ORIGINAL ARTICLE

The antioxidant and neuroprotective efects of the *Psychotria camptopus* **Verd. Hook. (Rubiaceae) stem bark methanol extract contributes to its antiepileptogenic activity against pentylenetetrazol kindling in male Wistar rats**

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Abstract

A substantial number of epileptic patients are resistant to the current medication thus necessitating the search for alternative therapies for intractable forms of the disease. Previous studies demonstrated the acute anticonvulsant properties of the methanol extract of the stem bark of *Psychotria camptopus* (MEPC) in rats. This study investigated the efects of MEPC on pentylenetetrazole-kindled Wistar rats. Kindling was induced by intraperitoneal injection of pentylenetetrazole (37.5 mg/ kg) on every alternate day, 1 h after each daily oral pretreatment of rats $(8 \le n \le 10)$ with MEPC (40, 80 and 120 mg/kg), vehicle or diazepam (3 mg/kg) for 43 days. The kindling development was monitored based on seizure episodes and severity. Rats' brains were collected on day 43 for the determination of oxidative stress parameters. The histomorphological features and neuronal cell viability of the prefrontal cortex (PFC) and hippocampus were also assessed using H&E and Cresyl violet stains. Chronic administration of pentylenetetrazole time-dependently decreased the latency to myoclonic and generalized seizures, and increased seizure scores and the number of kindled rats. MEPC and diazepam signifcantly increased the latencies to myoclonic jerks and generalized tonic-clonic seizures. These substances also reduced seizure score and the number of rats with PTZ-kindling. MEPC improved glutathione status and decreased lipid peroxidation in the brains of kindled rats. MEPC also exhibited neuroprotection against pentylenetetrazole-induced hippocampal and PFC neuronal damages. These results suggest that *P. camptopus* has antiepileptogenic activity, which might be related to the augmentation of antioxidant and neuroprotective defense mechanisms, and further confrm its usefulness in the management of epilepsy.

Keywords Epileptogenesis · Pentylenetetrazole kindling · Oxidative stress · Neuroprotection · *Psychotria camptopus*

Abbreviations

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Introduction

Epilepsy is a neurological disorder characterized by recurrent seizures, which has been ascribed to abnormal excessive and synchronous neuronal activity in the brain (Fisher et al. [2017](#page-11-0)). It has been described clinically as occurrence of at least two consecutive unprovoked seizures at interval of 24 h or one unprovoked seizure with the possibility of reoccurrence after ten years (Scheffer et al. [2017\)](#page-11-1). The etiological insult that converts normal brain into an epileptic brain entails a series of epileptogenic events known as epileptogenesis. The latent period between the initial insult and the development of spontaneous recurrent seizure has been reported to range from several weeks in animals to years in humans (Pitkänen et al. [2015](#page-11-2); Devinsky et al. [2018](#page-11-3)). This period is usually characterized by brain remodeling and other neurobiochemical changes in some brain regions that prime the organism for seizure development.

Cumulative evidences from rodent models of epilepsy showed that epileptogenic events and seizure activity are sustained by neuroinfammatory and oxidative stress mechanisms (Vezzani et al. [2012;](#page-12-0) Taiwe et al. [2016](#page-12-1); Ravizza et al. [2017](#page-11-4); Rana and Musto [2018;](#page-11-5) Singh et al. [2018,](#page-12-2) [2019](#page-12-3)). In a healthy brain, the levels of reactive oxygen species (ROS) and reactive nitrogen species (RNS), are known to be very low and well-regulated to prevent their harmful effects on biological molecules and cell structures (Singh et al. [2018](#page-12-2)). However, in an epileptic condition, activation of pro-infammatory pathways and the production of exponential amount of ROS and RNS, and loss of antioxidant mechanisms occurs (Nigar et al. [2016;](#page-11-6) Pearson-Smith and Patel [2017](#page-11-7)). This leads to accumulation of ROS and RNS in the brain, which in turns promote membrane lipid peroxidation, mitochondrial DNA damage, depletion of antioxidant defense system and apoptosis (Nigar et al. [2016;](#page-11-6) Pearson-Smith and Patel [2017](#page-11-7); Roganovic et al. [2019](#page-11-8)). These cellular and molecular changes result in increased seizure activity, neurodegeneration and abnormal synaptic rewiring, which might contribute to disease development and neurological complications associated with epilepsy (Devinsky et al. [2018](#page-11-3); Roganovic et al. [2019](#page-11-8)).

PTZ-kindling is a well-established model to elucidate the pathophysiology of epilepsy and for development of new antiepileptic drugs. One of the advantages of the PTZkindling is that it models epileptogenic events and chronic intractable epilepsy (Dhir [2012;](#page-11-9) Erkec [2015\)](#page-11-10), making it an ideal paradigm for novel antiepileptic drugs discovery. It is known that PTZ kindling induces seizures that are relevant to the human temporal lobe epilepsy (TLE) and generalized tonic-clonic convulsions (Kumar and Kumar [2017\)](#page-11-11). Studies revealed that PTZ-kindling induces neuronal hyperexcitability, neuroinfammation, oxidative stress and cell loss in specifc brain regions including the hippocampus and prefrontal cortex (Erkec [2015;](#page-11-10) Bascuñana et al. [2016;](#page-11-12) Kola et al. [2017](#page-11-13); Zhu et al. [2017;](#page-12-4) Samokhina and Samokhin [2018\)](#page-11-14).

Over last decade, epilepsy researches have emphasized on the elucidation of molecular pathways and cellular mechanisms that can be targeted in the development of novel therapies. Even with the availability of a stream of new antiepileptic drugs, of which some act on specifc molecular targets, many patients still do not achieve adequate seizure control. Indeed, more than 30% of epileptic patients fail to relieve under current medication (Wahab [2010](#page-12-5)). These drugs are generally anti-symptomatic than disease modifying molecules, and could be classifed as anti-seizures instead of antiepileptic drugs (Kubova [2016a;](#page-11-15) Saletti et al. [2019](#page-11-16)). The inability of these drugs to modify the core pathological abnormality in epileptic brains, coupled with numerous adverse efects, depressive and cognitive comorbidities indicate the need to search for new medicines for this disease. It is worthy to note that current research trends are directed towards the development of molecules with antioxidant, anti-infammatory and neuroprotective properties for patients with epilepsy (Tang et al. [2017;](#page-12-6) Saletti et al. [2019](#page-11-16)). Indeed, previous preclinical studies have shown that medicinal plants and plant-derived compounds with antioxidant and anti-infammatory properties exhibited promising antiepileptic efect (Taiwe et al. [2015](#page-12-7), [2016;](#page-12-1) Singh and Goel [2016](#page-12-8); Moto et al. [2018\)](#page-11-17).

Our previous studies have demonstrated that the stem bark extracts of *P. camptopus* has acute anticonvulsant properties against PTZ, strychnine, picrotoxin and thiosemicarbazide-induced seizure in Wistar rats (Fokoua et al. [2021](#page-11-18)). We proposed in this study to investigate the effects of the methanol extract of *Psychotria camptopus* on PTZ-kindling, a type of progressive epileptogenic process that closely replicate the human intractable temporal lobe epilepsy, in rats.

Materials and methods

Drug and chemicals

Sodium Hydrogen Phosphate (Cat# BDH9298-500G), Sodium Carbonate (Cat# BDH92284-500G), Formaldehyde (Cat# 10790–708), Potassium Carbonate (Cat# BDH9256- 500G) and Sodium Chloride (Cat# BDH9286-500G) were obtained from BDH Poole (England). Trichloroacetic acid (Cat# T6399-100G), Thiobarbituric acid (Cat# T5500- 100G), 5,5'-Dithiobis-2-nitrobenzoic acid (Cat# D8130- 10G), Sulfanilamide (Cat# S9251-100G), N-(1-Naphthyl) ethylenediamine dihydrochloride (Cat# 222488-25G), Sodium Nitrite (Cat# 237213-5G) and Pentylene tetrazole (Cat# P6500-100G) were purchased from Sigma Aldrich, St Louis, USA.

Plant extraction

The stem barks used in this study were harvested in Wabane highlands forest, South West region, Cameroon with the aid of an ethnobotanist, Dr. Tacham N.W. (Department of Biological Sciences, University of Bamenda). A voucher of the plant was authenticated at the Yaoundé National Herbarium as previously identifed by Focho et al. ([2009](#page-11-19)) under the identifcation number No: 56353/HNC (Focho et al. [2009\)](#page-11-19). The methanol extract was obtained as previously described (Fokoua et al. [2021\)](#page-11-18).

Experimental animals

Male Wistar rats weighting between 140 and 160 g (8–10 weeks old) were used in the study. They were obtained from the Central Animal House, College of Medicine, University of Ibadan, Nigeria and housed for two weeks in the Department of Pharmacology and Therapeutics, College of Medicine, University of Ibadan, Nigeria, prior to the beginning of the study. They were housed in $80 \times 50 \times 50$ cm plastic cages embedded with sawdust. The animals were maintained under natural dark/ light cycle and room temperature, with free access to water and standard rodent pellet (Oladekude Food Ltd., Nigeria) except during the treatment and video-recording periods. The animals were handled in accordance with the Guide for the Care and Use of Laboratory Animal published by the United States National Institutes of Health (NIH publication No: 85–23, revised in 1996) (Clark et al. [1997\)](#page-11-20). In addition, the experimental protocols were approved by the Animal Care, Use and Research Ethics Committee of the University of Ibadan (UI-ACUREC/19/00054). All the experiments were conducted between 7:00 and 11:30 AM every day to avoid bias from the circadian rhythm.

Study design

Fifty-six male Wistar rats were randomly divided into six groups. The naive group $(n=8)$ received vehicle (DMSO) 2%, 10 mL/kg). The control group $(n=10)$ received vehicle (DMSO 2%, 10 mL/kg/day) and intraperitoneal injection of a sub-convulsive dose of PTZ (37.5 mg/kg) on every alternate day till the end of the kindling process (43 days). The diazepam group $(n = 8)$ and the MEPC groups $(n = 10$ each) were treated orally everyday with Diazepam 3 mg/kg and *P. camptopus* methanol extract at the doses of 40, 80 or 120 mg/kg respectively. The sample size was determined based on the statistics conveniences, the study design and the ethical guidelines (UI-ACUREC/19/00054), with the prediction that the kindling induction could lead to animal death. The doses used in the present work are from our previous studies (Fokoua et al. [2021\)](#page-11-18). They were selected based on a screening and following the instructions given by the traditional healer.

On PTZ injection days, diazepam and MEPC were administered 1 h prior to PTZ. After the injection of PTZ, rats were individually placed in plexiglas boxes $(50 \times 40 \times 25$ cm) and video-monitored during 20 min for seizure behaviours.

Kindling procedure

The standard PTZ kindling protocol previously described by Davoudi et al (Davoudi et al. [2013\)](#page-11-21) was used in this study. Briefy, PTZ-induced kindling model of seizure was induced in adult male Wistar rats by intraperitoneal injection of PTZ (37.5 mg/kg) on alternate days for 43 days. The behaviours of the animals were video monitored for a period of 20 min after each PTZ injection. Seizure behaviour scoring was operated using a revised Racine's (0–6) scale. The seizure stages were scored and classifed as follows: no response (0); ear and facial twitching with sudden arrest behaviour (1); hyperactivity, rearing with convulsive waves through the body (2); rearing and myoclonic jerks (3); clonic-tonic convulsions, turn over into side position (4); generalized clonictonic seizures, loss of postural control and wild jumping (5); generalized tonic-clonic seizure followed by death (6) (Corda et al. [1991\)](#page-11-22). Rats were considered fully kindled after at least three consecutive manifestations of stage 4 and/or 5 seizures following injection of PTZ. Animals that did not show any sign of seizure activity within the period (20 min) of observation, received the highest latency corresponding to the duration of the observation.

Preparation for brain tissues for biochemical and histomorphological studies

After the behavioural testing on day 43, 5 animals per group were sacrifced by cervical dislocation and their brains were removed, rinsed and transferred into ice cold sodium phosphate buffer (PBS; 0.1 M , $pH = 7.4$). The hippocampus and the prefrontal cortex were dissected out and homogenized in PBS. After centrifugation (10,000 rpm, 4 °C, 10 min) (Olugbemide et al. [2021](#page-11-23)), the supernatant was collected into eppendorf tubes and stored at -20 °C for analysis of oxidative stress biomarkers (MDA, GSH, SOD, CAT and nitrite). The remaining animals $(n=3 \text{ or } 2)$ were trans-cardiacly perfused with normal saline followed by 10% neutral bufer formalin (NBF) and the brains were dissected out and stored in 10% NBF fxative for histological analysis.

Evaluation of the anti‑oxidant efects of *P. camptopus* **methanol extract in PTZ kindled rats**

The concentrations of reduced GSH were determined in the supernatants of hippocampus and prefrontal cortex tissues as described by Jollow et al (Jollow et al. [1974](#page-11-24)). Glutathione levels were extrapolated from a standard curve of glutathione $(0-200 \mu M)$ and expressed as μ mol/mg protein.

Catalase activity in the samples was estimated by a colorimetric assay based on the stable yellow complex formed with ammonium molybdate and hydrogen peroxide (Góth [1991\)](#page-11-25). Catalase activity in the samples was expressed U/ mg protein.

The malondialdehyde (MDA) level used as a marker of lipid peroxidation, was measured in the hippocampus and prefrontal cortex homogenates using the thiobarbituric reacting substances (TBARs) assay as previously described (Tseuguem et al. [2019\)](#page-12-9). The MDA contents were determined using the molar extinction coefficient 1.56×10^5 M/cm and expressed as nmol/mg protein.

The brain level of SOD activity was estimated by a modifed method previously described (Tseuguem et al. [2019](#page-12-9)) based on the superoxide dependent adrenaline autooxidation inhibition in a basic medium. The SOD activity expressed as Unit/mg protein, representing the amount of SOD necessary to cause 50% inhibition of the oxidation of adrenaline.

The concentrations of nitrite in the hippocampus and prefrontal cortex samples were used as a mean to evaluate the production of nitric oxide in the brain using the Greiss reagent colorimetric assay protocol (Green et al. [2000](#page-11-26)). The amount of nitrite in the homogenates were estimated from sodium nitrite standard curve and expressed as μmol/mg protein.

Histomorphological studies

The neutral buffer formalin fixed brains were processed and paraffin embedded. The blocks were then sagittaly sectioned into 4 μm thick, mounted on gelatine-coated charged slide and dried for 2 h at 60 °C in an oven. After dewaxing and rehydration, sections were stained with Haematoxylin & Eosin (H&E) or Cresyl violet stains, rehydrated and fnally cover-slipped and observed at 100 and 400X magnifcation using a Leica DM500 microscope equipped with a Leica ICC50 E camera for image acquisition. For each rat, we performed 3 diferent microphotographies per region of interest in each brain hemisphere. That means for each animal per region of interest, we had $3 \times 2 = 6$ different values. An average was made per region of interest and per animal and thereafter, the mean of the group was determined and plotted as bar graph. The Nissl staining sections were used for neuronal cell counting, using the automated cell counting plugin of the FIJI ImageJ software. Neuronal damages and alterations were examined on the H&E sections. The histomorphological alterations in the prefrontal cortex, CA1 and CA3 hippocampus regions were identifed by observing and quantifying one or more of the following damages: the cytoplasmic vacuolation and cytoplasmic eosinophilia, nuclear chromatin clumping and fragmentation, condensed cytoplasm and fragmentation of the neuronal cells (Singh et al. [2018](#page-12-2)).

Statistical analysis

The normality test was performed with the QQ plot for data with two variables. For data with one variable, normality was tested using Shapiro-Wilk and Kolmogorov-Smirnov tests (Results are reported in the supplementary fle). Parametric data are expressed as means \pm standard deviation (SD) and analyzed using one-way analysis of variance (ANOVA) followed by the Dunnett's multiple comparisons test. Non-parametric data are reported as median and interquartile range and analyzed using Dunn's test. All data analysis were performed with Graph Pad Prism Software, version 8.4. Values were considered statistically significant at $p \le 0.05$.

Results

P. camptopus **methanol extract reduces seizure scores in PTZ kindled rats**

This study was carried out to evaluate the antiepileptogenic efect of the plant extract. The seizure stages or scores were diferent in all the groups during the PTZ-kindling process as shown in Fig. [1.](#page-4-0) Repeated injections of PTZ (37.5 mg/kg, *i.p*) on alternate days for 43 days produced a gradual increase in seizure scores, cumulative seizure episodes in rats (Fig. [1a-b\)](#page-4-0) and up to 30% animal death (Supplementary Table S1). However, the administration of MEPC at all the tested doses and diazepam significantly [two way ANOVA main effects time (F $(11, 432) = 55.36, p < 0.0001$, treatment (F $(5, 432) = 308.3$, *p*<0.0001) and interaction (F (55, 432)=3.132, *p*<0.0001)] reduced the seizure scores in PTZ-kindled rats, with MEPC at 120 mg/kg being the most active (Fig. [1a\)](#page-4-0). Figure [1b](#page-4-0) represents the area under the curve plotted from the curve of the seizure scores and it clearly shows the dose-dependent anti-kindling efect of MEPC. Diazepam also signifcantly [Dunnett's multiple comparisons test (F $(5, 36) = 46.66$] ($p < 0.0001$) reduced the seizure severity and prevented kindling as evidenced by the reduced area under the curve.

P. camptopus **methanol extract increases the latency to convulsions and reduces the number of rats with PTZ‑kindling**

To determine the effect of repeated administration MEPC on the disease severity and the global response to treatment, we evaluated the latency to convulsion and the percentage of animal fully kindled. As shown in Fig. [2A,](#page-4-1) repeated intraperitoneal administration of PTZ (37.5 mg/kg) induced myoclonic jerks and tonic-clonic convulsions respectively at the stage 3 and 4 of the PTZkindling in rats. Daily pre-treatment of the rats with the plant extract or diazepam signifcantly [two way ANOVA

Fig. 1 Efects of *Psychotria camptopus* methanol extract on seizure stage during the PTZ-kindling in Wistar rats. (**a**) Seizure scores evolution during kindling. (**b**) Area under the curve of the seizure score graph, expressing the cumulative seizure scores for each group. Each point in panel A represents the mean \pm SD of 7 animals/

group. $*_{p}$ < 0.05, $*_{p}$ < 0.01 and $*_{p}$ < 0.001 relative to PTZ control (DMSO 2%). Panel (**a**) was analysed with two-way ANOVA followed by Dunnett's multiple comparisons test while panel (**b**) was analysed using one-way ANOVA followed by Dunnett's multiple comparisons test

Fig. 2 Efects of *Psychotria camptopus* extract on stage 3 or 4 latency (**a**) and the percentage of fully kindled rats (**b**) during the PTZ-kindling process. Each point or bar represents the mean \pm SD of 7 animals/group. $* p < 0.05$, $* p < 0.01$ and $* * p < 0.001$ relative to PTZ

control (DMSO2%). Panel A was analysed with two-way ANOVA followed by Dunnett's multiple comparisons test while panel B was analysed using Chi square test

main efects time (F (5, 216)= 26.78, *p* < 0.0001), treatment (F (5, 216) = 33.08, *p* < 0.0001) and interaction (F $(25, 216) = 1.760$, $p = 0.0175$) delayed the latencies to myoclonic jerks and tonic-clonic convulsions when compared with the PTZ control group (Fig. [2A\)](#page-4-1). With the time point analysis, only MEPC (120 mg/kg) and diazepam could signifcantly [Dunnett's multiple comaprisons test] $(p=0.0036)$ reduce the pathology occurrence.

As presented in Fig. [2B](#page-4-1), 85.7% of rats treated with intraperitoneal injection of PTZ developed kindling behaviour. However, the plant extract and diazepam significantly $[[chi]^2]$ (df, $N = 30$) = 1953] ($p < 0.001$) reduced the percentage of PTZ- kindled rats.

P. camptopus **methanol extract reduces MDA and nitrite contents in the brains of PTZ‑kindled rats**

The extension of the oxidative/nitrosative stress was evaluated by assessing the MDA and nitrite contents in the regions of interest of the brain. PTZ-kindling rats had increased **Fig. 3** Efects of the methanol extract of *P. camptopus* on MDA (**a-b**) and nitrite (**c-d**) levels in the prefrontal cortex (PFC) and hippocampus of PTZ kindled rats. Each bar represents the mean \pm SD of 5 animals/ group. **p*<0.05, ***p*<0.01 and ****p*<0.001 relative to PTZ control. Data were analysed using one-way ANOVA followed by Dunnett's multiple comparisons test

MDA and nitrite contents in the PFC and the hippocampus when compared with vehicle (Fig. [3\)](#page-5-0). Repeated oral administration of rats with MEPC signifcantly reduced the MDA (Fig. [3A-B](#page-5-0)) level in both pre-frontal cortex and hippocampus [Dunnett's multiple comparisons test] (*p=* 0.001). The nitrite content (Fig. [3C-D](#page-5-0)) was also significantly [Dunnett's multiple comparisons test $(p=0.0044)$ reduced by the MEPC treatment. Diazepam was only able to signifcantly [Dunnett's multiple comparisons test] $(p=0.0156)$ reduce the MDA content in the pre-frontal cortex (Fig. [3A](#page-5-0)).

P. camptopus **methanol extract boosts antioxidant profles in the brains of PTZ‑kindled rats**

To examine the effect of MEPC on the endogenous antioxidant system, we assessed the catalase and superoxide dismutase activities as well as the GSH content. As presented in Figs. [4,](#page-6-0) repeated injection of sub-convulsive dose of PTZ signifcantly decreased the antioxidant enzymes (CAT and SOD) and GSH contents in the PFC and the hippocampus [Dunnett's multiple comparisons test] (p from 0.0444 to $\langle 0.001 \rangle$ when compared with the naive control. Administration of the extract (120 mg/ kg) signifcantly augmented CAT activity in the hippocampus [Dunnett's multiple comparisons test] $(p=0.0039)$ but not in the PFC (Fig. [4A-B](#page-6-0)). MEPC did not exert any signifcant efect on the SOD activity (Fig. [4C-D](#page-6-0)) and GSH levels (Fig. [4E-F\)](#page-6-0) in the PFC and hippocampus of rats when compared with PTZ kindling group.

P. camptopus **exhibits neuroprotective efect against PTZ‑kindling in rats**

The H&E and Nissl staining were used to evaluate the neuroprotective efects of MEPC on brain cells and neurons, respectively. H&E staining of the PFC and the hippocampus (CA3) regions showed normal morphological features of the PFC and hippocampus in naive rats. In contrast, PTZ-kindling (control) resulted in marked neuropathological alterations in the PFC with increased number of nuclear pyknosis and cytoplasmic vacuolation. There were also distortions of the hippocampus CA3 and CA1 cells in PTZ-kindled rats, with cellular disorganization (Fig. [5A](#page-8-0)). These damages were reduced by the treatment with the plant extract at the dose of 120 mg/kg. Diazepam (3 mg/kg) failed to restore the cellular architecture of the hippocampal neurons both in the CA1 and CA3 (Fig. [5](#page-8-0)). Quantitative and qualitative analysis of the H&E sections revealed that PTZ kindling reduced cell density in the PFC and hippocampus CA3 of rats as compare to naive group. Also, this cell lost was correlated to an increase of cell damages as the percentage of altered **Fig. 4** Efects of the methanol extracts of *P. camptopus* on catalase activity (**a-b**), super oxide dismutase activity (**c-d**) and reduced-Glutathione (**e-f**) in the prefrontal cortex (PFC) and hippocampus of PTZkindled rats. Each bar represents the mean \pm SD of 5 animals/ group. **p*<0.05, ***p*<0.01 and ****p*<0.001 relative to PTZ control. Data were analysed using one-way ANOVA followed by Dunnett's multiple comparisons test

cells increased in the PTZ-control group as compare to the naive animals. As observed on Fig. [5B](#page-8-0), MEPC and DZP pretreatments prevented cell lost in the PFC and Hippocampus hCA1 and hCA3 regions as compare to the control group.

The Nissl staining was performed to focus on neuronal cells. This staining revealed normal cell morphology and distribution in the prefrontal cortex and the hippocampus from naive rats. Altered neuronal cells morphology and organization was observed in PTZ-kindling rat and these changes were attenuated by the methanol extract of *P. Camptopus* (120 mg/kg)*.* Quantitative analysis of these brain sections showed that PTZ kindling signifcantly reduced the number of viable neuronal cells in the PFC and hippocampus

CA3 when compared with Naive (Fig. [6A and B](#page-9-0)). However, treatment with the extract of *P. Camptopus* (120 mg/kg) signifcantly attenuated neuronal cell death in these brain regions. Diazepam signifcantly prevented cell death in the PFC and the hCA1. This substance instead tends to worsen the condition in the CA3 region (Fig. [6A](#page-9-0) and [B\)](#page-9-0).

Discussion

The present study was undertaken to evaluate the antiepileptogenic and neuroprotective efects of the methanol extract from the stem bark of *P. camptopus* (MEPC) in PTZ-induced

 (b)

Fig. 5 Efect of extract of *P. camptopus* on histomorphological ◂ changes of H&E stained sections of the prefrontal cortex and hippocampus of PTZ-kindled rats**. (a)-**Slices from naive group revealed normal histological open chromatin pattern and cytoarchitecture (white arrows) of viable neurons in all the brain regions observed. Control slices showed abnormal histomorphological features, with some pyknotic cells (black arrows), increased cytoplasmic vacuolations (arrow heads) and cell disorganization (red arrow) in the hCA3. Slices from DZP 3 mg/kg and MEPC 120 mg/kg treated rats presented ameliorated histomorphological distortions with reduced cellular damages. Original magnifcation X400, Calibration bar=50 μm for all microphotographs. **(b)-** Number of cells and damaged cells in the PFC and hippocampus CA1 and CA3 regions. Each bar represents the mean of microphotographs per group $(n=3 \text{ or } 2)$, for each animal 3 different microphotographs per region of interest \times 2 hemispheres)

kindled rats. Chronic administration of pentylenetetrazole time-dependently decreased the latency to myoclonic and generalized seizures. An increase in seizure scores and kindled rats number was also observed. MEPC and diazepam signifcantly increased the latencies to myoclonic jerks and generalized tonic-clonic seizures. These substances also reduced seizure score and the number of rats with PTZkindling. MEPC improved glutathione status and decreased lipid peroxidation in the brains of kindled rats. MEPC also exhibited neuroprotective activity against pentylenetetrazole-induced hippocampal and PFC neuronal damages.

Pentylenetetrazole-induced kindling is a type of progressive epileptogenic process and a well-established model in rodents that closely replicates the pathogenesis of refractory epilepsy in humans. It is also used to elucidate neurological complications associated with epilepsy, including oxidative stress (Taiwe et al. [2016](#page-12-1)) and neurodegeneration (Erkec [2015](#page-11-10); Singh et al. [2018\)](#page-12-2). These characteristics justify the use of pentylenetetrazole-induced kindling model in the present study. The inability of most antiepileptic drugs to modify the changes in oxidative stress in epileptic brain may perhaps contribute to their inefectiveness in certain patients, who still experienced seizures despite treatments (Kubova [2016b;](#page-11-27) Tang et al. [2017](#page-12-6)). Thus, phytochemicals with antioxidant and neuroprotective properties are increasingly being proposed as promising strategy for treatment of epilepsy par-ticularly the intractable forms (Ashrafi et al. [2007](#page-11-28); Dariani et al. [2013;](#page-11-29) Annamaria Vezzani [2014](#page-12-10); Tang et al. [2017](#page-12-6)). The results of this study showed that the methanol extract of *P. camptopus* exhibited antiepileptogenic efect against PTZ-induced kindling as evidenced by increased latency to myoclonic jerks and tonic-clonic convulsions, reduced seizure scores, and number of rats with seizure episodes. The extract also reduced MDA and nitrite levels in the PFC and hippocampus of PTZ-kindled rats. The altered histomorphological features of the PFC and the hippocampus of rats with PTZ-kindling were ameliorated by the plant extract.

The kindling process results in gradual increase in seizure susceptibility and amplifcation of seizure activity that often culminate in generalized tonic-clonic seizures (Davoudi et al. [2013;](#page-11-21) Shimada and Yamagata [2018\)](#page-11-30). In this model, antiepileptogenic activity is proven based on the ability of the test compound to prolong the latency to myoclonic and generalized seizures, to reduce seizure scores and the number of kindled rats. Our data showed that the extract of *P. camptopus* increased the latencies to myoclonic jerks and generalized tonic-clonic seizures, decreased seizure scores and reduced the number of rats with PTZ-kindling. Thus, suggesting that the extract of *P*. *camptopus* possesses antiepileptogenic activity and further support its use as a remedy for generalized tonic-clonic seizures in ethnomedicine. These fndings are in accordance with our previous study that demonstrated the anticonvulsant efects of the same extract on acute models of convulsion (Fokoua et al. [2021\)](#page-11-18).

It has been reported that PTZ-kindling results in an increase in brain levels of glutamate, increased NO-mediated activation of NMDA receptors and decreased brain GABA levels (De Luca et al. [2005](#page-11-31); Abdel-Zaher et al. [2017\)](#page-10-0) that play a vital role in epileptogenesis and seizure activity (Kumar and Kumar [2017\)](#page-11-11). The increase in the excitability of neurons due to antagonism of GABA and activation of NMDA receptors results in copious production of free radicals and oxidative stress-mediated neuronal cell loss (Roganovic et al. [2019\)](#page-11-8). Indeed, there are increasing evidences implicating oxidative stress in the development and progression of epilepsy (Waldbaum and Patel [2010](#page-12-11); Geronzi et al. [2018](#page-11-32)). In fact, the hyper-excitability of neurons in the process or during persistent seizure distorts the antioxidant-oxidant equilibrium. This imbalance causes neuronal cells injuries through membrane peroxidative activity of ROS and RNS (Ilhan et al. [2005](#page-11-33); Roganovic et al. [2019\)](#page-11-8). Brain tissues are extremely susceptible to oxidative stress due to high metabolism, increased blood perfusion, enriched membrane lipids and reduced antioxidant defense mechanisms (Ilhan et al. [2005](#page-11-33); Nigar et al. [2016](#page-11-6); Pearson-Smith and Patel [2017](#page-11-7)). The oxidative stress status will lead to membrane lipid peroxidation with increased MDA content, biological molecule oxidation, DNA alteration and other cell damages, which ultimately culminate into neuronal cell death (Waldbaum and Patel [2010;](#page-12-11) Snehunsu et al. [2015;](#page-12-12) Pearson-Smith and Patel [2017](#page-11-7)). The fndings that increased free radicals activity and lowered endogenous antioxidant molecules correlated with decreased seizure threshold and neuronal cell loss further support the role of oxidative stress in the pathophysiology of epilepsy (Nigar et al. [2016;](#page-11-6) Abdel-Zaher et al. [2017](#page-10-0); Pearson-Smith and Patel [2017;](#page-11-7) Roganovic et al. [2019](#page-11-8)). The cortex and limbic structures including the prefrontal cortex, amygdala and the hippocampus have been reported to be the most vulnerable brain regions during epileptogenesis (Roganovic et al. [2019\)](#page-11-8). These prompted us to evaluate

 (a)

Fig. 6 Efect of *P. camptopus* methanol extract on histomorphological changes of Nissl stained section of the prefrontal cortex and the hippocampus of PTZ-kindled rats. (**a)-**Microphotograhs of PFC and hippocampus sections. Slices from naive rats revealed normal histological open chromatin pattern and cytoarchitecture of viable neurons (white arrow) and a normal cellular organization of the hCA3. Control slices showed abnormal histomorphological features, with some pyknotic cells (black arrows) and loss of cellular organization

in the hCA3 (red arrow). Diazepam (3 mg/kg) and MEPC 120 mg/kg revealed ameliorative features of the alterations induced by kindling. Original magnifcation X40, Calibration bar=50 μm for all fgures**. (b)-** Number of Nissl stained neurons in the PFC and hippocampus CA1 and CA3 regions. Each bar represents the mean of microphotographs per group $(n=2 \text{ or } 3)$, for each animal 3 different microphotographs per region of interest \times 2 hemispheres

whether the antiepileptogenic efects of MEPC could be related to a probable antioxidant efect in target structures, namely the prefrontal cortex and the hippocampus.

In agreement with the literature, repeated administration of sub-convulsive dose of PTZ resulted in oxidative stress, as depicted by elevated MDA and nitrite levels accompanied by decreased antioxidant (GSH, CAT and SOD) contents in the PFC and hippocampus of PTZkindled rats. MEPC signifcantly reduced MDA in these brain regions. Besides, MEPC signifcantly increased the catalase activity in the hippocampus as compared to the PTZ-kindling rats. These results suggest in vivo antioxidant property of MEPC, which might contribute to its antiepileptogenic efect.

It is worth noting that MEPC did not have any signifcant efect on the GSH content, neither on the SOD activity, suggesting that the plant extract might be unable to boost these endogenous antioxidant parameters. The potentiation of the catalase activity alone might be insufficient to justify the potent efect of the plant extract against lipid peroxidation. It could be hypothesized that MEPC possess intrinsic antioxidant activity, serving as primary antioxidant substance.

Studies have revealed increased cell damages in the frontal cortex and hippocampus of animals after PTZ kindling (Samokhina and Samokhin [2018\)](#page-11-14). Specifcally, PTZ kindling has been shown to cause cell damages and reduced neuronal cell density in PFC, CA1 and CA3 brain regions (Snehunsu et al. [2015](#page-12-12); Vasil'ev et al. [2015;](#page-12-13) Aldawsari et al. [2017](#page-10-1); Erkec et al. [2018;](#page-11-34) Muke et al. [2018;](#page-11-35) Samokhina and Samokhin [2018;](#page-11-14) Singh et al. [2018,](#page-12-2) [2019\)](#page-12-3). Concordantly, results from the present study showed a close correlation between seizure severity, oxidative stress and neuronal cell damages in the PFC, CA1 and CA3 of rats with PTZ kindling. The fact that MEPC ameliorates the histomorphological distortions and reduced the loss of PFC and hippocampal neuronal cells suggest it has neuroprotective activity. Pentylenetetrazol initially blocks $GABA_A$ receptors and increases glutamate production by reducing the inhibitory GABAergic infuxes (Lin et al. [2019\)](#page-11-36). This results in excitotoxicityinduced cell damages. It is reported that establishment of functional GABAergic system and restoration of the inhibitory/excitatory balance attenuates seizure sensitivity and prevents neurons damages during epileptogenesis (Moto et al. [2018;](#page-11-17) Lin et al. [2019;](#page-11-36) Righes Marafga et al. [2020](#page-11-37); Sünnetçi et al. [2021\)](#page-12-14). It is worthy to note that previous studies have also established the presence in MEPC of bioactive compounds with neuroprotective and antioxidant properties. Furthermore, MEPC was able to inhibit thiosemicarbazideand picrotoxin-induced seizures in wistar rats (Fokoua et al. [2021](#page-11-18)). In this study, we found that MEPC protects nervous cells from PTZ-induced cell damages, suggesting a neuroprotective efect that may be mediated trough GABAergic modulation and antioxidant effects. However, the relevance of these bioactive compounds in the antiepileptogenic and neuroprotective activities of *P. camptopus* against PTZinduced kindling requires further investigations.

Strengths of the present study are the clear demonstration of the anti-epileptogenic and neuroprotective efects of the methanol extract of *P. camptopus* at the doses that are

in the pharmacological range. The study also demonstrated the antioxidant efect of the plant extract and linked it to the anti-epileptogenic and neuroprotective activities observed.

Its limitations are the lack of precise mechanism underlying the antiepileptogenic and neuroprotective efects of the plant extract. In addition, the efects of the plant extract on neuropsychiatric and cognitive alterations were not assessed. However, these limitations are currently under investigation in our laboratory.

Conclusion

The results of this study showed that the methanol extract of *Psychotria camptopus* stem bark exhibited antiepileptogenic and neuroprotective activities against PTZ-induced kindling in rats. The antioxidant efects of this plant extract may contribute to its antiepileptogenic and neuroprotective activities. These fndings support the potential of *Psychotria camptopus* in the management of epilepsy and especially the intractable forms.

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Data availability All the data and material from this study are available on demand.

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Declarations

Ethical approval The experimental protocols were approved by the Animal Care, Use and Research Ethics Committee of the University of Ibadan (UI-ACUREC/19/00054).

Competing interests The authors declare no competing interest.

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