# Characterisation and development of EST-SSR markers in tree peony using transcriptome sequences

Jing Wu · Changfu Cai · Fangyun Cheng · Huliang Cui · Hua Zhou

Received: 25 February 2014/Accepted: 30 June 2014/Published online: 8 July 2014 © Springer Science+Business Media Dordrecht 2014

Abstract Tree peony is a well-known ornamental and medicinal plant in China, as well as a new crop that has been recently demonstrated to exhibit a great potential for edible oil production. However, only limited molecular markers are currently available in this species, which hinders its genetic and breeding studies. The objective of this study was to develop expressed sequence tag (EST)-simple sequence repeat (SSR) markers based on transcriptome sequences in tree peony. A total of 4,373 EST-SSRs were identified from 3,787 unigene sequences with an average of one SSR per 9.24 kb. Di-nucleotide repeats (46.26 %) were the most abundant type followed by tri-, hexa-, tetra-, and penta-nucleotide repeats. Moreover, 788 primer pairs were synthesised, out of which 373 primer pairs (47.3 %) produced successful amplifications, and 149 primer pairs (39.9 %) were polymorphic in both parents of one mapping population. The 30 polymorphic EST-SSR markers were randomly selected to assess polymorphism further in 36 tree

**Electronic supplementary material** The online version of this article (doi:10.1007/s11032-014-0144-x) contains supplementary material, which is available to authorized users.

J. Wu · C. Cai · F. Cheng (⊠) · H. Cui · H. Zhou National Flower Engineering Research Centre, Beijing Key Laboratory of Ornamental Plants Germplasm Innovation and Molecular Breeding, College of Landscape Architecture of Beijing Forestry University, Beijing 100083, People's Republic of China e-mail: chengfy8@263.net peony accessions, in which the polymorphism information content values ranged from 0.36 to 0.85 with a mean of 0.69, thereby indicating a high level of informativeness. Transferability studies indicated that 95.3 % EST-SSR markers were transferable to nine other *Paeonia* species, thereby expanding their utility. In addition, as a demonstration of utility, a genetic relationship study was performed in 56 accessions. Cluster analysis using the neighbour-joining tree method demonstrated that major clusters corresponded to known pedigree trees. Thus, these newly developed markers will be valuable in facilitating marker-assisted selection breeding in tree peony and its related species.

**Keywords** *Paeonia suffruticosa* · Simple sequence repeats (SSRs) · Transcriptome sequences · Polymorphism · Transferability · Genetic relationships

# Introduction

The genus *Paeonia* includes three sections, namely sect. *Moutan*, sect. *Paeonia*, and sect. *Onaepia* (Stern 1946). Intersectional hybridization between the sect. *Moutan* and sect. *Paeonia* forms a new type of group which has been called Itoh Hybrid. Tree peony (*Paeonia suffruticosa* Andrews), which belongs to the section *Moutan*, is composed of the subsection *Vaginatae* and the subsection *Delavayanae* (Stern 1946). This species is a perennial woody flower plant

native to China with approximately 2,000 cultivars. China is also the origin and developmental centre of the cultivated tree peony with a cultivation history of 1,600 years. Due to its striking ornamental and medicinal value, as well as its unique culture symbolisation, the tree peony has become one of the most important horticultural flower crops in the world and is widely cultivated in many countries in Asia, America, Europe, and Australia (Cheng 2007). In addition, recent studies have shown that the tree peony seed has a high rate of oil yield (27-33 %), and seed oil contains abundant unsaturated fatty acids, such as linolenic acid (33.87 %), oleic acid (27.73 %), and linoleic acid (21.40 %) (Zhou et al. 2009). Thus, it has been proposed that the tree peony seed has a great potential for edible oil production as a new crop, which will have broad market prospects and is gaining increasing attention from the Chinese government (Li et al. 2012a).

Traditionally, the breeding of tree peony is based on hybridisation among species and cultivars and the phenotypic selection of novel or improved offspring, followed by vegetative propagation of released cultivars. Although traditional breeding still plays an important role in improving the quality of floral crops, it is limited by the long selection cycle and enormous resources required in the cultivation of new varieties. Thus, any method that offers a shortcut to breeding would be invaluable in the improvement of woody flower crops, such as the tree peony. Marker-assisted selection (MAS) has the great potential to accelerate the crop breeding; it will enable breeders to quickly select cultivars with desirable traits, thereby saving time and resources. However, the basic requirement of MAS is to find the molecular markers associated with important traits. In tree peony, MAS has been hindered due to lack of effective molecular markers.

Within the last two decades, many types of markers have been developed and used for crop breeding (Paux et al. 2012). Of these various markers, simple sequence repeats (SSRs) are widely used due to their co-dominant inheritance, multi-allelic nature, high reproducibility and transferability, extensive genome coverage and simple detection (Agarwal et al. 2008; Varshney et al. 2005). According to the original sequences used to identify SSR markers, SSRs can be divided into genomic SSRs and expressed sequence tag (EST) SSRs. The traditional development of genomic SSRs is both time-consuming and costintensive and involves the construction and sequencing of a genomic library (Eujayl et al. 2004). In contrast, EST-SSRs can be rapidly mined at a lower cost from expressed sequences, which are more conserved compared to non-coding sequences. Thus, EST-SSRs demonstrate a higher level of transferability across related species compared to genomic SSR markers (Zhou et al. 2014; Zeng et al. 2010). Moreover, due to their association with coding sequences, EST-SSRs have the potential for direct gene tagging for important horticultural traits (Bouck and Vision 2007).

To date, more than 200 SSRs markers have been developed in tree peony (Gao et al. 2013; Yu et al. 2013; Gai et al. 2012; Zhang et al. 2011, 2012; Hou et al. 2011a, b; Homolka et al. 2010; Wang et al. 2009), and they have been applied effectively to study hybrid origin (Yuan et al. 2010), genetic diversity (Yuan et al. 2012), cultivar identification (Zhang et al. 2012), genetic relationship (Yu et al. 2013) and the domestication of cultivated tree peonies (Yuan et al. 2014). However, compared with crop plants such as maize, rice, and wheat, the number of SSR markers for further molecular breeding of tree peony is inadequate. In addition, the size of the tree peony genome is very large, approximately 16 G (Gao et al. 2013). We speculate the developed SSR markers are very hard to cover and be evenly distributed to the entire tree peony genome, for the limited number of markers. Meanwhile, fewer SSRs have been developed for the herbaceous species of the genus Paeonia, P. lactiflora (Li et al. 2011; Sun et al. 2011; Gilmore et al. 2013). Thus, it is still very essential to develop more effective SSR markers for MAS in tree peony breeding.

In recent years, an increasing number of EST data sets have become available for model and non-model organisms; however, relatively few ESTs are currently available for tree peony. Transcriptome sequencing using next-generation sequencing (NGS) technology provides a simple and effective method to develop a large number of unigene-based SSR markers. In tree peony, our laboratory has previously performed *de novo* transcriptome sequencing for *P. suffruticosa* 'Luo Yang Hong' using the Illumina sequencing platform and identified a total of 59,275 unigene ESTs (Zhou et al. 2013). In this study, a comprehensive set of EST-SSRs for tree peony was identified and characterised in terms of frequency and distribution, informativeness, and transferability to related species. Moreover, the effectiveness of these SSR markers will be assessed in the study of genetic relationships between 56 accessions of *Paeonia*.

# Materials and methods

#### Plant material and DNA extraction

P. suffruticosa 'Luo Yang Hong' was first used to confirm the amplification specificity of the synthesised EST-SSR primers. Two other common cultivated varieties, P. ostii 'Feng Dan' and P. suffruticosa 'Hong Qiao', which were crossed to establish the mapping population in our laboratory, were used to examine polymorphic microsatellite loci. A set of 56 accessions, including 36 tree peonies (section *Moutan*) (8 species and 28 cultivars), 8 herbaceous peonies (section Paeonia) (2 species and 6 cultivars), and 12 intersectional hybrids between the two types (Itoh hybrids), was used to analyse the polymorphisms and transferability of 30 EST-SSR markers. And the genetic relationships of these 56 accessions were also analysed using 30 EST-SSR markers. The tree peony species used in this study included P. jishanensis, P. qiui, P. rockii, P. decomposita, P. ostii, P. delavayi var. lutea and P. ludlowii, all of which belong to the subsection Vaginatae except P. delavayi var. lutea and P. ludlowii which belong to the subsection Delavayanae. The herbaceous peony species included P. veitchii and P. lactiflora. The 46 cultivar accessions used (Table 2, 9-48, 51-56) included six distinct cultivar types, which consisted of Suffruticosa hybrid, Rockii hybrid, Lemoinei hybrid, Itoh hybrid, Lactiflora cultivar, and Hybrid peony. Detailed descriptions of the 56 accessions are listed in Table 2. All of these samples were collected from the Luoyang National Tree Peony Collection (Luoyang, Henan, China) and the Jiufeng Forestry Experiment Station of Beijing Forestry University (Beijing, China) in the spring of 2012 from fresh young leaves.

Genomic DNA was extracted from silica gel-dried leaves using a DNA secure plant kit (Tiangen Biotech, Beijing, China). The DNA quality and quantity were confirmed using 1 % agarose gels and Unico UV–visible Spectrophotometer (Unico, USA), respectively. The working concentration of DNA was adjusted to 25 ng/ $\mu$ L. EST-SSRs identification, primer design, PCR amplification

A total of 59,275 unigenes were obtained from tree peony 'Luo Yang Hong' transcriptome sequences of flower buds (Zhou et al. 2013). The simple sequence repeat identification tool program (SSRIT) (http:// www.gramene.org/db/markers/ssrtool) (Temnykh et al. 2001) was used to search for SSRs within this unigene ESTs database. In this study, only SSR loci were considered to contain di-, tri-, tetra-, penta- and hexa-nucleotides with minimum repeat numbers of 6, 5, 4, 4, and 3, respectively. Mononucleotide repeats and complex SSR types were excluded from the study. To predict the SSRs location and whether the SSR was present in the coding sequence (CDS), 5' untranslated region (5' UTR) or 3' untranslated region (3' UTR), the ORF Finder software (http://www.ncbi.nlm.nih.gov/ gorf/gorf.html) was used to identify the initiation and termination codons in the EST sequences.

Primers were designed to flank the SSRs using Primer Premier 5.0 software (Premier Biosoft International, Palo Alto, CA, USA). Primers were synthesised by Shanghai Sangon Biological Engineering Technology (Shanghai, China). Polymerase chain reactions (PCRs) were performed in 10-µl reaction volumes containing 25 ng of template DNA, 5  $\mu$ l of 1× Power Taq PCR MasterMix (BioTeke, Beijing, China), 0.5 µl of 10 µM each primer and 3 µl sterile distilled water. The following PCR program included an initial step at 94 °C for 3 min followed by 30 cycles of 94 °C for 30 s, the appropriate annealing temperature (see Table S1) for 30 s and 1 min at 72 °C, and a final extension for 10 min at 72 °C. Finally, the products were examined on 2 % agarose gels. Amplified products that showed a band of the expected size were separated on 6 % denaturing polyacrylamide gel electrophoresis and then visualised using silver staining. To confirm the reproducibility of the results, PCR amplification with each primer pair was performed three times.

### Putative functional annotation

To identify the putative function of the unigene sequences containing the polymorphic microsatellite loci, the sequences were subjected to a BLAST (http://blast.ncbi.nlm.nih.gov/Blast.cgi) search against the NCBI nonredundant protein database using the BLASTX program. Each unigene sequence was

assigned to its putative functions based on their highest scores to BLASTX. Sequences with *E*-value of less than  $10^{-6}$  and scores of more than 50 were designated as 'significant similarity matches' and were annotated.

# Data analysis

The SSR marker diversity was estimated using POPGENE version 1.31 (Yeh et al. 1999), which included the following parameters: the number of alleles ( $N_a$ ), the observed and expected heterozygosities ( $H_o$  and  $H_e$ ), polymorphic information content (PIC), and Shannon index. The availability of the EST-SSR was validated using cluster analyses to evaluate the genetic relationships between species and cultivars. A neighbour-joining (NJ) tree was constructed using the proportion of shared alleles coefficient from the program PowerMarker version 3.25 (Liu and Muse 2005). Eight herbaceous peony accessions were used as an outgroup in the NJ tree. The robustness of the dendrogram was subsequently tested using bootstrap analysis with 1,000 replications.

# Results

Frequency and distribution of EST-SSRs in the tree peony transcriptome

A total of 59,275 unigenes (19,288 contigs and 39,987 singletons) with a mean length of 698 bp were obtained to identify perfect SSRs, which represented approximately 39.5 Mb. Using the SSRIT tool, 4,373 potential EST-SSRs were identified from 3,787 (6.4 %) unigene sequences, of which 500 sequences (13.2 %) contained more than one SSR. On average, one SSR was identified for every 9.24 kb, or corresponded to one SSR for every 13.6 unigenes in the tree peony transcriptome. The EST-SSRs contained five diverse types of repeat motifs, and there was an uneven distribution of EST-SSRs. Among the motifs identified, di-nucleotide repeats were the most abundant type with a frequency of 46.26 % (2,023) followed by tri-nucleotide (27.30 %, 1,194), hexanucleotide (21.88 %, 957), tetra-nucleotide (3.06 %, 134) and penta-nucleotide repeats (1.49 %, 65) (Table 1).

 Table 1
 Summary of EST-SSRs identified in tree peony transcriptome

Searching items	Numbers
Total number of sequences examined	59,275
Total size of examined sequences (Mb)	39.5
Number of SSR-containing sequences	3,787
Total number of identified SSRs	4,373
Number of sequences containing more than 1 SSR	500
Frequency of SSRs	1/9.24 kb
Di-nucleotide	2,023
Tri-nucleotide	1,194
Tetra-nucleotide	134
Penta-nucleotide	65
Hexa-nucleotide	957

The frequency of 28 major types with di- and trinucleotide repeat motifs and repeat number of SSRs are shown in Fig. 1. Among the di-nucleotide motifs, the most common motifs were AG/CT (41.4 %) and GA/TC (39.9 %), which were also the most dominant motif of all SSRs, followed by the motif AT/TA (11 %), CA/TG (4.4 %), AC/GT (2.9 %), and CG/GC (0.4 %). Among the tri-nucleotide motifs, CCA/TGG (10.3 %) and GAA/TTC (10.3 %) were the richest motif (Fig. 1a, b). However, there were no obvious dominant motifs among the tetra-, penta- and hexanucleotide motifs. The SSR length was mostly distributed from 12 to 24 bp, which accounted for 99.1 % of the total SSRs, followed by 25-66 bp (0.9 %). Among them, the SSR loci with a length of 18 bp were the most common. The repeat unit number of SSR ranged from 3 to 17, and SSRs with six repeats were the most abundant followed by SSRs with five, three and seven repeats, while motifs of more than 11 reiterations were notably rare with a frequency of 0.2 % (Fig. 1c).

EST-SSR marker development, and polymorphic microsatellite loci screening and functional annotation

A total of 2,989 SSR-containing sequences were amenable for primer design, while the remaining 1,384 sequences did not qualify for primer design because the sequences flanking the SSRs were too short or inappropriate for primer design. Of the 2,989 Fig. 1 Characterisation of SSRs in tree peony transcriptome. a Frequency distribution of di-nucleotide SSRs based on motif type;
b frequency distribution of tri-nucleotide SSRs based on main motif type;
c number of different repeat motifs



primer pairs, 788 primer pairs were selected for primer synthesis on the basis of a software design score of more than 85. Of these SSR primers, 373 primers (47.3 %) were able to generate PCR products of the expected sizes. Details of these 373 EST-SSR primers are available in Table S1. In addition, 84 primer pairs amplified fragments that were larger than expected, suggesting that there may be an intron within the amplicons. In several cases, it is assumed that the examined polymorphism was caused by an intron polymorphism, which may overshadow a putative polymorphism of the microsatellite. Thus, these 84 primer pairs were excluded from further analyses. The remaining 331 primer pairs failed to amplify bands after subjected to various PCR programs or they generated multiple bands; thus, they were not selected for further analysis. Our analysis of the position of the 373 validated SSR markers within the transcriptome sequences showed that 219 markers (58.7 %) were present in the CDS, 147 markers (39.4 %) were present in the UTR (11.3 % in 3' UTR and 28.1 % in 5' UTR), and the remaining seven markers were present in unigenes without any hit to known proteins. In addition, 61.9 % of di-repeats, 75 % of tetra-repeats, 100 % of penta-repeats occurred within the UTR, whereas 82.1 % of the trirepeats and 73.5 % of the hexa-repeats occurred within the CDS.

All 373 validated primer pairs were first screened for polymorphisms in both parents of one mapping population. The amplification results suggested that 149 primer pairs (39.9 %) were polymorphic, including 54 di-, 41 tri-, 5 tetra-, 1 penta-, and 48 hexa-motif loci. The polymorphic proportion of the di-, tri-, tetra-, penta-, and hexa-motif loci were 38.8, 35.0, 31.3, 33.3, and 49.0 %, respectively. Analysis of the proportion of polymorphic loci among the three types of EST-SSR loci demonstrated that the proportion of polymorphic loci of those located in the 3' UTR (52.4 %) were the highest followed by those located in the 5'UTR (41.0%) and CDS (37.4%). For the 149 polymorphic loci, a total of 323 alleles were detected, and the average number of alleles per locus was 2.17 with a range of 2-4 (Table S2).

To explore the potential function of these polymorphic SSR-containing sequences, a search was performed in GenBank using BLASTX against 149 transcriptome sequences of the 149 polymorphic markers. These results showed that 108 sequences (72.5 %) of the 108 polymorphic EST-SSRs demonstrated a similarity to protein sequences, while the other 41 (27.5 %) did not show a significant match in the database and thus may encode proteins that were unique to the tree peony genome. Of the 108 annotated sequences, 50 sequences (46.3 %) exhibited putative functions in grape, 18 sequences (16.7 %) were matched to the Theobroma cacao protein, 11 sequences (10.2 %) were matched to the *Ricinus communis* protein, 9 sequences (8.3 %) were matched to genes in Prunus persica, and the remaining sequences (18.5 %) were matched to genes of other dicotyledonous plants, such as Poplar trichocarpa, Solanum lycopersicum, and among others. None of the sequences were matched to genes in monocotyledons (Table S2).

Polymorphic analysis and transferability of 30 polymorphic EST-SSR markers

In total, 30 EST-SSR markers, which demonstrated polymorphism when tested in both parents of one mapping population, were randomly selected and further used to assess polymorphism in 36 accessions of tree peonies (Table 2, 1–36). The results showed that 30 (100 %) markers were polymorphic in the genotypes examined, which revealed 254 alleles. The number of alleles varied from 3 to 12 with an average of 7.4 per locus. Furthermore, the observed heterozygosity ( $H_0$ )

and expected heterozygosity ( $H_e$ ) ranged from 0.19 to 0.81 (mean 0.52) and 0.43 to 0.87 (mean 0.74), respectively. The PIC values for 30 SSR markers ranged from 0.36 for PS290 to 0.85 for PS004 with an average value of 0.69. These findings indicated a high level of informativeness within these EST-SSRs loci. Furthermore, 28 markers showed PIC values greater than 0.5. In addition, the Shannon index, which can also reflect the polymorphism of markers, ranged from 0.83 to 2.14 and the mean value was 1.56 (Table 3).

These thirty primer pairs were also used to test their transferability across nine *Paeonias* species (Table 2, 1–7, 49–50). Thirty primer pairs revealed 100 % transferability rates in seven tree peony species, including *P. delavayi* var. *lutea*, *P. ludlowii*, *P. jishanensis*, *P. qiui*, *P. rockii*, *P. decomposita*, and *P. ostii*. Of these primer pairs, three primer pairs, PS099, PS106, and PS219 could not amplify a fragment in *P. veitchii*; two primer pairs, PS099 and PS166, could not amplify a fragment in *P. lactiflora*; and the remaining 26 primer pairs successfully amplified PCR products in *P. veitchii* and *P. lactiflora*. Thus, the transferability rate across nine species ranged from 90 to 100 %, with an average transferability ratio of 95.3 % (Table S3).

Evaluation of the genetic relationship within 56 genotypes using EST-SSR profiles

To evaluate the usefulness of 30 newly developed EST-SSRs, a genetic relationship study was performed in 56 accessions. A NJ tree was constructed based on the EST-SSR data (Fig. 2), in which the accessions in section Paeon (Table 2, 9-56) were distributed at the extreme of the dendrogram and were used as an outgroup. The P. lactiflora and cultivars of herbaceous peony first formed an independent cluster with a 92 % bootstrap value and were subsequently grouped together with P. veitchii with a 56 % bootstrap value. The wild species P. delavayi var. lutea and P. ludlowii with fleshy floral disks formed a separate clade from the other accessions examined in this study, with a bootstrap of 81 %. However, the remaining 34 accessions of tree peonies and 12 itoh hybrids formed another clade, and all their floral disks were leathery.

Next, thirty-four accessions of tree peonies and 12 itoh hybrids were primarily divided into two clusters. In cluster I, seven Rockii hybrid cultivars were first distributed in a separate branch with a 92 % bootstrap

 Table 2
 Abbreviation and origin of 56 accessions used in this study

Code	Accession	Abbreviation	Species or cultivar types/origin	
1	P. delavayi var. lutea	del	Species/Yunnan of China	
2	P. ludlowii	lud	Species/Yunnan of China	
3	P. jishanensis	jis	Species/Shanxi of China	
4	P. qiui	qiu	Species/Hubei of China	
5	P. rockii	roc	Species/Hubei of China	
6	P. decomposita	dec	Species/Sichuan of China	
7	P. ostii 1	ost1	Species/Henan of China	
8	P. ostii 2	ost2	Species//Henan of China	
9	Lan Bao Shi	suf1	Suffruticosa hybrid <sup>a</sup> /Luoyang of China	
10	Yao Huang	suf2	Suffruticosa hybrid <sup>a</sup> /Luoyang of China	
11	Shou An Hong	suf3	Suffruticosa hybrid <sup>a</sup> /Luoyang of China	
12	Cai Hui	suf4	Suffruticosa hybrid <sup>a</sup> /Luoyang of China	
13	Lu He Hong	suf5	Suffruticosa hybrid <sup>a</sup> /Luoyang of China	
14	Yin Hong Lv Bo	suf6	Suffruticosa hybrid <sup>a</sup> /Luoyang of China	
15	Kaoh	suf7	Suffruticosa hybrid <sup>a</sup> /Japan	
16	Kokuryu Nishiki	suf8	Suffruticosa hybrid <sup>a</sup> /Japan	
17	Yaezakura	suf9	Suffruticosa hybrid <sup>a</sup> /Japan	
18	Shima Nishiki	suf10	Suffruticosa hybrid <sup>a</sup> /Japan	
19	Kamata Nishiki	suf11	Suffruticosa hybrid <sup>a</sup> /Japan	
20	Qing Xin Bai	roc1	Rockii hybrid <sup>b</sup> /Gansu of China	
21	Gao Yuan Sheng Huo	roc2	Rockii hybrid <sup>b</sup> /Gansu of China	
22	Hong Hai Yin Bo	roc3	Rockii hybrid <sup>b</sup> /Gansu of China	
23	Ye Guang Bei	roc4	Rockii hybrid <sup>b</sup> /Gansu of China	
24	Hong Lou Cang Jiao	roc5	Rockii hybrid <sup>b</sup> /Gansu of China	
25	Han Hai Bing Xin	roc6	Rockii hybrid <sup>b</sup> /Gansu of China	
26	Hong Zhuang Su Guo	roc7	Rockii hybrid <sup>b</sup> /Gansu of China	
27	High Noon	lem1	Lemoinei hybrid <sup>e</sup> /America	
28	Huang Guan	lem2	Lemoinei hybrid <sup>c</sup> /Japan	
29	Souvenir de Maxime Cornu	lem3	Lemoinei hybrid <sup>c</sup> /France	
30	Renown	lem4	Lemoinei hybrid <sup>c</sup> /America	
31	Golden Era	lem5	Lemoinei hybrid <sup>c</sup> /America	
32	Chromatella	lem6	Lemoinei hybrid <sup>c</sup> /Australia	
33	Rosina	lem7	Lemoinei hybrid <sup>c</sup> /Australia	
34	Zephyrus	lem8	Lemoinei hybrid <sup>c</sup> /Australia	
35	Adelphi	lem9	Lemoinei hybrid <sup>c</sup> /Australia	
36	Iowa	lem10	Lemoinei hybrid <sup>c</sup> /Australia	
37	Callie's Memory	ito1	Itoh hybrid <sup>d</sup> /America	
38	Cora Louise	ito2	Itoh hybrid <sup>d</sup> /America	
39	First Arrival	ito3	Itoh hybrid <sup>d</sup> /America	
40	Lemon Dream	ito4	Itoh hybrid <sup>d</sup> /America	
41	Border Charm	ito5	Itoh hybrid <sup>d</sup> /America	
42	Morning Lilac	ito6	Itoh hybrid <sup>d</sup> /America	
43	Hillary	ito7	Itoh hybrid <sup>d</sup> /America	
44	Bartzella	ito8	Itoh hybrid <sup>d</sup> /America	

Code Accession		Abbreviation	Species or cultivar types/origin	
45	Lafayette Escadrille	ito9	Itoh hybrid <sup>d</sup> /America	
46	Viking Full Moon	ito10	Itoh hybrid <sup>d</sup> /America	
47	Copper Kettle	ito11	Itoh hybrid <sup>d</sup> /America	
48	Garden Treasure	ito12	Itoh hybrid <sup>d</sup> /America	
49	P. veitchii	vei	Species/Gansu of China	
50	P. lactiflora	lac1	Species/Chifeng of China	
51	Martha Washington	lac2	Lactiflora cultivar <sup>e</sup> /America	
52	Fen Yu Nu	lac3	Lactiflora cultivar <sup>e</sup> /Luoyang of China	
53	Huang Jin Lun	lac4	Lactiflora cultivar <sup>e</sup> /Luoyang of China	
54	Zi Feng Chao Yang	lac5	Lactiflora cultivar <sup>e</sup> /Luoyang of China	
55	Zi Ling	lac6	Lactiflora cultivar <sup>e</sup> /Luoyang of China	
56	Red Charm	lac7	Hybrid peonies <sup>f</sup> /America	

 Table 2
 continued

<sup>a</sup> All groups under *P. suffruticosa* composed of almost eight groups (Cheng 2007)

<sup>b</sup> Cultivars using mainly germpasm of *P. rockii* 

<sup>c</sup> Cultivars from intra-sectional hybridization between *P. delavayii* and *P. suffruticosa* 

<sup>d</sup> Cultivars from intersectional hybridization between *P. lactiflora* and Lemoinei hybrid

<sup>e</sup> The cultivars of *P. lactiflora* 

<sup>f</sup> A hybrid of the herbaceous peony was produced by crossing a variety of one species with a variety of a different species

value and then grouped together with P. rockii and finally grouped together P. decomposita with a 50 % bootstrap value. Cluster II in the NJ tree consisted of all of the cultivar accessions of the Suffruticosa hybrid, Lemoinei hybrid, Itoh hybrid and two P. ostii accessions, which can be explained based on their known origins and previous studies. Cluster II was later separated into two subclusters. In subcluster I, 13 samples fell into one large branch and were concentrated on two small groups with 84 % and 54 % bootstrap values. Six cultivars belonging to the P. suffruticosa Zhongyuan group were distributed in one group. Five cultivars belonging to the P. suffruticosa Japan group and two P. ostii samples comprised another group. In subcluster II, 'Souvenir de Maxime Cornu' formed a single branch, and other Lemoinei hybrids except for 'Chromatella' formed A, and all Itoh hybrids except for 'Lafayette Escadrille' formed B. These results indicated that the germplasms in each classified groups exhibited a great consistency with their origins.

# Discussion

Next-generation sequencing (NGS) technologies are becoming increasingly inexpensive and can provide

high-throughput data, and enormous quantities of sequencing data can now be generated exponentially and are publicly available. Transcriptome sequencing has been demonstrated to be an effective method in obtaining EST sequences, which are essential for mining a large number of functional SSR markers and in the identification of novel genes. Currently, a large number of EST-SSR markers, based on transcriptome sequences, have been developed in diverse species, such as vetch (Chung et al. 2014), rubber tree (Li et al. 2012b), linseed (Kale et al. 2012), and pigeonpea (Dutta et al. 2011). The development of these genebased SSR markers has directly or indirectly accelerated progress in MAS breeding (Zhai et al. 2014; Iorizzo et al. 2011).

The proportion of ESTs containing SSRs vary in different plant species. A survey on dicotyledonous species revealed that this ratio ranged from 2.65 to 16.82 % (Kumpatla and Mukhopadhyay 2005). In this study, approximately 6.4 % of the transcriptome sequences possessed at least one SSR, which was similar to reports for alfalfa (6.1 %) (Wang et al. 2013), bermudagrass (6.4 %) (Tan et al. 2012), and orchardgrass (7 %) (Bushman et al. 2011). Moreover, the frequency of the occurrence for EST-SSRs was one SSR in every 9.24 kb, which was much higher

**Table 3** Polymorphism information of 30 EST-SSR markers in 36 tree peony accessions, including the following parameters: the number of alleles  $(N_a)$ , observed and expected heterozygosities  $(H_o \text{ and } H_e)$ , PIC, and Shannon index

Primer name	Na	Ho	He	PIC	Shannon index
PS002	8	0.22	0.80	0.76	1.75
PS004	11	0.78	0.87	0.85	2.14
PS026	7	0.58	0.72	0.66	1.43
PS034	9	0.36	0.78	0.75	1.73
PS047	5	0.61	0.65	0.59	1.21
PS052	12	0.56	0.84	0.80	2.02
PS074	10	0.53	0.79	0.76	1.75
PS076	7	0.58	0.76	0.71	1.55
PS094	3	0.42	0.67	0.59	1.10
PS095	10	0.44	0.85	0.81	1.98
PS099	10	0.19	0.84	0.81	1.95
PS102	7	0.53	0.72	0.67	1.52
PS106	6	0.31	0.66	0.62	1.36
PS111	8	0.61	0.76	0.72	1.66
PS119	6	0.39	0.53	0.49	1.08
PS134	5	0.56	0.70	0.63	1.30
PS139	6	0.58	0.78	0.74	1.61
PS149	6	0.42	0.70	0.63	1.38
PS150	7	0.61	0.69	0.62	1.38
PS153	10	0.72	0.86	0.82	2.01
PS158	9	0.75	0.76	0.73	1.73
PS166	9	0.50	0.73	0.68	1.56
PS169	9	0.81	0.84	0.81	1.89
PS184	6	0.47	0.68	0.61	1.32
PS205	8	0.44	0.82	0.78	1.80
PS219	3	0.47	0.64	0.56	1.05
PS244	10	0.53	0.83	0.80	1.90
PS265	4	0.58	0.67	0.54	1.19
PS273	7	0.67	0.80	0.76	1.68
PS290	5	0.33	0.43	0.36	0.83
Mean	7.4	0.52	0.74	0.69	1.56

compared to alfalfa (1/12.06 kb) (Liu et al. 2013a), pineapple (1/13 kb) (Ong et al. 2011), and lotus (1/ 13.04 kb) (Pan et al. 2010). However, this value was lower compared to levant cotton (1/2.4 kb) (Jena et al. 2012), castor bean (1/1.77 kb) (Qiu et al. 2010), and gerbera (1/5.6 kb) (Gong and Deng 2010). These results indicated that the frequency of SSRs in EST sequences was highly variable with plant species, and this difference, to some extent, may be attributed to the size of the data set, SSR search criteria, and mining tool used for the SSR search (Varshney et al. 2005).

Until recently, both di- and tri-nucleotide repeats have been found to be predominantly repeat types in many plants, whereas the dominant repeat motifs were usually different (Varshney et al. 2005). In this study, di-nucleotide repeats were the most abundant type and were followed by tri-nucleotides, which were consistent with previous reports on tree peony (Shu et al. 2009; Gai et al. 2012) and other plants, such as pear (Zhang et al. 2014) and tea (Sharma et al. 2009). A potential explanation for the high ratio of di-nucleotide SSR loci in this study was that unigene sequences contained more information representing UTRs. Furthermore, we found that the shorter motifs appeared more frequently compared to longer motifs, except for hexa-nucleotides, which were more frequent than tetra- and penta-nucleotides in the transcriptome. As shown in Fig. 1, the most dominant di-nucleotide repeat motif was AG/CT, followed by GA/TC, and the same results were found in radish (Zhai et al. 2014) and sweet potato (Wang et al. 2011). The most prevalence tri-nucleotide motifs were CCA/TGG and GAA/TTC, which was consistent with the recent results obtained in tree peony (Gai et al. 2012), pineapple (Ong et al. 2011), and rubber tree (Feng et al. 2009). Furthermore, a common feature in most dicotyledonous plants was the rarity of GC/CG in dinucleotide motifs. There were no GC/CG repeat motifs observed in common bean (Chen et al. 2014) and alfalfa (Wang et al. 2013). However, the GC/CG repeat motif was determined in only 0.4 % of the dinucleotide repeats in this study, which was consistent with the results obtained in herb epimedii (Zeng et al. 2010) and coffee (Aggarwal et al. 2007).

In this study, 47.3 % of the EST-SSR primers amplified the target genomic DNA, which was lower than the results obtained in Gai's report in tree peony (Gai et al. 2012) (81.2 %), but higher than the results obtained in Wang's report in tree peony (Wang et al. 2009) (40.9 %). Such results indicated that the amplification efficiency of EST-SSRs was much higher than that of genomic SSRs, which may be due to the relatively highly conserved transcribed regions in the primer sequences. Compared to other plants, the success amplification rate is comparable to sugarcane, where 48.5 % of the primers amplified (Cordeiro et al. 2001), but higher than alfalfa (30 %) (Wang et al. 2013) and lower than rubber tree (59.8 %) (Feng et al. **Fig. 2** The NJ tree of 56 accessions was constructed using the PowerMarker version 3.25 based on the data of 30 EST-SSR markers. Bootstrap values over 50 % were displayed above the branch; based on 1,000 resamplings of the data set. The information of the abbreviations represented in the tree can be seen in Table 2



2009). The success amplification rate is relatively low in this study which may be due to the presence of introns, alternate splicing, or sequencing splicing technology. To the best of our knowledge, very few research has been performed on the development of so many EST-SSR markers in tree peony in a single study to date. In this study, we obtained 149 polymorphic EST-SSR markers from 373 valid EST-SSRs in the parents, *P. ostii* 'Feng Dan' and *P. suffruticosa* 'Hong Qiao', of our mapping population with a polymorphic ratio of 39.9 %, thereby providing a powerful tool for the construction of an SSR-based linkage map in further studies. Compared to other plants, the polymorphic ratio of EST-SSRs in this study was at the medium level (Varshney et al. 2005). The number of samples and samples obtained from a different geographic origin may result in a different ratio of polymorphism. For example, Wang et al. (2011) found 816 primers yielding amplification products, of which 195 (23.9 %) and 342 (41.9 %) primers exhibited polymorphism between the two mapping parents and among the eight cultivated sweet potatoes, respectively. Thus, if more samples were screened in our study, the number of polymorphic loci would most likely increase.

As previously described, di-, tetra-, and pentanucleotide repeats were primarily associated with the UTR, while tri- and hexa-nucleotide repeats primarily occurred within the CDS. Approximately 76.7 % of the SSRs that were located in the CDS were tri- and hexa-nucleotide repeats. This appeared to be a common feature of EST-SSRs and has often been found in other organisms (Gupta et al. 2014; Morgante et al. 2002). This may result from selection and evolution because tri- and hexa-nucleotide SSRs are thought to be required in the coding region to maintain the reading frame, while di-, tetra-, and penta-nucleotide SSRs can easily change the reading frame within coding regions and cause a negative mutation when SSR length variation occurs. Our study also revealed that the proportion of polymorphic SSR markers was considerably higher in the UTR compared to the CDS, which was consistent with previous reports in pigeonpea (Dutta et al. 2011) and castor bean (Qiu et al. 2010). The polymorphism detected by EST-SSRs from different EST regions (3' UTR, 5' UTR, and CDS) varied across the different taxonomic levels. In general, SSRs that were from the 3' UTR were most polymorphic at the cultivar level, 5' UTR SSRs were most polymorphic between cultivars and species, and CDS-SSRs tend to differentiate at higher taxonomic levels (Scott et al. 2000). This may be attributed to less tolerance for mutations in the CDS region compared to the UTR.

The PIC value of each EST-SSR, which reflects allelic diversity and frequencies among the sampled individuals, was used to assess the informativeness level of the markers and were categorised as high (PIC > 0.5), moderate (0.5 < PIC < 0.25), and low (PIC < 0.25) (Bostein et al. 1980). In this study, 28 out of 30 primers exhibited a high PIC value, which indicated good informativeness as a marker. In general, the EST-SSR markers revealed a lower level of polymorphism compared to the genomic SSR markers. However, in this study, it was higher compared to previous reports (Yu et al. 2013; Zhang et al. 2011). Some potential reasons include (1) employment of highly polymorphic EST-SSR markers for diversity assessment; and (2) use of a diverse set of genotypes derived from intra- and inter-specific hybridisation and wild species.

Located in transcript regions, EST-SSRs display a high degree of transferability to related species. Ellis and Burke (2007) summarised the transferability of EST-SSR among plant taxa and demonstrated a variation range of EST-SSR cross-genera transferability from 10 to 90 %. Our result showed that the EST-SSR identified in *P. suffruticosa* had a high transfer rate (95.3 %) in other species of Paeonia. A high transferability of EST-SSR was also reported in other plants, such as eucalyptus (Zhou et al. 2014) and citrus (Luro et al. 2008). This high success rate indicated that different species in the genus Paeonia may be evolutionarily closely related. Thus, these novel EST-SSR markers may be used as anchored markers for future comparative mapping and evolutionary studies in the Paeoniaceae family. The correlation between the index of transferability and PIC values of the SSR markers was not observed in this study.

In recent years, SSRs have been extensively applied in genetic diversity analysis, genetic relationship analysis, linkage mapping and association mapping studies (Wang et al. 2013; Liu et al. 2013b; Agrama et al. 2007). In this study, the validity of the developed EST-SSR markers were investigated for genetic relationships in the collected Paeonia germplasms of 56 accessions. Cluster analysis in the NJ tree demonstrated that major clusters corresponded to known pedigree trees. In the NJ tree, P. rockii and its seven cultivars group were closely relative to P. decomposita, which was consistent with recent molecular markers studies (Zhao et al. 2008; Yu et al. 2013). To the best of our knowledge, the ancestors of the Japanese cultivars were from China, particularly from the Zhongyuan groups. This close relationship between the Japanese and Chinese cultivars is supported by our clustering pattern. P. ostii is thought to contribute to the formation of the Suffruticosa hybrid (Cheng 2007). Indeed, it shares more genetic similarity and is distributed as a sister clade among the cultivars of the Japan group in this study. Subcluster II in the NJ tree consisted of all of the cultivar accessions of Lemoinei hybrid and Itoh hybrid. 'Souvenir de Maxime Cornu' formed a single branch, and other Lemoinei hybrids except for 'Chromatella' formed A, and all Itoh hybrids except for 'Lafayette Escadrille' formed B. 'Souvenir de Maxime Cornu' from France were the crossbred offspring of the P. delavayii and P.

suffruticosa Zhongyuan Group, whereas the other nine Lemoinei hybrids originated from similar crosses, but pollen was collected from the P. suffruticosa Japan Group (Cheng 2007; Wister 1995). Itoh hybrids in this study were the result of the intersectional hybridisation between P. lactiflora and the Lemoinei hybrid. We also found that there was some incongruence in a few branches. For example, the Itoh hybrid 'Lafayette Escadrille' and Lemoinei hybrids 'Chromatella' formed a branch. The incongruence between the dendrogram and the division of cultivars might be due to the limited number of samples. Clustering results also indicated that the EST-SSR markers were an informative and effective tool for the analysis of genetic relationships among a wide range of diverse accessions and exhibited promise for the development of additional molecular markers, which could reliably determine the relative genetic contributions of specific populations in existing and newly developed cultivars.

In conclusion, using transcriptome sequences to identify EST-SSR markers is a convenient and costeffective approach in tree peony. A large set of EST-SSR markers identified in this study has remarkably increased the molecular marker repository for tree peony. With its high polymorphism and transferability, these novel EST-SSR markers will represent a powerful molecular tool for germplasm identification, genetic diversity analyses, genetic relationship studies, and comparative mapping in tree peony and relative species. Moreover, these markers may also be valuable for the construction of genetic linkage maps, which are a prerequisite for the generation of markertrait relationships and the application of MAS in the future improvement of this important flower crop.

Acknowledgments We deeply thank Gaixiu Liu for her help during the sample collection. This study was funded by the National High Technology Research and Development Program of China (863 Program, 2011AA100207) and the National Science and Technology Support Program of China (2012BAD01B07).

# References

- Agarwal M, Shrivastava N, Padh H (2008) Advances in molecular marker techniques and their applications in plant sciences. Plant Cell Rep 27:617–631
- Aggarwal RK, Hendre PS, Varshney RK, Bhat PR, Singh L, Krishnakumar V (2007) Identification, characterization

and utilization of EST-derived genic microsatellite markers for genome analyses of coffee and related species. Theor Appl Genet 114(2):359–372

- Agrama HA, Eizenga GC, Yan W (2007) Association mapping of yield and its components in rice cultivars. Mol Breed 19(4):341–356
- Bostein D, White RL, Sholnick M, David RW (1980) Construction of a genetic linkage map in man using restriction fragment length polymorphism. Am J Hum Genet 32:314–331
- Bouck AMY, Vision T (2007) The molecular ecologist's guide to expressed sequence tags. Mol Ecol 16(5):907–924
- Bushman BS, Larson SR, Tuna M et al (2011) Orchardgrass (*Dactylis glomerata* L.) EST and SSR marker development, annotation, and transferability. Theo Appl Genet 123(1):119–129
- Chen M, Wu J, Wang L et al (2014) Development of mapped simple sequence repeat markers from common bean (*Phaseolus vulgaris* L.) based on genome sequences of a Chinese landrace and diversity evaluation. Mol Breed 33:489–496
- Cheng FY (2007) Advances in the breeding of tree peonies and a cultivar system for the cultivar group. Int J Plant Breed 1(2):89–104
- Chung JW, Kim TS, Sundan S et al (2014) New cDNA-SSR markers in the narrow-leaved vetch (*Vicia sativa* subsp. *nigra*) using 454 pyrosequencing. Mol Breed 33:749–754
- Cordeiro GM, Casu R, McIntyre CL, Manners JM, Henry RJ (2001) Microsatellite markers from sugarcane (*Saccharum* spp.) ESTs cross transferable to *Erianthus* and *Sorghum*. Plant Sci 160:1115–1123
- Dutta S, Kumawat G, Singh BP et al (2011) Development of genic-SSR markers by deep transcriptome sequencing in pigeonpea [*Cajanus cajan* (L.) Millspaugh]. BMC Plant Biol 11(1):17
- Ellis J, Burke J (2007) EST-SSRs as a resource for population genetic analyses. Heredity 99(2):125–132
- Eujayl I, Sledge MK, Wang L et al (2004) Medicago truncatula EST-SSRs reveal cross-species genetic markers for Medicago spp. Theor Appl Genet 108(3):414–422
- Feng SP, Li WG, Huang HS, Wang JY, Wu YT (2009) Development, characterization and cross-species/genera transferability of EST-SSR markers for rubber tree (*Hevea brasiliensis*). Mol Breed 23(1):85–97
- Gai SP, Zhang YX, Mu P et al (2012) Transcriptome analysis of tree peony during chilling requirement fulfillment: assembling, annotation and markers discovering. Gene 497(2):256–262
- Gao Z, Wu J, Wang L et al (2013) Rapid microsatellite development for tree peony and its implications. BMC Genomics 14(1):886
- Gilmore B, Bassil N, Nyberg A et al (2013) Microsatellite marker development in peony using next generation sequencing. J Am Soc Hortic Sci 138(1):64–74
- Gong L, Deng Z (2010) EST-SSR markers for gerbera (*Gerbera hybrida*). Mol Breed 26(1):125–132
- Gupta SK, Bansal R, Gopalakrishna T (2014) Development and characterization of genic SSR markers for mungbean (*Vigna radiata* (L.) Wilczek). Euphytica 195(2):245–258
- Homolka A, Berenyi M, Burg K, Kopecky D, Fluch S (2010) Microsatellite markers in the tree peony, *Paeonia suffruticosa* (Paeoniaceae). Am J Bot 97(6):e42–e44

- Hou XG, Guo DL, Cheng SP, Zhang JY (2011a) Development of thirty new polymorphic microsatellite primers for *Paeonia suffruticosa*. Biol Plant 55(4):708–710
- Hou XG, Guo DL, Wang J (2011b) Development and characterization of EST-SSR markers in *Paeonia suffruticosa* (Paeoniaceae). Am J Bot 11:e303–e305
- Iorizzo M, Senalik DA, Grzebelus D et al (2011) De novo assembly and characterization of the carrot transcriptome reveals novel genes, new markers, and genetic diversity. BMC Genomics 12:389
- Jena SN, Srivastava A, Rai KM et al (2012) Development and characterization of genomic and expressed SSRs for levant cotton (*Gossypium herbaceum* L.). Theor Appl Genet 124(3):565–576
- Kale SM, Pardeshi VC, Kadoo NY et al (2012) Development of genomic simple sequence repeat markers for linseed using next-generation sequencing technology. Mol Breed 30: 597–606
- Kumpatla SP, Mukhopadhyay S (2005) Mining and survey of simple sequence repeats in expressed sequence tags of dicotyledonous species. Genome 48:985–998
- Li L, Cheng F, Zhang Q (2011) Microsatellite markers for the Chinese herbaceous peony *Paeonia lactiflora* (Paeoniaceae). Am J Bot 98(2):e16–e18
- Li K, Zhou N, Li HY (2012a) Composition and function research of peony flowers and peony seeds. Food Res Dev 33(3):228–230
- Li DJ, Deng Z, Qin B, Liu XH, Men ZH (2012b) De novo assembly and characterization of bark transcriptome using Illumina sequencing and development of EST-SSR markers in rubber tree (*Hevea brasiliensis* Muell. Arg.). BMC Genomics 13(1):192
- Liu K, Muse SV (2005) PowerMarker: an integrated analysis environment for genetic marker analysis. Bioinformatics 21(9):2128–2129
- Liu Z, Chen T, Ma L et al (2013a) Global transcriptome sequencing using the Illumina platform and the development of EST-SSR markers in autotetraploid alfalfa. PLoS One 8(12):e83549
- Liu L, Huang Y, Punnuri S, Samuels T, Wu Y, Mahalingam R (2013b) Development and integration of EST-SSR markers into an established linkage map in switchgrass. Mol Breed 32(4):923–931
- Luro FL, Costantino G, Terol J et al (2008) Transferability of the EST-SSRs developed on Nules clementine (*Citrus clementina* Hort ex Tan) to other *Citrus* species and their effectiveness for genetic mapping. BMC Genomics 9:287
- Morgante M, Hanafey M, Powell W (2002) Microsatellites are preferentially associated with nonrepetitive DNA in plant genomes. Nat Genet 30(2):194–200
- Ong WD, Voo CLY, Kumar SV (2011) Development of ESTs and data mining of pineapple EST-SSRs. Mol Biol Rep 39(5):5889–5896
- Pan L, Xia Q, Quan Z, Liu H, Ke W, Ding Y (2010) Development of novel EST-SSRs from sacred lotus (*Nelumbo nucifera* Gaertn) and their utilization for the genetic diversity analysis of *N. nucifera*. J Hered 101(1):71–82
- Paux E, Sourdille P, Mackay I, Feuillet C (2012) Sequencebased marker development in wheat: advances and applications to breeding. Biotechnol Adv 30(5):1071–1088

- Qiu LJ, Yang C, Tian B, Yang JB, Liu AH (2010) Exploiting EST databases for the development and characterization of EST-SSR markers in castor bean (*Ricinus communis* L.). BMC Plant Biol 10:278
- Scott KD, Eggler P, Seaton G et al (2000) Analysis of SSRs derived from grape ESTs. Theor Appl Genet 100:723–726
- Sharma RK, Bhardwaj P, Negi R, Mohapatra T, Ahuja PS (2009) Identification, characterization and utilization of unigene derived microsatellite markers in tea (*Camellia* sinensis L.). BMC Plant Biol 9:53
- Shu QY, Wischnitzki E, Liu ZA et al (2009) Functional annotation of expressed sequence tags as a tool to understand the molecular mechanism controlling flower bud development in tree peony. Physiol Plant 135(4):436–449
- Stern FC (1946) A study of the genus *Paeonia*. Royal Horticulture Society, London
- Sun J, Yuan J, Wang B et al (2011) Development and characterization of 10 microsatellite loci in *Paeonia lactiflora* (Paeoniaceae). Am J Bot 98(9):e242–e243
- Tan C, Wu Y, Taliaferro CM, Anderson MP, Tauer C, Samuels T (2012) Development of simple sequence repeat markers for bermudagrass from its expressed sequence tag sequences and preexisting sorghum SSR markers. Mol Breed 29(1):23–30
- Temnykh S, DeClerck G, Lukashova A, Lipovich L, Cartinhour S, McCouch S (2001) Computational and experimental analysis of microsatellites in rice (*Oryza sativa* L.): frequency, length variation, transposon associations, and genetic marker potential. Genome Res 11(8):1441–1452
- Varshney RK, Graner A, Sorrells ME (2005) Genic microsatellite markers in plants: features and applications. Trends Biotechnol 23:48–55
- Wang JX, Xia T, Zhang JM, Zhou SL (2009) Isolation and characterization of fourteen microsatellites from a tree peony (*Paeonia suffruticosa*). Conserv Genet 10(4):1029–1031
- Wang Z, Li J, Luo Z et al (2011) Characterization and development of EST-derived SSR markers in cultivated sweetpotato (*Ipomoea batatas*). BMC Plant Biol 11(1):139
- Wang Z, Yan H, Fu X, Li X, Gao H (2013) Development of simple sequence repeat markers and diversity analysis in alfalfa (*Medicago sativa* L.). Mol Biol Rep 40(4): 3291–3298
- Wister JC (1995) The Peonies. American Peony Society, Hopkins
- Yeh FC, Yang RC, Boyle T et al (1999) POPGENE, version 1.32: the user friendly software for population genetic analysis. Molecular Biology and Biotechnology Centre, University of Alberta, Edmonton, AB, Canada
- Yu HP, Cheng FY, Zhong Y, Cai CF, Wu J, Cui HL (2013) Development of simple sequence repeat (SSR) markers from *Paeonia ostii* to study the genetic relationships among tree peonies (Paeoniaceae). Sci Hortic 164:58–64
- Yuan JH, Cheng FY, Zhou SL (2010) Hybrid origin of *Paeonia × yananensis* revealed by microsatellite markers, chloroplast gene sequences, and morphological characteristic. Int J Plant Sci 171(4):409–420
- Yuan JH, Cheng FY, Zhou SL (2012) Genetic structure of the tree Peony (*Paeonia rockii*) and the Qinling mountains as a geographic barrier driving the fragmentation of a large population. PLoS One 7(4):e34955

- Yuan JH, Cornille A, Giraud T, Cheng FY, Hu YH (2014) Independent domestications of cultivated tree peonies from different wild peony species. Mol Ecol 23(1):82–95
- Zeng SH, Xiao G, Guo J et al (2010) Development of a EST dataset and characterization of EST-SSRs in a traditional Chinese medicinal plant, *Epimedium sagittatum* (Sieb. Et Zucc.) Maxim. BMC Genomics 11:94–105
- Zhai L, Xu L, Wang Y et al (2014) Novel and useful genic-SSR markers from de novo transcriptome sequencing of radish (*Raphanus sativus* L.). Mol Breed 33:749–754
- Zhang JM, Liu J, Sun HL, Yu J, Wang JX, Zhou SL (2011) Nuclear and chloroplast SSR markers in *Paeonia delavayi* (Paeoniaceae) and cross-species amplification in *P. lud-lowii*. Am J Bot 98(12):e346–e348
- Zhang JJ, Shu QY, Liu ZA, Ren HX, Wang LS, De Keyser E (2012) Two EST-derived marker systems for cultivar identification in tree peony. Plant Cell Rep 31(2):299–310

- Zhang MY, Fan L, Liu QZ et al (2014) A novel set of ESTderived SSR markers for pear and cross-species transferability in Rosaceae. Plant Mol Biol Rep 32:290–302
- Zhao X, Zhou ZQ, Lin QB, Pan KY, Li MY (2008) Phylogenetic analysis of *Paeonia* sect. *Moutan* (Paeoniaceae) based on multiple DNA fragments and morphological data. Acta Phytotax Sin 46(4):563–572
- Zhou HM, Ma JQ, Miao CY et al (2009) Physicochemical indexes and fatty acid composition of peony seed oil. China Oils Fats 34(7):72–74
- Zhou H, Cheng FY, Wang R, Zhong Y, He CY (2013) Transcriptome comparison reveals key candidate genes responsible for the unusual reblooming trait in tree peonies. PLoS One 8(11):e79996
- Zhou CP, He XD, Li FG et al (2014) Development of 240 novel EST-SSRs in *Eucalyptus* L'Hérit. Mol Breed 33(1): 221–225