

Aspergillosis of Biliary Tract After Liver Transplantation: A Case Report

Chen Yuchong · Zhu Dingheng · Yuan Zhizhong ·
Yu Hongyu · He Jing · Chen Jianghan

Received: 15 July 2009 / Accepted: 8 March 2010 / Published online: 24 March 2010
© Springer Science+Business Media B.V. 2010

Abstract We reported a case of aspergillosis presented as cholangitis in a patient after liver transplantation, even with prophylactic use of fluconazole. The patient had multiple predisposing factors, such as leukocytopenia, immunosuppressive drug therapy. He died 2 days after an exploratory laparotomy was carried out. The histopathologic finding of biopsy specimen from biliary tract was positive for *Aspergillus*. And the pathogen was identified as *Aspergillus flavus* by mycological culture and PCR. The patient was confirmed as a case of Aspergillosis of biliary tract that was responded poorly to fluconazole. This indicated that azole should have been switched to more effective antifungal agents at earlier stage when the patients responded poorly to the original treatment.

Keywords Aspergillosis · Biliary tract · Organ transplantation · Leukocytopenia

Introduction

Aspergillus species have emerged as important causes of morbidity and mortality in immunocompromised patients [1–3]. *Aspergillus fumigatus* is the most common species recovered from cases of invasive aspergillosis [4]. The next most commonly recovered species are *Aspergillus flavus*, *Aspergillus niger*, and *Aspergillus terreus* [5]. Fungal infections of the biliary tract are uncommon and usually caused by *Candida* spp., which have been reported to cause biliary obstruction and cholangitis in the immunocompetent host [6–8]. Here, we report a case of Aspergillosis of biliary tract caused by *A. flavus* in a patient after liver transplantation.

Case Report

History

A 55-year-old man was admitted to the hospital with a 2-year history of recurrent episodes of debilitation, loss of appetite, epigastric discomfort, and dark urine and was diagnosed as chronic viral hepatitis type B. Two months prior to the admission, he developed hepatic encephalopathy and hepatorenal syndrome.

C. Yuchong · Z. Dingheng · C. Jianghan (✉)
Department of Dermatology and Mycology Center,
Changzheng Hospital, Second Military Medical
University, 415 Fengyang Road, 200003 Shanghai, China
e-mail: chenjianghan@126.com

Y. Zhizhong
VIP Ward, Changzheng Hospital, Second Military
Medical University, 415 Fengyang Road,
200003 Shanghai, China

Y. Hongyu · H. Jing
Department of Pathology, Changzheng Hospital,
Second Military Medical University, 415 Fengyang Road,
200003 Shanghai, China

During the admission, his condition deteriorated because of gastric perforation. The patient recovered after supportive treatment and antibiotics. But the liver function worsened consequently. After 3 months admission, he was referred for liver transplantation. The operation was successful. He was continuously treated with mycophenolate mofetil, prednisone, sulperazone, vancomycin, fluconazole as immunosuppressive and prophylactic drugs. The renal and hepatic function recovered gradually. After 24 days transplantation, the patient developed high fever, stomachache, abdominal tenderness abruptly. Diagnostic puncture showed bleeding in abdominal cavity. His conditions deteriorated quickly even after using a hemostatics and a transfusion treatment. An emergency exploratory laparotomy was then carried out. The patient died 2 days later.

Clinical Findings

The cavity was filled with about 700 ml bloody ascites. A large deposit of sludged blood appeared below the right lobe of liver with a slight odor. Transplanted liver was in situ with slight cholestasis. The bleeding was from porta hepatis that had a vomica with yellow staining after debriding the hematocele and adhesions. The common bile duct (2 cm upper and 1.5 cm lower anastomotic stoma) showed caseous necrosis, and the bile was leaking from the necrosis site. Small vessels around the biliary fistula showed active bleeding. The hepatic artery stoma, portal vein stoma, and caval vein stoma of transplantation operation were normal. A piece of common bile duct was biopsied for histopathological, mycological, and microbiological examinations. Ascites and bile were also sent for mycological and microbiological tests.

Investigations

Histopathologic Findings

There were 3–6 μm wide septate hyphae that typically branch at acute angles. A morphological yellowish mass around the hyphae were bile (Fig. 1).

Mycological Findings

Tissue from the bile was plated on Sabouraud dextrose agar with chloramphenicol and incubated

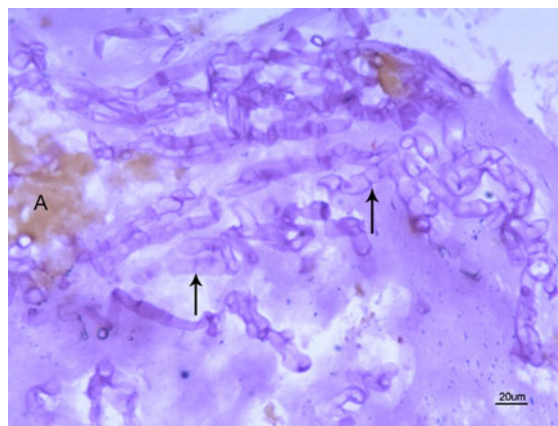


Fig. 1 Histopathology of the biopsy tissue. A bile juice; arrows showed the septate hyphae (PAS stain, $\times 400$)

at 25°. A filamentous fungus was recovered. Then, the colony was transferred to Czapek agar and malt extract agar (MEA), which was cultured at 25°. The strain showed good growth. Colonies on Czapek agar at 25° attained a diameter of 3 cm within 7 days and consisted of a dense felt of yellow–green conidiphores. Colonies on MEA grew faster otherwise, their characteristics were similar to those of colonies grown on Czapek agar.

Identification of the Pathogen

Mycological Morphology The conidial heads typically radiated to nearly globose. There were two kinds of vesicles which were nearly globose, fertile over almost the entire surface. The big one was up to 22.5–50 μm in diameter with sterigmata in two series, and the small one with a single row of phialides, 12.5–35 μm in diameter. Conidia were globose to subglobose, 3.0–5.0 μm (Figs. 2, 3).

Sequence analysis Culture and collection of mycelium were carried out as previously described [9]. For sequence analysis, DNA was extracted from pathogen and identified using methods previously described [9, 10]. The 18S rDNA and β -tubulin sequences were amplified by PCR with primer pair (18S rDNA forward primer: 5'-TCCGTAGGTG AACCTGCGG-3' and reverse primer: 5'-TCCTC CGCTTATTGATATGC-3'; β -tubulin gene forward primer: 5'-GGTAACCAAATCGGTGCTGCTTTC-3'



Fig. 2 The big conidia head, with sterigmata in two series ($\times 400$)



Fig. 3 The small one with a single row of phialides ($\times 400$)

and reverse primer: 5'-ACCCTCAGTGTAGTGAC CCTTGGC-3'). Sequencing of the PCR products were performed by the service of Sangon Company Ltd (Shanghai, China). The sequences obtained were submitted to GenBank for homology search with Blast (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>). After homology searching against the GenBank or the proprietary fungal DNA databases, the sequences of pathogen were found to share 100% similarity with that of *Aspergillus flavus* in 595-bp 18S rDNA and 545-bp β -tubulin gene.

Microbiological Findings

The Gram staining and microbiological culture both were negative (biopsy tissue, ascites, and bile).

Laboratory Evaluation

The routine blood test showed that the white blood cell counts were significantly reduced (less than 2×10^9). The biochemical tests showed remarkable elevations of total and direct bilirubin (up to 715 and 368.5 $\mu\text{mol/l}$), total bile acids levels (more than 100 $\mu\text{mol/l}$) above baseline at day before transplantation.

Discussion

According to the clinical manifestation and mycological findings, the patient was diagnosed as a case of Aspergillosis of biliary tract caused by *Aspergillus flavus*. *A. flavus* is ubiquitous in nature and can cause a variety of aspergilloses, including central nervous system infections [11]. It is the second most common species, produces aflatoxin, may be less susceptible to polyenes [12]. But the biliary is seldom infected by this pathogen. Steven H reported a case of cholangitis caused by *Aspergillus terreus* [13]. It was the first report of *Aspergillus* cholangitis with liver failure. As *Aspergillus* species can be readily found in the environment, invasive aspergillosis is widely believed to occur as a consequence of exogenous acquisition of the conidia (spores) of the species [14]. The most common route of transmission of *Aspergillus* infection is the airborne route. *Aspergillus* conidia are resilient and may survive for long periods in fomites (any substance that can absorb, retain, and transport infectious species, e.g., woolen clothes or bedding) [15]. However, the sources of *Aspergillus* may be broader than have traditionally been thought, as waterborne transmission of *Aspergillus* conidia through contaminated aerosols has been suggested [16]. The infestation route in this case was unclear. It may possibly include contaminated operation procedure, drainage tube, or ascending pathogen from gastrointestinal tract.

Invasive aspergillosis is extremely uncommon in immunocompetent hosts, although infection in apparently normal hosts can occur. The frequency and severity of invasive fungal infections in immunocompromised patients have increased steadily over the past two decades with the growing population of patients undergoing transplantation and the persistent challenges in preventing, diagnosing, and treating

these infections [1]. Mortality due to documented invasive aspergillosis approaches 80% to 100% in high-risk patients, including those with underlying hematologic malignancy or bone marrow or solid organ transplantation and may be related to several factors, including diagnostic and therapeutic inadequacies [17–19]. And in our case, the leukocytopenia also was an important predisposing factor. The changes in epidemiology of invasive aspergillosis may also be the result of growing awareness of aspergillosis among clinicians, the introduction of noninvasive diagnostic tools, and improved microbiological laboratory techniques.

Although the most frequent fungal infections are caused by *Candida* spp., infections due to *Aspergillus* spp. have the worst prognosis [20, 21]. Invasive aspergillosis has been reported in 1.5–6.5% of liver transplant patients [22–24], although other authors have reported higher frequencies [25]. A nationwide study carried out by the Spanish Study Group for Infections in the Transplant Patient documented the development of invasive aspergillosis in 2.8% of 1,719 liver transplant recipients, with a case fatality rate of 86% [26].

In China, fluconazole is the most extensively used prophylactically to treat fungal infection, such as in our case. The agent is effective against *Candida* species, *Cryptococcus neoformans*, and *Coccidioides immitis* but appears to be ineffective against other molds, including *Aspergillus* species and *Zygomycetes* [12]. As to this opinion, fluconazole prophylaxis significantly reduces invasive fungal infections in liver transplant recipients in the early postoperative period [25]. But prevention of infections caused by invasive molds remains a great challenge.

Treatment failure with currently available antifungal medication in patients with invasive aspergillosis has been reported to be at least 50% [18, 19]. New antifungal therapies with activity against *Aspergillus* have been developed, including lipid formulations of amphotericin B, the broad-spectrum triazoles (voriconazole, posaconazole, and ravuconazole), and the echinocandins (caspofungin, micafungin, and anidulafungin). But Fortun et al. [27] believed that the administration of L-AmpB in high-risk patients is independently associated with a reduction of invasive fungal infection. The successful therapy depends not only on an early diagnosis—which is often difficult to establish—but even more

importantly, on reversal of underlying host immune defects, such as neutropenia or high-dose immunosuppressive therapy [17].

References

- Patterson TF, Kirkpatrick WR, White M, et al. Invasive aspergillosis: disease spectrum, treatment practices, and outcomes. I3 Aspergillus Study Group. *Medicine (Baltimore)*. 2000;79:250–60.
- Denning DW. Invasive aspergillosis. *Clin Infect Dis*. 1998; 26:781–803.
- Marr KA, Patterson T, Denning D. Aspergillosis: pathogenesis, clinical manifestations, and therapy. *Infect Dis Clin North Am*. 2002;16:875–94.
- Perfect JR, Cox GM, Lee JY, et al. The impact of culture isolation of *Aspergillus* species: a hospital-based survey of aspergillosis. *Clin Infect Dis*. 2001;33:1824–33.
- Walsh TJ, Groll AH. Overview: non-fumigatus species of *Aspergillus*: perspectives on emerging pathogens in immunocompromised hosts. *Curr Opin Investig Drugs*. 2001;2: 1366–7.
- Christensen H, Nilsson KO, Nettelblad SC, et al. Common bile duct obstruction due to an intraluminal mass of candidiasis in a previously healthy child. *Pediatrics*. 1986;77: 858–61.
- Morris AB, Sands ML, Shiraki M, et al. Gallbladder and biliary tract candidiasis: nine cases and review. *Rev Infect Dis*. 1990;12:483–9.
- Noack KB, Osmon R, Patts KP, et al. Successful orthotopic liver transplantation in a patient with refractory biliary candidiasis. *Gastroenterology*. 1991;101:1728–30.
- Kabir S, Rajendran N, Amemiya T, et al. Quantitative measurement of fungal DNA extracted by three different methods using real-time polymerase chain reaction. *J Biosci Bioeng*. 2003;96:337–43.
- Zhao K, Ping W, Li Q, et al. *Aspergillus niger* var. *taxi*, a new species variant of taxol-producing fungus isolated from *Taxus cuspidata* in China. *J Appl Microbiol*. 2009; 107:1202–7.
- Kleinschmidt-DeMasters BK. Central nervous system aspergillosis: a 20-year retrospective series. *Hum Pathol*. 2002;33:116.
- Hospenthal DR, Rinaldi MG. Aspergillosis. In: Boucher HW, Patterson TF, editors. *Diagnosis and treatment of human mycoses*. New Jersey: Humana Press Inc; 2008. p. 181–94.
- Steven HE, Brent JB, Leslie LB. *Aspergillus* cholangitis: a late complication after Kasai portoenterostomy. *J Pediatr Surg*. 2002;37:923–5.
- Hajjeh RA, Warnock DW. Counterpoint: invasive aspergillosis and the environment rethinking our approach to prevention. *Clin Infect Dis*. 2001;33:1549–52.
- Woodcock AA, Steel N, Moore CB, Howard SJ, Custovic A, Denning DW. Fungal contamination of bedding. *Allergy*. 2006;61:140–2.
- Anaissie EJ, Stratton SL, Dignani MC, et al. Pathogenic moulds (including *Aspergillus* species) in hospital water distribution systems: a 3-year prospective study and

- clinical implications for patients with hematologic malignancies. *Blood*. 2003;101:2542–6.
17. Walsh TJ, Anaissie EJ, Denning DW, et al. Treatment of aspergillosis: clinical practice guidelines of the Infectious Diseases Society of America (IDSA). *Clin Infect Dis*. 2008;46:327–60.
 18. Lin SJ, Schranz J, Teutsch SM. Aspergillosis case-fatality rate: systematic review of the literature. *Clin Infect Dis*. 2001;32:358–66.
 19. McNeil MM, Nash SL, Hajjeh RA, et al. Trends in mortality due to invasive mycotic diseases in the United States, 1980–1997. *Clin Infect Dis*. 2001;33:641–7.
 20. Paterson DL, Singh N. Invasive aspergillosis in transplant recipients. *Medicine*. 1999;78:123–38.
 21. Kusne S, Torre-Cisneros J, Manos R, et al. Factors associated with invasive lung aspergillosis and the significance of positive *Aspergillus* culture after liver transplantation. *J Infect Dis*. 1992;166:1379–83.
 22. Castaldo P, Stratta RJ, Wood RP, et al. Clinical spectrum of fungal infections after orthotopic liver transplantation. *Arch Surg*. 1991;126:149–56.
 23. Singh N, Mieles L, Yu VL, et al. Invasive aspergillosis in liver transplant recipients: association with candidemia and consumption coagulopathy and failure of prophylaxis with low-dose amphotericin B. *Clin Infect Dis*. 1993;17:906–8.
 24. Paya CV, Hermans PE, Washington JA, et al. Incidence, distribution, and outcome of episodes of infection in 100 orthotopic liver transplantations. *Mayo Clin Proc*. 1989;64:555–64.
 25. Playford EG, Webster AC, Sorrell TC, Craig JC. Systematic review and meta-analysis of antifungal agents for preventing fungal infections in liver transplant recipients. *Eur J Clin Microbiol Infect Dis*. 2006;25:549–61.
 26. Muñoz P, Torre J, Bouza E, et al. Invasive aspergillosis in transplant recipients. A large multicentric study. In: Program and abstracts of the 36th interscience conference on antimicrobial agents and chemotherapy, New Orleans, Abstract J128. Washington: American Society for Microbiology; 1996. p. 242.
 27. Fortún J, Martín-Dávila P, Moreno S, et al. Prevention of invasive fungal infections in liver transplant recipients: the role of prophylaxis with lipid formulations of amphotericin B in high-risk patients. *J Antimicrob Chemother*. 2003;52:813–9.