



New insights into the emerging effects of inflammatory response on HDL particles structure and function

Xin Su¹ · Guoming Zhang¹ · Ye Cheng¹ · Bin Wang¹

Received: 13 May 2021 / Accepted: 8 July 2021 / Published online: 28 July 2021
© The Author(s), under exclusive licence to Springer Nature B.V. 2021

Abstract

According to the increasing results, it has been well-demonstrated that the chronic inflammatory response, including systemic lupus erythematosus, rheumatoid arthritis, and inflammatory bowel disease are associated with an increased risk of atherosclerotic cardiovascular disease. The mechanism whereby inflammatory response up-regulates the risk of cardio-metabolic disorder disease is multifactorial; furthermore, the alterations in high density lipoprotein (HDL) structure and function which occur under the inflammatory response could play an important modulatory function. On the other hand, the serum concentrations of HDL cholesterol (HDL-C) have been shown to be reduced significantly under inflammatory status with remarked alterations in HDL particles. Nevertheless, the potential mechanism whereby the inflammatory response reduces serum HDL-C levels is not simply defined but reduces apolipoprotein A1 production. The alterations in HDL structure mediated by the inflammatory response has been also confirmed to decrease the ability of HDL particle to play an important role in reverse cholesterol transport and protect the LDL particles from oxidation. Recently, it has been shown that under the inflammatory condition, diverse alterations in HDL structure could be observed which lead to changes in HDL function. In the current review, the emerging effects of inflammatory response on HDL particles structure and function are well-summarized to elucidate the potential mechanism whereby different inflammatory status modulates the pathogenic development of dyslipidemia.

Keywords Inflammatory response · Endotoxin · HDL · Structure · Function · Reverse cholesterol transport

Introduction

According to the results of previous studies, it has been shown that the chronic inflammatory disorders disease, including systemic lupus erythematosus (SLE), rheumatoid arthritis (RA), and inflammatory bowel disease (IBD) are associated with an increased risk of atherosclerotic cardiovascular disease [1]. For instance, in a meta-analysis which enrolled 24 clinical trial, the authors demonstrated that the patients with RA presented an approximately 48 % increase of the pathogenic development of cardio-metabolic disorder disease mortality [2]. However, within the patients under

the SLE status, the authors demonstrated an approximately 2.7 increase of the risk of cardio-metabolic disorder disease mortality via a meta-analysis which enrolled 12 clinical trials [3]. Consistent with these findings, another meta-analysis which contained 14 clinical trials revealed that in patients with severe psoriasis, as another important chronic inflammatory diseases, the risk of cardio-metabolic disorder diseases mortality was approximately 1.40 and the risk of acute myocardial infarction (AMI) was approximately 3.10 higher than those within the general population [4]. Furthermore, under the condition of chronic inflammatory status, the patients have been confirmed to have an up-regulated risk of thickening carotid intima media calculated by ultrasound and the coronary artery calcium measured by tomography (CT) [5–7]. Moreover, several other chronic infections diseases such as human immune-deficiency virus (HIV), periodontal disease, and chronic bronchitis have also been strongly correlated with an increased risk of cardio-metabolic disorder diseases, presumably owing to the chronic inflammatory condition which accompanies these inflammatory diseases

✉ Ye Cheng
chengyeheart@163.com

✉ Bin Wang
wangbinheart@163.com

¹ Department of Cardiology, The Xiamen Cardiovascular Hospital of Xiamen University, No. 2999 Jinshan Road, Xiamen 361000, Fujian, China

[8, 9]. On the other hand, the potential mechanisms whereby inflammatory condition modulates the risk of cardio-metabolic disorder diseases is likely to be multifactorial factor. In the current review, the emerging effects of inflammatory response on HDL particles structure and function are well-summarized to elucidate the potential mechanism whereby different inflammatory status modulates the pathogenic development of dyslipidemia.

Modulatory functions of inflammatory response on serum HDL-C concentrations

While inflammatory response induces a variety of alterations in serum concentrations of lipid profiles and lipoproteins, including increased serum concentrations of triglycerides (TG), one of the most consistent alterations is a reduced plasma concentrations of HDL-C [10]. In patients with severe RA, there is a similar reduced serum concentrations of HDL-C and apolipoprotein A1 (ApoA1) [11]. Meanwhile, small HDL are preferentially reduced significantly [12]. The degree of decreased in HDL particles is shown to be greatest within the patients under the condition of severe RA. Furthermore, there is a negative association of the serum levels of C-reactive protein (CRP) with the serum concentrations of HDL-C, including higher plasma CRP concentrations are associated with lower serum concentrations of HDL-C. Consistent alterations in HDL have also been found in patients with SLE, RA, and IBD [13]. Under the status of infection with HIV, the authors observed an early metabolic response was down-regulated in HDL particles, which occurs prior to an elevated in serum concentration of TG [14, 15].

In addition, a recent clinical trial demonstrated that the serum HDL-C concentrations were less than 10 mg/dl within the patient who suffered from severe sepsis, indicating that the status of chronic inflammatory condition could significantly modulate the serum HDL-C concentrations [16]. Moreover, the decreased plasma concentrations of HDL-C which occurs during the infection condition could be reproduced via using the animal models which was administrated by endotoxin to make an experimental animal model of gram (–) infection [13]. Additionally, several investigations have put forward that the extent of decrease in serum concentrations of HDL-C and ApoA1 could be used for the prediction of mortality within the patients who suffer from severe sepsis [17–19]. Notably, both animals and humans studies revealed that the lower serum concentrations of HDL-C were closely correlated with a melodramatic response to the administration of endotoxin [20–22].

Modulatory effects of inflammatory response on HDL particles function

As one of most vital proteins within circulation, HDL embrace several important effect which has been shown to inhibit the development of cardio-metabolic disorder diseases and to be adversely influenced by the progression of inflammatory response. In the present article, the impact of inflammatory response on modulating the functions of HDL particles, especially affecting the process of RCT and protecting LDL particles from oxidation, is well-summarized.

Modulatory effect of inflammatory response on RCT process

The step of excess cholesterol moving from the macrophage in reverse to the hepatocyte for subsequent excretion in the bile, not matter as the form of cholesterol or the form of bile acids, is a comprehensive progression that contains several steps. On the other hand, the vivo studies have revealed that the process of RCT could be measured by loading the macrophage with injecting the labelled cholesterol and next assessing the labelled cholesterol within the circulation, hepatocyte, and egesta [23]. Additionally, via this technical measurement, multiple research has demonstrated that intervention of the mice with endotoxin which lead to severe inflammatory response could further induce an inhibition of RCT [24–26]. Importantly, treatment with the relatively lower dose of endotoxin has been shown to induce a similar reduction of RCT process, revealing that this alteration in the progression of RCT is a sensitive response to the inflammatory status [25].

Recently, using the SAA gene-deficient mice with the administration of endotoxin, the authors showed that these mice had a relatively reduced extent of RCT, suggesting that the elevated serum levels of SAA is not vital for the reduction in RCT process [27]. Nevertheless, the mice with over-expressed SAA gene presented the similar extent of RCT compared to those in the mice which occurred under the inflammatory response, shedding light on that SAA is not vital in modulating RCT [20]. Furthermore, consistently elevating myeloperoxidase to the extents which is similar to that occurs under the inflammatory condition could also inhibit the process of RCT [20]. Similar with these findings, the circulating myeloperoxidase concentrations were shown to be increased significantly under the status of inflammation; meanwhile, the myeloperoxidase-induced oxidation of HDL particles were also reduced which lead to impaired RCT and the inhibition of LCAT [21, 22, 28]. On the contrary, elevated circulating levels

of solute PLA presented no significant function on RCT [20]. Interestingly, the functions of elevated serum levels of SAA or myeloperoxidase on modulating the process of RCT were relatively moderate compared with the regulatory function of severe inflammatory response, showing that several changes might facilitate the impaired extent of RCT [20].

Modulatory effect of inflammatory response on extracellular HDL metabolism

According to the reported article, the inflammatory condition could impair the activity of LCAT which plays an important role in inhibiting the exchange of cholesterol to EC within HDL particles [29]. Importantly, the above step has been considered as the most vital step of the HDL particles maturation which promote HDL to accept excessive cholesterol from macrophages. Reducing the LCAT activity in matured HDL particle has been shown to reduce the effect of HDL to mediate RCT [30]. Similarly, the impaired extent of RCT via inhibiting LCAT activity is almost the same as the impaired extent under the inflammatory status.

On the other hand, the inflammatory response is confirmed to inhibit the activity of CETP and as a consequence, inhibit the transportation of excessive cholesterol from HDL particles to Apo-B containing lipoproteins, including VLDL, LDL, and CM [31]. Notably, this process has been considered as an essential pathway of transferring the cholesterol to the hepatocytes in human beings [32]. Moreover, the inflammatory response could also reduce the expression levels of hepatic lipase (HL), as an essential rate-limiting enzyme within the hepatocyte which plays an important role in removing TG from HDL, and lead to an inhibition in serum HL activity which resultantly influenced the biological function of hepatocytes [33].

Modulatory effect of inflammatory response on anti-oxidant function of HDL

According to the published results, one of the essential properties of HDL particles is to prevent the LDL from oxidation and to inhibit the formation of ox-LDL. Via this function, HDL could improve the process of atherosclerosis [34]. Up to date, there is emerging evidence which showed that several HDL-related proteins, such as ApoA1, paraoxonase 1 (PON1), and transferrin protein, could affect the anti-oxidant functions of HDL proteins [35, 36]. Via the injection of croton oil into rabbits to lead to severe inflammatory response, the authors demonstrated that the HDL particle within the circulation of these rabbits presented loss-of-function alterations in protecting the LDL particles from oxidation [37, 38].

Aside from the croton oil, several other administration of medicine could also modulate the function of HDL. For instance, the mice with acute influenza infection presented a significantly impaired anti-oxidant function of HDL which were inclined to suffer from atherosclerosis [39]. In addition, the serum HDL particles isolated from the patients with chronic inflammatory diseases, such as SLE, RA, and IBD, has been shown to have a reduced function to inhibit LDL from formatting the ox-LDL. Indeed, the loss-of-function of HDL may reversely promote the LDL oxidation and facilitate the development of atherosclerosis [40, 41]. The anti-oxidative function of HDL has been also shown to be strongly associated with the severity of the disease. On the contrary, using the treatment which inhibit the pro-inflammatory response and improve the disease activity could significantly restore the anti-oxidant functions of HDL particles towards normal [42, 43]. With in-depth investigation, under the condition of inflammatory response, the pro-inflammatory cytokines could significantly reduce the expression levels of *PON1* gene and down-regulate the activity of PON1 protein, which could subsequently contribute to reduce the function of HDL particle in inhibiting LDL from oxidation [44]. By contrast, using the cultured cell lines, the authors found that the serum ceruloplasmin concentrations increased under the inflammatory status. As a pro-inflammatory proteins, ceruloplasmin could facilitate the oxidation of LDL particles, so we could infer from these findings that ceruloplasmin modulate the risk of dyslipidemia might via oxidation of LDL [43, 45].

On the other hand, the serum concentrations of transferrin proteins has been confirmed to reduce significantly under the progression of inflammatory response. Besides, multiple research have suggested that the transferrin protein could also protect LDL from oxidation [46, 47]. However, using the intervention of endotoxin to mice could induce the up-regulated concentrations of multiple bio-markers of per-oxidation in serum LDL particles, indicating that the inflammatory response could facilitate the risk of dyslipidemia [48, 49]. Additionally, LDL particles isolated from endotoxin-intervened mice is more inclined to oxidation and to foam cells [49]. In addition, other bio-markers of lipid peroxidation, such as conjugated dienes and lipid hydroperoxides, has been shown to up-regulated within circulation in patients with inflammatory diseases [50–52].

During the process of cardio-metabolic disorder diseases, the oxidant LDL particles within circulation could be taken up via endocytosis induced by macrophages. By this methods, the macrophage has been considered as the most important cell which plays an essential role within the formation of the foam cell. Furthermore, using the anti-bodies to the oxidant LDL particles isolated from the patients with IBD, SLE, and psoriasis, the authors found that these LDL particles presented the increased uptake of

the anti-body via the Fc-receptor on the surface of macrophages, indicating that under the condition of inflammatory status, the abnormal macrophages could affect the function of LDL metabolism [53, 54]. Consistently, several other research has demonstrated that the LDL particles isolated from individuals with periodontal disease presented the increased endocytosis of CE by macrophages [55]. However, due to the lack of literatures, the potential underlying mechanisms accounting for the enhanced oxidation of LDL particles under the inflammatory conditions is still needed to be further elucidated.

Modulatory effect of inflammatory response on exchange of cholesterol from macrophages to HDL

As is known to us, the first step in the reverse cholesterol transport (RCT) pathway is the movement of cholesterol from macrophages to HDL. Studies have demonstrated that inflammatory stimuli decrease macrophage expression of ABCA1, ABCG1, and SR-B1, transporters that mediate the efflux of cholesterol to HDL [56–58]. Additionally, Apo E expression in macrophages is also decreased by inflammatory stimuli, which could also impede cholesterol efflux [59, 60]. As noted earlier the levels of HDL and Apo A-I are decreased during inflammation, which could limit the acceptors available to promote efflux of cholesterol. Moreover, the structurally altered HDL formed during inflammation is a poor acceptor of cholesterol [61]. As described earlier, HDL enriched in SAA has a decreased ability to mediate cholesterol efflux [62]. In fact some studies have shown that inflammatory HDL instead of removing cholesterol from cells actually delivers cholesterol to cells [61]. HDL isolated from patients with RA, SLE, psoriasis, periodontal disease, HIV infection, and acute sepsis has been shown to be poor mediators of the efflux of cholesterol from macrophages [11, 19, 32, 63].

Furthermore, the administration of LPS to humans also results in the formation of HDL that is a poor facilitator of the efflux of cholesterol from macrophages [8]. Importantly, treatment that reduces inflammation has been shown to improve the ability of HDL from patients with RA, psoriasis, HIV infection, and periodontitis to mediate the efflux of cholesterol from macrophages [64].

Notably, under the condition of HIV infection, the circulating macrophage might also impair the process of excessive cholesterol efflux via down-modulating the ABCA1 concentrations through the NEF protein. It is worth noting that NEF protein could disrupt the exchange of ABCA1 from the endoplasmic reticulum (ER) to the plasma membrane and consequently activating the degradation of ABCA1 [65].

Modulatory effect of inflammatory response on cholesterol uptake by hepatocytes

As is shown in previous research, the cholesterol which is carried by HDL particles has been shown to be delivered into the hepatocyte by two methods. As reported, one of the methods is that the HDL particle could interact with scavenger receptor B1 (SR-B1), which could exchange the excessive cholesterol from the circulating HDL particles into the hepatocyte for the subsequently regenerating smaller HDL particles. It is worth noting that under the inflammatory conditions, several pro-inflammatory cytokines could down-regulate the SR-B1 expression within hepatocyte which resultantly induced an impaired endocytosis of HDL, indicating that the inflammatory process may promote the development of cardio-metabolic disorder disease via affecting the SR-B1 expression [66].

On the other hand, as reported in previous literatures, the cholesterol within HDL particle could be transferred into the ApoB containing lipoproteins, including LDL and VLDL, which subsequently delivered the excessive cholesterol into the hepatocyte by the LDL receptor (LDLR) induced by the endocytosis of ApoB. Concerning on this notion, several in vivo research has put forward that the inflammatory response could reduce the expression of LDLR on the surface of hepatocyte. However, this modulatory function which could not be accounted for by altering in *LDLR* gene concentrations [67]. Nevertheless, the expression levels of *PCSK9* gene in the hepatocyte has been activated by the inflammatory response. Since the PCSK9 within circulation could significantly inhibit the expression of LDLR and resultantly modulate the serum concentrations of LDL-C; as a consequence, this might explain the reduction in LDLR and the increase of serum LDL-C [68]. In addition, the circulating PCSK9 concentrations has been confirmed to be increased in individuals with periodontal inflammation [69]. A reduction in LDLR which is hand in hand with the reduction in serum contents of CETP could synergistically inhibit the HDL-C to be delivered into the hepatocyte by exchange with LDL and VLDL.

Modulatory effects of inflammatory response on HDL particles composition

Aside from the modulatory role in reducing the serum levels of HDL-C, emerging evidence has shown that the inflammatory response could also induce in significant changes in the intra-molecular composition of HDL particles [1]. To be more specific, in terms of the intra-molecular lipid components, several studies has indicated that the concentrations of cholesterol ester are reduced whereas the concentrations of free cholesterol, TG, and free fatty acids (FFA) are increased

remarkably. Additionally, there are remarked alterations in HDL-related proteins with some up-regulating and the others down-regulating. Among these altered components, the most profound alteration is an up-regulated concentrations of serum amyloid A (SAA) and a down-regulated concentrations of ApoA1. Importantly, according to the results, diverse modulatory functions on multiple enzymes and transfer proteins which are strongly correlated with HDL particle. In the following paragraph, the important alterations which occur in the essential proteins that are involved in HDL structure and catabolism has been well-documented.

Alterations in the concentrations of lecithin:cholesterol acyltransferase (LCAT)

Due to the technological advances, major breakthroughs have been made to elucidate the alterations in the concentrations of LCAT. As is known to us, the LCAT is one of the important proteins which has been confirmed to convert the free cholesterol within the HDL particles into cholesterol esters (CE). Importantly, this step has firmly been shown to be essential for the process of RCT and as a consequence, modulates the serum levels of lipid profiles. Multiple studies has used different animal models to further explore the modulatory functions of diverse interventions on the LCAT activity. For instance, administration of endotoxin or pro-inflammatory cytokines, such as tumor necrosis factor α (TNF- α), to hamsters, monkeys, and humans could lead to a significantly reduced LCAT activity within circulation [70]. In addition, in hamsters, the significantly reduction in LCAT activity has also been shown to be accompanied by an importantly reduction in the expression levels of *LCAT* gene within hepatocytes [71]. Furthermore, in patients with RA or severe sepsis, the circulating LCAT activity has been verified to be decreased importantly, suggesting that the inflammatory conditions could modulate the serum LCAT activity [72]. Nevertheless, an independent research demonstrated that under the condition of HIV infection, the circulating LCAT concentrations could be up-regulated importantly [73].

Alterations in the concentrations of cholesterol ester transfer protein (CETP)

CETP is an important protein within circulation which plays a vital role in mediating the exchange of intra-molecular cholesterol esters from the HDL particles for TG that was carried on apolipoprotein B (ApoB) lipoproteins. As a consequence, CETP could promote the process of RCT and resultantly modulate the serum levels of lipid profiles. Due to the potential role in modulating the risk and the development of dyslipidemia, the altered activity of CETP under the inflammatory condition has been given substantial attention

in recent year. Notably, an independent research using hamsters and found that via the administration of endotoxin, several pro-inflammatory cytokines, including TNF- α , Interleukin-1 β (IL-1 β), and IL-12, could significantly reduce the activity of CETP within circulation; meanwhile, the expression levels of *CETP* gene and CETP protein within different cells, such as adipocytes, myocardial cells, and skeletal muscle cells, were increased significantly, indicating that the pro-inflammatory cytokines could regulate the expression levels of CETP and subsequently influence the serum levels of lipids profiles [74].

Consistently, another important research demonstrated that via using endotoxin on the mice with over-expressing *CETP* gene, the mice presented a rapid remarkably decreased *CETP* gene within hepatocyte; simultaneously, the serum concentrations of CETP were also down-regulated significantly [75]. With in-depth investigations, the authors also revealed that in these mice, the reduced expression levels of *CETP* gene could be significantly regulated by the up-regulated glucocorticoid concentrations. However, the reduced CETP expression was confirmed to be independent of the alterations of glucocorticoids in hamsters [75]. Two independent research showed that in patients with sepsis, administration of endotoxin was found to reduce the serum CETP concentrations significantly, revealing that the pro-inflammatory status could also modulate the expression levels of CETP [70].

On the other hand, the cardiac surgery which also has the potentiality to facilitate the pro-inflammatory condition has been confirmed to down-regulate the serum CETP concentrations and its activity [76]. In addition, the patients under the status of RA presented the reduced plasma CETP activity. Nevertheless, the alterations of CETP activity could be only observed in the individuals who were under-taking the glucocorticoids [77]. Recently, another important research found that under the condition of HIV infection, the serum CETP concentrations has been shown to be up-regulated which was consistent with the paradoxical increase in the LCAT activity [73]. Taken together, we also needed further large-scale clinical trials and in-depth research which could elucidate the accurate relationship between the inflammatory condition with the altered activity of several proteins that play essential role in modulating the risk and the development of dyslipidemia.

Alterations in the concentrations of PON1

As reported in previous studies, the PON1 protein is one of the most important HDL-associated proteins which could protect LDL particles from oxidation and resultantly inhibit the development of atherosclerosis and its related cardio-metabolic disorder diseases [78]. The important function of PON1 in regulating lipid metabolism under the

inflammatory status has begun to gain appreciation since the PON1 could modulate the risk of dyslipidemia in mammals. To be more specific, one research used the mice with *PON1* gene deficiency and found that these mice presented reduction in anti-oxidant function of HDL particles; on the contrary, while used the mice with over-expression of *PON1* gene, the authors found that these mice presented increased anti-oxidant function of HDL particles and the increased protective function to atherosclerosis [78]. In addition, the mice with *PON1* gene deficiency were also shown to be more inclined to cardio-metabolic disorder disease and the lipoproteins within circulation isolated from these mice are more susceptible to be oxidized easily [78]. Another important research has also confirmed that under the inflammatory status in both humans and rabbits, the process of inflammation could significantly lead to a reduction of PON1 activity which inhibited the ability of PON1 to protect the LDL against oxidation and the risk of dyslipidemia [79]. Consistent with this notion, it has been verified that administration of endotoxin could lead to a significantly reduced expression of *PON1* gene in hepatocyte hand in hand with a decreased activity of PON1 within circulation in hamsters [80]. Similarly, several other pro-inflammatory cytokines, such as TNF- α and IL-1 β , has also confirmed to have the function to modulate the function in influencing serum lipid profiles [80]. It is also worth noting that the decreased expression levels of *PON1* gene in the hepatocyte and the activity precedes the appearance of LDL-oxidation which follows the administration of lipopolysaccharide (LPS), indicating that the down-regulated expression of *PON1* gene might contribute to the increased extent of LDL oxidation [49]. Likewise, the serum concentrations of PON1 has been found to be reduced in patients with several other infective diseases, such as SLE, IBD, and psoriasis [81]. Besides, after the treatment in patients with psoriasis for a period, the serum concentrations of PON1 increased significantly, putting forward the evidence that anti-inflammatory treatment could improve the functions of PON1 [82].

Alterations in the concentrations of apolipoprotein M (ApoM)

As shown in previous studies, apolipoprotein M (ApoM) is a novel apolipoprotein discovered in recent years which has been shown to combine to the HDL particles. Furthermore, ApoM is also confirmed to be predominantly expressed by the hepatocytes and renal cells [83]. In addition, using the LDL receptor (LDLR)-deficient mice with over-expression of ApoM, the authors found that the risk of cardio-metabolic disorder diseases decreased obviously [83]. On the other hand, the sphingosine-1-phosphate (S1P), which is a biologically active lipid that could combine with ApoM and has the functions on modulating the functions of endothelial cells,

has been shown to account for the protective functions of ApoM and have the anti-atherogenic functions [83]. Another important research which used the stimuli administration to induce pro-inflammatory progression in mice demonstrated that under the status of inflammatory, the mice presented reduced expression of *APOM* gene in the hepatocytes and renal cells, leading to a significantly reduced plasma ApoM concentrations [84]. Consistent with this notion, the participants who suffered from the bacterial infections or HIV displayed significantly reduced circulating concentrations of ApoM [84]. Besides, it is also found that the participants with sepsis, psoriasis, and SLE had a reduction in serum concentrations of ApoM. Furthermore, this alteration is confirmed to be strongly associated with the development of pro-inflammatory diseases. The serum ApoM concentrations are reduced in patients with [85].

Alterations in the concentrations of endothelial cell lipase (ECL)

ECL is one of the most vital members of the TG-catabolic lipase family which preferentially hydrolyzes phospholipids. In mammals, ECL is not only synthesized and secreted by the endothelial cells but also by the livers, smooth muscle cells, and macrophages [86]. It is firmly shown that over-expression of ECL significantly decreases serum HDL-C concentrations; however, in contrast, via inhibiting the activity of ECL could significantly elevate the serum concentrations HDL-C, suggesting that alterations of ECL might also influence the development of dyslipidemia and its related cardio-metabolic disorder diseases [86].

Multiple pro-inflammatory cytokines, including TNF- α , IL-1 β , and IL-6, have been confirmed to enhance the gene expression of ECL within endothelial cells. Using endotoxin intervention, it has been shown that the expression of ECL increased significantly within macrophages [87]. Other two independent research has also revealed that the ECL concentrations are strongly positively correlated with the serum concentrations of the bio-markers of inflammatory response, including CRP and IL-6 [88]. Furthermore, injection of endotoxin to the human volunteers could enhance the serum ECL concentrations which were hand in hand with the decreased concentrations HDL-C temporally [88].

Alterations in the concentrations of serum amyloid A (SAA)

SAA is an important protein within circulation which could be synthesized and secreted by several diverse tissues, such as the macrophages. However, the SAA is predominantly synthesized and secreted by the hepatocytes that could account for the majority of SAA within circulation [89]. The functions of SAA on modulating serum lipid

profiles have been given substantial attention in recent years. Under the status of inflammatory status, the serum concentrations of SAA has been shown to increase approximately 1,000 folds compared to those within healthy participants [89]. In addition, the increased serum concentrations of SAA could be found under the status of multiple inflammatory relate diseases including RA, IBD, and SLE.

In details, according to the previous studies, SAA could combine with HDL particles and could subsequently displace the ApoA1 protein within HDL particle, becoming the major intra-molecular protein which is closely correlated within HDL and influencing the biological functions of HDL [89]. Notably, SAA has been shown to displace several other intra-molecular proteins within HDL particles, such as PON1. Nevertheless, the biological effect of SAA in modulating the risk and the development of cardio-metabolic disorder diseases still remains uncertain and controversial [89].

The abnormal HDL which contains SAA presents a significantly reduced ability to facilitate the cholesterol efflux from macrophages [90]. Since the cholesterol efflux process mediated by macrophages is of great significance in the progression of atherosclerosis, it could be reasonably speculated that SAA could modulate the risk of coronary heart diseases via, at least partly, modulating the functions of macrophages. Currently, the functions of SAA on influencing the pathogenesis of coronary heart diseases has been reported by several research. In the mice with *APOE* gene-deficient mice, knockout of *SAA* gene presented no significant functions on the progression of atherosclerotic lesions and the severity of disease [91]. Consistently, in the mice with *LDLR*-deficiency, knockout of *SAA* gene lead to an elevation in atherosclerotic lesions but only seen in the ascending aorta. Due to the modulatory role of SAA on macrophage, we could infer from the research that alteration of *SAA* expression might lead to the development of atherosclerosis via influence the function of macrophages [92].

On the other hand, over-expression of *SAA* gene in *APOE* gene deficient mice has been demonstrated to induce an elevation in the atherosclerotic lesions [93]. Likewise, injecting adenovirus vector to overexpress *SAA* gene enhanced the development of atherosclerosis in *APOE* gene-deficient mice [94]. Intriguingly, it has been reported that a moderate increase of SAA could be sufficient to promote the development of atherosclerosis. By contrast, a prolonged increase of SAA were not required to facilitate the atherosclerotic progression, indicating that the acute episodes of inflammatory response which raise SAA concentrations might be pro-atherogenic functions [78]. Given the discordance of published results, the clearly additional research on the function of SAA on modulating atherosclerotic related cardiovascular diseases are still required to elucidate the important role of SAA in regulating the development of atherosclerosis.

Potential mechanism of the inflammatory response induced alterations of functions in HDL particles

Currently, increasing evidence has been put forward to elucidate the potential mechanism whereby the inflammatory response reduces the circulating HDL-C concentrations and the ApoA1 concentrations has not yet been well-established. However, it is possible that several potential pathways has been shown to be involved. For instance, it has been demonstrated that the expression levels of ApoA1 within the hepatocyte is reduced significantly by pro-inflammatory activation, including injection of endotoxin or the pro-inflammatory cytokines, including TNF- α , IL-1 β , and IL-12 [95]. Notably, the reduced circulating ApoA1 contents within circulation could contribute to the reduced in circulating contents of HDL-C which has been featured by inflammatory process. As mentioned above, excessive SAA within circulation significantly up-regulated under the inflammatory condition and displace ApoA1 within the HDL particles [89]. Via this method, the SAA-contained HDL particle has been confirmed to be eliminated from the circulation which might further reduce the serum levels of HDL-C, indicating that SAA modulated the risk of dyslipidemia and the related cardio-metabolic disorder diseases [89].

On the other hand, an independent research showed that ECL could induce a reduction of serum levels of HDL-C. Since the inflammatory response could up-regulate the expression levels of ECL, we could make a reasonable speculation that the inflammatory condition might facilitate the cardio-metabolic disorder disease via, at least partly, modulating the expression of ECL [86]. Consistently, the researcher also found that co-overexpressed SAA and ECL induced a relatively larger reduction in serum concentrations of HDL-C [96]. As a consequence, up-regulated ECL activity might further promote the reduction of circulating levels of HDL-C which occurs under the inflammatory response.

On the other hand, under the inflammatory, mice with over-expressed the PLA2 presented the ability of eliminating serum HDL particles more rapidly compared with that in healthy control mice, which resultantly induced a reduction of serum HDL-C concentrations [97]. Similarly, several research has demonstrated that PLA2 could cleave ApoA1 into multiple diverse protein fragment, which could be considered as an potential mechanism whereby PLA2 influenced the serum quantity and function of HDL [98].

More recently, as mentioned above, the serum concentrations and the activity of LCAT is shown to reduce under the inflammatory response. Finally, HDL particles which

contain abundant TG have been confirmed to be easily hydrolyzed by HL which could further promote the formation of smaller HDL particles and induce the clearance of ApoA1 [99]. However, in mouse models, the serum levels of HDL-C have been shown to reduce prior to the alterations of HDL particles, implying that there are still some other potential mechanisms which played an important role in reducing HDL-C under the inflammatory response [71]. Taken together, it is still necessary to conduct several large-scale clinical trials and animal experiment which could further elucidate the potential mechanism of the inflammatory response induced alterations of functions in HDL particles.

Conclusions and perspectives

By analyzing the results from previous studies, the serum HDL concentrations reduced significantly under the inflammatory response. Within the circulation, there are obvious remarkable alterations in HDL-associated proteins, including SAA, ApoA1, PON1, and CETP. Nevertheless, the accurate mechanism whereby inflammatory response reduced the serum HDL-C concentrations is still not elucidated. The alterations in HDL particle mediated by inflammatory response decrease the function of HDL particle in RCT and prevent the oxidation of LDL particles.

According to the findings mentioned above, it has been clearly indicated that inflammatory response could influence the structure and function of HDL particles. We could also make a reasonable speculation that these alterations were advantageous under the status of inflammatory response and acute/chronic impairments. For instance, under the condition of inflammatory status, the impaired process of RCT or elevated extent of LDL oxidation could up-regulate the serum cholesterol concentrations within macrophages where it could activate the host repair. Nevertheless, these alterations could increase the development of atherosclerosis over the long terms.

Author contributions XS and BW contributed to the study design; XS, and YC wrote the manuscript. All authors reviewed drafts and approved the final version of the manuscript.

Funding This work was supported by grants from the National Key Research and Development Program of China (No. 2016YFC1301202).

Declarations

Conflict of interests The authors have no other competing interests or conflict of interest to declare.

Ethical approval This article does not contain any studies with human participants performed by any of the authors.

References

1. Feingold KR, Grunfeld C (2000) The effect of inflammation and infection on lipids and lipoproteins. In: Feingold KR, Anawalt B, Boyce A, Chrousos G, de Herder WW, Dhatariya K, Dungan K, Grossman A, Hershman JM, Hofland J, Kalra S, Kaltsas G, Koch C, Kopp P, Korbonits M, Kovacs CS, Kuohung W, Laferrere B, McGee EA, McLachlan R, Morley JE, New M, Purnell J, Sahay R, Singer F, Stratakis CA, Trencle DL, Wilson DP (eds) *Endotext*, South Dartmouth
2. Avina-Zubieta JA, Choi HK, Sadatsafavi M, Etminan M, Esdaile JM, Laccaille D (2008) Risk of cardiovascular mortality in patients with rheumatoid arthritis: a meta-analysis of observational studies. *Arthritis Rheum* 59(12):1690–1697
3. Yurkovich M, Vostretsova K, Chen W, Avina-Zubieta JA (2014) Overall and cause-specific mortality in patients with systemic lupus erythematosus: a meta-analysis of observational studies. *Arthritis Care Res (Hoboken)* 66(4):608–616
4. Samarasekera EJ, Neilson JM, Warren RB, Parnham J, Smith CH (2013) Incidence of cardiovascular disease in individuals with psoriasis: a systematic review and meta-analysis. *J Invest Dermatol* 133(10):2340–2346
5. Liu T, Shi N, Zhang S, Silverman GJ, Duan XW, Zhang S, Niu H (2020) Systemic lupus erythematosus aggravates atherosclerosis by promoting IgG deposition and inflammatory cell imbalance. *Lupus* 29(3):273–282
6. Wu GC, Liu HR, Leng RX, Li XP, Li XM, Pan HF, Ye DQ (2016) Subclinical atherosclerosis in patients with systemic lupus erythematosus: a systematic review and meta-analysis. *Autoimmun Rev* 15(1):22–37
7. Yiu KH, Yeung CK, Zhao CT, Chan JC, Siu CW, Tam S, Wong CS, Yan GH, Yue WS, Khong PL, Chan HH, Tse HF (2013) Prevalence and extent of subclinical atherosclerosis in patients with psoriasis. *J Intern Med* 273(3):273–282
8. Ali M, Girgis S, Hassan A, Rudick S, Becker RC (2018) Inflammation and coronary artery disease: from pathophysiology to Canakinumab Anti-Inflammatory Thrombosis Outcomes Study (CANTOS). *Coron Artery Dis* 29(5):429–437
9. Gaudino M, Crea F (2019) Inflammation in coronary artery disease: which biomarker and which treatment? *Eur J Prev Cardiol* 26(8):869–871
10. Feillet-Coudray C, Fouret G, Vigor C, Bonafos B, Jover B, Blachnio-Zabielska A, Rieusset J, Casas F, Gaillet S, Landrier JF, Durand T, Coudray C (2019) Long-term measures of dyslipidemia, inflammation, and oxidative stress in rats fed a high-fat/high-fructose diet. *Lipids* 54(1):81–97
11. Robertson J, Peters MJ, McInnes IB, Sattar N (2013) Changes in lipid levels with inflammation and therapy in RA: a maturing paradigm. *Nat Rev Rheumatol* 9(9):513–523
12. Chung CP, Oeser A, Raggi P, Sokka T, Pincus T, Solus JF, Linton MF, Fazio S, Stein CM (2010) Lipoprotein subclasses determined by nuclear magnetic resonance spectroscopy and coronary atherosclerosis in patients with rheumatoid arthritis. *J Rheumatol* 37(8):1633–1638
13. Khovidhunkit W, Kim MS, Memon RA, Shigenaga JK, Moser AH, Feingold KR, Grunfeld C (2004) Effects of infection and inflammation on lipid and lipoprotein metabolism: mechanisms and consequences to the host. *J Lipid Res* 45(7):1169–1196
14. Njoroge A, Guthrie BL, Bosire R, Wener M, Kiarie J, Farquhar C (2017) Low HDL-cholesterol among HIV-1 infected and HIV-1 uninfected individuals in Nairobi, Kenya. *Lipids Health Dis* 16(1):110
15. Grunfeld C, Pang M, Doerrler W, Shigenaga JK, Jensen P, Feingold KR (1992) Lipids, lipoproteins, triglyceride clearance, and cytokines in human immunodeficiency virus infection and the

- acquired immunodeficiency syndrome. *J Clin Endocrinol Metab* 74(5):1045–1052
16. Silva BSA, Lira FS, Rossi FE, Ramos D, Uzeloto JS, Freire A, de Lima FF, Gobbo LA, Ramos EMC (2018) Inflammatory and metabolic responses to different resistance training on chronic obstructive pulmonary disease: a randomized control trial. *Front Physiol* 9:262
 17. Pirillo A, Catapano AL, Norata GD (2015) HDL in infectious diseases and sepsis. *Handb Exp Pharmacol* 224:483–508
 18. Fan Y, Chen J, Liu D, Li W, Wang H, Huang Y, Gao C (2020) HDL-S1P protects endothelial function and reduces lung injury during sepsis in vivo and in vitro. *Int J Biochem Cell Biol* 126:105819
 19. Tanaka S, Couret D, Tran-Dinh A, Duranteau J, Montravers P, Schwendeman A, Meilhac O (2020) High-density lipoproteins during sepsis: from bench to bedside. *Crit Care* 24(1):134
 20. Annema W, Nijstad N, Tolle M, de Boer JF, Buijs RV, Heeringa P, van der Giet M, Tietge UJ (2010) Myeloperoxidase and serum amyloid A contribute to impaired in vivo reverse cholesterol transport during the acute phase response but not group IIA secretory phospholipase A(2). *J Lipid Res* 51(4):743–754
 21. Delporte C, Van Antwerpen P, Vanhamme L, Roumeguere T, Zouaoui Boudjeltia K (2013) Low-density lipoprotein modified by myeloperoxidase in inflammatory pathways and clinical studies. *Mediators Inflamm* 971579
 22. El Samad G, Bazzi S, Karam M, Boudjeltia KZ, Vanhamme L, Daher J (2019) Effect of myeloperoxidase modified LDL on bovine and human aortic endothelial cells. *Exp Ther Med* 18(6):4567–4574
 23. Zhang Y, Zanotti I, Reilly MP, Glick JM, Rothblat GH, Rader DJ (2003) Overexpression of apolipoprotein A-I promotes reverse transport of cholesterol from macrophages to feces in vivo. *Circulation* 108(6):661–663
 24. Malik P, Berisha SZ, Santore J, Agatista-Boyle C, Brubaker G, Smith JD (2011) Zymosan-mediated inflammation impairs in vivo reverse cholesterol transport. *J Lipid Res* 52(5):951–957
 25. Millar CL, Duclos Q, Blesso CN (2017) Effects of dietary flavonoids on reverse cholesterol transport, HDL metabolism, and HDL function. *Adv Nutr* 8(2):226–239
 26. Tang SL, Zhao ZW, Liu SM, Wang G, Yu XH, Zou J, Wang SQ, Dai XY, Fu MG, Zheng XL, Zhang DW, Fu H, Tang CK (2019) Pregnancy-associated plasma protein-A accelerates atherosclerosis by regulating reverse cholesterol transport and inflammation. *Circ J* 83(3):515–523
 27. de Beer MC, Wroblewski JM, Noffsinger VP, Ji A, Meyer JM, van der Westhuyzen DR, de Beer FC, Webb NR (2013) The impairment of macrophage-to-feces reverse cholesterol transport during inflammation does not depend on serum amyloid A. *J Lipids* 283486
 28. Hewing B, Parathath S, Barrett T, Chung WK, Astudillo YM, Hamada T, Ramkhalawon B, Tallant TC, Yusufshaq MS, Didonato JA, Huang Y, Buffa J, Berisha SZ, Smith JD, Hazen SL, Fisher EA (2014) Effects of native and myeloperoxidase-modified apolipoprotein a-I on reverse cholesterol transport and atherosclerosis in mice. *Arterioscler Thromb Vasc Biol* 34(4):779–789
 29. Norum KR, Remaley AT, Miettinen HE, Strom EH, Balbo BEP, Sampaio C, Wiig I, Kuivenhoven JA, Calabresi L, Tesmer JJ, Zhou M, Ng DS, Skeie B, Karathanasis SK, Manthei KA, Retterstol K (2020) Lecithin:cholesterol acyltransferase: symposium on 50 years of biomedical research from its discovery to latest findings. *J Lipid Res* 61(8):1142–1149
 30. Petropoulou PI, Berbee JF, Theodoropoulos V, Hatziri A, Stamou P, Karavia EA, Spyridonidis A, Karagiannides I, Kypreos KE (2015) Lack of LCAT reduces the LPS-neutralizing capacity of HDL and enhances LPS-induced inflammation in mice. *Biochim Biophys Acta* 1852:2106–2115 (10 Pt A)
 31. Tall AR (2018) Plasma high density lipoproteins: therapeutic targeting and links to atherogenic inflammation. *Atherosclerosis* 276:39–43
 32. Trinder M, Boyd JH, Brunham LR (2019) Molecular regulation of plasma lipid levels during systemic inflammation and sepsis. *Curr Opin Lipidol* 30(2):108–116
 33. Andres-Blasco I, Herrero-Cervera A, Vinue A, Martinez-Hervas S, Piqueras L, Sanz MJ, Burks DJ, Gonzalez-Navarro H (2015) Hepatic lipase deficiency produces glucose intolerance, inflammation and hepatic steatosis. *J Endocrinol* 227(3):179–191
 34. Liu D, Ding Z, Wu M, Xu W, Qian M, Du Q, Zhang L, Cui Y, Zheng J, Chang H, Huang C, Lin D, Wang Y (2017) The apolipoprotein A-I mimetic peptide, D-4F, alleviates ox-LDL-induced oxidative stress and promotes endothelial repair through the eNOS/HO-1 pathway. *J Mol Cell Cardiol* 105:77–88
 35. Chistiakov DA, Melnichenko AA, Orekhov AN, Bobryshev YV (2017) Paraoxonase and atherosclerosis-related cardiovascular diseases. *Biochimie* 132:19–27
 36. Lioudaki S, Verikokos C, Kouraklis G, Ioannou C, Chatziioannou E, Perrea D, Klonaris C (2019) Paraoxonase-1: characteristics and role in atherosclerosis and carotid artery disease. *Curr Vasc Pharmacol* 17(2):141–146
 37. Villena C, Vivas JM, Villar AM (1999) Ocular inflammation models by topical application: croton-oil induced uveitis. *Curr Eye Res* 18(1):3–9
 38. Villena C, Vivas JM, Villar AM (2000) Suppression of croton oil-induced rabbit corneal edema by sideritis javalambrensis. *J Ethnopharmacol* 71(1–2):301–305
 39. Rosenson RS, Brewer HB Jr, Ansell BJ, Barter P, Chapman MJ, Heinecke JW, Kontush A, Tall AR, Webb NR (2016) Dysfunctional HDL and atherosclerotic cardiovascular disease. *Nat Rev Cardiol* 13(1):48–60
 40. Hahn BH, Grossman J, Chen W, McMahon M (2007) The pathogenesis of atherosclerosis in autoimmune rheumatic diseases: roles of inflammation and dyslipidemia. *J Autoimmun* 28(2–3):69–75
 41. Vivekanandan-Giri A, Slocum JL, Byun J, Tang C, Sands RL, Gillespie BW, Heinecke JW, Saran R, Kaplan MJ, Pennathur S (2013) High density lipoprotein is targeted for oxidation by myeloperoxidase in rheumatoid arthritis. *Ann Rheum Dis* 72(10):1725–1731
 42. Gomez Rosso L, Lhomme M, Merono T, Sorroche P, Catoggio L, Soriano E, Saucedo C, Malah V, Dauteuille C, Boero L, Lesnik P, Robillard P, John Chapman M, Brites F, Kontush A (2014) Altered lipidome and antioxidative activity of small, dense HDL in normolipidemic rheumatoid arthritis: relevance of inflammation. *Atherosclerosis* 237(2):652–660
 43. Popa C, van Tits LJ, Barrera P, Lemmers HL, van den Hoogen FH, van Riel PL, Radstake TR, Netea MG, Roest M, Stalenhoef AF (2009) Anti-inflammatory therapy with tumour necrosis factor alpha inhibitors improves high-density lipoprotein cholesterol antioxidative capacity in rheumatoid arthritis patients. *Ann Rheum Dis* 68(6):868–872
 44. Bains Y, Caccavello R, Kotani K, Gugliucci A (2019) Paraoxonase 1, HDL subclasses and post surgery acute inflammation: a pilot study. *Antioxidants (Basel)* 8(6)
 45. Rolla R, De Mauri A, Valsesia A, Vidali M, Chiarinotti D, Bellomo G (2015) Lipoprotein profile, lipoprotein-associated phospholipase A2 and cardiovascular risk in hemodialysis patients. *J Nephrol* 28(6):749–755
 46. Fleck A (1989) Clinical and nutritional aspects of changes in acute-phase proteins during inflammation. *Proc Nutr Soc* 48(3):347–354
 47. Jonsdottir IH, Sjors Dahlman A (2019) Mechanisms in endocrinology, endocrine and immunological aspects of burnout: a narrative review. *Eur J Endocrinol* 180(3):R147–R158

48. Tercan H, Riksen NP, Joosten LAB, Netea MG, Bekkering S (2021) Trained immunity: long-term adaptation in innate immune responses. *Arterioscler Thromb Vasc Biol* 41(1):55–61
49. Memon RA, Staprans I, Noor M, Holleran WM, Uchida Y, Moser AH, Feingold KR, Grunfeld C (2000) Infection and inflammation induce LDL oxidation in vivo. *Arterioscler Thromb Vasc Biol* 20(6):1536–1542
50. Frostegard J, Svenungsson E, Wu R, Gunnarsson I, Lundberg IE, Klareskog L, Horkko S, Witztum JL (2005) Lipid peroxidation is enhanced in patients with systemic lupus erythematosus and is associated with arterial and renal disease manifestations. *Arthritis Rheum* 52(1):192–200
51. Hileman CO, Turner R, Funderburg NT, Semba RD, McComsey GA (2016) Changes in oxidized lipids drive the improvement in monocyte activation and vascular disease after statin therapy in HIV. *AIDS* 30(1):65–73
52. Zidar DA, Juchnowski S, Ferrari B, Clagett B, Pilch-Cooper HA, Rose S, Rodriguez B, McComsey GA, Sieg SF, Mehta NN, Lederman MM, Funderburg NT (2015) Oxidized LDL levels are increased in HIV infection and may drive monocyte activation. *J Acquir Immune Defic Syndr* 69(2):154–160
53. Borba EF, Carvalho JF, Bonfa E (2006) Mechanisms of dyslipoproteinemias in systemic lupus erythematosus. *Clin Dev Immunol* 13(2–4):203–208
54. da Cunha J, Ferreira Maselli LM, Treitinger A, Monteiro AM, Gidlund M, Maranhao RC, Spada C, Bydlowski SP (2013) Serum levels of IgG antibodies against oxidized LDL and atherogenic indices in HIV-1-infected patients treated with protease inhibitors. *Clin Chem Lab Med* 51(2):371–378
55. Pussinen PJ, Vilkkuna-Rautiainen T, Alftan G, Palosuo T, Jauhiainen M, Sundvall J, Vesanen M, Mattila K, Asikainen S (2004) Severe periodontitis enhances macrophage activation via increased serum lipopolysaccharide. *Arterioscler Thromb Vasc Biol* 24(11):2174–2180
56. Baranova I, Vishnyakova T, Bocharov A, Chen Z, Remaley AT, Stonik J, Eggerman TL, Patterson AP (2002) Lipopolysaccharide down regulates both scavenger receptor B1 and ATP binding cassette transporter A1 in RAW cells. *Infect Immun* 70(6):2995–3003
57. Maitra U, Li L (2013) Molecular mechanisms responsible for the reduced expression of cholesterol transporters from macrophages by low-dose endotoxin. *Arterioscler Thromb Vasc Biol* 33(1):24–33
58. Park Y, Pham TX, Lee J (2012) Lipopolysaccharide represses the expression of ATP-binding cassette transporter G1 and scavenger receptor class B, type I in murine macrophages. *Inflamm Res* 61(5):465–472
59. Braesch-Andersen S, Paulie S, Smedman C, Mia S, Kumagai-Braesch M (2013) ApoE production in human monocytes and its regulation by inflammatory cytokines. *PLoS One* 8(11):e79908
60. Das A, Sudhakar V, Ushio-Fukai M, Fukai T (2019) Novel interaction of antioxidant-1 with TRAF4: role in inflammatory responses in endothelial cells. *Am J Physiol Cell Physiol* 317(6):C1161–C1171
61. Khovidhunkit W, Shigenaga JK, Moser AH, Feingold KR, Grunfeld C (2001) Cholesterol efflux by acute-phase high density lipoprotein: role of lecithin: cholesterol acyltransferase. *J Lipid Res* 42(6):967–975
62. Tsun JG, Shiu SW, Wong Y, Yung S, Chan TM, Tan KC (2013) Impact of serum amyloid A on cellular cholesterol efflux to serum in type 2 diabetes mellitus. *Atherosclerosis* 231(2):405–410
63. Morin EE, Guo L, Schwendeman A, Li XA (2015) HDL in sepsis: risk factor and therapeutic approach. *Front Pharmacol* 6:244
64. Liao KP, Playford MP, Frits M, Coblyn JS, Iannaccone C, Weinblatt ME, Shadick NS, Mehta NN (2015) The association between reduction in inflammation and changes in lipoprotein levels and HDL cholesterol efflux capacity in rheumatoid arthritis. *J Am Heart Assoc* 4(2)
65. Mukhamedova N, Brichacek B, Darwish C, Popratiloff A, Svirdov D, Bukrinsky M (2016) Analysis of ABCA1 and cholesterol efflux in HIV-infected cells. *Methods Mol Biol* 1354:281–292
66. Khovidhunkit W, Moser AH, Shigenaga JK, Grunfeld C, Feingold KR (2001) Regulation of scavenger receptor class B type I in hamster liver and Hep3B cells by endotoxin and cytokines. *J Lipid Res* 42(10):1636–1644
67. Li L, Thompson PA, Kitchens RL (2008) Infection induces a positive acute phase apolipoprotein E response from a negative acute phase gene: role of hepatic LDL receptors. *J Lipid Res* 49(8):1782–1793
68. Miyazawa H, Tabeta K, Miyauchi S, Aoki-Nonaka Y, Domon H, Honda T, Nakajima T, Yamazaki K (2012) Effect of *Porphyromonas gingivalis* infection on post-transcriptional regulation of the low-density lipoprotein receptor in mice. *Lipids Health Dis* 11:121
69. Miyazawa H, Honda T, Miyauchi S, Domon H, Okui T, Nakajima T, Tabeta K, Yamazaki K (2012) Increased serum PCSK9 concentrations are associated with periodontal infection but do not correlate with LDL cholesterol concentration. *Clin Chim Acta* 413(1–2):154–159
70. M de la L Moya, McGillicuddy FC, Hinkle CC, Byrne M, Joshi MR, Nguyen V, Tabita-Martinez J, Wolfe ML, Badellino K, Pruscino L, Mehta NN, Asztalos BF, Reilly MP (2012) Inflammation modulates human HDL composition and function in vivo. *Atherosclerosis* 222(2):390–394
71. Ly H, Francone OL, Fielding CJ, Shigenaga JK, Moser AH, Grunfeld C, Feingold KR (1995) Endotoxin and TNF lead to reduced plasma LCAT activity and decreased hepatic LCAT mRNA levels in Syrian hamsters. *J Lipid Res* 36(6):1254–1263
72. Charles-Schoeman C, Watanabe J, Lee YY, Furst DE, Amjadi S, Elashoff D, Park G, McMahon M, Paulus HE, Fogelman AM, Reddy ST (2009) Abnormal function of high-density lipoprotein is associated with poor disease control and an altered protein cargo in rheumatoid arthritis. *Arthritis Rheum* 60(10):2870–2879
73. Rose H, Low H, Dewar E, Bukrinsky M, Hoy J, Dart A, Svirdov D (2013) The effect of HIV infection on atherosclerosis and lipoprotein metabolism: a one year prospective study. *Atherosclerosis* 229(1):206–211
74. Hardardottir I, Moser AH, Fuller J, Fielding C, Feingold K, Grunfeld C (1996) Endotoxin and cytokines decrease serum levels and extra hepatic protein and mRNA levels of cholesteryl ester transfer protein in syrian hamsters. *J Clin Invest* 97(11):2585–2592
75. Masucci-Magoulas L, Moulin P, Jiang XC, Richardson H, Walsh A, Breslow JL, Tall A (1995) Decreased cholesteryl ester transfer protein (CETP) mRNA and protein and increased high density lipoprotein following lipopolysaccharide administration in human CETP transgenic mice. *J Clin Invest* 95(4):1587–1594
76. Jahangiri A, de Beer MC, Noffsinger V, Tannock LR, Ramaiah C, Webb NR, van der Westhuyzen DR, de Beer FC (2009) HDL remodeling during the acute phase response. *Arterioscler Thromb Vasc Biol* 29(2):261–267
77. Ferraz-Amaro I, Gonzalez-Gay MA, Garcia-Dopico JA, Diaz-Gonzalez F (2013) Cholesteryl ester transfer protein in patients with rheumatoid arthritis. *J Rheumatol* 40(7):1040–1047
78. Mackness M, Mackness B (2015) Human paraoxonase-1 (PON1): gene structure and expression, promiscuous activities and multiple physiological roles. *Gene* 567(1):12–21
79. Van Lenten BJ, Hama SY, de Beer FC, Stafforini DM, McIntyre TM, Prescott SM, La Du BN, Fogelman AM, Navab M (1995) Anti-inflammatory HDL becomes pro-inflammatory during the acute phase response. Loss of protective effect of HDL against LDL oxidation in aortic wall cell cocultures. *J Clin Invest* 96(6):2758–2767

80. Feingold KR, Memon RA, Moser AH, Grunfeld C (1998) Para-oxonase activity in the serum and hepatic mRNA levels decrease during the acute phase response. *Atherosclerosis* 139(2):307–315
81. Siegel MO, Borkowska AG, Dubrovsky L, Roth M, Welti R, Roberts AD, Parenti DM, Simon GL, Sviridov D, Simmens S, Bukrinsky M, Fitzgerald ML (2015) HIV infection induces structural and functional changes in high density lipoproteins. *Atherosclerosis* 243(1):19–29
82. Bacchetti T, Campanati A, Ferretti G, Simonetti O, Liberati G, Offidani AM (2013) Oxidative stress and psoriasis: the effect of antitumour necrosis factor-alpha inhibitor treatment. *Br J Dermatol* 168(5):984–989
83. Ren K, Tang ZL, Jiang Y, Tan YM, Yi GH, Apolipoprotein M (2015) *Clin Chim Acta* 446:21–29
84. Feingold KR, Shigenaga JK, Chui LG, Moser A, Khovidhunkit W, Grunfeld C (2008) Infection and inflammation decrease apolipoprotein M expression. *Atherosclerosis* 199(1):19–26
85. Holzer M, Wolf P, Curcic S, Birner-Gruenberger R, Weger W, Inzinger M, El-Gamal D, Wadsack C, Heinemann A, Marsche G (2012) Psoriasis alters HDL composition and cholesterol efflux capacity. *J Lipid Res* 53(8):1618–1624
86. Yasuda T, Ishida T, Rader DJ (2010) Update on the role of endothelial lipase in high-density lipoprotein metabolism, reverse cholesterol transport, and atherosclerosis. *Circ J* 74(11):2263–2270
87. Robert J, Lehner M, Frank S, Perisa D, von Eckardstein A, Rohrer L (2013) Interleukin 6 stimulates endothelial binding and transport of high-density lipoprotein through induction of endothelial lipase. *Arterioscler Thromb Vasc Biol* 33(12):2699–2706
88. Badellino KO, Wolfe ML, Reilly MP, Rader DJ (2008) Endothelial lipase is increased in vivo by inflammation in humans. *Circulation* 117(5):678–685
89. Prufer N, Kleuser B, van der Giet M (2015) The role of serum amyloid A and sphingosine-1-phosphate on high-density lipoprotein functionality. *Biol Chem* 396(6–7):573–583
90. Vaisar T, Tang C, Babenko I, Hutchins P, Wimberger J, Suffredini AF, Heinecke JW (2015) Inflammatory remodeling of the HDL proteome impairs cholesterol efflux capacity. *J Lipid Res* 56(8):1519–1530
91. De Beer MC, Wroblewski JM, Noffsinger VP, Rateri DL, Howatt DA, Balakrishnan A, Ji A, Shridas P, Thompson JC, van der Westhuyzen DR, Tannock LR, Daugherty A, Webb NR, De Beer FC (2014) Deficiency of endogenous acute phase serum amyloid A does not affect atherosclerotic lesions in apolipoprotein E-deficient mice. *Arterioscler Thromb Vasc Biol* 34(2):255–261
92. Krishack PA, Bhanvadia CV, Lukens J, Sontag TJ, De Beer MC, Getz GS, Reardon CA (2015) Serum amyloid A facilitates early lesion development in Ldlr^{-/-} mice. *J Am Heart Assoc* 4(7)
93. Ahlin S, Olsson M, Wilhelmson AS, Skalen K, Boren J, Carlsson LM, Svensson PA, Sjöholm K (2014) Adipose tissue-derived human serum amyloid a does not affect atherosclerotic lesion area in hSAA1^{+/-}/ApoE^{-/-} mice. *PLoS ONE* 9(4):e95468
94. Thompson JC, Jayne C, Thompson J, Wilson PG, Yoder MH, Webb N, Tannock LR (2015) A brief elevation of serum amyloid A is sufficient to increase atherosclerosis. *J Lipid Res* 56(2):286–293
95. Parseghian S, Onstead-Haas LM, Wong NC, Mooradian AD, Haas MJ (2014) Inhibition of apolipoprotein A-I expression by TNF-alpha in HepG2 cells: requirement for c-jun. *J Cell Biochem* 115(2):253–260
96. Wroblewski JM, Jahangiri A, Ji A, de Beer FC, van der Westhuyzen DR, Webb NR (2011) Nascent HDL formation by hepatocytes is reduced by the concerted action of serum amyloid A and endothelial lipase. *J Lipid Res* 52(12):2255–2261
97. Tietge UJ, Maugeais C, Lund-Katz S, Grass D, deBeer FC, Rader DJ (2002) Human secretory phospholipase A2 mediates decreased plasma levels of HDL cholesterol and apoA-I in response to inflammation in human apoA-I transgenic mice. *Arterioscler Thromb Vasc Biol* 22(7):1213–1218
98. Cavigliolo G, Jayaraman S (2014) Proteolysis of apolipoprotein A-I by secretory phospholipase A(2): a new link between inflammation and atherosclerosis. *J Biol Chem* 289(14):10011–10023
99. Lamarche B, Uffelman KD, Carpentier A, Cohn JS, Steiner G, Barrett PH, Lewis GF (1999) Triglyceride enrichment of HDL enhances in vivo metabolic clearance of HDL apo A-I in healthy men. *J Clin Invest* 103(8):1191–1199

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.