Transcriptome meta-analysis of peripheral lymphomononuclear cells indicates that gestational diabetes is closer to type 1 diabetes than to type 2 diabetes mellitus

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Abstract We performed a meta-analysis of the transcription profiles of type 1, type 2 and gestational diabetes to evaluate similarities and dissimilarities among these diabetes types. cRNA samples obtained from peripheral blood lymphomononuclear cells (PBMC) of 56 diabetes mellitus patients (type 1 = 19; type 2 = 20; gestational = 17) were hybridized to the same whole human genome oligomicroarray platform, encompassing 44,000 transcripts. The GeneSpring software was used to perform analysis and hierarchical clustering, and the DAVID database was used for gene ontology. The gene expression profiles showed more similarity between gestational and type 1 diabetes rather than between type 2 and gestational diabetes, a finding that was not influenced by patient gender and age. The meta-

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Faculty of Philosophy Sciences and Letters of Ribeirão Preto, University of São Paulo, Ribeirão Preto, SP, Brazil analysis of the three types of diabetes disclosed 3,747 differentially and significantly expressed genes. A total of 486 genes were characteristic of gestational diabetes, 202 genes of type 1, and 651 genes of type 2 diabetes. 19 known genes were shared by type 1, type 2 and gestational diabetes, highlighting *EGF*, *FAM46C*, *HBEGF*, *ID1*, *SH3BGRL2*, *VEPH1*, and *TMEM158* genes. The meta-analysis of PBMC transcription profiles characterized each type of diabetes revealing that gestational and type 1 diabetes were transcriptionally related.

Keywords Gene expression profiling · Microarray analysis · Gestational diabetes mellitus · Type 1 diabetes mellitus · Type 2 diabetes mellitus

Introduction

Four major groups of diabetes mellitus have been recognized, type 1, type 2, gestational and other types. Type 1 diabetes accounts for ~10 % of all cases, affecting individuals under the age of 30 years. Several genes and gene regions distributed throughout the genome have been associated with susceptibility to type 1 diabetes, including the regions designated as insulin dependent diabetes mellitus—IDDM1 to IDDM18 [1]; however, the strongest genetic contribution comes from the HLA (IDDM1) and insulin gene (IDDM2) regions [2].

Type 2 diabetes accounts for ~90 % of all cases, developing after the age of 30 years and exhibiting no important autoimmune component; however, multi-genetic and metabolic factors, and family history have been reported. Although insulin resistance and progressive pancreatic β -cell dysfunction have been recognized as fundamental factors in the pathogenesis of type 2 diabetes

Table 1 Demographical, laboratory, and treatment features of type 1 (T1DM), type 2 (T2DM), and gestational (GDM) diabetic patients

Subjects	Age (years)	Gender	Insulin regular/ NPH	Duration of diabetes (years)	Blood glucose (mg/dL)	Glycated hemoglobin (%)	Metformin (mg/day)	Duration of pregnancy (weeks)
T1DM-01	36	М	Yes	11	213	10.8	_	_
T1DM-02	23	М	Yes	13	197	8.3	_	_
T1DM-03	24	М	Yes	6	260	10	_	_
T1DM-04	18	М	Yes	8	23	7.2	_	_
T1DM-05	23	М	Yes	20	178	10.1	_	_
T1DM-06	21	F	Yes	8	223	7.8	_	_
T1DM-07	29	М	Yes	2	59	11.1	_	_
T1DM-08	30	М	Yes	14	47	8.9	_	_
T1DM-09	21	М	Yes	16	66	9	_	_
T1DM-10	28	F	Yes	5	193	12.5	_	_
T1DM-11	29	М	Yes	3	225	9.8	_	_
T1DM-12	27	F	Yes	10	257	10.4	_	_
T1DM-13	37	F	Yes	7	82	8.4	_	_
T1DM-14	24	F	Yes	14	293	8.5	_	_
T1DM-15	22	F	Yes	13	143	8.3	_	_
T1DM-16	18	М	Yes	5	60	9.5	_	_
T1DM-17	25	F	Yes	6	85	10.5	_	_
T1DM-18	23	М	Yes	11	123	10.3	_	_
T1DM-19	25	М	Yes	8	162	7.7	_	_
Mean \pm SD	25.42 ± 5.18			9.47 ± 4.7	152.05 ± 82.87	9.42 ± 1.36		
T2DM-01	62	М	Yes	15	109	7	2,550	_
T2DM-02	49	F	Yes	11	117	7.1	850	_
T2DM-03	47	F	No	10	92	5.8	1,700	_
T2DM-04	49	М	No	8	130	7.5	2,550	_
T2DM-05	42	М	Yes	11	53	5.1	1,700	_
T2DM-06	45	F	Yes	9	81	8.2	1,700	_
T2DM-07	64	М	Yes	5	118	7	2,550	_
T2DM-08	47	F	Yes	3	237	9.4	2,550	_
T2DM-09	52	М	No	10	100	6.6	1,700	_
T2DM-10	72	F	No	9	65	6.7	1,700	_
T2DM-11	41	F	Yes	3	92	10.2	2,550	_
T2DM-12	43	F	No	4	173	10.7	2,550	_
T2DM-13	60	F	Yes	20	306	10.9	0	_
T2DM-14	66	М	No	5	183	9.4	1,700	_
T2DM-15	51	F	Yes	10	219	10.2	0	_
T2DM-16	56	F	Yes	20	295	12	2,550	_
T2DM-17	61	F	Yes	20	101	7.8	2,550	_
T2DM-18	60	F	Yes	9	319	11.4	2,550	_
T2DM-19	45	F	Yes	1	254.1	9.6	2,550	_
T2DM-20	44	М	No	5	146	5.3	1,700	_
Mean \pm SD	52.8 ± 9.13			9.4 ± 5.69	159.5 ± 85.02	8.39 ± 2.09		
GDM-01	35	F	No		88	5.6	_	30
GDM-02	37	F	Yes		86	7.9	_	25
GDM-03	30	F	No		67	5.1	_	26
GDM-04	40	F	No		74	_	_	38
GDM-05	35	F	No		94	8.8	_	27
GDM-06	29	F	No		65	5.7	-	34

Table 1 continued

Subjects	Age (years)	Gender	Insulin regular/ NPH	Duration of diabetes (years)	Blood glucose (mg/dL)	Glycated hemoglobin (%)	Metformin (mg/day)	Duration of pregnancy (weeks)
GDM-07	24	F	No		74	5.4	_	29
GDM-08	23	F	No		86	5.4	_	34
GDM-09	33	F	No		80	5	_	35
GDM-10	29	F	Yes		89	5.8	_	37
GDM-11	37	F	No		80	_	_	37
GDM-12	39	F	No		72	5.7	_	32
GDM-13	29	F	Yes		92	5.2	_	28
GDM-14	30	F	No		82	5	_	34
GDM-15	33	F	Yes		94	9.2	_	18
GDM-16	38	F	Yes		59	9.1	_	34
GDM-17	28	F	No		81	6.1	_	34
$\text{Mean} \pm \text{SD}$	32.29 ± 5.08				80.17 ± 10.39	6.33 ± 1.56		31.29 ± 5.25

Duration of diabetes was defined as the period from diabetes onset until the enrollment in the study. The individual values and mean \pm standard deviation (SD) are expressed

M male, F female, NPH neutral protamine Hagedorn



Fig. 1 Dendrogram showing the clustering of samples according to the gene expression profile encompassing 3,747 differentially and significantly expressed genes in patients with type 1 (T1DM), type 2

[3], the specific molecular mechanisms that affect insulin sensitivity and β cell function remain unknown. The genomic scanning of susceptibility regions for type 2 diabetes included genes of cellular metabolism (*PPARG*, *KCNJ11* and *CAPN10*) and transcription factors (*HNF4A* and *TCF7L2*) [4].

Gestational diabetes has been defined as any degree of impaired glucose tolerance with onset during pregnancy, affecting ~ 7 % of pregnant women. The genetic risk for gestational diabetes has been reported to be heterogeneous [5] and no genomic scanning has been reported for this type of diabetes. Gestational diabetes shares with type 2 diabetes clinical features, life-style risk factors and genetic susceptibility genes, including *KCNJ11*, *TCF7L2*, *CDKAL1*, *CDKN2A/CDKN2B*, *HHEX/IDE*, *IGFBP2*, *SLC30A8*, and *FTO* [6]. On the other hand, 10 % of gestational diabetes patients show high frequency of *HLA-DRB1**03 and *HLA-DRB1**04 allele groups, which is similar to type 1 diabetes patients, and may produce autoantibodies against pancreas antigens during and after pregnancy [7].

(T2DM) and gestational diabetes mellitus (GDM). The hierarchical clustering discriminated each type of diabetes and clustered gestational diabetes closer to type 1 diabetes than to type 2 diabetes

Peripheral blood lymphomononuclear cells (PBMCs) have been widely used in microarray experiments evaluating patients with distinct disorders, including diabetes [8] and pre-diabetes [9], as reporters of the underlying pathogenic processes. Considering the intermingled gene susceptibility and transcription profiles among the major types of diabetes mellitus, in the present study we performed a meta-analysis of the transcription profiles observed in PBMCs of these three types of diabetes, using the same microarray platform.

Materials and methods

Subjects

Peripheral blood lymphomononuclear cells were obtained from 56 patients, i.e., 19 type 1 diabetes patients (7 women) aged 18–36 years, 20 type 2 diabetes patients (13 women) aged 41–72 years, and 17 patients with gestational diabetes aged 23–40 years, followed-up at the Outpatient Clinics of the Division of Endocrinology, Faculty of Medicine of Ribeirao Preto, University of Sao Paulo, Brazil. Major demographic, laboratory and treatment features of type 1, type 2, and gestational diabetes patients are shown in Table 1. The local Ethics Committee approved the protocol of the study (# 9153/2008) and informed written consent was obtained from all patients.

Total RNA extraction

Cells were isolated using Ficoll-Hypaque (Sigma, St. Louis, MO). Total RNA was extracted using Trizol reagent (Invitrogen, Carlsbad, CA), confirmed to be free of proteins or phenol using UV spectrophotometry, and RNA integrity was assessed by microfluidic electrophoresis (Bioanalyzer Model 2100 and RNA 6000 nanochips, both from Agilent Technologies, Palo Alto, CA).

Microarray hybridization

Hybridizations to whole human genome 4×44 K oligo microarrays (G4112F, Agilent) were performed using the one color (Cy3) Quick Amp labeling kit (Agilent). Samples of complementary RNA were hybridized for 17 h at 65 °C. Images were acquired using a DNA Microarray Scanner with Surescan High-Resolution Technology (Agilent). Hybridization quantitative numerical data and microarray quality control were performed using the Feature Extraction software (Agilent). All microarray data are available online at the MIAME database (type 1 diabetes: E-MEXP-3348; type 2 diabetes: E-MEXP-3287; gestational diabetes: E-MEXP-3349).

Microarray analysis

The GeneSpring GX version 11.0 software (Agilent) was used for background adjustment, quantile normalization data, logtransformation, and gene clustering. Gene ontology was performed using the GeneSpring or DAVID database (http:// david.abcc.ncifcrf.gov/). Meta-analyses were performed using one-way analysis of variance (ANOVA) (three types of diabetes) or the unpaired T test (two types of diabetes), with p values set at p < 0.05. In both situations, the Benjamini– Hochberg false discovery rate multiple testing corrections were performed to minimize selection of false-positive values. Overall, in the present study were considered differentially and significantly expressed genes exhibiting expression values greater than two-folds (fold change ≥ 2.0). Hierarchical clustering of differentially expressed genes was performed using average linkage and the Pearson uncentered distance metrics. In this kind of clustering, the relationship among genes is represented by a tree, whose branch lengths reflect the degree of similarity between genes.

Results

Hierarchical clustering analysis of the transcription profiles displayed by PBMCs of type 1, type 2 and gestational diabetic patients individualized each patient group. Overall, the transcription profile exhibited by the group of gestational diabetes was closer to type 1 diabetes than to type 2 diabetes (Fig. 1); however, one patient with type 2 diabetes (T2DM-10) clustered with the type 1 diabetes group. No particular clinical feature exhibited by this patient distinguished him from the type 2 diabetes group, except that patient was usually compensated in terms of glucose (65 mg/dL) and HbA1c (6.5 %) levels.

The meta-analysis encompassing type 1, type 2 and gestational diabetes revealed 3,747 differentially and significantly expressed genes, i.e., these genes exhibited foldchange ≥ 2.0 in at least one comparison between the diabetes groups. Among these genes, 805 were up-regulated and 570 were down-regulated. Overall, up-regulated genes were clustered into five major biological functions: (i) multicellular organism development (20.3 %), (ii) signal transduction (17.9 %), (iii) response to stress (12.2 %), (iv) cell differentiation (10.7 %), and (v) immune system processes (6.8 %). The down-regulated genes were primarily clustered into three processes: (i) metabolism regulation (30 %), (ii) biosynthetic pathways (26.9 %), and (iii) transcription machinery (22 %).

The multiple comparisons between the three types of diabetes revealed that: (i) 1,171 genes were differentially expressed between type 1 and gestational diabetes, (ii) 2,405 genes were differentially expressed between type 2 and gestational diabetes, and (iii) 1,509 genes were differentially expressed between type 1 and type 2 diabetes, of which, 486 genes were characteristic of gestational diabetes, 202 genes were specific for type 1, and 651 genes were typical of type 2 diabetes. Twenty-two transcripts were shared by type 1, type 2 and gestational diabetes, of which 19 have been defined by public gene banks. These data are shown in Fig. 2.

A more restrictive analysis of modulated genes exhibiting fold-change ≥ 2.0 in all comparisons of the three diabetes groups yielded seven genes, that were up-regulated in all comparisons between groups (GDM vs T1DM, GDM vs T2DM, and T1DM vs T2DM) (Table 2).

To circumvent the possible influence of gender on transcriptome profile, we performed a meta-analysis excluding men, and the expression profiles were maintained (Fig. 3). Considering that the median age of type 2 diabetes patients was higher than that of patients with other types, we reanalyzed data according to median age stratification of each group (GDM = 24 years; T1D = 33 years and T2D = 50 years), yielding four subgroups: (i) patients aged <24 years, (ii) patients aged between 24 and

Fig. 2 Sets of comparisons between type 1 (T1DM), type 2 (T2DM) and gestational diabetes mellitus (GDM) patients, showing private and shared genes in each type of diabetes. Twenty-two transcripts were shared and up-regulated in the three types of diabetes, of which 19 have been described in public databanks presenting known molecular and biological functions. The gene name, gene symbol and gene cytoband are also shown



Gene Symbol – Name	Cytoband
EGF- epidermal growth factor (beta- urogastrone)	4q25
MACROD1- MACRO domain containing 1	11q13.1
RPS4Y2- ribosomal protein S4, Y-linked 2	Yq11.223
CXCL3- chemokine (C-X-C motif) ligand 3	4q13.3
FAM46C- family with sequence similarity 46, member C	1p12
SH3BGRL2- SH3 domain binding glutamic acid-rich protein like 2	6q14.1
USP9Y- ubiquitin specific peptidase 9, Y- linked	Yq11.221
HBEGF- heparin-binding EGF-like growth factor	5q31.3
CYorf15A- chromosome Y open reading frame 15A	Yq11.223
HBD- hemoglobin, delta	11p15.4
VEPH1- ventricular zone expressed PH domain homolog 1 (zebrafish)	3q25.31
ZNF586- zinc finger protein 586	19q13.41
TMEM158- transmembrane protein 158	3p21.31
WDR5B- WD repeat domain 5B	3q13.33
ZNF738- zinc finger protein 738	19p12
THAP2- THAP domain containing, apoptosis associated protein 2	12q21.1
C8orf4- chromosome 8 open reading frame 4	8p11.21
ID1- inhibitor of DNA binding 1, dominant negative helix-loop-helix protein	20q11.21
ERB3- v-erb-b2 erythroblastic leukemia viral oncogene homolog 3 (avian)	12q14.1

Table 2 Meta-analysis of the transcriptional profiles observed in patients exhibiting type 1 (T1DM), type 2 (T2DM) and gestational diabetes mellitus (GDM) showed 3,747 differentially and significantly

expressed genes. Among these genes, seven were induced (fold change ≥ 2.0) in all comparisons between groups (GDM vs T1DM, GDM vs T2DM, and T1DM vs T2DM)

Symbol	Name	Cytoband	Modulation GDM \times T1DM	Modulation GDM \times T2DM	Modulation T1DM \times T2DM	Gene ID
EGF	Epidermal growth factor	4q25	↑ (2.04)	↑ (4.09)	↑ (2.00)	1950
FAM46C	Family with sequence similarity 46, member C	1p12	↑ (4.86)	↑ (16.08)	↑ (3.31)	54855
HBEGF	Heparin-binding EGF-like growth factor	5q23	↑ (2.06)	↑ (4.53)	↑ (2.19)	1839
ID1	Inhibitor of DNA binding 1, dominant negative helix-loop-helix protein	20q11	↑ (3.29)	↑ (32.66)	↑ (9.91)	3397
SH3BGRL2	SH3 domain binding glutamic acid-rich protein like 2	6q14.1	↑ (2.48)	↑ (5.17)	↑ (2.08)	83699
VEPH1	Ventricular zone expressed PH domain homolog 1 (zebrafish)	3q24-q25	↑ (2.052)	↑ (4.64)	↑ (2.26)	79674
<i>TMEM158</i>	Transmembrane protein 158 (gene/ pseudogene)	3p21.3	↑ (2.69)	↑ (5.57)	↑ (2.07)	25907

33 years, (iii) patients aged between 33 and 50 years, and (iv) patients aged more than 50 years. Hierarchical clustering was not influenced by age, as shown in Fig. 4.

Discussion

In this study, we assessed the hypothesis that each type of diabetes mellitus may display a specific expression profile signatures, and also may share sets of genes with similar expression profiles. To achieve this goal we performed a meta-analysis of the transcription profiles exhibited by each patient group, confirming that the overall gene expression profile is characteristic for each group of diabetic patients, and was not influenced by patient gender or age. Although these variables did not influence gene profiles, other variables including body mass index, disease duration, glucose and glycated hemoglobin levels, subset of peripheral lymphomononuclear cells, type of hypoglycemic agent, compensate or uncompensated disease, and





(T2DM) and gestational diabetes mellitus (GDM), exhibited a similar profile as that observed encompassing both genders



GROUPS					
G1 < 24 years	24 years <g2<33 years</g2<33 	33 years <g3<50 years</g3<50 	G4 >50 years		
T1DM-01	T1DM-03	GDM-02	T2DM-03		
T1DM-02	T1DM-07	GDM-04	T2DM-04		
T1DM-04	T1DM-08	GDM-05	T2DM-05		
T1DM-05	T1DM-10	GDM-09	T2DM-06		
T1DM-06	T1DM-11	GDM-11	T2DM-07		
T1DM-09	T1DM-12	GDM-12	T2DM-08		
T1DM-13	T1DM-14	GDM-15	T2DM-09		
T1DM-15	T1DM-17	GDM-16	T2DM-10		
T1DM-16	T1DM-19	T2DM-01	T2DM-17		
T1DM-18	GDM-06	T2DM-02	T2DM-19		
GDM-01	GDM-07	T2DM-11			
GDM-08	GDM-10	T2DM-12			
	GDM-13	T2DM-13			
	GDM-14	T2DM-14			
	GDM-17	T2DM-15			
		T2DM-16			
		T2DM-18			
		T2DM-20			

Fig. 4 The clustering of samples according to the gene expression profile in patients with type 1 (T1DM), type 2 (T2DM) and gestational diabetes mellitus (GDM), stratified according to age, still grouped gestational diabetes closer to type 1 diabetes in relation to type 2 diabetes

others, may also be confounder factors that need to be evaluated, and will be a matter of our further and larger studies. Despite these possible confounding factors, we unveiled shared genes in each set of comparison between the three types of diabetes.

Several lines of clinical evidence emphasize more similarities between gestational diabetes and type 2 diabetes, rather than between gestational and type 1 diabetes [5]; however, the present study showed that the overall gestational diabetes transcriptional profile was closer to type 1 than to type 2 diabetes. Considering the whole group of gestational diabetes, these patients exhibited 5 % of their modulated genes in susceptibility regions previously described in association with type 1 diabetes [insulindependent diabetes mellitus regions (IDDM-1, 2, 3, 4, 6, 7, 8, 10, 11, 12, 13, 15, 17, 18, X) and PTPN22], and gestational diabetes exhibited only 0.5 % of differentially expressed genes that coincided with or were close to previously described type 2 diabetes susceptibility genes/ regions. In addition, in the meta-analysis of the transcription profiles obtained for the three types of diabetes we observed that 50 (6.8 %) of the up-regulated genes were related to the immune response, including chemokines and chemokine receptors, interleukins, complement receptors, lymphocyte receptors among others. These findings corroborate the data of a recent study indicating that immune pathogenic mechanisms might be involved in gestational diabetes [7].

Considering the seven genes that were highly up-regulated in all patient comparisons, four of them (*EGF*, *FAM46C*, *HBEGF*, and *ID1*) have been previously described in association with at least one type of diabetes, but they have not been studied in all types of diabetes yet. Compared to healthy individuals, the expression of the *EGF* gene has been reported to be decreased after the onset of type 2 diabetes [10]. On the other hand, compared to non-diabetic pregnant women, it has been reported that gestational diabetes patients and their macrosomic children exhibited increased serum levels of EGF, and increased expression of the EGF receptor in placenta [11]. Taken together, the differential relative expression of EGF may be associated with an impairment of pancreas endocrine function and diabetes complications in gestational diabetes.

FAM46C encodes a type 1 interferon-stimulated protein that mediates the defense against invading viral pathogens and appears to contribute to the development of autoimmune diseases, including lupus, rheumatoid arthritis, multiple sclerosis, Sjögren's syndrome and type 1 diabetes [12, 13]. Since the expression of this gene was up-regulated in gestational diabetes, and since it has been associated with autoimmune diseases, *FAM46C* gene may be of interest regarding the follow-up of gestational diabetes patients who develop post-pregnancy diabetes.

The *HBEGF* gene encodes an adipocytokine that contributes to the development of vascular diseases, and plasma HBEGF levels increase with the extent of obesity and of visceral adiposity [14]. The differential expression of this gene in three types of diabetes may be used as a tool to correlate body mass index and disease complications. The *ID1* gene encodes a transcription factor that plays an important role during development and differentiation of many tissues and cells, and is essential for lymphocyte function and development. *ID1* gene has been reported to be up-regulated in patients with rheumatoid arthritis [15] and in type 1 diabetes [16]. Therefore, the differential modulation of this gene in gestational and type 1 diabetes in relation to type 2 diabetes (Table 1) further supports the role of immune system genes on the pathogenesis of gestational diabetes.

The other three up-regulated genes (*SH3BGRL2*, *VEPH1*, and *TMEM158*) have not been previously studied in diabetes, deserving further studies. The *SH3BGRL2* gene participates in mature red cell function and is important to protect erythrocytes against injurious oxygen effects [17]. The *VEPH1* gene encodes a protein that is expressed during the development of the central nervous system [18], and the *TMEM158* gene, which encodes a transmembrane protein, is up-regulated in response to the activation of the RAS pathway [19].

This is the first transcriptome study evaluating the three major types of diabetes using the same microarray platform. The meta-analysis of transcriptome profiles displayed by the three types of diabetes discriminated hybridization signatures, which were distinctive of each type of the disease, and showed that gestational diabetes hybridization signature is closer to type 1 than to type 2 diabetes. Besides, irrespective of the pathogenetic mechanisms involved in each type of diabetes, common sets of genes are similarly modulated.

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