

Long non‑coding RNAs in nasopharyngeal carcinoma: biological functions and clinical applications

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Abstract

Nasopharyngeal carcinoma (NPC) is one of the most common head and neck malignancies. It has obvious ethnic and regional specificity. Long non-coding RNAs (LncRNAs) are a class of non-protein coding RNA molecules. Emerging research shows that lncRNAs play a key role in tumor development, prognosis, and treatment. With the deepening of sequence analysis, a large number of functional LncRNAs have been found in NPC, which interact with coding genes, miRNAs, and proteins to form a complex regulatory network. However, the specifc role and mechanism of abnormally expressed lncRNAs in the pathogenesis of NPC is not fully understood. This article briefy introduced the concept, classifcation, and functional mechanism of lncRNAs and reviewed their biological functions and their clinical applications in NPC. Specifcally, we described lncRNAs related to the occurrence, growth, invasion, metastasis, angiogenesis, and cancer stem cells of NPC; discussed lncRNAs related to Epstein-Barr virus infection; and summarized the role of lncRNAs in NPC treatment resistance. We have also sorted out lncRNAs related to Chinese medicine treatment. We believe that with the deepening of lncRNAs research, tumor-specifc lncRNAs may become a new target for the treatment and a biomarker for predicting prognosis.

Keywords Long non-coding RNAs (lncRNAs) · Nasopharyngeal carcinoma (NPC) · Biological functions · Clinical applications

Introduction

Nasopharyngeal carcinoma (NPC) is a malignant tumor of the head and neck, which originates from the epithelium of nasopharyngeal mucosa, and its distribution is obviously regional and ethnic [\[1](#page-8-0)]. NPC is mainly concentrated in Southeast Asia and southern China. The pathogenesis of NPC is closely related to environmental factors, EBV infection, genetic susceptibility, and other factors. The latest data show that there are 129,079 new cases of NPC and 72,987 deaths worldwide. The incidence rate of males is much higher than that of females [[2\]](#page-8-1). At present, the morbidity and mortality of nasopharyngeal carcinoma in my country is relatively high, and it is highly invasive, prone to early metastasis, and has a poor prognosis. The population

is mainly middle-aged and elderly. As the population ages, the burden of nasopharyngeal cancer may further increase.

LncRNA (Long non-coding RNA) is a non-coding RNA with a length of more than 200 bp. LncRNA was initially regarded as the "noise" of genomic transcription, which is a by-product of RNA polymerase II transcription and has no biological function. However, recent studies have shown that lncRNA can regulate gene expression at many levels, including epigenetic regulation, transcriptional regulation, posttranscriptional regulation, and regulation of MiRNA [\[3\]](#page-8-2). The specifc mechanism is very complex and remains to be clarifed. According to the position of relative protein-coding genes in the genome, lncRNA can be divided into five categories, named Antisense lncRNA, Sense lncRNA, Intronic transcript, Long intergenic non-coding RNA (lincRNA), and Bidirectional lncRNA [[4\]](#page-8-3) (Fig. [1a](#page-1-0)). The cellular localization of diferent lncRNA is diferent. Nuclear lncRNA accounts for the majority of lncRNA, which plays a regulatory role by recruiting chromatin regulatory factors to DNA, or as scaffolding for some ribonucleoprotein. Cytoplasmic lncRNA acts on the level of post-transcriptional regulation of genes, such as MiRNA sponge, which inhibits the effect of miRNA

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on mRNA [[5](#page-8-4)]. In a classic review, Kevin C. Wang et al. distill the myriad functions of lncRNAs into four archetypes of molecular mechanisms, as signals, decoys, guides, and scaffolds [\[3,](#page-8-2) [5](#page-8-4)] (Fig. [1b](#page-1-0)). LncRNA is widely involved in a variety of biological processes, and the abnormal expression of LncRNA is closely related to a variety of diseases, including cancer.

At present, a large number of studies have confrmed that lncRNA is widely involved in the carcinogenesis, migration and invasion, epithelial-mesenchymal transformation, angiogenesis, and other malignant phenotypes of NPC and mediates radiotherapy resistance and chemotherapy resistance through a variety of pathways, which is an important biomarker for the diagnosis and prognosis of NPC.

Fig. 1 a According to the position of relative protein-coding genes in the genome, lncRNA can be divided into fve categories. ①. Intronic lncRNAs: transcribed from introns of protein-coding genes; ②. Intergenic region: transcribed from the intergenic region within two genes; ③. Bidirectional lncRNA: transcribed from diferent directions

and initiated by adjacent coding transcripts on the opposite strand; ④and⑤. Sense or antisense lncRNA: transcribed from the sense or antisense strand of the protein-coding gene and the overlapping exons of the protein-coding gene; **b** The lncRNA mechanisms of action can be divided into four categories: signal, decoy, guide, and scafold

Biological function of lncRNA in nasopharyngeal carcinoma

Oncogenic lncRNAs

A large number of lncRNA have been reported to be upregulated in NPC, which are likely to be involved in carcinogenesis, development, proliferation and migration, and other malignant biological phenotypes through diferent mechanisms as oncogenes. For example, the lncRNA differentiation antagonizing non-protein coding RNA (DANCR) has been reported to play a carcinogenic role in a variety of malignant tumors, including breast cancer [[6\]](#page-8-5), hepatocellular carcinoma [[7](#page-9-0)], and osteosarcoma [[8,](#page-9-1) [9](#page-9-2)]. With more and more in-depth research, the role of DANCR as a metastasis-specifc oncogenic lncRNA in NPC has gradually been revealed. Here, we try to briefy summarize the biological functions and molecular mechanisms of carcinogenic lncRNA in NPC (Table [1\)](#page-3-0).

Tumor suppressor lncRNAs

With the further study of lncRNA in tumor, some "beneficial" lncRNA have been widely reported. As tumor suppressors, they can inhibit tumor occurrence, development, migration, and invasion in vivo or in vitro. Maternal expression gene 3 (MEG3) is the frst lncRNA [[10](#page-9-3)], with tumor suppressor function [[11\]](#page-9-4). Its low expression can cause head and neck squamous cell carcinoma, breast cancer, and pancreatic ductal adenocarcinoma [\[12–](#page-9-5)[14](#page-9-6)]. MEG3 was also identifed as a low-expression tumor suppressor lncRNA in nasopharyngeal carcinoma (NPC) [\[15\]](#page-9-7). However, the understanding of tumor suppressor lncRNA still has a long way to go. The biological functions and mechanisms of known tumor suppressor lncRNA are summarized below (Tabl[e2](#page-4-0)).

Clinical application of lncRNA in NPC

Through the study of nasopharyngeal carcinoma-related lncRNA, it is not difficult to find that lncRNAs, as an oncogene or suppressor, play an important role in regulating the progression, metastasis, and invasion of NPC. The abnormal expression of LncRNA is closely related to the occurrence, development, and prognosis of NPC. Some of them can be used as efective diagnostic and prognostic markers of human tumors, and some lncRNA are involved in the treatment and resistance of NPC. The study of lncRNA provides a new idea for studying the pathogenesis of tumor and exploring new methods of diagnosis and treatment.

Long non‑coding RNAs and Epstein‑Barr virus infection in NPC

Epstein-Barr virus infection is a recognized risk factor for NPC. The establishment of EBV latent infection in the epithelium of nasopharyngeal precancerous lesions is considered to be an early and necessary step in the pathogenesis of NPC [[16\]](#page-9-8). The EB virus has three latent infection types, and the NPC cells are expressed as the II latency, and the viral protein associated with NPC is transcribed by viral RNA to the right transfer (BARTS) encoding. This transcription also encodes viruses microRNA and lncRNA [[17\]](#page-9-9). These viral products (EBV-related RNA EBER1 and EBER2) can change the host cell signal, establish a tumor microenvironment and promote the occurrence and development of tumors. BART RNA is usually highly expressed in EBV infection epithelial tumors. The products of these BART RNAs may be miRNAs or lncRNAs [[18\]](#page-9-10). EBV is also the frst virus to be discovered that encodes microRNAs [\[19](#page-9-11)]. Previous studies have also focused on the regulation of MiRNA's involvement in the occurrence and development of NPC [[20](#page-9-12)]. At present, there is evidence that lncRNA plays an important role in the occurrence, development, and progression of epithelial tumors associated with EBV infection [[21–](#page-9-13)[23](#page-9-14)]. Based on whole-genome RNA sequencing, some researchers have tested the lncRNA expression profles in four EBV genome-infected 293 cell lines and EBV-negative 293 cells. In the comparative analysis, a series of lncR-NAs expression disorders were found [[24](#page-9-15)]. The interaction between EBV and lncRNA has two main aspects: on the one hand, EBV regulates the expression of host lncRNAs, on the other hand, EBV itself encodes lncRNAs that have an impact on host cell-related proteins and signal pathways [[25\]](#page-9-16).

EBV‑encoded lncRNA

A 2015 study showed that a subset of the changes in gene expression induced by latent EBV infection in AGS cells is due to the expression of the BART nuclear RNAs and suggests that the spliced BART transcripts function as lncRNAs and from this point onward, they are referred to as the BART lncRNA [[26\]](#page-9-17). In NPC, EBV hardly expresses viral proteins, but Bam-HI A right transcript (BARTS) RNA levels are elevated, including viral microRNAs and LncRNAs. BART lncRNAs are located in the nucleus of EBV-infected cells [[27](#page-9-18)]. The knockdown of BART lncR-NAs signifcantly afects the expression of genes related to cell adhesion, oxidoreductase activity, infammation, and immunity. Bart lncRNAs also participate in the epigenetic regulation of host gene expression in nasopharyngeal carcinoma by interfering with histone methylation and acetylation processes [[28\]](#page-9-19). A study showed that the expression of BARTs is regulated by NF-κB signaling. EBV LMP1

LncRNA	Expression Targets		Biological functions	Refs
DANCR	Up	$HIF-1\alpha$ mRNA; SOX2 mRNA; PTEN	Promotes NPC cells proliferation and migration	$[90 - 93]$
XIST	Up	miR-491-5p; Notch3; miRNA-148a-3p/ ADAM17	Promotes hypoxia-induced glycolysis, migration and invasion, and tumor growth in vivo	$[94 - 97]$
ZFAS1	Up	miR-135a; miR-892b; PI3K/AKT signaling pathway	Promotes cell proliferation, tumorigenesis, and metastasis	$[98 - 101]$
H ₁₉	Up	EZH2 / miR-630; miR-675-5p/SFN; let-7	Promotes cell invasion, proliferation, and metastasis	$[102 - 104]$
HOTAIR	Up	miR-101/COX-2 axis; VEGFA	Promotes invasion and migration; mediated angiogenesis	$[105 - 107]$
SNHG7	Up	miR-140-5p; miR-514a-5p	Promotes proliferation and migration and inhibits apoptosis	[108, 109]
NEAT1	Up	miR-124/NF-κB pathway; miR-34a-5p/ Wnt/β- catenin signaling	Promotes cell proliferation, migration, invasion, and EMT; inhibits apoptosis	[110, 111]
MALAT1	Up	miR-124/Capn4	Promotes cell proliferation, migration, and inva- sion	$[112]$
SMAD5-AS1	Up	miR-106a-5p; miR-195	Promotes EMT, cell proliferation, migration, and invasion	[113, 114]
SRRM2-AS	Up	MYLK/cGMP-PKG signaling pathway	Mediated angiogenesis	[115]
FOXD3-AS1	Up	miR-185-3p;	Regulates NPC cell stemness invasion, viability, and migration potentials and apoptosis	[116, 117]
AFAP1-AS1	Up	KAT2B/H3K14ac/TIF1a/RBM3/YAP signaling pathway; miR-423-5p	Promotes proliferation and colony formation in vitro and tumorigenicity in vivo	[118, 119]
Gas5	Up	GAS5/ miR-4465/COX2 axis	Regulates proliferation and promotes apoptosis	[120]
RP11-624L4.1	Up	CDK4/6-CyclinD1-Rb-E2F1 Pathway	Induces tumor proliferation and cycle	$[121]$
LOC284454	Up	Rho/Rac signaling pathways	Promotes the migration and invasion capacity	$[122]$
LINC01503	Up	FOSL1	Promotes NPC cells growth, migration, and inva- sion	$[123]$
MSC-AS1	Up	miR-524-5p/NR4A2 axis	Promotes cell invasion and EMT process	[124]
AATBC	Up	miR-1237-3p-PNN-ZEB1 axis	Promotes migration, invasion, and metastasis	[125]
DRAIC	Up	microRNA-122/ SATB1 axis	Promotes migration and invasion	$[126]$
LINC00460	Up	miR-149-5p/ $IL6$ axis miR-30a-3p/RAP1A axis	Promotes migration, EMT, invasion, and metas- tasis	[127, 128]
FAM225A	Up	miR-590-3p/miR-1275 / ITGB3, and activate FAK/PI3K/Akt pathway	Promotes proliferation and metastasis	$\lceil 129 \rceil$
HOXC13-AS	Up	miR-383-3p/ HMGA2 axis	Regulates proliferation, migration, and invasion	$\lceil 130 \rceil$
SOX2-OT	Up	miR-146b-5p/HNRNPA2B1 pathway	Regulates cell proliferation, cell cycle, and metastasis	[131]
SNHG1	Up	miR-145-5p / $NUAK1$ axis	Promotes migration, EMT, and invasion	$\lceil 132 \rceil$
ANRIL	Up	mTOR signal	Induces the stem cell-like cells; promotes prolif- eration, colony formation, and transformation ability and reprograms the glucose metabolism	$\lceil 133 \rceil$
SWSAT1	Up	miR-326/330-5p / cyclin D1 axis	Promotes viability and growth	[134]

Table 1 Summary of tumorigenic lncRNA known to modulate nasopharyngeal carcinoma

is an efective activator of NF-κB signal. LMP1 can upregulate the expression of BARTs through NF-κB signal, but miR-BARTs can also down-regulate the expression of LMP1. The authors speculate that abnormal NF-κB signaling and BART expression form an auto-regulatory loop to maintain the EBV latency in NPC cells [[29](#page-9-20)].

Some lncRNAs encoded by EBV, such as BHLF1 and BHF3, can be used as a functional lncRNA to participate

EBV regulates the expression of host lncRNAs

The long intergenic non-coding RNA LINC00312, also called NAG7, was found that it could not only inhibit proliferation and induce apoptosis in NPC cells but also stimulate

in the regulation of the life cycle of EBV [[30\]](#page-9-21). But their

LncRNA	Expression	Targets	Biological functions	Refs
ZNF667-AS1	Down	miR-1290/ABLIM1 axis	Inhibits the proliferation, migration and invasion abilities, and tumor growth; promotes apoptosis	[135]
NKILA	Down	NF - κ B pathway	Inhibits tumorigenesis and metastasis	[136]
LET	Down	MAPK/ERK signaling pathway	Inhibited invasion and proliferation and induced cell apoptosis;	[137, 138]
LINC0086	Down	$miR-214$	Inhibited proliferation and promoted apoptosis	[139]
MEG3	Down	p53 Pathway	Suppresses cell proliferation and tumorigenicity	$\lceil 15 \rceil$
LOC401317	Up	Unknown	Inhibits cell growth and induces apoptosis	[140]

Table 2 Summary of tumor suppressor lncRNAs known to modulate nasopharyngeal carcinoma

NPC cell invasion [[31,](#page-9-22) [32](#page-9-23)]. The non-coding ribonucleic acid EBER-1 transcribed by the EBV is abundantly expressed in various cells latently infected with EBV and plays an important role in EBV-mediated tumorigenesis [[33\]](#page-9-24). In a study, the authors analyzed the relationship between LINC00312 and EBER-1 abnormal expression and found that LINC00312 and EBER-1 in nasopharyngeal carcinoma were signifcantly negatively correlated, suggesting that EBER-1 may regulate the expression of LINC00312 through certain genes and signal pathways [[34](#page-9-25)]. But its exact mechanism needs further study.

EBV can also exert oncogenic or tumor suppressor properties and participate in cancer progression by targeting host genes or its own viral genes by encoding microRNA. A study found that EBV-encoded microRNA, called EBVmiR-BART6-3p, inhibited the migration and invasion of nasopharyngeal carcinoma cells by reversing the epithelialmesenchymal transition (EMT) process. Through gene chip analysis, the authors found that EBV-miR-BART6-3p downregulated the expression of a new long non-coding RNA (LncRNA) LOC553103. EBV-miR-BART6-3p inhibits the migration and invasion of NPC cells by down-regulating LncRNA LOC553103 [[35\]](#page-9-26). Subsequently, the team further confrmed through proteomics analysis and luciferase report that Stathmin (STMN1) is afected by EBV-miR-BART6-3p and LOC553103, and that LOC553103 directly binds and stabilizes the 3'UTR region of STMN1 mRNA. STMN1 is an important tubulin, which can inhibit the stability of microtubules, thereby promoting tumor cell proliferation, invasion, and metastasis [[36\]](#page-9-27). These results indicate that the EBV-miR-BART6-3p/LOC553103/STMN1 axis regulates the expression of cell cycle-related proteins, thereby inhibiting the proliferation of EBV-related tumor cells [\[37](#page-9-28)].

Long non‑coding RNAs and radio‑resistance in NPC

The pathological classifcation of NPC is mainly poorly diferentiated squamous cell carcinoma, which accounts for more than 80% of all NPC cases [[38\]](#page-9-29). Studies have shown that poorly diferentiated squamous cell carcinoma is extremely sensitive to radiation, and NPC occurs in hidden parts such as the pharyngeal recess and the anterior parietal wall, making it difficult to perform surgery $[39, 40]$ $[39, 40]$ $[39, 40]$ $[39, 40]$ $[39, 40]$. This determines that the current main treatment for NPC is radiation therapy, supplemented by chemotherapy when neces-sary [\[41\]](#page-10-2). Radiotherapy for early NPC is effective, while radiotherapy for middle and late NPC is less efective, and long-term radiotherapy will lead to changes in certain genes and proteomics, resulting in radiotherapy resistance [[42](#page-10-3)]. Radiation resistance is the main obstacle to radiotherapy, which can lead to relapse and poor prognosis. Therefore, understanding the radio-resistance mechanism and developing new radio-sensitization strategies are of great signifcance for the treatment of NPC.

Recent research has shown that abnormal expression of lncRNAs in malignant tumor cells before and after radiotherapy may participate in the progression of cancers and afect the radiation sensitivity of malignant tumor cells mediated by specifc signaling pathways or cell cycle regulation [\[43](#page-10-4)[–45\]](#page-10-5). Similarly, in nasopharyngeal carcinoma, there are many lncRNAs involved in the regulation of radiosensitivity. On the one hand, some lncRNA can induce radiation resistance of nasopharyngeal carcinoma by adsorption with a specific miRNA through sponge. For example, the tumor suppressor FAM133B-2 is highly expressed in radioresistant NPC cells and can target the miR-34a-5p/CDK6 axis to inhibit radiotherapy resistance [[46\]](#page-10-6). There are also many lncRNAs such as ZFAS1 and PVT [\[47–](#page-10-7)[49](#page-10-8)] which can also be used to play anti-radiation through ceRNA networks. On the other hand, lncRNA can also modulate the radiation resistance by directly acting in certain cell's signal pathways or key molecules. For example, in one of the latest studies, researchers screened the lncRNA CASC19 as a candidate radiation resistance marker using RNA-SEQ analysis of NPC cell line CNE2 and its radiation resistance cell line CNE2R. The mechanism study found that CASC19 has an important contribution to the radio-resistance of NPC cells by AMPK / mTOR pathway [[50](#page-10-9)]. Evidence that lncRNA inhibits radiotherapy resistance of NPC by regulating DNA damage has also been found. A very important evidence is that Linc00312 directly binds to DNA-PKcs, inhibits the recruitment of Ku80 by DNA-PKcs, inhibits

the phosphorylation of AKT-DNA-PKcs axis, thus inhibits the perception and transduction of DNA damage signals in NHEJ repair pathway, and ultimately inhibits radiation resistance [[51\]](#page-10-10).

Radiation resistance is the main obstacle to radiotherapy, which can lead to relapse and poor prognosis. Therefore, understanding the radio-resistance mechanism of NPC and developing new radio-sensitization strategies are of great signifcance for the treatment of NPC. In this part, we summarize the published studies on lncRNAs in radiotherapy regarding the biological function and mechanism of NPC (Fig. [2\)](#page-5-0).

Long non‑coding RNAs and chemotherapy resistance in NPC

Chemotherapy is an effective treatment for advanced NPC. In a randomized controlled trial, through long-term

follow-up, it was found that cisplatin/fuorouracil/docetaxel (TPF) induction chemotherapy (IC) combined with concurrent radiotherapy (CCRT) could signifcantly improve the survival rate of patients with locally advanced NPC compared with CCRT alone, and there was no signifcant increase in advanced toxicity [[52](#page-10-11), [53](#page-10-12)]. Recently, the frst phase 3 trial of recurrent or metastatic nasopharyngeal carcinoma (RMNPC) has identifed gemcitabine plus cisplatin (GP) as the standard frst-line treatment [\[54\]](#page-10-13). Although single or combined chemotherapeutic drugs have a certain efect on recurrent or metastatic cancer, long-term use of chemotherapeutic drugs will produce resistance to chemotherapeutic drugs, which greatly limits the efficacy of chemotherapy.

More and more studies have shown that lncRNAs are new targets for the treatment of drug resistance in NPC. In the early stage, some researchers used next-generation sequencing technology to construct a diferential expression profle of lncRNA related to paclitaxel resistance in NPC, and selected lncRNA n375709 with the most significant expression diference for further research, and found that inhibiting lncRNA n375709 can increase the sensitivity of NPC 5-8F and 6-10B cells to paclitaxel [[55\]](#page-10-14). Subsequently, more and more lncRNAs have been confrmed to be involved in regulating the chemotherapy resistance of NPC, such as ROR [\[56](#page-10-15)], CCAT1 [\[57](#page-10-16)], and THOR [\[58](#page-10-17)] [[59–](#page-10-18)[63\]](#page-10-19). One of the most direct evidence comes from the latest research. KcNQ1OT1 signifcantly inhibited the viability of cisplatin-resistant NPC cells and increased the sensitivity to cisplatin by downregulating the sponge efect of USP47 on miR-454. It is suggested that KCNQ10T1/miR-454/USP47 axis is involved in the regulation of cisplatin resistance in NPC [[64](#page-10-20)]. At the same time, the researchers found that lncRNA is not only involved in the regulation of chemotherapy resistance, but also is an independent factor predicting the toxicity and efficacy of chemotherapy for NPC. For example, in the report of Youhong Wang et al., it was found that 6 potential SNPs were found in the 5 genes of the lncRNA-p53 regulatory network. They were signifcantly related to the toxicity and efficacy of CRT in 505 patients with NPC, thereby predicting the efficacy and acute toxicity provides new biomarkers [[65\]](#page-10-21). In addition, the LncRNA Gas5 gene polymorphisms rs2067079 and rs6790 may cause severe bone marrow suppression and severe neutropenia in patients with NPC and may be used as a predictive biomarker for the toxicity of radiotherapy and chemotherapy in NPC patients [[66\]](#page-10-22).

In the fgure below, we will summarize the lncRNAs related to chemotherapy resistance in NPC and briefy reveal their possible molecular mechanisms (Fig. [3\)](#page-6-0).

Long non‑coding RNAs and traditional Chinese medicine in NPC

Traditional Chinese medicine (TCM) has a long history in the treatment of tumors in China and has played an active role in modern tumor treatment due to its low toxicity, strong

Fig. 3 LncRNA related to chemotherapy resistance in NPC and its molecular mechanisms

specificity, and high curative effect [[67\]](#page-10-23). Recent studies have shown that many TCMs and monomer compounds are efective in breast cancer [\[68\]](#page-10-24). It has played a signifcant antitumor efect in the treatment of cancer. Not only that, compared with radiotherapy and/or chemotherapy alone, TCM and natural products as an adjuvant treatment of radiotherapy and/or chemotherapy have signifcant efects in improving the quality of life, reducing acute adverse reactions, and enhancing immune regulation [\[69,](#page-11-5) [70](#page-11-6)]. NPC is one of the most common tumors in China, and the application of TMCs is also very extensive. The clinical efect of Chinese medicine in adjuvant treatment of NPC is also defnite [\[71](#page-11-7)]. So, does lncRNA play a role in the adjuvant treatment of nasopharyngeal carcinoma with Chinese herbal medicine? Some researchers have focused their attention on this issue.

As early as 2013, researchers observed the LncRNA profle of nasopharyngeal carcinoma cells and the efect of curcumin (Cur) on the radiosensitivity of NPC cells [[72](#page-11-8)]. Cur is a kind of polyphenol extracted from turmeric rhizome [\[73\]](#page-11-9). In vivo and in vitro experiments have confrmed that Cur has anti-tumor activity that inhibits proliferation and induces apoptosis $[73, 74]$ $[73, 74]$ $[73, 74]$ $[73, 74]$ $[73, 74]$. In NPC, Cur also has the effect of inhibiting the proliferation and metastasis of NPC cells and enhancing radiosensitivity [[75](#page-11-11), [76](#page-11-12)]. In the study of the mechanism of curcumin radio-sensitization, the researchers found that Cur can signifcantly reverse the changes in lncRNA expression induced by radiotherapy, indicating that Cur can be used as a good radiosensitizer [[72\]](#page-11-8). In modern Chinese medicine, Chinese medicine preparations of Astragalus membranaceus (HangQi) are widely used to assist in the treatment of many diseases [[77\]](#page-11-13). Calycosin is one of the main bioactive components of Astragalus membranaceus [[78\]](#page-11-14). A large number of studies have shown that mullein has anti-tumor effects in a variety of tumors [[79,](#page-11-15) [80](#page-11-16)]. In NPC, Calycosin afects the growth of NPC cells by regulating lncRNA EWSAT1 and its downstream pathways. In the overexpression of EWSAT1, it was found that the increased expression of EWSAT1 weakened the growth inhibitory effect of Calycosin on NPC cells [[81](#page-11-17)]. There are some other active ingredients of TCM that interact with lncRNA to exert anti-cancer efects in NPC. For example, Triptonide, a small molecule monomer extract from the ancient Chinese herb Tripterygium wilfordii Hook, inhibits human NPC cell growth via disrupting LncRNA THOR-IGF2BP1 signaling [\[82\]](#page-11-18). Another evidence is that polyphylloside I, extracted from the natural herb Chonglou, has anticancer activity against nasopharyngeal carcinoma by regulating IncRNA ROR and p53 signaling [[83](#page-11-19)]. In summary, with the deepening of TCMs research, the role and mechanism of lncRNA in TCMs treatment will be gradually revealed. lncRNA is expected to become a new molecular target for TCMs therapy.

Long non‑coding RNAs and tumor diagnosis and prognosis biomarkers in NPC

In recent years, the research on diagnostic and prognostic biomarkers related to lncRNA has made great progress. The lncRNAs in circulating serum or plasma have great potential as diagnostic or prognostic markers of diseases, especially in certain types of cancer, because they are stable, available, and closely related to diseases [\[84\]](#page-11-20). Compared with classic biopsy, simple molecular biology techniques (PCR, sequencing) can be used for non-invasive detection [[85\]](#page-11-21). In addition, they have diferent expressions in biological fuids, and their expression is tissue-specifc. On this basis, combining diferent lncRNAs with conventional biomarkers can obtain an efective diagnosis and/ or prognosis. The abnormal expression of several lncR-NAs is closely related to the prognosis of NPC and can be used as potential prognostic indicators. A 2013 study confrmed that NPC patients with high levels of lncRNA HOTAIR have a poor overall survival prognosis. HOTAIR is an independent prognostic indicator of NPC progression and survival [[86\]](#page-11-22). In addition, the expression level of LINC01420 in NPC tissues were higher than nasopharyngeal epithelial (NPE) tissues [[87](#page-11-23)]. The overall survival rate of NPC patients with high expression of LINC01420 is lower. LINC01420 may play a key role in the progression of NPC and may become a potential prognostic biomarker for NPC patients. One study recruited 101 NPC patients, 20 chronic nasopharyngitis (CN) patients, 20 EBV carriers (EC), and 101 healthy controls. Through qRT-PCR and LncRNADisease database, three NPC diagnostic tools have been established. A profile of three circulating lncR-NAs (MALAT1, AFAP1-AS1, and AL359062) was established for NPC diagnosis. The high levels of these three lncRNAs are closely related to late NPC metastasis and EBV infection. After treatment, the serum levels of these three lncRNAs decreased signifcantly, indicating that circulating MALAT1, AFAP1AS1, and AL359062 may be new serum markers for the diagnosis and prognosis of NPC after treatment [\[88\]](#page-11-24).

With the deepening of people's understanding of lncRNA as a serum diagnosis and prognostic marker of NPC, more and more studies are focused on screening reliable serum lncRNA for NPC. A new study reveals the role of FOXP4-AS1 as a potential diagnostic marker and independent prognostic factor in NPC patients. The researchers found that the expression of FOXP4-AS1 was related to T stage, lymph node metastasis, tumor histological grade, and clinical stage, and the PFS and OS of patients with high expression of FOXP4-AS1 were lower than those with low expression of FOXP4-AS1 [\[89\]](#page-11-25).

Conclusion

Nasopharyngeal carcinoma is a common malignant tumor in Guangdong, China and other places. Its onset location is hidden, its early symptoms are not typical, it lacks specific serum diagnostic markers, and it is prone to recurrence and early metastasis, leading to its poor prognosis. The development of high-throughput sequencing technology has greatly promoted the identifcation and expression profle research of long non-coding RNAs in normal cells and tumor cells. Increasing evidence shows that the abnormal expression of lncRNA in diferent tumor types is related to the specifc biopathological characteristics of the tumor itself, the outcome of the disease, and the response to drug treatment.

This article focuses on the clinical application of lncR-NAs in NPC, especially the susceptibility factors (EB virus infection), the potential applications in NPC treatment resistance, and the efficacy and prognosis of traditional natural medicines. EB virus is a tumorigenic virus of nasopharyngeal carcinoma. EB virus can participate in the progression of NPC through its own encoding lncRNA. EB virus can also regulate the malignant biological behavior of NPC by encoding miRNA and then targeting lncRNA of the host. These fndings provide potential targets or strategies for new EBV-related cancer treatments, as well as new insights for understanding the carcinogenic mechanisms related to EBV infection. A variety of lncRNAs can promote or inhibit the efects of radiation or chemotherapy on NPC to varying degrees. Some lncRNA induce tumor cells to produce treatment resistance, but some lncRNA can enhance the sensitivity of tumor cells to treatment. Although the mechanism by which lncRNA regulates the sensitivity of treatment is still unclear, related lncRNA can be regarded as a breakthrough for patients with treatment resistance. For example, highly expressed lncRNAs can be specifcally silenced for treatment-resistant patients, and related signaling pathways can be monitored to predict the treatment efect and prognosis of cancer patients, so as to more efectively carry out individualized treatment. In China, the active ingredients of traditional natural herbal medicines are used in the treatment of NPC and have shown certain anti-tumor efects and reduce side efects of radiotherapy and chemotherapy. By observing the role of lncRNA in regulating the malignant phenotype of NPC cells by the efective components of Chinese herbal medicine, we believe that lncRNA has played a role in the auxiliary treatment of NPC by Chinese herbal medicine. This view provides a theoretical basis for traditional Chinese medicine to treat tumors, and it also enriches the complex network of lncRNA regulating tumors. LncRNA may be an emerging molecular target for traditional Chinese medicine to treat tumors. The specifc expression pattern of lncRNA provides a unique opportunity for fne regulation of lncRNA-targeting therapy for nasopharyngeal carcinoma. The treatment of tumor suppressor lncRNA will also provide a new dawn for the treatment of nasopharyngeal carcinoma. We believe that understanding the role of lncRNAs in the occurrence and development of NPC may help to use it as a diagnostic monitoring tool and a biological target for judging prognosis.

LncRNA has shown complex and powerful functions in NPC, but this is still not the whole truth. The next research plan of nasopharyngeal cancer research will focus on discovering new molecular biomarkers related to prognosis and diagnosis, improving existing screening and treatment strategies, and developing new treatment options for refractory relapsed diseases. lncRNA is considered to have considerable clinical value in oncology, and its potential clinical application is of great signifcance.

Author contribution YT: Data Curation, Visualization and Writing— Original Draft; XH: Writing—Review & Editing. All authors read and approved the fnal manuscript.

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Data availability The data used to support the fndings of this study are included in the article.

Declarations

Conflict of interest The authors declare that they have no conficts of interest.

References

- 1. Chua M, Wee J, Hui E, Chan A (2016) Nasopharyngeal carcinoma. Lancet (London, England) 387:1012–1024. [https://doi.](https://doi.org/10.1016/s0140-6736(15)00055-0) [org/10.1016/s0140-6736\(15\)00055-0](https://doi.org/10.1016/s0140-6736(15)00055-0)
- 2. Bray F, Ferlay J, Soerjomataram I et al (2018) Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA: Cancer J Clin 68:394–424. <https://doi.org/10.3322/caac.21492>
- 3. Wang K, Chang H (2011) Molecular mechanisms of long noncoding RNAs. Mol Cell 43:904–914. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.molcel.2011.08.018) [molcel.2011.08.018](https://doi.org/10.1016/j.molcel.2011.08.018)
- 4. Ransohof J, Wei Y, Khavari P (2018) The functions and unique features of long intergenic non-coding RNA. Nat Rev Mol Cell Biol 19:143–157. <https://doi.org/10.1038/nrm.2017.104>
- 5. Yao R, Wang Y, Chen L (2019) Cellular functions of long noncoding RNAs. Nat Cell Biol 21:542–551. [https://doi.org/10.](https://doi.org/10.1038/s41556-019-0311-8) [1038/s41556-019-0311-8](https://doi.org/10.1038/s41556-019-0311-8)
- 6. Zhang K, Tan X, Guo L (2020) The long non-coding RNA DANCR regulates the infammatory phenotype of breast cancer cells and promotes breast cancer progression via EZH2-dependent suppression of SOCS3 transcription. Mol Oncol 14:309–328. <https://doi.org/10.1002/1878-0261.12622>
- 7. Yuan S, Wang J, Yang F et al (2016) Long noncoding RNA DANCR increases stemness features of hepatocellular carcinoma by derepression of CTNNB1. Hepatology (Baltimore, Md.) 63:499–511. <https://doi.org/10.1002/hep.27893>
- 8. Jiang N, Wang X, Xie X et al (2017) lncRNA DANCR promotes tumor progression and cancer stemness features in osteosarcoma by upregulating AXL via miR-33a-5p inhibition. Cancer Lett 405:46–55.<https://doi.org/10.1016/j.canlet.2017.06.009>
- 9. Wang Y, Zeng X, Wang N et al (2018) Long noncoding RNA DANCR, working as a competitive endogenous RNA, promotes ROCK1-mediated proliferation and metastasis via decoying of miR-335-5p and miR-1972 in osteosarcoma. Mol Cancer 17:89. <https://doi.org/10.1186/s12943-018-0837-6>
- 10. Zhang X, Zhou Y, Mehta K et al (2003) A pituitary-derived MEG3 isoform functions as a growth suppressor in tumor cells. J Clin Endocrinol Metab 88:5119–5126. [https://doi.org/10.1210/](https://doi.org/10.1210/jc.2003-030222) [jc.2003-030222](https://doi.org/10.1210/jc.2003-030222)
- 11. Benetatos L, Vartholomatos G, Hatzimichael E (2011) MEG3 imprinted gene contribution in tumorigenesis. Int J Cancer 129:773–779.<https://doi.org/10.1002/ijc.26052>
- 12. Han T, Zhuo M, Yuan C et al (2020) Coordinated silencing of the Sp1-mediated long noncoding RNA MEG3 by EZH2 and HDAC3 as a prognostic factor in pancreatic ductal adenocarcinoma. Cancer Biol Med 17:953–969. [https://doi.org/10.20892/j.](https://doi.org/10.20892/j.issn.2095-3941.2019.0427) [issn.2095-3941.2019.0427](https://doi.org/10.20892/j.issn.2095-3941.2019.0427)
- 13. Ji Y, Feng G, Hou Y et al (2020) Long noncoding RNA MEG3 decreases the growth of head and neck squamous cell carcinoma by regulating the expression of miR-421 and E-cadherin. Cancer Med 9:3954–3963. <https://doi.org/10.1002/cam4.3002>
- 14. Zhang Y, Wu J, Jing H et al (2019) Long noncoding RNA MEG3 inhibits breast cancer growth via upregulating endoplasmic reticulum stress and activating NF-κB and p53. J Cell Biochem 120:6789–6797. <https://doi.org/10.1002/jcb.27982>
- 15. Chak W, Lung R, Tong J et al (2017) Downregulation of long non-coding RNA MEG3 in nasopharyngeal carcinoma. Mol Carcinog 56:1041–1054. <https://doi.org/10.1002/mc.22569>
- 16. Farrell P (2019) Epstein-Barr virus and cancer. Annu Rev Pathol 14:29–53. [https://doi.org/10.1146/annurev-pathm](https://doi.org/10.1146/annurev-pathmechdis-012418-013023) [echdis-012418-013023](https://doi.org/10.1146/annurev-pathmechdis-012418-013023)
- 17. You R, Liu Y, Lin M et al (2019) Relationship of circulating tumor cells and Epstein-Barr virus DNA to progression-free survival and overall survival in metastatic nasopharyngeal carcinoma patients. Int J Cancer 145:2873–2883. [https://doi.org/10.](https://doi.org/10.1002/ijc.32380) [1002/ijc.32380](https://doi.org/10.1002/ijc.32380)
- 18. Chan A, Lo Y, Zee B et al (2002) Plasma Epstein-Barr virus DNA and residual disease after radiotherapy for undiferentiated nasopharyngeal carcinoma. J Natl Cancer Inst 94:1614–1619. <https://doi.org/10.1093/jnci/94.21.1614>
- 19. Kanda T, Yajima M, Ikuta K (2019) Epstein-Barr virus strain variation and cancer. Cancer Sci 110:1132–1139. [https://doi.org/](https://doi.org/10.1111/cas.13954) [10.1111/cas.13954](https://doi.org/10.1111/cas.13954)
- 20. Lu T, Guo Q, Lin K et al (2020) Circulating Epstein-Barr virus microRNAs BART7-3p and BART13-3p as novel biomarkers in nasopharyngeal carcinoma. Cancer Sci 111:1711–1723. [https://](https://doi.org/10.1111/cas.14381) doi.org/10.1111/cas.14381
- 21. Xu M, Yao Y, Chen H et al (2019) Genome sequencing analysis identifes Epstein-Barr virus subtypes associated with high risk of nasopharyngeal carcinoma. Nat Genet 51:1131–1136. [https://](https://doi.org/10.1038/s41588-019-0436-5) doi.org/10.1038/s41588-019-0436-5
- 22. Li Z, Tsai M, Shumilov A et al (2019) Epstein-Barr virus ncRNA from a nasopharyngeal carcinoma induces an infammatory response that promotes virus production. Nat Microbiol 4:2475–2486.<https://doi.org/10.1038/s41564-019-0546-y>
- 23. Guo R, Tang L, Mao Y et al (2019) Proposed modifcations and incorporation of plasma Epstein-Barr virus DNA improve the TNM staging system for Epstein-Barr virus-related

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nasopharyngeal carcinoma. Cancer 125:79–89. [https://doi.org/](https://doi.org/10.1002/cncr.31741) [10.1002/cncr.31741](https://doi.org/10.1002/cncr.31741)

- 24. Zhang J, Zhang S, Zuo L et al (2019) Diferential expression profling of lncRNAs related to Epstein-Barr virus infection in the epithelial cells. J Med Virol 91:1845–1855. [https://doi.org/](https://doi.org/10.1002/jmv.25516) [10.1002/jmv.25516](https://doi.org/10.1002/jmv.25516)
- 25. Wang H, Liu W, Luo B (2021) The roles of miRNAs and lncR-NAs in Epstein-Barr virus associated epithelial cell tumors. Virus Res 291:198217. [https://doi.org/10.1016/j.virusres.2020.](https://doi.org/10.1016/j.virusres.2020.198217) [198217](https://doi.org/10.1016/j.virusres.2020.198217)
- 26. Marquitz A, Mathur A, Edwards R, Raab-Traub N (2015) Host gene expression is regulated by two types of noncoding RNAs transcribed from the Epstein-Barr virus BamHI A rightward transcript region. J Virol 89:11256–11268. [https://doi.org/10.](https://doi.org/10.1128/jvi.01492-15) [1128/jvi.01492-15](https://doi.org/10.1128/jvi.01492-15)
- 27. Park R, Miller G (2018) Epstein-Barr virus-induced nodules on viral replication compartments contain RNA processing proteins and a viral long noncoding RNA. J Virol. [https://doi.](https://doi.org/10.1128/jvi.01254-18) [org/10.1128/jvi.01254-18](https://doi.org/10.1128/jvi.01254-18)
- 28. Verhoeven R, Tong S, Mok B et al (2019) Epstein-Barr virus BART long non-coding RNAs Function as epigenetic modulators in nasopharyngeal carcinoma. Front Oncol 9:1120. [https://](https://doi.org/10.3389/fonc.2019.01120) doi.org/10.3389/fonc.2019.01120
- 29. Verhoeven R, Tong S, Zhang G et al (2016) NF-κB signaling regulates expression of Epstein-Barr virus BART MicroRNAs and long noncoding RNAs in nasopharyngeal carcinoma. J Virol 90:6475–6488. <https://doi.org/10.1128/jvi.00613-16>
- 30. Song K, Yang S, Hwang J, Kim J, Kang M (2015) The fulllength DNA sequence of Epstein Barr virus from a human gastric carcinoma cell line, SNU-719. Virus Genes 51:329–337. <https://doi.org/10.1007/s11262-015-1248-z>
- 31. Tan C, Peng C, Huang Y et al (2002) Efects of NPC-associated gene NAG7 on cell cycle and apoptosis in nasopharyngeal carcinoma cells. Ai zheng Chinese J Cancer 21:449–55
- 32. Huang C, Wu M, Tang Y et al (2009) NAG7 promotes human nasopharyngeal carcinoma invasion through inhibition of estrogen receptor alpha and up-regulation of JNK2/AP-1/ MMP1 pathways. J Cell Physiol 221:394–401. [https://doi.org/](https://doi.org/10.1002/jcp.21867) [10.1002/jcp.21867](https://doi.org/10.1002/jcp.21867)
- 33. Samanta M, Takada K (2010) Modulation of innate immunity system by Epstein-Barr virus-encoded non-coding RNA and oncogenesis. Cancer Sci 101:29–35. [https://doi.org/10.1111/j.](https://doi.org/10.1111/j.1349-7006.2009.01377.x) [1349-7006.2009.01377.x](https://doi.org/10.1111/j.1349-7006.2009.01377.x)
- 34. Zhang W, Huang C, Gong Z et al (2013) Expression of LINC00312, a long intergenic non-coding RNA, is negatively correlated with tumor size but positively correlated with lymph node metastasis in nasopharyngeal carcinoma. J Mol Histol 44:545–554.<https://doi.org/10.1007/s10735-013-9503-x>
- 35. He B, Li W, Wu Y et al (2016) Epstein-Barr virus-encoded miR-BART6-3p inhibits cancer cell metastasis and invasion by targeting long non-coding RNA LOC553103. Cell Death Dis 7:e2353.<https://doi.org/10.1038/cddis.2016.253>
- 36. Wu Y, Tang M, Wu Y et al (2014) A combination of paclitaxel and siRNA-mediated silencing of Stathmin inhibits growth and promotes apoptosis of nasopharyngeal carcinoma cells. Cell Oncol (Dordr) 37:53–67. [https://doi.org/10.1007/](https://doi.org/10.1007/s13402-013-0163-3) [s13402-013-0163-3](https://doi.org/10.1007/s13402-013-0163-3)
- 37. Wang D, Zeng Z, Zhang S et al (2020) Epstein-Barr virusencoded miR-BART6-3p inhibits cancer cell proliferation through the LOC553103-STMN1 axis. FASEB J 34:8012– 8027. [https://doi.org/10.1096/f.202000039RR](https://doi.org/10.1096/fj.202000039RR)
- 38. Chuang W, Chang S, Yu W et al (2020) Successful identifcation of nasopharyngeal carcinoma in nasopharyngeal biopsies using deep learning. Cancers (Basel). [https://doi.org/10.3390/](https://doi.org/10.3390/cancers12020507) [cancers12020507](https://doi.org/10.3390/cancers12020507)
- 39. Wu P, Zhao Y, Xiang L, Yang L (2020) Management of chemotherapy for stage II nasopharyngeal carcinoma in the intensity-modulated radiotherapy era: a review. Cancer Manag Res 12:957–963. <https://doi.org/10.2147/cmar.S239729>
- 40. Lee N, Harris J, Garden A et al (2009) Intensity-modulated radiation therapy with or without chemotherapy for nasopharyngeal carcinoma: radiation therapy oncology group phase II trial 0225. J Clin Oncol 27:3684–3690. [https://doi.org/10.1200/jco.2008.19.](https://doi.org/10.1200/jco.2008.19.9109) [9109](https://doi.org/10.1200/jco.2008.19.9109)
- 41. Lee A, Chow J, Lee N (2020) Treatment deescalation strategies for nasopharyngeal cancer: a review. JAMA Oncol. [https://doi.](https://doi.org/10.1001/jamaoncol.2020.6154) [org/10.1001/jamaoncol.2020.6154](https://doi.org/10.1001/jamaoncol.2020.6154)
- 42. Liu S, Sun X, Lu Z et al (2020) Nomogram predicting the benefts of adding concurrent chemotherapy to intensity-modulated radiotherapy after induction chemotherapy in stages II-IVb nasopharyngeal carcinoma. Front Oncol 10:539321. [https://doi.org/](https://doi.org/10.3389/fonc.2020.539321) [10.3389/fonc.2020.539321](https://doi.org/10.3389/fonc.2020.539321)
- 43. Zheng R, Yao Q, Ren C et al (2016) Upregulation of long noncoding RNA small nucleolar RNA host gene 18 promotes radioresistance of glioma by repressing semaphorin 5A. Int J Radiat Oncol Biol Phys 96:877–887. [https://doi.org/10.1016/j.ijrobp.](https://doi.org/10.1016/j.ijrobp.2016.07.036) [2016.07.036](https://doi.org/10.1016/j.ijrobp.2016.07.036)
- 44. Wang B, Zheng J, Li R et al (2019) Long noncoding RNA LINC02582 acts downstream of miR-200c to promote radioresistance through CHK1 in breast cancer cells. Cell Death Dis 10:764. <https://doi.org/10.1038/s41419-019-1996-0>
- 45. Xiao J, Lin L, Luo D et al (2020) Long noncoding RNA TRPM2- AS acts as a microRNA sponge of miR-612 to promote gastric cancer progression and radioresistance. Oncogenesis 9:29. <https://doi.org/10.1038/s41389-020-0215-2>
- 46. Huang D, Zhu X, Wang Y, Yu H, Pu Y (2020) Long non-coding RNA FAM133B-2 represses the radio-resistance of nasopharyngeal cancer cells by targeting miR-34a-5p/CDK6 axis. Aging 12:16936–16950.<https://doi.org/10.18632/aging.103600>
- 47. Peng J, Liu F, Zheng H, Wu Q, Liu S (2020) IncRNA ZFAS1 contributes to the radioresistance of nasopharyngeal carcinoma cells by sponging hsa-miR-7–5p to upregulate ENO2. Cell cycle (Georgetown, Tex). [https://doi.org/10.1080/15384101.2020.](https://doi.org/10.1080/15384101.2020.1864128) [1864128](https://doi.org/10.1080/15384101.2020.1864128)
- 48. Han Y, Li F, Xie J, Wang Y, Zhang H (2020) PVT1 mediates cell proliferation, apoptosis and radioresistance in nasopharyngeal carcinoma through regulating miR-515-5p/PIK3CA Axis. Cancer Manag Res 12:10077–10090. [https://doi.org/10.2147/cmar.](https://doi.org/10.2147/cmar.S257583) [S257583](https://doi.org/10.2147/cmar.S257583)
- 49. Han Y, Liu K, Xie J et al (2020) LINC00114 promoted nasopharyngeal carcinoma progression and radioresistance in vitro and in vivo through regulating ERK/JNK signaling pathway via targeting miR-203. Eur Rev Med Pharmacol Sci 24:2491–2504. https://doi.org/10.26355/eurrev_202003_20517
- 50. Liu H, Zheng W, Chen Q et al (2021) lncRNA CASC19 contributes to radioresistance of nasopharyngeal carcinoma by promoting autophagy via AMPK-mTOR pathway. Int J Mol Sci. [https://](https://doi.org/10.3390/ijms22031407) doi.org/10.3390/ijms22031407
- 51. Guo Z, Wang Y, Xu H et al (2021) LncRNA linc00312 suppresses radiotherapy resistance by targeting DNA-PKcs and impairing DNA damage repair in nasopharyngeal carcinoma. Cell Death Dis 12:69.<https://doi.org/10.1038/s41419-020-03302-2>
- 52. Zhang Y, Chen L, Hu G et al (2019) Gemcitabine and cisplatin induction chemotherapy in nasopharyngeal carcinoma. N Engl J Med 381:1124–1135.<https://doi.org/10.1056/NEJMoa1905287>
- 53. Li W, Chen N, Zhang N et al (2019) Concurrent chemoradiotherapy with/without induction chemotherapy in locoregionally advanced nasopharyngeal carcinoma: Long-term results of phase 3 randomized controlled trial. Int J Cancer 145:295–305. [https://](https://doi.org/10.1002/ijc.32099) doi.org/10.1002/ijc.32099
-
- 54. Chen Y, Ismaila N, Chua M et al (2021) Chemotherapy in Combination With Radiotherapy for Defnitive-Intent Treatment of Stage II-IVA Nasopharyngeal Carcinoma: CSCO and ASCO Guideline. J Clin Oncol.<https://doi.org/10.1200/jco.20.03237>
- 55. Ren S, Li G, Liu C et al (2016) Next generation deep sequencing identifed a novel lncRNA n375709 associated with paclitaxel resistance in nasopharyngeal carcinoma. Oncol Rep 36:1861– 1867.<https://doi.org/10.3892/or.2016.4981>
- 56. Li L, Gu M, You B et al (2016) Long non-coding RNA ROR promotes proliferation, migration and chemoresistance of nasopharyngeal carcinoma. Cancer Sci 107:1215–1222. [https://doi.](https://doi.org/10.1111/cas.12989) [org/10.1111/cas.12989](https://doi.org/10.1111/cas.12989)
- 57. Wang Q, Zhang W, Hao S (2017) LncRNA CCAT1 modulates the sensitivity of paclitaxel in nasopharynx cancers cells via miR-181a/CPEB2 axis. Cell cycle (Georgetown, Tex.) 16:795–801. <https://doi.org/10.1080/15384101.2017.1301334>
- 58. Gao L, Cheng X, Cao H (2018) LncRNA THOR attenuates cisplatin sensitivity of nasopharyngeal carcinoma cells via enhancing cells stemness. Biochimie 152:63–72. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.biochi.2018.06.015) [biochi.2018.06.015](https://doi.org/10.1016/j.biochi.2018.06.015)
- 59. Lin F, Lin X, Xu L, Zhu S (2020) Long noncoding RNA HOXA11-AS modulates the resistance of nasopharyngeal carcinoma cells to Cisplatin via miR-454-3p/c-Met. Mol Cells 43:856–869. <https://doi.org/10.14348/molcells.2020.0133>
- 60. Zheng Z, Li Z, Guan J et al (2020) Long noncoding RNA TINCR-mediated regulation of Acetyl-CoA metabolism promotes nasopharyngeal carcinoma progression and chemoresistance. Cancer Res 80:5174–5188. [https://doi.org/10.1158/0008-](https://doi.org/10.1158/0008-5472.Can-19-3626) [5472.Can-19-3626](https://doi.org/10.1158/0008-5472.Can-19-3626)
- 61. Zhu H (2020) Silencing long non-coding RNA H19 combined with paclitaxel inhibits nasopharyngeal carcinoma progression. Int J Pediatr Otorhinolaryngol 138:110249. [https://doi.org/10.](https://doi.org/10.1016/j.ijporl.2020.110249) [1016/j.ijporl.2020.110249](https://doi.org/10.1016/j.ijporl.2020.110249)
- 62. Liu F, Tai Y, Ma J (2018) LncRNA NEAT1/let-7a-5p axis regulates the cisplatin resistance in nasopharyngeal carcinoma by targeting Rsf-1 and modulating the Ras-MAPK pathway. Cancer Biol Ther 19:534–542. [https://doi.org/10.1080/15384047.2018.](https://doi.org/10.1080/15384047.2018.1450119) [1450119](https://doi.org/10.1080/15384047.2018.1450119)
- 63. Li H, Huang J, Yu S, Lou Z (2020) Long Non-Coding RNA DLEU1 Up-Regulates BIRC6 Expression by Competitively Sponging miR-381-3p to Promote Cisplatin Resistance in Nasopharyngeal Carcinoma. Onco Targets Ther 13:2037–2045. <https://doi.org/10.2147/ott.S237456>
- 64. Yuan F, Lou Z, Zhou Z, Yan X (2021) Long non-coding RNA KCNQ1OT1 promotes nasopharyngeal carcinoma cell cisplatin resistance via the miR-454/USP47 axis. Int J Mol Med 47:1. <https://doi.org/10.3892/ijmm.2021.4887>
- 65. Wang Y, Guo Z, Zhao Y et al (2017) Genetic polymorphisms of lncRNA-p53 regulatory network genes are associated with concurrent chemoradiotherapy toxicities and efficacy in nasopharyngeal carcinoma patients. Sci Rep 7:8320. [https://doi.org/](https://doi.org/10.1038/s41598-017-08890-2) [10.1038/s41598-017-08890-2](https://doi.org/10.1038/s41598-017-08890-2)
- 66. Guo Z, Wang Y, Zhao Y et al (2017) Genetic polymorphisms of long non-coding RNA GAS5 predict platinum-based concurrent chemoradiotherapy response in nasopharyngeal carcinoma patients. Oncotarget 8:62286–62297. [https://doi.org/10.18632/](https://doi.org/10.18632/oncotarget.19725) [oncotarget.19725](https://doi.org/10.18632/oncotarget.19725)
- 67. Liu Y, Yang S, Wang K et al (2020) Cellular senescence and cancer: Focusing on traditional Chinese medicine and natural products. Cell Prolif 53:e12894. [https://doi.org/10.1111/cpr.](https://doi.org/10.1111/cpr.12894) [12894](https://doi.org/10.1111/cpr.12894)
- 68. Liu Y, Hsiao C, Tzang B, Hsu T (2019) In vitro and in vivo efects of traditional Chinese medicine formula T33 in human breast cancer cells. BMC Complement Altern Med 19:211. <https://doi.org/10.1186/s12906-019-2630-5>
- 69. Bao H, Gao J, Huang T et al (2010) Relationship between traditional Chinese medicine syndrome diferentiation and imaging characterization to the radiosensitivity of nasopharyngeal carcinoma. Chin J Cancer 29:937–945. [https://doi.org/10.5732/](https://doi.org/10.5732/cjc.010.10209) [cjc.010.10209](https://doi.org/10.5732/cjc.010.10209)
- 70. Lou J, Yao P, Tsim K (2018) Cancer treatment by using traditional Chinese medicine: probing active compounds in antimultidrug resistance during drug therapy. Curr Med Chem 25:5128–5141. [https://doi.org/10.2174/092986732466617](https://doi.org/10.2174/0929867324666170920161922) [0920161922](https://doi.org/10.2174/0929867324666170920161922)
- 71. Cho W, Chen H (2009) Clinical efficacy of traditional Chinese medicine as a concomitant therapy for nasopharyngeal carcinoma: a systematic review and meta-analysis. Cancer Invest 27:334–344.<https://doi.org/10.1080/07357900802392683>
- 72. Wang Q, Fan H, Liu Y et al (2014) Curcumin enhances the radiosensitivity in nasopharyngeal carcinoma cells involving the reversal of diferentially expressed long non-coding RNAs. Int J Oncol 44:858–864.<https://doi.org/10.3892/ijo.2013.2237>
- 73. Shao M, Lou D, Yang J et al (2020) Curcumin and wikstrofavone B, a new bifavonoid isolated from Wikstroemia indica, synergistically suppress the proliferation and metastasis of nasopharyngeal carcinoma cells via blocking FAK/STAT3 signaling pathway. Phytomedicine 79:153341. [https://doi.org/](https://doi.org/10.1016/j.phymed.2020.153341) [10.1016/j.phymed.2020.153341](https://doi.org/10.1016/j.phymed.2020.153341)
- 74. Heger M, van Golen R, Broekgaarden M, Michel M (2014) The molecular basis for the pharmacokinetics and pharmacodynamics of curcumin and its metabolites in relation to cancer. Pharmacol Rev 66:222–307.<https://doi.org/10.1124/pr.110.004044>
- 75. Gao W, Chan J, Wong T (2014) Curcumin exerts inhibitory efects on undiferentiated nasopharyngeal carcinoma by inhibiting the expression of miR-125a-5p. Clinical science (London, England: 1979) 127:571–9. [https://doi.org/10.1042/cs201](https://doi.org/10.1042/cs20140010) [40010](https://doi.org/10.1042/cs20140010)
- 76. Yang J, Zhu D, Liu S et al (2020) Curcumin enhances radiosensitization of nasopharyngeal carcinoma by regulating circRNA network. Mol Carcinog 59:202–214. [https://doi.org/10.1002/](https://doi.org/10.1002/mc.23143) [mc.23143](https://doi.org/10.1002/mc.23143)
- 77. Wu X, Wang Y, Cheng J, Zhao Y (2006) Calcium channel blocking activity of calycosin, a major active component of Astragali Radix, on rat aorta. Acta Pharmacol Sin 27:1007–1012. [https://](https://doi.org/10.1111/j.1745-7254.2006.00349.x) doi.org/10.1111/j.1745-7254.2006.00349.x
- 78. Li J, Harata-Lee Y, Denton M et al (2017) Astragalus membranaceusLong read reference genome-free reconstruction of a fulllength transcriptome from reveals transcript variants involved in bioactive compound biosynthesis. Cell discovery 3:17031. <https://doi.org/10.1038/celldisc.2017.31>
- 79. Tian J, Wang Y, Zhang X et al (2017) Calycosin inhibits the in vitro and in vivo growth of breast cancer cells through WDR7-7-GPR30 Signaling. Journal of experimental & clinical cancer research : CR 36:153. [https://doi.org/10.1186/](https://doi.org/10.1186/s13046-017-0625-y) [s13046-017-0625-y](https://doi.org/10.1186/s13046-017-0625-y)
- 80. El-Kott A, Al-Kahtani M, Shati A (2019) Calycosin induces apoptosis in adenocarcinoma HT29 cells by inducing cytotoxic autophagy mediated by SIRT1/AMPK-induced inhibition of Akt/ mTOR. Clin Exp Pharmacol Physiol 46:944–954. [https://doi.org/](https://doi.org/10.1111/1440-1681.13133) [10.1111/1440-1681.13133](https://doi.org/10.1111/1440-1681.13133)
- 81. Kong L, Li X, Wang H, He G, Tang A (2018) Calycosin inhibits nasopharyngeal carcinoma cells by infuencing EWSAT1 expression to regulate the TRAF6-related pathways. Biomed Pharmacother 106:342–348. [https://doi.org/10.1016/j.biopha.2018.06.](https://doi.org/10.1016/j.biopha.2018.06.143) [143](https://doi.org/10.1016/j.biopha.2018.06.143)
- 82. Wang S, Lv Y, Xu X et al (2019) Triptonide inhibits human nasopharyngeal carcinoma cell growth via disrupting Lnc-RNA THOR-IGF2BP1 signaling. Cancer Lett 443:13–24. [https://doi.](https://doi.org/10.1016/j.canlet.2018.11.028) [org/10.1016/j.canlet.2018.11.028](https://doi.org/10.1016/j.canlet.2018.11.028)
- 83. Hong F, Gu W, Jiang J, Liu X, Jiang H (2019) Anticancer activity of polyphyllin I in nasopharyngeal carcinoma by modulation of lncRNA ROR and P53 signalling. J Drug Target 27:806–811. <https://doi.org/10.1080/1061186x.2018.1561887>
- 84. Zhou R, Chen K, Zhang J et al (2018) The decade of exosomal long RNA species: an emerging cancer antagonist. Mol Cancer 17:75.<https://doi.org/10.1186/s12943-018-0823-z>
- 85. Zuo Z, Hu H, Xu Q et al (2020) BBCancer: an expression atlas of blood-based biomarkers in the early diagnosis of cancers. Nucleic Acids Res 48:D789–D796. [https://doi.org/10.1093/nar/](https://doi.org/10.1093/nar/gkz942) [gkz942](https://doi.org/10.1093/nar/gkz942)
- 86. Nie Y, Liu X, Qu S et al (2013) Long non-coding RNA HOTAIR is an independent prognostic marker for nasopharyngeal carcinoma progression and survival. Cancer Sci 104:458–464. [https://](https://doi.org/10.1111/cas.12092) doi.org/10.1111/cas.12092
- 87. Yang L, Tang Y, He Y et al (2017) High expression of LINC01420 indicates an unfavorable prognosis and modulates cell migration and invasion in nasopharyngeal carcinoma. J Cancer 8:97–103. <https://doi.org/10.7150/jca.16819>
- 88. He B, Zeng J, Chao W et al (2017) Serum long non-coding RNAs MALAT1, AFAP1-AS1 and AL359062 as diagnostic and prognostic biomarkers for nasopharyngeal carcinoma. Oncotarget 8:41166–41177. <https://doi.org/10.18632/oncotarget.17083>
- 89. Yao L, Wang T, Wang X (2021) LncRNA FOXP4-AS1 serves as a biomarker for nasopharyngeal carcinoma diagnosis and prognosis. 3 Biotech 11:25. [https://doi.org/10.1007/](https://doi.org/10.1007/s13205-020-02593-8) [s13205-020-02593-8](https://doi.org/10.1007/s13205-020-02593-8)
- 90. Li Q, Jiang Y, Zhong G et al (2020) Long noncoding RNA DANCR regulates cell proliferation by stabilizing SOX2 mRNA in nasopharyngeal carcinoma. Am J Pathol 190:2343–2354. <https://doi.org/10.1016/j.ajpath.2020.09.005>
- 91. Zhang X, Yang J, Bian Z, Shi D, Cao Z (2019) Long noncoding RNA DANCR promotes nasopharyngeal carcinoma progression by interacting with STAT3, enhancing IL-6/JAK1/STAT3 signaling. Biomed Pharmacother 113:108713. [https://doi.org/10.](https://doi.org/10.1016/j.biopha.2019.108713) [1016/j.biopha.2019.108713](https://doi.org/10.1016/j.biopha.2019.108713)
- 92. Hao Y, Zhao H, Jin X et al (2019) Long non-coding RNA DANCR promotes nasopharyngeal carcinoma cell proliferation and migration. Mol Med Rep 19:2883–2889. [https://doi.org/10.](https://doi.org/10.3892/mmr.2019.9906) [3892/mmr.2019.9906](https://doi.org/10.3892/mmr.2019.9906)
- 93. Wen X, Liu X, Mao Y et al (2018) Long non-coding RNA DANCR stabilizes HIF-1 α and promotes metastasis by interacting with NF90/NF45 complex in nasopharyngeal carcinoma. Theranostics 8:5676–5689.<https://doi.org/10.7150/thno.28538>
- 94. Zhao C, Bai X, Hu X (2020) Knockdown of lncRNA XIST inhibits hypoxia-induced glycolysis, migration and invasion through regulating miR-381-3p/NEK5 axis in nasopharyngeal carcinoma. Eur Rev Med Pharmacol Sci 24:2505–2517. [https://doi.org/10.](https://doi.org/10.26355/eurrev_202003_20518) [26355/eurrev_202003_20518](https://doi.org/10.26355/eurrev_202003_20518)
- 95. Shi J, Tan S, Song L, Song L, Wang Y (2020) LncRNA XIST knockdown suppresses the malignancy of human nasopharyngeal carcinoma through XIST/miRNA-148a-3p/ADAM17 pathway in vitro and in vivo. Biomed Pharmacother 121:109620. [https://](https://doi.org/10.1016/j.biopha.2019.109620) doi.org/10.1016/j.biopha.2019.109620
- 96. Cheng Q, Xu X, Jiang H, Xu L, Li Q (2018) Knockdown of long non-coding RNA XIST suppresses nasopharyngeal carcinoma progression by activating miR-491-5p. J Cell Biochem 119:3936–3944. <https://doi.org/10.1002/jcb.26535>
- 97. Song P, Ye L, Zhang C, Peng T, Zhou X (2016) Long non-coding RNA XIST exerts oncogenic functions in human nasopharyngeal carcinoma by targeting miR-34a-5p. Gene 592:8–14. [https://doi.](https://doi.org/10.1016/j.gene.2016.07.055) [org/10.1016/j.gene.2016.07.055](https://doi.org/10.1016/j.gene.2016.07.055)
- 98. Peng J, Liu F, Zheng H, Wu Q, Liu S (2020) Long noncoding RNA ZFAS1 promotes tumorigenesis and metastasis in nasopharyngeal carcinoma by sponging miR-892b to up-regulate

LPAR1 expression. J Cell Mol Med 24:1437–1450. [https://doi.](https://doi.org/10.1111/jcmm.14823) [org/10.1111/jcmm.14823](https://doi.org/10.1111/jcmm.14823)

- 99. Wang X, Jin Q, Wang X, Chen W, Cai Z (2019) LncRNA ZFAS1 promotes proliferation and migration and inhibits apoptosis in nasopharyngeal carcinoma via the PI3K/AKT pathway in vitro. Cancer biomarkers : section A of Disease markers 26:171–182. <https://doi.org/10.3233/cbm-182080>
- 100. Wang M, Ji Y, Song Z et al (2019) Knockdown of lncRNA ZFAS1 inhibits progression of nasopharyngeal carcinoma by sponging miR-135a. Neoplasma 66:939–945. [https://doi.org/](https://doi.org/10.4149/neo_2018_181213N963) [10.4149/neo_2018_181213N963](https://doi.org/10.4149/neo_2018_181213N963)
- 101. Chen X, Li J, Li C, Lu X (2018) Long non-coding RNA ZFAS1 promotes nasopharyngeal carcinoma through activation of Wnt/ β-catenin pathway. Eur Rev Med Pharmacol Sci 22:3423–3429. https://doi.org/10.26355/eurrev_201806_15165
- 102. Li X, Lin Y, Yang X, Wu X, He X (2016) Long noncoding RNA H19 regulates EZH2 expression by interacting with miR-630 and promotes cell invasion in nasopharyngeal carcinoma. Biochem Biophys Res Commun 473:913–919. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.bbrc.2016.03.150) [bbrc.2016.03.150](https://doi.org/10.1016/j.bbrc.2016.03.150)
- 103. Zhang T, Lei F, Jiang T et al (2019) H19/miR-675-5p targeting SFN enhances the invasion and metastasis of nasalpharyngeal cancer cells. Curr Mol Pharmacol 12:324–333. [https://doi.org/](https://doi.org/10.2174/1874467212666190719120446) [10.2174/1874467212666190719120446](https://doi.org/10.2174/1874467212666190719120446)
- 104. Zhang Y, Zhu R, Wang J et al (2019) Upregulation of lncRNA H19 promotes nasopharyngeal carcinoma proliferation and metastasis in let-7 dependent manner. Artif Cells Nanomed Biotechnol 47:3854–3861. [https://doi.org/10.1080/21691401.2019.](https://doi.org/10.1080/21691401.2019.1669618) [1669618](https://doi.org/10.1080/21691401.2019.1669618)
- 105. Hu W, Xu W, Shi Y, Dai W (2018) lncRNA HOTAIR upregulates COX-2 expression to promote invasion and migration of nasopharyngeal carcinoma by interacting with miR-101. Biochem Biophys Res Commun 505:1090–1096. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.bbrc.2018.09.190) [bbrc.2018.09.190](https://doi.org/10.1016/j.bbrc.2018.09.190)
- 106. Ma D, Yuan L, Lin L (2017) LncRNA HOTAIR contributes to the tumorigenesis of nasopharyngeal carcinoma via up-regulating FASN. Eur Rev Med Pharmacol Sci 21:5143–5152. [https://](https://doi.org/10.26355/eurrev_201711_13831) doi.org/10.26355/eurrev_201711_13831
- 107. Fu W, Lu Y, Hu B et al (2016) Long noncoding RNA Hotair mediated angiogenesis in nasopharyngeal carcinoma by direct and indirect signaling pathways. Oncotarget 7:4712–4723. <https://doi.org/10.18632/oncotarget.6731>
- 108. Hu W, Li H, Wang S (2020) LncRNA SNHG7 promotes the proliferation of nasopharyngeal carcinoma by miR-514a-5p/ ELAVL1 axis. BMC Cancer 20:376. [https://doi.org/10.1186/](https://doi.org/10.1186/s12885-020-06775-8) [s12885-020-06775-8](https://doi.org/10.1186/s12885-020-06775-8)
- 109. Dai Y, Zhang X, Xing H et al (2020) Downregulated long noncoding RNA SNHG7 restricts proliferation and boosts apoptosis of nasopharyngeal carcinoma cells by elevating microRNA-140–5p to suppress GLI3 expression. Cell cycle (Georgetown, Tex.) 19:448–463. [https://doi.org/10.1080/15384101.2020.17120](https://doi.org/10.1080/15384101.2020.1712033) [33](https://doi.org/10.1080/15384101.2020.1712033)
- 110. Cheng N, Guo Y (2017) Long noncoding RNA NEAT1 promotes nasopharyngeal carcinoma progression through regulation of miR-124/NF-κB pathway. Onco Targets Ther 10:5843–5853. <https://doi.org/10.2147/ott.S151800>
- 111. Ji Y, Wang M, Li X, Cui F (2019) The long noncoding RNA NEAT1 targets miR-34a-5p and drives nasopharyngeal Carcinoma progression via Wnt/β-catenin signaling. Yonsei Med J 60:336–345. <https://doi.org/10.3349/ymj.2019.60.4.336>
- 112. Shi B, Wang Y, Yin F (2017) MALAT1/miR-124/Capn4 axis regulates proliferation, invasion and EMT in nasopharyngeal carcinoma cells. Cancer Biol Ther 18:792–800. [https://doi.org/](https://doi.org/10.1080/15384047.2017.1373214) [10.1080/15384047.2017.1373214](https://doi.org/10.1080/15384047.2017.1373214)
- 113. Zheng Y, Zhao J, Liang T et al (2019) Long noncoding RNA SMAD5-AS1 acts as a microRNA-106a-5p sponge to promote epithelial mesenchymal transition in nasopharyngeal carcinoma. FASEB J 33:12915–12928. [https://doi.org/10.1096/f.20190](https://doi.org/10.1096/fj.201900803R) [0803R](https://doi.org/10.1096/fj.201900803R)
- 114. Li S, Zhao B, Zhao H et al (2019) Silencing of long non-coding RNA SMAD5-AS1 reverses epithelial mesenchymal transition in nasopharyngeal carcinoma via microRNA-195-dependent inhibition of SMAD5. Front Oncol 9:1246. [https://doi.org/10.3389/](https://doi.org/10.3389/fonc.2019.01246) [fonc.2019.01246](https://doi.org/10.3389/fonc.2019.01246)
- 115. Chen S, Lv L, Zhan Z et al (2020) Silencing of long noncoding RNA SRRM2-AS exerts suppressive efects on angiogenesis in nasopharyngeal carcinoma via activating MYLK-mediated cGMP-PKG signaling pathway. J Cell Physiol 235:7757–7768. <https://doi.org/10.1002/jcp.29382>
- 116. Zhang E, Li C, Xiang Y (2020) LncRNA FOXD3-AS1/miR-135a-5p function in nasopharyngeal carcinoma cells. Open medicine (Warsaw, Poland) 15:1193–1201. [https://doi.org/10.1515/](https://doi.org/10.1515/med-2020-0177) [med-2020-0177](https://doi.org/10.1515/med-2020-0177)
- 117. Hu J, Pan J, Luo Z, Duan Q, Wang D (2020) Long non-coding RNA FOXD3-AS1 silencing exerts tumor suppressive efects in nasopharyngeal carcinoma by downregulating FOXD3 expression via microRNA-185-3p upregulation. Cancer Gene Ther. <https://doi.org/10.1038/s41417-020-00242-z>
- 118. Fang M, Zhang M, Wang Y et al (2020) Long noncoding RNA AFAP1-AS1 Is a critical regulator of nasopharyngeal carcinoma tumorigenicity. Front Oncol 10:601055. [https://doi.org/10.3389/](https://doi.org/10.3389/fonc.2020.601055) [fonc.2020.601055](https://doi.org/10.3389/fonc.2020.601055)
- 119. Lian Y, Xiong F, Yang L et al (2018) Long noncoding RNA AFAP1-AS1 acts as a competing endogenous RNA of miR-423-5p to facilitate nasopharyngeal carcinoma metastasis through regulating the Rho/Rac pathway. J Exp Clin Cancer Res 37:253. <https://doi.org/10.1186/s13046-018-0918-9>
- 120. Zhong Q, Wang Z, Liao X, Wu R, Guo X (2020) LncRNA GAS5/ miR-4465 axis regulates the malignant potential of nasopharyngeal carcinoma by targeting COX2. Cell Cycle (Georgetown, Tex.) 19:3004–3017. [https://doi.org/10.1080/15384101.2020.](https://doi.org/10.1080/15384101.2020.1816280) [1816280](https://doi.org/10.1080/15384101.2020.1816280)
- 121. Zhou L, Liu R, Liang X et al (2020) lncRNA RP11-624L4.1 Is Associated with Unfavorable Prognosis and Promotes Proliferation via the CDK4/6-Cyclin D1-Rb-E2F1 Pathway in NPC. Molecular therapy Nucleic acids 22:1025–1039. [https://doi.org/](https://doi.org/10.1016/j.omtn.2020.10.017) [10.1016/j.omtn.2020.10.017](https://doi.org/10.1016/j.omtn.2020.10.017)
- 122. Fan C, Wang J, Tang Y et al (2020) Upregulation of long noncoding RNA LOC284454 may serve as a new serum diagnostic biomarker for head and neck cancers. BMC Cancer 20:917. <https://doi.org/10.1186/s12885-020-07408-w>
- 123. He S, Xu C, Li Y et al (2020) AR-induced long non-coding RNA LINC01503 facilitates proliferation and metastasis via the SFPQ-FOSL1 axis in nasopharyngeal carcinoma. Oncogene 39:5616– 5632.<https://doi.org/10.1038/s41388-020-01388-8>
- 124. Yao H, Yang L, Tian L, Guo Y, Li Y (2020) LncRNA MSC-AS1 aggravates nasopharyngeal carcinoma progression by targeting miR-524-5p/nuclear receptor subfamily 4 group A member 2 (NR4A2). Cancer Cell Int 20:138. [https://doi.org/10.1186/](https://doi.org/10.1186/s12935-020-01202-1) [s12935-020-01202-1](https://doi.org/10.1186/s12935-020-01202-1)
- 125. Tang T, Yang L, Cao Y et al (2020) LncRNA AATBC regulates Pinin to promote metastasis in nasopharyngeal carcinoma. Mol Oncol 14:2251–2270.<https://doi.org/10.1002/1878-0261.12703>
- 126. Liao B, Wang Z, Zhu Y, Wang M, Liu Y (2019) Long noncoding RNA DRAIC acts as a microRNA-122 sponge to facilitate nasopharyngeal carcinoma cell proliferation, migration and invasion via regulating SATB1. Artifcial cells, nanomedicine, and biotechnology 47:3585–3597. [https://doi.org/10.1080/21691401.](https://doi.org/10.1080/21691401.2019.1656638) [2019.1656638](https://doi.org/10.1080/21691401.2019.1656638)
- 127. Hu X, Liu W, Jiang X et al (2019) Long noncoding RNA LINC00460 aggravates invasion and metastasis by targeting miR-30a-3p/Rap1A in nasopharyngeal carcinoma. Hum Cell 32:465–476. <https://doi.org/10.1007/s13577-019-00262-4>
- 128. Kong Y, Cui M, Chen S et al (2018) LncRNA-LINC00460 facilitates nasopharyngeal carcinoma tumorigenesis through sponging miR-149-5p to up-regulate IL6. Gene 639:77–84. [https://doi.org/](https://doi.org/10.1016/j.gene.2017.10.006) [10.1016/j.gene.2017.10.006](https://doi.org/10.1016/j.gene.2017.10.006)
- 129. Zheng Z, Li Z, Zhou G et al (2019) Long noncoding RNA FAM225A promotes nasopharyngeal carcinoma tumorigenesis and metastasis by acting as ceRNA to sponge miR-590-3p/miR-1275 and upregulate ITGB3. Cancer Res 79:4612–4626. [https://](https://doi.org/10.1158/0008-5472.Can-19-0799) doi.org/10.1158/0008-5472.Can-19-0799
- 130. Gao C, Lu W, Lou W, Wang L, Xu Q (2019) Long noncoding RNA HOXC13-AS positively affects cell proliferation and invasion in nasopharyngeal carcinoma via modulating miR-383-3p/ HMGA2 axis. J Cell Physiol 234:12809–12820. [https://doi.org/](https://doi.org/10.1002/jcp.27915) [10.1002/jcp.27915](https://doi.org/10.1002/jcp.27915)
- 131. Zhang E, Li X (2019) LncRNA SOX2-OT regulates proliferation and metastasis of nasopharyngeal carcinoma cells through miR-146b-5p/HNRNPA2B1 pathway. J Cell Biochem 120:16575– 16588.<https://doi.org/10.1002/jcb.28917>
- 132. Lan X, Liu X (2019) LncRNA SNHG1 functions as a ceRNA to antagonize the efect of miR-145a-5p on the down-regulation of NUAK1 in nasopharyngeal carcinoma cell. J Cell Mol Med 23:2351–2361.<https://doi.org/10.1111/jcmm.13497>
- 133. Zou Z, Ma C, Medoro L et al (2016) LncRNA ANRIL is upregulated in nasopharyngeal carcinoma and promotes the cancer progression via increasing proliferation, reprograming cell glucose metabolism and inducing side-population stem-like cancer cells. Oncotarget 7:61741–61754. [https://doi.org/10.18632/oncot](https://doi.org/10.18632/oncotarget.11437) [arget.11437](https://doi.org/10.18632/oncotarget.11437)
- 134. Song P, Yin S (2016) Long non-coding RNA EWSAT1 promotes human nasopharyngeal carcinoma cell growth in vitro by

targeting miR-326/-330-5p. Aging 8:2948–2960. [https://doi.org/](https://doi.org/10.18632/aging.101103) [10.18632/aging.101103](https://doi.org/10.18632/aging.101103)

- 135. Chen X, Huang Y, Shi D et al (2020) ZNF667-AS1LncRNA promotes expression by adsorbing to suppress nasopharyngeal carcinoma cell progression. Onco Targets Ther 13:4397–4409. <https://doi.org/10.2147/ott.S245554>
- 136. Zhang W, Guo Q, Liu G et al (2019) NKILA represses nasopharyngeal carcinoma carcinogenesis and metastasis by NF-κB pathway inhibition. PLoS Genet 15:e1008325. [https://doi.org/10.](https://doi.org/10.1371/journal.pgen.1008325) [1371/journal.pgen.1008325](https://doi.org/10.1371/journal.pgen.1008325)
- 137. Sun Q, Liu H, Li L et al (2015) Long noncoding RNA-LET, which is repressed by EZH2, inhibits cell proliferation and induces apoptosis of nasopharyngeal carcinoma cell. Medical Oncol (Northwood, London, England) 32:226. [https://doi.org/](https://doi.org/10.1007/s12032-015-0673-0) [10.1007/s12032-015-0673-0](https://doi.org/10.1007/s12032-015-0673-0)
- 138. Chen L, Sun L, Dong L et al (2017) The role of long noncoding RNA-LET in cell proliferation and invasion of nasopharyngeal carcinoma and its mechanism. Onco Targets Ther 10:2769–2778. <https://doi.org/10.2147/ott.S126907>
- 139. Guo J, Ma J, Zhao G et al (2017) Long Noncoding RNA LINC0086 Functions as a Tumor Suppressor in Nasopharyngeal Carcinoma by Targeting miR-214. Oncol Res 25:1189–1197. <https://doi.org/10.3727/096504017x14865126670075>
- 140. Gong Z, Zhang S, Zeng Z et al (2014) LOC401317, a p53-regulated long non-coding RNA, inhibits cell proliferation and induces apoptosis in the nasopharyngeal carcinoma cell line HNE2. PLoS One 9:e110674. [https://doi.org/10.1371/journal.](https://doi.org/10.1371/journal.pone.0110674) [pone.0110674](https://doi.org/10.1371/journal.pone.0110674)

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