#### **ORIGINAL ARTICLE**



# **Multicomponent reaction for synthesis, molecular docking, and anti‑infammatory evaluation of novel indole‑thiazole hybrid derivatives**

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#### **Abstract**

In this article, novel thiazol-indolin-2-one derivatives **4a–f** have been synthesized via treatment of thiosemicarbazide (**1**) with some isatin derivative **2a–f** and *N*-(4-(2-bromoacetyl)phenyl)-4-tolyl-sulfonamide (**3)** under refux in ethanol in the presence of triethyl amine (TEA). The structures of new products were elucidated by elemental and spectral analyses. Moreover, all compounds were investigated for their in vivo anti-infammatory activity using celecoxib as a reference drug. The target compound **4b** was the most active anti-inflammatory candidate and exhibited higher edema inhibition ( $EI = 38.50\%$ ) than that recorded by celecoxib (EI=34.58%) after 3 h. Furthermore, the most active compounds **4b** and **4f** were subjected to a molecular docking study inside COX-2 enzyme to show their binding interactions**.** Both compounds **4b** and **4f** showed good fitting into COX-2 binding site with docking energy scores − 11.45 kcal/mol and − 10.48 kcal/mol, respectively which indicated that compound **4b** revealed the most promising and efective anti-infammatory potential.

#### **Graphical abstract**



**Keywords** Indole derivatives · Thiazole · Multicomponent reaction · Anti-infammatory activity · Histopathological examination

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### **Introduction**

Infammation is a biological reaction to a disturbance in tissue homeostasis and body defense chemicals in which cells penetrate the affected tissue causing increasing blood flow, vascular permeability, and vasodilatation  $[1, 1]$  $[1, 1]$ [2](#page-9-1)]. Non-steroidal anti-infammatory drugs (NSAIDs) are the most commonly used medications for relieving pain and infammation by inhibiting Cyclooxygenase (COX) enzymes [\[3,](#page-9-2) [4\]](#page-9-3). The constitutive COX-1 performs numerous physiological activities as protecting gastric mucosa, vascular homeostasis and platelet aggregation, while the other isoform, the inducible COX-2 is concerned with prostaglandins that promote infammation and modulate pain [[5](#page-9-4)[–7\]](#page-10-0).

The use of traditional NSAIDs as aspirin, indomethacin and phenazone causes gastrointestinal side efects due to the inhibition of both COX isoforms  $[8-10]$  $[8-10]$  $[8-10]$ . Selective COX-2 inhibitor medications as celecoxib, valdecoxib, and rofecoxib have been prepared to avoid the side efects produced by traditional NSAIDs [[11](#page-10-3), [12](#page-10-4)]. Unfortunately, rofecoxib and valdecoxib were taken off the market due to their cardiovascular side efects including myocardial infarction and the occurrence of high blood pressure [[13–](#page-10-5)[15\]](#page-10-6). So, there is a great demand for selective COX-2 inhibitors with diminished side efects. Indole is one of the most widely used scafolds in a broad range of anti-infammatory agents [[16](#page-10-7)[–18\]](#page-10-8). Many research investigations have focused on indole-based NSAIDs such as indomethacin (**I**) to enhance their COX-2 selectivity and decrease the ulcerogenic adverse efects that linked to their strong COX-1 selectivity and drugs acidic properties [[19–](#page-10-9)[21](#page-10-10)]. Knaus and co-workers synthesized a new set of indole derivatives substituted at N-1 and C-3 [[22](#page-10-11)]. From the prepared indole derivatives, compound **II** was the most selective  $(SI > 312)$  and potent (COX-2 IC<sub>50</sub> = 0.32 µM) COX-2 inhibitor. In 2021, new indole derivatives having thiosemicarbazone moiety were prepared and screened for their anti-infammatory efect using carrageenan-induced paw edema assay [[23](#page-10-12)]. Compound **III** recorded superior COX-2 selectivity (SI=23.06) than displayed by celecoxib  $(SI = 11.88)$ .

Thiazole is a fve-membered heterocyclic ring [[24](#page-10-13)[–28\]](#page-10-14) with many pharmacological utilities as anticancer [[29](#page-10-15), [30](#page-10-16)], antioxidant [[31](#page-10-17), [32\]](#page-10-18), antimicrobial [[33](#page-10-19), [34\]](#page-10-20), antidiabetic [\[35\]](#page-10-21), anthelmintic [[36\]](#page-10-22) and anti-inflammatory [[37,](#page-10-23) [38\]](#page-10-24). For example, the thiazole derivative **IV** signifcantly inhibited edema (60.82%) in carrageenan-induced edema compared with indomethacin (53.21%) [[39\]](#page-10-25). Also, thiazole derivative **V** recorded comparable edema inhibition  $(EI = 87%)$  to that registered by indomethacin ( $EI = 91\%$ ) after 4 h [[40](#page-10-26)].

Considering the aforesaid data and as an extension and development of our previous studies [\[41–](#page-10-27)[50](#page-11-0)], we present the design and construction of novel indole-thiazole hybrids and biologically screened for their anti-infammatory effect. This work aims to get new compounds with selective COX-2 inhibition, favorable anti-infammatory potential and minimized gastric side efects. This aim has been achieved by hybridization of the privileged indole ring with the thiazole nucleus in one chemical entity (Fig. [1\)](#page-1-0).

### **Result and discussion**

### **Chemistry**

In this work, treatment of thiosemicarbazide (**1**) with isatin derivatives **2a–f** and 4-(bromoacetyl)-*N*-(4-methylphenyl) benzenesulfonamide (**3**) via three-component reaction under refux in ethanol/tiethylamine (TEA). Firstly, compound **1** was allowed to react with some isatin derivatives **2a–f** namely; isatin (**2a**), 5-chloro-isatin (**2b**), 5-nitro-isatin (**2c**), *N*-methyl-isatin (**2d**), 5-chloro-*N*-methyl-isatin (**2e**), 5-nitro-*N*-methyl-isatin (**2f**), then compound **3** was added until the



<span id="page-1-0"></span>**Fig. 1** Examples of some reported indoles **(I–III)** and thiazoles **(IV, V)** and the rationale for the design of target compounds **4a–f**



<span id="page-2-0"></span>**Scheme 1.** Synthesis of indole-thiazole hybrid derivatives **4a–f**

reaction completed (TLC), to obtain thiazol-indolin-2-one derivatives **4a–f** (Scheme [1](#page-2-0)).

The chemical structure of compounds **4a–f** was determined using diferent elemental and spectroscopic analyses such as  ${}^{1}$ H-NMR,  ${}^{13}$ CNMR, as well as infrared spectroscopy.

Their infrared spectra revealed the existence of new bands in the range 3363–3124  $cm^{-1}$  corresponding to NH groups. <sup>1</sup>H NMR spectra showed, as well as the aromatic signals, new singlet signals in the region δ 13.30–9.03 ppm for NH groups. The N-CH3 proton in compounds **4d–f** appeared as a singlet signal in the range δ 3.73–3.84 ppm, respectively. Moreover, their  $^{13}$ C NMR spectra matched the accurate chemical structure which showed the carbonyl groups

in the range 190.6–161.2 ppm and the N-CH<sub>3</sub> groups in compounds **4d**–**f** appeared in the region δ 34.5–30.6 ppm, and the CH<sub>3</sub> groups at range  $\delta$  22.4–21.3 ppm, respectively. Furthermore, elemental analyses of thiazoles **4a**–**f** provided the correct structure of the new products (cf. experimental).

#### **Biological screening**

#### **Anti‑infammatory activity**

The anti-inflammaory potential of indol-3-ylidenehydrazino-1,3-thiazole derivatives **4a–f** was estimated applying the carrageenan-induced rat paw edema method using

<span id="page-3-0"></span>**Table 1** Anti-infammatory potential of test compounds (4a-f) using celecoxib reference drug

Compound	Diameter inflammation (mm)			% Edema inhibition		
	1 <sub>h</sub>	3 <sub>h</sub>	5 h	1 <sub>h</sub>	3 h	5 h
Control	$3.90 + 0.14$	$3.90 + 0.14$	$3.90 + 0.14$			
Carrageenan	$5.76 \pm 0.08$	$7.20 + 0.08$	$7.91 + 0.15$			
Celecoxib	$4.09 + 0.06$	$4.71 + 0.18$	$4.01 + 0.14$	28.99	34.58	49.30
4a	$5.70 + 0.14$	$5.90 + 0.12$	$6.90 + 0.15$	01.04	18.05	12.76
4b	$4.90 + 0.51$	$4.46 + 0.23$	$4.30 + 0.33$	12.72	38.50	45.63
4c	$5.53 + 0.08$	$5.52 + 0.09$	$4.81 + 0.23$	03.99	23.33	39.19
4d	$5.60 \pm 0.96$	$6.09 + 0.22$	$6.03 \pm 0.59$	02.77	15.41	23.76
4e	$5.74 \pm 0.23$	$6.79 + 0.19$	$7.22 + 0.23$	0.34	5.69	8.72
4f	$5.36 + 0.30$	$4.90 + 0.20$	$4.60 + 0.06$	6.94	31.94	41.84



<span id="page-3-1"></span>**Fig. 2** The % of edema inhibition in response to test compounds

celecoxib as a standard. Each target compound was taken immediately before infammation induction by carrageenan injection. The anti-infammatory potential was recorded according to paw volume changes after 1, 3 and 5 h as displayed in Table [1](#page-3-0). The obtained outcomes disclosed that 4-{2-[2-(5-chloro-2-oxo-1,2-dihydro-3*H*-indol-3-ylidene) hydrazino]-1,3-thiazol-4-yl}-*N*-phenyl-4-tolylsulfonamide (**4b**) was the most active candidate with edema inhibition percent equal to 12.72–45.63%. Furthermore, this compound **4b** showed higher edema inhibition  $(EI = 38.50%)$  than that exhibited by celecoxib ( $EI = 34.58\%$ ) after 3 h. Moreover, compound **4f** revealed comparable edema inhibition (3 h;  $EI = 31.94\%, 5 h$ ;  $EI = 41.84\%$ ) to that recorded by celecoxib  $(3 h; EI = 34.58\%, 5 h; EI = 49.30\%).$  In addition, within 1*H*-indole derivatives **4a–c**, the 5-chloroindole derivative (**4b**) was the most active anti-infammatory candidate (3 h;  $EI = 38.50\%, 5 \text{ h}; EI = 45.63\%)$  followed by 5-nitro analogue (**4c**) (3 h; EI=23.33%, 5 h; EI=39.19%) while compound **4a** with no substitution at positions 1 and 5 of indole moiety exhibited the least anti-infammatory activity (3 h;  $EI = 18.05\%, 5 \text{ h}; E = 12.76\%$ . In case of *N*-methylindole derivatives (**4d–f**), the 5-nitroindole candidate (**4f**) showed the highest edema inhibition percent  $(3 h; EI = 31.94\%, 5 h;$  $EI = 41.84\%$ ) followed by compound **4d** (3 h;  $EI = 15.41\%$ , 5 h; EI=23.76%) then the 5-chloro analogue (**4e**) (3 h;  $EI = 5.69\%, 5$  h;  $EI = 8.72\%$ ). From the recorded data in (Table [1](#page-3-0) and Fig. [2\)](#page-3-1), it is clear that compound **4b** has the



<span id="page-3-2"></span>**Fig. 3** Microscopic examination of the impact of 4b on paw tissues following carrageenan injection. **A** control group with normal paw tissue, **B** carrageenan group disclosed an acute infammation (black arrow) with edema (black star), **C** Celecoxib group with remarkable attenuation of edema and neutrophils migration, **D** compound **4b** treated group with signifcant attenuation of infammation and edema. Sections stained with H&E,  $(\times 200)$ 

most promising anti-infammatory potential in comparison with celecoxib.

#### **Histological investigation**

The impact of compound **4b** on paw tissue after carrageenan injection, comparing the results to those observed with indomethacin. It has been found that compound **4b** is the most efective of the tested products. As shown in Fig. [3](#page-3-2), the paw tissues of control rats (2, A) are not infamed. The carrageenan model demonstrated neutrophil migration in addition to acute infammation (black arrow) and hemorrhagic edema (black star) (Fig. [2B](#page-3-1)). Nevertheless, the rats given Celecoxib showed a notable decrease in infammation Fig. [2C](#page-3-1). Figure [2](#page-3-1)D demonstrated the impact of test chemical **4b** on paw tissue infammation, demonstrating a notable reduction in infammatory cells and edema.

Compounds	COX-1 IC <sub>50</sub> ( $\mu$ M)	COX-2 IC <sub>50</sub> ( $\mu$ M)	SI
4b	$21.97 \pm 1.56$	$8.26 + 0.77$	2.65
4f	$99.50 + 3.34$	$23.03 + 0.75$	4.32
Indomethacin	$0.52 + 0.06$	$0.77 + 0.041$	0.67
Celecoxib	$7.11 + 0.28$	$1.60 + 0.036$	4.44

<span id="page-4-0"></span>**Table 2** I*n vitro* inhibitory efects of compounds **4b** and **4f** on COX-1 and COX-2 enzymes

#### **In vitro COX inhibition screening**

Using a colorimetric enzyme immunoassay (EIA), the most active candidates **4b** and **4f** inhibitory efficacy against COX-1/COX-2 enzymes was ascertained. For each hybrid, the effectiveness was measured using serial dilutions to determine the concentration that inhibited 50% of the enzyme  $(IC_{50})$  (100, 10, 1, and 0.1 M). Additionally, for COX-1/COX-2, selectivity indexes (SI values) against COX-2 were computed and compared with reference drugs celecoxib and indomethacin.

The results obtained from the **4b** and **4f** hybrids are displayed in (Table [2\)](#page-4-0), which indicates that both compounds **4b** and **4f** exhibited higher COX-2 inhibitory efect than COX-1 enzyme.

Compound **4b** was the most potent COX-2 inhibitor  $(IC_{50} = 8.26 \mu M)$ , while **4f** was the most COX-2 selective  $(SI = 4.32)$  compared with indomethacin  $(COX - 2)$  $IC_{50} = 0.77$  µM,  $SI = 0.67$ ) and celecoxib (COX-2 IC<sub>50</sub> = 1.60  $\mu$ M, SI = 4.44). Furthermore, both **4b** (SI = 2.65) and  $4f(SI = 4.32)$  were more COX-2 selective than indomethacin  $(SI = 0.67)$ . In addition, the 5-nitroindole candidate (4f) exhibited comparable selectivity index  $(SI = 4.32)$  to that recorded by celecoxib  $(SI = 4.44)$ .

#### **Histological evaluation of ulcers**

Most NSAIDs have been found to induce stomach ulcers. Finding innovative anti-infammatory medicines that helps to avoid this problem is crucial because the unfavorable gastrointestinal adverse reactions of current anti-infammatory therapies is a major disadvantage Thus, to fnd any gastroprotective efects of the most potent anti-infammatory compounds **4b** and **4f**, we investigated how hybrids afected rat stomach ulcers. Figure [4](#page-4-1) clearly depicts the changes in the histology of the stomachs of rats. While the indomethacintreated group (Fig. [4](#page-4-1)B) showed an interfered with glandular mucosal layer with numerous focal ulcerative zones, marked by the bringing of the epithelial lining and an accumulation of necrotic tissue, the control group (Fig. [4](#page-4-1)A) showed a normal gastric mucosa without any indication of erosion or infammation. Furthermore, after administration of both compounds **4b** and **4f** (Fig. [4C](#page-4-1), [D\)](#page-4-1) the glandular mucosa and



<span id="page-4-1"></span>**Fig. 4** Macroscopical evaluation efect of compounds **4b** and **4f** on the integrity of gastric mucosal membranes. **A** control group. **B** Indomethacin-induced peptic ulcer. **C 4b** treated group, **D 4f** treated group. (H&E stain) magnification power  $\times 20$ 

submucosa in the several regions investigated in the stomach mucosa exhibited largely intact histological structure without any defects. More information from a new study with long term treatment is required to clarify the gastric efects of these compounds at long term of usage.

#### **Infammation induced by LPS in the RAW 264.7 cell line**

The efficacy of synthesized compounds and their cytotoxicity on LPS-induced RAW 264.7 cells were evaluated at diferent concentrations by using the MTT assay. As seen in Fig. [5,](#page-5-0) as compared to LPS-induced macrophages, none of the two compounds were able to stop the growth of macrophages at 5 µM.

#### **Structure–activity relationship**

From the anti-infammatory activity of the newly prepared compounds, some relations between their structure and activity could be concluded as outlined in Fig. [6](#page-5-1).

In case of 1*H*-indole derivatives **4a-c**, the presence of electron-withdrawing groups  $(Cl, NO<sub>2</sub>)$  at C-5 of indole moiety markedly improved the in *vivo* anti-infammatory potential. This is obvious upon comparing the anti-infammatory activity of compound **4b** (3 h; EI=38.50%, 5 h; EI=45.63%) and **4c** (3 h; EI=23.33%, 5 h; EI=39.19%) with unsubstituted analog **4a** (3 h; EI=18.05%, 5 h; EI=12.76%). In addition, methylation of N-1 of indole derivatives attenuated the anti-infammatory potential (except **4c)** as observed in methylation of **4b** (3 h; EI = 38.50%, 5 h; EI = 45.63%) to **4e** (3 h; EI=5.69%, 5 h; EI=8.72%). While, methylation



<span id="page-5-0"></span>**Fig. 5** Cytotoxic evaluation of **4b** and **4f** hybrids in RAW 264.7 cells stimulated with lipopolysaccharides (LPS). The MTT assay was used to measure cell viability. The data is shown as mean $\pm$ SD (n=3). Relative to the LPS-induced cells, the statistical analysis showed no significant differences ( $p > 0.05$ )

of 5-nitroindole derivative (**4c**) (3 h; EI = 23.33%, 5 h; EI=39.19%) to 5-nitro-*N*-methylindole derivative **4f** (3 h;  $EI = 31.94\%, 5 \text{ h}; EI = 41.84\%)$  enhanced the anti-inflammatory potential.

#### **Molecular docking study**

To gain insights into the fundamental mechanism of action of newly prepared indol-3-ylidenehydrazino-1,3-thiazole derivatives, molecular docking of the most active candidates (**4b** and **4f**) was conducted inside COX-2 active region. The results of docking including docking score (Kcal/mol), types of interactions and the binded amino acids are listed in Table [3](#page-6-0) and Figs. [7](#page-6-1) and [8.](#page-7-0)

Compound **4b** revealed good binding within COX-2 with binding energy score = -11.45 kcal/mol. Conventional hydrogen bonds with ARG376, TRP139, ASP229 and GLY235 amino acids were detected (Fig. [7\)](#page-6-1). In addition, this compound **4b** displayed other Pi-Cation interactions with LYS333, Pi-Alkyl binding with LEU145, PRO538; Amide-Pi Stacked with LEU224 and Van der Waals interactions with SER143 and ASN375 (Fig. [7](#page-6-1)).

Furthermore, compound **4f** exhibited three hydrogen bonding interactions with ARG44, CYS41 and GLY135 amino acids with a binding energy score equal to − 10.48 kcal/mol. also, other binding interactions were registered as Pi-Alkyl binding with LYS468, PRO153 and ALA156; Amide-Pi binding with VAL155 and Van der Waals interaction with ASN34, ALA156 and ARG469 (Fig. [8\)](#page-7-0).

<span id="page-5-1"></span>**Fig. 6** SAR study of in vivo anti-infammatory activity of target compounds **4a–f**

**The presence of electron withdrawing groups at C-5 (except 4e) recorded higher AI activity than unsubstituted analogues**





<span id="page-6-1"></span>**Fig. 7** The proposed binding mode of compound **4b** within COX-2 enzyme. **A** 2D binding form, **B** 3D binding form

#### Molecular Diversity

<span id="page-6-0"></span>**Table 3** Outcomes of docking study for target candidates **4b** and **4f** inside COX-2 enzyme





<span id="page-7-0"></span>**Fig. 8** The proposed binding mode of compound **4f** within COX-2 enzyme. **A** 2D binding form, **B** 3D binding form

# **Experimental**

### **Chemistry**

All melting points were recorded on Melt-Temp II melting point apparatus. IR spectra were measured as KBr pellets on a Shimadzu DR-8001 spectrometer. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were recorded on a Bruker at 400 MHz and 100 MHz using TMS as an internal reference, DMSO- $d_6$  as solvent. The elemental analyses were carried out on a Perkin-Elmer 240C Micro analyzer. All reactions were monitored by thin layer chromatography (TLC) using precoated plates of silica gel G/UV-254 of 0.25 mm thickness (Merck 60F254), with R*f* value 0.58–64 (eluent solvent, Hexane/EtOH, 5:1).

### **General Procedure for the Synthesis of thiazole derivatives 4a–f**

An equimolar mixture of thiosemicarbazide (**1**) (0.01 mol, 0.91 g) with the appropriate isatin derivative (**2a–f**) (0.01 mol) was allowed to refux in ethanol (20 ml., 5 drops TEA). Then 4-(bromoacetyl)-*N*-(4-methylphenyl)benzenesulfonamide (**3**) (0.01 mol, 0.35 g) was added to the reaction mixture, and refuxing was continued for 20–30 min until the reaction was complete (TLC) to afford the corresponding thiazole derivatives (**4a–f**). The reaction mixture was allowed to cool to room temperature and the solid precipitate was fltrated, and recrystallized from ethanol.

### **4‑{2‑[2‑(2‑Oxo‑1,2‑dihydro‑1H‑indol‑3‑ylidene) hydrazino]‑1,3‑thiazol‑4‑yl}‑***N***‑phenyl‑4‑tolyl sul‑ fonamide (4a)**

Yield, 85%; Mp 335–337 °C; R<sub>f</sub> value = 0.59; IR cm<sup>-1</sup>: 3363, 3178, 3120 (3NH), 3064 (C–H<sub>arom.</sub>), 1682 (C=O), <sup>1</sup>H NMR *δ* 13.30 (br,1H, NH), 11.20 (s, 1H, NH), 10.29 (s, 1H, NH), 7.76–6.85 (m, 13H, CH<sub>arom.</sub>), 2.33 (s, 3H, -CH<sub>3</sub>); 13C NMR; 166.5 (C=O), 150.9, 143.8, 141.7, 137.9, 137.0, 132.5, 130.9, 130.3, 130.2, 130.1, 127.1, 125.0, 122.9, 120.4, 120.2, 120.1, 111.5, 106.4, 21.3; Anal. Calcd. For  $C_{24}H_{19}N_5O_3S_2$  (489.56) C (58.88%), H (3.91%), N (14.31%), S (13.10%); Found C (58.95%), H (3.98%), N (14.26%), S  $(13.16\%).$ 

### **4‑{2‑[2‑(5‑Chloro‑2‑oxo‑1,2‑dihydro‑3H‑in‑ dol‑3‑ylidene)hydrazino]‑1,3‑thiazol‑4‑yl}‑***N***‑phenyl‑ tolylsulfonamide (4b)**

Yield, 88%; Mp 341–343 °C; R<sub>f</sub> value = 0.58; IR cm<sup>-1</sup>: 3308, 3124 (3NH), 3061 (CH- arom.), 1671 (C=O), <sup>1</sup>H NMR *δ:* 13.24 (s,1H, NH), 11.26 (s, 1H, NH), 10.29 (s, 1H, NH), 8.39–6.81 (m, 12H, CH<sub>arom.</sub>), 2.33 (s, 3H, -CH<sub>3</sub>); <sup>13</sup>C

NMR δ: 163.3 (C=O), 150.9, 143.9, 143.8, 141.0, 140.2, 138.7, 137.9, 137.0, 131.3, 130.2, 129.9, 127.1, 127.0, 127.0, 125.8, 121.8, 120.4, 120.1, 119.4, 21.3; Anal. Calcd. For  $C_{24}H_{18}CIN_5O_3S_2$  (524.01) C (55.01%), H (3.46%), N (13.36%), S (12.24%); Found; C (54.98%), H (3.39%), N (13.29%), S (12.18%); MS: *m/z*: 524 [*M*] +.

### **4‑{2‑[2‑(5‑Nitro‑2‑oxo‑1,2‑dihydro‑3H‑in‑ dol‑3‑ylidene)hydrazino]‑1,3‑thiazol‑4‑yl}‑***N***‑phenyl‑ tolysulfonamide (4c)**

Yield, 78%; Mp 312–314 °C; R<sub>f</sub> value = 0.60; IR cm<sup>-1</sup>: 3247- 3154 (3NH), 3085 (C–H<sub>arom.</sub>), 1654 (C=O), <sup>1</sup>H NMR *δ:* 10.28 (s,1H, NH), 9.14 (s, 1H, NH), 8.64–7.11 (m, 13H, CH<sub>arom</sub> + NH  $)$ , 2.32 (s, 3H, CH<sub>3</sub>), <sup>13</sup>C NMR δ: 163.5 (C=O), 146.6, 146.2, 144.9, 134.1, 133.5, 133.0, 131.5, 130.0, 130.7, 130.3, 128.9, 128.7, 128.6, 127.2, 123.8, 123.6, 123.4, 22.4; Anal. Calcd. For  $C_{24}H_{18}N_6O_5S_2$  (534.56); C (53.92%), H (3.39%), N (15.72%), S (12.00%) Found: C (53.86%), H (3.46%), N (15.65%), S (12.06%); MS: *m/z*: 534 [*M*] +.

### **(4‑(2‑(2‑(1‑methyl‑2‑oxoindolin‑3‑ylidene)hydrazi‑ nyl)thiazol‑4‑yl)‑***N***‑(p‑tolyl) benzene sulfonamide (4d)**

Yield, 82%; Mp 328–330 °C; R<sub>f</sub> value = 0.64; IR cm<sup>-1</sup>: 3217, 3161 (2NH), 3078 (C–H<sub>arom.</sub>), 1658 (C=O), <sup>1</sup>H NMR *δ* 13.16 (br,1H, NH), 10.45 (s, 1H, NH), 7.75–7.02 (m, 13H, CH<sub>arom.</sub>), 3.73 (s, 3H, N-CH<sub>3</sub>), 2.30 (s, 3H, CH<sub>3</sub>); <sup>13</sup>C NMR δ: 161.2 (C=O), 152..4, 143.8, 142.1, 138.7, 137.1, 133.0, 131.2, 130.2, 129.8,128.9, 127.2, 123.3, 122.2, 120.5, 119.5, 119.3, 115.4, 34.5, 21.3; Anal. Calcd. For  $C_{25}H_{21}N_{5}O_{3}S_{2}$ (503.36); C (59.62%), H (4.20%), N (13.91%), S (12.73%) Found; C (59.69%), H (4.16%), N (13.84%), S (12.85%); MS: *m/z*: 503 [*M*] +.

### **4‑{2‑[2‑(5‑Chloro‑2‑oxo‑1,2‑dihydro‑***N***‑me‑ thyl‑3H‑indol‑3‑ylidene)hydrazino]‑1,3‑thia‑ zol‑4‑yl}‑N‑phenyl‑4 tolylsulfonamide (4e)**

Yield, 92%; Mp 330–335 °C; R<sub>f</sub> value = 0.62; IR cm<sup>-1</sup>: 3190, 3167 (2NH), 3047 (C–H<sub>arom.</sub>), 1661 (C=O), <sup>1</sup>H NMR *δ* 12.27 (s,1H, NH), 10.33 (s, 1H, NH), 7.79–6.99 (m, 12H,  $CH_{arom}$ ), 3.75 (s, 3H, N-CH<sub>3</sub>), 2.33 (s, 3H, CH<sub>3</sub>), <sup>13</sup>C NMR δ (ppm): 190.6 (C=O), 146.6, 146.5, 140.5, 137.3, 134.2, 132.8, 131.8, 131.1, 130.8, 129.0, 128.6, 128.5, 127.5, 126.0, 123.3, 122.7, 122.3,121.8, 118.7, 31.6, 21.6; Anal. Calcd. For  $C_{25}H_{20}CIN_5O_3S_2(538.04)$  C (55.81%), H (3.75%), N (13.02%), S (11.92%) Found C (55.86%), H (3.83%), N (12.97%), S (11.86%); MS: *m/z*: 538 [*M*] +.

### **4‑{2‑[2‑(5‑Nitro‑2‑oxo‑***N***‑methyl‑3H‑indol‑3‑ylidene) hydrazino]‑1,3‑thiazol‑4‑yl}‑***N***‑phenyl‑4‑tolylsulfon‑ amide (4f)**

Yield, 86%; Mp 305–307 °C; R<sub>f</sub> value = 0.60; IR cm<sup>-1</sup>: 3251, 3126 (2NH), 3085 (C–Harom.), 1654 (C=O), 1 H NMR *δ* 9.18  $(s, 1H, NH)$ , 9.03  $(s, 1H, NH)$ , 8.64–7.11 (m, 12H, CH<sub>arom</sub>), 3.84 (s, 3H, N-CH<sub>3</sub>), 2.34 (s, 3H, CH<sub>3</sub>), <sup>13</sup>C NMR δ: 162.3 (C=O), 154.2, 153.0, 151.0, 150.3, 146.3, 133.2, 132.7, 132.6, 132.3, 131.7, 130.7, 129.8, 129.7, 129.3, 128.6, 124.4, 123.0, 30.6, 22.0; Anal. Calcd. For  $C_{25}H_{20}N_6O_5S_2$ (548.59); C (54.73%), H (3.67%), N (15.32%), S (11.69%) Found: C (54.69%), H (3.73%), N (15.26%), S (11.78%). MS: *m/z*: 548 [*M*] +.

### **Biological evaluation**

#### **In vivo anti‑infammatory activity**

For the in vivo assessment of the test compounds' antiinflammatory activity, male Wister rats weighing  $180 \pm 10$  g each were employed, with celecoxib serving as the reference drug. All animals had to acclimate to the criteria set by the Institutional Animals Ethics Committee (IAEC) of the Faculty of Science at Sohag University for at least one week prior to the investigations (permit No;). For this in vivo evaluation, 40 adult male Westar rats  $(n=4)$  were randomly assigned. The selected agents were suspended in 1% newly prepared carboxy methyl cellulose (CMC) prior to being administered orally by gavage. Following a sub plantar injection of 100  $\mu$ L of freshly prepared carrageenan gel (1%) distilled water) into each rat's left hind paw, changes in paw thickness were observed [\[46](#page-11-1)]. Rats were administered test compounds orally via gavage one hour before the injection of carrageenan. Paw thickness was measured one, three, and fve hours after the development of infammation. The tested compound's efects were quantifed as a percentage of edema inhibition. The anti-infammatory potential is expressed as a percentage suppression of paw edema and quantifed [\[52](#page-11-2)].

#### **Histopathological analysis of the tissues in the paws**

Prior to being embedded in paraffin, the tissues from the paws were stored in a 10% formalin-neutral buffer. Hematoxylin and eosin (H&E) were used to stain the slides after thin sections of 5–6 µm were cut using a microtome. The slides that were made with a light microscope exhibit pathological changes in them.

#### **In vitro COX inhibition Screening**

By utilizing the appropriate Human enzyme immune assay (ELIA) kits (Cayman Chemical, USA) and adhering to the

manufacturer's instructions, the inhibitory activities of the test hybrids **4b** and **4f** versus COX-1, and COX-2 enzymes were assessed. The results were expressed as an  $IC_{50}$  value  $(\mu M)$ . The data for studies conducted in triplicate are shown as  $IC_{50} \pm SD$ . As positive controls, we employed the wellknown COX-1, and COX-2 inhibitors indomethacin and celecoxib [\[53](#page-11-3)].

### **Histological evaluation of ulcers**

To assess the ulcerogenic potential of diferent hybrids, twenty-four adult male Albino Wistar rats weighing  $180 \pm 20$  g were divided into four groups. Water was supplied to the animals as needed throughout their overnight fast before treatment. The chosen hybrids were given orally at a dose of 50 mg/kg, whereas the positive control was indomethacin (30 mg/kg suspension in 1% CMC orally). Four hours later, the rats were slaughtered, and to assess the extent of gastrointestinal injury, their stomachs were promptly removed, cleaned with ice-cold saline, and preserved in 10% formalin saline [\[54\]](#page-11-4). The stomach tissues from each group were thinly sliced and prepared into slides, which were then stained with hematoxylin and eosin (H&E) for microscopic examination [\[55](#page-11-5)].

### **Assessment of the cytotoxicity of selected hybrids on RAW cells**

RAW 264.7 cells were seeded at 5000 cells per well in a 96-well plate and incubated for 24 h. Following this, the cells were exposed to the investigated hybrids (**4b** and **4f**) at five concentrations  $(5, 10, 20, 40, \text{ and } 80 \mu\text{M})$  for two hours before being stimulated with 1 µg/mL lipopolysaccharide (LPS) for 48 h. The cytotoxic activity of the produced hybrids was determined using the MTT assay. The hybrids were used at subtoxic concentrations to the cells in all sub-sequent tests to prevent any cytotoxic influence [[56,](#page-11-6)57].

### **Docking study**

The crystal structure of COX-2 was downloaded from Protein Data Bank (PDB:1CX2) and the molecular docking was performed following our previously reported work [\[52](#page-11-2)].

### **Statistical analysis**

The obtained data were statistically analyzed using Graph-Pad Prism version 9, and the mean values and standard deviations (mean  $\pm$  SD) were presented as a result. The significance of mean differences was evaluated using the Tukey–Kramer test and one-way analysis of variance (ANOVA), with p-values of less than 0.05 being considered statistically signifcant.

### **Conclusion**

New series of novel indole-thiazole hybrids derivatives **4a–f** were synthesized via multi-components of thiosemicarbazide with some isatine derivatives **a–f** and *N*-(4-(2 bromoacetyl)phenyl)-4-methylbenzenesulfonamide (**3**) under refux in ethanol. The chemical structures of novel compounds were elucidated by elemental and spectral analyses. All new compounds have been screened for their anti-infammatory activity using celecoxib as a reference drug. It has been found that compound **4b** (3 h;  $EI = 38.50\%, 5 \text{ h}; EI = 45.63\%)$  has the most promising and efective anti-infammatory potential. Furthermore, molecular docking study of compounds **4b** and **4f** displayed that these compounds ftted into the COX-2 binding site with good docking energy scores.

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**Author contribution** All authors wrote the main manuscript text and reviewed the manuscript.

**Data availability** No datasets were generated or analysed during the current study.

### **Declarations**

**Conflict of interest** The authors declare no competing interests.

**Ethical approval** This study has been carried on rats and the authors take personal responsibility for knowingtheir statutory responsibility under the Animal (Scientifc Procedures) Act 1986, under the acceptance of Committee for Scientifc Research Ethics (CSRE), code no. (CSRE-23-24)-Sohag University.

### **.References**

- <span id="page-9-0"></span>1. Silva LA (2015) literature review of infammation and its relationship with the oral cavity. Glob J Infect Dis Clin Res 1:21–27
- <span id="page-9-1"></span>2. Abdellatif KRA, Abdelall EKE, Bakr RB (2017) Nitric oxide-NASIDS donor prodrugs as hybrid safe anti-inflammatory agents. Curr Top Med Chem 17:941–955
- <span id="page-9-2"></span>3. Jahnavi K, Reddy PP, Vasudha B, Narender B (2019) Non-steroidal anti-infammatory drugs: an overview. J Drug Deliv Ther 9:442–448
- <span id="page-9-3"></span>4. Hil $\tilde{A}$ ; rio, MOE, Terreri, MT, and Len, CuA. (2006) Nonsteroidal anti-infammatory drugs: cyclooxygenase 2 inhibitors. J Pediatr 82:S206–S212
- <span id="page-9-4"></span>5. Gudis K, Sakamoto C (2005) The role of cyclooxygenase in gastric mucosal protection. Dig Dis Sci 50:S16–S23
- 6. Martinez-Gonzalez J, Badimon L (2007) Mechanisms underlying the cardiovascular efects of COX-inhibition: benefts and risks. Curr Pharm Des 13:2215–2227
- <span id="page-10-0"></span>7. Perrone G, Scilimati M (2010) Selective COX-1 inhibition: a therapeutic target to be reconsidered. Curr Med Chem 17:3769–3805
- <span id="page-10-1"></span>8. Vitale P, Panella A, Scilimati A, and Perrone MG (2016) COX-1 Inhibitors: beyond structure toward therapy. Med Res Rev 36:641–671
- 9. Abdelgawad MA, Bakr RB, Azouz AA (2018) Novel pyrimidinepyridine hybrids: synthesis, cyclooxygenase inhibition, antiinfammatory activity and ulcerogenic liability. Bioorg Chem 77:339–348
- <span id="page-10-2"></span>10. Bakr RB, Azouz AA, Abdellatif KR (2016) Synthesis, cyclooxygenase inhibition, anti-infammatory evaluation and ulcerogenic liability of new 1-phenylpyrazolo [3, 4-d] pyrimidine derivatives. J Enzyme Inhib Med Chem 31:6–12
- <span id="page-10-3"></span>11. McMurray RW, Hardy KJ (2002) Cox-2 inhibitors: today and tomorrow. Am J Med Sci 323:181–189
- <span id="page-10-4"></span>12. Bakr RB, Ghoneim AA, Azouz AA (2019) Selective cyclooxygenase inhibition and ulcerogenic liability of some newly prepared anti-infammatory agents having thiazolo [4, 5-d] pyrimidine scaffold. Bioorg Chem 88:102964
- <span id="page-10-5"></span>13. Dogn J-M, Supuran CT, Pratico D (2005) Adverse cardiovascular efects of the coxibs. J Med Chem 48:2251–2257
- 14. Burnier M (2005) The safety of rofecoxib. Expert Opin Drug Saf 4:491–499
- <span id="page-10-6"></span>15. PraticoÌ D, DogneÌ J-M (2005) Selective cyclooxygenase-2 inhibitors development in cardiovascular medicine. Circulation 112:1073–1079
- <span id="page-10-7"></span>16. Hemalatha K, Madhumitha G, Roopan SM (2013) Indole as a core anti-infammatory agent: a mini review. Chem Sci Rev Lett 2:287–292
- 17. Singh S, Sharma N, Chandra R (2022) The indole nucleus as a selective COX-2 inhibitor and anti-infammatory agent (2011– 2022). Org Chem Front 9:3624–3639
- <span id="page-10-8"></span>18. Kumar D, Kumar RR, Pathania S, Singh PK, Kalra S, Kumar B (2021) Investigation of indole functionalized pyrazoles and oxadiazoles as anti-infammatory agents: synthesis, in-vivo, in-vitro and in-silico analysis. Bioorg Chem 114:105068
- <span id="page-10-9"></span>19. Amin NH, El-Saadi MT, Hefny AA, Abdelazeem AH, Elshemy HA, Abdellatif KR (2018) Anti-infammatory indomethacin analogs endowed with preferential COX-2 inhibitory activity. Future Med Chem 10:2521–2535
- 20. Nagesh KM, Prashanth T, Khamees HA, Khanum SA (2022) Synthesis, analgesic, anti-infammatory, COX/5-LOX inhibition, ulcerogenic evaluation, and docking study of benzimidazole bearing indole and benzophenone analogs. J Mol Struct 1259:132741
- <span id="page-10-10"></span>21. Sravanthi T, Manju S (2015) Indoles: a promising scafold for drug development. Eur J Pharm Sci 91:1–10
- <span id="page-10-11"></span>22. Kaur J, Bhardwaj A, Huang Z, Knaus EE (2012) N-1 and C-3 substituted indole Schiff bases as selective COX-2 inhibitors: synthesis and biological evaluation. Bioorg Med Chem Lett 22:2154–2159
- <span id="page-10-12"></span>23. ÃrT J, Gomes FO, de Miranda MD, de Almeida SM, da Cruz-Filho IJ, Peixoto CA, da Silva TG, Moreira DR, de Melo CM, de Oliveira JF (2023) Anti-infammatory activity of novel thiosemicarbazone compounds indole-based as COX inhibitors. Pharmacol Rep 73:907–925
- <span id="page-10-13"></span>24. Hassan M, Ghafari R, Sardari S, Farahani YF, Mohebbi S (2020) Discovery of novel isatin-based thiosemicarbazones: synthesis, antibacterial, antifungal, and antimycobacterial screening. Res Pharm Sci 15(3):281–290
- 25. Rahim F, Taha M, Iqbal N, Hayat S, Qureshi F, Uddin I, Zaman K, Rab A, Wadood A, Uddin N, Nawaz M (2020) Isatin based thiosemicarbazide derivatives as potential inhibitor of

α-glucosidase, synthesis and their molecular docking study. J Mol Struct 1222:128922

- 26. Cheng R, Shi W, Yuan Q, Tang R, Wang Y, Yang D, Xiao X, Zeng J, Chen J, Wang Y (2021) 5-Substituted isatin thiosemicarbazones as inhibitors of tyrosinase: Insights of substituent efects. Spectrochim Acta Part A 255:119669
- 27. Freitas LAB, Santos AC, Silva GC, Albuquerque FN, Silva ED, Simone CA, Pereira VR, Alves LC, Brayner FA, Leite AC, Gomes PA (2021) Structural improvement of new thiazolylisatin derivatives produces potent and selective trypanocidal and leishmanicidal compounds. Chem Biol Interact 345:109561
- <span id="page-10-14"></span>28. Hasan Y, Mohammed A, Sevgi K, Halit M, Mustafa E, Parham T, Ümit MK, Muhammet K, Saud A, Kim M (2023) Isatin/thiosemicarbazone hybrids: facile synthesis, and their evaluation as anti-proliferative agents and metabolic enzyme inhibitors. Bull Chem Soc Ethiop 37(5):1221–1236
- <span id="page-10-15"></span>29. Patel S, Patle R, Parameswaran P, Jain A, Shard A (2019) Design, computational studies, synthesis and biological evaluation of thiazole-based molecules as anticancer agents. Eur J Pharm Sci 134:20–30
- <span id="page-10-16"></span>30. Pawar S, Kumar K, Gupta MK, Rawal RK (2021) Synthetic and medicinal perspective of fused-thiazoles as anticancer agents. Anticancer Agents Med Chem 21:1379–1402
- <span id="page-10-17"></span>31. Djukic M, Fesatidou M, Xenikakis I, Geronikaki A, Angelova VT, Savic V, Pasic M, Krilovic B, Djukic D, Gobeljic B (2018) In vitro antioxidant activity of thiazolidinone derivatives of 1, 3-thiazole and 1, 3, 4-thiadiazole. Chem Biol Interact 286:119–131
- <span id="page-10-18"></span>32. Muluk M, Patil PS, Kasare SL, Kulkarni RS, Dixit PP, Choudhary P, Haval KP (2020) Synthesis and molecular docking studies of novel pyridine-thiazole-hydrazone conjugates as antimicrobial and antioxidant agents. Eur Chem Bull 9:184–192
- <span id="page-10-19"></span>33. Borcea A-M, IonuÈ I, CriÈan O, Oniga O (2021) An overview of the synthesis and antimicrobial, antiprotozoal, and antitumor activity of thiazole and bisthiazole derivatives. Molecules 26:624
- <span id="page-10-20"></span>34. Mishra I, Mishra R, Mujwar S, Chandra P, Sachan N (2020) A retrospect on antimicrobial potential of thiazole scafold. J Heterocycl Chem 57:2304–2329
- <span id="page-10-21"></span>35. Khatik GL, Datusalia AK, Ahsan W, Kaur P, Vyas M, Mittal A, Nayak SK (2018) A retrospect study on thiazole derivatives as the potential antidiabetic agents in drug discovery and developments. Curr Drug Discov Technol 15:163–177
- <span id="page-10-22"></span>36. Amnerkar ND, Bhusari KP (2011) Synthesis of some thiazolyl aminobenzothiazole derivatives as potential antibacterial, antifungal and anthelmintic agents. J Enzyme Inhib Med Chem 26:22–28
- <span id="page-10-23"></span>37. Kamat V, Santosh R, Poojary B, Nayak SP, Kumar BK, Sankaranarayanan M, Faheem Khanapure S, Barretto DA, Vootla SK (2020) Pyridine-and thiazole-based hydrazides with promising anti-infammatory and antimicrobial activities along with their in silico studies. ACS Omega 5:25228–25239
- <span id="page-10-24"></span>38. Pattan SR, Hullolikar R, Dighe NS, Ingalagi B, Hole M, Gaware V, Chavan P (2009) Synthesis and evaluation of some new phenyl thiazole derivatives for their anti-infammatory activities. J Pharm Sci Res 1:96
- <span id="page-10-25"></span>39. Tratrat C, Haroun M, Tsolaki E, Petrou A, Gavalas A, Geronikaki A (2021) Thiazole-based chalcone derivatives as potential anti-infammatory agents: Biological evaluation and molecular modeling. Curr Top Med Chem 21:257–268
- <span id="page-10-26"></span>40. Manju S (2020) Identifcation and development of thiazole leads as COX-2/5-LOX inhibitors through in-vitro and in-vivo biological evaluation for anti-infammatory activity. Bioorg Chem 100:103882
- <span id="page-10-27"></span>41. Helal M, Salem M, El-Gaby M, Aljahdali M (2013) Synthesis and biological evaluation of some novel thiazole compounds as potential anti-infammatory agents. Eur J Med Chem 65:517–526

<span id="page-11-3"></span><span id="page-11-2"></span>51. Arooj B (2023) Anti-infammatory mechanisms of eucalyptol rich Eucalyptus globulus essential oil alone and in combination with furbiprofen. Infammopharmacology 31(4):1849–1862 52. Rudrapal M, Eltayeb WA, Rakshit G, El-Arabey AA, Khan J, Aldosari SM (2023) Dual synergistic inhibition of COX and LOX by potential chemicals from Indian daily spices investigated through detailed computational studies. Sci Rep 13:8656 53. Subhawa S, Arpornchayanon W, Jaijoy K, Chansakaow S, Soonthornchareonnon N, Sireeratawong S (2023) Anti-infammatory, antinociceptive, antipyretic, and gastroprotective efects of Eurycoma longifolia Jack ethanolic extract. Life 13(7):1465 54. Shaik RA (2024) Parthenolide alleviates indomethacin-induced gastric ulcer in rats via antioxidant, anti-infammatory, and antiapoptotic activities. Naunyn Schmiedebergs Arch Pharmacol 4:1–

<span id="page-11-4"></span>13.<https://doi.org/10.1007/s00210-024-03110-x>

Chem 105:104439

<span id="page-11-5"></span>55. Mohamed MFA, Marzouk AA, Nafady A, El-Gamal DA, Allam RM, Abuo-Rahma GE-DA, El Subbagh HI, Moustafa AH (2020) Design, synthesis and molecular modeling of novel aryl carboximidamides and 3-aryl-1,2,4-oxadiazoles derived from indomethacin as potent anti-infammatory iNOS/PGE2 inhibitors. Bioorgan

<span id="page-11-6"></span>56. Ghoneim MM, Abdelgawad MA, Elkanzi NA, Parambi DGT, Alsalahat I, Farouk A, Bakr RB (2024) A literature review on pharmacological aspects, docking studies, and synthetic approaches of quinazoline and quinazolinone derivatives. Arch Pharm 357(8):e2400057. <https://doi.org/10.1002/ardp.202400057>

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- 42. Elkanzi NA, Abdelhamid AA, Ali AM (2022) Designing and antiinfammatory efectiveness of novel phenytoin derivatives *via* one pot multicomponent reaction. Chem Select 7:e202201293
- 43. Abdelgawad MA, Bakr RB, El-Gendy AO, Kamel GM, Azouz AA, Bukhari SNA (2017) Discovery of a COX-2 selective inhibitor hit with anti-infammatory activity and gastric ulcer protective efect. Future Med Chem 9:1899–1912
- 44. Abdelgawad MA, Elkanzi NA, Musa A, Ghoneim MM, Ahmad W, Elmowafy M, Ali AM, Abdelazeem AH, Bukhari SN, El-Sherbiny M (2022) Optimization of pyrazolo [1, 5-a] pyrimidine based compounds with pyridine scafold: synthesis, biological evaluation and molecular modeling study. Arab J Chem 15:104015
- 45. Abdelgawad MA, Al-Sanea MM, Musa A, Elmowafy M, El-Damasy AK, Azouz AA, Ghoneim MM, Bakr RB (2022) Docking study, synthesis, and anti-infammatory potential of some new pyridopyrimidine-derived compounds. J Inflammat Res 5:451–463
- <span id="page-11-1"></span>46. Elkanzi NA, AlHazmi AKG, Bakr RB, Gad MA, Abd ElLateef HM, Ali AM (2023) Design and synthesis of pyridine and thiazole derivatives as ecofriendly insecticidal to control olive pests. Chem Biodivers 20:e202300559
- 47. Shaker ME, Goma HA, Alsalahat I, Elkanzi NA, Azouz AA, Abdel-Bakky MS, Ghoneim MM, Hazem SH, El-Mesery ME, Farouk F (2023) Design and construction of novel pyridinepyrimidine hybrids as selective COX-2 suppressors: anti-infammatory potential, ulcerogenic profle, molecular modeling and ADME/Tox studies. J Biomol Struct Dyn 9:1–14. [https://doi.org/](https://doi.org/10.1080/07391102.2023.2293257) [10.1080/07391102.2023.2293257](https://doi.org/10.1080/07391102.2023.2293257)
- 48. Khodairy A, Ali AM, ElWassimy M (2018) Synthesis and reactions of new thiazoles and pyrimidines containing sulfonate moiety. J Heterocycl Chem 55:964–970
- 49. Elkanzi NA, Kadry AM, Ryad RM, Bakr RB, Ali El-Remaily MAEAA, Ali AM (2022) Efficient and recoverable bio-organic catalyst cysteine for synthesis, docking study, and antifungal activity of new bio-active 3, 4-dihydropyrimidin-2 (1 H)-ones/thiones under microwave irradiation. ACS Omega 7:22839–22849
- <span id="page-11-0"></span>50. Winter CA, Risley EA, Nuss GW (1962) Carrageenin-induced edema in hind paw of the rat as an assay for antiinfammatory drugs. Proc Soc Exp Biol Med 111(3):544–547

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