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A novel framework for real-time diseased coral reef image classification

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Abstract

The new challenge in image processing is in processing submarine coral reef images. The coral reef disease classification from such submarine coral reef images has become an important research activity that helps marine biologist. An automated system is required to extract texture features so as to classify coral reef diseases from captured images. The proposed framework encompasses an efficient feature descriptor that classifies different submarine images of coral reef with diseases. The proposed framework employs most excellent image processing and machine learning techniques for classification. At first, the diseased coral reef images are segmented using Gradient-based sobel operator. Then, texture features are extracted from HSV color space using the proposed Mean Direct Code Pattern (MDCP) and from RGB space using proposed Diagonal Direction Value Pattern (DDVP). The proposed feature descriptors provide codes considering elements in diagonal directions. The resultant feature vector is then given as input to various classifiers to classify the diseased images. The efficiency of the proposed framework is demonstrated using real-time coral reef diseased images. The performance of various classifiers such as Decision Tree (DT), Classification And Regression Tree (CART), C4.5, Adaboost, Rotation Forest (RoF), Random Forest (RF), SVM, KNN, CNN, PCCNN and Naive Bayes is analysed. Performance results of the proposed framework for diseased coral reef image classification show that the framework outperforms recent works where feature descriptors such as LBP, LDP, CLBP, ILDP, DLBP, LTxXORP, CS-LBP, RLTP, $Z \oplus TZLBP$, OC-LBP, LTrP and PRI-CoLBP are used. Classification results are validated by marine biologists.

Keywords Classification · Feature extraction · Feature descriptor · Classifiers · HSV · RGB

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1 Introduction

Coral reefs are one of the wealthy and assorted ecosystems on earth [[24,](#page-37-0) [34\]](#page-37-0). They occupy a smaller area of just 0.1% of the ocean surface although they are the territory for at least 25% of the entire marine species. Healthy coral reefs are one of the most important and the most precious ecosystems on planet Earth. In recent days, coral diseases are a major threat to the coral ecosystem worldwide [[6\]](#page-36-0). Coral diseases generally occur due to increased sea surface temperatures, ultraviolet radiation and pollutants which can increase the susceptibility of corals to disease, leading to outbreaks where corals are abundant in diversity. Coral diseases can cause significant changes in coral reproduction rates, growth rates, community structure, species diversity, and abundance of reef-associated organisms. Besides these, corals span a wide area, and their abundance supports biodiversity since it is a natural barrier to deadly tsunami and is also a habitat to many creatures. Coral diseases present in a particular area at a given period (month) if reported early will enable government to take countermeasures so as to save the precious ecosystem. Marine biologists are still looking for an automated system to provide earlier reporting of solutions to protect marine ecosystem.

Corals are generally classified as healthy, unhealthy and dead as shown in Fig. 1. Healthy coral appears in colors of olive green, brown, tan and pale yellow. Healthy coral colony is not affected by disease or bleaching. Healthy coral provides shelter for many species that rely on the composition provided by corals for their homes. Corals are affected by a form of narrow band intruding on living tissues of a healthy coral, with the dead white coral skeletons above. The narrow band's color comes from deliberations of bacteria that produce sulphur compounds and low-oxygen conditions, which infect the coral [\[34\]](#page-37-0). For example, black band disease is caused by a bacterium called phormidium corallyticum. Healthy and diseased coral images have to be classified as any onset of disease can be curbed in the bud to protect coral for the future. An automated system reports coral diseases earlier so that the government can take countermeasures so as to prevent further damage to coral reef. There are a lot of existing works on classification that are applicable to texture data sets and some works for coral reef image classification, but only a few works are reported on coral reef disease classification. An automated system is the need of the hour for coral reef disease classification. An efficient feature vector must be devised to reduce the computational and time complexities and to improve accuracy of the classification process.

Fig. 1 a Healthy coral. b Diseased coral. c Dead coral

This paper is divided into five sections and starts with introduction. Section two presents research works that have been carried out on coral reef classification. Section three describes the overall framework of the proposed system. Section four describes the proposed novel feature descriptors. Experimental results are presented in section five, and the last section presents the conclusion and future work.

1.1 The contributions in this work are summarized as follows

- 1. Both HSV and RGB color spaces are considered with feature extraction for diseased coral reef image classification.
- 2. A novel feature descriptor MDCP is proposed which considers every element for mean estimation and only the diagonal elements for assigning direct code during feature extraction.
- 3. A novel feature descriptor DDVP is proposed which considers diagonal directional elements for feature extraction.
- 4. Feature vector is constructed, by concatenating both HSV-MDCP and RGB-DDVP.
- 5. The proposed framework reduces the computational and time complexities effectively by considering the diagonal elements only.
- 6. Since diagonal elements are considered, classification results are improved which is proven with various classifiers like Decision Tree (DT), Classification And Regression Tree (CART), C4.5, Adaboost, Rotation Forest (RoF), Random Forest (RF), SVM, CNN, PCCNN, KNN and Naive Bayes.
- 7. Performance comparison of the proposed feature descriptors with other feature descriptors such as LBP [\[23\]](#page-37-0), LDP [[38\]](#page-38-0), CLBP [\[33\]](#page-37-0), ILDP [[2](#page-36-0)], DLBP [\[16\]](#page-37-0), LTxXORP [[4](#page-36-0)], CS-LBP [\[11](#page-37-0)], RLTP [[31\]](#page-37-0), Z ⊕ TZLBP [[1](#page-36-0)], OC-LBP [\[39](#page-38-0)], LTrP [[25\]](#page-37-0), PRI-CoLBP [[30\]](#page-37-0), OPT [\[3\]](#page-36-0), is made to highlight the superiority of the proposed framework in terms of classification accuracy.

2 Related works

Coral disease is a major threat to coral reef ecosystems [\[14\]](#page-37-0). With the available literature it is observed that there are some works that have reported coral reef image classification, but no work has reported on coral reef disease classification. Pérez et al. [[28\]](#page-37-0) have enhanced coral reef images using Contrast Limited Adaptive Histogram Equalization (CLAHE), segmented coral reef images using Gaussian mixture models [[7\]](#page-36-0) and extracted texture features using Gabor. Shiela et al. [[23](#page-37-0)] have classified corals in videos as living and nonliving by extracting texture features using Local Binary Pattern (LBP) descriptor and classified them using Linear Discriminant Analysis (LDA). According to Shiela et al. [\[23\]](#page-37-0) living coral is considered as smooth textures, and non living coral is considered as irregular textures. For enhancing coral reef images, Shihavuddin et al. [[33\]](#page-37-0) have utilised Normalization, color correction, color stretching and Contrast Limited Adaptive Histogram Specification (CLAHS) whereas Ani et al. [[2](#page-36-0)] have utilised CLAHE and Contrast Stretching (CS). Multi-task learning and temporal pattern mining [[19\]](#page-37-0) are mostly applied for feature representation for activities and selecting discriminant features [\[21](#page-37-0)]. Generic human motions [\[18\]](#page-37-0) are tracked using fusion of low and high dimensional Approaches [\[37\]](#page-38-0). Ye Liu et al. [\[20](#page-37-0)] have used multi-task multi-view learning for urban water quality prediction.

For extracting features of coral reef images, Shihavuddin et al. [\[33\]](#page-37-0) have utilised Gabor, GLCM, CLBP, opponent angle and hue histogram whereas Ani et al. [\[2,](#page-36-0) [3\]](#page-36-0) have utilised Improved Local Derivative Pattern (ILDP) and Octa-angled Pattern for triangular sub region (OPT). For coral reef classification, Shihavuddin et al. [[33\]](#page-37-0) have utilised KNN, SVM, and NN classifiers whereas Ani et al. [[2](#page-36-0), [3](#page-36-0)] have utilised Support Vector Machine (SVM), K-Nearest Neighbor (KNN), Pulse Coupled Convolutional Neural Network (PCCNN) and Convolutional Neural Network (CNN) classifiers. Ani et al. [\[1\]](#page-36-0) have segmented coral reef images using zctive contour and classified coral videos using KNN classifier with various distance metrices whereas Mohammad et al. [\[32](#page-37-0)] have utilised KNN with euclidean distance for classification. For extracting features, Ani et al. [[1](#page-36-0)] have utilised Z with Tilted Z Local Binary Pattern ($Z \oplus$ TZLBP) feature descriptor whereas Mohammad et al. [[32\]](#page-37-0) have utilised Completed Local Binary Pattern (CLBP). To enhance coral reef images, Ani et al. [\[1\]](#page-36-0) have utilised CLAHE and contrast stretching whereas Mohammad et al. [[32](#page-37-0)] have utilised contrast normalization. Beijbom et al. [[5](#page-36-0)] have provided Moorea Labeled Corals (MLC) data set, which is a benchmark data set for coral reef image classification. Maximum Response (MR) filter bank is used for extracting features, and Library SVM is applied for classification. Not only is coral reef, fish that lives in coral reef also classified using SVM and CNN classifiers by extracting HOG features [\[35](#page-37-0)].

2.1 Observation

Just as coral reef image is classified into any one of the classes, Coral reef diseased images have to be classified into any one of the nine classes with class types being Aspergillosis (Asper), Bacterial Bleaching (BaB), Black Band (BB), Black Spot Dark Spot (BSDS), White Plague (WPl), White Band (WB), Yellow Band (YB), White Pox (WPo), and Pink Spot (PS). An automated system is expected to classify the diseased coral reef images into any one of the classes. For texture, a lot of standard data sets are available. But there are not many data sets available for coral reef diseases. Coral reef and texture data sets are classified with their textural features, but texture combined with colour spaces is important for diseased coral reef image classification.

3 System architecture

The proposed framework consists of the following steps for diseased coral reef image classification, and it is shown in Fig. [2](#page-4-0). The steps in the proposed framework are segmentation, feature extraction and classification.

3.1 Segmentation using gradient-based sobel operator

In a submarine environment, the edges of the coral reef must be correctly estimated. To detect the coral edges perfectly, coral reef has to be correctly separated from its background. K-Means is the mostly used clustering technique for image segmentation [\[36](#page-37-0)]. The difficulty with K-Means is that it considers global cluster where results are not accepted. For various initial divisions, K-Means results in different final clusters. It is also not simple to calculate K-Value. On the other hand, Fuzzy C-Means method poses difficulty in managing outlier points [[9\]](#page-37-0). It allocates high membership values for the **OFFLINE**

Proposed Feature Descriptor

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Fig. 2 System Architecture of the proposed framework

outlier points so that the membership of data points depends directly on the membership values of other cluster centres. This directs to unwanted results.

Coral Reef Image

Classification using various Classifiers

ATABASE

Gradient-based approach is applied on the diseased coral reef images for efficient segmentation. Accurate edges of the coral image are identified using Sobel operator. The number of white points are estimated from the edge extracted coral image. Gradient-based image segmentation technique is used to separate the coral reef image from its background efficiently as shown in Fig. [3.](#page-5-0) b. The Gradient-based image segmentation identifies significant local changes in the intensity level of a coral image. Consider a coral image $f(x, y)$, where magnitude and the direction of the gradient are computed as shown in Eqs. (1) and (2) respectively.

$$
|G| = \sqrt{G_x^2 + G_y^2} \tag{1}
$$

$$
\alpha(x, y) = \tan^{-1}\left(\frac{G_y}{G_x}\right) \tag{2}
$$

Accurate edges of the coral reef image are identified using Sobel operator as shown in Fig. [3](#page-5-0). c. Sobel operator is based on convolving the coral reef image with a small, separable, and integer-valued filter in the horizontal and the vertical directions. It is inexpensive in terms of computations. The kernels of sobel operator are shown in Eq. (3):

$$
G_x = \begin{matrix} -1 & 0 & 1 & 1 & 2 & 1 \\ -2 & 0 & 2 & G_y & 0 & 0 \\ -1 & 0 & 1 & -1 & -2 & -1 \end{matrix}
$$
 (3)

Where G_x and G_y are the horizontal and the vertical derivative approximations respectively.

(a) (b) (c)

Fig. 3 a Input coral reef b Background removed coral reef c Edges detected coral reef

3.2 Feature extraction

DDVP extracts features by finding the local difference between the elements among the three planes in four diagonal directions in RGB format. Then, the original image is transformed to HSV color space because it is more perceptually uniform than other color spaces. MDCP so as to extract features considers all elements for mean estimation and diagonal elements for applying direct codes.

A feature descriptor can be efficient only when it improves accuracy besides reducing computational complexity. For feature extraction, it is not important to use all pixels in an image as it becomes a time consuming process if all the pixels are considered. It is a fact that diagonal elements contribute more than other elements for extracting features [\[2](#page-36-0), [3](#page-36-0)]. Novel DDVP from RGB plane and MDCP from HSV plane is concatenated to generate feature vector which is explained in detail in Section [4.](#page-8-0)

However deep learning might together deal with the feature extraction and classification step, the purpose of this work is to develop a feature descriptor that might pair with existing classifiers for the classification of coral reef or its diseases with greater accuracy. However one more objective of this work is to create it appropriate for real time implementations where computational resources and time are constraints. The drawback of deep learning is its computational and time complexity, which makes it incompatible for real time implementation where there is time constraint [[3\]](#page-36-0).

3.3 Classification

For diseased coral reef image classification, texture feature vector is provided as input for various classifiers. A Decision Tree (DT) classifier is a non-parametric supervised learning technique applied for classification and regression. Still now, DT classifiers have provided good results for large data sets and imbalanced data sets. So, in this work the performances of different DT classifiers are evaluated and assessed. The classifiers used for classification are Decision Trees (DT), Classification And Regression Trees (CART), Random Forest (RF), C4.5, AdaBoost, Rotation Forest (RoF), SVM, KNN, CNN, PCCNN [\[3\]](#page-36-0) and Naive Bayes.

Classification using DT classifier is very easy. A DT is a non-parametric supervised learning technique used for classification. The DT [[22](#page-37-0)] constructs a model that classifies an image by learning simple decision rules inferred from the features trained. Random forest [\[15\]](#page-37-0) is an ensemble method which considers a subset of annotations and a subset of variables to build a decision tree. It builds several such decision trees and combines them collectively to result in accurate classification. Adaboost improves the classification results by combining weak predictors collectively. CART considers every possible subset of features for classification.

CART is divided into Regression Trees and Classification Trees. Both the trees divide the feature space into distinct and non-overlapping regions. Both the trees apply recursive binary splitting technique that uses a top-down greedy approach. CART makes the tree grow for the entire data and then prunes back. C4.5 [\[17](#page-37-0)] is a statistical classifier which is an extension of ID3 algorithm. It uses an information entropy evaluation function for feature selection. In Rotation Forest (RoF), every decision tree is trained by first applying Principal Component Analysis (PCA) on a random subset of the input features. This uses a number of decision trees in order to improve the classification accuracy. Each tree depends on the values of a random vector sampled independently in the forest.

Experiment is started by classifying using DT with default parameters to get a baseline. By default, the Minimum sample per leaf node is set to 1, that logically makes the tree over-fit and contains all the data points, including outliers. By default, Greedy approach is used to reduce similar trees. The features that are considered for decision making is common for all classifiers such as bin which contains the maximum pixel value, bin which contains the minimum pixel value and bin which contains the median pixel value. Features considered for diseased coral reef image data sets with nine classes are shown in Fig. [4,](#page-7-0) where FV represents Feature Vector of the proposed framework. The bin with maximum, minimum and median pixel values are set to 24 because the bin size of histogram is 24. This is similar for all classifiers. Here maximum tree depth is set to 12, Hence every features considered will get a chance to participate in becoming a decision node.

In Random forest, the bin with maximum, minimum and median pixel values are set to 24. The node estimators is set to 9, to find the optimum classification. In Adaboost, number of weak learners and the maximum depth of tree are the same as other classifiers.

Fig. 4 Decision Tree Classifier for coral reef diseased image

Here learning rate is an important parameter that is set to fit the weak learners perfectly. By default, learning rate is set to 0.01, that fits more weak learners. The two important parameters in C4.5 are Minimum cases (M) and Confidence Level (CL). M is preferred to be a high value and is set as 9 and the CL should be a lower value and is set as 1 for classification. The number of iterations and the number of splits are the two main default parameters in RoF which are set to 9 and 2.

4 The proposed framework

4.1 Background

There are a lot of differences between normal coral reef images and diseased coral reef images. For coral reef classification, texture features alone are considered. Each species of coral reef has its unique texture features. In the existing literature for feature extraction, texture features alone are considered. In order to improve the accuracy of classification of diseased coral reef images, features extracted from RGB and HSV color spaces are combined with textural feature. Usually for texture feature extraction, Ojala et al. [[26,](#page-37-0) [27](#page-37-0)] and Heikkila et al. [\[12](#page-37-0), [13\]](#page-37-0) have used Local Binary Pattern (LBP), which is called as 'Curse of dimensionality', due to the size of feature vector.

4.2 Diagonal direction value pattern (DDVP)

Several feature descriptors are available in the literature for texture classification. Most of the feature descriptors make use of only the gray scale values of the pixel in an image. But the relationship between the diagonal directional elements in three planes of an image is not considered.

DDVP extracts features diagonally along four $(45^0, 135^0, 225^0, 315^0)$ directions from three planes efficiently. The speciality of DDVP is that diagonal direction difference among pixel values in the three planes of the image are considered while reducing the bin size of histogram used in the classification stage. The steps in the proposed feature descriptor are summarized as follows:

In an image I, let $R_{i,j}$, $G_{i,j}$ and $B_{i,j}$ be the centre pixels of a 3 \times 3 local region termed as Block B_k , in each of the three planes as shown in Fig. 5, where $1 \ge k \le N$, N is the total number of 3 ×3 blocks in the image I. $R_{i-1,j+1}$, $R_{i,j+1}$, $R_{i+1,j+1}$, $R_{i+1,j}$, $R_{i+1,j-1}$, $R_{i,j-1}$, $R_{i-1,j-1}$ and $R_{i-1,j}$ are the eight neighbors of the centre pixel $R_{i,j}$ in the Red plane. $G_{i-1,j+1}$, $G_{i,j+1}$, $G_{i+1,j+1}$, $G_{i+1,j+1}$ $1, j, G_{i+1,j-1}, G_{i,j-1}, G_{i-1,j-1}$ and $G_{i-1,j}$ are the eight neighbors of the centre pixel $G_{i,j}$ in the Green Plane. $B_{i-1,j+1}, B_{i,j+1}, B_{i+1,j+1}, B_{i+1,j}, B_{i+1,j-1}, B_{i,j-1}, B_{i-1,j-1}$ and $B_{i-1,j}$ are the eight neighbors of the centre pixel $B_{i,j}$ in the Blue plane.

4.2.1 Direction difference estimation

For a Block B_K , R_θ , B_θ , G_θ , RG_θ , RB_θ , GR_θ , BR_θ and BG_θ are constructed as shown in Eq. (4) to Eq. (12) from the neighbors of the centre pixel in three planes along the directions θ = < 45°, 135°, 225° *and* 315°> as shown in Fig. [6](#page-9-0)

$R_{i-1,j+1}$	$R_{i,j+1}$	$R_{i+1,j+1}$		$G_{i-1,j+1}$	$G_{i,j+1}$	$G_{i+1,j+1}$	$B_{i-1,j+1}$	$B_{i,j+1}$	$B_{i+1,j+1}$
$R_{i-1,j}$	$R_{i,j}$	$R_{i+1,j}$		$G_{i-1,j}$	$G_{i,j}$	$G_{i+1,j}$	$B_{i-1,j}$	$B_{i,j}$	$B_{i+1,j}$
$R_{i-1,j-1}$	$R_{i,j-1}$	$R_{i+1,j-1}$		$G_{i-1,j-1}$	$G_{i,j-1}$	$G_{i+1,j-1}$	$B_{i-1,j-1}$	$B_{i,j-1}$	$B_{i+1,j-1}$
(a) R-plane				(b) G-plane		(c) B-plane			

Fig. 5 RGB plane images

$$
R_{B_k,\theta}[m] = R_{i,j} - R_{i+u,j+v} \tag{4}
$$

$$
G_{B_k,\theta}[m] = G_{i,j} - G_{i+u,j+v} \tag{5}
$$

$$
B_{B_k,\theta}[m] = B_{i,j} - B_{i+u,j+v} \tag{6}
$$

$$
RG_{B_k, \theta}[m] = R_{i,j} - G_{i+u,j+v} \tag{7}
$$

$$
RB_{B_k,\theta}[m] = R_{i,j} - B_{i+u,j+v} \tag{8}
$$

$$
BR_{B_k,\theta}[m] = B_{i,j} - R_{i+u,j+v} \tag{9}
$$

$$
BG_{B_k,\theta}[m] = B_{i,j} - G_{i+u,j+v} \tag{10}
$$

$$
GR_{B_k, \theta}[m] = G_{i,j} - R_{i+u, j+v} \tag{11}
$$

$$
GB_{B_k,\theta}[m] = G_{i,j} - B_{i+u,j+v} \tag{12}
$$

where ' B_k ' is of size 3×3 ,

1≥m≤4;

 $1 \geq k \leq N$

and

$$
\begin{aligned} \n\text{(u, v)} &= \n\begin{cases} \n u = 1, v = 1, \text{if } \theta = 45^{\circ} \\ \n u = -1, v = 1, \text{if } \theta = 135^{\circ} \\ \n u = -1, v = -1, \text{if } \theta = 225^{\circ} \\ \n u = 1, v = -1, \text{if } \theta = 315^{\circ} \n\end{cases} \n\end{aligned}
$$

4.2.2 Vector generation using four diagonal directions in three planes

For each R, G and B plane, the vectors are generated along diagonal directions. Vector $R_{B_k,\theta}$ is constructed using R, RB and RG values, whereas vector $G_{B_k,\theta}$ is constructed using G, GR and GB values and vector $B_{B_k,\theta}$ is constructed using B, BR and BG values. The vectors generated for three planes are shown in Eqs. (13) to (15) .

$$
R_{B_k, \theta}[i] = \langle R_{\theta}, R B_{\theta}, R G_{\theta} \rangle_{B_k} \tag{13}
$$

$$
G_{B_k,\theta}[i] = \langle G_{\theta}, GB_{\theta}, GR_{\theta} \rangle_{B_k} \tag{14}
$$

$$
B_{B_k,\theta}[i] = \langle B_{\theta}, BR_{\theta}, BG_{\theta} \rangle_{B_k} \tag{15}
$$

where $\theta = <45^{\circ}$, 135°, 225° and 315°>,

 $1 \geq i \leq 4$ and

Now, $R_{B_k, \theta}$, $G_{B_k, \theta}$, and $B_{B_k, \theta}$ vectors may contain negative and non-negative values. Binary codes (0 and 1) are assigned depending on negative and non-negative values.

4.2.3 Binary codes assignment

The binary codes are assigned for three planes as given by Eqs. (16) to [\(18\)](#page-11-0). Negative and nonnegative values are converted into binary codes either 0 or 1.

$$
BCR_{B_k, \theta}[a] = \begin{cases} 1, R_{B_k, \theta}[i] > 0\\ 0, Otherwise \end{cases}
$$
 (16)

$$
\text{BCG}_{B_k,\theta}[a] = \begin{cases} 1, G_{B_k,\theta}[i] > 0\\ 0, Otherwise \end{cases}
$$
 (17)

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$$
\text{BCB}_{B_k,\theta}[a] = \begin{cases} 1, B_{B_k,\theta}[i] > 0\\ 0, Otherwise \end{cases}
$$
 (18)

where $\theta = <45^{\circ}$, 135°, 225° and 315°>,

$$
1 \ge i \le 3,
$$

$$
1 \ge k \le N \text{ and}
$$

$$
1 \ge a \le 3.
$$

4.2.4 Diagonal direction value assignment

DDV is assigned for three plane vectors estimated. Depending on the position of the binary codes of three planes, the diagonal directional value is assigned for diagonal directions as shown in Eq. (19) to Eq. (21) .

$$
DDV_BCR\left[_{B_k,\theta}\right] = \begin{cases} 1, \text{if } BCR_{B_k,\theta}[1] = 0 \text{ and } BCR_{B_k,\theta}[2] = 0 \text{ and } BCR_{B_k,\theta}[3] = 0 \\ 2, \text{if } BCR_{B_k,\theta}[1] = 0 \text{ and } BCR_{B_k,\theta}[2] = 0 \text{ and } BCR_{B_k,\theta}[3] = 1 \\ 3, \text{if } BCR_{B_k,\theta}[1] = 0 \text{ and } BCR_{B_k,\theta}[2] = 1 \text{ and } BCR_{B_k,\theta}[3] = 0 \\ 4, \text{if } BCR_{B_k,\theta}[1] = 0 \text{ and } BCR_{B_k,\theta}[2] = 1 \text{ and } BCR_{B_k,\theta}[3] = 1 \\ 5, \text{if } BCR_{B_k,\theta}[1] = 1 \text{ and } BCR_{B_k,\theta}[2] = 0 \text{ and } BCR_{B_k,\theta}[3] = 0 \\ 6, \text{if } BCR_{B_k,\theta}[1] = 1 \text{ and } BCR_{B_k,\theta}[2] = 0 \text{ and } BCR_{B_k,\theta}[3] = 1 \\ 7, \text{if } BCR_{B_k,\theta}[1] = 1 \text{ and } BCR_{B_k,\theta}[2] = 1 \text{ and } BCR_{B_k,\theta}[3] = 0 \\ 8, \text{if } BCR_{B_k,\theta}[1] = 1 \text{ and } BCR_{B_k,\theta}[2] = 1 \text{ and } BCR_{B_k,\theta}[3] = 1 \end{cases}
$$

$$
DDV_BCG\Big[_{B_k,\theta}\Big] = \begin{cases} 1, \text{if } BCG_{B_k,\theta}[1] = 0 \text{ and } BCG_{B_k,\theta}[2] = 0 \text{ and } BCG_{B_k,\theta}[3] = 0 \\ 2, \text{if } BCG_{B_k,\theta}[1] = 0 \text{ and } BCG_{B_k,\theta}[2] = 0 \text{ and } BCG_{B_k,\theta}[3] = 1 \\ 3, \text{if } BCG_{B_k,\theta}[1] = 0 \text{ and } BCG_{B_k,\theta}[2] = 1 \text{ and } BCG_{B_k,\theta}[3] = 0 \\ 4, \text{if } BCG_{B_k,\theta}[1] = 0 \text{ and } BCG_{B_k,\theta}[2] = 1 \text{ and } BCG_{B_k,\theta}[3] = 1 \\ 5, \text{if } BCG_{B_k,\theta}[1] = 1 \text{ and } BCG_{B_k,\theta}[2] = 0 \text{ and } BCG_{B_k,\theta}[3] = 0 \\ 6, \text{if } BCG_{B_k,\theta}[1] = 1 \text{ and } BCG_{B_k,\theta}[2] = 0 \text{ and } BCG_{B_k,\theta}[3] = 1 \\ 7, \text{if } BCG_{B_k,\theta}[1] = 1 \text{ and } BCG_{B_k,\theta}[2] = 1 \text{ and } BCG_{B_k,\theta}[3] = 0 \\ 8, \text{if } BCG_{B_k,\theta}[1] = 1 \text{ and } BCG_{B_k,\theta}[2] = 1 \text{ and } BCG_{B_k,\theta}[3] = 1 \end{cases}
$$

$$
DDV_BCB\left[_{B_k, \theta}\right] = \begin{cases} 1, \text{if } BCB_{B_k, \theta}[1] = 0 \text{ and } BCB_{B_k, \theta}[2] = 0 \text{ and } BCB_{B_k, \theta}[3] = 0 \\ 2, \text{if } BCB_{B_k, \theta}[1] = 0 \text{ and } BCB_{B_k, \theta}[2] = 0 \text{ and } BCB_{B_k, \theta}[3] = 1 \\ 3, \text{if } BCB_{B_k, \theta}[1] = 0 \text{ and } BCB_{B_k, \theta}[2] = 1 \text{ and } BCB_{B_k, \theta}[3] = 0 \\ 4, \text{if } BCB_{B_k, \theta}[1] = 0 \text{ and } BCB_{B_k, \theta}[2] = 1 \text{ and } BCB_{B_k, \theta}[3] = 1 \\ 5, \text{if } BCB_{B_k, \theta}[1] = 1 \text{ and } BCB_{B_k, \theta}[2] = 0 \text{ and } BCB_{B_k, \theta}[3] = 0 \\ 6, \text{if } BCB_{B_k, \theta}[1] = 1 \text{ and } BCB_{B_k, \theta}[2] = 0 \text{ and } BCB_{B_k, \theta}[3] = 1 \\ 7, \text{if } BCB_{B_k, \theta}[1] = 1 \text{ and } BCB_{B_k, \theta}[2] = 1 \text{ and } BCB_{B_k, \theta}[3] = 0 \\ 8, \text{if } BCB_{B_k, \theta}[1] = 1 \text{ and } BCB_{B_k, \theta}[2] = 1 \text{ and } BCB_{B_k, \theta}[3] = 1 \end{cases}
$$

where $\theta = 45^{\circ}$, 135°, 225° and 315°.

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4.2.5 Summed diagonal direction value

The obtained DDV values are summed together along the four diagonal directions. Summed Diagonal Direction Value (SDDV) is estimated using Eq. (22).

$$
SDDV(B_k, \theta) = \sum_{k=1}^{N} |DDV_BCR[B_k, \theta] + DDV_BCG[B_k, \theta] + DDV_BCB[B_k, \theta]| (22)
$$

where $\theta = <45^{\circ}$, 135°, 225° and 315°>.

Summed Diagonal Direction Value Vector (SDDVV) is estimated using Eq. (23)

$$
SDDVV = \langle SDDV(B_{1_k}, \theta), SDDV(B_{2_{k+1}}, \theta), ..., SDDV(B_{N-1}, \theta), SDDV(B_N, \theta) \rangle \qquad (23)
$$

where $\theta_i = \langle 45^\circ, 135^\circ, 225^\circ \text{ and } 315^\circ \rangle$ and

 $1 \geq k \leq N$.

For an entire image, SDDVV is considered as the feature vector. This is shown in Eq. (24).

i.e.Feature vector
$$
(I) = SDDVV
$$
 (24)

Figure 7 explains the algorithm for feature extraction using DDVP.

4.3 Mean direct code pattern (MDCP)

Many feature descriptors utilize the relationship between the neighbor pixels in the gray scale image based on co-occurrence matrix, but the relationship between the diagonal neighbors in an image is not considered. In the proposed feature descriptor, the relationship between the diagonal neighbors is considered by providing a direct code pattern. The new feature descriptor, named MDCP with HSV extracts texture features of diseased coral reef images. RGB color space cannot represent color in terms of human interception, pixel variations due to illumination problem and intensity variations are overcome by HSV. Hence HSV is considered in this work. First the color image is converted into HSV color space. The steps in the proposed feature descriptor are summarized in the following sub sections:

4.3.1 Mean estimation for HSV images

The image is transformed to HSV color space as shown in Fig. [9](#page-14-0). In a diseased coral reef image, consider a Block B_k of size 3 ×3 where $1 \le k \le N$ and N is the total number of blocks in a frame. Let $E_{i,j}$ be the centre pixel of a 3 ×3 Block B_k , as shown in Fig. 8. $E_{i-1,j+1}, E_{i,j+1}, E_{i+1,j+1}$ $j_{i+1}, E_{i+1,j}, E_{i+1,j-1}, E_{i,j-1}, E_{i-1,j-1},$ and $E_{i-1,j}$ are the eight neighbors of the centre pixel. $E_{i,j}$ j . $E_{i-1, j+1}$, $E_{i+1, j+1}$, $E_{i-1, j-1}$ and $E_{i+1, j-1}$ are the four diagonal elements. Here, the input corresponds to three channels, i.e. H-plane, S-plane and V-plane as shown in Fig. [9.](#page-14-0)

The mean value of Block B_k is computed as in Eq. (25) separately for each plane as in Eq. (26), Eq. (27) and Eq. (28).

Mean = average of nine elements in a block
$$
(25)
$$

$$
HMean[k] = \frac{1}{n} \sum_{i=1}^{n} H[i]_{(B_k)}
$$
\n(26)

$$
SMean[k] = \frac{1}{n} \sum_{i=1}^{n} S[i]_{(B_k)}
$$
 (27)

$$
VMean[k] = \frac{1}{n} \sum_{i=1}^{n} V[i]_{(B_k)}
$$
 (28)

where $1 \ge k \le N$, N represents the elements in a Block B_k .

As shown in Fig. [10,](#page-14-0) more importance is provided to diagonal elements, where the diagonal directions are covered very efficiently along 45°, 135°, 225° and 315° as shown in Fig. [10a](#page-14-0). To improve accuracy, diagonal elements are given more importance compared to the other vertical and horizontal neighbors. Depending on the mean value, diagonal elements are alone

Fig. 8 Diagonal elements in a block are highlighted

Fig. 9 HSV plane images

considered for assigning the direct code. Diagonal elements are compared with the mean value obtained. Then, a direct code is assigned as feature vector for that block. This pattern is efficient because it depends on the diagonal elements much.

4.3.2 Diagonal neighbors estimation for HSV images

In a Block B_k , for H-plane, S-plane and V-plane the mean value is compared with four diagonal elements. Let the four diagonal elements be N_i , where $i = 1$ to 4 as shown in Eq. (29) to Eq. ([31\)](#page-15-0), mean value is compared with the four diagonal elements. If the value of the diagonal element is equal to or greater than mean value of the block, it is assigned a directional code as 1 or 0, otherwise as shown in Eq. (29) to Eq. [\(31](#page-15-0)).

$$
BCH_{B_K}[j] = \begin{cases} 1, [N_i]_{B_K} \geq HMean[k] \\ 0, [N_i]_{B_K} < HMean[k] \end{cases}
$$
 (29)

Fig. 10 (a) Diagonal elements are covered

$$
BCS_{B_K}[j] = \begin{cases} 1, [N_i]_{B_K} \geq SMean[k] \\ 0, [N_i]_{B_K} < SMean[k] \end{cases} \tag{30}
$$

$$
BCV_{B_K}[j] = \begin{cases} 1, [N_i]_{B_K} \geq VMean[k] \\ 0, [N_i]_{B_K} < VMean[k] \end{cases} \tag{31}
$$

where $j = 1 - N$, $1 \ge j \le 4$

where
$$
i = \begin{cases} i-m, j+n \\ i-m, j-n \\ i+m, j+n \\ i+m, j-n \end{cases}
$$
, $(m, n) = (1, 1)$

4.3.3 Assigning direct code for HSV images

Direct codes are assigned for a block B_k depending on the values of the diagonal neighbors. Now, the four neighbors value will be either 0 or 1. Depending on these values, a particular direct code is assigned for H color space as shown in Eq. [\(32\)](#page-16-0), for S color space as shown in Eq. [\(33](#page-16-0)), for V color space as shown in Eq. ([34\)](#page-16-0) and in Fig. 11.

Fig. 11 Direct code pattern

$$
\begin{pmatrix}\n1, if & BCH_{B_k} = 0000 \\
2, if & BCH_{B_k} = 0001 \\
3, if & BCH_{B_k} = 0010 \\
4, if & BCH_{B_k} = 1100 \\
5, if & BCH_{B_k} = 1100 \\
6, if & BCH_{B_k} = 1001 \\
7, if & BCH_{B_k} = 1001 \\
8, if & BCH_{B_k} = 1001 \\
9, if & BCH_{B_k} = 0101 \\
10, if & BCH_{B_k} = 0011 \\
11, if & BCH_{B_k} = 0011 \\
12, if & BCH_{B_k} = 0111 \\
13, if & BCH_{B_k} = 1110 \\
14, if & BCH_{B_k} = 1110 \\
15, if & BCH_{B_k} = 1111 \\
16, if & BCH_{B_k} = 1111 \\
17, if & BCH_{B_k} = 1001 \\
18, if & BCH_{B_k} = 1001 \\
2, if & BCH_{B_k} = 0001 \\
3, if & BCH_{B_k} = 0010 \\
4, if & BCH_{B_k} = 0010 \\
5, if & BCH_{B_k} = 0010 \\
6, if & BCH_{B_k} = 1100 \\
7, if & BCH_{B_k} = 1010 \\
8, if & BCH_{B_k} = 1010 \\
9, if & BCH_{B_k} = 0101 \\
11, if & BCH_{B_k} = 0011 \\
12, if & BCH_{B_k} = 0011 \\
13, if & BCH_{B_k} = 0111 \\
14, if & BCH_{B_k} = 0111 \\
15, if & BCH_{B_k} = 0111 \\
16, if & BCH_{B_k} = 0111 \\
17, if & BCH_{B_k} = 0111 \\
18, if & BCH_{B_k} = 1111 \\
19, if & BCH_{B_k} = 1110 \\
11, if & BCH_{B_k} = 0100 \\
5, if & BCH_{B_k} = 0100 \\
6, if & BCH_{B_k} = 0100 \\
7, if & BCH_{B_k} = 0100 \\
8, if & BCH_{B_k} = 0100 \\
9, if & BCH_{B_k} = 0110 \\
10, if & BCH_{B_k} = 0110 \\
11, if & BCH_{B_k} = 0111 \\
1
$$

4.3.4 Summed direct code pattern

The texture feature vector size obtained will be of 16 bins for each H-plane, S-plane and Vplane. The three planes feature vectors are summed as shown in Eq. (35).

$$
SDC[B_k] = \sum_{k=1}^{N} |DCH[B_k] + DC \cdot S[B_k] + DC \cdot V[B_k]| \tag{35}
$$

The resultant feature vector size for a particular Block B_k , after combining the three planes will be of 48 bins. These 48 bins of histogram is then normalised to half of its bin size as 24 bin histogram, which provides efficient feature extraction.

For an entire image, *SDC* is considered as the feature vector. This is shown in Eq. (36).

$$
SDDC = \frac{1}{2} \, SDC[B_k] \tag{36}
$$

Feature vector $(I) = SDDC$

4.4 Feature vector concatenation

The obtained feature vectors for DDVP and MDCP are concatenated to provide an efficient feature descriptor as shown in Eq. (37)

$$
Feature vector (I) = (37)
$$

Figure [12](#page-18-0) explains the algorithm of MDCP method. [Appendix 1](#page-30-0) explains the proposed method.

5 Experimental evaluation

5.1 Data sets description

The real-time diseased coral reef database was obtained from Suganthi Devadasan Marine Research Institute (SDMRI) (<http://www.sdmri.in>), Beach Road in Tuticorin coast. Diseased coral reef images are taken in a shallow water reef area of two metres depth in Shingle Island. The images are taken during the months between September 2017 and February 2018. Diseased coral reef images are classified into nine classes as shown in Fig. [13.](#page-19-0) The data set consists of 878 images at a resolution of 640×480 . The images in data set are all diseased images.

5.2 Quantitative comparison

Implementation is executed using MATLAB 2016a with Intel® Pentium® Processor (Dual Core), 4GB Memory and 2 TB Hard drive. The performance of various classifiers is compared using different metrics such as specificity, sensitivity, accuracy, F-measure, time and Mathews' Correlation Coefficient. To estimate the effectiveness of the proposed technique, the samples in the data set are separated in the following ratio: 90% for training and 10% for testing, 75% for training and 25% for testing, and 50% for training and 50% for testing.

Algorithm 2: Feature Vector Generation using HSV with MDCP								
Input: Segmented coral reef diseased image								
Output: Feature vector (I)								
Do for all pixels in an image								
Block B_k is separated into H, S and V Planes								
For every block B_k of size 3 \times 3 in the H-Plane								
Calculate Mean of pixels of B_k .								
Difference is estimated between Mean values and diagonal neighbors.								
Direct codes are assigned depending on difference values.								
End								
For every block B_k of size 3 \times 3 in the S-Plane								
Calculate Mean of pixels of B_k .								
Difference is estimated between Mean values and diagonal neighbors.								
Direct codes are assigned depending on difference values.								
End								
For every block B_k of size 3 \times 3 in the V-Plane								
Calculate Mean of pixels of B_k .								
Difference is estimated between Mean values and diagonal neighbors.								
Direct codes are assigned depending on difference values.								
End								
Summed Direct Code is estimated using Eq. (35).								
After Normalization, feature descriptor for a particular Block B_k is estimated using Eq.(36).								
End								

Fig. 12 MDCP Algorithm

5.2.1 F-measure

F-measure is a computation method used to test accuracy. It considers both precision and recall values. In Table [1,](#page-20-0) the variables are represented as TE to denote Testing, TR to denote Training, Prec to denote Precision, Recal to denote Recall and Fm to denote F-measure.

$$
F-measure = \frac{2*Prec*Recal}{Prec+Recal}
$$
 (38)

$$
Prec = \frac{Number\ of\ images\ classified\ accurately}{Total\ number\ of\ images\ classified} \tag{39}
$$

$$
Recal = \frac{Number\ of\ images\ classified\ accurately}{Total\ number\ of\ images\ in\ the\ database}
$$
\n
$$
(40)
$$

Table [1](#page-20-0) presents the results of classification accuracy for diseased coral reef data sets in comparison with existing feature descriptors. The proposed feature descriptor provides better classification

Fig. 13 SDMRI coral diseases

accuracy compared to existing feature descriptors. Since the data sets depend on both color and texture features, so the proposed feature descriptor has resulted in an improvement in accuracy by 9 and 10% respectively over existing feature descriptors. In SDMRI, real-time diseased coral reef data set, following the proposed, OPT, $Z \oplus TZLBP$ and ILDP feature descriptors provide improved results compared to CS-LBP, LBP, DLBP, PRI-CoLBP, LTxXORP, CLBP and OC-LBP. In OPT, pixels are covered in eight diagonal directions so as to improve accuracy. In ILDP, diagonal directions and diagonal neighbors are utilised for estimation which covers all pixels in an image efficiently to improve accuracy. In $Z \oplus TZLBP$, all the pixels are covered using Z and TZ (Tilted Z) pattern, where the centre pixel is also considered as one of the neighbors. In ILDP, OPT and $Z \oplus Z$ TZLBP only texture features are considered, but in the proposed feature descriptor textural features from RGB and HSV color planes are considered which provides efficient results.

5.2.2 Accuracy

$$
Accuracy = \frac{Images \ classified \ correctly}{All \ images \ classified} \tag{41}
$$

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Data set Feature		Division of Samples										
Descriptor			TE-10%, TR-90%			TE-25%, TR-75%			TE-50%, TR-50%			
SDMRI CORAL	Metrices	Pr	Re	Fm	Pr	Re	Fm	Pr	Re	Fm		
DISEASES	LBP	84.9	81.61	83.22	78.27	77.11	77.68	67.2	65.33	66.25		
EILAT	CLBP	85.21	83.9	84.54	76.1	74.95	75.52	68.32	66.4	68.34		
	DLBP	83.9	84.99	84.44	78.54	79.43	78.98	71.16	69.99	70.57		
	LTxXORP	85.6	86.91	86.25	79.67	78.4	79.02	71.9	69.03	70.43		
	CS -LBP	86	85.3	85.64	79.2	78.81	79	68.23	67.8	68.01		
	OC-LBP	85.91	86	85.95	78.31	79.68	78.98	68.5	67.5	67.99		
	PRI-CoLBP	89.45	86.9	88.15	83.5	82.78	83.13	72.86	74.12	73.48		
	RLTP	86.7	85.78	86.23	83.09	78.86	80.19	72.99	69.64	71.27		
	LTrP	86.85	85.9	86.37	83.54	79.36	81.39	72.6	70.39	71.47		
	LDP	87.9	88.19	88	83.67	82.93	83.29	72.9	74.73	73.80		
	ILDP	90.08	88.93	89.5	83.8	82.98	83.38	72.93	73.71	73.31		
	$Z \oplus TZLBP$	89.76	88.91	89.33	84.92	82.84	83.86	72.86	74.12	73.48		
	OPT	90.4	91.47	90.93	84.23	83.8	84.01	74.56	73.97	74.26		
	PROPOSED	96.19	94.6	95.38	89.54	87.41	88.46	78.25	80.17	79.19		
CORAL DISEASES	LBP	94.01	92.71	93.35	83.63	81.9	82.75	76.39	73.12	74.71		
RSMAS	CLBP	95.99	94.7	95.34	84.8	82.07	83.41	77	74.27	75.61		
	DLBP	94.62	92.98	93.79	83.39	82.7	83.04	77.45	73.88	75.62		
	LTxXORP	96.6	95.19	95.88	85.91	84.8	85.35	79.43	76.91	78.14		
	CS -LBP	94.68	92.85	93.75	84.71	83.9	84.3	75.49	75.21	75.34		
	OC-LBP	95.2	94.09	94.64	84.7	85.94	85.31	77.33	78.12	77.80		
	PRI-CoLBP	96.53	94.79	95.65	85.78	84.1	84.93	78.93	76.12	77.49		
	RLTP	95.90	92.69	94.26	85.1	83.95	84.52	78.32	76.4	77.34		
	LTrP	96.53	95.23	95.87	85.45	84.24	84.84	79.4	77.91	78.64		
	LDP	96.98	94.17	95.55	85.04	86.19	85.61	79.90	77.2	78.52		
	ILDP	96.96	95.45	96.19	84.7	85.94	85.31	79.07	78.69	78.87		
	$Z \oplus TZLBP$	97.78	96.36	97.06	86.47	85.27	85.86	80.24	78.51	79.36		
	OPT	97.98	98.32	98.14	87.61	86.85	87.22	79.69	80.13	79.90		
	PROPOSED	99.31	97.36	98.32	87.58	86.91	87.24	81.52	80.43	80.97		

Table 1 Comparison of feature descriptors with F-measure (Fm) on various coral Diseases data sets. The highest OA (%) obtained for each data set is shown in bold

In Table [2,](#page-21-0) the accuracy results of the proposed framework is compared with previous submarine image classification approaches. The proposed framework is not directly comparable with the results obtained by other previous approaches in literature as the experimental setups differ. Even though no pre-processing techniques are used, the proposed framework achieves good results for various data sets. The recent techniques of Shihavuddin et al. [[33\]](#page-37-0), Ani et al. [[2](#page-36-0)], Ani et al. [[1](#page-36-0)], Ani et al. [[3](#page-36-0)], Mohammad et al. [\[32\]](#page-37-0) and the proposed method are shown in Table [2.](#page-21-0) This table indicates that the proposed framework and OPT uses a quicker and simpler classification framework. This simpler classification framework provides outstanding results in coral reef image classification compared to existing coral reef classification frameworks.

For EILAT coral data set, DT classifier provides the highest classification accuracy as shown in Table [3](#page-22-0). DT is the well-known classifier for performing the multi-class classification on a data set. For EILAT2 coral data set, PCCNN classifier provides the highest classification accuracy. For MLC 2012 coral data set and SDMRI coral diseased data set, Random Forest provides the highest classification accuracy. For RSMAS coral data set and SDMRI real time data set, SVM, PCCNN and KNN provide highest classification accuracy compared to other classifiers. The dimensionality of the proposed feature vector is very low to reduce the complexity during classification Table [4.](#page-22-0)

Data set Approach	EILAT2	EILAT	MLC 2012	RSMAS	SDMRI	SDMRI diseased
$MDCP + DDVP + SVM$	98.67	97.23	93.83	97.66	88.45	90.21
$MDCP + DDVP + CNN$	97.53	96.92	96.21	96.18	91.35	93.88
$MDCP + DDVP + PCCNN$	99.83	98.85	98.47	97.66	92.80	95.29
$MDCP + DDVP + KNN$	98.05	97.88	93.09	94.9	92.80	90.18
$MDCP + DDVP + CART$	97.74	96.83	94.11	94.77	87.91	89.31
$MDCP + DDVP + RF$	96.25	97	98.54	93.81	88	95.38
$MDCP + DDVP + C4.5$	96.95	96.91	93	94.73	86.39	88
$MDCP + DDVP + AdaBoost$	97.53	96	94.02	93	87.03	88.93
$MDCP + DDVP + RoF$	98.6	97.1	93.64	94.19	88.54	89.47
$MDCP + DDVP + DT$	98.7	98.92	93.84	95	88.69	89.1
$MDCP + DDVP + NB$	99.7	98	93.2	94.04	88.99	89.95

Table 3 Testing of the classification algorithms for coral data sets. The highest accuracy (%) acquired for every data set is highlighted in bold

5.2.3 Specificity

Specificity is defined as the true negative rate that estimates the percentage of correct rejection with diseased coral reef images.

$$
Specificity = \frac{TN}{TN + FP}
$$
\n(42)

where TN represents True Negative rate and FP represents False Positive rate.

Specificity of classifiers is excellent since the true negatives are mostly identified as shown in Table [5](#page-23-0). The specificity for the tree classifiers ranges from 94 to 100% because the true negative rate is good, i.e. misclassification of diseased images is less. PCCNN, SVM and KNN have shown excellent results due to majority voting and support vectors. It also indicates that tree classifiers are selective. DT classifier has better overall specificity compared to other classifiers since it tries to maintain some prediction strength. RF also has good specificity value closer to DT classifier because there is limited generalization error in the classification. The other tree classifiers such as Adaboost, RoF, CART and C4.5 have obtained values that range from 20 to 70% due to fewer trees grown. For Naive Bayes, the specificity values range from

Table 4 Accuracy calculation for SDMRI coral diseased images with DT Classifiers. The highest accuracy (%) acquired for every data set is highlighted in bold

Classifiers	Average	Accuracy percentage for various coral disease classes								
		Asper	BaB	BB	BSDS	WB	WPL	WPO	YB	PS
$MDCP + DDVP + SVM$	86.25	86.7	87.9	86.22	88.92	85.12	86.11	84.38	85.12	85.81
$MDCP + DDVP + KNN$	88.58	91.23	86.08	92.36	86.98	85.18	88.58	91.56	84.79	90.49
$MDCP + DDVP + CNN$	86.72	85.96	86.48	87.21	87.40	85.99	87.20	88.43	83.77	88.06
$MDCP + DDVP + PCCNN$	90.91	93.21	87.2	91.9	90.86	91.3	87.25	90.99	93.62	91.93
$MDCP + DDVP + CART$	84.66	84.85	86.73	83.93	88.39	82.38	84.97	83.53	83.97	83.21
$MDCP + DDVP + RF$	89.55	92.91	86.86	85.82	90.1	87.83	86.83	90.75	93.22	91.64
$MDCP + DDVP + C4.5$	86.90	80.98	86.97	88.4	89.74	90.51	83.02	85.84	86.04	90.61
$MDCP + DDVP + AddBoost$	85.16	85.9	87	82.87	89.32	84.71	84.59	80.98	82.73	88.39
$MDCP + DDVP + RoF$	85.94	84.96	85.99	85.36	88	83.93	82.63	88.61	86.18	87.88
$MDCP + DDVP + DT$	89.85	93.21	87.31	85.92	90.86	87.9	87	90.95	93.62	91.93
$MDCP + DDVP + NB$	85.92	84.79	86.11	88.38	85.27	87.76	85.60	82.59	86.93	85.93

Classifiers	Average	Specificity percentage for various coral disease classes								
		Asper	BaB	BB	BSDS	WB	WPL	WPO.	YB	PS
$MDCP + DDVP + SVM$	95.05	90.32	88.19	100	94.14	99.74	98.5	92.8	99.79	91.98
$MDCP + DDVP + CNN$	92.16	88.95	87.61	100	96.32	91.83	87.62	90.44	97.47	89.23
$MDCP + DDVP + PCCNN$	96	90.61	88.95	100	100	99.27	99.16	93.21	99.18	93.67
$MDCP + DDVP + KNN$	95.55	90.62	88.5	100	100	97.41	97.97	93	98.99	93.51
$MDCP + DDVP + CART$	55.29	48.65	16.67	65.7	55.13	16.67	86.96	26.32	92.34	89.2
$MDCP + DDVP + RF$	92.03	88.56	77.48	100	97.4	90.42	96.59	86.32	97.9	93.67
$MDCP + DDVP + C4.5$	68.58	60.90	21.83	98.6	80.1	71.53	70.75	73.27	71.6	68.67
$MDCP + DDVP + AddBoost$	66.59	58.61	23.64	97.5	77.82	72.66	70.48	71.26	-76.54	50.81
$MDCP + DDVP + RoF$	65.51	77.49	19.23	77.48	86.96	70.53	69.52	73.26	66.67	48.51
$MDCP + DDVP + DT$	95.33	89.79	88.57	99.76	98.56	97.64	97.8	93.84	98.63	93.45
$MDCP + DDVP + NB$	41.47	56.74	19.23	66.67	48.51	48.65	48.51	41.67	23.91	19.37

Table 5 Specificity calculation for various coral diseases with DT Classifiers. The highest Specificity (%) acquired for every data set is highlighted in bold

16 to 49% only which is less due to misclassification rate. For the class Black band, SVM, PCCNN, KNN and Random Forest have yielded full results. Also PCCNN and SVM have obtained the highest results for white Plague, white band and yellow band.

5.2.4 Sensitivity

Sensitivity is a metric that reveals how correctly a classifier classifies the true positives. It estimates the percentage of correctly classified diseased coral reef images.

$$
Sensitivity = \frac{TP}{TP + FN}
$$
\n(43)

where TP represents the True Positive rate and FN represents the False Negative rate.

Sensitivity is good for classes with large sample images, and it is fair for classes with fewer sample images. Table [6](#page-24-0) shows that diseased image classes with large number of samples like black spot dark spot, white band and pink spot, KNN gives highest sensitivity value, but for the least sample class such as Aspergillosis and Black Band, they are not sensitive. DT has the highest overall sensitivity compared to other decision tree classifiers because of the procedure of creating many trees. It is able to classify a few sample images from the classes having fewer samples, i.e. White Pox whereas other classifiers are not able to classify as observed in column 9 of Table [6](#page-24-0). Also, SVM and PCCNN have the highest sensitivity for the classes with fewer samples such as White Plague and Yellow band. Naive Bayes, AdaBoost, CART, C4.5 and Random Forest classifiers are also sensitive to classes with fewer sample especially for Aspergillosis. But Random Forest classifier and PCCNN have the highest sensitivity for Black band class among all the classifiers implemented because they does not need a lot of features to perform well.

5.2.5 Mathew's correlation coefficient (MCC)

It is considered as one of the most effective and accurate performance parameters for any purpose of classification. MCC takes values in the interval (−1and 1), whereby 1 indicates that the classifier classifies the diseased images correctly and − 1 indicates that the classifier classifies the diseased images incorrect by MCC is estimated as shown in Eq. [\(13\)](#page-10-0), for real time diseased data set [\(http://www.sdmri.in\)](http://www.sdmri.in),

Classifiers	Average	Sensitivity percentage for various coral disease classes								
		Asper	BaB	BB	BSDS	WB	WPL	WPO	YB	PS
$MDCP + DDVP + SVM$	83.64	77.34	78.44	90.68	85	74.71	89.22	78.4	89.46	89.58
$MDCP + DDVP + KNN$	84.11	76.9	79.61	91.28	85.33	76.13	88.9	79	89.25	90.51
$MDCP + DDVP + CNN$	80.59	75.94	77.96	88.73	80.47	73.86	86.55	77.42	82.34	82.06
$MDCP + DDVP + PCCNN$	84.58	80.19	81.72	91.66	84.81	75.42	89.19	79.39	89.5	89.37
$MDCP + DDVP + CART$	39.66	40.82	33.33	36.48	43.47	16.67	62.61	30.43	44.64	48.51
$MDCP + DDVP + RF$	81.32	75.12	80.21	91.67	81.39	72.09	85.91	74.38	85.05	86.12
$MDCP + DDVP + C4.5$	38.72	41.42	41.03	39.74	25	65.7	26.32	23.91	76.13	9.23
$MDCP + DDVP + AdaBoost$	46.58	56.28	52.15	34.43	38.61	54.91	61.47	43.47	49.55	28.38
$MDCP + DDVP + RoF$	49.50	52.1	63.81	42.10	39.65	53.88	59.05	48.36	53.76	32.84
$MDCP + DDVP + DT$	84.18	80.18	81.76	90.65	84.71	75.99	88	79.38	88.5	88.51
$MDCP + DDVP + NB$	49.96	64.9	59.23	45.66	37.91	51.04	58.91	47.22	54.81	30

Table 6 Sensitivity calculation for various coral classes with DT classifiers. The highest Sensitivity (%) acquired for every data set is highlighted in bold

$$
MCC = \frac{TP \times TN - FP \times FN}{\sqrt{(TP + FP)(TP + FN)(TN + FP)(TN + FN)}}
$$
(13)

where FN represents False Negative, TP represents True Positive, TN represents True Negative and FP represents False Positive.

The overall highest Mathew's Correlation Coefficient (MCC) value is obtained for SVM, CNN, PCCNN and KNN because they are accurate and powerful classifiers as shown in Table 7. C4.5 and Naive Bayes classifiers have the lowest MCC values due to the problem of over fitting in C4.5 and strong assumption on the shape of the feature distribution in Naive Bayes. From the study, it is found that RF classifier, PCCNN and DT classifier perform better compared to other classifiers based on the performance measures accuracy, specificity, Fmeasure, time and Mathew's Correlation Coefficient for coral disease classification.

5.2.6 Time

It is essential to analyze the time taken by various feature descriptors, classifiers and existing techniques to classify the diseased coral reef images. So as to make it usable for real time

implementations. Tables 8, [9](#page-26-0), and [10](#page-26-0) show the time estimated for feature descriptors, classifiers and comparison with existing techniques. When the feature descriptor's feature size is less it helps in fast and efficient classification. In Table 8, the proposed feature descriptor is about two times faster to compute than LBP, CLBP RLTP and LTrP. Even though LDP, ILDP, OPT and $Z \oplus TZLBP$ take less time, the proposed has improved accuracy by 2% than them.

5.2.7 Computational complexity

LBP, LDP, CLBP, DLBP, LTxXORP, CS-LBP, OC-LBP, PRI-CoLBP, ILDP, Z ⊕ TZLBP and the proposed feature descriptors have a linear Complexity. If there is an iteration, and the iterative variables are incrementing linearly, then time complexity is determined as O(n). RLTP and LTrP techniques have a Quadratic Complexity. This is shown in Table [11](#page-27-0).

5.2.8 Space complexity

The space complexity of an algorithm is the amount of storage it uses. The feature vector that is obtained has the histogram bin size in the range 0 to 256 for LBP, LDP, CLBP, DLBP, LTxXORP and LTrP feature descriptor whereas for RLTP it is between 0 to 512 which is devastating. Hence the size of feature vector and its bin size has an impact on storage requirement. In ILDP and OC-LBP, the bin size is between 0 to 32. For $Z \oplus TZLBP$ and CS-LBP, the bin size is between 0 to 16. For OPT and the proposed technique, the bin size is between 0 to 24. This is presented in Table [11](#page-27-0).

6 Results and discussions

6.1 Accuracy

The accuracy of various classifiers implemented using the same SDMRI diseased coral data set with different feature classes is compared in Fig. [14](#page-27-0). It is observed from Table [4](#page-22-0) that the DT and PCCNN produce higher classification rate, i.e. approximately 89–92% which is greater when compared to other classifiers. RF performs well because it can produce accurate predictions that

Feature Descriptors	Average Time Taken (in secs)
LBP	1.912
CLBP	1.988
DLBP	1.123
LTxXORP	1.008
CS-LBP	0.9865
OC-LBP	0.9736
PRI-CoLBP	1.113
RLTP	1.486
LTrP	1.389
LDP	0.9923
ILDP	0.7581
$Z \oplus TZLBP$	0.6229
OPT	0.6224
Proposed	0.6119

Table 8 Average time reported for the different feature descriptors using SDMRI Diseased data set

Classifiers	Time taken for training and testing (in secs)
SVM	28.65
CNN	95.81
KNN	21.64
PCCNN	37.11
CART	2.82
RF	15.73
C4.5	1.52
AdaBoost	73.81
RoF	64.6
DT	2.03
NB	0.29

Table 9 Time taken by various classifiers for SDMRI diseased data set

do not overfit the data. SVM (RBF kernel), CNN and KNN also provide better results compared to other classifiers. It is also noticed that DT and PCCNN classifiers provide the highest overall percentage of accuracy of 90.91 and 89.85%. Also, it provides highest accuracy for the classification of diseased samples of Pink spot, Aspergillosis, Black spot dark band and Yellow band classes. It is also found that KNN classifier performs well, i.e. it provides the highest accuracy for White plague, Black band and White pox disease classes. This is because White plague, Black band and White pox disease have larger samples compared to other classes and KNN classifier is having the capability to classify problems using majority voting.

The accuracy of various classifiers implemented using SDMRI real-time diseased coral data set with different feature classes are compared in Fig. [14.](#page-27-0) Among various classifiers, DT, PCCNN and RF provide good classification accuracy. KNN also provides better classification results compared to other classifiers. The other classifiers such as Adaboost and CART provide very less classification results because AdaBoost is sensitive to noisy data and outliers. Compared to other classifiers, AdaBoost has overfitting problem.

6.2 Specificity

As shown in Fig. [15,](#page-28-0) SVM, PCCNN, KNN, DT and RF show the highest results of 90 - 96%. Naive Bayes shows the very least results in specificity due to strong feature independence

SDMRI diseased DATA SET	Average Time Taken (in secs)
SHIELA [23] et al. (2008)	0.8747
STOKES [8] et al. (2009)	0.8845
OSCAR [29] et al. (2008)	0.9861
BEIJBOM $[5]$ et al. (2012)	1.199
GUO $[10]$ et al. (2010)	1.988
Mohammad $[32]$ et al. (2017)	1.929
SHIVAHUDIN $[33]$ et al. (2013)	2.776
ANI [2] et al. (2017)	0.7581
ANI [1] et al. (2017)	0.6229
ANI [3] et al. (2018)	0.6224
PROPOSED	0.6119

Table 10 Comparison of the total execution time (in sec.) of SDMRI diseased data set with the existing and the proposed approaches

Feature Descriptors	Bin Sizes used	Computational Complexity		
LBP	256	O(n)		
CLBP	256	O(n)		
DLBP	256	O(n)		
LTxXORP	256	O(n)		
CS-LBP	16	O(n)		
OC-LBP	32	O(n)		
PRI-CoLBP	256	O(n)		
RLTP	512	$O(n^2)$		
LTrP	256	$O(n^2)$		
LDP	256	O(n)		
ILDP	32	O(n)		
$Z \oplus TZLBP$	16	O(n)		
OPT	24	O(n)		
Proposed	24	O(n)		

Table 11 Computational Complexity and Bin Size for various feature descriptors

assumptions. The other classifiers such as AdaBoost, C4.5, CART and Rotation Forest produce average results of 60-70% results.

6.3 Sensitivity

As shown in Fig. [16,](#page-28-0) PCCNN, CNN, SVM, KNN, DT and Random Forest report sensitivity of 80 - 88%. CART and C4.5 show the very least results in sensitivity due to the unstable DT formation and empty branches. The other classifiers such as AdaBoost, Naive Bayes and Rotation Forest produce average results of 45-52%.

Fig. 14 Overall Accuracy of classifiers

Fig. 15 Specificity

6.4 Time

The time taken by various classifiers is shown in Table [9](#page-26-0). Naive Bayes classifier takes less time compared to all the other classifiers studied, but it is not so good in other measures. The other three classifiers such as Adaboost, CNN, Rotation forest except Random forest take more time compared to these classifiers, since it takes more time to build the tree. CART takes less time

Fig. 16 Sensitivity

but other measures are not good. Random forest and PCCNN have a good performance in all aspects because boot strapping reduces bias and variance both which make RF more accurate and robust. Similarly, Adaboost has an average performance in all aspects but in case of time taken it takes more time compared to Random Forest.

The time taken by existing approaches are shown in Table [10](#page-26-0). Even though Shivahudin [\[33\]](#page-37-0) have proved good contribution in accuracy for various coral data sets, their time complexity is more due to utilization of various feature descriptors such as GLCM, Gabor and CLBP. Shiela [[23\]](#page-37-0), Stokes [[8](#page-37-0)], Oscar [[29\]](#page-37-0), Beijbom [[5](#page-36-0)], Guo [[10\]](#page-37-0) and Mohammad [\[32\]](#page-37-0) have provided good classification accuracy with reasonable time complexity. But they did not take any steps to reduce time complexity. Ani [[1](#page-36-0)–[3](#page-36-0)] have concentrated on both improving accuracy and reducing time complexity by reducing the feature vector size of the proposed method.

6.5 Conclusion and future work

The proposed feature descriptors are competent for submarine diseased coral reef images. This paper presents the test results with real-time diseased coral reef data sets, i.e. SDMRI real-time diseased coral image data sets have been used. The proposed feature descriptor extracts the important diagonal information by reducing the size of the feature vector. The combination of the proposed feature descriptor, segmentation and various classifiers attains the best results. MDCP assists in achieving the highest overall classification accuracy and the smallest execution time. Moreover, the proposed feature descriptor is about two times faster to compute than LBP. Therefore, it can produce promising results for a large class of problems.

Even though many feature descriptors are available to improve the classification accuracy, very few works have been reported to reduce the size of feature vectors. CS-LBP, ILDP, $Z \oplus TZLBP$ and OC-LBP operators reduce the size of original LBP histogram with improvement in accuracy. But these feature vectors cover every pixel in an image which is a time-consuming process. Compared to these techniques, the proposed feature descriptor shows its efficiency in classification, feature vector size and time complexity. It outperforms OPT, CS-LBP, OC-LBP, ILDP and $Z \oplus TZLBP$ patterns with the smallest histogram size and maximum accuracy for coral reef image classification.

In this work, various classifiers are studied, and their performances are analysed with various performance measures. Also, the performances of various classifiers are compared. It is concluded that Random Forest and Decision Tree have a good performance to handle the classes with fewer sample images and larger sample images. Naive Bayes classifier is having the least performance compared with the other classifiers.

Experimental results indicate that the proposed feature extraction method achieves the highest overall classification accuracy with minimum execution time when compared to other state-of-the-art methods. From the computational complexity point of view, the proposed feature descriptor provides min size when compared to the other previous feature descriptors, which makes it a faster classifiers. In addition, the speed of the proposed framework is considerably higher than some advanced approaches of coral reef classification. The classification of diseased coral images would enhance the study of coral reef ecosystem, dynamics and processes by reducing the runtime.

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Appendix 1: To calculate the feature extraction using the proposed method do the following: Consider a 3 \times 3 Block B_k in image I as shown below

Step 1: Calculate the diagonal directional difference between four directions, namely 45° , 135° , 225° and 315° on three channels, namely red, green and blue as shown in Eqs. ([4](#page-8-0))–[\(12](#page-10-0)).

$$
G_{45}^{\circ} = 11 - 22 = -11
$$

$$
G_{135}^{\circ} = 11 - 8 = 3
$$

$$
G_{225}^{\circ} = 11 - 12 = -1
$$

$$
G_{315}^{\circ} = 11 - 5 = 6
$$

Step 2: Calculate the diagonal direction difference between three channels along 45° , in the following combinations, namely RG, RB, GR, GB, BG, BR as shown in Eqs. [\(4](#page-8-0))–([12\)](#page-10-0).

Step 3: Calculate the diagonal direction difference between three channels along 135°, in the following combinations, namely RG, RB, GR, GB, BG, BR as shown in Eqs. [\(4](#page-8-0))–([12\)](#page-10-0).

Step 4: Calculate the diagonal direction difference between three channels along 225° , in the following combinations, namely RG, RB, GR, GB, BG, BR as shown in Eqs. [\(4](#page-8-0))–([12\)](#page-10-0).

Step 5: Calculate the diagonal direction difference between three channels along 315° , in the following combinations, namely RG, RB, GR, GB, BG, BR as shown in Eqs. [\(4](#page-8-0))–([12\)](#page-10-0).

Step 6: Vector Generation using four diagonal directions in the three planes is as shown in Eqs. (13) (13) – (15) .

For 45°

$$
R_{45^\circ} = \langle 8, 7, -7 \rangle
$$

$$
G_{45^\circ} = \langle -11, 3, 4 \rangle
$$

$$
B_{45^\circ} = \langle 4, 5, -10 \rangle
$$

For 135°

For 225°

For 315°

$$
R_{135^{\circ}} = \langle 10, 7, 6 \rangle
$$
\n
$$
G_{135^{\circ}} = \langle 3, 3, 6 \rangle
$$
\n
$$
B_{135^{\circ}} = \langle 3, 3, 6 \rangle
$$
\n
$$
B_{135^{\circ}} = \langle 3, 7, 4 \rangle
$$
\nFor 225°\n
$$
R_{225^{\circ}} = \langle 5, 3, 4 \rangle
$$
\n
$$
G_{225^{\circ}} = \langle -1, 0, 1 \rangle
$$
\n
$$
B_{225^{\circ}} = \langle 1, 2, 0 \rangle
$$
\nFor 315°\n
$$
R_{315^{\circ}} = \langle 6, 10, 11 \rangle
$$
\n
$$
G_{315^{\circ}} = \langle 6, 7, 2 \rangle
$$
\n
$$
B_{315^{\circ}} = \langle 8, 3, 7 \rangle
$$
\nStep 7: Binary Codes Assignment is as shown in Eqs. (16)–(18)\nFor 45°\n
$$
BCR_{45^{\circ}} = \langle 1, 1, 0 \rangle
$$
\n
$$
BCB_{45^{\circ}} = \langle 0, 1, 1 \rangle
$$
\n
$$
BCB_{45^{\circ}} = \langle 1, 1, 0 \rangle
$$
\nFor 135°

 $\text{BCR}_{135^\circ} = \langle 1, 1, 1 \rangle$

 $BCG_{135^{\circ}} = \langle 1, 1, 1 \rangle$

 $BCB_{135^{\circ}} = \langle 1, 1, 1 \rangle$

For 135°

² Springer

For 225°

```
BCR_{225^\circ} = \langle 1, 1, 1 \rangleBCG_{225}^{\circ} = \langle 0, 1, 1 \rangleBCB_{225^\circ} = \langle 1, 1, 0 \rangleBCR_{315^{\circ}} = \langle 1, 1, 1 \rangleBCG_{315^\circ} = \langle 1, 1, 1 \rangleBCB_{315^\circ} = \langle 1, 1, 1 \rangleStep 8: Diagonal Direction Value Assignment is as shown in Eqs. (19)–(21).
                                                                DDV_BCR<sub>45</sub>° = \langle 7 \rangleDDV_BCG<sub>45</sub>° = \langle 4 \rangleDDV\_BCB_{45}^{\circ} = \langle 7 \rangleDDV_BCR<sub>135</sub>° = \langle 8 \rangleDDV_BCG<sub>135</sub>° = \langle 8 \rangleDDV_BCB_{135^\circ} = \langle 8 \rangleDDV BCR<sub>225</sub>° = \langle 8 \rangleDDV_BCG<sub>225</sub><sup>\epsilon</sup> = \langle 4 \rangleDDV_BCB_{225} = \langle 7 \rangle
```
For 315°

For 45°

For 135°

For 225°

For 315°

```
DDV_BCR<sub>315</sub>° = \langle 8 \rangleDDV BCG<sub>315</sub>° = \langle 8 \rangleDDV BCB<sub>315</sub>° = \langle 8 \rangleStep 9: Summed Diagonal Direction Value is as shown in Eq. (22).
                                                     SDDV_{45}^{\circ} = 18SDDV_{135} = 24
```
 $SDDV_{225^{\circ}} = 19$

$$
\mathrm{SDDV}_{315^\circ}=24
$$

Finally feature vector for DDVP is estimated as follows $SDDV = < 18, 24, 19, 24>$ Step 10: Given image I is converted from RGB to HSV color space

H-Plane S-Plane V-Plane

Step 11: Mean Estimation for three planes

As shown in Eq. (26) HMean is estimated for H-Plane, HMean = 5

As shown in Eq. [\(27\)](#page-13-0) SMean is estimated for S-Plane, SMean = 10

As shown in Eq. [\(28\)](#page-13-0) VMean is estimated for V-Plane, VMean = 9

Step 12: Diagonal Neighbors Estimation, As shown in Fig. [11](#page-15-0), Diagonal neighbors are chosen for comparing with the Mean Value in three planes.

Step 14: Summed Direct Code, After comparison of diagonal elements with neighbors. Depending on the binary values 0, 1 the codes are assigned. For this 3×3 Block B_k , the feature vector is estimated in three planes and summed together as shown in Eq. ([35\)](#page-17-0).

 $SDC = 7 + 14 + 11 = 32$

Step 15: After Normalization as shown in Eq. (36) , SDDC = 16 Step 16: Feature Vector Concatenation as shown in Eq. [\(37\)](#page-17-0) Feature Vector (I) = <18, 24, 19, 24, 16>

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References

- 1. Ani Brown Mary N, Dejey D (2017) Classification of coral reef submarine images and videos using a novel Z with tilted Z local binary pattern (Z⊕TZLBP). Wirel Pers Commun. [https://doi.org/10.1007/s11277-017-](https://doi.org/10.1007/s11277-017-4981-x) [4981-x](https://doi.org/10.1007/s11277-017-4981-x)
- 2. Ani Brown Mary N, Dharma D (2017) Coral reef image classification employing improved LDP for feature extraction. Elsevier, Journal of Visual Communication and Image Representation 49:225–242
- 3. Ani Brown Mary N, Dharma D (2018) Coral reef image/video classification employing novel octa-angled pattern for triangular sub region and pulse coupled convolutional neural network (PCCNN). Springer, Multimed Tools Appl. <https://doi.org/10.1007/s11042-018-6148-5>
- 4. Bala A, Kaur T (2016) Local texton XOR patterns: a new feature descriptor for content based image retrieval. ELSEVIER, Engineering Science and Technology, an International Journal 19:101–112
- 5. Beijbom O, Edmund PJ, Kline DI, Greg Mitchell B, Kriegman D (2012) Automated annotation of coral reef survey images. IEEE Conference on Computer Vision and Pattern Recognition (CVPR), Providence, pp 16–21
- 6. Bourne DG, Ainsworth TD, Pollock FJ, Willis BL (2014) Towards a better understanding of white syndromes and their causes on Indo-Pacific coral reefs. Coral Reefs
- 7. Cheng HD, Jiang XH, Wang J (2002) Color image segmentation based on homogram thresholding and region merging. Elsevier, Pattern Recognition 35:373–393
- 8. Dale Stokes M, Deane GB (2009) Automated processing of coral reef benthic images. Limnol Oceanogr Methods 7:157–168
- 9. Grover N (2014) A study of various fuzzy clustering algorithms. International Journal of Engineering Research 3(3):177–181
- 10. Zhenhua Guo, Lei Zhang, and David Zhang (2010) A Completed Modeling of Local Binary Pattern Operator for Texture Classification. IEEE Trans Image Proc 19(6)
- 11. Heikkil M, Pietikainena M, Schmid C (2009) Description of interest regions with local binary patterns. Elsevier, Pattern Recognition 42:425–436
- 12. Heikkila M, Pietikainen M (2006) A texture-based method for modelling the background and detecting moving objects. IEEE Trans Pattern Anal Mach Intell 28(4):657–662
- 13. Heikkila M, Pietikainen M, Schmid C (2009) Description of interest regions with local binary patterns. Elsevier, Pattern recognition 42(3):425–436
- 14. Joseph Pollock F, Morris PJ, Willis BL, Bourne DG (2011) The urgent need for robust coral disease diagnostics. PLoS Pathog 7(10):e1002183
- 15. Kavzoglu T, Colkesen I, Yomralioglu T (2015) Object-based classification with rotation forest ensemble learning algorithm using very-high-resolution WorldView-2 image. Remote Sensing Letters 6(11):834–843. <https://doi.org/10.1080/2150704X.2015.1084550>
- 16. Liao S, Law MWK, Chung ACS (2009) Dominant Local Binary Patterns for Texture Classification. IEEE Trans Image Proc 18(5)
- 17. Lin S-W, Chen S-C (2012) Parameter determination and feature selection for C4.5 algorithm using scatter search approach. Springer Soft Computing 16:63–75. <https://doi.org/10.1007/s00500-011-0734-z>
- 18. Liu Y, Zhang L, Nie L, Yan Y, Rosenblum DS (2016) Fortune teller: predicting your career path. Proc Thirtieth AAAI Conf Artif Intel (AAAI-16)
- 19. Liu Y, Nie L, Liu L, Rosenblum DS (2016) From action to activity: sensor-based activity recognition. Elsevier, Neuro computing 181:108–115
- 20. Liu Y, Zheng Y, Liang Y, Liu S, Rosenblum DS (2016) Urban water quality prediction based on multi-task multi-view learning. Proc Twenty-Fifth Int Joint Conf Artif Intel (IJCAI-16), 2576–2582
- 21. Liuy Y, Niey L, Hanx L, Zhangy L, Rosenblum DS (2015) action2activity: recognizing complex activities from sensor data. IJCAI
- 22. Mantovani RG Horváth T, Cerriz R, Vanschorenx J, de Carvalho ACPLF (2016) Hyper-parameter Tuning of a Decision Tree Induction Algorithm", 5th Brazilian Conference on Intelligent Systems
- 23. Marcos MSA, David L, Peñaflor E, Ticzon V, Soriano M (2008) Automated benthic counting of living and non-living components in Ngedarrak reef, Palau via subsurface underwater video. Environ Monit Assess 145:177–184
- 24. Moberg F, Folke C (1999) Ecological goods and services of coral reef ecosystems. Ecol Econ 29:215–233
- 25. Murala S, Maheshwari RP, Balasubramanian R (2012) Local tetra patterns: a new feature descriptor for content-based image retrieval, IEEE Trans Image Process 21(5)
- 26. Ojala T, Pietikainen M, Maenpaa T (2002) Multiresolution gray-scale and rotation invariant texture classification with local binary patterns. IEEE Trans Pattern Anal Mach Intel 24(7)
- 27. Ojala T, Pietikainen M, Harwood D (1996) A comparative study of texture measures with classification based on featured distributions. Elsevier, Pattern Recognition 29(1):51–59
- 28. Pérez JF, Gómez A, Giraldo JH, Guzmán S, Fernández DS (2015) Automatic segmentation of coral reefs implementing textures analysis and color features with Gaussian mixtures models. 6th Latin-American Conference on Networked and Electronic Media, LACNEM
- 29. Pizarro O, Rigby P, Colquhoun J (2008) Towards image-based marine habitat classification. Oceans 1–7
- 30. Qi X, Xiao R, Guo J, Zhang L (2013) Pairwise rotation invariant co-occurrence local binary pattern
- 31. Satpathy A, Jiang X, Eng H-L (2014) LBP-based edge-texture features for object recognition, IEEE Transactions on Image Processing 23(5)
- 32. Shakoor MH, Boostani R (2017) A novel advanced local binary pattern for image-based coral reef classification. SPRINGER, Multimedia Tools and Applications
- 33. Shihavuddin ASM, Gracias N, Garcia R, Gleason ACR, Gintert B (2013) Image-based coral reef classification and thematic mapping. Remote Sens 5:1809–1841
- 34. Spalding MD, Grenfell AM (1997) New estimates of global and regional coral reef areas. Coral Reefs 16: 225–230
- 35. Villon S, Chaumont M, Subsol G, Villéger S, Claverie T, Mouillot D (2016) Coral reef fish detection and recognition in underwater videos by supervised machine learning : Comparison between Deep Learning and HOG+SVM methods. Springer, 17th International Conference on Advanced Concepts for Intelligent Vision Systems, Image Processing, Computer Vision, Pattern Recognition, and Graphics (10016)
- 36. Wagsta K, Cardie C, Rogers S, Schroedl S (2001) Constrained K-means Clustering with Background Knowledge. Eighteenth Int Conf Mach Learn 577–584
- 37. Xu Y, Cui J, Zhao H, Zha H (2011) Tracking Generic Human Motion via Fusion of Low- and High-Dimensional Approaches. BMVC
- 38. Zhang B, Gao Y, Zhao S, Liu J (2010) Local derivative pattern versus local binary pattern: face recognition with high-order local pattern descriptor, IEEE Transactions on Image Processing 19(2)
- 39. Zhu C, Bichot C-E, Chen L (2013) Image region description using orthogonal combination of local binary patterns enhanced with color information. Elsevier, Pattern Recognition 46:1949–1963

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