



New tetrazolopyrrolidine-1,2,3-triazole analogues as potent anticancer agents: design, synthesis and molecular docking studies

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Abstract

1,2,3-Triazole and tetrazole derivatives bearing pyrrolidines are found to exhibit notable biological activity and have become useful scaffolds in medicinal chemistry for application in lead discovery and optimization. We report design, synthesis and molecular docking studies of tetrazolyl-1,2,3-triazole derivatives (**7a-i**) bearing pyrrolidine moiety and evaluating their anticancer activity against four cancer cell lines viz. Hela, MCF-7, HCT-116 and HepG2. The structures of the new compounds were ascertained by spectral means IR, NMR: ¹H & ¹³C and Mass spectrum. From the studies compounds **7a** and **7i** exhibited significant anticancer activity against the Hela cell line with IC₅₀ = 0.32 ± 1.00, 1.80 ± 0.22 μM when compared to reference drug Doxorubicin (IC₅₀ = 2.34 ± 0.11 μM), whereas **7h**, **7i**, and **7b** were found to be active against MCF-7, HCT-116 and HepG2 cell lines with IC₅₀ = 3.20 ± 1.40, 1.38 ± 0.06 and 0.97 ± 0.12 μM respectively. Notably **7a** exhibited highest conventional hydrogen bondings TyrA:40, SerA:17, LysA:117, AlaA:146, Tyr218 with 3HB4 and SerA:17, LysA:117, AlaA:146, TyrA:40 with 6IBZ and docking energy – 10.85, – 8.21 kcal/mol respectively. These compounds were further evaluated for their ADMET and physicochemical properties by using SwissADME. The results of the in vitro and in silico studies suggest that the tetrazole incorporated pyrrolidine-triazoles may possess the ideal structural requirements for further developing new anticancer agents.

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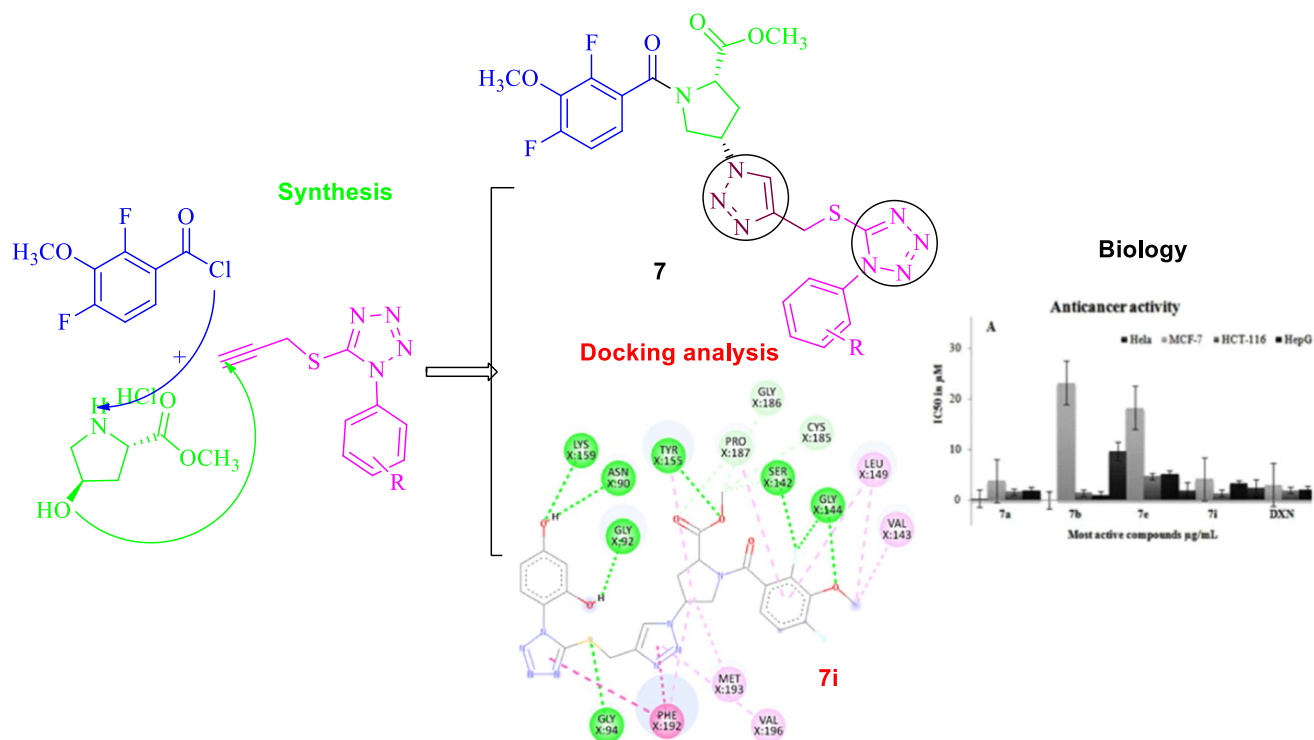
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Graphical Abstract



Keywords 1,2,3-Triazoles · Tetrazoles · Docking analysis · Anticancer activity · SwissADME · PDB

Abbreviations

CYP2C19	Cytochrome P450 2C19 gene family
LGA	Lamarckian Genetic Algorithm
ADMET	Absorption, distribution, metabolism, excretion and toxicology
TPSA	Topological polar surface area
ADT	Auto dock tools
PDBQT	Protein Data Bank includes partial charges ('Q') and atom types ('T')

Introduction

The uncontrolled growths of abnormal cells lead to cause deadly diseases. Cancer is the second biggest cause of illness and mortality after heart disorders according to the World Health Organization. Though there is tremendous growth in the screening of various types of cancer, emergence of effective drugs and prevention, cancer is still the leading cause of death in people [1]. Most of the currently used chemotherapeutic drugs are ineffective because of the development of drug resistance during treatment, in spite of advances in the understanding of the molecular

biology of cancer and the ensuing rise in the development of anticancer compounds [2]. According to GLOBOCAN 2018, approximately 18.1 million new cases of cancer have been recognized and among them, lung cancer (18.4%), followed by breast (11.6%), prostate (7.1%), colorectal (6.1%), stomach and liver cancer are the most common [3]. Breast cancer is the most frequently diagnosed cancer and the leading cause of death among females, accounting for 23% of the total cancer cases and 14% of cancer deaths; thus, research in this field is important to overcome both economical and psychological burden [4]. As for breast cancer, MCF-7 cells represent a very important candidate as they are used ubiquitously in research for estrogen receptor (ER)-positive breast cancer cell experiments and many sub-clones, which have been established, represent different classes of ER-positive tumours with varying nuclear receptor expression levels [5]. The second most frequent female malignant tumour in the world, cervical cancer poses a major threat to the health of women [6]. HeLa cell line was a particularly aggressive strain of cervical cancer cells acquired during a normal biopsy from a 30-year-old mother of five. It has been determined that persistent infection of high-risk human papillomavirus (HPV) is the cause of cervical cancer. Therefore, the need

for new and effective anticancer agents increased for the treatment of new increasing number of cancer patients [3].

5-membered heterocycles with nitrogen are structural motifs that have undergone substantial research in the creation of molecules with a range of biological potential. The tetrazole pharmacophore is present in several current medications [7, 8], in particular, 1,5-substituted tetrazole derivatives have been employed as peptidomimetics and have a wide range of applications in medicinal chemistry as substitutes for cis-amide linkages [9]. Additionally, tetrazole-bearing derivatives are claimed to have biological properties that are anti-hypertensive [10], antibacterial [11], anticonvulsant [12], analgesic [13], antiproliferative [14], anti-fungal [15], anti-tuberculosis [16], anti-malarial, anti-leishmaniasis [17], anti-diabetic [18], anticancer [19], and many other biological activities. Another well-known heterocycles 1,2,3-triazoles, which have a wide range of biological applications, including anticonvulsant [20], antiviral [21], anti-tubercular [22], antibacterial [23], antifungal [24], anticancer [25], antioxidant [26], antimalarial [27], anti-alzheimer effect [28], antidepressant [29], anti-inflammatory [30], antiplatelet [31], anti-HIV activities [32]. Because of their outstanding biological activity profiles, these heterocyclic compounds are well documented in the literature for the treatment of various diseases and deadly tumours as shown in (Fig. 1). Based on the information above and ongoing work, we have designed a small number of hybrid molecules that combine the pyrrolidine, tetrazole, and 1,2,3-triazole pharmacophores in a single framework.

Additionally, the pharmacokinetic properties of the final 1,2,3-triazolyl tetrazoles (**7a–i**) were predicted using absorption, distribution, metabolism excretion, and toxicity

(ADMET) descriptors by a SwissADME (<https://www.swissadme.ch/>), ADMETlab2.0 (<https://admetmesh.scbdd.com>) server. In silico ADMET is currently used widely to determine whether it is possible for a drug candidate to reach its site of action. The synthesized compounds may have fascinating biological features that are effective against many cancer cell lines. To our knowledge, the targeted pyrrolidine-tetrazoles **7a–i** and the applied methodology (Fig. 2) are not reported previously. Therefore, we performed the synthesis and docking studies of tetrazole–triazole linked pyrrolidine derivatives, and these results suggested that they might be lead compounds for treating cancer infections that are resistant to treatment.

Results and discussion

Chemistry

In the present study, we reported the synthesis and structural characterization of a novel series of tetrazolyl 1,2,3-triazoles bearing pyrrolidines and their derivatives as lead compounds via 4-steps. (Scheme 1, Fig. 3). Compounds 2,4-difluoro-3-methoxybenzoyl chloride **1** and (2*S*,4*R*)-methyl-4-hydroxypyrrolidine-2-carboxylate hydrochloride **2** were synthesized by following the previously reported synthetic procedures and detailed experimental procedures are provided in the supporting information [33, 34]. Synthesis of key intermediate (2*S*,4*R*)-methyl 1-(2,4-difluoro-3-methoxybenzoyl)-4-hydroxy pyrrolidine-2-carboxylate **3** was attained by following the reaction conditions reported in the literature with slight modifications [34].

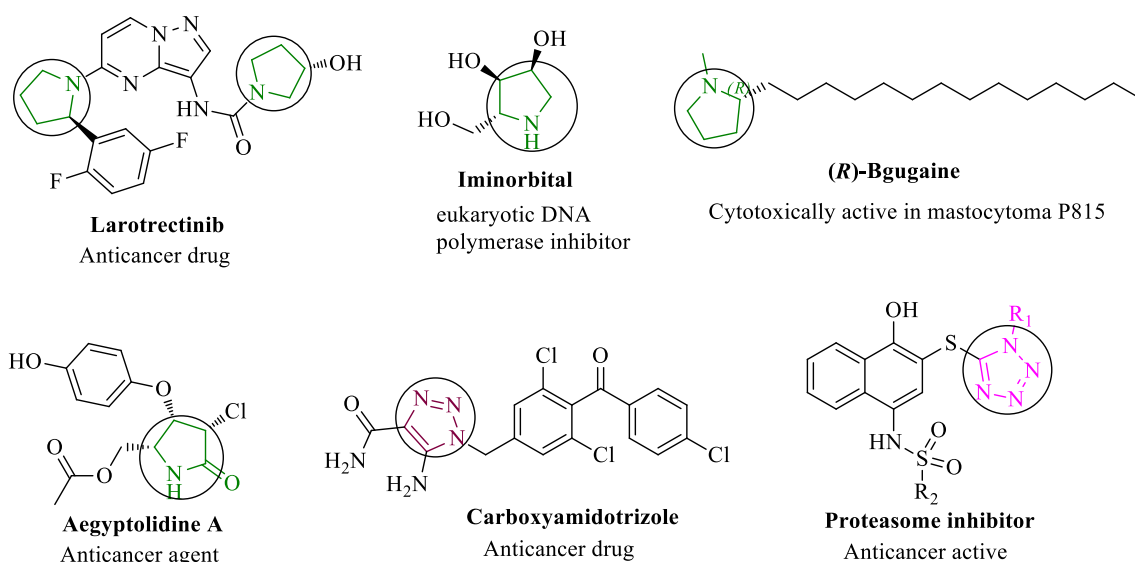


Fig. 1 Chemical structures of the reported anticancer active molecules bearing pyrrolidine, 1,2,3-triazole and tetrazole

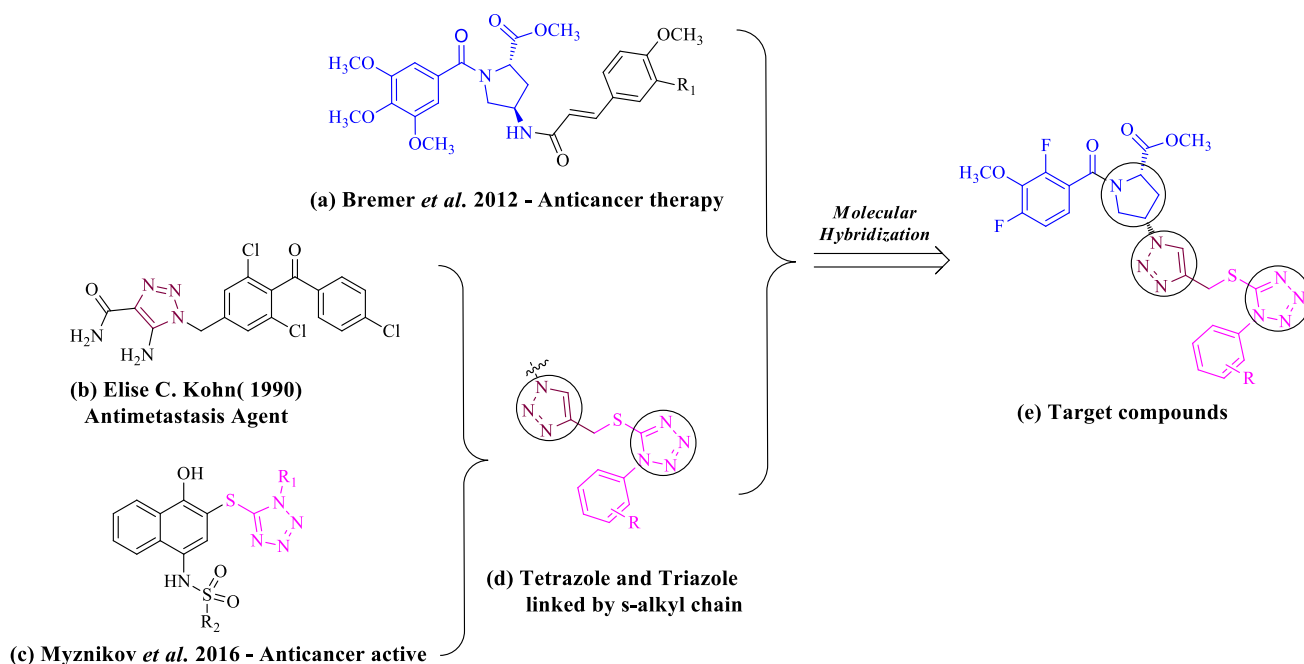
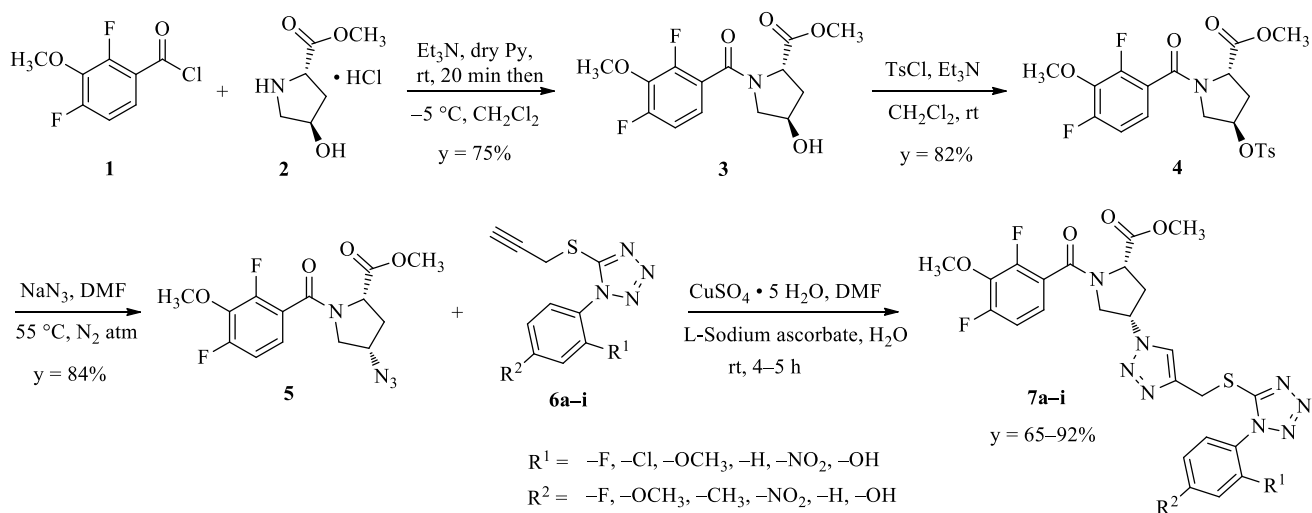


Fig. 2 Rational design strategy of tetrazole incorporated pyrrolidine-triazoles, **a** structure of the previously reported Anticancer compound with pyrrolidine ring, **b** example of 1,2,3-triazole derivative used in

Anticancer therapy, **c** representative example of tetrazole molecule with sulfur that exhibits Anticancer activity



Scheme 1 Synthetic pathway of tetrazole-triazole linked pyrrolidine derivatives

(2*S*,4*R*)-methyl-4-hydroxypyrrolidine-2-carboxylate hydrochloride **2** was suspended in pyridine and triethylamine for 20 min at ambient temperature.

Then it was reacted with 2,4-difluoro-3-methoxy benzoyl chloride **1** in dichloromethane for 4 h, followed by filtration and dried to provide pale yellow solid (key intermediate **3**, 75%). Intermediate **3** was reacted with 4-tolylsulfonyl chloride in dichloromethane and triethylamine to provide Tosylate **4** as a white crystalline solid. The Tosylate was

treated with sodium azide in dry DMF at 50–55 °C to provide pale yellow oil, which solidified upon standing to provide the azide intermediate **5** (S_N² reaction with sodium azide and resulted in the inversion of stereocenter, 83%). A series of 1-phenyl-5-(prop-2-yn-1-ylthio)-1*H*-tetrazoles **6a-i** were synthesized as per the reported literature starting from phenyl isothiocyanate which was reacted with sodium azide to give 1-phenyl-1*H*-tetrazole-5-thiol [35]. Then the solution of 1-phenyl-1*H*-tetrazole-5-thiol, propargyl bromide, and

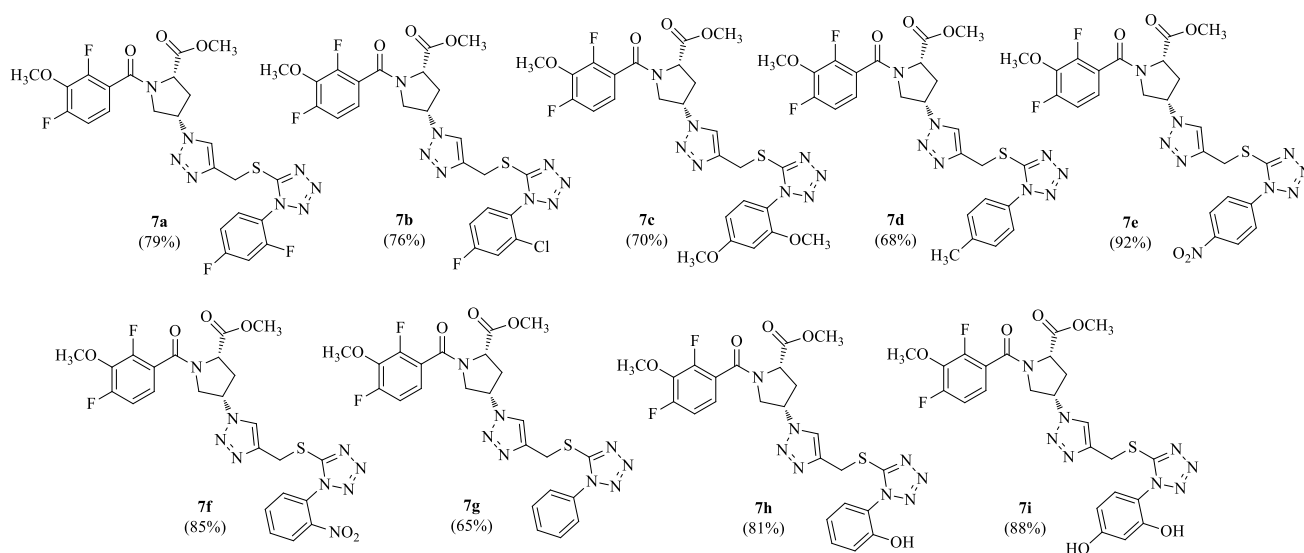


Fig. 3 List of synthesized tetrazole–triazole linked pyrrolidine derivatives with yield

tetrabutylammonium bromide in a mixture of triethylamine and dichloromethane was stirred for 4 h to obtain the tetrazole derivatives (80–87%). The synthesis of new tetrazolyl triazole pyrrolidine derivatives was accomplished via Click chemistry of appropriate substituted tetrazole alkynes **6a–i** with pyrrolidine azide **5**, L-sodium ascorbate in mixture of DMF–H₂O stirred for 4–5 h at room temperature. The synthesized tetrazolyl triazole derivatives **7a–i** were obtained in good yields of up to 92%.

Spectral analysis

All the synthesized compounds **7a–i** were characterized by spectral techniques prior to screening for anticancer activity. The IR spectrum of **7a** has shown the characteristic absorption bands for =CH (aromatic), –CH (alkanes) at 3037.75, 2927 cm⁻¹ respectively. Two strong peaks at 1699.86 cm⁻¹ and 1637.56 cm⁻¹ were pertaining to stretching frequency of the carbonyl group (C=O); the higher frequency band was allocated to ester carbonyl and the lower frequency band was allocated to amide carbonyl vibrations. The bands at 1597.22, 1514.65 cm⁻¹ and 1406.41 cm⁻¹ being assigned to the valence bonds Ar–C=C and –C=N groups. The C–S–C, C–O–C and C–F functional groups were characterized by the presence of the vibrational peaks at 1209.47 cm⁻¹, 1103.49 cm⁻¹ and 1040.23 cm⁻¹ respectively. The ¹H NMR spectrum of compound **7a** exhibited two doublets of doublets at δ 7.37, 8.03 ppm and three doublets at 6.81, 7.54, and 8.44 ppm for aromatic protons respectively. The singlet at 8.62 ppm could be attributed to the triazole proton, whereas the peaks at 3.77, 2.88 and 4.88 ppm were assigned to methoxy (Ar–OCH₃ & –C–OCH₃) and –SCH₂ groups respectively. The pyrrolidine moiety presents four

types of protons, three multiplets, two doublets and a triplet. The triplet at δ 2.65 ppm is attributable to the proton CH whereas, the doublets at δ 3.31, 2.98 ppm are attributed to the protons CH₂. The protons of the methylene moiety exhibited multiplet between δ 0.93–1.65 ppm overlapping with the protons of the adjacent nucleus. The ¹³C NMR of this compound has shown two carbonyl signals at δ 176.2, 172.0 ppm, which are assigned to the carbons of amide and ester groups. Moreover, the aromatic carbons revealed that the signals at δ 160.0, 161.4, 156.2 ppm were attributed to carbon phenolic fragment bearing fluoro benzene groups. Furthermore, one can notice an overlapping of the signals due to aromatic carbons in the range δ 102.5–142.3 ppm whereas the signals exhibited at δ 151.5, 150.0 ppm due to tetrazole, triazole carbons respectively. For compound **7a**, the molecular ion peak m/z at 593 corresponding to [M+H]⁺ was observed by the mass spectrum (ESI–MS) from which the molecular weight is consistent with the molecular formula C₂₄H₂₀F₄N₈O₄S. The chemical structures of all the synthesized compounds are presented in Fig. 3.

Anticancer activity

Anticancer activity of these compounds has been assessed in vitro by MTT 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay against four HeLa, MCF-7, HCT-116, and HepG2 cell lines [36–38]. The percentage of cell death was measured for the new pyrrolidines linked to dual heterocycles **7** exhibited higher anticancer activities at various concentrations along with IC₅₀ values (Table 1, Fig. 4). Among the tested compounds **7a**, **7c**, **7h**, and **7i** showed excellent anticancer activity against both HeLa (human cervix epithelioid carcinoma), MCF-7

Table 1 Cytotoxicity of selected compounds with their IC₅₀ in μM

Entry	Hela (IC ₅₀)	MCF-7 (IC ₅₀)	HCT-116 (IC ₅₀)	HepG2 (IC ₅₀)
7a	1.00 ± 0.32	3.83 ± 0.37	1.66 ± 0.64	1.92 ± 0.06
7b	–	23.17 ± 1.07	1.48 ± 0.08	0.97 ± 0.12
7c	4.85 ± 1.15	10.77 ± 1.12	–	14.62 ± 0.35
7d	14.88 ± 2.01	5.06 ± 0.17	19.20 ± 0.09	8.17 ± 0.29
7e	9.71 ± 0.09	18.3 ± 0.84	4.72 ± 0.03	5.08 ± 1.49
7f	4.83 ± 1.02	8.11 ± 2.12	10.73 ± 0.52	–
7g	6.94 ± 1.16	–	11.90 ± 0.19	10.67 ± 0.62
7h	8.09 ± 0.45	3.20 ± 1.40	–	13.78 ± 1.13
7i	1.80 ± 0.22	4.15 ± 0.55	1.38 ± 0.06	3.25 ± 0.07
DXN	2.34 ± 0.11	3.02 ± 0.39	1.96 ± 0.20	2.08 ± 0.46

(–): IC₅₀ value above 50 μM; these results were expressed as mean value ± standard deviation (SD)

(human breast adenocarcinoma) cell lines. From screening results compounds displayed promising activity and **7a**, **7i** exhibited the highest cytotoxicity against Hela cell line with IC₅₀ 0.32 ± 1.00, 1.80 ± 0.22 μM and **7f**, **7c** shows good activity against Hela cell line with IC₅₀ 4.83 ± 1.02, 4.85 ± 1.15 μM respectively. Compounds **7a**, **7h**, **7i** with difluorophenyl pyrrolidine tetrazole, orthohydroxyphenyl triazolyl-tetrazole, dihydroxyphenyl pyrrolidine elicited good activity against MCF-7 cell line with IC₅₀ value 3.83 ± 0.37, 3.20 ± 1.40, 4.15 ± 0.55 μM respectively. Moreover the compounds **7c**, **7d** and **7f** displayed moderate to good anticancer activity against MCF-7 cell line with IC₅₀ range 10.77 ± 1.12, 5.06 ± 0.17, 8.11 ± 2.12 μM respectively. Compounds **7i**, **7b**, **7a** and **7e** displayed excellent anticancer activity against human colon carcinoma (HCT-116) cell line with IC₅₀ values of 1.38 ± 0.06, 1.48 ± 0.08, 1.66 ± 0.64, and 4.72 ± 0.03 μM where as the dual heterocyclic compounds **7f** and **7g** exhibited the superior anticancer activity

against HCT-116 cell line with IC₅₀ value of 10.73 ± 0.52, 11.90 ± 0.19 μM respectively. Preliminary cytotoxicity was performed with the synthesized heterocyclic compounds and the positive control, doxorubicin towards (human caucasian hepatocyte carcinoma) HepG2 cell line. All selected compounds **7b**, **7a**, **7i**, and **7e** had cytotoxic effect in the tested hepatocyte carcinoma cell line with IC₅₀ values 0.97 ± 0.12, 1.92 ± 0.06, 5.08 ± 1.49, and 3.25 ± 0.07 μM separately. In the present study, it was noted that dual heterocyclic compounds **7a**, **7f**, **7i**, **7e**, **7b**, and **7h** had IC₅₀ values below or around 10 μM on all the four selected cancer cell lines including drug-sensitive and multidrug-resistant phenotypes. Amongst them, all the title compounds and doxorubicin displayed IC₅₀ values below 20 μM, **7b** had an IC₅₀ value of 23.17 ± 1.07 μM meanwhile other compounds **7b** (Hela), **7c**, **7h** (HCT-116), **7f** (HepG2), **7g** (MCF-7) were not active at up to 50 μM (Table 1).

Regarding the structure–activity relationship, it appears that the synthesized 2,4-fluoro phenyl tetrazole derivative **7a** and 2-chloro-4-fluoro phenyl tetrazolyl triazole derivative **7b** had excellent or having good cytotoxic effects against Hela (IC₅₀ values below 10 μM) (Fig. 5). In contrast, o-nitro **7f**, 2, 5- dimethoxy pyrrolidines **7c** with tetrazole and 1,2,3-triazole substituents has the best cytotoxic effect with IC₅₀ values below 10 μM against Hela cancer cell line (Table 1). This is an indication that electron-donating substituents decrease the anticancer activity of pyrrolidines whereas electron-withdrawing substituents increase the activity against all the tested cell lines. Within the tetrazolyl triazole pyrrolidines, it was observed that the numbers and positions of both methoxy (**7b**) methyl (**7d**), and hydroxy (**7h**) functional groups did not significantly influence the cytotoxic activity. The presence of fluoro (**7a**), chloro (**7b**) and nitro (**7e**) substituents significantly increases the anticancer activity against all the tested cell lines. However, the presence

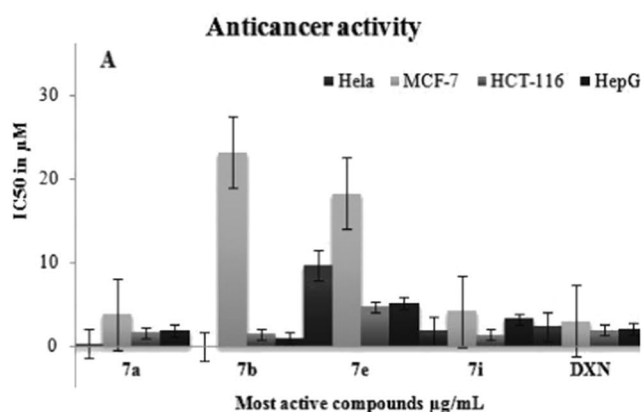
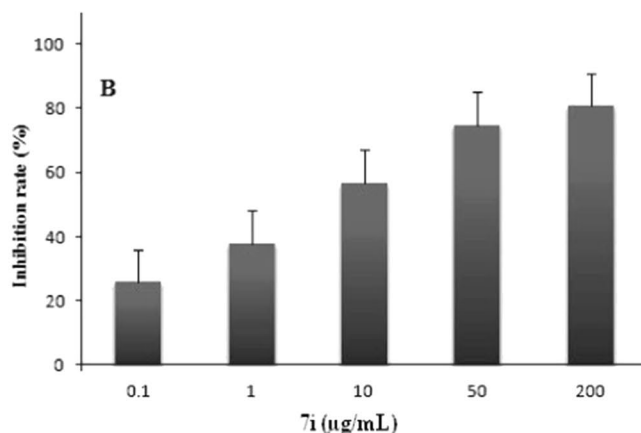


Fig. 4 **A** IC₅₀ values for the most active pyrrolidines against human cervix epithelioid carcinoma (Hela), human breast adenocarcinoma (MCF-7), human colon carcinoma (HCT-116), and human cauca-



sian hepatocyte carcinoma (HepG2) cell lines. **B** Rates of Inhibition of cell invasion by various concentrations of **7i** (HCT-116). Data are mean ± SD (n = 5)

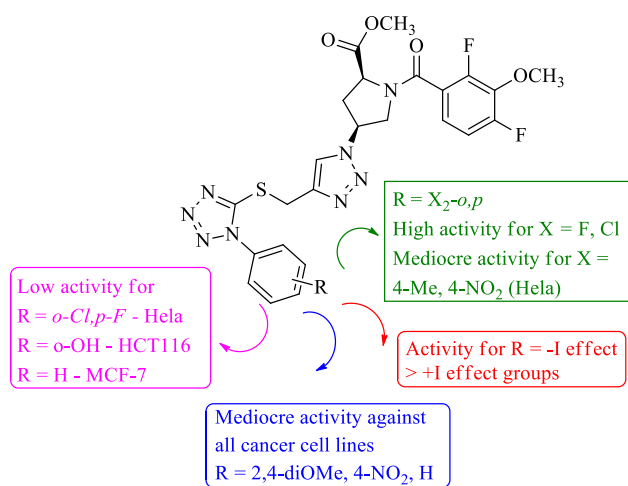


Fig. 5 SAR summary for anticancer activities of tetrazolyl-triazole pyrrolidine derivatives (**7**)

of nitro and hydroxy substituents at *ortho* position of the phenyl ring (**7f** & **7h**) has minimized the biological activity, probably due to the steric hindrance that might prevent the compounds to reach its biological target.

Molecular docking studies

Breast and carcinoma cancer cell lines contain a diverse set of enzymes that are considered necessary for MCF-7, HCT-116 cell lines against 17β -HSD type 1, PFKFB3 inhibitors and reproduction. One of these enzymes that have been recognized as an anticancer target is breast cancer and carcinoma cancer therapy. To validate the accuracy of Autodock 4.2 is an appropriate docking tool for the present purpose, the co-crystal structure of 17β -hydroxy steroid dehydrogenase type 1 complexed with E2B (3HB4), and the co-crystallized structure of Human PFKFB3 in complex with a N-Aryl 6-Aminoquinoxaline inhibitor **7** (6IBZ) having 2.21 Å resolution [39, 40]. According to the method of validation cited in the literature, the successful scoring function is the one in which the RMSD of the best-docked conformation is ≤ 2.0 Å from the experimental one [41–43]. Pyrrolidines **7e**, **7i**, **7a**, **7h** have shown highest docking energies of -11.83 , -11.50 , -10.85 , -9.28 kcal/mol with 17β -hydroxysteroid dehydrogenase protein and formed various π -interactions such as π - π , π -cation, sigma-sigma n - π and π -sigma, alkyl contacts (Table 2, Fig. 6). The ligand **7a** displayed five conventional H-bondings TyrA:40 (O–F) (3.101 Å), SerA:17 (triazole pocket) (2.942 Å), LysA:117 (sulfur pocket) (2.016 Å), AlaA:146 (fluoro benzene pocket) (2.633 Å), Tyr218 (2.165 Å), three π -alkyl residues with ValA:29, AlaA:18 (b/n triazole and tetrazole), PheA:28, one π -anion residue with AspA:38, two fluorine interactions with AspA:119, AsnA:116. Whereas the new 1,2,3-triazolyl

tetrazole **7a** disclosed the diverse noteworthy hydrogen bonding interaction with the active site of amino acid radius SerA:17 (2.346 Å), LysA:117 (1.948 Å), AlaA:146 (2.795 Å), TyrA:40 (2.648 Å), and Π -cation, anion bondings with ArgA:75, AsnA:163 (F), CysA:154, GluA:166. For a most active compound **7e** reveals a dissociation constant of 84.30 nM and exhibited the highest hydrogen binding modes with LysA:147, AlaA:146, LysA:117, SerA:17, AsnA:116 at 1.348, 1.678, 2.037, 1.975 and 2.084 Å respectively. Moreover, this compound possesses carbon hydrogen bondings with AspA:33, ProA:34, ThrA:35, one π - π interaction with PheA:28 (2.348 Å), and two Π -alkyl residues ValA:29, TyrA:40 against 6IBZ protein. Whereas the ligand **7e** reveals significant H-bond interactions with GlyX:15 (2.112 Å), ValX:143 (2.120 Å), Ile:14 (2.004 Å), and notable π -cation, π -alkyl interactions like LysX:159, IleX:14, ValX:225, PheS:259 the hydrogen bonding stackings (Green), carbon-hydrogen bond (Aqua), π - π (pink), Π -halogen (blue), π -anion (dark orange), π -sigma (violet), π -sulphur (yellow), π -alkyl and sigma stackings (rose) were found. The most important derived ligand **7i** exhibits seven stronger H-bond interactions with amino acid LysX:159, AsnX:90, TyrX:155, GlyX:92, SerX:142, GlyX:144, GlyX:94 with bond distance 2.183, 2.165, 2.013, 1.326, 2.015, 1.954, 2.103 Å and the other interactions like carbon-hydrogen bondings exhibits GlyX:186, CysX:185, ProX:187, one Π - Π bonding PheX:192, and four π -alkyl bondings with amino acids MetX:193, ValX:196, ValX:143, LeuX:149 against 3HB4. Compound **7i** formed π - π , n - π , π -alkyl amino acids AspA:33, AlaA:146, LysA:147, PheA:28, LysA:117, AlaA:18, TyrA:32 with substituted aromatic ring and pyrrolidine pocket residue respectively and also formed a hydrogen binding interactions with SerA:17 (2.018 Å), ProA:34 (2.394 Å), AspA:38 (2.236 Å), TyrA:40 (2.102 Å) via sulfur, phenol, tetrazole & phenol, tetrazole substituents in the active site of Human PFKFB3 in complex with an N-aryl 6-aminoquinoxaline inhibitor (Fig. 7). Based on the docking energies, the tested compounds have more effective inhibitors in following PFKFB3, E2B receptor interactions with modern drug design.

ADMET predictions

Drug-likeness assessment

Further investigation, these analogues have been predicted for their drug-likeness scores and pharmacokinetic properties including ADMET parameters to verify that the designed molecules are viable drugs (Table 3, Fig. 8). Ghose, Veber, Egan, and Muegge filters, as well as Lipinski's rule of five, were not projected to be satisfied by the chosen compounds [44]. They had good lipophilic

Table 2 Docking confirmations with the active site of human breast adenocarcinoma (3HB4) and colon carcinoma (6IBZ)

Entry	$\Delta G/kJ$ (Kcal/mol)/nM ^{a,b}	Interacting residues (3HB4)/Bond distances (Å°)	Interacting residues (6IBZ)/Bond distances (Å°)
7a	– 10.85/11.07 – 8.21/53.94	Conventional Hydrogen bondings TyrA:40 (3.101), SerA:17 (2.942), LysA:117 (2.016), AlaA:146 (2.633), Tyr218 (2.165) Other interactions ValA:29, AlaA:18, PheA:28 (Pi-alkyl), AspA:38 (Pi- anion), AspA:119, AsnA:116 (F)	Hydrogen bondings SerA:17 (2.346), LysA:117 (1.948), AlaA:146 (2.795), TyrA:40 (2.648) Pi-cation bondings ArgA:75, AsnA:163 (F) CysA:154, GluA:166 (Pi-anion)
7e	– 11.83/2.14 – 7.15/84.30	Conventional Hydrogen bondings GlyX:15 (2.112), ValX:143 (2.120), IleX:14 (2.004), LysX: 159 (Pi-cation), IleX:14, ValX:225, PheS:259 (Pi-alkyl) Other amino acid residues Leu149, Tyr155, Gly144, Val143, Ser142, Asn90	Hydrogen bondings LysA:147 (1.348), AlaA:146 (1.678), LysA:117 (2.037), SerA:17 (1.975), AsnA:116 (2.084) Carbon hydrogen bondings AspA:33, ProA:34, ThrA:35, PheA:28 (Pi-Pi), ValA:29, TyrA:40 (Pi-alkyl)
7h	– 9.28/4.86 – 8.17/40.37	Conventional Hydrogen bondings AlaA:423 (2.947), AsnA:163 (2.395) Pi-anion bondings CysA:154 (2.846), GluA:166 (2.641) Pi-sigma bondings ValA:217, ValA:214, IleA:50, ValA:159, ValA:243, ValA:167	Hydrogen bondings Leu145, Gly90, Ala26, Carbon hydrogen condings Leu93, Gly92, Val29, Tyr66 (Pi-Pi), Asp33 (Pi-alkyl)
7i	– 11.50/3.72 – 10.12/29.34	Conventional Hydrogen bondings LysX:159 (2.183), AsnX:90 (2.165), TyrX:155 (2.013), GlyX:92 (1.326), SerX:142 (2.015), GlyX:144 (1.954), GlyX:94 (2.103) Carbon hydrogen bondings GlyX:186, CysX:185, ProX:187 Pi-Pi bonding PheX:192 Pi-alkyl bondings MetX:193, ValX:196, ValX:143, LeuX:149,	Hydrogen bondings SerA:17 (2.018), ProA:34 (2.394), AspA:38 (2.236), TyrA:40 (2.102) Carbon hydrogen bondings AspA:119, GlyA:15, AspA:30, SerA:145 Pi-alkyl bondings AlaA:146, LysA:147, PheA:28, LysA:117, AlaA:18, TyrA:32 AspA:33 (Pi-anion)
DXN	– 6.15/15.02	Conventional hydrogen bondings ThrA:48 (2.018); AsnA:69 (1.761); GluA:72 (2.430), AspA:124 (1.952); ArgA:75 (2.018); ArgA:189 (1.971) Pi-cation; LysA:168 (3.107) AlaA:125 (Pi-alkyl)	Gly130 (2.037), Met270 (1.679), Phe220 (1.554), Phe192, (1.990) Pi-cation Lys145, Asn60, Glu25, Pi-alkyl Asp124

^{a,b}Docking energy, dissociation constant values for 3HB4, 6IBZ proteins

properties, a consensus log Po/w value of 3.89, a high GI absorption score (17%), and were expected not to pass through the blood–brain barrier (BBB) or act as a P-gp substrate. The designed compounds are inhibitors of the cytochrome P450 isoenzymes CYP1A2 and CYP2C19, but not of CYP2D6 or, more specifically, CYP3A4, which is known to be the most abundant in the liver and an active participant in the metabolism of about 45% of known drugs [45, 46]. This was revealed by predictive data. The chosen compounds' skin permeation rates (Log K_p) ranged from – 8.44 to – 7.54 cm/s. Brain Or Intestinal estimated permeability approach (BOILED-Egg) was presented as an effective prediction model based on small molecule lipophilicity and polarity calculations in order to attain this objective. The BOILED-Egg model offers a rapid, simple, easily reproducible, and statistically unmatched method for predicting the excellent gastrointestinal absorption and

brain permeability of tiny compounds that may be used in drug development and discovery. The white part of the egg (the yolk) seems to be the physicochemical zone of substances that are likely to be absorbed by the GI tract. The Blood–Brain Barrier, shown in yellow, is a physical and chemical zone where chemicals that are likely to get into the brain (BBB permeation) are kept. As shown in Fig. 8, the compound **7b** with apparent oral bioavailability was superimposed over the BROILED-Egg. According to medicinal chemistry qualities, none of the compounds displayed any pain or brink alerts, and the bulk of them had generally good synthetic accessibility. The bioavailability radar plot, which took into account the following characteristics, shows that the chosen analogues exceed the pink area zone by one parameter, indicating that their predicted oral bioavailability is good. These characteristics include flexibility, lipophilicity, saturation, size, polarity, and solubility.

Experimental

Chemistry

All reactions were performed in anhydrous conditions using dry, recently distilled solvents that were obtained from SD Fine Chemicals or Sigma-Aldrich (India). Dichloromethane was distilled from CaH_2 just before use, while THF was produced from salt just before use. On silica gel plates (60F-254), thin-layer chromatography (TLC) was carried out under UV light (254 and 365 nm). On silica gel, flash chromatography was carried out (230–400 mesh). A Finnigan LCQ advantage max spectrometer was used to record the ESI mass spectra and a PerkinElmer GX FTIR spectrometer was used to record the IR spectrum of these substances. In DMSO-d_6 , CDCl_3 , NMR spectra (300–400 and 100–125 MHz for ^1H and ^{13}C , respectively) were captured with TMS serving as the internal standard. The following information is recorded for ^1H NMR: Hertz calculates the coupling constant J , chemical shift (ppm), multiplicity (s, singlet; d, doublet; t, triplet; q, quartet); and m, multiplet (dd, doublet of doublet) (Hz).

Synthesis of (2*S*,4*R*)-methyl-1-(2,4-difluoro-3-methoxybenzoyl)-4-hydroxy pyrrolidine-2-carboxylate **3**

(2*S*,4*R*)-methyl-4-hydroxypyrrolidine-2-carboxylate hydrochloride **2** (3.0 g, 16.7 mmol) was suspended in a mixture of pyridine (25 mL) and triethylamine (4.3 mL, 30.5 mmol). The resulting mass was stirred for 20 min at ambient temperature and filtered. The filtrate was cooled to $-5\text{ }^\circ\text{C}$, then a solution of 2,4-difluoro-3-methoxybenzoyl chloride **1** (3.0 g, 14.5 mmol) in dichloromethane DCM (15 mL) was added dropwise under nitrogen atmosphere. The reaction mass was stirred for 4 h at ambient temperature. After completion of the reaction (TLC), it was filtered and the filtrate was concentrated to dryness under reduced pressure. The residue was purified by silica gel chromatography using petroleum ether-ethyl acetate (8:2) as eluent to provide **3** (3.44 g, 75%) as pale yellow solid; mp: 75–77 $^\circ\text{C}$.

Synthesis of (2*S*,4*R*)-methyl-1-(2,4-difluoro-3-methoxybenzoyl)-4-((methylsulfonyl)oxy) pyrrolidine-2-carboxylate **4**

(2*S*,4*R*)-Methyl 1-(2,4-difluoro-3-methoxybenzoyl)-4-hydroxypyrrolidine-2-carboxylate **3** (3.00 g, 9.51 mmol) was taken in DCM (15 mL) and triethylamine (3.3 mL, 23.7 mmol) was added at 0 $^\circ\text{C}$ under nitrogen atmosphere. Then a solution of *p*-tolylsulfonyl chloride (3.02 g, 16 mmol)

in DCM (10 mL) was added dropwise and the resulting mixture was stirred at room temperature overnight. The reaction mixture was diluted with dichloromethane and washed with 10% aq. HCl (10 mL), saturated NaHCO_3 solution (10 mL), water (10 mL) followed by brine (15 mL) and dried over anhydrous sodium sulfate. The organic layer was separated and evaporated to obtain a light yellow oil, which was purified by flash chromatography over silica gel with petroleum ether-acetone (1:1) to provide **4** (3.66 g, 82%) as a white solid; mp: 65–67 $^\circ\text{C}$.

Synthesis of (2*S*,4*S*)-methyl-4-azido-1-(2,4-difluoro-3-methoxybenzoyl)pyrrolidine-2-carboxylate (**5**)

(2*S*,4*R*)-methyl-1-(2,4-difluoro-3-methoxybenzoyl)-4-((methoxy sulfonyl)oxy)pyrrolidine-2-carboxylate **4** (3.0 g, 6.39 mmol) was taken in anhydrous DMF (15 mL) under nitrogen atmosphere and sodium azide (5.2 g, 8.0 mmol) was added to the reaction mixture and stirred at 55 $^\circ\text{C}$ for overnight. After completion of the reaction, the reaction mass was partitioned between water (30 mL) and EtOAc (25 mL). After separation of the layers, the organic phase was washed with water followed by 0.1 M HCl (25 mL). The organic layer was then washed with brine, dried over MgSO_4 , separated, and evaporated to dryness under reduced pressure. The resulting oily residue was chromatographed with hexane–EtOAc (3:1) to provide **5** (1.82 g, 84%) as a yellow oil, which solidified upon standing; mp: 145–147 $^\circ\text{C}$.

4.5 General procedure for the synthesis of (2*S*,4*S*)-methyl-1-(2,4-difluoro-3-methoxybenzoyl)-4-(((1-substitutedphenyl-1*H*-tetrazol-5-yl)thio)methyl)-1*H*-1,2,3-triazol-1-yl)pyrrolidine-2-carboxylate (**7**)

A series of 1-phenyl-5-(prop-2-yn-1-ylthio)-1*H*-tetrazoles **6a–i** were prepared starting from phenyl isothiocyanate which was reacted with sodium azide to give 1-phenyl-1*H*-tetrazole-5-thiol. Then the solution of 1-phenyl-1*H*-tetrazole-5-thiol (2.1 mmol), propargyl bromide (1.2 mmol), and tetrabutylammonium bromide in a mixture of triethylamine (4 mL) and DCM (6 mL) was stirred at ambient temperature for 4 h. After completion of the reaction (TLC), the reaction mixture was poured into ice-cold water (15 mL), and the solid product was filtered off, dried, and purified by column chromatography using ethyl acetate–hexane (2:8) (87%). A mixture of tetrazole **6** (1.2 mmol), pyrrolidine azide **5** (1.2 mmol), copper sulfate, and L-sodium ascorbate in $\text{DMF-H}_2\text{O}$ (10 mL, 3:2 ratio) was stirred at room temperature for 4 h. After completion of the reaction, the mixture was poured into ice water (30 mL) and extracted

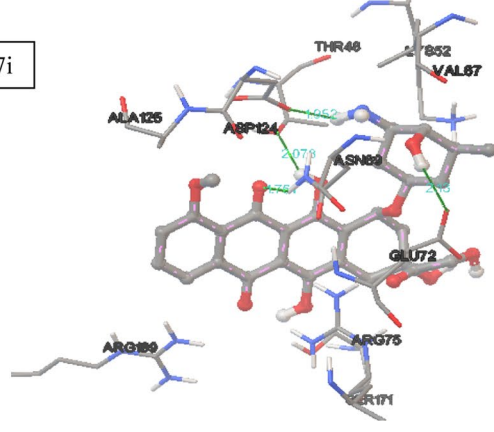
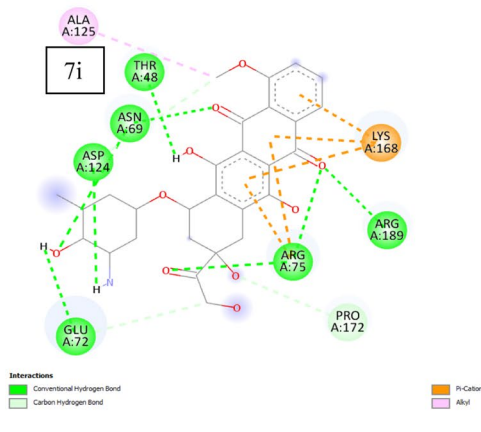
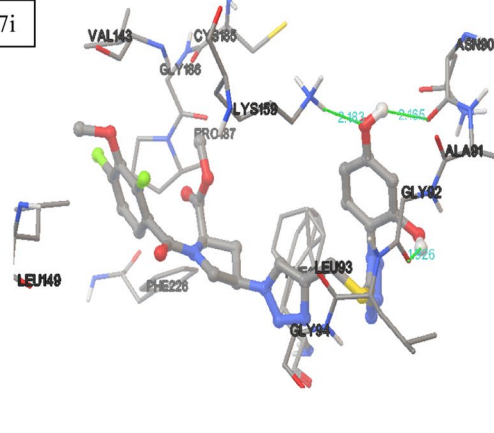
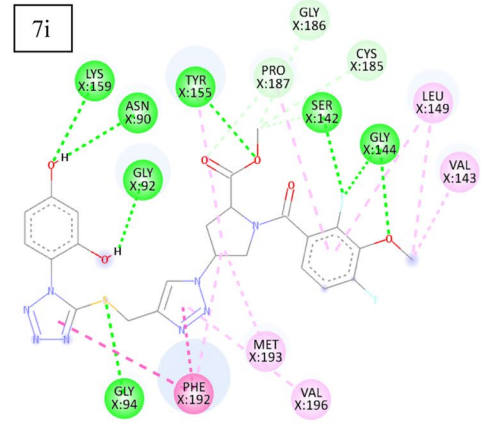
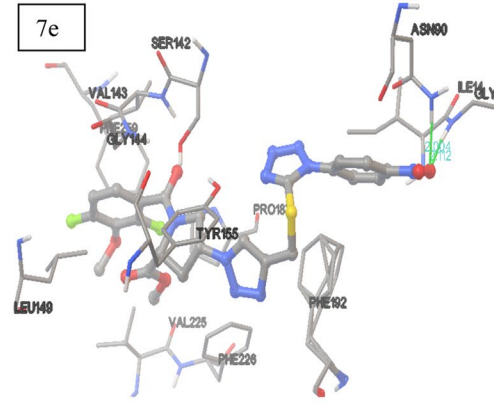
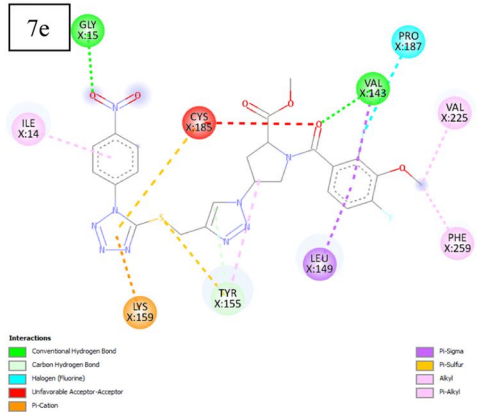
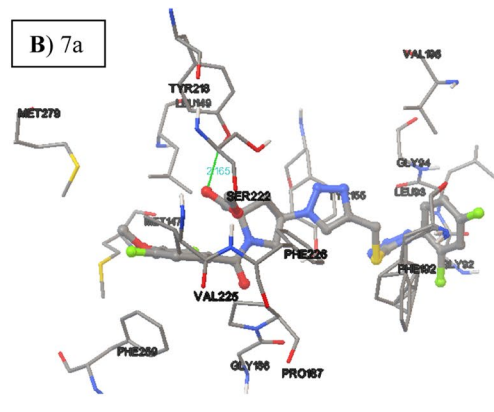
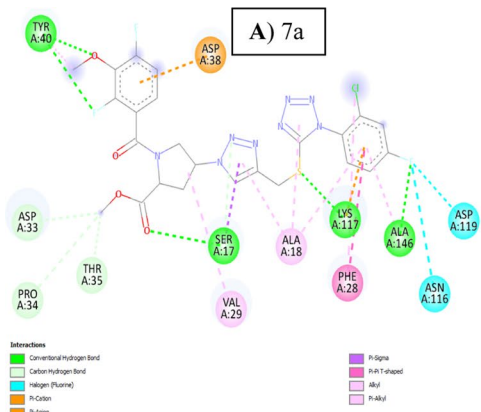


Fig. 6 Validation of the docking study into active site of proteins 3HB4; A) 2D-structures of **7b**, **7e**, **7i**, **DXN**; B) 3D-side chine flexibility of **7b**, **7e**, **7i**, **DXN** against human breast adenocarcinoma (**3HB4**). Probable binding mode of compounds (lime color) in the active site of a subunit from *M. tuberculosis*; the hydrogen bond interactions are shown in light green color dotted lines, the important residues are shown in cyan/aqua color, and protein cartoon is represented according to the secondary structure of the protein

with 2 X 25 mL of ethyl acetate. The combined organic extract was washed twice with a saturated ammonium chloride (20 mL) and with brine (20 mL), dried over Na₂SO₄, and concentrated to dryness under reduced pressure. The residue was purified by column chromatography on silica gel using *n*-hexane: ethyl acetate (7:3) as eluent.

(2S, 4S)-methyl-1-(2,4-difluoro-3-methoxybenzoyl)-4-(4-(((1-(2,4-difluorophenyl)-1H-tetrazol-5-yl)thio)methyl)-1H-1,2,3-triazol-1-yl)pyrrolidine-2-carboxylate (7a)

White solid; yield 79.8%; mp: 165–167 °C. ¹H NMR (DMSO-*d*₆, 400 MHz): δ 0.93 (1H, m, CH₂), 1.16 (1H, m, CH₂), 1.65 (1H, m, CH), 2.81 (3H, s, OCH₃), 3.31 (2H, d, *J* = 8.4 Hz, CH₂), 3.72 (3H, s, OCH₃), 2.65 (1H, t, *J* = 7.8 Hz, CH), 4.88 (2H, s, SCH₂), 6.81 (1H, d, *J* = 7.2 Hz, ArH), 7.37 (1H, dd, *J* = 7.2, 3.0 Hz, ArH), 7.54 (1H, d, *J* = 7.2 Hz, ArH), 8.03 (1H, dd, *J* = 7.2, 3.0 Hz, ArH), 8.44 (1H, d, *J* = 7.4 Hz, ArH), 8.62 (1H, s, triazole H). ¹³C NMR (DMSO-*d*₆, 100 MHz): δ 24.02, 34.90, 48.56, 51.79, 55.27, 61.98, 62.28, 102.47, 119.78, 123.59, 125.66, 128.26, 128.40, 135.78, 142.35, 150.03, 151.48, 156.23, 160.06, 161.41, 172.03, 176.17. IR (KBr, cm⁻¹): 3037.75 (=CH), 2927.23, 2908.34 (CH), 1699.86 (ester C=O), 1637.56 (amide C=O), 1597.22, 1514.65 (Ar–C=C), 1406.41 (C=N), 1368.12 (N=N), 1209.47 (C–S–C), 1103.49 (C–O–C), 1040.23 (C–F); *m/z*: 593 [M + H]⁺.

(2S,4S)-methyl-4-(4-(((1-(2-chloro-4-fluorophenyl)-1H-tetrazol-5-yl)thio)methyl)-1H-1,2,3-triazol-1-yl)-1-(2,4-difluoro-3-methoxybenzoyl)pyrrolidine-2-carboxylate (7b)

White crystalline solid; yield 76.1%; mp: 160–162 °C. ¹H NMR (DMSO-*d*₆, 400 MHz): δ 1.57 (1H, m, CH₂), 1.79 (1H, t, *J* = 7.8 Hz, CH₂), 2.14 (1H, m, CH), 2.91 (2H, d, *J* = 8.0 Hz, CH₂), 3.23 (3H, s, OCH₃), 3.43 (3H, s, OCH₃), 3.67 (1H, t, *J* = 7.8 Hz, CH), 4.77 (2H, s, SCH₂), 7.24 (1H, d, *J* = 7.3 Hz, ArH), 7.75 (1H, dd, *J* = 7.4, 3.2 Hz, ArH), 7.79 (1H, d, *J* = 7.3 Hz, ArH), 7.85 (1H, dd, *J* = 7.4, 3.2 Hz, ArH), 8.79 (1H, d, *J* = 7.3 Hz, ArH), 9.46 (1H, s, triazole H). ¹³C NMR (DMSO-*d*₆, 100 MHz): δ 24.71, 35.41, 40.86, 54.98, 57.98, 61.03, 62.50, 102.56, 113.34, 126.03, 128.40, 128.78, 135.68, 139.82, 147.92,

150.19, 151.32, 155.51, 165.06, 166.02, 171.20. IR (KBr, cm⁻¹): 3042.49 (=CH), 2966.64, 2931.91 (CH), 1699.22 (ester C=O), 1672.11 (amide, C=O), 1594.50, 1539.93 (Ar–C=C), 1427.03 (C=N), 1279.37 (C–S–C), 1173.97 (C–O–C), 1104.69 (C–F), 1021.49 (C–F) *m/z*: 609 [M + H]⁺.

(2S, 4S)-methyl-1-(2,4-difluoro-3-methoxybenzoyl)-4-(4-(((1-(2,4-dimethoxyphenyl)-1H-tetrazol-5-yl)thio)methyl)-1H-1,2,3-triazol-1-yl)pyrrolidine-2-carboxylate (7c)

Pale yellow oil; yield 70.2%; ¹H NMR (DMSO-*d*₆, 400 MHz): δ 2.12 (1H, m, CH₂), 2.27 (1H, m, CH₂), 2.69 (1H, m, CH), 2.78 (1H, d, *J* = 7.3 Hz, CH₂), 2.89 (1H, d, *J* = 7.3 Hz, CH₂), 3.07 (6H, s, OCH₃), 3.24 (3H, s, OCH₃), 3.55 (3H, s, OCH₃), 3.84 (1H, t, *J* = 7.4 Hz, CH), 4.95 (2H, s, SCH₂), 7.03 (1H, s, ArH), 7.24 (1H, d, *J* = 7.1 Hz, ArH), 7.41 (1H, t, *J* = 7.4 Hz, ArH), 7.86 (1H, d, *J* = 7.1 Hz, ArH), 8.23 (2H, d, *J* = 7.4 Hz, ArH), 8.83 (1H, s, triazole H). ¹³C NMR (DMSO-*d*₆, 100 MHz): δ 23.81, 36.62, 46.20, 51.27, 56.31, 59.60, 62.37, 64.83, 109.37, 113.21, 116.39, 124.38, 127.64, 128.02, 131.58, 137.22, 146.85, 154.39, 157.45, 158.88, 172.63, 174.14. IR (KBr, cm⁻¹): 3051.26 (=CH), 2935.33, 2875.15 (CH), 1706.45 (ester C=O), 1653.91 (amide C=O), 1585.62, 1524.29 (Ar–C=C), 1422.47 (C=N), 1215.10 (C–S–C), 1166.01 (C–O–C), 1034.77 (C–F). *m/z*: 617 [M + H]⁺.

(2S,4S)-methyl-1-(2,4-difluoro-3-methoxybenzoyl)-4-(4-(((1-(*p*-tolyl)-1H-tetrazol-5-yl)thio)methyl)-1H-1,2,3-triazol-1-yl)pyrrolidine-2-carboxylate (7d)

Colourless gummy liquid; yield: 68%; ¹H NMR (DMSO-*d*₆, 400 MHz): δ 1.49 (1H, m, CH₂), 1.78 (1H, m, CH), 2.20 (3H, s, CH₃), 2.57 (1H, dd, *J* = 7.4, 2.7 Hz, CH), 3.37 (3H, s, OCH₃), 3.62 (1H, d, *J* = 7.5 Hz, CH₂), 3.70 (1H, d, *J* = 7.5 Hz, CH₂), 4.03 (3H, s, OCH₃), 4.37 (1H, t, *J* = 7.8 Hz, CH), 4.90 (2H, s, SCH₂), 7.33 (1H, d, *J* = 7.4 Hz, ArH), 7.72 (2H, d, *J* = 7.3 Hz, ArH), 8.81 (2H, d, *J* = 7.3 Hz, ArH), 8.87 (1H, dd, *J* = 7.4, 3.1 Hz, ArH), 9.37 (1H, s, triazole H). ¹³C NMR (DMSO-*d*₆, 100 MHz): δ 22.92, 29.10, 35.27, 48.79, 51.56, 56.54, 62.22, 63.50, 104.01, 118.12, 119.28, 121.43, 122.73, 124.06, 130.18, 132.77, 142.07, 151.10, 155.66, 163.22, 171.45, 172.38. IR (KBr, cm⁻¹): 3032.59 (=CH), 2956.22, 2896.71 (CH), 1713.31 (ester C=O), 1671.06 (amide C=O), 1551.05, 1472.97 (Ar–C=C), 1408.22 (C=N), 1323.46 (N=N), 1258.64 (C–S–C), 1171.07 (C–O–C), 1026.60 (C–F), 992.61 (C–F) *m/z*: 571 [M + H]⁺.

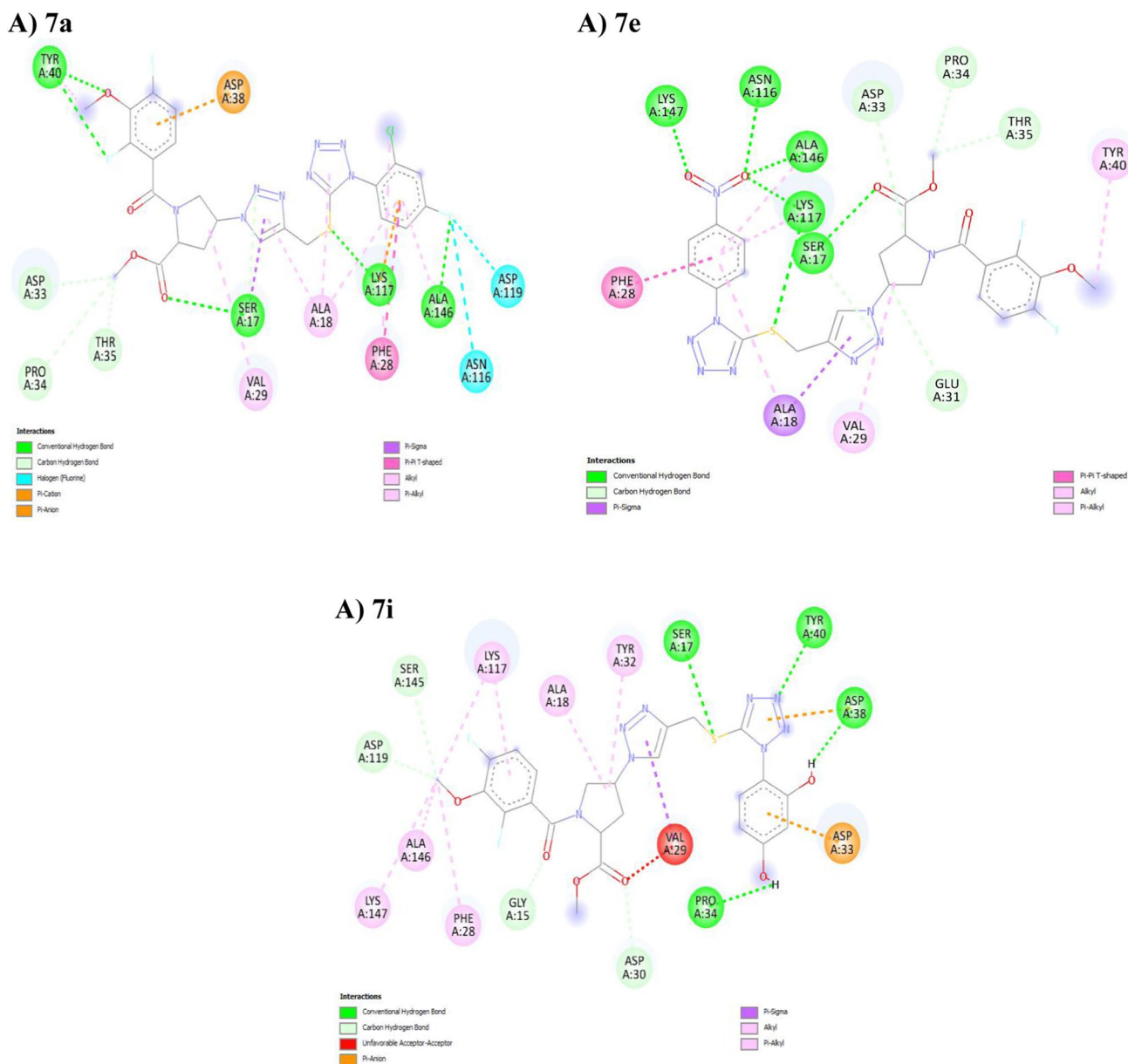


Fig. 7 A 2D-structures of **7b**, **7e**, and **7i** against human colon carcinoma (**6IBZ**)

(2S, 4S)-methyl-1-(2,4-difluoro-3-methoxybenzoyl)-4-(4-(((1-(4-nitrophenyl)-1H-tetrazol-5-yl)thio)methyl)-1H-1,2,3-triazol-1-yl)pyrrolidine-2-carboxylate (7e**)**

Yellow crystalline solid; yield 92%; mp: 145–147 °C. ^1H NMR (DMSO- d_6 , 400 MHz): δ 2.06 (1H, m, CH_2), 2.25 (1H, m, CH_2), 2.74 (1H, m, CH), 2.71 (1H, d, $J=7.4$ Hz, CH_2), 2.79 (1H, d, $J=7.4$ Hz, CH_2), 3.26 (3H, s, OCH_3), 3.58 (3H, s, OCH_3), 3.79 (1H, t, $J=7.8$ Hz, CH), 4.62 (2H, s, SCH_2), 7.01 (1H, d, $J=7.0$ Hz, ArH), 7.36 (1H, t, $J=7.5$ Hz, ArH), 7.84 (1H, s, triazole H), 8.21 (2H, d, $J=7.4$ Hz, ArH), 8.51 (2H, d, $J=7.4$ Hz, ArH). ^{13}C NMR (DMSO- d_6 , 100 MHz): δ

22.97, 25.79, 37.79, 49.81, 51.36, 58.83, 60.75, 114.28, 114.47, 118.71, 125.56, 127.76, 128.02, 130.85, 137.66, 142.35, 145.58, 157.36, 158.50, 172.68, 176.13. IR (KBr, cm^{-1}): 3022.99 (=CH), 2927.05, (CH), 1730.56 (ester C=O), 1640.15 (amide C=O), 1601.39, 1512.78 (Ar-C=C), 1429.07 (C=N), 1336.73 (N=N), 1261.23 (C-S-C), 1135.37 (C-O-C), 1030.78 (C-F), 983.11 (CF) m/z : 602 [M+H] $^+$.

Table 3 Pharmacokinetic predictions of final triazolyl-tetrazole ligands **7a–i**

S. no	7a	7b	7c	7d	7e	7f	7g	7h	7i
No of heavy atoms	41	41	43	40	42	42	39	40	41
Num. atom. heavy atoms	22	22	22	22	22	22	22	22	22
Fraction Csp ³	0.29	0.29	0.35	0.32	0.29	0.29	0.29	0.29	0.29
Num. rotatable bonds	10	10	12	10	11	11	10	10	10
Num. H-bond acceptors	13	12	13	11	13	13	11	12	13
Num. H-bond donors	0	0	0	0	0	0	0	1	2
TPSA (Å ²)	155.54	155.54	173.91	155.45	201.27	201.27	155.45	175.68	195.91
Lipophilicity [Log Po/w]	3.84	3.87	3.88	3.76	3.17	3.46	3.89	3.65	3.06
Water solubility/Log S	− 5.88	− 6.43	− 5.99	− 6.04	− 6.25	− 6.46	− 5.67	− 5.72	− 5.78
Lipinski	No	No	No	No	No	No	No	No	No
Ghose	No	No	No	No	No	No	No	No	No
Veber	No	No	No	No	No	No	No	No	No
Egan	No	No	No	No	No	No	No	No	No
Muegge	No	No	No	No	No	No	No	No	No
Bioavailability Score	0.17	0.17	0.17	0.17	0.17	0.17	0.17	0.17	0.17
Synthetic accessibility	4.60	4.64	4.97	4.70	4.70	4.83	4.58	4.64	4.70
Pharmacokinetics									
GI absorption	Low	Low	Low	Low	Low	Low	Low	Low	Low
BBB permeant	No	No	No	No	No	No	No	No	No
P-gp substrate	No	No	No	No	No	Yes	No	Yes	Yes
CYP1A2 inhibitor	No	No	No	No	No	No	No	No	No
CYP2C19 inhibitor	No	No	No	No	No	No	No	No	No
CYP2C9 inhibitor	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	No
CYP2D6 inhibitor	No	No	No	No	No	No	No	No	No
CYP3A4 inhibitor	No	No	Yes	Yes	Yes	Yes	Yes	Yes	No
Log K _p (cm/s)	− 7.82	− 7.54	− 8.15	− 7.57	− 8.14	− 8.14	− 7.74	− 8.10	− 8.44

For the classification endpoints, the prediction probability values are transformed into six symbols that were divided into three empirical-based decision states visually represented with different colours, including (i) excellent/green: 0–0.1 and 0.1–0.3, (ii) medium/yellow: 0.3–0.5 and 0.5–0.7, (iii) red/poor: 0.7–0.9 and 0.9–1.0

TPSA topological polar surface area, BBB blood–brain barrier (0=log BBB ≤ − 1, 1=log BBB > − 1), Pgp P-glycoprotein (0=non inhibitor, 1=inhibitor), CYP1A2 inhibitor (0=non inhibitor, 1=inhibitor), CYP2C19 inhibitor (0=non inhibitor, 1=inhibitor), CYP2C9 inhibitor (0=non inhibitor, 1=inhibitor), CYP2D6 inhibitor (0=non inhibitor), CYP3A4 inhibitor (0=non inhibitor, 1=inhibitor), Lipinski rule < 2 violations

(2S, 4S)-methyl-1-(2,4-difluoro-3-methoxybenzoyl)-4-(4-(((1-(2-nitrophenyl)-1H-tetrazol-5-yl)thio)methyl)-1H-1,2,3-triazol-1-yl)pyrrolidine-2-carboxylate (7f)

Yellow crystalline solid, yield: 85%, mp: 132–134 °C. ¹H NMR (DMSO-*d*₆, 400 MHz) δ ppm: 2.03 (1H, m, CH₂), 2.21 (1H, m, CH₂), 2.73 (1H, m, CH), 2.78 (1H, d, *J* = 7.3 Hz, CH₂), 2.96 (1H, d, *J* = 7.3 Hz, CH₂), 3.21 (3H, s, OCH₃), 3.45 (3H, s, OCH₃), 3.80 (1H, t, *J* = 7.7 Hz, CH), 4.68 (2H, s, SCH₂), 7.06 (1H, d, *J* = 6.8 Hz, ArH), 7.30 (1H, t, *J* = 7.7 Hz, ArH), 7.73 (1H, dd, *J* = 7.5, 3.2 Hz, ArH), 7.88 (1H, dd, *J* = 7.5, 3.1 Hz, ArH), 8.05 (2H, m, ArH), 8.50 (1H, s, triazole H). ¹³C NMR (DMSO-*d*₆, 100 MHz): δ 24.13, 26.37, 35.60, 48.44, 50.37, 57.66, 61.91, 114.17,

115.33, 118.45, 124.88, 127.30, 128.21, 130.92, 135.55, 142.37, 145.68, 156.69, 157.11, 171.37, 173.94. IR (KBr, cm^{−1}): 3081.31 (=CH), 2934.79, 2834.30 (CH), 1719.60 (ester C=O), 1646.18 (amide C=O), 1578.90 (Ar–C=C), 1431.90 (C=N), 1328.11 (N=N), 1224.83 (C–S–C), 1142.66 (C–O–C), 1026.81 (C–F), 977.52 (C–F) *m/z*: 602 [M+H]⁺.

(2S, 4S)-methyl-1-(2,4-difluoro-3-methoxybenzoyl)-4-(4-(((1-phenyl-1H-tetrazol-5-yl)thio)methyl)-1H-1,2,3-triazol-1-yl)pyrrolidine-2-carboxylate (7g)

Brown color solid; yield 65%; mp: 161–163 °C. ¹H NMR (CDCl₃, 400 MHz): δ 2.20 (1H, m, CH₂), 2.41 (1H, m, CH₂), 2.82 (1H, d, *J* = 7.0 Hz, CH₂), 3.09 (1H, m, CH), 3.30 (1H,

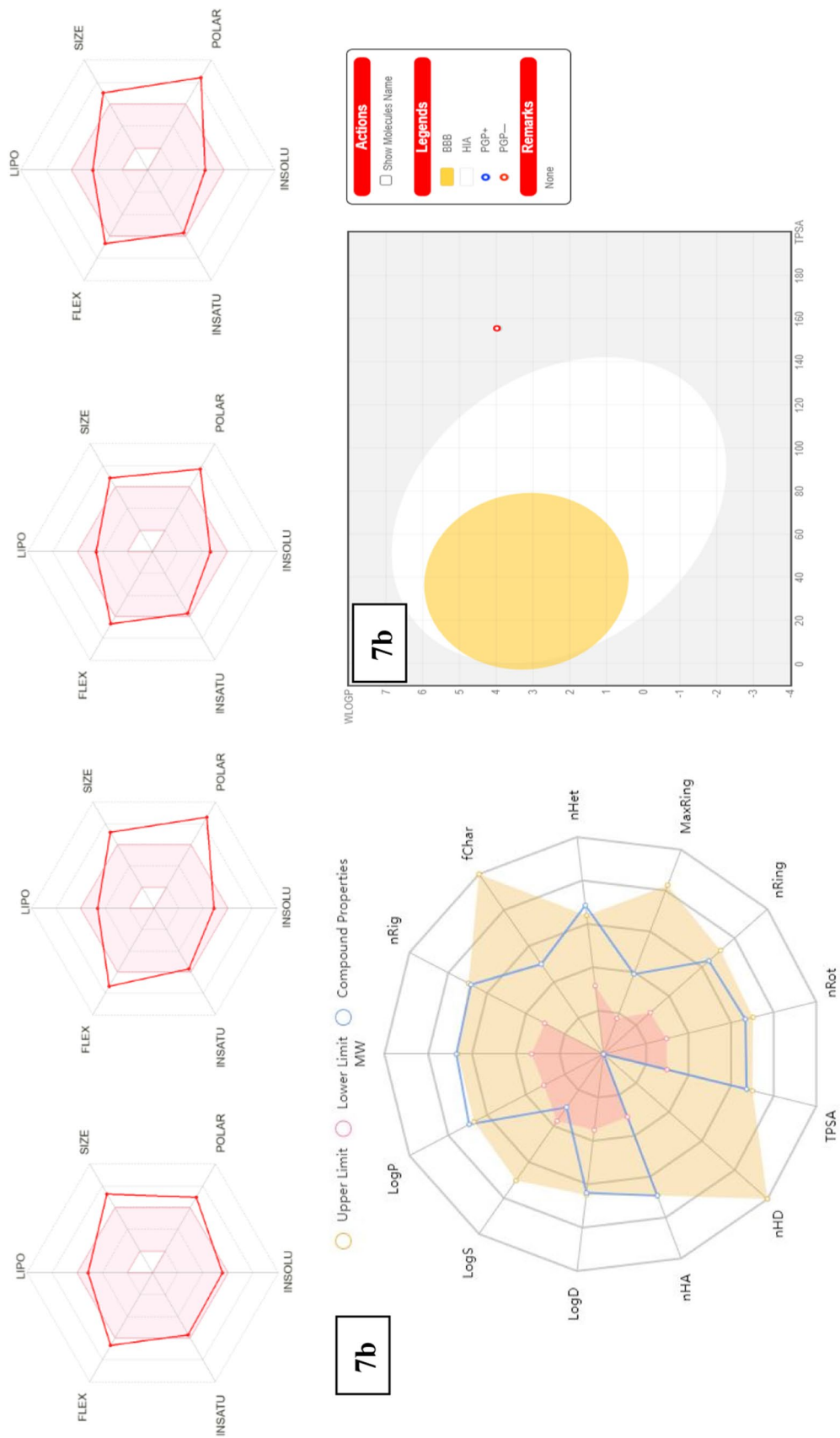


Fig. 8 SwissADME platform of **7b**, **7f**, **7h** and **7i** respectively. The colored zone (pink) was the suitable physicochemical space for oral bioavailability and lines and dots representing each compound are shown in the figure. BOILED-Egg predictive model for the compounds **7b**

d, $J=7.0$ Hz, CH₂), 3.55 (3H, s, OCH₃), 3.83 (3H, s, OCH₃), 3.96 (1H, t, $J=7.5$ Hz, CH), 4.93 (2H, s, SCH₂), 6.83 (1H, d, $J=7.2$ Hz, ArH), 7.12 (3H, m, ArH), 7.28 (1H, d, $J=7.2$ Hz, ArH), 7.62 (1H, t, $J=7.4$ Hz, ArH), 7.94 (1H, d, $J=7.2$ Hz, ArH), 8.77 (1H, s, triazole H). ¹³C NMR (CDCl₃, 100 MHz): δ 23.21, 35.60, 43.67, 52.75, 55.33, 62.50, 63.97, 116.33, 118.43, 122.80, 124.55, 127.28, 135.61, 148.67, 152.34, 155.81, 157.33, 161.73, 170.39, 173.61. IR (KBr, cm⁻¹): 3043.11 (=CH), 2931.37, (CH), 1727.34 (ester C=O), 1638.91 (C=O), 1606.46, 1534.24 (Ar-C=C), 1416.27 (C=N), 1343.44 (N=N), 1254.37 (C-S-C), 1144.27 (C-O-C), 1025.77 (C-F), 979.24 (C-F) m/z : 557 [M+H]⁺.

4.13 (2S, 4S)-Methyl-1-(2,4-difluoro-3-methoxybenzoyl)-4-((1-(2-hydroxyphenyl)-1H-tetrazol-5-yl)thio)methyl)-1H-1,2,3-triazol-1-ylpyrrolidine-2-carboxylate (7h) Colourless gummy liquid; yield 81%; ¹H NMR (DMSO-*d*₆, 400 MHz): δ 2.16 (1H, m, CH₂), 2.31 (1H, m, CH₂), 2.76 (1H, m, CH), 3.13 (1H, d, $J=7.1$ Hz, CH₂), 3.25 (1H, d, $J=7.1$ Hz, CH₂), 3.50 (3H, s, OCH₃), 3.71 (3H, s, OCH₃), 3.89 (1H, t, $J=7.4$ Hz, CH), 5.10 (2H, s, SCH₂), 5.67 (1H, brs, OH), 6.86 (1H, d, $J=7.0$ Hz, ArH), 7.06 (1H, dd, $J=7.8$, 3.1 Hz, ArH), 7.20 (1H, dd, $J=7.8$, 3.1 Hz, ArH), 7.45 (1H, d, $J=7.0$ Hz, ArH), 7.64 (1H, t, $J=7.5$ Hz, ArH), 7.94 (1H, d, $J=7.0$ Hz, ArH), 8.86 (1H, s, triazole H). ¹³C NMR (DMSO-*d*₆, 100 MHz): δ 22.31, 36.20, 44.67, 52.66, 56.45, 61.56, 63.41, 106.70, 118.72, 119.94, 123.85, 124.61, 127.11, 135.62, 148.91, 153.44, 154.26, 156.70, 157.92, 161.46, 171.04, 173.37. IR (KBr, cm⁻¹): 3458.24 (OH), 3084.34 (=CH), 2963.55, 2874.23 (CH), 1716.94 (ester C=O), 1661.18 (amide C=O), 1555.60 (Ar-C=C), 1428.63 (C=N), 1320.83 (N=N), 1204.77 (C-S-C), 1166.18 (C-O-C), 1020.61 (C-F) m/z : 573 [M+H]⁺.

(2S, 4S)-methyl-1-(2,4-difluoro-3-methoxybenzoyl)-4-(((1-(2,4-dihydroxyphenyl)-1H-tetrazol-5-yl)thio)methyl)-1H-1,2,3-triazol-1-ylpyrrolidine-2-carboxylate (7i)

Pale yellow solid; yield 88%; mp: 128–130 °C. ¹H NMR (DMSO-*d*₆, 400 MHz): δ 1.48 (1H, m, CH₂), 1.62 (1H, m, CH₂), 2.07 (1H, m, CH), 2.72 (1H, d, $J=7.2$ Hz, CH₂), 2.81 (1H, d, $J=7.2$ Hz, CH₂), 2.97 (3H, s, OCH₃), 3.52 (3H, s, OCH₃), 3.66 (1H, t, $J=7.5$ Hz, CH), 4.90 (2H, s, SCH₂), 5.50 (2H, brs, OH), 6.58 (1H, s, ArH), 6.75 (1H, d, $J=7.0$ Hz, ArH), 7.21 (1H, t, $J=7.4$ Hz, ArH), 7.42 (1H, d, $J=7.0$ Hz, ArH), 7.69 (1H, d, $J=7.0$ Hz, ArH), 8.21 (1H, s, triazole H). ¹³C NMR (DMSO-*d*₆, 100 MHz): δ 25.16, 37.26, 41.70, 51.79, 56.69, 60.50, 65.27, 105.76, 117.02, 119.78, 123.59, 128.45, 135.78, 141.54, 148.82, 153.45, 154.38, 156.87, 157.56, 161.46, 172.80, 175.67. IR (KBr, cm⁻¹): 3486.30 (OH), 3055.09 (=CH), 2982.59, 2896.08 (CH), 1712.63 (ester C=O), 1668.10 (ester C=O),

1598.62, 1552.60 (Ar-C=C), 1467.59 (C=N), 1325.73 (N=N), 1256.93 (C-S-C), 1173.20 (C-O-C), 1103.32 (C-F), 1026.40 (C-F) m/z : 589 [M+H]⁺.

General experimental details of biological activity evaluation, computational docking studies and copies of ¹H NMR, ¹³C NMR, and IR spectrums are included in supporting information.

Conclusion

New series of 1,2,3-triazoles & tetrazoles incorporated pyrrolidine derivatives have been prepared and evaluated for their anticancer activity. In the current study, novel pyrrolidine coupled to dual heterocycles was conveniently synthesised, and all the title compounds were examined using ¹H NMR, ¹³C NMR, IR, and mass spectrometry. The results of this study demonstrate that the compounds **7a**, **7b**, **7e**, **7f**, **7h**, and **7i** have a promising anticancer activity against Hela (human cervical epithelioid carcinoma), MCF-7 (human breast adenocarcinoma), HCT-116 (human colon carcinoma), and HepG2 cells (human caucasian hepatocyte carcinoma). In particular, the prepared scaffolds **7b** and **7i**, showed remarkable anticancer activity against HCT-116 cancer cell line with IC₅₀ values 1.48 ± 0.08 μM, 1.38 ± 0.06 μM respectively. Molecular docking simulations of targeting the active site of human breast and colon carcinoma proteins (3HB4, 6IBZ) were used in addition to the in vitro analysis to explore potential interactions of these analogues with the receptor. On the other hand, the docking results also correlate well with the biological activity and compounds with potent inhibitory activities towards anticancer growths were further evaluated for their ADMET and physicochemical properties. Upon keen observation of the obtained results, it can be concluded that these molecules become lead molecules for further synthetic and biological evaluation.

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Author contributions SKG, AAK, AJ in-charge of synthesis, write up and characterization studies; TRA and KC write up the computational molecular docking calculations and biological activities.

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Data availability Not Applicable.

Declarations

Conflict of interest The authors declare no conflict of interest.

Consent for publication Not applicable.

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