

Regucalcin and cell regulation: role as a suppressor protein in signal transduction

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Abstract Regucalcin was discovered in 1978 as a calcium-binding protein that does not contain EF-hand motif of calcium-binding domain (Yamaguchi and Yamamoto Chem Pharm Bull 26:1915–1918, 1978). The name regucalcin was proposed for this calcium-binding protein, which can regulate various Ca^{2+} -dependent enzyme activations in liver cells. The regucalcin gene is localized on the chromosome X, and the organization of the regucalcin gene consists of seven exons and six introns. AP-1, NF1-A1, and RGPR-p117 bind to the promoter region of the rat regucalcin gene and enhance transcription activity of regucalcin gene expression that is mediated through calcium signaling. Regucalcin plays a pivotal role in the keep of intracellular calcium ion (Ca^{2+}) homeostasis due to activating Ca^{2+} pump enzymes in the plasma membrane (basolateral membrane), microsomes (endoplasmic reticulum), mitochondria, and nuclei of many cell types. Regucalcin has a suppressive effect on calcium signaling from the cytoplasm to the nucleus in the proliferative cells. Regucalcin has also been demonstrated to transport to the nucleus, and it can inhibit Ca^{2+} -dependent protein kinase and protein phosphatase activities, Ca^{2+} -activated deoxyribonucleic acid (DNA) fragmentation, and DNA and ribonucleic acid (RNA) synthesis in the nucleus. Overexpression of regucalcin suppresses cell death and apoptosis in the cloned rat hepatoma cells induced by various signaling factors. Regucalcin can inhibit the enhancement of

cell proliferation due to hormonal stimulation. Regucalcin plays an important role as a regulatory protein in cell signaling system, and it is proposed to play a pivotal role in keep of cell homeostasis and function.

Keywords Regucalcin · RGN · Gene expression · Calcium signaling · Calmodulin · Protein kinase C · Nuclear function · Apoptosis · Cell proliferation

Introduction

Calcium ion (Ca^{2+}) plays an important role in the regulation of many cell functions. Ca^{2+} can regulate muscle contraction, neurotransmission, hormone secretion, cell mitosis, and gene expression. A role as second messengers of Ca^{2+} in cells for hormonal stimulation comes into notice. Calcium signal is transmitted to intracellular responses, which are mediated through a family of calcium-binding protein and protein kinase C [1, 2]. Liver metabolism is regulated by an increase in Ca^{2+} in the cytoplasm of liver cells due to hormonal stimulation [3–5]. The effect of Ca^{2+} is amplified through calmodulin and protein kinase C [1–5]. Calcium signaling is important in the regulation of liver metabolism.

Liver has been shown to participate in the regulation of calcium metabolism through hepatic bile system in rats, and bile calcium excretion is increased by hormonal stimulation [5, 6]. On the basis of this finding, it was found that a novel calcium-binding protein, which differs from calmodulin and other calcium-related proteins, was present in the hepatic cytoplasm of rats [7–9]. The name regucalcin was proposed for this calcium-binding protein, which regulates various Ca^{2+} - or Ca^{2+} /calmodulin-dependent enzyme activations in liver cells [10–15].

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Regucalcin and its gene (RGN) are identified in over 15 species consisting of regucalcin family [16–21]. Comparison of the nucleotide sequences of regucalcin from vertebrate species is highly conserved in their coding region with throughout evolution. The regucalcin gene is localized on the chromosome X, and the organization of the regucalcin gene consists of seven exons and six introns [21–23].

AP-1, NF1-A1, and RGPR-p117, which is a transcription factor, bind to the promoter region of the regucalcin gene and enhance transcription activity of regucalcin gene expression that is mediated through calcium and other signalings [21]. Regucalcin mRNA expression and its protein content are pronounced in the liver and kidney cortex of rats, although it is present only slightly in other tissues (including the duodenum, testis, spleen, lung, smooth muscle, heart, and brain) [17, 24–29]. The role of regucalcin is investigated in the liver and kidney cells in detail [reviewed in Ref. 30–34]. After finding of regucalcin, the identical protein to regucalcin was also reported as senescence marker protein-30 (SMP30) [35, 36].

There are growing evidences that regucalcin plays an important role as a regulatory protein for calcium signaling from the cytoplasm to the nuclei in liver cells [37]. Over-expression of regucalcin has been demonstrated to inhibit cell apoptosis and cell proliferation induced by various signaling factors. This review has been written to outline the recent advances that have been made concerning the role of regucalcin as a regulatory protein in cell signaling in liver, kidney, and other tissues: its role in regulation of intracellular Ca^{2+} homeostasis, inhibition of Ca^{2+} -dependent enzyme activations, regulation of nuclear calcium signaling, and inhibitory effects in cell apoptosis and cell proliferation induced by various signaling factors.

Properties of calcium-binding in regucalcin

The molecular weight of rat regucalcin is estimated as 33,388 Da, composing of 299 amino acid residues from the cloning of rat regucalcin cDNA [16]. The regucalcin molecule does not contain the EF-hand motif as a calcium-binding domain [16]. The isoelectric point of regucalcin is 5.20 [16].

From the experimental data by Scatchard plot, the apparent association constant (Kf) for calcium ion (Ca^{2+}) of regucalcin is found to be $4.19 \times 10^5 \text{ M}^{-1}$ by equilibrium dialysis with a correlation coefficient of 0.99, and there are 6.52 high-affinity sites per molecule of protein [7–9]. Regucalcin appears to have six or seven high-affinity binding sites for Ca^{2+} per molecule of protein [8]. Extrapolation of the regression line to infinite Ca^{2+} concentration indicated a maximal Ca^{2+} -binding of 2.28×10^{-4} mol of Ca^{2+} per gram of protein. It is known

that calmodulin exists as a monomer with a molecular weight of 17,000 and contains four Ca^{2+} -binding sites [1].

The conformational changes induced by binding of Ca^{2+} to regucalcin have been investigated by means of the ultraviolet (UV) absorption spectrum [9]. UV of regucalcin showed a maximum at 278 nm in the range from 240 to 330 nm. In the presence of Ca^{2+} (0.1 and 1.0 mM), a decrease in absorption at 278 nm was observed [9]. Such a negative UV difference is similar to the Ca^{2+} -induced absorption changes in calmodulin. The spectrum can be attributed to charges in both tyrosine and tryptophan residues. Changes in the environment of both aromatic amino acids occur upon Ca^{2+} -binding [9].

Fluorescence spectroscopy is used to study the effect of Ca^{2+} on the conformation of regucalcin [9]. The spectral emission was quenched after the addition of 1.0 mM Ca^{2+} . Known fluorescence emission properties of isolated tyrosine and tryptophan residues suggest, as indicated above, that changes in the environment of these two aromatic amino acids occur upon Ca^{2+} -binding. These observations demonstrate that Ca^{2+} -binding induces conformational changes in regucalcin. These changes may result in increasing the hydrophobicity of regucalcin [9, 16].

The conformation of the polypeptide backbone of regucalcin has been studied by circular dichroism (CD) spectroscopy [9]. The presence of 1.0 mM Ca^{2+} caused clear alterations in the CD spectrum. The apparent α -helical content of regucalcin in Ca^{2+} -free buffer was estimated to be 34%, and the presence of 1.0 mM Ca^{2+} decreased this by 4.5%. Thus, conformational changes are induced by Ca^{2+} binding to regucalcin. This binding also loosens the conformation of regucalcin. The apparent α -helical content of calmodulin is 30%, and it is increased after Ca^{2+} binding and its conformation is tightened [9, 16]. The increase in hydrophobicity after Ca^{2+} binding represents the mechanism that calmodulin activates its target proteins. However, regucalcin reverses the activation of many enzymes by Ca^{2+} /calmodulin. The role of regucalcin may be different from that of calmodulin in cells.

Regucalcin differs entirely from other calcium-binding proteins of the EF-hand type: calmodulin, calcineurin, parvalbumin, S-100a, S-100b proteins, caligulin, calregulin, calbindin, calreticulin, and annexins. This is supported from the results of the molecular cloning and sequencing of the cDNA coding for a regucalcin from various mammalian livers [16, 20]. The nucleotide and amino acid sequences of regucalcin does not have statistically significant homology, as compared with the registered sequences which are found in the EMBL and GenBank databases (D14327 and D86217) [16].

The hydropathy profile of regucalcin shows that there is a hydrophobic sequence in both N-terminal and C-terminal regions of the regucalcin molecules [16]. Regucalcin shows

a hydrophilic character as molecule [16, 20]. The most common EF-hand is composed of the helix-loop-helix-domain. The prototype loop consists of 12 amino acids, of which five have a carboxyl (or a hydroxyl group) in their side chain, precisely spaced so as to coordinate the Ca^{2+} . Analysis of the structure of the EF-hand from the regucalcin sequence did not give the expected pattern of amino acids conforming to the typical EF-hand structure of a calcium-binding site.

Amino acid analysis shows that regucalcin has a relatively high content of glycine and much lower amounts of glutamic acid, valine, aspartic acid, and lysine [16, 20]. Regucalcin contains about 20% (mol%) glutamic and aspartic acids and about 17% amide residues (lysine, histidine, and arginine) and thus a high proportion of charged residues: regucalcin molecule contains aspartic acid (24 residues) and glutamic acid (16 residues) [16]. Acidic amino acids comprise approximately one-fifth of the regucalcin molecule, a prevalence that appears to be characteristic of the composition of all Ca^{2+} -binding proteins. The significance of the high di-carboxylic acid content is that it permits binding of Ca^{2+} to protein. These amino acids may be related to Ca^{2+} binding.

The result of crystal structure with X-ray diffraction data shows that regucalcin contains the metal site bound with either a Ca^{2+} or a Zn^{2+} atom, suggesting that the Ca^{2+} -bound form may be physiologically relevant for stressed cells with an elevated Ca^{2+} level [38]. This supports our finding for regucalcin as a calcium-binding protein.

Regulation of regucalcin gene expression

The rat regucalcin gene is localized on the proximal end of the rat chromosome Xq11.1-12 [22], and the gene is demonstrated in human, mouse, cow, monkey, dog, rabbit, and chicken but not in yeast [17]. The amino acid sequence of mouse regucalcin had 94% homology as compared with that of rat regucalcin [20]. Regucalcin may be a protein that is highly differentiated. The organization of the rat regucalcin gene seems to be about 18 kb in size, and consisted of seven exons and six introns [39]. There are many regulatory elements (AP-1, NF1-A1, RGPR-p117, β -catenin, and NF- κ B) in the 5'-flanking region [21, 40–48]. The promoter activity of the rat regucalcin gene is enhanced by treatment with Bay K 8644, dibutyryl cyclic AMP, phorbol esters, insulin, and dexamethasone [42]. Using gel mobility shift assays, it is found that nuclear proteins from rat liver cells and rat hepatoma H4-II-E cells specifically bind to the 5'-flanking region of the rat regucalcin gene [40–42]. Treatment with Bay K 8644, dibutyryl cyclic AMP, phorbol esters, and insulin stimulates the binding of nuclear

factors to the 5'-flanking region of the rat regucalcin gene in H4-II-E cells. These factor-inducible nuclear proteins are related to enhance promoter activity of the regucalcin gene [42].

Regucalcin mRNA expression has been demonstrated to mediate through signaling pathway of Ca^{2+} /calmodulin-dependent protein kinase, protein kinase C, and tyrosine kinase in the cells [49–51]. AP-1 factor binds to the 5'-flanking region of the rat regucalcin gene that is mediated through the Ca^{2+} response [41]. AP-1 factor is complex of c-fos/c-jun that is phosphorylated by protein kinases [52, 53]. Calcium signaling system is an important pathway in the stimulation of regucalcin mRNA expression.

Regucalcin mRNA is mainly present in liver and renal cortex with a size of 1.8 kb [24]. The expression of regucalcin mRNA in the liver and renal cortex is clearly stimulated through an increase in the cellular Ca^{2+} levels following an oral administration of calcium chloride in rats in vivo [54–56]. Hepatic regucalcin mRNA expression is increased with fetal development and its expression is stimulated after the intake of dietary calcium to maternal rats in vivo [57]. Liver regucalcin concentration is increased after an oral administration of calcium in rats [55].

Hepatic regucalcin mRNA expression is also stimulated after a single subcutaneous administration of calcitonin [58], insulin [59], and estrogen [60], suggesting that the expression of regucalcin mRNA is enhanced through various hormonal stimulation. Regucalcin mRNA expression is increased in regenerating rat liver, suggesting its role in the proliferation of liver cells [61]. Aging has been shown to decrease liver regucalcin mRNA expression [62].

Rat regucalcin immunoreactivity is most pronounced in the liver of rats; it is not seen in the duodenum, testicle, spleen, lung, and smooth muscle (bladder) and is barely visible in the kidney, heart, and brain [25, 26]. Regucalcin is primarily located in the rat liver. Thus, the tissue specific distribution of regucalcin is demonstrated by northern blotting analysis or enzyme immunoassay.

Role of regucalcin in intracellular Ca^{2+} homeostasis

Intracellular Ca^{2+} homeostasis is regulated through plasma membrane (Ca^{2+} - Mg^{2+})-adenosine 5'-triphosphatase (ATPase), microsomal Ca^{2+} -ATPase, mitochondrial Ca^{2+} uptake, and nuclear Ca^{2+} transport in the cells. Regucalcin has been demonstrated to regulate Ca^{2+} -transporting systems in the liver, renal cortex cells, heart, and brain tissues, suggesting its role in the regulation of intracellular Ca^{2+} homeostasis.

Role of regucalcin in Ca^{2+} homeostasis in liver cell

The high-affinity $(\text{Ca}^{2+}\text{-Mg}^{2+})\text{-ATPase}$ is located on the plasma membranes of liver cells [63, 64]. This enzyme acts as a Ca^{2+} pump to exclude the metal ion from the cytoplasm of liver cells. Addition of regucalcin into the reaction mixture *in vitro* caused an increase in $(\text{Ca}^{2+}\text{-Mg}^{2+})\text{-ATPase}$ activity in the plasma membranes isolated from rat liver, suggesting a role in the regulation of Ca^{2+} pump activity [65]. Regucalcin directly activates $(\text{Ca}^{2+}\text{-Mg}^{2+})\text{-ATPase}$ independently of Ca^{2+} -stimulated phosphorylation of the enzyme [65–67], and it has been shown to stimulate ATP-dependent calcium transport across the plasma membrane vesicles of rat liver after addition of $^{45}\text{Ca}^{2+}$ into the reaction mixture *in vitro* [68]. Regucalcin-enhanced ATP-dependent $^{45}\text{Ca}^{2+}$ uptake in the plasma membrane vesicles is completely inhibited in the presence of N-ethylmaleimide or digitonin [67]. Regucalcin has been shown to bind the lipid components of liver plasma membrane, and it acts on the sulfhydryl (SH) groups that are an active site of $(\text{Ca}^{2+}\text{-Mg}^{2+})\text{-ATPase}$ [67]. The mechanism of regucalcin in activating $(\text{Ca}^{2+}\text{-Mg}^{2+})\text{-ATPase}$ may be not involved on GTP-binding protein that modulates the receptor-mediated hormonal effect (including calcitonin, epinephrine, phenylephrine, and insulin) in liver plasma membranes [69].

The effect of hormonal signaling factors (inositol-glycan, dibutyryl cyclic AMP, and inositol 1,4,5-trisphosphate) on the regucalcin-increased $(\text{Ca}^{2+}\text{-Mg}^{2+})\text{-ATPase}$ activity in rat liver plasma membranes is examined [70]. Inositol-glycan, which is generated by insulin [70], can directly activate the plasma membrane $(\text{Ca}^{2+}\text{-Mg}^{2+})\text{-ATPase}$, and its effect is modulated by regucalcin. Cross talk with signaling factors may be seen in the regulation of Ca^{2+} pump activity in the plasma membranes of liver cells.

The physiological role of regucalcin in the regulation of $(\text{Ca}^{2+}\text{-Mg}^{2+})\text{-ATPase}$ activity in liver plasma membranes is examined after an oral administration of calcium chloride solution in rats [71]. Calcium administration caused an increase in $(\text{Ca}^{2+}\text{-Mg}^{2+})\text{-ATPase}$ activity in liver plasma membranes [71]. This increase was abolished in the presence of anti-regucalcin antibody, suggesting an involvement of endogenous regucalcin that is distributed in the cytoplasm. Regenerating rat liver with a proliferative cells significantly increased liver calcium content and plasma membrane $(\text{Ca}^{2+}\text{-Mg}^{2+})\text{-ATPase}$ activity between 12 and 48 h after partial hepatectomy [72]. This increase was completely abolished in the presence of anti-regucalcin antibody, indicating an involvement of endogenous regucalcin [72]. Activatory effect of regucalcin on hepatic plasma membrane $(\text{Ca}^{2+}\text{-Mg}^{2+})\text{-ATPase}$ was impaired in liver injury with carbon tetrachloride administration in rats

[73]. Regucalcin plays a role as an activator protein for Ca^{2+} pump enzyme in the hepatic plasma membranes.

Regucalcin binds Ca^{2+} in the cytoplasm of liver cells, and the metal is subsequently transported into the organelle dependent on ATP [74]. Regucalcin may not tightly bind cytosolic Ca^{2+} of lower levels, since its calcium-binding constant is $4.19 \times 10^5 \text{ M}^{-1}$ [8]. Regucalcin may regulate cytoplasmic Ca^{2+} levels by activating Ca^{2+} pump enzyme in the plasma membranes of liver cells.

Regucalcin can stimulate the uptake of Ca^{2+} by rat liver mitochondria [75, 76]. The effect of regucalcin on mitochondrial Ca^{2+} uptake is inhibited in the presence of ruthenium red or lanthanum chloride [75], which is an inhibitor of mitochondrial Ca^{2+} transport. Regucalcin may have a role in the reduction of cytoplasmic Ca^{2+} levels due to activating mitochondrial Ca^{2+} uptake.

Regucalcin has also been demonstrated to activate Ca^{2+} pump enzymes ($\text{Ca}^{2+}\text{-ATPase}$) and to stimulate ATP-dependent $^{45}\text{Ca}^{2+}$ uptake by liver microsomes [77, 78], suggesting a role in the regulation of cytoplasmic Ca^{2+} levels. The effect of regucalcin in increasing $\text{Ca}^{2+}\text{-ATPase}$ activity in the microsomes is inhibited in the presence of thapsigargin, a specific inhibitor of microsomal Ca^{2+} pump enzyme. Regucalcin acts on the SH groups of microsomal $\text{Ca}^{2+}\text{-ATPase}$ due to the binding on the membranous lipids [78].

The components of Ca^{2+} uptake ($\text{Ca}^{2+}\text{-ATPase}$) and Ca^{2+} release ($\text{Ca}^{2+}\text{-channels}$) are located at separate sites on liver microsomes. Interestingly, regucalcin has been found to stimulate Ca^{2+} release from rat liver microsomes [79]. The mechanism is related to the inositol 1,4,5-trisphosphate (IP3)-induced Ca^{2+} release [80]. Regucalcin may bind to IP3 receptors on the microsomes. This cell physiological significance of regucalcin is unknown. Presumably, regucalcin stimulates microsomal Ca^{2+} uptake when cytosolic Ca^{2+} concentration is raised. Also, regucalcin regulates Ca^{2+} storage in the endoplasmic reticulum of liver cells: it stimulates Ca^{2+} release from the microsomes to restore the microsomal calcium accumulation to regulate Ca^{2+} -related microsomal functions. Regucalcin has been shown to have the reversible effect on liver microsomal glucose-6-phosphatase activity increased by Ca^{2+} addition [11].

The existence of an ATP-stimulated Ca^{2+} -sequestration system is also found in liver nuclei, and it generates a net increase in nuclear matrix free Ca^{2+} concentration [81]. This system may play an important role in the regulation of intranuclear Ca^{2+} -dependent processes [82]. ATPase, which is stimulated by Ca^{2+} in the presence of Mg^{2+} , exists in the nuclei of rat liver, and the Ca^{2+} -stimulated ATPase activity is involved in the nuclear Ca^{2+} uptake [83]. Regucalcin increases $\text{Ca}^{2+}\text{-ATPase}$ activity in rat liver nuclei [84]. Regucalcin has also been shown to

stimulate Ca^{2+} release from liver nuclei [85]. Presumably, regucalcin has a role in the regulation of liver nuclear function through the effect on Ca^{2+} transporting system in the nuclei.

Regucalcin (SMP30) has been shown to lower intracellular Ca^{2+} levels by modulating plasma membrane Ca^{2+} -pumping activity in the cloned human hepatoma HepG2 cells that overexpress regucalcin [86].

Role of regucalcin in Ca^{2+} homeostasis in kidney cells

Regucalcin is largely present in the kidney cortex of rats [26]. Kidney plays a physiological role in the regulation of calcium homeostasis in blood through re-absorption of urinary calcium [87, 88]. Renal cortex cells play a role in the re-absorption of urinary calcium.

Regucalcin mRNA is expressed in the kidney cortex but not in the medulla of rats [56], and its expression is stimulated after calcium administration in vivo [56]. The binding of kidney nuclear proteins to the 5'-flanking region of the rat gene for regucalcin has been shown to be enhanced through Ca^{2+} /calmodulin signaling [89, 90]. Regucalcin mRNA expression is stimulated after the administration of dexamethasone in rats [91], and its expression is suppressed in hypertensive state in rats [92–94]. The specific nuclear factor binds to the NF1-like sequence in the promoter region of regucalcin gene in the kidney cortex of rats [90], and the nuclear factor binding and regucalcin mRNA expression are suppressed after administration of cisplatin that induces kidney damage [90, 95].

Ca^{2+} -ATPase system has been shown to exceed the capacity of the $\text{Na}^+/\text{Ca}^{2+}$ exchanger, and it plays a primary role in Ca^{2+} homeostasis of rat kidney cortex cells [96, 97]. Regucalcin has been demonstrated to play a role as an activator of the ATP-dependent Ca^{2+} pumps in the basolateral membranes isolated from rat kidney cortex [98]. The effect of regucalcin in increasing Ca^{2+} pump enzyme (Ca^{2+} -ATPase) activity in the basolateral membranes is completely inhibited in the presence of N-ethylmaleimide, indicating that regucalcin may act on the SH groups of Ca^{2+} -ATPase [98].

Regucalcin has also been shown to increase Ca^{2+} -ATPase activity and ATP-dependent calcium uptake in the microsomes of rat kidney cortex [99]. These increases are clearly decreased in the presence of N-ethylmaleimide, suggesting that regucalcin acts on the SH groups of Ca^{2+} -ATPase in the microsomes [97].

The finding, that regucalcin increases Ca^{2+} -ATPase activity in the basolateral membranes and microsomes of rat renal cortex, suggests a physiological role of regucalcin in the regulation of the Ca^{2+} homeostasis in renal cells. Regucalcin may be responsible for ATP-dependent transcellular Ca^{2+} transport, and it participates in the promotion

of Ca^{2+} re-absorption in the nephron tubule of kidney cortex. Regucalcin may play a physiological role in the regulation of calcium metabolism in blood through re-absorption of urinary calcium in kidney.

Role of regucalcin in Ca^{2+} homeostasis in heart cells

The Ca^{2+} current is one of the most important components in cardiac excitation–contraction coupling [100]. This coupling mechanism is based on the regulation of intracellular Ca^{2+} concentration by Ca^{2+} pump in the sarcoplasmic reticulum of heart muscle [100]. The role of regucalcin in the regulation of heart muscle function is shown. Regucalcin mRNA is expressed in rat heart [27]. The result with western blot analysis indicates that regucalcin is present in the cytoplasm of heart muscle cells [27]. Regucalcin concentration in the heart muscle tissues has been shown to be about 3.86×10^{-8} M [26]. Regucalcin mRNA expression in the hearts of rats is decreased with increasing age [101], and free radical stress has a suppressive effect on its gene expression [101]. Overexpression of regucalcin in transgenic rats has been found to accelerate free radical stress-induced death of rats [101].

Regucalcin has been found to increase Ca^{2+} -ATPase activity and ATP-dependent Ca^{2+} uptake, which regulates intracellular Ca^{2+} concentration related to cardiac excitation–contraction coupling, in rat heart microsomes, suggesting its role in the regulation of heart muscle function [27]. The effect of regucalcin in increasing heart microsomal Ca^{2+} -ATPase activity is inhibited in the presence of thapsigargin [27], a specific inhibitor of the sarcoplasmic reticulum Ca^{2+} pump enzyme (Ca^{2+} -ATPase) [102], indicating that regucalcin activates Ca^{2+} pump enzyme in the sarcoplasmic reticulum. It is suggested that regucalcin binds to the lipids at the close site of Ca^{2+} -ATPase in heart microsomes, and that it acts on the SH group which may be an active site of the enzyme and stimulates Ca^{2+} -dependent phosphorylation of Ca^{2+} -ATPase [27].

Phospholamban has been known to inhibit Ca^{2+} pump enzyme (Ca^{2+} -ATPase) in the sarcoplasmic reticulum (microsomes) of heart muscle [103]. Ca^{2+} -ATPase is activated through cAMP-dependent phosphorylation of phospholamban following hormonal stimulation [103]. The endogenous activatory protein of sarcoplasmic reticulum Ca^{2+} -ATPase is unknown. Regucalcin, which is present in the cytoplasm of heart muscle, may play an important role as an endogenous activator in the regulation of sarcoplasmic reticulum Ca^{2+} -ATPase activity in rat heart muscle [100]. Regucalcin may play a physiological role in the regulation of cardiac excitation–contraction coupling.

The role of regucalcin in the regulation of Ca^{2+} -ATPase activity in the heart mitochondria of rats is examined, moreover [104]. Regucalcin is found to be present in the

mitochondria of normal rat heart [104], and this localization is increased in the heart of regucalcin transgenic rats as compared with that of normal rats [104]. The addition of regucalcin (10^{-11} to 10^{-8} M) in the enzyme reaction mixture caused a significant increase in Ca^{2+} -ATPase activity in the heart mitochondria in the presence of $50 \mu\text{M}$ CaCl_2 [104]. Ca^{2+} -ATPase activity was also increased in the heart mitochondria of regucalcin transgenic rats [104]. Regucalcin has an activating effect on Ca^{2+} -ATPase in rat heart mitochondria, suggesting its role in the regulation of heart mitochondrial function.

Regucalcin may play a role in the regulation of cytoplasmic Ca^{2+} levels due to activating Ca^{2+} pump activity in the sarcoplasmic reticulum and mitochondria in heart cells.

Role of regucalcin in Ca^{2+} homeostasis in brain tissues

Intracellular Ca^{2+} concentration in the neuronal cells of brain is regulated by various buffering and transport systems such as the membrane Na^+ - Ca^{2+} exchanges, the membranous Ca^{2+} -ATPase, Ca^{2+} -binding proteins, and intracellular Ca^{2+} uptake systems [105–109]. The changes in the neuronal Ca^{2+} homeostasis with aging may be implicated in age-related disturbance in cognitive functions [109]. There is growing evidence that the alteration in the neuronal Ca^{2+} regulation is also implicated in the pathology of Alzheimer's disease [109].

The expression of regucalcin mRNA is demonstrated in brain tissues [28, 110]. Regucalcin concentration in the brain tissues has been shown to be about 5×10^{-9} M as measured using enzyme-linked immunoadsorbent assay, and this level is lowered with aging [26]. Regucalcin is localized in the neurons isolated from rat brain tissues [28], suggesting its role in brain function.

Brain calcium accumulation has been shown to increase after oral administration of calcium in rats, and fasting enhances its accumulation [111]. The supply of glucose may be required in the regulation of brain Ca^{2+} homeostasis in rats in vivo [111], suggesting a physiological significance of energy-dependent mechanism in brain calcium metabolism. Fasting caused a significant increase in brain calcium content and Ca^{2+} -ATPase activity in the microsomes and mitochondria of brain tissues in young and aged rats, and these increases were restored after the supply of glucose, supporting a physiologic significance of energy-dependent mechanism in the regulation of brain calcium in rats with different ages [112].

Aging causes a decrease in Ca^{2+} -ATPase activity in the brain plasma membranes of rats [113], suggesting that aging enhances the entry of Ca^{2+} into brain neuronal cells across the plasma membranes. In addition, aging induces an attenuation of Ca^{2+} -sequestering system in the brain microsomes, supporting the view that a disturbance of the

neuronal Ca^{2+} regulation is brought with increasing age [114]. Protein kinase C activates brain microsomal Ca^{2+} -ATPase in aged rats [115]. It is speculated that aging-induced increase in Ca^{2+} -ATPase activity results from the translocation to the microsomes of protein kinase C in brain cytosol [115]. The development of brain disease with aging may be partly related to the toxicity of brain calcium raised by the increase in microsomal Ca^{2+} -ATPase activity with aging. The disturbance of brain Ca^{2+} homeostasis may play a pivotal role in the revelation of brain disease.

Regucalcin has been found to have an inhibitory effect on Ca^{2+} -ATPase activity in rat brain microsomes [110], suggesting that regucalcin plays a role in the regulation of microsomal Ca^{2+} -ATPase activity in rat brain. Interestingly, the concentration of regucalcin in the cerebral cortex and hippocampus of brain tissues is decreased with aging [110]. The suppressive effect of regucalcin on brain microsomal Ca^{2+} -ATPase activity was weakened in aged rats [110]. Presumably, the aging-induced elevation of brain microsomal Ca^{2+} -ATPase activity is partly resulted from an attenuation of regucalcin action on the enzyme activity with aging. There may be a possibility that the translocation of protein kinase C to the brain microsomes of aged rats [115] is a cause of the attenuation of regucalcin effect on the enzyme activity. Regucalcin may play a physiological and pathophysiological role in the regulation of intracellular Ca^{2+} concentration in the brain tissues.

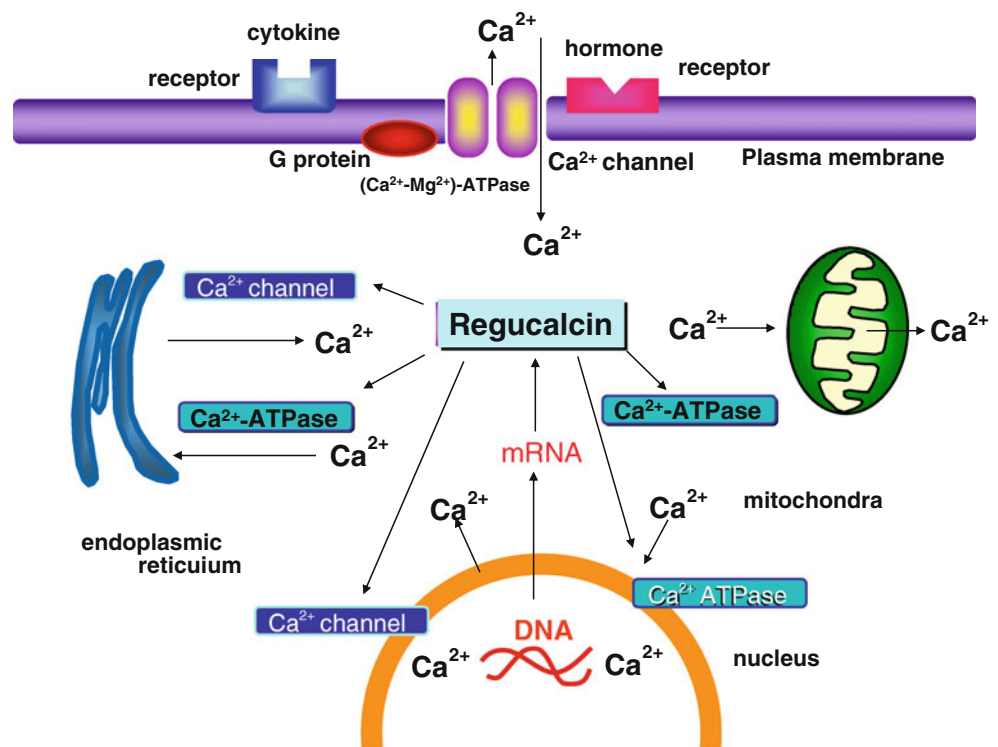
Regucalcin has an activatory role in the regulation of Ca^{2+} -ATPase activity in the mitochondria of brain tissues of rats [116]. The addition of regucalcin (10^{-10} to 10^8 M), which is a physiological concentration in rat brain tissues, into the enzyme reaction mixture containing $25 \mu\text{M}$ calcium chloride caused a significant increase in Ca^{2+} -ATPase activity, while it did not significantly change in Mg^{2+} -ATPase activity [116].

Regucalcin levels are increased in the brain tissues or the mitochondria obtained from regucalcin transgenic rats. The mitochondrial Ca^{2+} -ATPase activity has been found to increase in regucalcin transgenic rats as compared with that of wild-type rats [116]. Endogenous regucalcin plays a role in the regulation of Ca^{2+} -ATPase activity in the brain mitochondria of rats.

Conclusion

Regucalcin plays a cell physiological role as a regulatory protein that is involved in the regulation of Ca^{2+} homeostasis in various cell types. The low cytoplasmic Ca^{2+} concentration of living cells is maintained through energy-requiring pumps. These pumps either remove Ca^{2+} to the extracellular space by transport across the plasma membrane or accumulate it inside of intracellular organelles such as the mitochondria and endoplasmic reticulum

Fig. 1 Regucalcin has a pivotal role in keeping intracellular Ca^{2+} homeostasis that is attenuated with various stimulating in cells. Regucalcin increases plasma membrane $(\text{Ca}^{2+}-\text{Mg}^{2+})$ -ATPase, mitochondrial Ca^{2+} -ATPase and microsomal Ca^{2+} -ATPase activities in cells. Regucalcin also stimulates Ca^{2+} release from the microsomes (endoplasmic reticulum). Regucalcin has an inhibitory effect on nuclear Ca^{2+} -ATPase and a stimulatory effect on Ca^{2+} release from the nucleus. Through thus mechanism, regucalcin plays a part in regulating the rise of cytosolic Ca^{2+} concentration and nuclear matrix Ca^{2+} levels in cells that suppresses Ca^{2+} -dependent cellular events



(microsomes). Regucalcin stimulates the activity of these pumps to lower the cytoplasmic Ca^{2+} levels, as shown in Fig. 1. This may provide the cellular mechanism of the inhibitory effect of regucalcin in the regulation of cell functions related to Ca^{2+} signaling.

Regucalcin reverses Ca^{2+} effect in enzyme regulation

Ca^{2+} and calmodulin systems generally activate various enzymes in many cells. There are many evidences that regucalcin has an inhibitory effect on enzyme activation by Ca^{2+} /calmodulin [24–28]. This section outlines the findings on the inhibitory effect of regucalcin on action of Ca^{2+} in cell metabolism.

Liver metabolism is regulated through an increase in Ca^{2+} level in the cytoplasm of liver cells due to hormonal stimulation [3–5]. Regucalcin has an inhibitory effect on Ca^{2+} /calmodulin-dependent enzyme activity in vitro. The hormonal effect on fructose-1,6-diphosphatase, which promotes the conversion from fructose-1,6-diphosphate to glucose-6-phosphate in the hepatic cytoplasm of rats, is mediated through Ca^{2+} . This enzyme activity is activated through Ca^{2+} /calmodulin [10]. The activation of fructose-1,6-diphosphatase by Ca^{2+} /calmodulin was completely reversed after the addition of regucalcin in the enzyme reaction mixture [10].

Phosphorylase *a* activity in the liver particulate glycogen is increased after addition of Ca^{2+} (10 μM) [13]. This

increase was completely reversed after addition of regucalcin in the enzyme reaction mixture [13]. Regucalcin (1.0 μM) reversed activations of pyruvate kinase [12] and glucose-6-phosphatase [11] after addition of Ca^{2+} in the enzyme reaction mixture. These findings suggest that regucalcin regulates glycogenolysis and gluconeogenesis that is stimulated by Ca^{2+} in liver cells.

The reversible effect of regucalcin is also shown in Ca^{2+} -induced inhibition of 5'-nucleotidase activity in liver plasma membranes [15] and deoxyuridine 5'-triphosphatase activity in hepatic cytosol [117].

Thus, regucalcin has a reversible effect on the activation and inhibition of many enzymes by Ca^{2+} .

The controlled use of energy to maintain homeostasis and cellular function is a basic property of all cells. The free energy of ATP is believed to be the major cytosolic intermediate in this process. ATP is produced from the energy obtained by the oxidation of metabolic substrates in glycolysis and oxidative phosphorylation. ATPase produces the energy from ATP in the cell cytosol. The cytosolic factors regulating ATPase activity is important. Regucalcin has been shown to play an inhibitory role in the regulation of ATPase activity in the brain cytosol of young and aged rats [118], suggesting a role in the regulation of energy conversion in brain tissues.

The mechanism of the reversible effect of regucalcin on various enzyme activities, which are regulated through Ca^{2+} , has not been well known. However, action of regucalcin may be partly based on Ca^{2+} binding, because the

protein has 6–7 high-affinity binding sites per molecule ($K_f = 4.19 \times 10^5 \text{ M}^{-1}$) [8]. Regucalcin may directly bind to Ca^{2+} and/or calmodulin. In addition, it is possible that regucalcin may bind to enzymes and that affects enzyme activity.

Regucalcin inhibits Ca^{2+} /calmodulin-dependent enzyme activation

Ca^{2+} /calmodulin-dependent enzymes are localized in many tissues and cells. Regucalcin has been found to have an inhibitory effect on various Ca^{2+} /calmodulin-dependent enzyme activations.

Cyclic adenosine monophosphate (AMP) is a second messenger for hormonal stimulation in many cells. Cyclic AMP is degraded by cyclic AMP phosphodiesterase in liver cytosol [119]. The enzyme activity is increased through Ca^{2+} /calmodulin [1]. Regucalcin has been found to inhibit the activation of cyclic AMP phosphodiesterase by Ca^{2+} /calmodulin in the cytosols of liver and renal cortex [120, 121], suggesting a role of regucalcin in the regulation of cyclic AMP level in the cells.

Nitric oxide (NO) may be important as a signaling factor in many cells [122]. NO, which has an unpaired electron reacts with protein, targets primarily through their thiol or heme groups, and acts as a messenger or modulator molecule in many biological systems. NO is produced from L-arginine with L-citrulline as a coproduct in a reaction catalyzed by NO synthase that requires Ca^{2+} /calmodulin [122].

Regucalcin has been shown to have a suppressive effect in the enhancement of NO synthase activity in the cytosols of liver [123, 124] and kidney cortex [125] of rats. Over-expression of regucalcin did not cause a significant alteration of NO synthase activity in the kidney cortex cytosol of regucalcin transgenic rats as compared with that of wild-type rats [125]. However, the effect of calcium chloride (10 μM) in increasing NO synthase activity in the kidney cortex cytosol of wild-type rats was weakened in regucalcin transgenic rats [125]. The presence of anti-regucalcin monoclonal antibody (25 or 50 ng/ml) in the reaction mixture caused a significant increase in NO synthase activity, and this increase was completely abolished after the addition of regucalcin (10^{-7} M). Endogenous regucalcin has a suppressive effect on NO synthetase activity in the cytosol of various tissues of rats.

Regucalcin has also been found to suppress Ca^{2+} /calmodulin-dependent NO synthase activity in the heart cytosol of rats [126]. Regucalcin had an inhibitory effect on NO synthase activity in the presence of antagonist for calmodulin [126], indicating a direct effect of regucalcin on the enzyme independent of Ca^{2+} /calmodulin. The physiological significance of regucalcin inhibition of NO

synthase in heart muscle cytosol is unknown. However, regucalcin may participate in the regulation of NO production in heart muscle cells. NO acts as a messenger or modulator molecule in heart muscle. NO production may be stimulated through Ca^{2+} signaling due to hormonal stimulation in heart muscle cells. Regucalcin may have a suppressive effect on over-production of NO due to inhibiting NO synthase in heart muscle cells.

Nitric oxide (NO) acts as a messenger or modulator molecule in brain neurons. Regucalcin has also been shown to reveal a suppressive effect on NO synthase activity in the brain cytosol of young and aged rats, even though regucalcin levels are reduced with increasing age [127]. A remarkable expression of regucalcin protein was seen in the cytosol and nucleus of the brain tissues of regucalcin transgenic rats as compared with that of wild-type rats [128]. NO synthetase activity was decreased in the brain cytosol of transgenic rats, and the presence of anti-regucalcin monoclonal antibody (50 ng/ml) in the enzyme reaction mixture caused a significant increase in cytosolic NO synthase activity in the cytosol of brain tissues of wild-type rats [128]. Endogenous regucalcin plays a suppressive role in the regulation of brain neuronal NO synthase activity in rats.

Thus, regucalcin has been demonstrated to have a suppressive role on Ca^{2+} /calmodulin-dependent NO synthase activity in various tissues. Regucalcin may play a role as a suppressor protein in NO production in many cell types, and it may regulate many cellular events that are involved in NO signaling.

Superoxide dismutase (SOD) plays a role in the prevention of cell death and apoptosis in the heart. The decrease in Mn-SOD activity is associated with increased mitochondrial oxidative damage as demonstrated by a decrease in the activities of iron sulfhydryl proteins sensitive to oxygen stress [129]. Cu/Zn-SOD has been shown to play the role of protector against doxorubicin-induced cardiotoxicity in mice [130]. Meanwhile, NO has a role in the suppression of myocardial O_2 consumption in rats [131].

Regucalcin has been found to increase SOD activity in the cytosol of rat liver [132] and heart [133]. Regucalcin has an inhibitory effect on NO synthase activity in the heart cytosol [133]. Production of superoxide radicals is widely accepted as the cause of the cardiodamage. Presumably, regucalcin participates in the control of production of superoxide radicals in rat heart muscle cells.

The multifunctional Ca^{2+} /calmodulin-dependent protein kinases play an important role in the response of the cells to a calcium signal [1, 134]. Regucalcin has been shown to inhibit Ca^{2+} /calmodulin-dependent protein kinase activity in the cytosol of rat liver [135], kidney cortex [136], and brain tissues of rats [137, 138]. An appreciable effect of

regucalcin is seen at 0.5 μM , which is a cell physiological concentration. As Ca^{2+} /calmodulin-dependent protein kinase in the cytoplasm is activated through calcium signal, regucalcin may regulate a signal transduction for Ca^{2+} . The mechanism of action of regucalcin may be partly based on its binding to Ca^{2+} /calmodulin and/or enzyme. Regucalcin has been demonstrated to bind on calmodulin in analysis with sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) using calmodulin-agarose beads [139].

Nishizuka [2] discovered a diacylglycerol-activated Ca^{2+} and phospholipid-dependent protein kinase (protein kinase C). Protein kinase C is distributed widespread in the body, with amounts in the liver being intermediate between the high levels found in brain and spleen [2]. Protein kinase C is capable of phosphorylating cytoplasmic proteins. It is found that regucalcin inhibits protein kinase C activity in the cytoplasm of rat liver [140], kidney cortex [141], and brain cytosol and brain neurons [137, 138], supporting the view that regucalcin plays a role in the regulation of Ca^{2+} -dependent cellular functions. The presence of anti-regucalcin monoclonal antibody in the enzyme reaction mixture caused a significant elevation of protein kinase activity, indicating that the endogenous regucalcin has an inhibitory effect on the enzyme activity [137, 138].

The regulatory effect of regucalcin in rat brain function may be attenuated with aging. Increasing age enhances protein kinase activity in rat brain cytosol [137]. This enhancement may be partly involved in aging-decreased regucalcin in rat brain tissues [110]. It is speculated that the endogenous regucalcin plays a suppressive role in the activation of Ca^{2+} -dependent protein kinase in the brain cytosol, and that aging may weaken the effect of regucalcin. Regucalcin may play a pivotal role in the regulation of phosphorylation of the cytosolic proteins in brain tissues. Interestingly, it has been reported that regucalcin gene is localized on human chromosome X that encompasses the map location for a growing number of diseases with a genetic basis; these include syndromic and non-syndromic forms of X-linked mental retardation and X-linked neuromuscular diseases [23]. Regucalcin may have a pathophysiological role in brain disease with aging.

As mentioned above, regucalcin plays an inhibitory role in signaling pathway that is mediated through cyclic AMP, NO, and Ca^{2+} -dependent protein kinases in many tissues and cell types.

Protein phosphorylation–dephosphorylation is a universal mechanism by which numerous cellular events are regulated [142]. It has become apparent that there may exist many phosphatases that, like the kinases, are just elaborately and rigorously controlled [142, 143]. Protein phosphatase plays an important role in intracellular signal transduction due to hormonal stimulation [142].

Calcineurin, a calmodulin-binding protein, has been shown to possess a Ca^{2+} -dependent and calmodulin-stimulated protein phosphatase activity [144]. Regucalcin has been demonstrated to inhibit calcineurin activity in the cytosol of liver and renal cortex after its binding to calmodulin [145, 146]. Protein phosphatases, which endogenous regucalcin acts in liver cytoplasm, may be insensitive to okadaic acid [147]. Protein phosphatase activity toward phosphotyrosine, phosphoserine, and phosphothreonine in the cytosol of rat liver was elevated in the presence of anti-regucalcin monoclonal antibody in the enzyme reaction mixture *in vitro*, suggesting its role of endogenous regucalcin [145]. Regucalcin may be a unique protein, which has inhibitory effects on protein tyrosine phosphatase and protein serine/threonine phosphatase. Regucalcin, which was localized in rat liver nuclei, has been shown to inhibit nuclear protein phosphatase activity [148].

Endogenous regucalcin plays a role in the regulation of protein phosphatase activity in the cytosol and nuclei of rat renal cortex [149–151]. Regucalcin has been found to be present in the cytosol and nuclei of rat kidney cortex using western blot analysis [151]. The addition of regucalcin (50–250 nM) in the enzyme reaction mixture obtained from the cytoplasm and nuclei from rat kidney cortex caused a decrease in protein phosphatase activity toward phosphotyrosin, phosphoserine, and phosphothreonine [149, 150]. The effect of calcium (25 μM) and calmodulin (2.5 $\mu\text{g/ml}$) in increasing protein phosphatase activity was decreased after the addition of regucalcin. Protein phosphatase activity in the cytosol and nuclei was increased in the presence of anti-regucalcin monoclonal antibody (10–50 ng/ml) in the enzyme reaction mixture [149, 150]. Regucalcin plays a suppressive role in the regulation of protein phosphatase activity in the cytoplasm and nucleus of rat kidney cortex.

Kidney cortex calcium content and the cytosolic and nuclear regucalcin levels were increased at 0.5–5 h after a single intraperitoneal administration of calcium chloride solution (10 mg Ca/100 g body weight) in rats [151]. The cytosolic and nuclear protein phosphatase activity, which is raised in calcium-administered rats, was found to enhance when anti-regucalcin monoclonal antibody was added in the enzyme reaction mixture [151]. The effect of antibody was completely abolished after the addition of regucalcin in the enzyme reaction mixture. Thus, endogenous regucalcin has a suppressive effect on the enhancement of protein phosphatase activity in the cytosol and nucleus of kidney cortex in calcium-administered rats.

Cardiac hypertrophy is induced by calcineurin, which dephosphorylates the transcription factor NF-A3, enabling it to translocate to the nucleus [152]. Transgenic mice, that express activated forms of calcineurin or NF-AT3 in the heart, develops cardiac hypertrophy and heart failure that

mimic human heart disease [152], suggesting a hypertrophic signaling pathway. If regucalcin has a suppressive effect on calcineurin activity in the heart cytosol of normal and transgenic rats [153], overexpression of regucalcin may have a pathophysiological role in the prevention of development of cardiac hypertrophy and heart failure.

Regucalcin has also been shown to have an inhibitory effect on Ca^{2+} /calmodulin-dependent protein phosphatase activity toward phosphotyrosine, phosphoserine, and phosphothreonine in rat brain cytosol [154] and neurons [28]. The presence of anti-regucalcin monoclonal antibody in the enzyme reaction mixture caused a significant elevation of protein phosphatase activity in the brain cytosol, indicating that the endogenous regucalcin has a suppressive effect on the cytosolic enzyme activity [154]. Regucalcin has been shown to have an inhibitory role in the regulation of protein phosphatase activity in rat brain cytosol.

Regucalcin is localized in the microsomes of rat brain, and aging causes a decrease in its protein levels [155]. Regucalcin has been also shown to have a suppressive effect on protein tyrosine phosphatase activity in rat brain microsomes [155]. Aging caused an increase in protein tyrosine phosphatase activity in rat brain microsomes and the suppressive effect of regucalcin on the enzyme activity was weakened in aged rats [155], suggesting that the decrease in microsomal regucalcin with aging is partly involved in the enhancement of microsomal protein tyrosine phosphatase activity with aging [155].

Regucalcin has been found to be present in the nucleus of rat brain and the endogenous regucalcin has a suppressive effect on the nuclear protein tyrosine phosphatase activity [156]. Increasing age has also been found to induce a reduction in rat brain nucleus and may lead to attenuation of the suppressive effect of regucalcin on the nuclear protein tyrosine phosphatase activity [156]. Regucalcin may play a pivotal role as a regulatory protein in the regulation of brain function that relates to protein phosphorylation–dephosphorylation.

Thus, regucalcin may play a physiological role in the intracellular control of the hormonal stimulation for phosphorylation and dephosphorylation of many proteins in various cell types.

As mentioned above, regucalcin has been demonstrated to reverse the activity of many Ca^{2+} -activated enzymes (phosphorylase *a*, glucose-6-phosphatase, fructose-1,6-bisphosphatase, pyruvate kinase, protein kinase C, Ca^{2+} /calmodulin-dependent protein kinase, protein phosphatase, and Ca^{2+} /calmodulin-dependent cyclic AMP phosphodiesterase) and of Ca^{2+} -inhibited enzymes (5'-nucleotidase and dUTPase).

The first action is that regucalcin binds Ca^{2+} and that inhibits the metal's effect on many enzymes. The effect of

regucalcin, that reverses Ca^{2+} action on many enzymes, may be based on its binding of Ca^{2+} , since the protein has 6–7 high-affinity binding sites per molecule, and a Ca^{2+} -binding constant of $4.19 \times 10^5 \text{ M}^{-1}$ [8]. The intrinsic significance of regucalcin action may be the binding of Ca^{2+} by its protein.

The second is that Ca^{2+} -binding regucalcin directly inhibits the function of enzymes and somewhat stimulates enzyme function. The direct action of Ca^{2+} -binding regucalcin or the protein itself may be decided by the protein structure of enzymes. Spectroscopical studies have clearly demonstrated that Ca^{2+} -binding induces conformational changes in regucalcin, which may then result in increased hydrophobicity of the protein, and loosening of the conformation of regucalcin [9].

Regucalcin can directly inhibit the enzymes that are activated through Ca^{2+} -calmodulin [10]; this also results from regucalcin that affects the binding of Ca^{2+} to calmodulin. Which of the two proteins binds Ca^{2+} may be decided by their relative concentrations of Ca^{2+} in hepatic cytosol, since the Ca^{2+} -binding constant of regucalcin is greater than that of calmodulin [8]. Calmodulin exists as a monomer of molecular weight 17,000 and contains four Ca^{2+} -binding sites [1]. In the enzyme assay system of Ca^{2+} /calmodulin-dependent cyclic AMP phosphodiesterase activity, the inhibitory effect of regucalcin on the enzyme activation through Ca^{2+} -calmodulin is completely blocked with the addition with increasing concentrations of Ca^{2+} [117, 118]. This further supports the view that the mechanism by which regucalcin inhibits Ca^{2+} action is based on the binding of Ca^{2+} .

Moreover, in the enzyme assay system of protein kinases (protein kinase C and Ca^{2+} /calmodulin-dependent protein kinase), the effect of regucalcin, which inhibits the activation of enzymes by Ca^{2+} , is seen with increasing concentrations of Ca^{2+} . This suggests that regucalcin directly inhibits the enzyme activation in addition to Ca^{2+} -binding. Also, it is possible that Ca^{2+} -binding regucalcin or the protein itself can directly inhibit the enzyme activity.

There may be many enzymes that are regulated by regucalcin and/or Ca^{2+} -binding regucalcin in various cell types. This remains to be elucidated.

Regucalcin regulates protein synthesis and degradation

Regucalcin has been shown to have a regulatory effect on protein synthesis and protein degradation, suggesting that regucalcin plays a role in the regulation of protein turnover in cells.

Role of regucalcin in protein synthesis

Protein synthesis is depressed in a variety of eukaryotic cell types exposed to conditions depleting Ca^{2+} but not Mg^{2+} [157]. It has been also proposed that hormones (vasopressin and α -adrenergic agonist), which are known to mobilize sequestered Ca^{2+} within liver cells, inhibit amino acid incorporation by influencing a Ca^{2+} requirement associated with protein synthesis [158]. Moreover, vasopressin inhibits the rate of protein synthesis in isolated hepatocytes partially depleted of Ca^{2+} [159]. These investigations propose the hypothesis that a sequestered pool of intracellular Ca^{2+} is required for the maintenance of high rates of protein synthesis in liver cells [158, 159]. On the other hand, vasopressin and α -adrenergic agonist cause an increase in the intracellular free Ca^{2+} concentration of hepatocytes that are not depleted of Ca^{2+} [160, 161]. Whether the increase in intracellular Ca^{2+} influences hepatic protein synthesis is well undefined, however, it may be important to clarify the effect of Ca^{2+} addition on hepatic protein synthesis *in vitro*.

It has been demonstrated that Ca^{2+} , of various metals, can uniquely inhibit *in vitro* protein synthesis using the 5,500g supernatant fraction (the microsomes and cytosol) of rat liver homogenate [162]. Its inhibition was seen after addition of 1.0 μM Ca^{2+} . Ca^{2+} addition caused a remarkable decrease in the activity of aminoacyl (leucyl)-tRNA synthetase, which is a rate-limiting enzyme of protein synthesis at translational process, in hepatic cytosol [162]. Ca^{2+} may directly inhibit hepatic protein synthesis in subcellular fraction of liver cells. Ca^{2+} is required for protein synthesis in hepatocytes exposed to conditions depleting the cation [162]. It is not clarified whether protein synthesis in hepatocytes not depleted of Ca^{2+} requires exogenous Ca^{2+} . The mechanism by which Ca^{2+} is required in protein synthesis of hepatocytes depleted of Ca^{2+} may be complex.

Calmodulin, which can amplify Ca^{2+} effects on enzymes [1], did not have an appreciable effect on *in vitro* protein synthesis using the 5,500g supernatant fraction of liver homogenate in the presence of Ca^{2+} (10 μM) [162]. The activity of aminoacyl-tRNA synthetase in hepatic cytosol was not altered through calmodulin [163]. Presumably, the protein synthesis is inhibited by Ca^{2+} , which is not bound to calmodulin.

The role of regucalcin in the regulation of *in vitro* protein synthesis using the 5,500g supernatant fraction of rat liver homogenate is investigated [162]. Regucalcin caused a remarkable inhibition of hepatic protein synthesis *in vitro* [162]. Regucalcin could not reverse the Ca^{2+} -induced inhibition of protein synthesis [162], although it has been shown that regucalcin can reverse the Ca^{2+} effect on many enzymes in liver cells. Since regucalcin can bind

to liver cytosolic proteins and the binding is slightly enhanced with the coexistence of 0.1 μM Ca^{2+} [65], Ca^{2+} -binding regucalcin and/or Ca^{2+} free regucalcin may be able to inhibit hepatic protein synthesis. In fact, the presence of regucalcin (1 and 2 μM) could fairly decrease hepatic protein synthesis that was reduced after the addition of 10 μM Ca^{2+} [162]. Regucalcin itself may play a role in the regulation of protein synthesis in liver cells.

Regucalcin has been shown to inhibit hepatic aminoacyl-tRNA synthase activity [162]. The inhibitory effect of regucalcin was seen in the presence of Ca^{2+} (10 μM). The inhibitory effect of regucalcin on hepatic protein synthesis may be partly based on a remarkable decrease of aminoacyl-tRNA synthetase activity caused by regucalcin. Regucalcin may bind aminoacyl (leucyl)-tRNA synthetase in hepatic cytosol, since iodinated regucalcin can bind the proteins in hepatic cytosol [65]. Regucalcin may be able to regulate liver cell function that is not affected by cellular Ca^{2+} .

The role of endogenous regucalcin on protein synthesis is examined using anti-regucalcin monoclonal antibody, moreover [163]. The presence of anti-regucalcin monoclonal antibody in the reaction mixture caused a significant increase in protein synthesis and [^3H] leucyl-tRNA synthetase activity in normal rat liver. These increases were completely prevented in the addition of exogenous regucalcin (1.0 μM). Liver cytosol contained about 16 μg of regucalcin per 1 mg of the cytosolic protein; the reaction mixture contained about 0.17–0.19 μM of endogenous regucalcin, because the cytosolic protein in the range 360–390 μg was added into the mixture of 1.0 ml. Endogenous regucalcin may have a suppressive effect on protein synthesis in liver cells.

Hepatic protein synthesis has been shown to enhance regeneration of rat liver, which induces a proliferation of liver cells after partial hepatectomy [163]. This enhancement was remarkable at 24 and 48 h after partial hepatectomy. Hepatic protein synthesis in regenerating liver was further enhanced in the presence of anti-regucalcin monoclonal antibody in the reaction mixture. Endogenous regucalcin has a suppressive role on the enhancement of protein synthesis in regenerating liver.

Role of regucalcin in protein degradation

Evidence for the role of Ca^{2+} -activated protease (calpains) is implicated in signal transduction [164]. Two neutral Ca^{2+} -requiring proteinases, differing in molecular size, have been isolated from rabbit liver cytosol [165]. Both are recovered as inactive proenzymes that can be converted to the active forms by high (0.1–1.0 mM) concentrations of Ca^{2+} in the absence of substrate or, in the presence of a protein substrate, by low (1–5 μM) concentrations of Ca^{2+}

[165]. The activated proteinases required only 1–5 μM Ca^{2+} for maximal activity [165].

The addition of regucalcin (0.25–2.0 μM) into the enzyme reaction mixture has been found to induce a remarkable increase of neutral proteinase activity in the presence of 5.0 μM Ca^{2+} [166]. The effect of regucalcin was seen at 0.25 μM . Regucalcin may activate Ca^{2+} -requiring proteinase in rat liver cytosol. The effect of regucalcin in increasing liver cytosolic proteinase activity is also seen in the absence of Ca^{2+} [166]. This increase was remarkable as compared with that of Ca^{2+} addition. Regucalcin may activate Ca^{2+} -not requiring neutral proteinase in rat liver cytosol. Regucalcin has a reversible effect on the activation of various enzymes by Ca^{2+} and/or calmodulin [120, 121, 147]. The finding, that regucalcin can activate neutral proteinase in rat liver cytosol in the presence or absence of Ca^{2+} , was novel.

The activatory effect of regucalcin on liver cytosolic proteinase activity was not seen in the presence of anti-regucalcin antiserum in the enzyme reaction mixture [166], suggesting a role of endogenous regucalcin in the activation of cytosolic proteinase.

The activatory effect of regucalcin on neutral proteases in the liver cytosol is characterized [167]. Leupeptin is a potent inhibitor of SH-proteinase. The regucalcin-increased proteinase activity was inhibited in the presence of leupeptin in the enzyme reaction mixture [167]. Regucalcin may activate neutral cysteinyl-proteinase in the liver cytosol. The effect of regucalcin in increasing proteolytic activity in rat liver cytosol was not abolished in the presence of diisopropylfluorophosphate (DFP), an inhibitor of serine-protease, although DFP alone had an inhibitory effect on the proteolytic activity [167]. Regucalcin does not act on serine-proteases in liver cytosol. The effect of regucalcin was also seen in the presence of a chelator of metal ions, suggesting that regucalcin does not activate metal-related proteases in liver cytosol.

The proteolytic activity in liver cytosol was markedly increased after the addition of dithiothreitol (DTT), a protecting reagent for SH group, in the enzyme reaction mixture and this increase was completely inhibited in the presence of N-ethylmaleimide (NEM), a SH group-modifying agent [167]. The effect of regucalcin in increasing proteolytic activity was seen in the presence of DTT in liver cytosol, although it was completely inhibited in the presence of NEM [167]. Regucalcin may act on the SH group of cysteinyl-proteases in liver cytosol.

Two forms of Ca^{2+} -activated neutral proteases (calpains) have been identified in hepatocytes and other cells, *m*-calpain and *μ* -calpain [165]. Isolated *μ* -calpain requires micromolar concentrations of Ca^{2+} for activation, while *m*-calpain requires millimolar concentrations [168]. The activation of proteases in rabbit liver cytosol required only

1–5 μM Ca^{2+} for maximal activity [167]. Ca^{2+} (10 μM)-increased proteolytic activity in rat liver cytosol was inhibited in the presence of NEM, indicating that neutral proteases (including calpains), which are activated by Ca^{2+} , exist in liver cytoplasm [167].

The activity of calpains, which are thiol protease [169], increases in hepatocytes following addition of ATP [170]. The proteolytic activity in liver cytosol was decreased after the addition of calpastatin, a specific inhibitor of calpains, indicating the existence of calpains in the cytosol [167]. Regucalcin-increased proteolytic activity was abolished in the presence of calpastatin. Regucalcin may be an activator for calpains. *m*-Calpain isolated from rabbit skeletal muscle was activated by regucalcin independent on Ca^{2+} . This activation was inhibited after the addition of NEM [167]. Regucalcin acts on the SH group in *m*-calpain. Presumably, regucalcin may be able to activate both *m*- and *μ* -calpains.

The role of regucalcin in the regulation of neutral proteolytic activity in rat kidney cortex cytosol is also examined [171, 172]. Regucalcin had an activatory effect on neutral proteolytic activity in the kidney cortex cytosol [172]. This increase was abolished in the presence of anti-regucalcin monoclonal antibody, supporting the view that endogenous regucalcin plays a role as activator of proteases in the renal cortex cytosol [172]. The effect of regucalcin on proteolytic activity was not altered in the presence of calcium chloride (0.01 and 1.0 mM) or EGTA (1.0 mM), indicating that the effect of regucalcin was independent on Ca^{2+} . Regucalcin may activate both calpains and other proteases in the kidney cortex cytosol. Regucalcin has been also shown to activate SH proteases in rat renal cortex cytosol. Presumably, regucalcin acts on the SH groups of protease in rat renal cortex cytosol.

The activatory effect of regucalcin on proteases is seen in the concentrations of 0.01–0.25 μM [171]. The concentration of regucalcin in rat kidney tissue is found to be present at about 5.3 μM . Regucalcin plays a physiological role in the activation of thiol proteases in renal cortex cells.

Regucalcin uniquely activates thiol proteases independent on Ca^{2+} in the liver cytosol, whereas it has no effect on serine proteases and metalloproteases [166, 167]. Such an effect of regucalcin was also found in the renal cortex cytosol [172]. Regucalcin may be an activator on thiol proteases in liver and kidney cells. Regucalcin also plays a role as an activator in other tissues that express regucalcin.

Calpains are ubiquitous, non-lysosomal, calcium-dependent proteases that may play important roles in Ca^{2+} -mediated intracellular processes [164, 165, 170]. The ability of calpain to alter the limited proteolysis, the activity or function of numerous cytoskeletal proteins, protein kinases, receptors and transcription factors suggests the involvement of the protease in various Ca^{2+} -regulated cellular functions [173]. Calpains may also play an integral

role in modulating the activity of protein kinase C, a key protein in many signal transduction processes [174], since they convert the native Ca^{2+} /phospholipids-dependent kinase to a soluble form that does not require Ca^{2+} or phospholipids for activity [2]. Regucalcin can increase the activity of thiol proteases including calpain in rat liver and renal cortex cytosols. Regucalcin may play a pivotal role in the regulation of cellular functions related to Ca^{2+} mediated through thiol proteases. Presumably, regucalcin plays an important role in the regulation of signal transduction that is involved in proteases.

Role of regucalcin in the regulation of nuclear function

Mounting evidence suggests that Ca^{2+} is active in liver nuclear function [81–83]. Calmodulin exists in rat liver nuclei [82]. The existence of an ATP-stimulated Ca^{2+} -sequestration system in rat liver nuclei that requires calmodulin and generates a net increase in nuclear matrix free Ca^{2+} concentration has been reported [81]. Calmodulin stimulates DNA synthesis in liver cells [175], and the effect of calmodulin is mediated through α -adrenergic stimulation [176, 177]. There are many evidences that regucalcin plays an important role in the regulation of nuclear functions.

Nuclear localization of regucalcin and its binding to nuclear protein or DNA

Regucalcin has been shown to localize in the nuclei of rat liver [37, 148]. Regucalcin can bind on calmodulin-agarose beads [139]. Liver nuclear extract were incubated with calmodulin-agarose beads, and calmodulin-agarose beads were applied to SDS-PAGE. Band that coincides with regucalcin was found on SDS-PAGE [139], suggesting that regucalcin is present in the nucleus [148].

Exogenous regucalcin has been shown to transport into the nucleus isolated from normal rat liver [37]. Endogenous regucalcin is present in the nuclei of rat liver using western blotting analysis [37]. When isolated liver nuclei were incubated in the presence of exogenous regucalcin (50 $\mu\text{g}/\text{ml}$; 1.5 μM), potent band for regucalcin was found in the nucleus [37], supporting that the protein is translocated into the nucleus. This translocation was seemed to be an early event, since potent band for regucalcin was seen with only 10 min of incubation. A part of regucalcin, which is localized in the cytoplasm of liver cells, is translocated to the nucleus [37].

Nuclear regucalcin translocation was not appreciably changed in the presence of ATP (2 mM), guanosine 5'-triphosphate (GTP, 2 mM), and calcium chloride (0.1 mM), suggesting that its translocation is not mediated through nuclear localization signal [37]. ATP and GTP are

required for nuclear import of proteins that are localized in the nuclei. ATP or GTP does not regulate the translocation of regucalcin into liver nuclei. Regucalcin is translocated independently of Ca^{2+} .

Nuclear protein transport is blocked in the presence of the lectin wheat germ agglutinin (WGA) [83]. Nuclear regucalcin translocation was not appreciably changed in the presence of WGA in the reaction mixture [37]. This finding suggests that the nuclear translocation of regucalcin is not related to nuclear localization signal that is responsible for selection for intranuclear active transport. Presumably, regucalcin is passively transported to the nucleus through nuclear pore in liver cells, since a molecular weight of regucalcin is about 33 kDa [16].

Regucalcin has been also shown to localize in the nuclei of the cloned normal rat kidney proximal tubular epithelial NRK52E cells with immunocytochemical analysis [178]. The nuclear localization of regucalcin is enhanced through hormonal signaling process that is involved in protein kinase C [178].

Regucalcin has been shown to bind proteins in isolated rat liver nuclei using Far-western blot analysis [179]. The results of the Far-western analysis showed the existence of protein components that bind to regucalcin in the nucleus isolated from rat liver [179]. Regucalcin has been also demonstrated to bind DNA using western blot analysis for regucalcin with DNA cellulose-binding assay [179]. These findings show that regucalcin binds proteins and DNA in liver nucleus. Regucalcin may have a regulatory effect on signaling pathways that modulate transcriptional activity in liver cells.

Regucalcin inhibits Ca^{2+} -stimulated nuclear DNA fragmentation

Isolated rat liver nucleus contains a DNA endonuclease activity dependent upon Ca^{2+} and Ca^{2+} results in extensive DNA hydrolysis [180]. The Ca^{2+} dependence of this endogenous DNA fragmentation process is based on DNA endonuclease activity dependent upon sub-micromolar Ca^{2+} when the nucleus is reconstituted with NAD^+ and ATP [180]. This endogenous endonuclease activity may be responsible for the DNA fragmentation occurring during programmed cell death (apoptosis) and certain forms of chemically induced cell killing [180, 181].

To explore the regulatory role of regucalcin in liver nuclear function, it was first examined whether the protein has an effect on Ca^{2+} -activated DNA fragmentation in isolated rat liver nuclei [182]. Among various metals, Ca^{2+} has been shown to stimulate uniquely in vitro DNA fragmentation in isolated rat liver nuclei [182]. This increase was seen after the addition of 1.0 μM Ca^{2+} , in agreement with previous work [182]. The presence of regucalcin

(0.5–2.0 μM) completely inhibited the activation of liver nuclear DNA fragmentation when 10 μM Ca^{2+} was added. This inhibition was not seen in the presence of Ca^{2+} at 25 or 50 μM . Thus, regucalcin had an inhibitory effect on DNA fragmentation when a comparatively lower concentration of Ca^{2+} (5.0 and 10 μM) was added [182]. The inhibitory effect of regucalcin on DNA fragmentation may be partly based on binding of Ca^{2+} [9].

DNA fragmentation in rat liver nucleus has been reported to be stimulated through Ca^{2+} -calmodulin [180], which exists in liver nuclei [82]. Addition of calmodulin (10 and 20 $\mu\text{g/ml}$) did not enhance Ca^{2+} (10 μM)-activated DNA fragmentation in liver nuclei [182]; however, nuclear endogenous calmodulin may be able to enhance Ca^{2+} -activated DNA fragmentation in the nucleus. Regucalcin has been found to inhibit Ca^{2+} -activated DNA fragmentation after Ca^{2+} addition [182]. Such an inhibition was also seen in the presence of exogenous calmodulin [182]. Presumably, regucalcin can inhibit Ca^{2+} /calmodulin-dependent DNA fragmentation in liver nuclei, since radioiodinated regucalcin has been found in the nuclei isolated from rat liver in the absence or presence of 1.0 mM Ca^{2+} [65].

Several studies have shown that Ca^{2+} plays an important role in the regulation of nuclear functions [81, 82]. Also, it has been found that a sustained increase in cytosolic Ca^{2+} level precedes the activation of DNA fragmentation that is characteristic of programmed cell death (apoptosis) and in certain forms of chemically induced cell killing [180, 181]. The finding, that regucalcin inhibits the activation of DNA fragmentation by Ca^{2+} , was the first time for a role of regucalcin in liver nuclear functions.

Regucalcin suppresses nuclear enzyme activity

Small GTPase Ran (ras-related nuclear protein) is required for protein export from the nucleus and protein import into the nucleus [183]. The role of regucalcin in the regulation of GTPase activity in the nuclei of rat liver is shown [184]. We found the existence of GTPase activity in the nuclei isolated from rat liver [184]. Liver nuclear GTPase activity was increased after calcium addition with a comparatively higher concentration in the enzyme reaction mixture, and this increase was not seen in the presence of TFP, an antagonist of calmodulin [184]. The effect of calcium in increasing nuclear GTPase activity may be related to endogenous calmodulin. Calcium in liver cytoplasm is transported through an energy-dependent mechanism to the nucleus, and the comparatively higher concentration of calcium is found in the nucleus [184]. Calmodulin is shown to be present in liver nucleus [82]. GTPase, which is activated by Ca^{2+} /calmodulin, may also be localized in liver nucleus.

The presence of exogenous regucalcin (0.5 μM) used in the enzyme reaction mixture caused an inhibitory effect on GTPase activity in liver nucleus [184]. This effect was also seen in the presence of EGTA, a chelator of Ca^{2+} . Presumably, the inhibitory effect of regucalcin on liver nuclear GTPase activity is revealed independent of Ca^{2+} /calmodulin in the nucleus. Regucalcin has been shown to have an inhibitory effect on the activation of enzymes by Ca^{2+} /calmodulin due to binding Ca^{2+} and/or calmodulin [135–138]. Regucalcin can inhibit the activity of various enzymes through the mechanism by which it binds directly to the enzyme [30, 32]. Regucalcin may directly inhibit GTPase activity in liver nucleus.

The physiological significance of the inhibitory effect of regucalcin on GTPase activity in liver nucleus is unknown. However, the presence of anti-regucalcin monoclonal antibody in the enzyme reaction mixture caused a significant increase in this activity in liver nucleus [184]. This increase was completely blocked after regucalcin addition, suggesting that endogenous regucalcin has a suppressive effect on GTPase activity in liver nucleus. Endogenous regucalcin, which is localized in liver nucleus, may participate in the regulation of nuclear functions that are related to hydrolysis of GTP.

Regucalcin may be able to regulate a process of signal transduction from the cytoplasm to nucleus in liver cells. This process is mediated through various protein kinases. The role of regucalcin in the regulation of Ca^{2+} -dependent protein kinase and protein tyrosine phosphatase activities in isolated liver nucleus is examined [37, 147, 148]. Nuclear Ca^{2+} -dependent protein kinase and protein tyrosine phosphatase activities were increased in the presence of anti-regucalcin monoclonal antibody in the enzyme reaction mixture, and these increases were completely abolished after the addition of regucalcin [37]. The translocation of regucalcin to the nucleus may play a suppressive role in the regulation of protein kinase and protein tyrosine phosphatase in liver nucleus [37].

Endogenous regucalcin has been demonstrated to have a suppressive effect on the enhancement of protein kinase activity with a proliferation of liver cells [185]. Protein kinase activity is enhanced in the cytosol and nucleus of regenerating rat liver [185]. Regucalcin had a suppressive effect on tyrosine kinase, protein kinase C, and Ca^{2+} /calmodulin-dependent protein kinase in the cytoplasm and nucleus of regenerating rat liver [120, 135, 136, 185].

Phosphatase activity toward phosphotyrosine, phosphoserine, and phosphothreonine is found in the liver nucleus [148]. Nuclear phosphotyrosine phosphatase activity was increased after Ca^{2+} addition in the enzyme reaction mixture, although the enzyme activity was not altered by TFP, an inhibitor of calmodulin, or cyclosporine A, an inhibitor of calcineurin [148]. Nuclear phosphatase activity

toward phosphotyrosine may be independent of calmodulin. Meanwhile, nuclear phosphatase activity toward phosphoserine was elevated after the addition of Ca^{2+} , while the enzyme activity was appreciably decreased by TFP and cyclosporine A, suggesting that the enzyme activity is partly involved in Ca^{2+} /calmodulin-dependent protein phosphatase (calcineurin). Nuclear phosphatase activity toward phosphothreonine was not altered after the addition of Ca^{2+} , TFP, and cyclosporine A in the enzyme reaction mixture. Vanadate caused an inhibition of nuclear phosphatase activity toward phosphotyrosine and phosphoserine but not phosphothreonine. Thus, different protein phosphatases toward phosphotyrosine, phosphoserine, and phosphothreonine have been shown to be present in liver nucleus.

The addition of regucalcin in the enzyme reaction mixture caused a significant decrease in phosphatase activity toward phosphotyrosine, phosphoserine, and phosphothreonine in the liver nuclei [148]. Liver nuclear phosphatase activity toward phosphoamino acids was assayed using 5–6 mg of nuclear protein per milliliter of reaction mixture; it contained 275–330 ng of nuclear regucalcin [148]. The concentration of endogenous nuclear regucalcin was estimated to be about 82.4–98.8 nM. Further addition of exogenous regucalcin (0.25 μM) caused a significant decrease in nuclear phosphatase activity, although the effect was saturated with increasing concentrations of regucalcin (0.5 μM) [148].

Nuclear phosphatase activity was elevated in the presence of anti-regucalcin monoclonal antibody (25 and 50 ng/ml of reaction mixture) [148]. This elevation was completely abolished after addition of regucalcin. Endogenous regucalcin may regulate protein phosphatase activity toward phosphotyrosine, phosphoserine, and phosphothreonine in liver nucleus. Regucalcin may have an inhibitory effect on various protein phosphatases in liver nucleus.

Regucalcin suppresses nuclear DNA and RNA synthesis

Regucalcin has been shown to have an inhibitory effect on DNA synthesis activity in the nuclei of normal rat liver [186]. The inhibitory effect of regucalcin was seen in the presence of EGTA, a chelator of Ca^{2+} , in the reaction mixture. Ca^{2+} is present in liver nucleus [186]. Liver nuclear DNA synthesis activity was increased in the presence of EGTA in the reaction mixture, suggesting that Ca^{2+} suppresses DNA synthesis activity in the nucleus. The effect of regucalcin in inhibiting nuclear DNA synthesis may be not related to Ca^{2+} in liver nucleus.

Liver nuclear DNA synthesis has been shown to stimulate in regenerating rat liver [187]. Nuclear DNA synthesis was markedly increased at 1 day after hepatectomy,

and this increase was also seen at 3 days [187]. Nuclear DNA synthesis was enhanced in the presence of EGTA (0.4 mM) in the incubation mixture. The presence of Ca^{2+} (1.0–25 μM) caused a significant decrease in the nuclear DNA synthesis of normal rat liver. Regucalcin (0.25 and 0.5 μM) caused an inhibition of nuclear DNA synthesis of normal rat liver [187]. This inhibition was also seen in the presence of Ca^{2+} (1.0 μM). The inhibitory effect of regucalcin was remarkable in regenerating rat liver nuclei in comparison with that of normal rat liver. Regucalcin has been shown to have a suppressive effect on nuclear DNA synthesis in regenerating rat liver. Regucalcin may have a suppressive role in the enhancement of nuclear DNA synthesis in liver cell proliferation.

Regucalcin has been also shown to have a suppressive effect on DNA synthesis activity in the nuclei isolated from rat renal cortex [188]. The addition of regucalcin (0.1–0.5 μM) in the reaction mixture containing either EGTA (1 mM) or calcium chloride (50 μM) had an inhibitory effect on nuclear DNA synthesis activity [188]. The presence of anti-regucalcin monoclonal antibody (10–50 ng/ml) in the reaction mixture caused a significant increase in nuclear DNA synthesis activity [188]. This increase was completely abolished in the presence of regucalcin (0.5 μM). Endogenous regucalcin has been found to have a suppressive effect on DNA synthesis in the nuclei of rat renal cortex [188].

Regucalcin has been shown to have an inhibitory effect on RNA synthesis in the nuclei isolated from control rat liver [189] and regenerating rat liver [190]. RNA synthesis in rat liver nuclei was stimulated after Ca^{2+} addition with a comparatively lower concentration, although the submicromolar concentration of Ca^{2+} evoked an inhibition of nuclear RNA synthesis [189]. The addition of Ca^{2+} with higher concentrations has been reported to have an inhibitory effect of nuclear RNA synthesis in rat liver cells [191]. Since liver nuclei contain a DNA endonuclease activity dependent upon Ca^{2+} in the submicromolar range and Ca^{2+} causes extensive DNA hydrolysis [180, 182], nuclear RNA synthesis may be suppressed with higher Ca^{2+} concentrations. Inactivation of RNA polymerase III transcription has been shown to be calcium dependent; the changes in Ca^{2+} concentration, the activation of calpains, and the consequent proteolytic degradation of RNA of transcription factors has been suggested to be involved in the regulation of RNA polymerase III transcription in the presence of 1 mM Ca^{2+} [192].

The effect of regucalcin in decreasing nuclear RNA synthesis activity in normal rat liver was not seen in the presence of α -amanitin, an inhibitor of RNA polymerase II and III [189], suggesting that the suppressive effect of regucalcin on nuclear RNA synthesis activity is partly resulted from its inhibitory action on RNA polymerase II

and III [189, 190]. Meanwhile, it has been reported that Ca^{2+} has a stimulatory effect on RNA synthesis in liver nucleus [191, 192]. This effect may be partly mediated through Ca^{2+} -dependent protein kinase [191, 192]. The stimulatory effect of Ca^{2+} on nuclear RNA synthesis activity was completely blocked in the presence of regucalcin [189, 190]. Regucalcin has been shown to inhibit Ca^{2+} -dependent protein kinases in rat liver nucleus [185]. Presumably, the effect of regucalcin in decreasing RNA synthesis activity in liver nucleus is partly involved in its inhibitory action on the activities of both RNA polymerase II and III and Ca^{2+} -dependent protein kinases. Further mechanism remains to be elucidated. Regucalcin has been proposed to have a role as a transcriptional factor in liver nucleus.

Regucalcin has been found to have a suppressive effect on liver nuclear DNA and RNA synthesis [186–190]. The mechanism by which regucalcin inhibits nuclear DNA and RNA synthesis is not well known. It is speculated that regucalcin has an inhibitory effect on DNA and RNA polymerase activity. However, the effect of regucalcin in inhibiting nuclear RNA synthesis activity is observed in the presence of α -amanitin, an inhibitor of RNA polymerase II. Regucalcin can directly bind DNA [179]. Which base pairs of DNA bind regucalcin remains to be elucidated. It is possible that regucalcin binds DNA and it has an inhibitory effect on nuclear DNA and RNA synthesis activity.

Regucalcin suppresses apoptosis mediated through cell signaling

Regucalcin inhibits NO synthase activity that is related to apoptosis

Nitric oxide (NO) may be important as a signaling factor in many cells [122], and it plays a role in apoptosis of hepatoma cells [193]. NO mediates apoptosis by D-galactosamine in a primary culture of rat hepatocytes [194]. Regucalcin has been shown to inhibit NO synthase that is related to cell apoptosis [123], suggesting that regucalcin has a suppressive role in apoptosis [195].

Regucalcin has a suppressive effect on Ca^{2+} /calmodulin-dependent NO synthase in the cloned rat hepatoma H4-II-E cells [123]. The effect of regucalcin in decreasing NO synthase activity was also seen in the presence of TFP or EGTA. Presumably, regucalcin has an inhibitory effect on NO synthase activity due to binding to calmodulin and/or the enzyme independently of Ca^{2+} in proliferative cells.

Overexpression of regucalcin has been also shown to have a suppressive effect on NO synthase activity in H4-II-E cells (transfectants) [123]. This decrease was completely abolished in the presence of anti-regucalcin monoclonal

antibody in the reaction mixture. Moreover, the effect of Ca^{2+} /calmodulin addition in increasing NO synthase activity in H4-II-E cells (wild type) was completely prevented in transfectants. Endogenous regucalcin had a suppressive effect on NO synthase activity the cloned rat hepatoma H4-II-E cells.

Nitric oxide (NO) synthase activity was enhanced in H4-II-E cells cultured with 10% FBS as compared with that of 1% FBS [123], suggesting that the enzyme is induced in proliferative cells. The enhancement of NO synthase activity in H4-II-E cells cultured with 10% FBS was abolished in the presence of anti-regucalcin monoclonal antibody [123]. Regucalcin levels were elevated in H4-II-E cells with 10% FBS-culture [123]. Endogenous regucalcin may have a suppressive effect on the enhancement of NO synthase activity with proliferation of H4-II-E cells.

A high concentration of NO, which is produced from inducible NO synthase, has been shown to inhibit cell proliferation [195] and to induce cell apoptosis [196]. It is reported that a low concentration of NO, which is produced from endothelial NO synthase, protects against the cytotoxic effects of reaction oxygen species in cells [197]. Whether endogenous regucalcin suppresses NO production in H4-II-E cells is unknown at present. It is speculated, however, that regucalcin may inhibit NO production in H4-II-E cells, since regucalcin can decrease NO synthase activity in the cells [123]. Endogenous regucalcin may have an inhibitory effect on inducible and endothelial NO synthetases in hepatoma cells. Alternatively, regucalcin may have a physiological role in the regulation of NO-related cell functions.

Regucalcin suppresses various factors-induced cell death and apoptosis in liver cells

Tumor necrosis factor α (TNF- α) and NO mediate apoptosis by D-galactosamine in a primary culture of rat hepatocytes [194, 195]. TNF- α induces apoptosis in mammary adenocarcinoma cells by an increase in intranuclear free Ca^{2+} concentration and DNA fragmentation [195]. H4-II-E cells with subconfluent monolayer cells were cultured in a medium without FBS in the presence of TNF- α . TNF- α (0.1–10 ng/ml) caused a significant decrease in the number of H4-II-E cells (wild type), inducing cell death. Overexpression of regucalcin in H4-II-E cells (transfectants) has been found to prevent the effect of TNF- α in decreasing cell number [198]. Overexpression of regucalcin had a preventive effect on cell death induced with the higher concentration of TNF- α (10 ng/ml). This finding demonstrates that overexpression of regucalcin has a suppressive effect on cell death induced by stimulation of TNF- α [198].

Culture with NAME, an inhibitor of NO synthase, had a significant preventive effect on TNF- α -induced cell death.

Regucalcin inhibits Ca^{2+} /calmodulin-dependent NO synthase activity in H4-II-E cells [123]. The suppressive effect of regucalcin on cell death may be partly resulted from the inhibition of NO production, which can induce apoptosis, stimulated after TNF- α stimulation in H4-II-E cells.

The effect of caspase inhibitor on TNF- α -mediated cell death in H4-II-E cells is examined [198]. TNF- α -induced cell death was prevented in culture with caspase inhibitor in wild-type cells and transfectants, suggesting that TNF- α -induced cell death is partly involved activation of caspases in H4-II-E cells. Regucalcin may have an inhibitory effect on activation of caspases in the cells.

Lipopolysaccharide (LPS) has been shown to induce cell apoptosis [199, 200]. H4-II-E cells with the subconfluent monolayer were cultured in a medium without FBS in the presence of LPS. LPS caused a decrease in the number of H4-II-E cells (wild-type), inducing cell death and apoptosis [201]. This decrease was completely prevented in the regucalcin cDNA-transfected hepatoma cells overexpressing regucalcin with culture for 12–48 h [201]. Overexpression of regucalcin has a suppressive effect on LPS-stimulated cell death and apoptosis.

LPS acts to modulate the expression of a large number of genes that favor apoptosis of fibroblastic cells that are dependent upon activation of caspase-8 [200]. There is evidence that LPS-induced cell death is mediated through accumulation of reactive oxygen species and activation of p38 in rat cortex and hippocampus [200]. Culture with LPS caused a significant decrease in Ca^{2+} /calmodulin-dependent NO synthase activity in H4-II-E (wild type) cells [201]. LPS-induced decrease in NO synthase activity was found to prevent in the transfectants overexpressing regucalcin [201]. LPS-induced cell death may be not result from NO production in hepatoma cells, and the suppressive effect of regucalcin on LPS-induced cell death is not involved in NO in the cells. Moreover, LPS-induced cell death was prevented in culture with caspase-3 inhibitor [201]. The effect of regucalcin in suppressing LPS-induced cell death is partly related to the inhibitory effect on caspase-3 in hepatoma cells.

An induction of apoptosis is partly mediated through pathway of protein kinase. The death of H4-II-E cells (wild type) has been found to be induced in culture with PD98059, a ERK inhibitor, dibucaine, an inhibitor of Ca^{2+} -dependent protein kinase, or staurosporine, a potent inhibitor of protein serine/threonin kinases (protein kinase C), suggesting that various inhibitors-induced cell deaths are partly involved in the inhibition of protein kinases [201]. Overexpression of regucalcin in H4-II-E cells rescued cell death with PD98059 or dibucaine [201]. Such an effect was not observed with staurosporine. PD98059 induces apoptosis, which is in part due to the inactivation of Bcl-2 by increasing phosphorylated Bcl-2 in human

prostate cancer cells [202]. Dibucaine has been shown to activate various caspases, such as caspase-3, -6, -8, and -9 (-like) activities, but not caspase-1 (-like) activity, and to induce mitochondrial membrane depolarization and the release of cytochrome C from mitochondria into the cytosol in leukemia cells (HL-60) [203]. Staurosporine induces apoptosis in Chang liver cells by a mitochondria-caspase-dependent pathway, which is closely correlated with a decrease in Bcl-2 and Bcl-XL levels in cancer cells [204]. Regucalcin may partly inhibit the inactivation of Bcl-2 or the activation of caspases in signaling mechanism that PD98059 or dibucaine induces apoptosis.

Calcium channel blockers, the endoplasmic reticulum Ca^{2+} -ATPase inhibitor thapsigargin and calcium ionophores are potent to lead several cell types to apoptosis [205, 206]. Thapsigargin is an inhibitor of Ca^{2+} -ATPase in the endoplasmic reticulum (Ca^{2+} store) in cells, and treatment with thapsigargin causes an elevation of sustained Ca^{2+} concentration in cells and induces apoptosis in the hepatoma cells [207]. Experiments on nucleus isolated from cells clearly demonstrate the induction of Ca^{2+} -dependent endonuclease activity during triggering apoptosis events [207]. Rises in intracellular Ca^{2+} concentration are believed to activate this nuclease and to mediate DNA cleavages into oligonucleosome fragments [208]. Regucalcin has been shown to have an inhibitory effect on Ca^{2+} -activated DNA fragmentation in isolated rat liver nucleus [182], suggesting that the protein has an inhibitory effect on apoptosis in liver cells. Thapsigargin induces cell death and apoptosis causing DNA fragmentation [202]. Thapsigargin-induced DNA fragmentation in the hepatoma cells is not altered in culture with caspase inhibitor, suggesting that thapsigargin-mediated apoptosis is independent of activation of caspases [202]. Overexpression of regucalcin in the hepatoma cells has been found to suppress DNA fragmentation induced by thapsigargin [202]. This effect was not further enhanced in culture with caspase inhibitor [202]. Presumably, regucalcin has a suppressive effect on thapsigargin-mediated cell death due to preventing the rise in intracellular Ca^{2+} concentration in the hepatoma cells, since the protein can keep intracellular Ca^{2+} homeostasis due to activating Ca^{2+} pump enzymes in the plasma membranes, mitochondria, and endoplasmic reticulum of rat liver cells [30–33].

Calcium entry into cells induces cell death [202, 209]. Culture with Bay K 8644, an antagonist of Ca^{2+} entry in cells, caused a significant increase in the death of hepatoma H4-II-E cells (wild-type) [202]. Culture with Bay K 8644 did not induce cell death of transfectants (H4-II-E cells) overexpressing regucalcin [202]. Overexpression of regucalcin in H4-II-E cells was found to suppress DNA fragmentation induced by Bay K 8644. Regucalcin may have a suppressive effect on Ca^{2+} entry-induced stimulation of

apoptosis in the hepatoma cells [202]. Regucalcin has a suppressive effect on Ca^{2+} entry-mediated cell death due to preventing the rise in intracellular Ca^{2+} concentration in the hepatoma cells. In addition, regucalcin may suppress the effect of Ca^{2+} on DNA fragmentation in the nucleus of H4-II-E cells.

The effect of insulin or IGF-I on cell death and apoptosis in H4-II-E cells has not been well known. H4-II-E cells were cultured in a medium containing, either vehicle, insulin, insulin-like growth factor-I (IGF-I), epinephrine, or transforming growth factor- β in the absence of FBS [210]. The number of wild-type cells was decreased in the presence of insulin or IGF-I [210]. Agarose gel electrophoresis showed the presence of low-molecular-weight DNA fragments of adherent wild-type cells cultured with insulin or IGF-I [210]. The effect of insulin or IGF-I in stimulating cell death and DNA fragmentation H4-II-E cells (wild type) was prevented in transfectants overexpressing regucalcin [210].

The effect of insulin in decreasing the number of H4-II-E cells was prevented in the presence of caspase-3 inhibitor [210]. The effect of IGF-I on cell death, however, was also observed in the presence of caspase-3 inhibitor [210]. These observations suggest that the effect of insulin on cell death is involved in activation of caspase-3, and that the effect of IGF-I is not dependent on caspase-3 in H4-II-E cells. The effect of IGF-I in inducing cell death in the presence of caspase-3 inhibitor was completely blocked in transfectants overexpressing regucalcin [210], suggesting that regucalcin inhibits signaling pathway of IGF-I-induced cell death that is not mediated through caspase-3 in H4-II-E cells.

The effect of insulin or IGF-I in inducing cell death and apoptosis of H4-II-E cells was not observed in the presence of NAME, an inhibitor of NO synthase [210], suggesting that insulin- or IGF-induced cell death is partly involved in production of NO in H4-II-E cells. Overexpression of regucalcin has been shown to have a suppressive effect on activation of Ca^{2+} /calmodulin-dependent NO synthase in H4-II-E cells [123].

The effect of IGF-I in inducing cell death of H4-II-E cells was also observed in the presence of Bay K 8644 [210]. Such an effect was not seen in the case of insulin [210]. The mode of IGF-I action differs from that of insulin. It is assumed that insulin induces cell death that is partly mediated through intracellular calcium-dependent signaling pathway in H4-II-E cells, and that IGF-I may not be mediated through calcium-dependent signaling pathway in H4-II-E cells. The effect of IGF-I in inducing cell death in the presence of Bay K 8644 was not observed in transfectants overexpressing regucalcin [210].

Genistein has an inhibitory effect on protein tyrosine kinases and it can produce cell cycle arrest and apoptosis in

leukemic cells [211]. Genistein was found to induce cell death of H4-II-E cells, and the effect was not seen in the transfectants overexpressing regucalcin [210]. Genistein-induced cell death is partly mediated through inhibition of protein tyrosine kinase in H4-II-E cells. Regucalcin has an inhibitory effect on protein tyrosine kinase activity in the cytoplasm and nucleus of rat liver [185].

The effect of insulin in inducing cell death of H4-II-E cells was not seen in the presence of genistein [210], although such an effect was seen in the case of IGF-I. The effect of IGF-I on cell death in the presence of genistein was prevented in transfectants overexpressing regucalcin [210]. Regucalcin has a suppressive effect on cell apoptosis that is mediated through signaling pathways with dependent or independent on protein tyrosine kinase.

Vanadate is an inhibitor of protein tyrosine phosphatase in cells [142]. Regucalcin has been shown to have an inhibitory effect on protein tyrosine phosphatase activity in the cytoplasm and nucleus of rat liver [147]. Vanadate was found to induce cell death of H4-II-E cells [210], suggesting that cell death is not caused by mechanism that is mediated through inhibition of protein tyrosine phosphatase activity. Vanadate induced cell death for transfectants overexpressing regucalcin [210], suggesting that the suppressive effect of regucalcin on cell death of H4-II-E cells is independent on protein phosphatase. IGF-I had a stimulatory effect on cell death of H4-II-E cells overexpressing regucalcin in the presence of vanadate [210], suggesting that the effect of IGF-I is not mediated through protein tyrosine phosphatase in the transfectants.

The effect of insulin in inducing cell death may be partly mediated through signaling pathway which is involved in caspase-3, calcium, NO, protein tyrosine kinase, or protein tyrosine phosphatase in H4-II-E cells. The effect of IGF-I on cell death of H4-II-E cells may be mediated through NO and other molecules. Overexpression of regucalcin may have a suppressive effect on signaling mechanism by which insulin or IGF-I induces cell death of H4-II-E cells.

Sulforaphane is an isothiocyanate that is present naturally in widely consumed vegetables and has a particularly high concentration in broccoli. This compound has been shown to block the formation of tumors initiated by chemicals in the rat [212]. Sulforaphane has been shown to induce a cell cycle arrest, followed by cell death in HT29 human colon cancer cells [212]. Sulforaphane increases expression of the pro-apoptotic protein Bax, the release of cytochrome C from the mitochondria to the cytosol, and the proteolytic cleavage of poly (ADP-ribose) polymerase in HT29 human colon cancer cells [212]. In human T-cell leukemia, sulforaphane induces apoptosis due to increased p53 and Bax protein expression, and slightly affected Bcl-2 expression [213]. In cultured PC-3 human prostate cancer cells, moreover, sulforaphane-induced apoptosis is associated with up-regulation of Bax,

down-regulation of Bcl-2 and activation of caspase-3, -9, and -8 [214]. Sulforaphane has been found to induce cell death and apoptosis in H4-II-E cells [215]. Caspase-3 inhibitor prevented the effect of sulforaphane, while it was not inhibited by NAME, an inhibitor of NO synthase, in H4-II-E cells [215]. Sulforaphane-induced cell death and apoptosis partly result from activation of caspase-3 in the hepatoma cells.

Overexpression of regucalcin had a suppressive effect on cell death and apoptosis induced by sulforaphane in H4-II-E cells [215]. The suppressive effect of regucalcin on sulforaphane-induced cell death and apoptosis in H4-II-E cells may be partly involved in the molecules of Bax, cytochrome C, caspase, and Bcl-2. In addition, regucalcin may have an inhibitory effect on NO synthase and Ca^{2+} -dependent endonuclease activities in H4-II-E cells [123]. Regucalcin has a suppressive effect on many signaling pathways that mediate apoptotic cell death.

As mentioned above, regucalcin has been shown to play a role in the regulation of cell death and apoptosis in H4-II-E cells [198, 201, 210]. Overexpression of regucalcin has been demonstrated to have a suppressive effect on cell death and apoptosis induced by TNF- α , LPS, thapsigargin, Bay K 8644, dibucaine, or PD98059, an inhibitor of protein tyrosine kinase, insulin, or IGF-I in H4-II-E cells [198, 201, 210]. The signaling mechanisms that TNF- α , LPS, or other factors mediate cell death and apoptosis may be different. The suppressive effect of regucalcin on apoptotic cell death is related to its inhibitory effect on the activities of various protein kinases, NO synthase, caspase-3, or Ca^{2+} -dependent endonuclease, and its activatory effect on Bcl-2. Regucalcin has a suppressive effect on many signaling pathways that mediate cell death and apoptosis, and the protein suppresses cell death and apoptosis mediated through many different signaling pathways in H4-II-E cells.

Regucalcin has a suppressive effect on cell apoptosis in kidney cells

Regucalcin has been shown to express in the cloned normal rat kidney proximal tubular epithelial NRK52E cells and its expression is enhanced after hormonal stimulation [216]. Nuclear localization of regucalcin is enhanced after hormone stimulation in NRK52E cells [178]. The role of regucalcin in cell death and apoptosis is examined using NRK52E cells overexpressing regucalcin [217]. The number of wild-type cells was decreased with culture for 42–72 h in the presence of TNF- α , LPS, Bay K 8644, or thapsigargin [217]. These effects were prevented in transfectants overexpressing regucalcin. DNA fragmentation induced after culture with LPS, Bay K 8644, or thapsigargin were prevented in transfectants overexpressing regucalcin [217]. Thus, overexpression of regucalcin has a

suppressive effect on apoptotic cell death induced by TNF- α , LPS, Bay K 8644, or thapsigargin in kidney NRK52E cells. The effect of regucalcin in suppressing apoptotic cell death may be mediated through its action on many intracellular signaling pathways in NRK52E cells.

Bcl-2 is a suppressor in apoptotic cell death [218]. Apaf-1 participates in activation of caspase-3 [219]. Akt-1 involves in survival signaling pathway for cell death [220]. Overexpression of regucalcin caused a remarkable elevation of Bcl-2 mRNA expression in NRK52E cells, and it slightly stimulated Akt-1 mRNA expression in the cells. Apaf-1, caspase-3, or G3PDH mRNA expressions were not significantly altered in transfectants [217]. Presumably, the enhancement of Bcl-2 mRNA expression contributes to the suppression of apoptotic cell death in NRK52E cells overexpressing regucalcin. Regucalcin may play a role in the regulation of Bcl-2 gene expression in NRK52E cells.

TNF- α enhanced the expression of caspase-3 mRNA in NRK52E cells [217]. This enhancement was found to suppress in transfectants [217], suggesting that the mechanism by which regucalcin suppresses TNF- α -induced cell death is partly related to the decrease in caspase-3 mRNA expression in transfectants.

The presence of LPS caused a significant decrease in Bcl-2 mRNA levels in NRK52E cells, suggesting that this decrease is partly related to LPS-induced cell death [217]. The enhancement of Bcl-2 mRNA expression induced by overexpression of regucalcin was also seen in the presence of LPS [217]. LPS-stimulated expression of Apaf-1 mRNA was suppressed after overexpression of regucalcin [217]. This may partly involve in the suppression of LPS-induced cell death in NRK52E cells overexpressing regucalcin.

Culture with Bay K 8644 or thapsigargin was found to cause a significant increase in caspase-3 mRNA levels in wild-type cells, indicating that the increased gene expression partly contributes to inducing apoptotic cell death [217]. This increase was completely prevented in transfectants. Regucalcin may have a suppressive effect on caspase-3 mRNA expression enhanced by Bay K 8644 or thapsigargin in NRK52E cells. Thus, regucalcin was found to regulate the expression of Bcl-2, caspase-3, and Akt-1 mRNAs in the cloned normal rat kidney NRK52E cells. The change in protein levels, however, remains to be elucidated.

Toxic factors have been reported to induce renal failure due to stimulating apoptotic cell death [221]. Overexpression of regucalcin was found to have a suppressive effect on apoptotic cell death induced by various factors (including TNF- α , LPS, Bay K 8644, or thapsigargin) in NRK52E cells. Regucalcin may have a role as a suppressor in the development of apoptotic cell death in kidney proximal tubular epithelial cells. Presumably, regucalcin plays a physiological role in the maintenance of homeostasis of cellular response for cell stimulation.

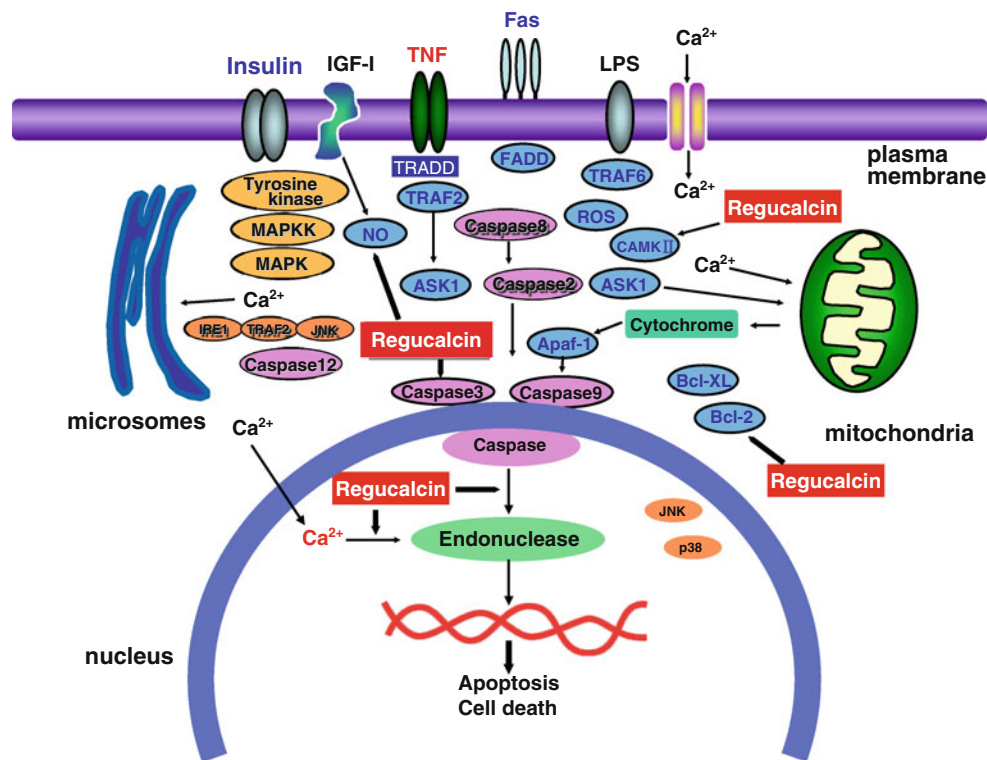


Fig. 2 Regucalcin has a role as suppressor in cell death and apoptosis induced by various factors. Regucalcin suppresses cell death induced by various factors (including TNF- α , insulin, IGF-I, LPS, PD98059, dibucaine, thapsigargin, Bay K 8644, or sulforaphane). The

suppressive effect of regucalcin on cell death and apoptosis is mediated due to inhibiting the activities of NO synthase, caspase-3, and Ca²⁺-dependent endonuclease and activating Bcl-2 in the cells

Conclusion

Overexpression of regucalcin rescues cell death and apoptosis induced in culture with various factors in the hepatoma cells and normal kidney cells. The cell signaling mechanisms, that these factors mediate cell death and apoptosis, may be different. Regucalcin may have a suppressive effect on many signaling pathways that mediate cell death and apoptosis, as is summarized in Fig. 2. The suppressive effect of regucalcin on cell death and apoptosis may be related to the inhibitory effect on the activities of NO synthase, caspase-3, or Ca²⁺-dependent endonuclease and its activatory effect on Bcl-2. Moreover, regucalcin has regulatory effects on gene expression of many molecules that are related to cell apoptosis. Regucalcin plays an important role as a regulatory protein in intracellular signaling pathway which is related to cell death and apoptosis.

Regucalcin suppresses cell proliferation

Regucalcin mRNA and its protein are expressed in the cloned rat hepatoma H4-II-E cells, although these expressions show low levels as compared to that in the cytosol of normal rat liver [50, 51]. The expression of regucalcin

mRNA is stimulated in H4-II-E cells after culture with addition of serum (10% FBS) [50, 51], suggesting that regucalcin plays a role in the proliferation of cells. Regucalcin has been demonstrated to have a role as suppressor in the enhancement of proliferation of liver cells in vitro. This section describes the mechanism by which regucalcin has a suppressive effect on cell proliferation using H4-II-E cells.

Regucalcin suppresses the enhancement of protein kinase activity in cell proliferation

The role of endogenous regucalcin in the regulation of protein kinase activity in the proliferation of H4-II-E cells is examined [222]. H4-II-E cells were cultured for 6–72 h in the presence of FBS (1 or 10%). The number of cells and protein kinase activity in the 5,500g supernatant of cell homogenate was increased 24 and 48 h after the culture with FBS (1 or 10%); the culture with 10% FBS had potent effect as compared with that of 1% FBS [222]. The culture with FBS produced an increase in protein kinase activity and a corresponding elevation of cell number in H4-II-E cells [222]. The increase in protein kinase activity of the 5,500g supernatant of cell homogenate preceded a significant elevation of cell number, suggesting that serum factors

(including growth factors and hormones) stimulate cell proliferation that is partly mediated through cascades of protein kinases.

Serum-stimulated protein kinase activity in H4-II-E cells was further enhanced in the presence of calmodulin or dioctanoylglycerol in the presence of calcium chloride, and this increase was inhibited in the presence of trifluoperazine, staurosporine, or genistein in the enzyme reaction mixture [222], indicating that Ca^{2+} /calmodulin-dependent protein kinase, protein kinase C, and protein tyrosine kinase are present in H4-II-E cells [222]. Various protein kinases may be involved in the enhancement of cell proliferation with serum stimulation.

The presence of anti-regucalcin monoclonal antibody in the enzyme reaction mixture containing the 5,500g supernatant of cell homogenate of H4-II-E cells with FBS caused a significant increase in protein kinase activity [222]. This effect was completely abolished after the addition of exogenous regucalcin, which has an inhibitory effect on the enzyme activity [222]. This finding indicates that endogenous regucalcin plays a suppressive role in the enhancement of protein kinase activity in the cytoplasm in H4-II-E cells with cell proliferation. The anti-regucalcin monoclonal antibody-increased protein kinase activity in H4-II-E cells was inhibited in the presence of trifluoperazine, staurosporine, or genistein, suggesting that endogenous regucalcin inhibits Ca^{2+} /calmodulin-dependent protein kinase, protein kinase C, or protein tyrosine kinase activities [222].

Serum stimulation may lead to an increase in cell proliferation that is partly mediated through cascade for various protein kinases in H4-II-E cells [222]. Regucalcin may have a suppressive effect for overexpression of cell proliferation due to inhibiting various protein kinases in the cytoplasm and nucleus of H4-II-E cells. Presumably, regucalcin plays a role as suppressor protein in cell proliferation of normal liver cells and hepatoma cells.

Regucalcin suppresses the enhancement of protein phosphatase activity in cell proliferation

Regucalcin has been shown to have an inhibitory effect on protein phosphatase activity in the cytoplasm of rat liver [147, 148]. H4-II-E cells were cultured for 3 days in a medium containing serum (10% FBS). After subconfluency, the cells were used for the assay of protein phosphatase activity toward phosphotyrosine. Ca^{2+} /calmodulin-dependent protein tyrosine phosphatases were present in H4-II-E cells [223]. Regucalcin had an inhibitory effect on Ca^{2+} /calmodulin-dependent protein tyrosine phosphatase activity in the cells [223]. The culture with Bay K 8644, an agonist of Ca^{2+} channels, caused an elevation of protein

tyrosine phosphatase in H4-II-E cells, whereas dibutyryl cyclic AMP had no effect [223]. Bay K 8644-induced increase in protein phosphatase activity was inhibited in the presence of TFP, indicating that this increase is mediated through Ca^{2+} /calmodulin in hepatoma cells [223]. The effect of antibody was enhanced in the presence of TFP [223]. This enhancement may result from an increase in endogenous regucalcin in H4-II-E cells, since the expression of regucalcin mRNA in H4-II-E cells is stimulated after the culture with Bay K 8644 [223]. This finding may support the view that endogenous regucalcin, which is enhanced through Ca^{2+} signaling, has a suppressive effect on Ca^{2+} /calmodulin-activated protein phosphatase activity in proliferative cells.

The presence of anti-regucalcin monoclonal antibody in the enzyme reaction mixture caused a remarkable elevation of protein tyrosine phosphatase activity in the cell homogenate (5,500g supernatant) of H4-II-E cells cultured with serum addition (1 or 10% FBS) [224]. This elevation was completely prevented after the addition of regucalcin [224]. Endogenous regucalcin may have a suppressive effect on the enhancement of protein tyrosine phosphatase activity in the proliferative cells.

There may be many protein phosphatases in hepatoma H4-II-E cells. The antibody-increased protein tyrosine phosphatase activity was also inhibited by okadaic acid or vanadate, which is an inhibitor of protein phosphatases [142], although cyclosporine A, an inhibitor of calcineurin (protein phosphatase) [225], had no effect [224]. Endogenous regucalcin may also act on okadaic acid or vanadate-sensitive protein tyrosine phosphatases in H4-II-E cells.

The culture with serum addition (10% FBS) caused an increase in proliferation of H4-II-E cells and a corresponding elevation of protein tyrosine phosphatase activity in the cells [224]. This finding suggests that the augmentation of protein tyrosine phosphatase activity is partly involved in the proliferation of H4-II-E cells, although protein phosphatase activity toward phosphoserine and phosphothreonine was not raised in the proliferative cells cultured with serum addition [224].

Processes that are reversibly controlled by protein phosphorylation require not only a protein kinase but also a protein phosphatase [142, 226]. Target proteins are phosphorylated at specific sites by one or more protein kinases and these phosphoproteins are removed by specific protein phosphatase [225].

As mentioned above, regucalcin may play an important role as a suppressor for the enhancement of cell proliferation due to inhibiting various protein kinases and protein phosphatases activities that are raised in the proliferation of H4-II-E cells.

Regucalcin suppresses the enhancement of DNA synthesis in cell proliferation

Endogenous regucalcin has a suppressive effect on the enhancement of DNA synthesis in the nuclei of H4-II-E with cell proliferation [227]. Cells were cultured for 6–96 h in medium containing FBS (1 or 10%). Cell number was increased between 24 and 96 h; cell proliferation was markedly stimulated after culture with 10% FBS as compared with that of 1% FBS [227]. Nuclear DNA synthesis activity *in vitro* was elevated 6 h after culture with 10% FBS and its elevation was remarkable at 12 and 24 h after the culture [227]. The increase in nuclear DNA synthesis activity preceded an elevation of the number of the cloned rat hepatoma cells H4-II-E cells cultured with FBS (1 or 10%) [227].

The increase in nuclear DNA synthesis activity at 12 and 24 h after culture with FBS was inhibited in the presence of PD98059, an inhibitor of MAP kinase, staurosporine, an inhibitor of protein kinase C, and TFP, an antagonist of Ca²⁺/calmodulin-dependent protein kinase, in the reaction mixture [227]. The increase in nuclear DNA synthesis activity after serum stimulation may be partly mediated through action of various protein kinases in the nuclei of H4-II-E cells. However, serum-stimulated increase in nuclear DNA synthesis activity was not related to various protein phosphatases [227].

Regucalcin has been shown to transport in the nucleus of rat liver [37, 148], and it can inhibit nuclear DNA synthesis of normal rat liver [186, 187]. The presence of regucalcin in the reaction mixture caused a decrease in DNA synthesis activity in the nuclei of H4-II-E cells cultured with FBS [227]. This effect was not altered in the presence of various protein kinase inhibitions. Regucalcin has been demonstrated to inhibit the activity of various protein kinases in the nuclei of liver cells [185, 222]. The effect of regucalcin in decreasing nuclear DNA synthesis activity may be partly mediated through the pathway of various protein kinases in H4-II-E cells.

Nuclear DNA synthesis activity was increased in the presence of anti-regucalcin monoclonal antibody in the reaction mixture containing the nucleus of H4-II-E cells cultured for 24 h with 10% FBS [227]. This elevation was inhibited after addition of various protein kinase inhibitors in the reaction mixture [227]. These findings support the view that endogenous regucalcin suppresses DNA synthesis activity through mechanism by which it inhibits nuclear protein kinases, and that it has a suppressive effect on DNA synthesis activity in the nuclei of H4-II-E cells with proliferation.

The transcriptional activity for regucalcin gene has been shown to enhance in H4-II-E cells cultured with Bay K 8644, an agonist of Ca²⁺ entry in cells, in the presence of

10% FBS [44, 51]. Culture with Bay K 8644 caused an increase in regucalcin levels in H4-II-E cells that were cultured in the presence of FBS (1 and 10%) [227]. In this case, nuclear DNA synthesis activity was not changed after culture with Bay K 8644 [227]. The presence of anti-regucalcin monoclonal antibody in the reaction mixture containing the nuclei of H4-II-E cells cultured with Bay K 8644 resulted in an elevation of nuclear DNA synthesis activity [227], suggesting that nuclear DNA synthesis activity is suppressed through endogenous regucalcin that is increased in the nuclei of H4-II-E cells cultured with Bay K 8644.

As mentioned above, regucalcin may have a suppressive effect on the enhancement of nuclear DNA synthesis activity in proliferative cells, and it may play a suppressive role for overexpression of cell proliferation. This was further supported in H4-II-E cells overexpressing regucalcin stably [228].

The regucalcin content of regucalcin/pCXN2-transfected cells used in this study was 19.7-fold as compared with that of the parental wild-type H4-II-E cells and pCXN2 vector-transfected cells (mock type) [228]. Regucalcin/pCXN2 vector-transfected cells (transfectants) were cultured for 72 h in the presence of FBS (10%) [228]. Cell numbers and DNA synthesis activity in the transfectants were found to suppress as compared with those of wild and mock type, suggesting that overexpression of regucalcin has a suppressive effect on cell proliferation [228].

The presence of anti-regucalcin monoclonal antibody in the reaction mixture caused an increase in DNA synthesis activity in the nuclei obtained from wild-type H4-II-E cells, mock-type cells, and transfectants with overexpression of regucalcin [228]. However, the augmentation of nuclear DNA synthesis activity was remarkable in the transfectants [228]. This may support the view that endogenous regucalcin has a great suppressive effect on nuclear DNA synthesis activity.

The expression of regucalcin mRNA has been shown to stimulate through a Ca²⁺-signaling mechanism in H4-II-E cells [41, 51]. Regucalcin is translocated to the nucleus of liver cells [37, 149]. Regucalcin inhibits nuclear protein kinase and protein phosphatase activities [222, 223], which are involved in signal transduction to the nucleus, and it causes an inhibition in nuclear DNA synthesis in proliferative liver cells [227]. Regucalcin may play a suppressive role for the overexpression of proliferation of liver cells.

Regucalcin suppresses cell cycle-related genes in proliferative cells

Regucalcin has a suppressive effect on liver cell proliferation [222–224]. Whether regucalcin suppress cell cycle-related genes is examined in proliferative cells [229]. H4-II-E cells

(wild type) and stable regucalcin/pCXN2 transfectants were cultured for 72 h in a medium containing 10% FBS to obtain subconfluent monolayers. The proliferation of cells was suppressed in transfectants cultured for 24–72 h. The proliferation of wild-type cells was inhibited when the cells were cultured for 72 h in a medium containing an inhibitor of transcriptional activity or protein synthesis. Such an effect was not seen in transfectants [229]. Regucalcin has a suppressive effect on cytosolic protein synthesis [162, 163] and nuclear RNA synthesis [189, 190] in rat liver. If overexpression of regucalcin inhibits protein and RNA synthesis in transfectants, the inhibitors of transcriptional activity or protein synthesis may not have additional effect. This suggests that the effect of regucalcin in suppressing cell proliferation is partly mediated through its suppressive effect on protein and RNA synthesis in the cells.

Regucalcin has an inhibitory effect on various protein kinases in rat liver cytosol and nucleus [135, 140, 185, 222]. The proliferation of H4-II-E cells (wild type and transfectants) was inhibited in the presence of PD98059, dibucaine, staurosporine, or genistein, which is an inhibitor of various protein kinases [229]. The effect of regucalcin in suppressing cell proliferation may be partly related to its inhibitory effect on MAP kinase, Ca^{2+} /calmodulin-dependent kinase, and protein tyrosine kinase in H4-II-E cells.

Wortmannin is known to have an inhibitory effect on PI3-kinase. The proliferation of H4-II-E cells (wild type) was inhibited in the presence of wortmannin, an inhibitor of PI3-kinase, or vanadate, an inhibitor of protein tyrosine phosphatase [229]. These effects were not observed in transfectants [230]. Regucalcin may inhibit PI3-kinase and protein tyrosine phosphatase activities, and those inhibitory effects may partly contribute to the suppression of proliferation in H4-II-E cells.

Bay K 8644 is an agonist of calcium entry into cells. The proliferation of H4-II-E (wild type) was inhibited in the presence of Bay K 8644 [229]. This effect of Bay K 8644 was not seen in transfectants [229], because regucalcin has a role in the maintenance of intracellular calcium homeostasis in many cell types [Review in Ref. 230].

Overexpression of regucalcin has been found to suppress the inhibitory effect of various factors, which induce cell cycle arrest, on the proliferation of H4-II-E cells (wild type) [229]. The effect of roscovitine, a potent and selective inhibitor of the cyclin-dependent kinase cdc2, cdk2m, and cdk5 [231], or sulforaphane, which can induce G2/M phase cell cycle arrest [232], in inhibiting the proliferation of wild-type cells was not observed in transfectants [229]. Sulforaphane with a higher concentration caused a decrease in cell number of transfectants, suggesting that the chemical induces cell death and apoptosis (unpublished data). Butyrate induced an inhibition of the proliferation of

wild-type cells and transfectants. Roscovitine can arrest in G1 and accumulate in G2 of cell cycle [231]. Butyrate induces an inhibition of G1 progression [233]. The inhibitory effect of roscovitine or sulforaphane on cell proliferation may not be seen in transfectants, if regucalcin has a suppressive effect on the same pathway which roscovitine or sulforaphane has an inhibitory effect on cell proliferation. Presumably, regucalcin induces G1 and G2/M phase cell cycle arrest in H4-II-E cells.

The expression of p21 mRNA has been found to enhance in transfectants overexpressing regucalcin, although *cdc2a* and *chk2* (checkpoint-kinase 2) mRNA levels are not changed in transfectants [230]. p21 is an inhibitor of cyclin-dependent kinases (cdk) [234]. Regucalcin may enhance p21 expression and it inhibits G1 progression in H4-II-E cells. It cannot exclude the possibility, however, that regucalcin directly inhibits cdk activity in the cells.

Overexpression of regucalcin suppressed the expression of IGF-I mRNA in H4-II-E cells [229]. IGF-I is a growth factor in cell proliferation. Regucalcin may have a suppressive effect on IGF-I expression in H4-II-E cells, and this suppression of IGF-I expression leads to retardation of cell proliferation.

Regucalcin modulates tumor-related gene expression in proliferative cells

It is known that *c-myc*, *c-fos*, *c-jun*, and *Ha-ras* are tumor stimulator genes [235]. *p53* and *Rb* are tumor suppressor genes and *c-src* is oncogene [233]. The expression of *c-myc*, *Ha-ras*, or *c-src* mRNAs was found to suppress transfectants overexpressing regucalcin [179, 236]. The expression of *p53* and *Rb* mRNAs was markedly enhanced in transfectants overexpressing regucalcin [179]. Presumably, the suppression of *c-myc*, *Ha-ras*, and *c-src* mRNAs expressions and the enhancement of *p53* and *Rb* mRNAs expression in transfectants overexpressing regucalcin is partly involved in the retardation of proliferation of hepatoma H4-II-E cells.

The mechanism by which regucalcin regulates the expression of genes related to tumor is unknown. Regucalcin has been shown to translocate into the nucleus of rat liver [37, 148], and the protein has an inhibitory effect on RNA synthesis in isolated rat liver nucleus [189, 190]. Regucalcin can bind DNA and modulates nuclear transcriptional activity [179]. Regucalcin may bind to promoter region of tumor-related genes and it may suppress the expression of tumor stimulator gene or stimulate the expression of tumor suppressor gene in H4-II-E cells overexpressing regucalcin. As a result, the proliferation of hepatoma cells overexpressing regucalcin may be suppressed.

The expression of regucalcin is reduced in the cloned rat hepatoma H4-II-E cells as compared with that of normal rat liver interestingly [222], suggesting an involvement of regucalcin in the suppression of carcinogenesis. The down-regulation of regucalcin expression in liver cells may lead to stimulation of cell proliferation with alteration of various tumor-related gene expressions. Regucalcin may play an important role as suppressor in cell proliferation and tumorigenesis of liver cells.

Regucalcin suppresses cell proliferation in normal kidney cells

Regucalcin has been also shown to have a role in the regulation of the proliferation of rat kidney proximal tubular epithelial NRK52E cells [237]. The regucalcin content of regucalcin/pCXN2-transfected cells was about 21-fold as compared with that of the parental wild-type cells. The enhancement in cell proliferation was suppressed in the transfectants overexpressing regucalcin [237]. The decrease in cell number of NRK52E (wild type) cells after culture with butyrate, rescovitine, and sulforaphane, which is an inhibitor of cell cycle, was not observed in transfectants, suggesting that regucalcin induces G1 and G2/M phase cell cycle arrest in NRK52E cells [237].

The inhibition of the proliferation of NRK52E cells induced by PD98059, staurosporine, or dibucaine that is an inhibitor of various protein kinase inhibitors was not seen in the transfectants [238–240]. The suppressive effect of regucalcin on cell proliferation may result from the inhibitory effect of regucalcin on various protein kinases that are involved in stimulation of cell proliferation. The inhibition in proliferation of NRK52E cells by wortmannin, an inhibitor of PI3-kinase, was not observed in transfectants [241], suggesting that regucalcin inhibits PI3-kinase and that it partly contributes suppression of cell proliferation in NRK52E cells.

Bay K 8644 is an agonist of calcium entry into cells. The proliferation of NRK52E cells (wild type) was inhibited in the presence of Bay K 8644. Overexpression of regucalcin had a preventive effect on Bay K 8644-induced inhibition of cell proliferation [237]. Regucalcin has a role in the maintenance of intracellular calcium homeostasis in many cell types.

The effect of regucalcin on the gene expression of proteins that are related to cell proliferation and cell cycle is examined. The expression of c-jun and chk2 (checkpoint-kinase 2) mRNAs was found to suppress in the transfectants [237]. The expression of p53 mRNA was enhanced in transfectants, while the expression of c-myc, c-fos, cdc2, and p21mRNA was not changed in the transfectants [237]. The decrease in c-jun and chk2 mRNA expressions may partly contribute to suppress cell

proliferation induced in NRK52E cells overexpressing regucalcin. The expression of the tumor suppressor gene p53 mRNA, which was enhanced with overexpression of regucalcin, may have a partial role in the retardation of proliferation of NRK52E cells. Regucalcin has been shown to localize in the nucleus of NRK52E cells [242]. Regucalcin may have a suppressive effect on cell proliferation due to regulating many gene expressions that is related to cell proliferation in normal kidney NRK52E cells.

Regucalcin, moreover, has been shown to have a regulatory effect on gene expression of proteins that regulate calcium transport system in kidney cells. Overexpression of regucalcin has been found to have suppressive effects on the gene expression of L-type Ca^{2+} channel and calcium-sensing receptor (CaR), which regulate intracellular Ca^{2+} signaling in NRK52E cells [242]. Overexpression of regucalcin caused an increase in rat outer medullary K^{+} channel (ROMK) mRNA expression in NRK52E cells, while it did not have an effect on Na, K-ATPase, and epithelial sodium channel (ENaC) mRNA expressions [242]. The expression of Type II Na-Pi cotransporter (NaPi-IIa) and angiotensinogen mRNAs was not changed in NRK52E cells overexpressing regucalcin [242], suggesting that regucalcin does not have effects on NaPi-IIa and angiotensinogen mRNA expressions in kidney NRK52E cells.

The blockade of calcium influx through L-type calcium channels has been shown to attenuate mitochondrial injury and apoptosis in hypoxia renal tubular cells [243]. The entry of calcium through L-type Ca^{2+} channels induces mitochondrial disruption and cell death [243]. CaR participates in the regulation of renal Ca^{2+} transport [244]. It is speculated that regucalcin regulates intracellular Ca^{2+} -signaling pathway through its suppressive effect on L-type Ca^{2+} channel or CaR mRNA expression in the kidney proximal tubular epithelial cells.

The expression of regucalcin mRNA in NRK52E cells has been shown to enhance after the treatment of PTH [216], suggesting that regucalcin partly mediates cellular response for PTH in kidney cells. Overexpression of regucalcin did not attenuate the expression of L-type Ca^{2+} channel or CaR mRNAs, which is decreased after PTH treatment, in NRK52E cells. Regucalcin decreased L-type Ca^{2+} channel or CaR mRNA expressions in NRK52E cells. Regucalcin may partly contribute as a mediator in cellular response for stimulation of PTH in NRK52E cells.

Whether handling of calcium in NRK52E cells is changed in transfectants overexpressing regucalcin is unknown. Overexpression of regucalcin has been shown to have suppressive effects on apoptosis with culture of Bay K 8644 in NRK52E cells [217]. Presumably, the effects of regucalcin on gene expression are not mediated through

change in calcium handling in transfectants. Regucalcin has been shown to play a role in the regulation of intracellular Ca^{2+} transport; the protein activates Ca^{2+} -pumping enzymes (Ca^{2+} -ATPase) in the basolateral membranes, mitochondria, and microsomes in rat kidney cortex [98, 99, 245]. Regucalcin may regulate intracellular Ca^{2+} homeostasis in kidney proximal tubular epithelial cells so that Ca^{2+} is passed through transcellular transport. Moreover, regucalcin was found to suppress the expression of L-type Ca^{2+} channel or CaR mRNAs in NRK52E cells, supporting the view that regucalcin plays a physiological role in the regulation of intracellular Ca^{2+} homeostasis in kidney proximal tubular epithelial cells.

Overexpression of regucalcin has a suppressive effect on cell responses that are mediated through signaling process following stimulation with TNF- α or transforming growth factor- β 1 (TGF- β 1) in NRK52E cells [246]. Overexpression of regucalcin had a suppressive effect on apoptotic cell death induced by TNF- α or TGF- β 1 that is mediated through caspase-3 in NRK52E cells [246]. Culture with TNF- α or TGF- β 1 caused a remarkable increase in α -smooth muscle actin level in NRK52E cells. Such an increase was not seen in transfectants. In addition, the expression of α -smooth muscle actin was markedly suppressed in transfectants cultured without TNF- α or TGF- β 1. These findings demonstrate that overexpression of regucalcin has a suppressive effect on the expression of α -smooth muscle actin in NRK52E cells cultured with TNF- α or TGF- β 1, and that regucalcin regulates signaling pathway that is mediated through TNF- α or TGF- β 1 to stimulate the expression of α -smooth muscle actin in NRK52E cells. TGF- β 1 is a key mediator that regulates transdifferentiation of NRK52E cells into myofibroblasts expressing α -smooth muscle actin [247]. This may contribute to renal fibrosis associated with overexpression of TGF- β 1 within the diseased kidney [247]. Regucalcin may regulate transdifferentiation to renal fibrosis in NRK52E cells with TGF- β 1 or TNF- α .

Overexpression of regucalcin caused a remarkable increase in the expression of mRNA of Smad 2, which is involved in signal transduction of TGF- β 1 [248], or NF- κ B, which is related to signaling of TNF- α [249], in NRK52E cells. Such an increase was not seen in Smad 3 mRNA expression in transfectants. This finding suggests that regucalcin stimulates the gene expression of Smad 2 or NF- κ B, which is related to signaling mechanism of TNF- α or TGF- β 1. Regucalcin may have a suppressive effect on signaling pathway by which TNF- α or TGF- β 1 stimulates gene expression of NF- κ B or Smad 2 in NRK52E cells. These cytokines may not have enhancing effects on gene expression of NF- κ B or Smad 2 in transfectants. Presumably, the suppressive effects of regucalcin on apoptotic cell death and α -smooth muscle actin expression may be not

involved in the expressions of NF- κ B or Smad 2 that is stimulated by TNF- α or TGF- β 1 in NRK52E cells.

Conclusion

As mentioned above, regucalcin has been demonstrated to have a suppressive effect on hepatoma H4-II-E cells and normal kidney NRK52E cells. The suppressive effect of regucalcin on cell proliferation is related to its inhibitory effect on the activities of various protein kinases and protein phosphatases, calcium-dependent signaling factors, protein synthesis, nuclear DNA and RNA synthesis, and IGF-I expression and its activatory effect on p21, an inhibitor of cell cycle-related protein kinases. Moreover, regucalcin has been shown to suppress the expressions of *c-myc*, *Ha-ras*, and *c-src* mRNAs and to enhance the expressions of *p53* and *Rb* mRNAs, which are related to tumorigenesis of liver cells. *p53* is also known to stimulate p21 mRNA expression to induce cell cycle arrest. Regucalcin has a suppressive effect on many intracellular signaling pathways that is related to cell proliferation due to hormonal stimulation, as summarized in Fig. 3. The suppressive effect of regucalcin on cell apoptosis and cell proliferation is based on the mechanism by which the protein inhibits cellular events that are mediated through many intracellular signaling factors. Regucalcin may play an important role as suppressor protein in the maintenance of homeostasis of cellular response for cell stimulation.

Regucalcin has been demonstrated to have a suppressive effect on cell death and apoptosis and cell proliferation induced by stimulation of various factors in hepatoma H4-II-E cells and normal kidney NRK52E cells. Presumably, regucalcin plays a physiological role in maintaining cell homeostasis of cellular response for various stimulating factors. Regucalcin may be a key molecule as a suppressor protein in cell regulation.

Role of regucalcin in regenerating liver in vivo

Adult rat hepatocytes are normally quiescent in vivo. However, 20–30 h after partial hepatectomy (about 70%) they undergo a synchronous wave of DNA synthesis and cell division and continue to divide until the original mass of the liver is regenerated 3–7 days later [250]. The liver weight at 1 day after hepatectomy is increased about 50% of that of sham-operated rats, and it reached to the same levels as sham operated at 3 days after hepatectomy, indicating that the increase in cell proliferation is induced at 1 day after hepatectomy [251]. Regenerating liver is a good model of liver cell proliferation in vivo.

Hepatocyte growth factor, which can promote liver regenerating after partial hepatectomy [248], has been

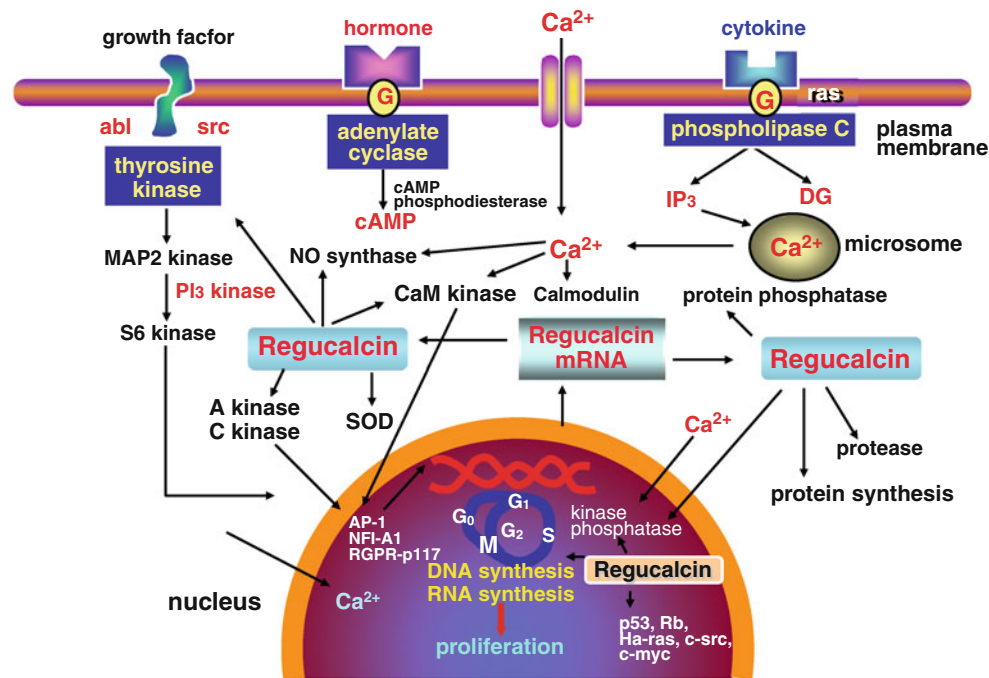


Fig. 3 Regucalcin has a suppressive effect on the enhancement of cell proliferation. Regucalcin mRNA expression is stimulated through the pathway of signaling mechanism concerning Ca^{2+} /calmodulin (CaM)-dependent protein kinase (CaM kinase), protein kinase C (C kinase), protein kinase A (A kinase), and tyrosine kinase due to hormonal stimulation. Regucalcin inhibits the activities of various protein kinases and protein phosphatases in the cytoplasm and nucleus of cells, and it also can inhibit Ca^{2+} /calmodulin-dependent enzyme activity (including cyclic AMP phosphodiesterase, NO synthase, superoxide dismutase (SOD), and others). Cytoplasmic regucalcin

translocates into nucleus. Regucalcin inhibits nuclear DNA and RNA synthesis. Regucalcin has an inhibitory effect on the expression of *c-myc*, *Ha-ras*, and *c-src* mRNAs, which are tumor stimulator genes. Regucalcin also stimulates the expression of *p53* and *Rb* mRNAs that are tumor suppressor genes. Moreover, regucalcin can inhibit protein synthesis and it can stimulate protein degradation. Regucalcin induces G1 and G2/M phase cell cycle arrest in cells. The suppressive effect of regucalcin on cell proliferation is mediated through regulating many signaling systems

shown to increase calcium concentration in rat hepatocytes [249]. Calmodulin synthesis in regenerating rat liver is increased at 8 h after hepatectomy [252].

Regenerating rat liver has been shown to enhance Ca^{2+} -ATPase activity in the nuclei between 1 and 5 days after hepatectomy [251]. Liver nuclear Ca^{2+} -ATPase is related to Ca^{2+} uptake by the nuclei and the metal accumulates nuclear matrix [81, 84, 85]. The increase in the nuclear Ca^{2+} -ATPase activity in regenerating liver caused a corresponding augmentation of the nuclear calcium content [251]. Liver calcium content was increased between 1 and 5 days after hepatectomy, although the nuclear calcium content was a slight increase at 1 day after hepatectomy [251]. Calcium, which increased in the cytoplasm of regenerating liver, may be transported into the nuclei with the enhancement of nuclear Ca^{2+} -ATPase activity.

Protein kinase C and Ca^{2+} /calmodulin-dependent protein kinase exists in liver nuclei [186]. Nuclear Ca^{2+} -ATPase activity of regenerating rat liver was decreased after the addition of protein kinase inhibitor into the enzyme reaction mixture [251]. The increase in nuclear

Ca^{2+} -ATPase activity in regenerating liver may be partly activated through Ca^{2+} -dependent kinases. DNA content in the nuclei of regenerating rat liver was increased from 1 day after hepatectomy, while the nuclear DNA fragmentation activity was decreased in regenerating liver [251]. The nuclear DNA fragmentation activity in regenerating liver was not altered in the presence of Ca^{2+} chelator (EGTA) [251]. Presumably, the increased nuclear calcium in regenerating liver is involved in the regulation of nuclear DNA synthesis rather than the activation of nuclear DNA fragmentation.

Liver regucalcin mRNA expression has been shown to enhance 1–5 days after hepatectomy as compared with that of sham-operated rats, which is at the time point during activation of liver proliferation with a longer time (5 days) [62]. The expression of regucalcin mRNA in regenerating liver has been suggested to mediate through Ca^{2+} /calmodulin in rat liver, since hepatocyte growth factor, which promotes liver regenerating after partial hepatectomy, has been shown to increase calcium concentration in rat hepatocytes [252]. The increase in regucalcin mRNA expression in regenerating liver was independent on the

decomposition of mRNA [252]. The process that transcripts to regucalcin mRNA may be stimulated in regenerating liver. The increase in regucalcin may be related to the suppression of cell proliferation in regenerating rat liver.

The role of regucalcin in the regulation of the nuclear function of regenerating rat livers is examined. Protein kinase activity was elevated in the liver nuclei obtained at 6–48 h after partial hepatectomy [185], suggesting that protein kinases, which are involved in Ca^{2+} signaling, participate in the enhancement of nuclear phosphorylation in regenerating liver with proliferative liver cells. Regucalcin was found to have a suppressive effect on the enhancement of Ca^{2+} /calmodulin-dependent protein kinase and protein kinase C activities in the nuclei of regenerating rat livers [185].

The phosphorylation in the nuclei of regenerating rat livers was increased in the presence of anti-regucalcin antibody in the reaction mixture [185]. Such stimulation was completely abolished in the presence of exogenous regucalcin [185], suggesting that endogenous regucalcin is involved in the suppression of protein kinase activity in the nuclei. The net increase in protein kinase activity after anti-regucalcin antibody addition was about two-fold in the nuclei of regenerating rat liver as compared with that of normal rat liver [185]. Endogenous regucalcin, which is enhanced in regenerating liver, may be translocated to the nucleus *in vivo*.

Protein phosphatase activity toward phosphotyrosine is also found in the liver nuclei, whereas the nuclear enzyme activity toward phosphoserine and phosphothreonine is only slight [147, 148]. Protein phosphotyrosine phosphatase activity in the cytoplasm and nuclei of liver cells was increased in regenerating liver in rats [147]. This increase was enhanced in the presence of anti-regucalcin monoclonal antibody in the enzyme reaction mixture from regenerating liver [147].

Regucalcin has an inhibitory effect on Ca^{2+} /calmodulin-dependent protein kinases, protein kinase C, and protein phosphatases [139, 140]. Presumably, regucalcin participates in the regulation of cellular function due to inhibiting processes of phosphorylation and dephosphorylation of various proteins in proliferative liver cells. This role may be important intensively in the suppression of over-proliferation of liver cells *in vivo*.

The suppressive effect of regucalcin on protein synthesis in regenerating rat liver is also shown [163]. Regenerating liver induced an increase in protein synthesis. This increase was enhanced in the presence of anti-regucalcin monoclonal antibody [163]. This elevation was completely prevented after the addition of regucalcin. Endogenous regucalcin has a suppressive effect on the enhancement of protein synthesis in regenerating rat liver with proliferative

cells [163]. Regucalcin may have a role as an inhibitor in protein synthesis in proliferative liver cells *in vivo*.

Liver nuclear DNA synthesis has been shown to stimulate in regenerating rat liver [186, 187]. Liver nuclear DNA synthesis was markedly enhanced 1 and 3 days after partial hepatectomy [186]. Regucalcin was found to have an inhibitory effect on nuclear DNA synthesis in regenerating rat liver [187]. This effect was not resulted from nuclear DNA hydrolysis, because regucalcin can inhibit Ca^{2+} activated DNA fragmentation in rat liver nuclei [187]. DNA synthesis activity in the nucleus of regenerating rat liver was enhanced in the presence of anti-regucalcin monoclonal antibody in the reaction mixture [187]. This increase was completely abolished after the addition of regucalcin [187]. The endogenous regucalcin in liver nucleus may play a suppressive role in the enhancement of nuclear DNA synthesis activity of regenerating liver.

Nuclear functions in regenerating liver may be stimulated through many signaling factors. The increase in nuclear DNA synthesis activity in regenerating rat liver was prevented in the presence of strurosporine, TFP, and PD98059, which are various protein kinase inhibitors, in the reaction mixture [187]. This suggests that the nuclear DNA synthesis activity in regenerating liver with proliferative cells is partly stimulated through signaling pathway, which is related to protein kinase C, Ca^{2+} /calmodulin-dependent protein kinase, and MAPK kinase in the nucleus. The effect of anti-regucalcin monoclonal antibody in enhancing nuclear DNA synthesis in regenerating liver was blocked in the presence of strurosporine, TFP, and PD98059 [187]. This finding suggests that the suppressive effect of regucalcin on nuclear DNA synthesis activity in regenerating liver may be partly mediated through the inhibitory action of regucalcin on various protein kinases. In addition, regucalcin may directly inhibit nuclear DNA synthesis activity in liver cells.

Regucalcin has also been shown to inhibit RNA synthesis in the nuclei isolated from control rat liver [189] and from regenerating rat liver [190]. The presence of anti-regucalcin monoclonal antibody in the reaction mixture caused a significant increase in RNA synthesis activity in the nuclei of normal rat liver [190]. This increase was completely abolished after the addition of regucalcin. The effect of anti-regucalcin monoclonal antibody in increasing nuclear RNA synthesis activity in regenerating rat liver was enhanced in the presence of the antibody in the reaction mixture [190].

The enhancement of nuclear RNA synthesis activity in regenerating rat liver is partly dependent on the activation of protein kinases and protein phosphatases, which are involved intracellular signaling pathway of hormone and growth factors [190]. The effect of anti-regucalcin monoclonal antibody in enhancing RNA synthesis activity in the

nuclei of regenerating rat liver was inhibited in the presence of α -amanitin, PD98059, staurosporine, or vanadate in the reaction mixture [190]. The enhancement of nuclear RNA synthesis activity in regenerating liver is partly suppressed through signaling pathways that endogenous regucalcin inhibits RNA polymerase II and III, MAPK kinase, protein kinase C, and protein phosphatases in the liver nucleus.

Regucalcin could inhibit RNA synthesis activity in the nuclei of normal and regenerating rat livers. Regucalcin may have a role as the transcription-related factor in liver nucleus. Whether regucalcin can directly bind to the promoter region in the gene is unknown, however.

As mentioned above, regucalcin mRNA expression in liver cells is stimulated through the pathway of signaling mechanism concerning protein kinases and protein phosphatases that are increased in regenerating liver with proliferative cells *in vivo*, and endogenous regucalcin has a suppressive effect on protein, DNA, and RNA synthesis, which is mediated through calcium signaling, in the cells. Presumably, regucalcin plays a suppressive role in the overexpression of cell proliferation *in vivo*. Regucalcin may have an important role as a suppressor protein in the differentiation and proliferation of liver cells.

Interestingly, new markers for the liver pre-neoplastic foci are investigated in rats treated with diethylnitrosamine and then 2-acetylaminofluorene combined with partial hepatectomy [253]. Transaldolase, rat aflatoxin B1 aldehyde reductase, and gamma-glutamylcysteine synthetase are found as up-regulated gene and regucalcin is found as a down-regulated gene, in line with findings for hepatocellular carcinomas [253]. The suppression of regucalcin expression may play a role in the development of carcinogenesis in liver cells. If the chemical feeding-induced suppression of regucalcin expression influences on Ca^{2+} -signaling regulation in liver cells, it may generate a possible circumstance for carcinogenesis. In this aspect, the pathophysiological role of regucalcin in carcinogenesis with chemical feeding may be significant *in vivo*.

Role of endogenous regucalcin in animals with transgene

The role of endogenous regucalcin in cellular regulation is shown using regucalcin transgenic (TG) rats *in vivo*. Regucalcin homozygote male and female rats express a prominent regucalcin protein in the tissues [254]. The expression of regucalcin was remarkable in many tissues of TG female rats: its expression of male rats was seen in the liver, kidney cortex, heart, lung, and stomach [254].

Exogenous regucalcin has been shown to have the regulatory effect on liver nuclear function *in vitro* [185–188]. Regucalcin levels were increased in the liver nuclei of

regucalcin TG male and female rats [255]. The effect of endogenous regucalcin in suppressing protein tyrosine phosphatase and RNA synthesis activities, which are inhibited by exogenous regucalcin in normal rat liver nucleus *in vitro* [256], is also demonstrated in regucalcin TG rats *in vivo* using anti-regucalcin monoclonal antibody. Nuclear protein tyrosine phosphatase activity was elevated in the presence of anti-regucalcin monoclonal antibody in the reaction mixture containing liver nuclear protein obtained from normal (wild type) rats [256]. The effect of the antibody was not seen in the liver nuclei of regucalcin TG rats [256]. Moreover, RNA synthesis was suppressed in the liver nuclei of regucalcin TG rats as compared with that of normal rats [256]. The effect of anti-regucalcin monoclonal antibody in increasing RNA synthesis was blocked in the liver nuclei of the TG rats [256]. The enhancement of endogenous regucalcin expression plays an important role in the suppression of RNA synthesis in the liver nucleus *in vivo*, supporting the view that regucalcin may have a role as a negative transcriptional factor in liver nucleus. Endogenous regucalcin plays a suppressive role in the regulation of liver nuclear function in rats *in vivo*.

Regucalcin has been shown to inhibit protein phosphatase activity in the cytoplasm [145] and nucleus [148, 149] in rat liver cells. Regucalcin has been found to have a suppressive effect on protein tyrosine phosphatase activity in rat liver microsomes in the range of its physiological concentration of liver cytoplasm [257]. The increase in endogenous regucalcin has been also shown to inhibit liver microsomal protein tyrosine phosphatase activity using regucalcin TG rats [257].

Protein tyrosine phosphatases have been postulated to play a key role in the regulation of the insulin action pathway. The inhibition of protein tyrosine phosphatase activity mimics insulin action [258]. It has been shown that the increase in protein tyrosine phosphatase activity in skeletal muscle, adipocytes, and liver is associated with insulin resistance and diabetic state [255, 259, 260]. The expression of regucalcin mRNA in rat liver has been shown to enhance after feeding and insulin treatment [59]. Regucalcin, which can inhibit protein tyrosine phosphatase activity in rat liver microsomes, may have a pathophysiological role in hepatic insulin resistance. Overexpression of regucalcin has been demonstrated to enhance glucose utilization and lipid production in the cloned rat hepatoma H4-II-E cells *in vitro* and it is involved in insulin resistance [261], and it suppresses the enhancing effect of insulin or glucose on the gene expression of insulin signaling-related proteins in the H4-II-E cells [262].

Regucalcin has been found to inhibit NO synthase activity in the cytosols of liver [124], kidney cortex [125], heart [126], and brain [127]. NO may act as a messenger or modulator molecule in many cells [122]. NO synthase is

activated by Ca^{2+} /calmodulin which is related to Ca^{2+} signaling. Overproduction of NO may lead to the damage of many cells. Regucalcin may have a suppressive effect on overproduction of NO in the cells, and the protein may have a protective effect on NO-induced damage of cells. Overexpression of regucalcin in TG rats has a suppressive effect on NO synthase activity in the cytosol of liver [124], kidney cortex [125], heart [126], and brain [128]. The presence of anti-regucalcin monoclonal antibody in the reaction mixture containing the cytosolic protein of various tissues obtained from regucalcin TG rats caused an increase in NO synthetase activity, and this increase was completely abolished after the addition of regucalcin. This finding supports the view that endogenous regucalcin has a suppressive effect on NO synthase activity *in vivo*.

The role of endogenous regucalcin in the regulation of Ca^{2+} -ATPase activity in the mitochondria of brain tissues is examined using regucalcin TG rats [116]. The addition of regucalcin, which is a physiological concentration in rat brain tissues, into the enzyme reaction mixture caused a significant increase in Ca^{2+} -ATPase activity, while it did not have an effect on Mg^{2+} -ATPase activity [116]. Regucalcin was increased in the brain tissues or the mitochondria of regucalcin TG rats [116]. The mitochondrial Ca^{2+} -ATPase activity was increased in the TG rats as compared with that of wild-type rats. Thus, endogenous regucalcin has an enhancing effect on Ca^{2+} -ATPase activity in the mitochondria of rat brain tissues *in vivo*.

Thus, the increase in endogenous regucalcin has been demonstrated to have a suppressive effect on many functions in various tissues, which the effect of exogenous regucalcin are found *in vitro*, using regucalcin TG rats *in vivo*.

Interestingly, osteoporosis and hyperlipidemia are induced in regucalcin TG rats [reviewed in Ref. 263], indicating a pathophysiological role of regucalcin. This may be resulted from the regulatory effect of regucalcin on cell signaling systems in cells that are related to bone cells [264–267] and liver cells [261, 262].

One party, regucalcin (SMP30) deficiency in mice causes an accumulation of neutral lipids and phospholipids in the liver and shortens the life span [268]. Regucalcin (SMP30) knockout mouse liver is highly susceptible to TNF- α - and Fas-mediated apoptosis [269], supporting the view that regucalcin has a suppressive effect on cell apoptosis in cell culture system [201, 215]. The deficiency of regucalcin (SMP30) induces a decrease in L-ascorbic acid (vitamin C) in mice [270], although this may not be significant because of vitamin C is not synthesized in human.

β -Catenin loss and regucalcin decrease have been shown to be contributing to apoptosis [271]. β -Catenin-null hepatocytes or regucalcin small interfering RNA-transfected HepG2 cells were cultured, and these cells exhibited

significant apoptosis that was alleviated by the addition of ascorbic acid, which in turn may be one of the mechanisms contributing to the role of β -catenin in cell survival through regucalcin expression [271].

The exploration of other roles of regucalcin *in vivo* is expected.

Prospects

Regucalcin, which its gene is localized on the chromosome X in human and rats, is thought as a protein that is highly differentiated because of a great conservation of the regucalcin genes throughout evolution in vertebrates species. Regucalcin plays a multifunctional role in cellular regulation; a role in keeping intracellular Ca^{2+} homeostasis, an inhibitory role on various Ca^{2+} -dependent enzyme activations, protein kinases and protein phosphatases. Regucalcin regulates due to suppressing protein synthesis and stimulating protein degradation, and it has a suppressive effect on DNA and RNA synthesis that are mediated through various signaling systems in the nucleus. Regucalcin suppresses an enhancement of cell proliferation and apoptotic cell death that are induced by various signaling factors. Regucalcin has been proposed to play a pivotal role as a suppressor protein of intracellular signaling system in maintaining cell homeostasis. Regucalcin is the first time finding in protein molecule that has a role as a suppressor protein in cell signaling, although it is well known that many proteins enhance cell signal transduction so far.

There are growing evidences that regucalcin may be a key molecule in metabolic disease. The overexpression of regucalcin gene has been known to induce osteoporosis and hyperlipidemia in the transgenic rats and the deficiency of regucalcin (SMP30) induces a decrease in vitamin C (it is not synthesized in human), which is related to regulation of oxidative stress in mice and to inducing of cell apoptosis. The expression of regucalcin gene is downregulated in the development of carcinogenesis in liver cells, suggesting that its suppression induces promotion of tumor cells.

The gene therapy, that targets the regucalcin gene, may be useful as a therapeutic tool for disease with the attenuation of regucalcin gene expression. Development of drug, which modulates regucalcin molecule, may have a clinical significance in the restoration of metabolic disorder that is implicated to regucalcin. Clinical studies of regucalcin for disease are expected.

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References

- Cheung WY (1980) Calmodulin plays a pivotal role in cellular regulation. *Science* 202:19–27
- Nishizuka Y (1986) Studies and perspectives of protein kinase C. *Science* 233:305–331
- Williamson JR, Cooper RK, Hoek JB (1981) Role of calcium in the hormonal regulation of liver metabolism. *Biochim Biophys Acta* 639:243–295
- Kraus-Friedman N, Feng L (1996) The role of intracellular Ca^{2+} in the regulation of gluconeogenesis. *Metabolism* 48:389–403
- Yamaguchi M, Takei Y, Yamamoto Y (1975) Effect of thyrocalcitonin on calcium concentration in liver of intact and thyro-parathyroidectomized rats. *Endocrinology* 96:1004–1008
- Yamaguchi M, Yoshida H (1985) Participation of Ca^{2+} channel in liver calcium regulation by calcitonin in rats. *Acta Endocrinol* 110:239–243
- Yamaguchi M, Yamamoto T (1978) Purification of calcium binding substance from soluble fraction of normal rat liver. *Chem Pharm Bull* 26:1915–1918
- Yamaguchi M, Sugii K (1981) Properties of calcium-binding protein isolated from the soluble fraction of normal rat liver. *Chem Pharm Bull* 29:567–570
- Yamaguchi M (1988) Physicochemical properties of calcium-binding protein isolated from rat liver cytosol: Ca^{2+} -induced conformational changes. *Chem Pharm Bull* 36:286–290
- Yamaguchi M, Yoshida H (1985) Regulatory effect of calcium-binding protein isolated from rat liver cytosol on activation of fructose 1,6-diphosphatase by Ca^{2+} -calmodulin. *Chem Pharm Bull* 33:4489–4493
- Yamaguchi M, Mori S, Suketa Y (1989) Effects of Ca^{2+} and V^{5+} on glucose-6-phosphatase activity in rat liver microsomes: the Ca^{2+} effect is reserved by regucalcin. *Chem Pharm Bull* 37:388–390
- Yamaguchi M, Shibano H (1987) Calcium-binding protein isolated from rat liver cytosol reverses activation of pyruvate kinase by Ca^{2+} . *Chem Pharm Bull* 35:2025–2029
- Yamaguchi M, Shibano H (1987) Effect of calcium-binding protein on the activation of phosphorylase *a* in rat hepatic particulate glycogen by Ca^{2+} . *Chem Pharm Bull* 35:2581–2584
- Yamaguchi M, Shibano H (1987) Reversible effect of calcium-binding protein on the Ca^{2+} -induced activation of succinate dehydrogenase in rat liver mitochondria. *Chem Pharm Bull* 35:3766–3770
- Yamaguchi M, Mori S (1988) Effect of Ca^{2+} and Zn^{2+} on 5'-nucleotidase activity in rat liver plasma membranes: hepatic calcium-binding protein (regucalcin) reverses the Ca^{2+} effect. *Chem Pharm Bull* 36:321–325
- Shimokawa N, Yamaguchi M (1993) Molecular cloning and sequencing of the cDNA coding for a calcium-binding protein regucalcin from rat liver. *FEBS Lett* 327:251–255
- Shimokawa N, Isogai M, Yamaguchi M (1995) Specific species and tissue differences for the gene expression of calcium-binding protein regucalcin. *Mol Cell Biochem* 143:67–71
- Misawa H, Yamaguchi M (2000) Transcript heterogeneity of the human gene for Ca^{2+} -binding protein regucalcin. *Int J Mol Med* 5:283–287
- Murata T, Yamaguchi M (1997) Molecular cloning of the cDNA coding for regucalcin and its mRNA expression in mouse liver: the expression is stimulated by calcium administration. *Mol Cell Biochem* 173:127–133
- Misawa H, Yamaguchi M (2000) The gene of Ca^{2+} -binding protein regucalcin is highly conserved in vertebrate species. *Int J Mol Med* 6:191–196
- Yamaguchi M (2011) The transcriptional regulation of regucalcin gene expression. *Mol Cell Biochem* 346:147–171
- Shimokawa N, Matsuda Y, Yamaguchi M (1995) Genomic cloning and chromosomal assignment of rat regucalcin gene. *Mol Cell Biochem* 151:157–163
- Thiselton DL, McDowall J, Brandau O, Ramser J, d'Esposito F, Bhattacharya SS, Ross MT, Hardcastle AJ, Meindl A (2002) An integrated, functionally annotated gene map of the DXS8026-ELK1 interval on human Xp11.3-Xp11.23: potential hotspot for neurogenetic disorders. *Genomics* 79:560–572
- Shimokawa N, Yamaguchi M (1992) Calcium administration stimulates the expression of calcium-binding protein regucalcin mRNA in rat liver. *FEBS Lett* 305:151–154
- Yamaguchi M, Isogai M, Kato S, Mori S (1991) Immunohistochemical demonstration of calcium-binding protein regucalcin in the tissues of rats: the protein localizes in liver and brain. *Chem Pharm Bull* 36:1601–1603
- Yamaguchi M, Isogai M (1993) Tissue concentration of calcium-binding protein regucalcin in rats by enzyme-linked immunoadsorbent assay. *Mol Cell Biochem* 122:65–68
- Yamaguchi M, Nakajima R (2002) Role of regucalcin as an activator of sarcoplasmic reticulum Ca^{2+} -ATPase activity in rat heart muscle. *J Cell Biochem* 86:184–193
- Yamaguchi M, Hamano T, Misawa H (2000) Expression of Ca^{2+} -binding protein regucalcin in rat brain neurons: inhibitory effect on protein phosphatase activity. *Brain Res Bull* 52:343–348
- Yamaguchi M, Misawa Y, Uchiyama S, Morooka Y, Tsurusaki Y (2002) Role of endogenous regucalcin in bone metabolism: bone loss is induced in regucalcin transgenic rats. *Int J Mol Med* 10:377–383
- Yamaguchi M (1992) A novel Ca^{2+} -binding protein regucalcin and calcium inhibition. Regulatory role in liver cell function. In: Kohama K (ed) Calcium inhibition. Japan Science Society Press and CRC Press, Tokyo, Boca Raton, pp 19–41
- Yamaguchi M (1998) Role of calcium-binding protein regucalcin in regenerating rat liver. *J Gastroenterol Hepatol* 13(Suppl):S106–S112
- Yamaguchi M (2000) Role of regucalcin in calcium signaling. *Life Sci* 66:1769–1780
- Yamaguchi M (2000) The role of regucalcin in nuclear regulation of regenerating liver. *Biochem Biophys Res Commun* 276:1–6
- Yamaguchi M (2002) Impact of aging on calcium channels and pumps. In: Mattson MP (ed) Calcium homeostasis and signaling in aging. Elsevier, Amsterdam, pp 47–65
- Fujita T, Uchida K, Maruyama N (1992) Purification of senescence marker protein-30 (SMP30) and its androgen-independent decrease with age in the rat liver. *Biochim Biophys Acta* 1116:122–128
- Fujita T, Shirasawa T, Uchida K, Maruyama N (1992) Isolation of cDNA clone encoding rat senescence marker protein-30 (SMP30) and its tissue distribution. *Biochim Biophys Acta* 1132:297–305
- Tsurusaki Y, Misawa H, Yamaguchi M (2000) Translocation of regucalcin to rat liver nucleus: involvement of nuclear protein kinase and protein phosphatase regulation. *Int J Mol Med* 6:655–660
- Chakraborti S, Bahnson BJ (2010) Crystal structure of human senescence marker protein 30: insights linking structural, enzymatic, and physiological functions. *Biochemistry* 49:3436–3444
- Yamaguchi M, Makino R, Shimokawa N (1996) The 5' end sequences and exon organization in rat regucalcin gene. *Mol Cell Biochem* 165:145–150

40. Murata T, Yamaguchi M (1998) Tissue-specific binding of nuclear factors to the 5'-flanking region of the rat gene for calcium-binding protein regucalcin. *Mol Cell Biochem* 178:305–310
41. Murata T, Yamaguchi M (1998) Ca²⁺ administration stimulates the binding of AP-1 factor to the 5'-flanking region of the rat gene for the Ca²⁺-binding protein regucalcin. *Biochem J* 329:157–163
42. Murata T, Yamaguchi M (1999) Promoter characterization of the rat gene for Ca²⁺-binding protein regucalcin. Transcriptional regulation by signaling factors. *J Biol Chem* 274:1277–1285
43. Misawa H, Yamaguchi M (2000) Involvement of hepatic nuclear factor I binding motif in transcriptional regulation of Ca²⁺-binding protein regucalcin gene. *Biochem Biophys Res Commun* 269:270–278
44. Misawa H, Yamaguchi M (2000) Intracellular signaling factors-enhanced hepatic nuclear protein binding to TTGGC sequence in the rat regucalcin gene promoter: involvement of protein phosphorylation. *Biochem Biophys Res Commun* 279:275–281
45. Misawa H, Yamaguchi M (2002) Identification of transcription factor in the promoter region of rat regucalcin gene: binding of nuclear factor I-A1 to TTGGC motif. *J Cell Biochem* 84:795–802
46. Misawa H, Yamaguchi M (2001) Molecular cloning and sequencing of the cDNA coding for a novel regucalcin gene promoter region-related protein in rat, mouse and human liver. *Int J Mol Med* 8:513–520
47. Misawa H, Yamaguchi M (2002) Gene expression for a novel protein RGPR-p117 in various species: the stimulation by intracellular signaling factors. *J Cell Biochem* 87:188–193
48. Yamaguchi M, Misawa H, Ma ZJ (2003) Novel protein RGPR-p117: the gene expression in physiologic state and the binding activity to regucalcin gene promoter region in rat liver. *J Cell Biochem* 88:1092–1100
49. Murata T, Shinya N, Yamaguchi M (1997) Expression of calcium-binding protein regucalcin mRNA in the cloned human hepatoma cells (Hep G2): stimulation by insulin. *Mol Cell Biochem* 175:163–168
50. Nakajima M, Murata T, Yamaguchi M (1999) Expression of calcium-binding protein regucalcin mRNA in the cloned rat hepatoma cells (H4-II-E) is stimulated through Ca²⁺ signaling factors: involvement of protein kinase C. *Mol Cell Biochem* 198:101–107
51. Yamaguchi M, Nakajima M (1999) Involvement of intracellular signaling factors in the serum-enhanced Ca²⁺ binding protein regucalcin mRNA expression in the cloned rat hepatoma cells (H4-II-E). *J Cell Biochem* 74:81–89
52. Vinson CR, Sigler PB, McKnight SL (1989) Scissors-grip model for DNA recognition by a family of leucine zipper proteins. *Science* 246:911–916
53. O'Shea EK, Rutkowski R, Stafford WF III, Kim PS (1989) Preferential heterodimer formation by isolated leucine zippers from fos and jun. *Science* 245:646–648
54. Shimokawa N, Yamaguchi M (1993) Expression of hepatic calcium-binding protein regucalcin mRNA is mediated through Ca²⁺/calmodulin in rat liver. *FEBS Lett* 316:79–84
55. Isogai M, Yamaguchi M (1995) Calcium administration increases calcium-binding protein regucalcin concentration in the liver of rats. *Mol Cell Biochem* 143:53–58
56. Yamaguchi M, Kurota H (1995) Expression of calcium-binding protein regucalcin mRNA in the kidney cortex of rats: the stimulation by calcium administration. *Mol Cell Biochem* 146:71–77
57. Yamaguchi M, Ueoka S (1998) Expression of calcium-binding protein regucalcin mRNA in fetal rat liver is stimulated by calcium administration. *Mol Cell Biochem* 178:283–287
58. Yamaguchi M, Kanayama Y, Shimokawa N (1994) Expression of calcium-binding protein regucalcin mRNA in rat liver is stimulated by calcitonin: the hormonal effect is mediated through calcium. *Mol Cell Biochem* 136:43–48
59. Yamaguchi M, Oishi K, Isogai M (1995) Expression of hepatic calcium-binding protein regucalcin mRNA is elevated by refeeding of fasted rats: involvement of glucose, insulin and calcium as stimulating factors. *Mol Cell Biochem* 142:35–41
60. Yamaguchi M, Oishi K (1995) 17 β -Estradiol stimulates the expression of hepatic calcium-binding protein regucalcin mRNA in rats. *Mol Cell Biochem* 143:137–141
61. Yamaguchi M, Kanayama Y (1995) Enhanced expression of calcium-binding protein regucalcin mRNA in regenerating rat liver. *J Cell Biochem* 57:185–190
62. Ueoka S, Yamaguchi M (1998) Sexual difference of hepatic calcium-binding protein regucalcin mRNA expression in rats with different ages: effect of ovarian hormone. *Biol Pharm Bull* 21:405–407
63. Lotersztajn S, Hanoune J, Pecker F (1981) A high affinity calcium-stimulated magnesium-dependent ATPase in rat liver plasma membranes. Dependence on an endogenous protein activator distinct from calmodulin. *J Biol Chem* 256:11209–11215
64. Chen K-M, Junger KD (1983) Calcium transport and phosphorylated intermediate of (Ca²⁺-Mg²⁺)-ATPase in plasma membranes of rat liver. *J Biol Chem* 258:4404–4410
65. Yamaguchi M, Mori S, Kato S (1988) Calcium-binding protein regucalcin is an activator of (Ca²⁺-Mg²⁺)-adenosine triphosphatase in the plasma membranes of rat liver. *Chem Pharm Bull* 36:3532–3539
66. Takahashi H, Yamaguchi M (1993) Regulatory effect of regucalcin on (Ca²⁺-Mg²⁺)-ATPase in rat liver plasma membranes: comparison with the activation by Mn²⁺ and Co²⁺. *Mol Cell Biochem* 124:169–174
67. Takahashi H, Yamaguchi M (1994) Activatory effect of regucalcin on (Ca²⁺-Mg²⁺)-ATPase in rat liver plasma membranes: relation to sulfhydryl group. *Mol Cell Biochem* 136:71–76
68. Takahashi H, Yamaguchi M (1997) Stimulatory effect of regucalcin on ATP-dependent calcium transport in rat liver plasma membranes. *Mol Cell Biochem* 168:149–153
69. Takahashi H, Yamaguchi M (1993) Regucalcin modulates hormonal effect on (Ca²⁺-Mg²⁺)-ATPase activity in rat liver plasma membranes. *Mol Cell Biochem* 125:171–177
70. Takahashi H, Suzuki S, Yamaguchi M (1995) Stimulatory effect of hormonal signaling factors on (Ca²⁺-Mg²⁺)-ATPase activity in rat liver plasma membranes: cross talk with regucalcin. *Mol Cell Biochem* 151:1–7
71. Takahashi H, Yamaguchi M (1995) Increase of (Ca²⁺-Mg²⁺)-ATPase activity in hepatic plasma membranes of rats administered orally calcium: the endogenous role of regucalcin. *Mol Cell Biochem* 144:1–6
72. Takahashi H, Yamaguchi M (1996) Enhancement of plasma membrane (Ca²⁺-Mg²⁺)-ATPase activity in regenerating rat liver: involvement of endogenous activating protein regucalcin. *Mol Cell Biochem* 162:133–138
73. Takahashi H, Yamaguchi M (1996) Activatory effect of regucalcin on hepatic plasma membrane (Ca²⁺-Mg²⁺)-ATPase is impaired by liver injury with carbon tetrachloride administration in rats. *Mol Cell Biochem* 158:9–16
74. Yamaguchi M (1985) Mitochondrial uptake of ⁴⁵Ca²⁺ bound to calcium-binding protein isolated from rat liver cytosol. *Chem Pharm Bull* 33:3390–3394
75. Mori S, Yamaguchi M (1991) Calcium-binding protein regucalcin stimulates the uptake of Ca²⁺ by rat liver mitochondria. *Chem Pharm Bull* 39:224–226

76. Takahashi H, Yamaguchi M (2000) Stimulatory effect of regucalcin on ATP-dependent Ca^{2+} uptake activity in rat liver mitochondria. *J Cell Biochem* 78:121–130
77. Yamaguchi M, Mori S (1989) Activation of hepatic microsomal Ca^{2+} -adenosine triphosphatase by calcium-binding protein regucalcin. *Chem Pharm Bull* 37:1031–1034
78. Takahashi H, Yamaguchi M (1999) Role of regucalcin as an activator of Ca^{2+} -ATPase activity in rat liver microsomes. *J Cell Biochem* 74:663–669
79. Yamaguchi M, Mori S (1989) Effect of the calcium-binding protein regucalcin on the Ca^{2+} transport system in rat liver microsomes: the protein stimulates Ca^{2+} release. *Chem Pharm Bull* 37:3037–3041
80. Yamaguchi M, Mori S (1990) Regucalcin-induced Ca^{2+} release from rat liver microsomes: the effect is inhibited by heparin. *Chem Pharm Bull* 38:2305–2307
81. Nicotera P, McConkey DJ, Jones DP, Orrenius S (1989) ATP stimulates Ca^{2+} uptake and increases the free Ca^{2+} concentration in isolated rat liver nuclei. *Proc Natl Acad Sci USA* 86:453–457
82. Bachs O, Carafolli E (1995) Calmodulin and calmodulin-binding proteins in liver cell nuclei. *J Biol Chem* 262:10786–10790
83. Csermely P, Schnaider T, Szanto I (1995) Signalling and transport through the nuclear membrane. *Biochim Biophys Acta* 1241:425–452
84. Tsurusaki Y, Yamaguchi M (2000) Role of endogenous regucalcin in the regulation of Ca^{2+} -ATPase activity in rat liver nuclei. *J Cell Biochem* 78:541–549
85. Yamaguchi M (1992) Effect of calcium-binding protein regucalcin on Ca^{2+} transport system in rat liver nuclei: stimulation of Ca^{2+} release. *Mol Cell Biochem* 113:63–70
86. Fujita T, Inoue H, Kitamura T, Sato N, Shimosawa T, Maruyama N (1998) Senescence marker protein-30 (SMP30) rescues cell death by enhancing plasma membrane Ca^{2+} -pumping activity in Hep G2 cells. *Biochem Biophys Res Commun* 250:374–380
87. Van Os CH (1985) Transcellular calcium transport in intestinal and renal epithelial cells. *Biochim Biophys Acta* 906:195–222
88. Taylor CW (1985) Calcium regulation in vertebrates: an overview. *Comp Biochem Physiol* 82A:249–255
89. Murata T, Yamaguchi M (1999) Binding of kidney nuclear proteins to the 5'-flanking region of the rat gene for Ca^{2+} -binding protein regucalcin: involvement of Ca^{2+} /calmodulin signaling. *Mol Cell Biochem* 199:35–40
90. Misawa H, Yamaguchi M (2001) Involvement of nuclear factor-1 (NF1) binding motif in the regucalcin gene expression of rat kidney cortex: the expression is suppressed by cisplatin administration. *Mol Cell Biochem* 219:29–37
91. Kurota H, Yamaguchi M (1996) Steroid hormonal regulation of calcium-binding protein regucalcin mRNA expression in the kidney cortex of rats. *Mol Cell Biochem* 155:105–111
92. Shinya N, Kurota H, Yamaguchi M (1996) Calcium-binding protein regucalcin mRNA expression in the kidney cortex is suppressed by saline ingestion in rats. *Mol Cell Biochem* 162:139–144
93. Shinya N, Yamaguchi M (1997) Alteration in Ca^{2+} -ATPase activity and calcium-binding protein regucalcin mRNA expression in the kidney cortex of rats with saline ingestion. *Mol Cell Biochem* 170:17–22
94. Shinya N, Yamaguchi M (1998) Stimulatory effect of calcium administration on regucalcin mRNA expression is attenuated in the kidney cortex of rats ingested with saline. *Mol Cell Biochem* 178:275–281
95. Kurota H, Yamaguchi M (1995) Suppressed expression of calcium-binding protein regucalcin mRNA in the renal cortex of rats with chemically induced kidney damage. *Mol Cell Biochem* 151:55–60
96. Agus ZS, Chiu PJS, Goldberg M (1997) Regulation of urinary calcium-excretion in the rat. *Am J Physiol* 232:F545–F549
97. Kennedy MB, Bennett MK, Erondy NE, Miller SG (1987) Calcium/calmodulin-dependent protein kinases. In: Cheung WY (ed) Calcium and cell function. Academic Press Inc, New York, pp 61–107
98. Kurota H, Yamaguchi M (1997) Activatory effect of calcium-binding protein regucalcin on ATP-dependent calcium transport in the basolateral membranes of rat kidney cortex. *Mol Cell Biochem* 169:149–156
99. Kurota H, Yamaguchi M (1997) Regucalcin increases Ca^{2+} -ATPase activity and ATP-dependent calcium uptake in the microsomes of rat kidney cortex. *Mol Cell Biochem* 177:201–207
100. Langer GA (1992) Calcium and the heart: exchange at the tissue, cell, and organelle levels. *FASEB J* 6:893–902
101. Akhter T, Nakagawa T, Kobayashi A, Yamaguchi M (2007) Suppression of regucalcin mRNA expression in the hearts of rats administered with free radical compound: the administration-induced death is accelerated in regucalcin transgenic rats. *Int J Mol Med* 19:653–658
102. Thastrup O, Culler PJ, Drbbak BK, Hanley MR, Dawson AP (1990) Thapsigargin, a tumor promoter, discharges intracellular Ca^{2+} stores by specific inhibition of the endoplasmic reticulum Ca^{2+} -ATPase. *Proc Natl Acad Sci USA* 87:2466–2470
103. Tada M, Kadoma M (1989) Regulation of the Ca^{2+} pump ATPase by cAMP-dependent phosphorylation of phospholamban. *Bioessays* 10:157–163
104. Akhter T, Sawada N, Yamaguchi M (2006) Regucalcin increases Ca^{2+} -ATPase activity in the heart mitochondria of normal and regucalcin transgenic rats. *Int J Mol Med* 18:171–176
105. Dahan D, Spanier R, Rahaaminoff H (1991) The modulation of rat brain Na^{+} - Ca^{2+} exchange by K^{+} . *J Biol Chem* 266:2067–2075
106. MacDermott AB, Dale N (1986) Receptors, ion channels and synaptic potential underlying the integrative actions of excitatory amino acids. *Trends Neurosci* 10:280–284
107. Cambray-Deakin MA, Burgoyne RD (1992) Intracellular Ca^{2+} and N-methyl-D-aspartate-stimulated neuriteogenesis in rat cerebellar granule cell cultures. *Dev Brain Res* 66:25–32
108. Treves S, De Mattei M, Lanfred M, Villa A, Green NM, MacLennan DH, Meldolesi J, Pozzan T (1990) Calreticulin is a candidate for a calsequestrin-like function in Ca^{2+} -storage compartment (calciosomes) of liver and brain. *Biochem J* 271:473–480
109. Hartman H, Eckert A, Muller WE (1994) Disturbances of the neuronal calcium-homeostasis in the aging nervous system. *Life Sci* 55:2011–2018
110. Yamaguchi M, Hanahisa Y, Murata T (1999) Expression of calcium-binding protein regucalcin and microsomal Ca^{2+} -ATPase regulation in rat brain: attenuation with increasing age. *Mol Cell Biochem* 200:43–49
111. Hanahisa Y, Yamaguchi M (1996) Characterization of calcium accumulation in the brain of rats administered orally calcium: the significance of energy-dependent mechanism. *Mol Cell Biochem* 158:1–7
112. Hanahisa Y, Yamaguchi M (1997) Increase in calcium content and Ca^{2+} -ATPase activity in the brain of fasted rats: comparison with different ages. *Mol Cell Biochem* 171:127–132
113. Hanahisa Y, Yamaguchi M (2001) Decrease in Ca^{2+} -ATPase activity in the brain plasma membrane of rats with increasing age: involvement of brain calcium accumulation. *Int J Mol Med* 7:407–411

114. Hanahisa Y, Yamaguchi M (1999) Brain microsomal calcium accumulation in rats with increasing age: involvement of thapsigargin sensitive Ca^{2+} -ATPase. *Int J Mol Med* 4:627–631
115. Hanahisa Y, Yamaguchi M (1998) Increase of Ca^{2+} -ATPase activity in the brain microsomes of rats with increasing ages: involvement of protein kinase C. *Brain Res Bull* 46:329–332
116. Yamaguchi M, Takakura Y, Nakagawa T (2008) Regucalcin increases Ca^{2+} -ATPase activity in the mitochondria of brain tissues of normal and transgenic rats. *J Cell Biochem* 104:795–804
117. Yamaguchi M, Sakurai T (1992) Reversible effect of calcium-binding protein regucalcin on the Ca^{2+} -induced inhibition of deoxyuridine 5'-triphosphatase activity in rat liver cytosol. *Mol Cell Biochem* 110:25–29
118. Hanahisa Y, Yamaguchi M (1999) Effect of calcium-binding protein on adenosine 5'-triphosphatase activity in the brain cytosol of rats of different ages: the inhibitory role of regucalcin. *Biol Pharm Bull* 22:313–316
119. Rasmussen J (1970) Cell communication, calcium ion, and cyclic adenosine monophosphate. *Science* 170:404–412
120. Yamaguchi M, Tai H (1991) Inhibitory effect of calcium-binding protein regucalcin on Ca^{2+} /calmodulin-dependent cyclic nucleotide phosphodiesterase activity in rat liver cytosol. *Mol Cell Biochem* 106:25–30
121. Yamaguchi M, Kurota H (1997) Inhibitory effect of regucalcin on Ca^{2+} /calmodulin-dependent cyclic AMP phosphodiesterase activity in rat kidney cytosol. *Mol Cell Biochem* 177:209–214
122. Lowenstein CJ, Dinerman JL, Snyder SH (1994) Nitric oxide: a physiologic messenger. *Ann Intern Med* 120:227–237
123. Izumi T, Tsurusaki Y, Yamaguchi M (2003) Suppressive effect of endogenous regucalcin on nitric oxide synthase activity in cloned rat hepatoma H4-II-E cells overexpressing regucalcin. *J Cell Biochem* 89:800–807
124. Yamaguchi M, Takahashi H, Tsurusaki Y (2003) Suppressive role of endogenous regucalcin in the enhancement of nitric oxide synthase activity in liver cytosol of normal and regucalcin transgenic rats. *J Cell Biochem* 88:1226–1234
125. Ma ZJ, Yamaguchi M (2003) Regulatory effect of regucalcin on nitric oxide synthase activity in rat kidney cortex cytosol: role of endogenous regucalcin in transgenic rats. *Int J Mol Med* 12:201–206
126. Ma ZJ, Yamaguchi M (2002) Suppressive role of endogenous regucalcin in the regulation of nitric oxide synthase activity in heart muscle cytosol of normal and regucalcin transgenic rats. *Int J Mol Med* 10:761–766
127. Tobisawa M, Yamaguchi M (2003) Inhibitory role of regucalcin in the regulation of nitric oxide synthase activity in rat brain cytosol: involvement of aging. *J Neurol Sci* 209:47–54
128. Tobisawa M, Yamaguchi M (2003) Role of endogenous regucalcin in brain function: suppression of cytosolic nitric oxide synthase and nuclear protein tyrosine phosphatase activities in brain tissue of transgenic rats. *Int J Mol Med* 12:581–585
129. Van Remmen H, Williams MD, Guo Z, Estlack L, Yang H, Carlson EJ, Epstein CJ, Huang TT, Richardson A (2001) Knockout mice heterozygous for Sod2 show alterations in cardiac mitochondrial function and apoptosis. *Am J Physiol Heart Circ Physiol* 281:H1422–H1432
130. Den Hartog GJ, Haenen GR, Boven E, van der Vijgh WJ, Bast A (2004) Lecithinized copper, zinc-superoxide dismutase as a protector against dexorubicin-induced cardiotoxicity in mice. *Toxicol Appl Pharmacol* 194:180–188
131. Adler A, Messina E, Sherman B, Wang Z, Huang H, Linke A, Hintze TH (2003) NAD(P)H oxidase-generated superoxide anion accounts for reduced control of myocardial O_2 consumption by NO in old Fisher 344 rats. *Am J Physiol Heart Circ Physiol* 285:H1015–H1022
132. Fukaya Y, Yamaguchi M (2004) Regucalcin increases superoxide dismutase activity in rat liver cytosol. *Biol Pharm Bull* 27:1444–1446
133. Ichikawa E, Yamaguchi M (2004) Regucalcin increases superoxide dismutase activity in the heart cytosol of normal and regucalcin transgenic rats. *Int J Mol Med* 14:691–695
134. Connelly PA, Sisk RB, Schulman H, Garrison JC (1987) Evidence for the activation of the multifunction Ca^{2+} /calmodulin-dependent protein kinase in response to hormones that increase intracellular Ca^{2+} . *J Biol Chem* 262:10154–10163
135. Mori S, Yamaguchi M (1990) Hepatic calcium-binding protein regucalcin decreases Ca^{2+} /calmodulin-dependent protein kinase activity in rat liver cytosol. *Chem Pharm Bull* 38:2216–2218
136. Kurota H, Yamaguchi M (1997) Inhibitory effect of regucalcin on Ca^{2+} /calmodulin-dependent protein kinase activity in rat renal cortex cytosol. *Mol Cell Biochem* 177:239–243
137. Hamano T, Hanahisa Y, Yamaguchi M (1999) Inhibitory effect of regucalcin on Ca^{2+} -dependent protein kinase activity in rat brain cytosol: involvement of endogenous regucalcin. *Brain Res Brain* 50:187–192
138. Hamano T, Yamaguchi M (2001) Inhibitory role of regucalcin in the regulation of Ca^{2+} -dependent protein kinase activity in rat brain tissues. *J Neurol Sci* 183:33–38
139. Omura M, Yamaguchi M (1998) Inhibition of Ca^{2+} /calmodulin-dependent phosphatase activity by regucalcin in rat liver cytosol: involvement of calmodulin binding. *J Cell Biochem* 71:140–148
140. Yamaguchi M, Mori S (1990) Inhibitory effect of calcium-binding protein regucalcin on protein kinase C activity in rat liver cytosol. *Biochem Med Metab Biol* 43:140–146
141. Kurota H, Yamaguchi M (1998) Inhibitory effect of calcium-binding protein regucalcin on protein kinase C activity in rat renal cortex cytosol. *Biol Pharm Bull* 21:315–318
142. Hunter T (1995) Protein kinases and phosphatases: the Yin and Yang of protein phosphorylation and signaling. *Cell* 80:225–236
143. Wang Y, Santini F, Qin K, Huang CY (1995) A Mg^{2+} -dependent, Ca^{2+} -inhibitable seruin/throsine protein phosphatase from bovine brain. *J Biol Chem* 270:25607–25612
144. Pallen CJ, Wang JH (1983) Calmodulin-stimulated dephosphorylation of *p*-nitrophenylphosphate and free phosphotyrosine by calcineurin. *J Biol Chem* 258:850–855
145. Omura M, Yamaguchi M (1999) Effect of anti-regucalcin antibody on neutral phosphatase activity in rat liver cytosol: involvement of endogenous regucalcin. *Mol Cell Biochem* 197:25–29
146. Omura M, Kurota H, Yamaguchi M (1998) Inhibitory effect of regucalcin on Ca^{2+} /calmodulin-dependent phosphatase activity in rat renal cortex cytosol. *Biol Pharm Bull* 21:440–443
147. Omura M, Yamaguchi M (1999) Enhancement of neutral phosphatase activity in the cytosol and nuclei of regenerating rat liver: role of endogenous regucalcin. *J Cell Biochem* 73:332–341
148. Omura M, Yamaguchi M (1999) Regulation of protein phosphatase activity by regucalcin localization in rat liver nuclei. *J Cell Biochem* 75:437–445
149. Morooka Y, Yamaguchi M (2001) Suppressive role of endogenous regucalcin in the regulation of protein phosphatase activity in rat renal cortex cytosol. *J Cell Biochem* 81:639–646
150. Morooka Y, Yamaguchi M (2001) Inhibitory effect of regucalcin on protein phosphatase activity in the nuclei of rat kidney cortex. *J Cell Biochem* 83:111–120
151. Morooka Y, Yamaguchi M (2002) Endogenous regucalcin suppresses the enhancement of protein phosphatase activity in the cytosol and nucleus of kidney cortex in calcium-administered rats. *J Cell Biochem* 85:553–560

152. Molkenkin JD, Lu JR, Antos CL, Markham B, Richardson J, Robbins J, Grant SR, Olson EN (1998) A calcineurin-dependent transcriptional pathway for cardiac hypertrophy. *Cell* 17:215–228
153. Ichikawa E, Tsurusaki Y, Yamaguchi M (2004) Suppressive effect of regucalcin on protein phosphatase activity in the heart cytosol of normal and regucalcin transgenic rats. *Int J Mol Med* 13:289–293
154. Hamano T, Yamaguchi M (1999) Inhibitory effect of regucalcin on Ca^{2+} /calmodulin-dependent protein phosphatase activity in rat brain cytosol. *Int J Mol Med* 3:615–619
155. Tobisawa M, Tsurusaki Y, Yamaguchi M (2003) Decrease in regucalcin level and enhancement of protein tyrosine phosphatase activity in rat brain microsomes with increasing age. *Int J Mol Med* 12:577–580
156. Tobisawa M, Yamaguchi M (2003) Suppressive effect of endogenous regucalcin on protein tyrosine phosphatase activity in the nucleus of rat brain: attenuation with increasing age. *Int J Mol Med* 11:205–210
157. Brostrom CO, Bocckino SB, Brostrom MA, Galuska EM (1983) Identification of a Ca^{2+} requirement for protein synthesis in eukaryotic cells. *J Biol Chem* 258:14390–14399
158. Brostrom CO, Bocckino SB, Brostrom MA, Galuska EM (1986) Regulation of protein synthesis in isolated hepatocytes by calcium-mobilizing hormones. *Mol Pharmacol* 29:104–111
159. Menaya J, Parvilla R, Ayuso MS (1988) Effect of vasopressin on the regulation of protein synthesis initiation in liver cells. *Biochem J* 254:773–779
160. Thomas AP, Alexander J, Williamson JR (1984) Relationship between inositol polyphosphate production and the increase of cytosolic free Ca^{2+} induced by vasopressin in isolated hepatocytes. *J Biol Chem* 259:5574–5584
161. Berthon B, Binet A, Mauger J-P, Claret M (1984) Cytosolic free Ca^{2+} in isolated rat hepatocytes as measured by quin 2. Effects of noradrenalin and vasopressin. *FEBS Lett* 167:19–24
162. Yamaguchi M, Mori S (1990) Effect of calcium-binding protein regucalcin on hepatic protein synthesis: inhibition of aminoacyl-tRNA synthetase activity. *Mol Cell Biochem* 99:25–32
163. Tsurusaki Y, Yamaguchi M (2000) Suppressive effect of endogenous regucalcin on the enhancement of protein synthesis and aminoacyl-tRNA synthetase activity in regenerating rat liver. *Int J Mol Med* 6:295–299
164. Wang KK, Villalobo A, Ronfogalis BD (1989) Calmodulin-binding protein as calpain substrates. *Biochem J* 262:693–706
165. Pontremoli S, Melloni E, Salamino F, Sparator B, Michetti M, Horecker BL (1984) Cytosolic Ca^{2+} -dependent neutral proteinases from rabbit liver: activation of the proenzymes by Ca^{2+} and substrate. *Proc Natl Acad Sci USA* 81:53–56
166. Yamaguchi M, Tai H (1992) Calcium-binding protein regucalcin increases calcium-independent proteolytic activity in rat liver cytosol. *Mol Cell Biochem* 12:89–95
167. Yamaguchi M, Nishina N (1995) Characterization of regucalcin effect on proteolytic activity in rat liver cytosol: relation to cysteinyl-proteases. *Mol Cell Biochem* 148:67–72
168. Mellgren RI (1987) Calcium-dependent proteases: an enzyme system active at cellular membranes? *FASEB J* 1:110–115
169. Bode W, Huber R (1992) Natural protein proteinase inhibitors and their interaction with proteinase. *Eur J Biochem* 204:433–451
170. Rosser BG, Powers SP, Gores GJ (1993) Calpain activity increases in hepatocytes following addition of ATP. Demonstration by a novel fluorescent approach. *J Biol Chem* 268:23593–23600
171. Baba T, Yamaguchi M (1999) Stimulatory effect of regucalcin on proteolytic activity in rat renal cortex cytosol: involvement of thiol proteases. *Mol Cell Biochem* 195:87–92
172. Baba T, Yamaguchi M (2000) Stimulatory effect of regucalcin on proteolytic activity is impaired in the kidney cortex cytosol of rats with saline ingestion. *Mol Cell Biochem* 206:1–6
173. Croall DE, Demartino GN (1991) Calcium-activated neutral protease (calpain) system: structure, function, and regulation. *Physiol Rev* 71:813–847
174. Melloni E, Pontremoli S, Michetti M, Sacco O, Sparatore B, Horecker BL (1986) The involvement of calpain in the activation of protein kinase C in neutrophils stimulated by phorbol myristic acid. *J Biol Chem* 261:4101–4105
175. Boynton AL, Whitfield JF, MacManus JP (1980) Calmodulin stimulates DNA synthesis by rat liver cells. *Biochem Biophys Res Commun* 95:745–749
176. Cruise J, Houck KA, Michalopoulos GK (1985) Induction of DNA synthesis in cultured rat hepatocytes through stimulation of alpha 1 adrenoreceptor by norepinephrine. *Science* 227:749–751
177. Pujol MJ, Soriano M, Alique R, Carafoli E, Bachs O (1989) Effect of alpha-adrenergic blockers on calmodulin associate with the nuclear matrix of rat liver cells during proliferative activation. *J Biol Chem* 264:18863–18865
178. Nakagawa T, Yamaguchi M (2008) Nuclear localization of regucalcin is enhanced in culture with protein kinase C activation in cloned normal rat kidney proximal tubular epithelial NRK52E cells. *Int J Mol Med* 21:605–610
179. Tsurusaki Y, Yamaguchi M (2004) Role of regucalcin in liver nuclear function: binding of regucalcin to nuclear protein or DNA and modulation of tumor-related gene expression. *Int J Mol Med* 14:277–281
180. Jones DP, McConkey DJ, Nicotera P, Orrenius S (1989) Calcium-activated DNA fragmentation in rat liver nuclei. *J Biol Chem* 264:6398–6403
181. Farber JL (1981) The role of calcium in cell death. *Life Sci* 29:1289–1295
182. Yamaguchi M, Sakurai T (1991) Inhibitory effect of calcium-binding protein regucalcin on Ca^{2+} -activated DNA fragmentation in rat liver nuclei. *FEBS Lett* 279:281–284
183. Moroianu J, Blobel G (1995) Protein export from the nucleus requires the GTPase Ran and GTP hydrolysis. *Proc Natl Acad Sci USA* 92:4318–4322
184. Tsurusaki Y, Yamaguchi M (2001) Suppressive effect of endogenous regucalcin guanosine triphosphatase activity in rat liver nucleus. *Biol Pharm Bull* 24:958–961
185. Katsumata T, Yamaguchi M (1998) Inhibitory effect of calcium-binding protein regucalcin on protein kinase activity in the nuclei of regenerating rat liver. *J Cell Biochem* 71:569–576
186. Yamaguchi M, Kanayama Y (1996) Calcium-binding protein regucalcin inhibits deoxyribonucleic acid synthesis in the nuclei of regenerating rat liver. *Mol Cell Biochem* 162:121–126
187. Tsurusaki Y, Yamaguchi M (2002) Suppressive role of endogenous regucalcin in the enhancement of deoxyribonucleic acid synthesis activity in the nucleus of regenerating rat liver. *J Cell Biochem* 85:516–552
188. Morooka Y, Yamaguchi M (2002) Suppressive effect of endogenous regucalcin on deoxyribonucleic acid synthesis in the nuclei of rat renal cortex. *Mol Cell Biochem* 229:157–162
189. Yamaguchi M, Ueoka S (1997) Inhibitory effect of calcium-binding protein regucalcin on ribonucleic acid synthesis in isolated rat liver nuclei. *Mol Cell Biochem* 173:169–175
190. Tsurusaki Y, Yamaguchi M (2002) Role of endogenous regucalcin in nuclear regulation of regenerating rat liver: suppression of the enhanced ribonucleic acid synthesis activity. *J Cell Biochem* 87:450–457
191. Pardo JP, Fernandez F (1982) Effect of calcium and calmodulin on RNA synthesis in isolated nuclei from rat liver cells. *FEBS Lett* 143:157–160

192. Sturges MR, Peck LJ (1994) Calcium-dependent inactivation of RNA polymerase III transcription. *J Biol Chem* 269:5712–5719
193. Liu S, Shia D, Liu G, Chen H, Liu S, Hu Y (2000) Roles of Se and NO in apoptosis of hepatoma cells. *Life Sci* 68:603–610
194. Abou-Elella AM, Siendones E, Padillo J, Montero JL, De la Meta M, Relat JM (2002) Tumour necrosis factor- α and nitric oxide mediate apoptosis by D-galactosamine in a primary culture of rat hepatocytes: exacerbation of cell death by cocultured Kupffer cells. *Can J Gastroenterol* 16:791–799
195. Belloma G, Perotti M, Taddei F, Mirabelli F, Finardi G, Nicoletta P, Orrenius S (1992) Tumor necrosis factor α induces apoptosis in mammary adenocarcinoma cells by an increase in intranuclear free Ca^{2+} concentration and DNA fragmentation. *Cancer Res* 52:1342–1346
196. Hukkanen M, Hughes FJ, Buttery LD, Gross SS, Evans TJ, Seddon S, Riveros-Moreno V, MacIntyre I, Polak JM (1995) Cytokine stimulated expression of inducible nitric oxide synthase by mouse, rat, and human osteoblast-like cells and its functional role in osteoblast metabolic activity. *Endocrinology* 136:5445–5453
197. Wink DA, Hanbauer I, Laval F, Cook JA, Krishna MC, Mitchell JB (1994) Nitric oxide protects against the cytotoxic effects of reactive oxygen species. *Ann NY Acad Sci* 738:265–278
198. Izumi T, Yamaguchi M (2004) Overexpression of regucalcin suppresses cell death in cloned rat hepatoma H4-II-E cells induced by tumor necrosis factor- α or thapsigargin. *J Cell Biochem* 92:296–306
199. Alikhani M, Alikhani Z, He H, Liu R, Popek BI, Graves DT (2003) Lipopolysaccharides indirectly stimulate apoptosis and global induction of apoptotic genes in fibroblasts. *J Biol Chem* 278:52901–52908
200. Nalan Y, Vereker E, Lynch AM, Lynch MA (2003) Evidence that lipopolysaccharide-induced cell death is mediated by accumulation of reactive oxygen species and activation of p38 in rat cortex and hippocampus. *Exp Neurol* 184:794–804
201. Izumi T, Yamaguchi M (2004) Overexpression of regucalcin suppresses cell death and apoptosis in cloned rat hepatoma H4-II-E cells induced by lipopolysaccharide, PD98059, dibucaine, or Bay K 8644. *J Cell Biochem* 93:598–608
202. Zelivianshi S, Spellman M, Kellerman M, Kakitelashvili V, Zhou XW, Lugo E, Lee MS, Taylor R, Daris TL, Hauke R, Lin MF (2003) ERK inhibitor PD98059 enhances docetaxel-induced apoptosis of androgen-independent human prostate cancer cells. *Int J Cancer* 107:478–485
203. Arita K, Utsumi T, Kato A, Kanno T, Kobuchi H, Inoue B, Akiyama J, Utsumi M (2000) Mechanism of dibucaine-induced apoptosis in promyelocytic leukemia cells (HL-60). *Biochem Pharmacol* 60:905–915
204. Giuliano M, Bellacchia G, Lauricella M, D'Anneo A, Vassallo B, Vento R, Tesoriere G (2004) Staurosporine-induced apoptosis in Chang liver cells is associated with down-regulation of Bcl-2 and Bcl-XL. *Int J Mol Med* 13:565–571
205. Christensen SB, Andersen A, Kromann H, Treiman M, Tombal B, Denmeads S, Isaacs JT (1999) Thapsigargin analogues for targeting programmed death of androgen-independent prostate cancer cells. *Bioorg Med Chem* 7:1273–1280
206. Tombal B, Weeraratna AT, Denmeade SR, Isaacs JT (2000) Thapsigargin induces a calmodulin/calcineurin-dependent apoptotic cascade responsible for the death of prostatic cancer cells. *Prostate* 43:303–317
207. Cohen JJ, Duke RC (1984) Glucocorticoid activation of a calcium-dependent endonuclease in thymocytes nuclei leads to cell death. *J Immunol* 132:38–42
208. Pereira M, Millot J-M, Seville S, Manfait M (2002) Inhibitory effect of extracellular Mg^{2+} on intracellular Ca^{2+} dynamic changes and thapsigargin-induced apoptosis in human cancer MCF7 cells. *Mol Cell Biochem* 229:163–171
209. Cano-Abad MF, Villarroja M, Garcia AG, Gabilan NH, Lopez MG (2001) Calcium entry through L-type calcium channels causes mitochondrial disruption and chromaffin cell death. *J Biol Chem* 276:39695–39704
210. Fukaya Y, Yamaguchi M (2005) Overexpression of regucalcin suppresses cell death and apoptosis in cloned rat hepatoma H4-II-E cells induced by insulin or insulin-like growth factor-I. *J Cell Biochem* 96:145–154
211. Spinozzi F, Pagliacci MC, Migliorati G, Moraca R, Grignani F, Riccardi C, Nicoletti I (1994) The natural tyrosine kinase inhibitor genistein produces cell cycle arrest and apoptosis in Jurkat T-leukemia cells. *Leuk Res* 18:431–439
212. Gamet-Payrastra L, Li P, Lumeau S, Cassar G, Duport NA, Chevolleau S, Gasc N, Tulliez J, Terce F (2000) Sulforaphane, a naturally occurring isothiocyanate, induces cell cycle arrest and apoptosis in HT29 human colon cancer cells. *Cancer Res* 60:1426–1433
213. Fimognari C, Nusse M, Cesari R, Iori R, Cantelli-Forti G, Hrelia P (2002) Growth inhibition, cell-cycle arrest and apoptosis in human T-cell leukemia by the isothiocyanate sulforaphane. *Carcinogenesis* 23:581–586
214. Singh AV, Xiao D, Lew KL, Dhir R, Singh SV (2004) Sulforaphane induces caspase-mediated apoptosis in cultured PC-3 human prostate cancer cells and retards growth of PC-3 xenografts in vivo. *Carcinogenesis* 25:83–90
215. Fukaya Y, Yamaguchi M (2005) Overexpression of regucalcin suppresses apoptotic cell death in the cloned rat hepatoma H4-II-E cells induced by a naturally occurring isothiocyanate sulforaphane. *Int J Mol Med* 15:853–857
216. Nakagawa T, Yamaguchi M (2005) Hormonal regulation on regucalcin mRNA expression in cloned normal rat kidney proximal tubular epithelial NRK52E cells. *J Cell Biochem* 95:589–597
217. Nakagawa T, Yamaguchi M (2005) Overexpression of regucalcin suppresses apoptotic cell death in cloned normal rat kidney proximal tubular epithelial NRK52E cells: change in apoptosis-related gene expression. *J Cell Biochem* 96:1274–1285
218. Vogelstein B, Lane D, Levine AJ (2000) Surfing the p53 network. *Nature* 408:307–310
219. Zou H, Hanzel WJ, Liu X, Lutschg A, Wang X (1997) Apaf-1, a human protein homologous to *C. elegans* CED-4, participates in cytochrome c-dependent activation of caspase-3. *Cell* 90:405–413
220. Widmann C, Gibson S, Johnson GL (1988) Caspase-dependent cleavage of signaling proteins during apoptosis. A turn-off mechanism for anti-apoptotic signals. *J Biol Chem* 273:7141–7147
221. Dieguez-Acuna FJ, Polk WW, Ellis ME, Simmonds PL, Kushleika JV, Woods JS (2004) Nuclear factor kappaB activity determines the sensitivity of kidney epithelial cells to apoptosis: implications for mercury-induced renal failure. *Toxicol Sci* 82:1114–1123
222. Inagaki S, Yamaguchi M (2001) Suppressive role of endogenous regucalcin in the enhancement of protein kinase activity with proliferation of cloned rat hepatoma cells (H4-II-E). *J Cell Biochem (Supplement)* 36:12–18
223. Inagaki S, Misawa H, Yamaguchi M (2000) Role of endogenous regucalcin in protein tyrosine phosphatase regulation in the cloned rat hepatoma cells (H4-II-E). *Mol Cell Biochem* 213:43–50
224. Inagaki S, Yamaguchi M (2000) Enhancement of protein tyrosine phosphatase activity in the proliferation of cloned rat hepatoma H4-II-E cells: suppressive role of endogenous regucalcin. *Int J Mol Med* 6:323–328

225. Mackintosh C, Malintosh RW (1997) Inhibitors of protein kinases and phosphatases. *Trends Biochem Sci* 19:444–448
226. Cohen P, Cohen PTW (1989) Protein phosphatase come of age. *J Biol Chem* 264:21435–21438
227. Inagaki S, Yamaguchi M (2001) Regulatory role of endogenous regucalcin in the enhancement of nuclear deoxyribonucleic acid synthesis with proliferation of cloned rat hepatoma cells (H4-II-E). *J Cell Biochem* 82:704–711
228. Misawa H, Inagaki S, Yamaguchi M (2002) Suppression of cell proliferation and deoxyribonucleic acid synthesis in cloned rat hepatoma H4-II-E cells overexpressing regucalcin. *J Cell Biochem* 84:143–149
229. Yamaguchi M, Daimon Y (2005) Overexpression of regucalcin suppresses cell proliferation in cloned rat hepatoma H4-II-E cells: involvement of intracellular signaling factors and cell cycle-related genes. *J Cell Biochem* 95:1169–1177
230. Yamaguchi M (2005) Role of regucalcin in maintaining cell homeostasis and function (Review). *Int J Mol Med* 15:371–389
231. Meijer L, Borgne A, Mulner O, Chhng JP, Blow JJ, Inagaki N, Inagaki M, Delcros JG, Moulinoux JP (1997) Biochemical and cellular effects of roscovitine, a potent and selective inhibitor of the cyclin-dependent kinases cdc2, cdk2 and cdk5. *Eur J Biochem* 243:527–536
232. Singh SV, Herman-Antosiewicz A, Singh AV, Lew KL, Srivastava SK, Kamath R, Brown KD, Zhang L, Baskaran R (2004) Sulforaphane-induced G2/M phase cell cycle arrest involves checkpoint kinase 2-mediated phosphorylation of cell division cycle 25C. *J Biol Chem* 279:25813–25822
233. Hulla JE, Schneider RP (1993) Structure of the rat *p53* tumor suppressor gene. *Nucleic Acids Res* 21:713–717
234. Charollais RH, Buquet C, Mester J (1990) Butyrate blocks the accumulation of Cdc2 mRNA in late G1 phase but inhibits both early and late G1 progression in chemically transformed mouse fibroblasts BP-A31. *J Cell Physiol* 145:46–52
235. Curran T (1991) Fos and Jun: intermediary transcription factors. In: Cohen P, Foulkes JG (eds) *The hormonal control of gene transcription*. Elsevier Science Publisher, New York, pp 295–308
236. Tsurusaki Y, Yamaguchi M (2003) Overexpression of regucalcin modulates tumor-related gene expression in cloned rat hepatoma H4-II-E cells. *J Cell Biochem* 90:619–626
237. Nakagawa T, Sawada N, Yamaguchi M (2005) Overexpression of regucalcin suppresses cell proliferation of cloned normal rat kidney proximal tubular epithelial NRK52E cells. *Int J Mol Med* 16:637–643
238. Dang ZC, Lowik CW (2004) Differential effects of PD98059 and UO126 on osteogenesis and adipogenesis. *J Cell Biochem* 92:525–533
239. Tamaoki T, Nomoto H, Takahashi I, Kato Y, Morimoto M, Tomita E (1986) Staurosporine, a potent inhibitor of phospholipids/ Ca^{2+} -dependent protein kinase. *Biochem Biophys Res Commun* 135:397–402
240. Vincenzi FF (1982) Pharmacology of calmodulin antagonism. In: Godfrani T, Albertini A, Paoletti R (eds) *Calcium modulators*. Elsevier Biomedical Press, Amsterdam, pp 67–80
241. Park YC, Lee CH, Kang HS, Chung HT, Kim HD (1997) Wortmannin, a specific inhibitor of phosphatidylinositol-3-kinase, enhances LPS-induced NO production from murine peritoneal macrophages. *Biochem Biophys Res Commun* 240:692–696
242. Nakagawa T, Yamaguchi M (2006) Overexpression of regucalcin enhances its nuclear localization and suppresses L-type Ca^{2+} channel and calcium-sensing receptor mRNA expressions in cloned normal rat kidney proximal tubular epithelial NRK52E cells. *J Cell Biochem* 99:1064–1077
243. Tanaka T, Nangaku M, Miyata T, Inagi R, Ohse T, Ingelfinger JR, Fujita T (2004) Blockade of calcium influx through L-type calcium channels attenuates mitochondrial injury and apoptosis in hypoxic renal tubular cells. *J Am Soc Nephrol* 15:2320–2333
244. Ba J, Friedman PA (2004) Calcium-sensing receptor regulation of renal mineral ion transport. *Cell Calcium* 35:229–237
245. Xue JH, Takahashi H, Yamaguchi M (2000) Stimulatory effect of regucalcin on mitochondrial ATP-dependant calcium uptake activity in rat kidney cortex. *J Cell Biochem* 80:285–292
246. Nakagawa T, Yamaguchi M (2007) Overexpression of regucalcin suppresses cell response for tumor necrosis factor- α or transforming growth factor- β 1 in cloned normal rat kidney proximal tubular epithelial NRK52E cells. *J Cell Biochem* 100:1178–1190
247. Fan JM, Ng Y-Y, Hill PA, Nikolic-Paterson DJ, Mu W, Atkins RC, Lan HY (1999) Transforming growth factor- β regulates tubular epithelial-myofibroblast transdifferentiation in vitro. *Kidney Int* 56:1455–1467
248. Higuchi O, Nakamura T (1991) Identification and change in the receptor for hepatocyte growth factor in rat liver after partial hepatectomy or induced hepatitis. *Biochem Biophys Res Commun* 176:599–607
249. Baffy G, Yang L, Michalopoulos GK, Williamson JR (1992) Hepatocyte growth factor induces calcium mobilization and inositol phosphate production in rat hepatocytes. *J Cell Physiol* 153:332–339
250. Higgins GM, Anderson RM (1931) Experimental pathology of the liver. Restoration of the liver of the white rat following partial surgical removal. *Arch Pathol* 12:186–202
251. Kanayama Y, Yamaguchi M (1995) Enhancement of nuclear Ca^{2+} -ATPase activity in regenerating rat liver: involvement of nuclear DNA increase. *Mol Cell Biochem* 146:179–186
252. Pinol MR, Berchtold MW, Backs O, Heizmann CW (1988) Increased calmodulin synthesis in the pre-replicative phase of rat liver regeneration. *FEBS Lett* 231:445–450
253. Suzuki S, Asamoto M, Tsujimura K, Shirai T (2004) Specific differences in gene expression profile revealed by cDNA microarray analysis of glutathione S-transferase placental form (GST-P) immunohistochemically positive rat liver foci and surrounding tissue. *Carcinogenesis* 25:439–443
254. Yamaguchi M, Morooka Y, Misawa H, Tsurusaki Y, Nakajima R (2002) Role of endogenous regucalcin in transgenic rats: suppression of kidney cortex cytosolic protein phosphatase activity and enhancement of heart muscle microsomal Ca^{2+} -ATPase activity. *J Cell Biochem* 86:520–529
255. Carpentier A, Taghibiglou C, Leung N, Szeto L, van Iderstine SC, Vffelman KD, Buckingham R, Adeli K, Lewis GF (2002) Ameliorated hepatic insulin resistance is associated with normalization of microsomal triglyceride transfer protein expression and reduction in very low density lipoprotein assembly and secretion in the fructose-fed hamster. *J Biol Chem* 277:28795–28802
256. Tsurusaki Y, Yamaguchi M (2003) Role of endogenous regucalcin in transgenic rats: suppression of protein tyrosine phosphatase and ribonucleic acid synthesis in liver nucleus. *Int J Mol Med* 12:207–211
257. Fukaya Y, Yamaguchi M (2004) Characterization of protein tyrosine phosphatase activity in rat liver microsomes: suppressive effect of endogenous regucalcin in transgenic rats. *Int J Mol Med* 14:427–432
258. Posner BI, Faure R, Burgess JW, Bevan AP, Lachance D, Zhand-Sun G, Fantus IG, Ng JB, Hall DA, Lum BS, Shaver A (1994) Peroxovanadium compounds: a new class of potent phosphotyrosine phosphatase inhibitors which are insulin mimetics. *J Biol Chem* 269:4596–4604

259. Ahmad F, Goldstein BJ (1995) Increased abundance of specific skeletal muscle protein-tyrosine phosphatases in a genetic model of insulin-resistance obesity and diabetes mellitus. *Metabolism* 44:1175–1184
260. Dylla SJ, Williams JP, Williford J, Hardy RW (2000) Phosphatase activity in rat adipocytes: effects of insulin and insulin resistance. *J Cell Biochem* 77:445–454
261. Nakashima C, Yamaguchi M (2006) Overexpression of regucalcin enhances glucose utilization and lipid production in cloned rat hepatoma H4-II-E cells: involvement of insulin resistance. *J Cell Biochem* 99:1582–1592
262. Nakashima C, Yamaguchi M (2007) Overexpression of regucalcin suppresses gene expression of insulin signaling-related proteins in cloned rat hepatoma H4-II-E cells: involvement of insulin resistance. *Int J Mol Med* 20:709–716
263. Yamaguchi M (2010) Regucalcin and metabolic disorder: osteoporosis and hyperlipidemia are induced in regucalcin transgenic rats. *Mol Cell Biochem* 341:119–133
264. Yamaguchi M, Kobayashi M, Uchiyama S (2005) Suppressive effect of regucalcin on cell differentiation and mineralization in osteoblastic MC3T3–E1 cells. *J Cell Biochem* 96:543–554
265. Yamaguchi M, Uchiyama S (2005) Regucalcin stimulates osteoclast-like cell formation in mouse marrow cultures. *J Cell Biochem* 94:794–803
266. Yamaguchi M, Otomo Y, Uchiyama S, Nakagawa T (2008) Hormonal regulation of regucalcin mRNA expression in osteoblastic MC3T3–E1 cells. *Int J Mol Med* 21:771–775
267. Otomo Y, Yamaguchi M (2006) Regulatory effect of exogenous regucalcin on cell function in osteoblastic MC3T3–E1 cells: involvement of intracellular signaling factor. *Int J Mol Med* 18:321–327
268. Ishigami A, Fujita T, Handa S, Shirasawa T, Koseki H, Kitamura T, Enomoto N, Sato N, Shimosawa T, Maruyama N (2002) Senescence marker protein-30 knockout mouse liver is highly susceptible to tumor necrosis factor- α -and Fas-mediated apoptosis. *Am J Pathol* 161:1273–1281
269. Ishigami A, Kondo Y, Nanba R, Ohsawa T, Handa S, Kubo S, Akita M, Maruyama N (2004) SMP30 deficiency in mice causes an accumulation of neutral lipids and phospholipids in the liver and shortens the life span. *Biochem Biophys Res Commun* 315:575–580
270. Kondo Y, Inai Y, Sato Y, Handa S, Kubo S, Shimokado K, Goto S, Nishikimi M, Maruyama N, Ishigami A (2006) Senescence marker protein 30 functions as gluconolactonase in L-ascorbic acid biosynthesis, and its knockout mice are prone to scurvy. *Proc Natl Acad Sci USA* 103:5723–5728
271. Nejak-Bowen KN, Zeng G, Tan X, Cieply B, Monga SP (2009) β -Catenin regulates vitamin C biosynthesis and cell survival in murine liver. *J Biol Chem* 284:28115–28127