braak, H. Braak, et al., "Distribution of active glycogen synthase kinase 3beta (GSK-3beta) in brains staged for Alzheimer disease neurofi brillary changes," *J. Neuropathol. Exp. Neurol.*, **58**, No. 9, 1010–1019 (1999).
- 99. D. P. Pelvig, H. Pakkenberg, A. K. Stark, and B. Pakkenberg, "Neocortical glial cell numbers in human brains," *Neurobiol. Aging*, **29**, No. 11, 1754–1762 (2008).
- 100. P. N. Pompl, S. Yemul, Z. Xiang, et al., "Caspase gene expression in the brain as a function of the clinical progression of Alzheimer disease," *Arch. Neurol.*, **60**, No. 3, 369–376 (2003).
- 101. J. L. Price, A. I. Ko, M. J. Wade, et al., "Neuron number in the entorhinal cortex and ca1 in preclinical Alzheimer disease," *Arch. Neurol.*, **58**, No. 9, 1395–1402 (2001).
- 102. A. Quist, I. Doudevski, H. Lin, et al., "Amyloid ion channels: A common structural link for protein-misfolding disease," *Proc. Natl. Acad. Sci. USA*, **102**, No. 30, 10,427–10,432 (2005).
- 103. C. A. Raji, O. L. Lopez, L. H. Kuller, et al., "Age, Alzheimer disease, and brain structure," *Neurology*, **73**, No. 22, 1899–1905 (2009).
- 104. C. G. Rasool, C. N. Svendsen, and D. J. Selkoe, "Neurofibrillary degeneration of cholinergie and noncholinergic neurons of the basal forebrain in Alzheimer's disease," *Ann. Neurol.*, **20**, No. 4, 482–488 (1986).
- 105. J. J. Rodriguez and A. Verkhratsky, "Neurogenesis in Alzheimer's disease," *J. Anat.*, **219**, No. 1, 78–89 (2011).
- 106. L. Saavedra, A. Mohamed, V. Ma, et al., "Internalization of β-amyloid peptide by primary neurons in the absence of apolipoprotein E," *J. Biol. Chem.*, **282**, No. 49, 35 722–35 732 (2007).
- 107. N. Sanai, T. Nguyen, R. A. Ihrie, et al., "Corridors of migrating neurons in the human brain and their decline during infancy," *Nature*, **478**, No. 7369, 382–386 (2011).
- 108. E. A. Schon and E. Area-Gomez, "Mitochondria-associated ER membranes in Alzheimer disease," *Mol. Cell. Neurosci.*, **55**, 26–36 (2013).
- 109. I. F. Sevrioukova, "Apoptosis-inducing factor: Structure, function, and redox regulation," *Antioxid. Redox Signal.*, **14**, No. 12, 2545–2579 (2010).
- 110. S. Shimohama, H. Tanino, and S. Fujimoto, "Changes in caspase expression in Alzheimer's disease: comparison with development and aging," *Biochem. Biophys. Res. Commun.*, **256**, No. 2, 381–384 (1999).
- 111. S.-H. Shi, T. Cheng, L. Jan, and Y.-N. Jan, "APC and GSK-3β are involved in mPar3 Targeting to the nascent axon and establishment of neuronal polarity," *Curr. Biol.*, **14**, No. 22, 2025–2032 (2004).
- 112. T. J. Shors, D. A. Townsend, M. Zhao, et al., "Neurogenesis may relate to some but not an types of hippocampal-dependent learning," *Hippocampus*, **12**, No. 5, 578– 584 (2002).
- 113. G. Šimić and N. Bogdanović, "Volume and number of neurons of the human hippocampal formation in normal aging and Alzheimer's disease," *J. Comp. Neurol.*, **379**, No. 4, 482–494 (1997).
- 114. G. Smale, N. R. Nichols, D. R. Brady, et al., "Evidence for apoptotic cell death in Alzheimer's disease," *Exp. Neurology*, **133**, No. 2, 225– 230 (1995).
- 115. K. J. Smillie and M. A. Cousin, "The role of GSK3 in presynaptic function," *Int. J. Alzheimer Dis.* (2011), doi: 10.4061/2011/263673, www.hindawi.com/journals/journals/ijad/2011/263673, publ. March 14, 2011, acces. May 27, 2013.
- 116. S. Snigdha, E. D. Smith, G. A. Prieto, and C. W. Cotman, "Caspase-3 activation as a bifurcation point between plasticity and cell death," *Neurosci. Bull.*, **28**, No. 1, 14–24 (2012).
- 117. K. L. Spalding, O. Bergmann, K. Alkass, et al., "Dynamics of hippocampal neurogenesis in adult humans," *Cell*, **153**, No. 6, 1219–1227 (2013).
- 118. W. Sun, A. Winseck, S. Vinsant, et al., "Programmed cell death of adult-generated hippocampal neurons is mediated by the proapoptotic gene bax," *J. Neurosci.*, **24**, No. 49, 11205–11213 (2004).
- 119. C. Supnet and I. Bezprozvanny, "The dysregulation of intracellular calcium in Alzheimer disease," *Cell Calcium*, **47**, No. 2, 183–189 (2010).
- 120. H. J. Ter Laak, K. Renkawek, and F. P. van Workum, "The olfactory bulb in Alzheimer disease: a morphologic study of neuron loss, tangles, and senile plaques in relation to olfaction," *Alzheimer Dis. Assoc. Disord.*, **8**, No. 1, 38–48 (1994).
- 121. R. D. Terry, R. DeTeresa, and L. A. Hansen, "Neocortical cell counts in normal human adult aging," *Ann. Neurol.*, **21**, No. 6, 530–539 (1987).
- 122. G. Tesco, Y. H. Koh, E. L. Kang, et al., "Depletion of GGA3 stabilizes BACE and enhances β-secretase activity," *Neuron*, **54**, No. 5, 721–737 (2007).
- 123. C. M. Troy, S. A. Rabacchi, W. J. Friedman, et al., "Caspase-2 mediates neuronal cell death induced by β-amyloid," *J. Neurosci.*, **20**, No. 4, 1386–1392 (2000).
- 124. T. H. L. G. Vereecken, O. J. M. Vogels, and R. Nieuwenhuys, "Neuron loss and shrinkage in the amygdala in Alzheimer' s disease," *Neurobiol. Aging*, **15**, No. 1, 45–54 (1994).
- 125. B. Vincent and F. Checler, "α-Secretase in Alzheimers disease and beyond: mechanistic, regulation and function in the shedding of membrane proteins," *Curr. Alzheimer Res.*, **9**, No. 2, 140–156 (2012).
- 126. J. M. Wang, P. B. Johnston, B. G. Bail, and R. D. Brinton, "The neurosteroid allopregnanolone promotes proliferation of rodent and human neural progenitor cells and regulates cell-cycle gene and protein expression," *J. Neurosci.*,**25**, No. 19, 4706–4718 (2005).
- 127. J. M. Wang, C. Singh, L. Liu, et al., "Allopregnanolone reverses neurogenic and cognitive deficits in mouse model of Alzheimer's disease," *Proc. Natl. Acad. Sci. USA*, **107**, No. 14, 6498–6503 (2010).
- 128. D. Weinshenker, "Functional consequences of locus coeruleus degeneration in Alzheimer's disease," *Curr. Alzheimer Res.*, **5**, No. 3, 342–345 (2008).
- 129. W. Wei, L. N. Nguyen, H. W. Kessels, et al., "Amyloid beta from axons and dendrites reduces local spine number and plasticity," *Nature Neurosci.*, **13**, No. 2, 190–196 (2010).
- 130. M. J. Wes and H. J. G. Gundersen, "Unbiased stereological estimation of the number of neurons in the human hippocampus," *J. Comp. Neurol.*, **296**, No. 1, 1–22 (1990).
- 131. P. J. Whitehouse, D. L. Price, A. W. Clark, et al., "Alzheimer disease: Evidence for selective loss of cholinergic neurons in the nucleus basalis," *Ann. Neurol.*, **10**, No. 2, 122–126 (1981).
- 132. P. J. Whitehouse, D. L. Price, R. G. Struble, et al., "Alzheimer's disease and senile dementia: loss of neurons in the basal forebrain," *Science*, **215**, 1237–1239 (1982).
- 133. R. W. Williams and K. Herrup, "The control of neuron number," *Annu. Rev. Neurosci.*, **11**, No. 1, 423–453 (1988).
- 134. L. Wiskott, M. J. Rasch, and G. Kempermann, "A functional hypothesis for adult hippocampal neurogenesis: Avoidance of catastrophic interference in the dentate gyrus," *Hippocampus*, **16**, No. 3, 329–343 (2006).
- 135. T. Yamatsuji, T. Matsui, T. Okamoto, et al., "G Protein-mediated neuronal DNA fragmentation induced by familial Alzheimer' s disease-associated mutants of APP," *Science*, **272**, No. 5266, 1349–1352 (1996).
- 136. T. Yoshimura, Y. Kawano, N. Arimura, et al., "GSK-3β regulates phosphorylation of CRMP-2 and neuronal polarity," *Cell*, **120**, No. 1, 137–149 (2005).
- 137. C. Zarow, S. A. Lyness, J. A. Mortimer, and H. C. Chui, "Neuronal loss is greater in the locus coeruleus than nucleus basalis and substantia nigra in Alzheimer and Parkinson diseases," *Arch. Neurol.*, **60**, No. 3, 337–341 (2003).
- 138. L. Zhang, G. Kokkonen, and G. S. Roth, "Identification of neuronal programmed cell death in situ in the striatum of normal adult rat brain and its relationship to neuronal death during aging," *Brain Res.*, **677**, No. 1, 177–179 (1995).
- 139. Y. Zhang, "Caspases in Alzheimer's disease," in: *Neurodegenerative Diseases*, U. Kishore (ed.), InTech (2013), pp. 125–150 (2013).
- 140. X. Zhu, G. Perry, M. A. Smith, and X. Wang, "Abnormal mitochondrial dynamics in the Pathogenesis of Alzheimer's disease," *J. Alzheimers Dis.*, **33**, Suppl. 1, S253–S262 (2013).
- 141. X. Zhu, A. K. Raina, G. Perry, and M. A. Smith, "Apoptosis in Alzheimer disease: A mathematical improbability," *Curr. Alzheimer Res.*, **3**, No. 4, 393–396 (2006).
- 142. I. Ziabreva, E. Perry, R. Perry, et al., "Altered neurogenesis in Alzheimer's disease," *J. Psychosom. Res.*, **61**, No. 3, 311–316 (2006).
- 143. J. Zumbrunn, K. Kinoshita, A. Hyman, and I. S. Näthke, "Binding of the adenomatous polyposis coli protein to microtubules increases microtubule stability and is regulated by GSK3β phosphorylation," *Curr. Biol.*, **11**, No. 1, 44–49 (2001).

516 Mukhin, Pavlov, and Klimenko