ORIGINAL ARTICLE



TRAILR1 (rs20576) and *GRIA3* (rs12557782) are not associated with interferon- β response in multiple sclerosis patients

Parham Jazireian¹ · Soheila Talesh Sasani² · Farhad Assarzadegan³ · Mojtaba Azimian⁴

Received: 25 September 2020 / Accepted: 19 November 2020 / Published online: 2 December 2020 © Springer Nature B.V. 2020

Abstract

Multiple sclerosis (MS) is an autoimmune-type inflammatory disorder in human central nervous system. Recombinant interferon beta (IFN- β) decreases the number of relapses and postpones disability progression in MS. However, up to 50% of patients treated with interferon beta continue experiencing relapses and/or worsening disability. Single nucleotide polymorphisms in different genes have been known to show significant associations with response to IFN- β in MS patients. In the present work, we examined the potential role of *TRAILR1* and *GRIA3* genes polymorphisms on response to IFN- β therapy in Iranian MS patients. The DNA was extracted from blood samples by standard procedures from 73 patients diagnosed with Multiple Sclerosis that were either responded to IFN- β or did not. We carried out RFLP -PCR and tetra-primer ARMS-PCR methods to study of *rs20576* and *rs12557782*, respectively. All results were analyzed using the SPSS software. *TRAILR1 rs20576* genotype frequencies in responders and non-responders were similar ($\chi^2 = 0.26$, P = 0.87, Fisher's Exact test). Our results showed that response to IFN- β has not association with sex (p = 0.73). Also, genotypic frequencies of *GRIA3 rs12557782* had no significant differences between two groups of female population ($\chi^2 = 3.75$, p = 0.15). Furthermore, it had not been any statistical differences between responder and non-responder males ($\chi^2 = 0.7$, p = 0.4) related to the SNP. Our results analysis revealed no significant association between the studied SNPs (*TRAILR1 rs20576* and *GRIA3rs 12,557,782*) and response to IFN- β in Iranian MS patients.

Keywords Pharmacogenetics \cdot Multiple sclerosis \cdot Gene polymorphisms \cdot Interferon- β

Introduction

Multiple sclerosis (MS) is an autoimmune, demyelinating neurodegenerative disorder which affects central nervous system (CNS) and it is widely assumed to be responsible for non-traumatic disability in youngsters. Like most autoimmune diseases, MS occurs three times more common in women than men [1, 2]. MS can cause demyelination, oligodendrocyte destruction and eventually axonal injury. The

Soheila Talesh Sasani sasani@guilan.ac.ir

- ¹ Department of Biology, University Campus 2, University of Guilan, Rasht, Iran
- ² Department of Biology, Faculty of Sciences, University of Guilan, Rasht, Iran
- ³ Department of Neurology, Imam Hossein Hospital, Shahid Beheshti University of Medical Sciences, Tehran, Iran
- ⁴ Rofeydeh Rehabilitation Center, University of Social Welfare and Rehabilitation Sciences, Tehran, Iran

exact etiology of MS is still unknown; however, based on the recent researches and evidences it is clear both genetic and environmental factors are involved in the disease. Human leukocyte antigen- (HLA-) *DRB1*1501* has the strongest association with MS susceptibility [3, 4]. Various environmental factors have been identified to be associated with MS; these factors include vitamin D level, usage of tobacco, geographical latitude and exposure to infectious organisms such as Epstein-Bar virus [5–8].

Multiple sclerosis is classified into four types due to the disease progression process. Relapsing-remitting MS(RRMS) that is characterized by clearly defined attacks (relapses) which is followed by partial or complete recovery (remission). Secondary progressive MS (SPMS), relapsingremitting MS often proceeds to secondary progressive MS and SPMS patients continuously experience the symptoms. In third kind, primary progressive MS patients show the disease' s symptoms which slowly worsen without any remission periods. Fourth type of MS contains relapsing progressive MS which the symptoms gradually worsen with possibility of one or more relapses [9]. No therapies have been found to reverse neurodegenerative process of MS however within the last two decades, disease-modifying therapies such as interferon beta, glatiramer acetate, mitoxantrone, natalizumab have been able to reduce the amount of relapses and reduce the progression of the disease [10]. Clinical trials have revealed that interferon beta can prevent the progression of the disease and reduce the number of relapses in all relapsing forms of MS when compared with placebo [11–13]. The precise molecular mechanism for preventing of the disease progression is still unknown but it is assumed that it inhibits the activation and proliferation of T cells and also decreases the production of proinflammatory cytokines through activating Jak-Stat signaling pathway [14]. However, up to 50% of MS patients do not respond to this drug properly [15, 16].

Some genetic variations may be associated with response to interferon beta and these variations identifying may help physicians to proper choice of treatment and also prevention of drug's adverse effects on patients who do not respond to the drug. The recent genome wide association study (GWAS) on interferon beta pharmacogenetics has demonstrated polymorphisms in multiple genes which might be associated with response to treatment with interferon beta in RRMS patients [17]. TRAIL is a tumor necrosis factor which induces apoptosis in various cells. All TRAIL receptors are expressed in oligodendrocytes, neurons and astrocytes [18]. TRAIL and TRAIL-R1 genes are related to Multiple Sclerosis. While treating with IFN-B, the enhancement of TRAIL expression might both induce apoptosis in granulocytes or monocytes and also it could perish or inhibit the activation of auto-reactive T cells, or even promote the production of regulatory T cells. Several studies have shown that single nucleotide polymorphisms in the genes are effective in the response to IFN- β therapy. *TRAILR-1* single nucleotide polymorphism rs20576 is in the exon 5 of the gene. The rs20576-C allele causes substitution of pGlu228Ala adjacent to the membrane anchoring and it might affect TRAIL binding and the apoptotic signaling. The SNP, rs20576-C, has been associated with susceptibility to various cancer types [19-22] as well as with an enhanced risk of metastasis [23]. Rs20576-C allele has also been studied in different diseases, it has been demonstrated that this polymorphism is associated with mantle cell lymphoma and chronic lymphocytic leukemia [21]. GRIA3 encodes an AMPA-type glutamate receptor, an excitatory neurotransmitter that takes part in learning, memory, neural formation, habit formation and sensory gating [24]. Excessive activation of this receptor may cause central neurons degeneration to die. Glutamate receptors play a pivotal role in the central nervous system and are associated with various brain disorders. They play a crucial role in the pathogenesis of different neurodegenerative diseases (NDDs) such as Parkinson's disease, Alzheimer's disease, and amyotrophic lateral sclerosis [25]. A positive association was also found between GRIA3 with schizophrenia, bipolar disorder and it has been suggested that GRIA3 might play a crucial role in the development of the aforementioned diseases [26]. It has also been reported to be associated with mental retardation and sexual dysfunction in major depression [27, 28]. Moreover, a variant of GRIA3 gene, rs3761555, has been found to be associated with migraine phenotype in Italian population [29]. The SNP rs12557782 resides on the x chromosome, in the second intron of GRIA3 gene and has been selected as a potential candidate for determining response to treatment with IFN- β in RRMS patients by the GWAS study [17]. In the current research we have studied the association of two candidate genes polymorphisms, TRAILR1rs20576 A > C and GRIA3rs12557782 G > A, with response to interferon beta therapy in Iranian patients.

Materials and methods

Subjects

In this study, 73 relapsing – remitting MS patients including 19 men and 54 women met McDonald's criteria and were treated with interferon beta [9]. Written informed consent was gathered from the patients and the project had been approved by local ethics committee.

The 37 patients who were classified as responders to the treatment did not have increase in relapses during the first 2 years of treatment's initiation. The 36 patients who were categorized as non-responders to the treatment had experienced at least one or more relapses during the first 2 years of treatment usage [30]. The patients were not related by blood.

Genotyping

Peripheral blood samples were collected in tubes contained EDTA and DNA was extracted from blood nucleated cells using Rapid Genomic DNA Isolation kit (Gen Fanavaran, Iran) according to manufacturer's guideline. The purity and amount of isolated DNA was checked by agarose gel electrophoresis (1%) and a Nanodrop spectrophotometer (Thermo Scientific, USA), respectively.

Primers and PCR Conditions

The specific primers were designed for the studied genes using Primer3 online software v4.1.0 and were analyzed by NCBI Primer- BLAST tool (http://www.ncbi.nlm.nih. gov/). The *TRAILR1*rs20576 primers were as follow: 5'-GTGTGCATGCTCAGACCCTT -3' and 5'- CGGAAC AACCAAAGTCACAAC -3'. Restriction fragment length polymorphism- polymerase chain reaction (RFLP-PCR) method was done. PCR reaction was carried out in 25 uL mixture, containing 30 ng of template DNA, 1 µl of each 10 pmol/µl primer (Sinaclone, Iran), 5 µl of 2x Master mix red (Ampliqon, Denmark) and 17 µl dH₂O. PCR amplification was conducted by an automated thermocycler (TC512 TECHNE, United Kingdom). The following conditions were used: 94 °C for the first denaturation at 5 min, 35 cycles of 30s for second denaturation at 94 °C, 30s for primers annealing at 60 °C and 30s DNA extension at 72 °C, respectively, ending with 7 min at 72 °C to complete extension. The PCR product was electrophoresed on 2% agarose gel and a 390 bp replicon was detected. The PCR products digestion was carried out by Taq1(Sinaclone, Iran) restriction enzyme and two DNA fragments, 239 bp and 151 bp, were obtained related to 683AA genotype for TRAILR1. However, the enzyme did not digest the amplified fragment obtained from 683CC genotype and after digestion reaction a 390 bp DNA segment was detected for this variant (Fig. 1).

To clarify the relationship of the GRIA3 rs12557782 and IFN-β resistance, DNA samples from both responder and non-responder groups were amplified by tetra ARMS- PCR method using two pairs of specific primers to amplification of different alleles. The outer primers for GRIA3 rs12557782 were: 5'-AGAACGTGATAATCACACATTGGTATGATT -3' and 5'- TACTTGAATGTAACTGCTAGCAAATGCA -3' as forward and reverse, respectively. Additionally, 5'-TGCTTAACCATATTATCTCGATATAAACGG -3' and 5'- TGGGCTAAGAGGGTGCTTTTTAGTCT -3' were used as inner forward and reverse primers, respectively. Polymerase chain reaction was performed in 25 µL volume mixture containing 30 ng of template DNA, 1 µl of each 10 pmol/ul primer (Sinaclone, Iran), 5 µl of 2x Master mix blue (Ampliqon, Denmark) and 17 µl dH₂O. The amplification reaction was done as follows: 5 min at 94 °C for

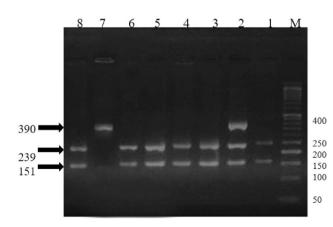


Fig. 1 RFLP-PCR genotyping of *TRAILR1rs20576* A>C by *Taq1* digestion Lanes: M, DNA ladder; 1, 3, 4, 5, 6 and 8 show AA; 2 indicates AC; 7 represents CC genotypes

first denaturation, 35 cycles of 40 s at 94 °C, 58 °C and 72 °C, respectively, ending with 10 min at 72 °C to complete extension. 5 μ L of the PCR product was electrophoresed on an agarose gel (2%). The SNP, *GRIA3 rs12557782*, was identified based on the PCR products lengths. Outer primers amplified a 410 nucleotide amplicon containing *GRIA3 rs12557782*, while inner primers led to amplification of different replicons based on the allele types. For *rs12557782-G* allele a 194 bp length fragment was amplified while a 273 bp for *rs12557782-A* allele were obtained (Fig. 2).

Statistical analysis

We performed a Mann–Whitney U test, one non-parametric test, to compare number of relapses between responder and non-responder patients. Genotypic and allelic frequencies were compared between the two groups using Fishers Exact test with GraphPad Prism program (version 7.05 for Windows, GraphPad Software, La Jolla California USA, www. graphpad.com). Chi-square and *P* values were calculated. We used SPSS v.19.0 software (SPSS, Chicago, IL, USA) for data statistical analysis. For all statistics, *p* values less than 0.05 was considered as statistically significant.

Results

In this association study, we investigated the relationship of two genetic polymorphisms, *TRAILR1* rs20576 A > C and *GRIA3* rs12557782 G > A, with response to interferon beta therapy in patients diagnosed with relapsing-remitting multiple sclerosis (RRMS). The subjects included responder group and non-responders with 37(27 women and 10 men) and 36(27 women and 9 men) patients, respectively. All studied patients had been using interferon beta and follow-up

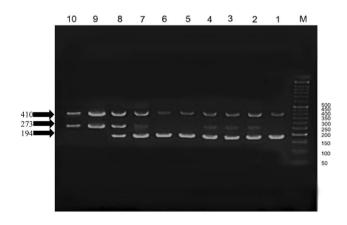


Fig. 2 Agarose gel electrophoresis pattern of *GRIA3* rs4938723 G > A genotyping by tetra ARMS-PCR. Lanes: M, DNA ladder; 1, 5 and 6, GG; 2, 3, 4, 7 and 8 represent AG; 9 and 10 indicates AA genotype

for at least 2 years during from June 22, 2017 to September 30, 2019. Some of the important clinical characteristics of the studied patients, concerned to our study, are shown in Table 1. No significant differences were seen between two groups about the age at disease onset and the age at treatment onset, p=0.09 and p=0.1, respectively. After taking the IFN- β , the number of relapses was significantly lowered in responder group than to non-responders (p=0.03). Therefore, responder subjects had fewer or no relapses in the 2 years while non-responders had more relapses within the 2 years (Table 1).

The genotypic frequencies for *TRAILR1 rs20576* were as follows: AA (67%), AC (30%) and CC (3%) in responder population while the same were 69%, 28% and 3% in nonresponders, respectively. However, the frequencies of *TRAILR1 rs20576* genotypes had no statistical differences between responder and non-responder groups ($\chi^2 = 0.26$, P = 0.87). Also, the frequencies of A and C alleles had no significant differences between two groups (P > 0.05). In the other hand, because *GRIA3* is a sex-linked gene, genotypic and allelic frequencies of *GRIA3 rs12557782* were separately assessed in both responder and non-responder patients based on patients' gender. Our results showed that response to IFN- β had not association with sex (p = 0.73). Also, genotypic frequencies had no significant differences between two groups of female population ($\chi^2 = 3.75$, p = 0.15). The

Table 1 Clinical features of the examined population

Characteristic	Responders	Non-responders
Female	72.9%	75%
Male	27.1%	25%
Age at disease onset, y, mean, SD	24.70 ± 7.6	24.44 ± 3.9
Age at treatment Onset, y, mean, SD	28 ± 8.7	26.69 ± 3.6
Relapses in 2 y prior to treatment, mean, SD	2.3 ± 1.3	3.4 ± 2.2
Relapses during 2 y of treatment, mean, SD	0.35 ± 0.58	3.3 ± 2.1

frequencies were GG (15%), AG (81%) and AA (4%) in responder females while the proportions were 26%, 70% and 4%, respectively, in non-responders. Furthermore, it had not been any statistical differences between responders and non-responder males ($\chi^2 = 0.7$, p = 0.4). The detailed distribution of genotypic frequencies of the two studied SNPs are presented in Table 2. Combination analysis was carried out to investigation of dominant and recessive genetic models for both SNPs, rs20576 A>C and *rs12557782* G>A. The combined genotypes were not associated with response to IFN- β treatment (Table 3).

Discussion

Multiple sclerosis (MS) is an autoimmune disease that leads to neurons demyelination in central nervous system (CNS) and it is suggested to be one of the causes of disability at a young age [1, 2]. The precise pathogenesis of the disease is still unknown; however, genetic factors are involved in MS disease. Relapsing-remitting MS (RRMS) is determined by defined attacks (relapses) which is followed by partial or complete recovery (remission). Clinical experiments have revealed that interferon beta can prevent the progression of the disease and reduce the number of relapses [11-13]. But half of the patients do not respond suitably [15, 16]. Some genes and genetic variations may be related to the response to interferon beta therapy. We have studied the association of two candidate gene polymorphisms, TRAILR1 rs20576 A>C and GRIA3 rs12557782 G>A, with responsiveness to interferon beta treatment in Iranian patients. According to our results, TRALR1 rs20576 and GRIA3 rs12557782 did not show any significant association with response to IFN-β treatment in RRMS patients.

It has been identified that TRAIL/TRAIL receptor system takes part in various steps in immune cell activation, migration, proliferation, differentiation and might be associated with different autoimmune disorders [31–34]. TRAIL can attach to its receptor and initiate apoptosis. In contrast to

Gene	Genotype	Responders n(%)	Non-responders n(%)	χ2	P- value
	AA	25(67)	25(69)		
TRAILR1 (rs20576)	AC	11(30)	10(28)	0.26	0.87
Female	CC	1(3)	1(3)		
	GG	4(15)	7(26)		
	AG	22(81)	19(70)	3.75	0.15
	AA	1(4)	1(4)		
GRIA3 rs12557782					
Male	G	6(60)	7(78)		
	А	4(40)	2(22)	0.69	0.40

Table 2 Genotypic frequencies
of TRAILR1 rs20576 and
GRIA3 rs12557782

Table 3Different models ofthe genotypic distributionof rs20576 and rs12557782polymorphisms

Genetic models	Genotype	Responders N(%)	Non-Respond- ers N(%)	OR (95% CI)	P Value
TRAILR1 rs20576					
Dominant	AA	25 (%67)	25 (%69)	1.00	-
	AC+CC	12 (%33)	11 (%31)	0.91(0.34-2.46)	0.86
Recessive	AA+AC	36 (%97)	35(%97)	1.00	-
	CC	1 (%3)	1 (%3)	1.02 (0.06–17.09)	0.98
GRIA3 rs12557782					
Dominant	GG	4(15)	7 (26)	1.00	_
	AG+AA	23 (85)	20 (74)	0.49 (0.12–1.94)	0.31
Recessive	AG+GG	26 (%96)	26(%96)	1.00	_
	AA	1 (4)	1 (4)	1.00 (0.05–16.85)	0.31

our results, López-Gómez et al. indicated TRALR1 rs20576 to be statistically associated with response to IFN- β in an original cohort of Spanish MS patients. They found the SNP frequency significantly differed between responders and non-responder patients in the cohort, although the results were not obtained in the validation cohort [35]. The research showed relationship between rs20576-CC genotype with response to IFN- β while we did not obtain any association. These different results may be related to restricted number of the patients in current study because we detected only one responder patient with CC-genotype. However, we could not study a validation cohort due to limited number of the subjects. The genetic variant has also been studied in other neurological diseases such as Alzheimer's disease (AD) but no association was found between the mentioned polymorphism and AD [36]. Morales-Lara and et al. indicated that TRAILR1 rs20575 affects rheumatoid arthritis (RA) and ankylosing spondylitis (AS) patient's response to infliximab treatment [37]. Additionally, Taghavi et al. conducted a case-control study in Azeri patients in Iran. In the study, frequencies of genotypes and alleles of TRAILR1 rs4872077 (T > C), and TRAILR2 rs1001793 (G > A), were assessed. The frequencies were not different between MS patients and healthy controls ($\chi^2 = 0.42$, p = 0.514 for rs4872077 and $\chi^2 = 2.3$, p = 0.127 for rs1001793). They did not obtain any association between the two polymorphisms and multiple sclerosis disease [38]. Taheri et al. (2016) evaluated TRAIL gene expression changes in relapsing- remitting MS patients who respond to IFN- β therapy. They found no significant difference between the patients and healthy controls [39].

GRIA3 encodes an AMPA-type glutamate receptor, so excessive activation of this receptor may cause central neurons degeneration and death. Our results showed that RRMS patients reflex to IFN- β treatment has no significant relationship with *GRIA3 rs12557782* in both males and females. Little research has been done on the association of the SNP with MS. Comabella et al. demonstrated different

association between *GRIA3* rs12557782 and response to IFN- β therapy in MS patients due to gender. The study showed significant relationship between the polymorphism and IFN- β therapy response in women (p < 0.001, OR 8.5) but not in men (p = 0.56, OR = 1.7) [40]. We report the minor allele frequencies (MAFs) of *TRAILR1* (rs20576) and *GRIA3* (rs12557782) in Iranian population for the first time. The MAFs were obtained 0.18 for rs20576 A > C and 0.22 for rs12557782 G > A single nucleotide polymorphisms.

Conclusion

This is the first study about association between *TRAL-R1rs20576* and *GRIA3*rs12557782 genetic variations with response to IFN- β treatment in Iranian RRMS patients. We have not found any association between the two SNPs and response to IFN- β therapy. The possible explanations for our outcomes might be due to the limited number of studied patients. Due to this limitation we could not validate our results through a validation cohort. Also, ethnic characteristics can lead to different results in association studies. Therefore, further investigations with larger number of subjects is recommended. Also, future studies on different racial populations will be necessary to get further insight into the relationship between *TRAILR1 rs20576* and *GRIA3 rs12557782* with responsiveness to interferon beta treatment.

Acknowledgments We thank all subjects who participated in this study and Iranian Multiple Sclerosis Community for their cooperation. Also, we would like to thank University of Guilan for supporting the current research.

Funding Not applicable.

Data availability The datasets generated and analysed during the current study are available from the corresponding author on reasonable request.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

Ethics approval This study was approved by Iran National Committee for Ethics in Biomedical Research (IR. GUMS. REC. 1397.08) and the study was performed in accordance with the ethical standards as laid down in the 1964 Declaration of Helsinki and its later amendments or comparable ethical standards.

Consent to participate Informed consent was obtained from all participants of the study.

Consent to publish The participant was informed that the results of this research would be published in the journal.

References

- Dilokthornsakul P, Valuck RJ, Nair KV, Corboy JR, Allen RR, Campbell JD (2016) Multiple sclerosis prevalence in the United States commercially insured population. Neurology 86(11):1014–1021
- Debouverie M, Pittion-Vouyovitch S, Louis S, Guillemin F, Group L (2008) Natural history of multiple sclerosis in a populationbased cohort. Eur J Neurol 15(9):916–921
- Risch N, Merikangas K (1996) The future of genetic studies of complex human diseases. Science 273(5281):1516–1527
- Ramagopalan SV, Knight JC, Ebers GC (2009) Multiple sclerosis and the major histocompatibility complex. Curr Opin Neurol 22(3):219–225
- 5. Pantazou V, Schluep M, Du Pasquier R (2015) Environmental factors in multiple sclerosis. Presse Med 44(4):e113–e120
- Beecham AH, Patsopoulos NA, Xifara DK, Davis MF, Kemppinen A, Cotsapas C et al (2013) Analysis of immune-related loci identifies 48 new susceptibility variants for multiple sclerosis. Nat Genet 45(11):1353–1360
- Dobson R, Giovannoni G, Ramagopalan S (2013) The month of birth effect in multiple sclerosis: systematic review, metaanalysis and effect of latitude. J Neurol Neurosurg Psychiatry 84(4):427–432
- Santiago O, Gutierrez J, Sorlozano A, Jd DL, Villegas E, Fernandez O (2010) Relation between Epstein-Barr virus and multiple sclerosis: analytic study of scientific production. Eur J Clin Microbiol Infect Dis 29(7):857–866
- McDonald WI, Compston A, Edan G, Goodkin D, Hartung H-P, Lublin FD, McFarland HF, Paty DW, Polman CH, Reingold SC, Sandberg-Wollheim M, Sibley W, Thompson A, Svd N, Weinshenker BY, Wolinsky JS (2001) Recommended diagnostic criteria for multiple sclerosis: guidelines from the international panel on the diagnosis of multiple sclerosis. Ann Neurol 50(1):121–127
- Maroney M, Hunter SF (2014) Implications for multiple sclerosis in the era of the affordable care act: a clinical overview. Am J Manag Care 20(11 Suppl):S220–S227
- Ebers GC (1998) Randomised double-blind placebo-controlled study of interferon beta-1a in relapsing/remitting multiple sclerosis. PRISMS (prevention of relapses and disability by interferon beta-1a subcutaneously in multiple sclerosis) study group. Lancet 352(9139):1498–1504
- Jacobs LD, Cookfair DL, Rudick RA, Herndon RM, Richert JR, Salazar AM et al (1996) Intramuscular interferon beta-1a for disease progression in relapsing multiple sclerosis. Ann Neurol 39(3):285–294

- Kinkel RP, Dontchev M, Kollman C, Skaramagas TT, O'Connor PW, Simon JH, Controlled High-Risk Avonex Multiple Sclerosis Prevention Study in Ongoing Neurological Surveillance Investigators (2012) Association between immediate initiation of intramuscular interferon beta-1a at the time of a clinically isolated syndrome and long-term outcomes: a 10-year follow-up of the controlled high-risk avonex multiple sclerosis prevention study in ongoing neurological surveillance. Arch Neurol 69(2):183–190
- 14. Markowitz CE (2007) Interferon-beta mechanism of action and dosing issues. Neurology 68(24 suppl 4):8–11
- Weinstock-Guttman B, Badgett D, Patrick K, Hartrich L, Santos R, Hall D, Baier M, Feichter J, Ramanathan M (2003) Genomic effects of IFN-β in multiple sclerosis patients. J Immunol 171(5):2694–2702
- 16. Taniguchi T, Takaoka A (2002) The interferon- α/β system in antiviral responses: a multimodal machinery of gene regulation by the IRF family of transcription factors. Curr Opin Immunol 14(1):111–116
- 17. Coyle PK (2017) Pharmacogenetic biomarkers to predict treatment response in multiple sclerosis : current and future perspectives. Mult Scler Int 2017:1–10
- Cannella B, Gaupp S, Omari KM, Raine CS (2007) Multiple sclerosis: death receptor expression and oligodendrocyte apoptosis in established lesions. J Neuroimmunol 188(1–2):128–137
- Frank B, Hemminki K, Shanmugam KS, Meindl A, Klaes R, Schmutzler RK, Wappen schmidth B, Untch M, Bugert P, Bartram CR, Burwinkel B (2005) Association of death receptor 4 haplotype 626C–683C with an increased breast cancer risk. Carcinogenesis 26(11):1975–1977
- Frank B, Shanmugam KS, Beckmann L, Hemminki K, Brenner H, Hoffmeister M, Chang-Claude J, Burwinkel B (2006) Death receptor 4 variants and colorectal cancer risk. Cancer Epidemiol Prev Biomarkers 15(10):2002–2005
- 21. Wolf S, Mertens D, Pscherer A, Schroeter P, Winkler D, Gröne H-J, Hofele C, Hemminki K, Kumar R, Steineck G, Dohner H, Stilgenbauer S, Lichter P (2006) Ala228 variant of trail receptor 1 affecting the ligand binding site is associated with chronic lymphocytic leukemia, mantle cell lymphoma, prostate cancer, head and neck squamous cell carcinoma and bladder cancer. Int J Cancer 118:1831–1835
- Chen B, Liu S, Wang X-L, Xu W, Li Y, Zhao WH, Wu JQ (2009) TRAIL-R1 polymorphisms and cancer susceptibility: an evidence-based meta-analysis. Eur J Cancer 45(14):2598–2605
- 23. Langsenlehner T, Langsenlehner U, Renner W, Kapp KS, Krippl P, Hofmann G, Clar H, Pummer K, Mayer R (2008) The Glu-228Ala polymorphism in the ligand binding domain of death receptor 4 is associated with increased risk for prostate cancer metastases. Prostate 68(3):264–268
- 24. Riedel G, Platt B, Micheau J (2003) Glutamate receptor function in learning and memory. Behav Brain Res 140(1–2):1–47
- 25. Jakaria M, Park S-Y, Haque M, Karthivashan G, Kim I-S, Ganesan P, Choi D-K (2018) Neurotoxic agents-induced injury in neurode-generative disease model: focusing on involvement of glutamate receptors. Front Mol Neurosci 11:307
- 26. Magri C, Gardella R, Valsecchi P, Barlati SD, Guizzetti L, Imperadori L, Bonvicini C, Tura GB, Gennarelli M, Sacchetti E, Barlati S (2008) Study on GRIA2, GRIA3 and GRIA4 genes highlights a positive association between schizophrenia and GRIA3 in female patients. AM J Med Genet B 147B:745–753
- 27. Gécz J, Barnett S, Liu J, Hollway G, Donnelly A, Eyre H, Eshkevari HS, Baltazar R, Grunn A, Nagaraja R, Gilliam C, Peltonen L, Sutherland GR, Baron M, Mulley JC (1999) Characterization of the human glutamate receptor subunit 3 gene (GRIA3), a candidate for bipolar disorder and nonspecific X-linked mental retardation. Genomics 62(3):356–368

- Perlis RH, Laje G, Smoller JW, Fava M, Rush AJ, McMahon FJ (2009) Genetic and clinical predictors of sexual dysfunction in citalopram-treated depressed patients. Neuropsychopharmacology 34:1819–1824
- 29. Formicola D, Aloia A, Sampaolo S, Farina O, Diodato D, Griffiths LR, Gianfrancesco F, Iorio GD, Esposito T (2010) Common variants in the regulative regions of GRIA1 and GRIA3 receptor genes are associated with migraine susceptibility. BMC Med Genet 11:103
- Río J, Nos C, Tintoré M, Téllez N, Galán I, Pelayo R, Manuel C, Montalban X (2006) Defining the response to interferon-β in relapsing-remitting multiple sclerosis patients. Ann Neurol
- Song K, Chen Y, Göke R, Wilmen A, Seidel C, Göke A, Hilliard B, Chen Y (2000) Tumor necrosis factor–related apoptosis-inducing ligand (TRAIL) is an inhibitor of autoimmune inflammation and cell cycle progression. J Exp Med 191(7):1095–1103
- 32. Mi Q-S, Ly D, Lamhamedi-Cherradi S-E, Salojin KV, Zhou L, Grattan M, Meagher C, Zucker P, Chen YH, Nagle J, Taub D, Delovitch TL (2003) Blockade of tumor necrosis factor-related apoptosis-inducing ligand exacerbates type 1 diabetes in NOD mice. Diabetes 52(8):1967–1975
- Gourraud P, Harbo HF, Hauser SL, Baranzini SE (2012) The genetics of multiple sclerosis: an up-to-date review. Immunol Rev 248(1):87–103
- Huang W-X, Huang P, Gomes A, Hillert J (2000) Apoptosis mediators fasL and TRAIL are upregulated in peripheral blood mononuclear cells in MS. Neurology 55(7):928–934
- 35. Lopez-Gomes C, Pino-Angeles A, Orpez-Zafra T, Pinto-Medel MJ, Oliver-Martos B, Ortega-Pinazo J, Araiz C, Guijarro-Castro C, Varade J, Alvarez-Lafuente R, Urcelay E, Sanchez-Jimenez F, Fernandez O, Leyva L (2013) Candidate gene study of TRAIL

and TRAIL receptors: association with response to interferon beta therapy in multiple sclerosis patients. PLoS One 8(4):e62540

- 36. Gökdogan T, Aynur E, Osnnan Ö, Yalin Ö, Kup S, Erdal ME (2013) A study of the impact of death receptor 4 (DR4) Gene polymorphisms in Alzheimer 's disease. Balkan Med J 30(3):268–273
- 37. Morales-Lara MJ, Conesa-Zamora P, Santaclara V, Torres-Moreno D, Pedero F, Corral J, Perez-Guillermo M, Rodriguez-Martinez FJ, Soriano-Navarro E (2010) Role of TRAILR1 and TNFR1A polymorphisms in the susceptibility and pharmacogenetics of rheumatoid arthritis and ankylosing spondylitis patients treated with infliximab. J Transl Med 8(Suppl 1):P50
- Taghavi BA, Hashemi M, Niaei G, Rahmani SA (2016) Association of TNF-related apoptosis inducing ligand receptor (TRAIL-R) gene polymorphisms in Iranian Azeri patients with multiple sclerosis. Arch Adv Biosci 7(4):39–44
- 39. Taheria M, Nematia S, Movafagh A, Saberi M, Mirfakhraie R, Eftekharianc MM, Arsang-Jang S, Rezagholizadeh SA (2016) TRAIL gene expression analysis in multiple sclerosis patients. Hum Antibodies 24(1–2):33–38
- 40. Comabella M, Craig DW, Morcillo-Suarez C, Rio J, Navarro A, Fernandez M, Rolan M, Montalban X (2009) Genome-wide scan of 500 000 single-nucleotide polymorphisms among responders and nonresponders to interferon Beta therapy in multiple sclerosis. Arch Neurol 66(8):972–978

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.