

# **Protein Hydrogels: A Concise Review of Properties and Applications**

**Bhawna Malik<sup>1</sup> · Rekha Chawla1 · Sunil Kumar Khatkar1**

Accepted: 2 September 2023 / Published online: 25 September 2023 © The Author(s), under exclusive licence to Springer Nature B.V. 2023

## **Abstract**

Proteins are renewable resources generated from residues and byproducts of the agriculture and livestock industries. Numerous in-depth studies have been conducted on proteins for their utilisation as protein-based hydrogels. Henceforth, for maximum enrichment of these dietary proteins in the diet, available proteins are transformed into hydrogels using enzymes, chemicals, or physical means. In addition to being nontoxic, biocompatible, biodegradable, accessible, and renewable, protein hydrogels are also afordable. They come in a wide range of physicochemical states dissolved in emulsions or liquids, like particle hydrogels, macro hydrogels, a form of gel colloid, and suspensions. However, composite or hybrid hydrogels are developed by mixing proteins with polysaccharides and other biomolecules to enhance mechanical and functional properties for specifc applications. Additionally, these biomaterials have attracted increasing interest in the past few years owing to their improved tensile strengths and can enclose preserved and released biomolecules. Thus, hydrogels can manage the release of bioactive substances and boost the bioavailability of such ingredients, in addition to their capacity to heal wounds and their use in the food and agriculture industries. This article thoroughly analyses basic polymer hydrogel categorization, diverse protein hydrogel forms, production techniques, physicochemical and functional properties, and applications in various felds. This review also sheds light on methods for determining the shape, thermal stability, and rheological characteristics of protein-based hydrogels. Therefore, will provide a comprehensive reference for the numerous aspects of protein hydrogels that are important to multiple academic and industrial disciplines.

 $\boxtimes$  Bhawna Malik bhawnamalik7721@gmail.com

 $\boxtimes$  Rekha Chawla mails4rekha@gmail.com

<sup>1</sup> College of Dairy Science & Technology, Guru Angad Dev Veterinary & Animal Sciences University, Ludhiana, Punjab, India

#### **Graphical Abstract**



**Keywords** Hydrogel · Protein · Polysaccharide · Biomaterials · Material science

## **Introduction**

The term "hydrogel" was frst applied in 1894 in a publication citing its usage as a protein-rich source. Also, the article referred it to be a colloidal gel consisting of inorganic salts rather than a hydrogel as we understand it today. Furthermore, a polyhydroxy ethyl methacrylate (pHEMA) hydrogel was invented a long time afterwards, in 1960, intending to employ them in long-term applications involving human tissues, and was the frst cross-linked network material that was refected in the scientifc literature. It was characterized by its standard hydrogel properties, one of which is high water affinity (Chirani et al. [2015\)](#page-16-0). Since then, numerous studies have been conducted on hydrogels and presented their vast applications. The term hydrogel is composed of two words, i.e., hydro (water) and gels (thick, soft-cum-semirigid, and transparent colloidal dispersion) and has been well-defned in various manners by scientists over the years. The most prevalent is "hydrogels or hydrophilic gels," having a threedimensional structure composed of a single monomer or polymer chain of networks. The hydrogel is fexible and can be made into a variety of forms and sizes. It can also absorb up to a thousand of its dry weight in water (Seliktar [2012](#page-19-0)). Another description is that it is a polymeric substance that can absorb a lot of water inside its network but won't dissolve (Ahmed [2015](#page-16-1)). A hydrogel can be produced using any method that produces a cross-linked polymer. When hydrophilic monomers and a multipurpose cross-linker are combined, copolymerization/cross-linking free-radical polymerizations are frequently utilised to create hydrogels (Ullah et al. [2015](#page-19-1)). Due to counter-balanced capillary, osmotic, and hydration forces, polymeric chain networks interact with water or biological fuids, causing the chain network to expand. This equilibrium state in the hydrogel is determined by the degree of these opposing efects, which dictates some of the hydrogel's inherent qualities, such as internal transport, difusion properties, and mechanical strength (Varaprasad et al. [2017\)](#page-19-2). Based on certain characteristics such as swelling rate, mechanical capabilities (for food packaging and pharmaceutical applications), and biocompatibility (essential for edible hydrogels), hydrogels can also be employed in a variety of industries (Ali and Ahmed [2018](#page-16-2)). Hydrogels impart a wide range of food and non-food applications such as carriers for favour, bioactive & nutritional compounds; enhance bioavailability; provide stable network structures; improve stability; calorie reduction; trans & saturated fat replacements in dessert; oral delivery of lipophilic active ingredients; tissue engineering; wound dressing; contact lens; disposable diapers; pharmaceutical industry; agriculture; etc. (Caló and Khutoryanskiy [2015;](#page-16-3) Li et al. [2016](#page-18-0); Pyarasani et al. [2019](#page-19-3); Sabbagh et al. [2019](#page-19-4)). Moreover, the characteristics of hydrogels depend upon several factors like polymer concentration, the degree of crosslinking, temperature, pH, age, salt concentrations, etc. (Bae and Kurisawa [2016](#page-16-4); Saini [2017;](#page-19-5) Gun'ko et al. [2017](#page-17-0)). Synthetic polymers including poly acrylamide, poly hydroxyalkyl methacrylates, poly acrylamide, and polymethacrylamide, as well as its derivates poly N-vinyl-2-pyrolidone and polyvinyl alcohol (PVA), are used to make hydrogels. Despite having good mechanical strength and great water absorption capabilities, synthetic hydrogels have certain unpredictability of their toxicity references caused by the unresponsive monomers in the gel's structure, which refrains their usage.

In addition, the shortcomings ofered by synthetic hydrogels in terms of their poor biodegradability and biocompatibility could also pose environmental issues (Ni and Dumont [2017](#page-18-1)). Henceforth, it is preferable to utilize natural biopolymers like protein, which can quickly produce functional colloids since they are safe for humans and environmentfriendly. Polysaccharides and proteins are the constituents of the majority of natural hydrogels. When creating hydrogels, proteins have an advantage over polysaccharides because they have more functional groups available for modifcation (amino, thiol, hydroxyl, and carboxyl), are delivered more sensitively to external stimuli, have a unique ability to recognise some peptides and can self-assemble (Ren et al. [2017;](#page-19-6) Cai et al. [2017](#page-16-5); Zhang et al. [2013](#page-20-0)). Edible proteins have been considered prominent in the emergence of functional colloidal gels, including targeted specifc delivery of bioactive substances and modifcation of food-body interactions to strengthen nutritional absorption rate, digestion, and oral perspective (McClements [2017](#page-18-2)). The functional groups inside proteins include carboxyl, amino, hydroxyl, phenolic, and sulfhydryl, which offer many potential applications in the food and drug industries. These may serve as reactive sites for chemical reactions and widen the food, non-food and other biomedical applications. The cross-proteins are safe to eat, compatible with other organisms, and break down naturally. Peptides and proteins are consistently included in hydrogels for creating imitation substances for tissue engineering and medication delivery since the polypeptides are an integral component of the extracellular matrix (Singhal and Gupta [2016\)](#page-19-7). Also, bestowing unique properties, such as foaming, water binding, and thickening, protein-based hydrogels are extensively utilized in various food applications (Gorji et al. [2014](#page-17-1); Zhang et al. [2020](#page-20-0); Abaee et al. [2017\)](#page-15-0). Food protein-based hydrogels from casein, whey protein, keratin, gelatin, collagen, plant protein, etc., can be made unaided or combined with other food-grade polymers. However, protein also imparts biocompatibility, non-toxicity, and availability of reactive sites for alteration, as they contain most hydrophilic functional groups. This review aims to determine protein-based hydrogels, initiated with a brief description of native hydrogels and their classifcation based on diferent characteristics followed by diferent types of proteins utilised for hydrogel preparation and their properties are discussed at length. As hydrogels have tunable viscoelasticity and are biocompatible and injectable, based on these features, their applications in biomedical, food and agricultural felds are discussed.

## **Classifcation of Hydrogels Based on Diferent Parameters**

Diferent categories of hydrogels exist (Fig. [1\)](#page-3-0) based on their physical qualities, manufacturing process, swelling characteristics, ionic charges, origin and sources, cross-linking nature, rate of biodegradation, etc. (Ullah et al. [2015\)](#page-19-1).

#### **Based on the Source**

There are three broad groups of hydrogels based on the source, i.e., natural, synthetic and hybrid. Protein-based materials, polysaccharide-based materials, and those formed from de-cellularized tissue are examples of naturally derived hydrogels wherein natural gels are primarily composed of proteins. The extracellular matrix components like chitin make them naturally biocompatible, bioactive, and potentially suitable for various biomedical applications (Catoira et al. [2019\)](#page-16-6), Whereas synthetic polymers like polyamide (PA) and polyethylene glycol, also known as PEG, are the source of synthetic hydrogels. These are the further diferent kinds of synthetic monomers: anionic, like those made from acrylic acid or crotonic acid; and cationic, like vinyl pyridine, which comes from aminoethyl methacrylate; and neutral, like hydroxyl alkyl methacrylates made from acrylamide, N-vinyl pyrrolidone, and hydrophobic acrylic (Li et al. [2007](#page-18-3)). These biocompatible polymers are hydrolysed either enzymatically or non-enzymatically, or both, in vivo to disintegrate them. The biocompatibility and biodegradability of such polymers also often depend on altering their surface hydrophilicity (Gyles et al. [2017](#page-17-2)). Hybrid/composite hydrogels are produced by combining synthetic polymers with natural ones to overcome the aforementioned problems, enhance current formulations, and broaden the scope of potential uses (Zhu et al. [2018\)](#page-20-1). Generally speaking, proteins and peptides incorporated into networks react with synthetic polymers through polymerization or conjugation to create hybrid hydrogels for a variety of experimental applications

<span id="page-3-0"></span>

gels based on various attributes

#### Conventional **Smart** Copolymeric **Chemical** hydrogels hydrogels hydrogels stimulation **Bio-chemical** Homopolymeric stimulation hydrogels **Physical Properties** Semi-**Stimuli** Physical **Preparation** interpenetrating responsive stimulation hydrogels Interpenetrating **Physically** network crosslinked **HYDROGEL** Cross linking Ampholytic Chemically hydrogels crosslinked Charge **Ionic hydrogels** Biodegradable **Degradability** Source Non-Non ionic biodegradable hydrogels Synthetic **Natural Hybrid**

(cell proliferation, diferentiation, and migration investigations) and in-vivo (wound healing, tissue restoration, and drug delivery) (Palmese et al. [2019\)](#page-18-4).

#### **Based on the Charge**

Hydrogels are also available based on their non-ionic, ionic and ampholytic nature wherein the backbone and side groups of the non-ionic hydrogels are chargeless (Fig. [1](#page-3-0)). Water-polymer interactions cause these non-ionic hydrogels to swell in aqueous medium wherein Poly acrylamide, polyhydroxy ethyl methacrylate, PVA, and PEG are examples of non-ionic hydrogels (Singhal and Gupta [2016](#page-19-7)). Cationic (positive charge carrying) and anionic (negative charge carrying) hydrogels are examples of ionic hydrogels. The pH of the aqueous medium, which controls the level of ionic chain dissociation, controls swelling in the frst scenario. These gels exhibit improved swelling in an acidic media, and low pH values favour the expansion of their chain dissociation. Vinyl pyridine, diethylamino ethyl methacrylate, aminoethyl methacrylate, and dimethyl aminoethyl methacrylate are a few examples of monomers employed in the creation of cationic hydrogels (Ramos et al. [2013\)](#page-19-8). Anionic hydrogels, on the other hand, have a negatively charged backbone. Because these hydrogels dissolve more at higher pH, they swell better in neutral to basic liquids. Maleic acid, acrylic acid, itaconic acid, p-styrene sulphonic acid, crotonic acid, and methacrylic acid are anionic hydrogel monomers (Singhal and Gupta [2016](#page-19-7)). Ampholytic hydrogels, the third type of charge hydrogels, can have both positive and negative charges on one single polymer chain. Every structural repetitive unit in these hydrogels has both basic and acidic groups, and they are all balanced at an isoelectric point. These hydrogels' overall ionic characteristics can be altered by a small pH adjustment. A monomer for the creation of ampholytic hydrogels is N-isopropylacrylamide/ [[3- (methacryloylamino) propyl] dimethy (3-sulfopropyl) ammonium hydroxide] (Singhal and Gupta [2016\)](#page-19-7).

#### **Based on Cross‑Linking**

As hydrogels are mostly made up of cross-linking networks, they are divided into two types depending on cross-linking i.e., chemically crosslinked and physically crosslinked or self-assembled hydrogels (Slaughter et al. [2009\)](#page-19-9). The crosslinking process in physical gels is physically complex like hydrogen bonding, chain aggregation, crystallization, hydrophobic association, and polymer chain complexion (Ullah et al. [2015](#page-19-1)). To create a chemically crosslinked hydrogel, a chemical covalent crosslinking stage is used (either simultaneously with polymerization or post-polymerization). Chemical hydrogels are permanent and irreversible due to persistent confgurational changes in the molecular structure, but physically crosslinked hydrogels are reversible due to modifcations in conformation (Ullah et al. [2015](#page-19-1)).

#### **Based on Methods of Preparation**

Hydrogels are also classed on their fabrication/preparational methods, like (1) homopolymers, (2) copolymers, (3) semiinterpenetrating networks, and (4) interpenetrating networks (Fig. [1](#page-3-0)). Homopolymers are networks of polymers created from a single monomer and a fundamental structural component for a polymeric structure. It might have a crosslinked framework depending on the ingredient and polymerization technique. As a result of their responsiveness to outside stimuli, PEG-based hydrogels are frequently utilised in systems for drug delivery (Iizawa et al. [2007](#page-17-3)). Copolymeric hydrogels are composed of a pair of distinct kinds of monomer units, at least one of which is hydrophilic. To transport drugs, a biodegradable tri-block poly (ethylene glycol) poly(-caprolactone)-poly (ethylene glycol) co-polymeric hydrogel can be used in this study (Gong et al. [2009](#page-17-4)). When a linear polymer penetrates a cross-linked structure lacking any other chemical interactions, it creates a semi-interpenetrating network. For example, semi-IPNs can better handle pH or temperature changes while providing benefts such as fexible pores and progressive drug release because there is no limiting interpenetrating elastic network. The retention of linear cationic polyallyl ammonium chloride in acrylamide/ acrylic acid copolymer hydrogels, which increases mechanical strength and allows for fully reversible pH switching of theophylline release, is one illustration to support the situation (Zhang et al. [2009](#page-20-2)). By dipping a pre-polymerized hydrogel into a mixture of monomers and a polymerization catalyst, these networks develop. The presence of the permanent interlocking of network segments, allows the Interpenetrating network (IPN) to accomplish controlled phase separation and get around thermodynamic incompatibility. The integrity of both bulk and surface characteristics is thought to be ensured by the interlocking structure of the cross-linked IPN components (Maolin et al. [2000\)](#page-18-5). IPNs are primarily described as the close union of two polymers in the presence of at least one being instantly synthesised in the presence of the other one (Ullah et al. [2015](#page-19-1)).

#### **Advances in Hydrogels (Based on Stimuli)**

Environmental-specifc, intelligent hydrogels are hydrogels that react to external stimuli and undergo unanticipated alterations to their growth processes, the structure of networks, mechanical properties, and permeability (Gil and Hudson [2004](#page-17-5)). Chemical stimuli that change the interactions among polymeric chains and solvents as well as polymer networks at the molecular scale include the pH, ionic factors, and chemical substances. A biochemical stimulation responds to ligands, enzymes, antigens, and other biochemical agents. Damiri et al. ([2022b\)](#page-16-7) created glucose and pH-responsive hydrogels with chitosan and 4- 4-formylphenylboronic acid for the controlled release of insulin. Physical stimuli, which alter molecular level interactions at crucial points, include light, temperature, pressure, mechanical stress, magnetic and electric felds, and intensity of diferent energy sources (Ullah et al. [2015](#page-19-1)). Using UV light irradiation (PVA-MA), thermally sensitive poly(N-isopropyl acrylamide) PNIPAM was attached to the robust polymer backbone of poly (vinyl alcohol) (PVA) and its methacrylate derivatives. After the one-pot polymerization, the samples were submerged within a sodium sulphate salt solution to further strengthen the hydrogel network, where the PVA clumps and crystallises under these conditions (Pardeshi et al. [2022](#page-18-6)). Other than these basic parameters, hydrogels can also be classifed as conventional and smart based on physical properties, and non-biodegradable, biodegradable hydrogels, depending upon the degradation pattern.

## **Properties of Hydrogels**

Hydrogel has numerous properties which contribute to feld applications. Polymer hydrogels are often fragile, soft, and brittle and are unable to sustain signifcant deformations. A hydrogel's structure and composition are crucial factors in determining its mechanical strength (Shibayama [2012\)](#page-19-10). The medical and healthcare sectors can use hydrogels because of their mechanical properties, which include maintaining the physical texture of therapeutic ingredients during administration. This is important for biomedical applications like ligament and tendon repair, drug delivery matrix, dressing material for wounds, repair of tissue, and cartilage replacement material (Mishra et al. [2018\)](#page-18-7). Future applications of the matrix of hydrogel in the drug, medical, eye disease, and tissue repair sectors will be characterized by its swelling properties (Zhang et al. [2020\)](#page-20-0). Biodegradation is an essential component of biomedical applications, which calls for controlled absorption in vivo and targeted dispersion to support cell morphogenesis and motility. However, a variety of mechanisms (including the process of hydrolysis, the breakdown of proteins, disentanglement, or environmental

stimuli) could cause local or bulk breakdown of hydrogels; it is difficult to engineer the spatiotemporal aspects. Connecting massive hydrolytic dissolution to particular periodic occurrences within the human body, like the regeneration of bones, can improve the repair of tissues by regulating the amount of hydrolytically responsive linkages in the network of polymers (Seliktar [2012\)](#page-19-0). Hydrogels can be used as tissue adhesives in surgical wound healing or as inductive scaffolds for the regeneration of tissue because of the signifcant property of bio adhesion, which also allows cells and tissues to attach to hydrogels. The majority of other naturally occurring and manufactured hydrogels do not have bio-adhesive qualities, even though some hydrogels, such as collagen or fbrin, do. Crosslinking molecules that permit covalent or non-covalent molecular connections between the implant and the surrounding environment, such as cell-adhesive oligopeptides generated from fbronectin's core cell-binding domains, can be used to build cell-adhesive characteristics into a hydrogel network (Censi et al. [2012\)](#page-16-8). Hydrogels must be both toxic-free and compatible to be employed by the biomedical industry. To be employed for this purpose, the majority of polymeric substances have to undergo in-vivo toxicity tests. Biosafety and bio-functionality are two factors that go into a biocompatibility examination. The former indicates a sufficient host reaction, including both local and systemic (i.e., surrounding tissue) responses, as well as the lack of cytotoxicity, mutant development, and carcinogenesis, whereas the latter shows the material's ability to carry out the desired task (Dasgupta et al. [2013\)](#page-16-9). Nowadays seeing the importance of proteins, these are utilized for hydrogel formation and have various biomedical and food applications. The following section provides a detailed explanation of proteins utilized for the preparation of hydrogels.

#### **Protein Hydrogels**

All organisms, from microorganisms to primates and larger animals, depend on proteins as key macromolecules. Protein can make up as much as 50% of the dry mass of some cells. The proteins connect to diferent biomolecules such as lipids, carbohydrates, phosphate bonds, DNA and metal ions by covalent or non-covalent interactions (Panahi and Baghban-Salehi [2019\)](#page-18-8). DNA is the genetic blueprint for protein synthesis for a cell's genetic material and all living cells synthesize proteins using replication, transcription, and translation. The basic primary structure of a protein molecule consists of a linear sequence of polypeptides which may contain up to 20 diferent amino acids. These amino acids possess other functional groups that can be chemically modifed (Whitford [2005](#page-20-3)). Several biomaterials, including hydrogels, flms, and composites, have been developed (Ashfaq et al. [2022\)](#page-16-10). Depending on their applications, hydrogels are manufactured in various shapes, including cubes, hollow tubes, rods, sheets, and flms (Shi et al. [2014](#page-19-11)). According to Totosaus et al. [\(2002](#page-19-12)), protein-based hydrogels are made through (Table [1\)](#page-6-0) enzymatic, chemical, or physical processes (heating, cooling, high pressure, etc.). The unfolding of the native protein structure and development of a gel network is the most signifcant process in the protein's gelation process. The constructed network can store water within its framework. Hydrophobic, electrostatic, and hydrogen bonding are examples of non-covalent cross-links that can keep these gel networks stable. Additionally, these could also be stabilized through covalent cross-linking (Le et al. [2017](#page-18-9)).

#### **Animal Protein**

The extracellular matrix contains collagen as a natural polymer. Around 30% of all the proteins in animals are made up of animal collagen, which is present in the dermis, bones, and articular tissues of animals. Collagen is primarily a fbrous protein, responsible for critical mechanical functions throughout the body (Gyles et al. [2017\)](#page-17-2). Bone and cartilage contain high concentrations of collagen, which is essential to the structure and function of these tissues. Additionally, collagen can be used to connect and release cell regulators like cytokine and growth elements. Biomaterials such as collagen are frequently employed in tissue engineering (Cen et al. [2008\)](#page-16-11) which is utilized both in its original fbrillar and in denatured state. The latter produces several collagen shapes, such as sheets, tablets, pellets, and sponges. In addition to its utility in tissue engineering, collagen has been utilized as a protein, medication, and gene delivery method (Aigner and Stöve [2003](#page-16-12)). Gelatin is created when collagen's triple-helix structure is broken down into single-strand molecules. There are two varieties of gelatin, types A and B. The former is produced after acidic pre-treatments of collagen and thermal decomposition, while the latter is produced after alkaline treatment. This process transforms glutamine and asparagine residues into glutamic and aspartic acids, yielding a greater concentration of carboxylic acid in gelatin B (Jonker et al. [2012\)](#page-17-6). A simple temperature change is all it takes to gel a gelatin solution. Gels made from gelatin are non-immunogenic, biodegradable, and biocompatible; making them good candidates for use in biomedicine. On the other hand, these have poor thermal and mechanical stability. Chemical changes, such as cross-linking, improve the stability of these for long-term biological uses and several tissue engineering applications employ gelatin-based materials (Kuijpers et al. [2000](#page-18-10)). The ability of collagen-based hydrogels to replace or temporarily repair injured or diseased tissues makes them appealing for use in hemostasis devices and wound dressings (Doillon et al. [1997\)](#page-17-7). However, the application possibilities for collagen-based hydrogels are constrained by their rapid in-vivo breakdown and low mechanical strength. Because of

## <span id="page-6-0"></span>**Table 1** Hydrogel materials and their methods of production



**Table 1** (continued)



this, recent research has concentrated on developing composite hydrogels that incorporate collagen composites with other polymers that have improved mechanical strength and degradation (Sahiner et al. [2014\)](#page-19-16). It is possible to create collagen-based hydrogels by physical processes. The reforming of fbrils and the formation of a hydrogel network is caused by the aggregation of collagen through covalent bonding, which is initiated by heating and an increase in pH. However, collagen in real tissues forms inter and intramolecular covalent connections via hydroxylysine as well as lysine residues, these hydrogels are weak in terms of their mechanical strength wherein additional cross-linking is often regarded as the most efective strategy for overcoming this hindrance (Antoine et al. [2014\)](#page-16-14). Similarly, these were additionally cross-linked with gamma radiation, carbodiimide, genipin, glutaraldehyde, and these substances. Several compounds were used to synthesize standard, hybrid or composite collagen hydrogels to modify the properties of conventional hydrogels (Chen et al. [2016](#page-16-15)).

The majority of the cytoplasmic epithelium and the outermost layer of the epidermis is made of the fbrous protein group keratin, which is abundant in animal hair, fngernails, fbres such as wool, horns, and feathers—all of which contain over 90 per cent by weight. These products can be used in a wide range of industrial settings. It can be divided into two categories: hard keratin and soft keratin. The stratum corneum layer in the skin, which is an example of epidermal keratin, is known as soft keratin and includes 1% sulphur, whereas the latter is found in strands of hair nails, horns, and feathers and has about 5% sulphur. The high cysteine content of the hard keratin gives it greater durability and

structural stability (Shavandi et al. [2017\)](#page-19-17). Keratin-based materials, in contrast to collagen-based ones, are mechanically strong and biodegradable in vivo. As a result, it can be used to build scafolds for permanent cell culture. For example, keratin-based hydrogels have drawn a lot of interest in the discipline of peripheral regeneration of nerves. In this instance, keratin-based hydrogels can be introduced inside a nerve channel as a fller to direct nerve regeneration. Regenerative scafolds and injectable drug delivery devices are two more uses for keratin-based hydrogels (Rouse & Van-Dyke [2010\)](#page-19-4). The keratin needs to be frst cross-linked, either chemically or by the formation of disulfde bonds, to produce these hydrogels. For instance, to hasten the formation of these bonds, the keratin solution could be combined with a hydrogen peroxide solution. The combination that results is then incubated for an additional day to create a gel. In a diferent study, both potassium persulfate, as well as sodium bisulfte, were added to a solution of hydrolyzed feather keratin as initiators. (Wattie et al. [2018\)](#page-19-18).

#### **Milk Protein**

Casein, the most abundant protein in milk, includes approximately 94 per cent of protein combined with colloidal calcium phosphate. Milk-based hydrogels can also be made with casein (proline-rich open-structured phosphoproteins from milk) only or it can be combined with other food-grade polymers. Caseins may self-assemble or aggregate into various protein structures (Dickinson [2006](#page-17-10)) and are sensitive to pH fuctuations. It is generally insoluble, however, becomes solubilized above pH 5.5 and below pH 3.5, while it is insoluble between 4.5 and 5.0 pH (Nascimento et al. [2020\)](#page-18-14). In addition to other positive traits like biocompatible, non-toxic, and reactive sites are available for alteration, caseins, & casein derivatives contain a majority of hydrophilic functional groups, such as lysine and glutamine residues, which may impart signifcant functional attributes to the hydrogel (Gorji et al. [2014](#page-17-1)). The addition of chemicals such as crosslinking agents can generate a variety of casein-based hydrogels wherein the amount of agent makes a profound impact (Song et al. [2009](#page-19-20)). Casein hydrogels can also be made via enzymatic cross-linking. To gel the milk proteins, enzymes such as transglutaminase could be utilized at relatively mild conditions resulting in a hydrogel suitable for biomedical applications (Elzoghby et al. [2011](#page-17-11)).

The remaining milk proteins constituting whey proteins have many beneficial features, including highly functional, biological, and nutritional. Whey protein's primary constituents are globular proteins, such as lactoglobulin and lactalbumin. For the creation of protein-based hydrogels, numerous studies have documented the use of whey protein products, particularly whey protein concentrate and isolate, or other whey constituents (especially lactoglobulin and bovine serum albumin), to encapsulate and distribute bioactive molecules and micronutrients (Deeth and Bansal [2019\)](#page-16-18). Therefore, cold set hydrogel-forming properties of whey proteins are well-known wherein acidulating agents like glucono-d-lactone (GDL) or salt-induced cold gelation are used to create soluble aggregates through a two-step process that begins with heating and culminates with cooling (acid-induced cold gelation) (Brodkorb et al. [2016\)](#page-16-19). Hydrogels produced by the cold-set gelation method are capable of providing efficient delivery of heat-sensitive nutraceutical ingredients like vitamins and probiotics better than heat-set gels can. More recently, Alavi et al. [\(2018](#page-16-20)) created a new gelation technique based on radical protein cross-linking, doing away with the heating stage and causing non-heated whey proteins to gel. whey protein-based hydrogels created by cold-set gelation technique are expected to have poor mechanical strength, which makes them susceptible to enzyme degradation.

#### **Plant Protein**

Plant proteins are receiving more attention in the food industry as a result of consumers' growing concerns about the safety of products made from animal products and their evolving dietary habits and culinary preferences. Being unstable at around their isoelectric point, these proteins do not function well in acidic pH conditions (Hertzler et al. [2020\)](#page-17-12). Corn (also known as maize) is one of the most extensively produced crops on the planet. In addition to proteins like glutelin, globulins, and albumins, zein is a vital storage protein present in the endosperm of maize kernels. In maize

kernels, it accounts for 44–79 per cent of the total protein content (on a wet basis) (Shukla and Cheryan [2001](#page-19-21)). The signifcant percentage of non-polar amino acids in nature (about 55%) makes it a hydrophobic protein. Zein has a regular geometry and is amphiphilic, which allows it to assemble itself into chains, sections, or foam by various processing techniques. It is possible to encourage the controlled aggregation of zein protein molecules, which results in a zein layer, on a surface that is hydrophobic (such as metal, glass slides, or manufactured polymer). This aggregation tendency is caused by the link between the hydrophobic groups of zein and the hydrophobic region (Dong et al. [2013\)](#page-17-13). Zein protein molecules remain to form aggregates on a hydrophobic surface after that, eventually forming a flm layer or a gel structure. Zein's ability to self-assemble into a variety of microstructures, such as flms and fbres, is a result of its amphiphilic nature. Because of its one-of-a-kind quality, it is possible to synthesize materials (hydrogels) that have the potential to be used in several areas, particularly in the encapsulation of drugs and controlled drug delivery (Sousa et al. [2012](#page-19-22)).

Soy protein is a signifcant by-product of the soybean oil industry. Owing to its excellent nutritional content, emulsifying capacities, and gel formation properties, it bestows good functional properties for food preparation. Soy protein has an isoelectric point of pH 4.8, which is the prime reason for its limited applications in most foods that are acidic in nature. Typically, soybean comprises 15–25 per cent oil, 35–45 per cent protein, 33–35 per cent carbohydrate (soluble and insoluble), and 5–6 per cent ash on a dry basis (Liu et al. [1997](#page-18-15)). It is distinguished by its propensity for forming gel and foam, which is crucial for creating porous hydrogels. These porous hydrogels are useful for the development of tissues, nutrient transportation, and the creation of hygienic products because they feature large and interconnected pores, a larger surface domain, and an excellent swelling rate (Kang et al. [2005\)](#page-17-14) (Santin and Ambrosio [2008](#page-19-23)).

Furthermore, hydrogels made from soy protein have signifcant applications in controlled compound release, orthopaedic implants, and superabsorbent materials (Zohuriaan-Mehr et al. [2009](#page-20-6)). Soy proteins can gel by a variety of methods, including heat gelation, cold setting gelation, chemical and enzymatic crosslinking, and others. Because of the unfolding and exposing of hydrophobic areas following thermal treatment, soy protein molecules can interact and combine to produce stable hydrogels (Puppo and Anon [1998](#page-19-24)). Another method of gelation, known as the cold gelation process, globular protein mixture (heat-denatured) after inducing gelation at room temperature by reducing pH or adding calcium is created. The quantities of soy and calcium used determine the properties of the resultant hydrogel (Alting et al. [2002\)](#page-16-21). For example, a larger protein concentration can enhance the gel's ability to hold water, whereas

a higher  $CaCl<sub>2</sub>$  (calcium chloride) concentration can raise the gel's opacity and elastic modulus. But soy hydrogels created through thermal gelation or cold gelation are weak mechanically and lose their strength soon. Enzymatic or chemical crosslinking can be used to improve this (Maltais et al. [2005](#page-18-2)). The removal of toxic substances from industrial effluents is one potential application of hydrogels made of soy. To create hydrogels with greater stability and cellular afnities, soy protein isolate (SPI) is also combined with collagen, zein, and other proteins. To get the appropriate hydrogel characteristics and reduce the amount of soy protein contaminants, extraction conditions should be optimised for usage as biomaterials (Panahi and Baghban-Salehi [2019](#page-18-8)).

## **Composite Hydrogels**

Hydrogels made from natural biomaterials or synthetic polymers cannot accurately duplicate all of the characteristics of the original extracellular matrix for tissue engineering and offer some disadvantages in other food and biomedical applications because of the inherent limitations of both types of hydrogels. Hydrogels comprise both synthetic and natural components. Combining the advantages of both synthetic and natural polymers (for example, cell-adhesive compounds and fbrous structure) will result in a composite hydrogel (Afewerki et al. [2019](#page-16-22)). Furthermore, naturally generated polymers, which include collagen and gelatin, are prone to change from batch to batch, have subpar mechanical properties, and have a low level of biochemical stability (Pradhan et al. [2016\)](#page-19-25). Several studies (Chen et al. [2015](#page-16-23); Nonoyama et al. [2016;](#page-18-16) Zhang et al. [2016](#page-20-2)) addressed the functions and advantages of double networks or interpenetrating networks made of natural-natural, synthetic-natural, and syntheticsynthetic hydrogels. Marine-derived collagen and agarose were recently added to alginate in an investigation including bioactive gel for 3-D cell culturing to develop a physically crosslinked hydrogel. With a range of cell types, this composite hydrogel showed outstanding cytocompatibility. The multicellular structures that developed within this composite hydrogel were what produced exceptional yields (Shin et al. [2016](#page-19-26)).

Furthermore, polysaccharides can be combined with gelatin to increase their bioactivity (Afewerki et al. [2019\)](#page-16-22). Due to its antibacterial qualities, biocompatibility, and biodegradability, chitosan is a desired polysaccharide (Mohanty et al. [2018](#page-18-17)), although can be improved by adding gelatin because it has poor bioactivity and sluggish gelation. At pH 6.5, positively charged chitosan can combine with gelatin (negatively charged) to produce polyelectrolyte complexes. The bioprinting of a structure used a mixture of alginate and gelatin as bio-ink. This printed construct's mechanical characteristics were comparable with those of pre-crosslinked alginate, but it was superior in promoting cell growth (Chung et al. [2013\)](#page-16-24). Li et al. ([2021](#page-18-18)) investigated the gels of the sodium alginate (Alg)/casein composite produced by glucono-dlactone (GDL) and their physical characteristics. The results showed that as the Alg concentration increased, the gel hardness initially increased but then decreased, corresponding to the structural transition of the composite gels. The gels had 8.0 per cent, w/v casein concentration and 0–1.0 per cent, w/v Alg concentration. At low Alg concentrations (0.1 per cent), the composite gel had a casein-dominant network; however, when Alg concentrations rose (e.g., 0.2–0.5 per cent), the caseins took over as the continuous gelling phase and the Alg served as the dispersed phase. Wei et al. ([2016\)](#page-20-7) studied the correlation of structure and pH in sodium caseinate (SC) and N, O-carboxymethyl chitosan (NOCC) based composite hydrogel. According to rheological experiments, the optimal hydrogel ratio was  $SC/NOCC = weight$  ratio 3/7. AFM confrmed the well-developed three-dimensional network structure of hydrogel. The resultant hydrogel also exhibited outstanding pH sensitivity. Temperature variations had a signifcant impact on the gelation process; a high temperature led to an earlier start to gelation.

## **Methods for the Preparation of Protein Hydrogels**

#### **Heat‑Induced Hydrogels**

When protein solutions are heated, the unfolding and disruption of protein molecules are triggered, leading to the exposure of hydrophobic groups and the creation of disulfde bonds. In conclusion, the production of heat-set gels is caused by the association of the unfolded protein molecules and the involvement of intermolecular-sheet topologies (Oztop et al. [2014\)](#page-18-19). A network is generated in whey protein heat-induced gels which include hydrogen bonds, hydrophobic and ionic interactions, along with covalent (intermolecular-and intramolecular disulfde bonding) and non-covalent (ionic and hydrophobic) interactions (Oztop et al. [2014\)](#page-18-19). The whey protein isolate (WPI) and lotus root amylopectin (LRA) composite gels were made utilising a similar heat-induced method and possessed high values for the strength of the gel, capacity for holding water, storage modulus, and better rheological properties (Liu et al. [2017](#page-18-11)). LRA was heated to 80 $\degree$ C in this method after being kept for 12 h at  $4 \,^{\circ}\text{C}$ . After that, to stabilise the matrix within the WPI-LRA composite hydrogels, the LRA solution was combined with the WPI solution and heated at 95 °C for 30 min in a water bath. Using cellulose nanocrystals that were derived from wheat bran, Xiao et al. ([2020](#page-20-4)) improved the characteristics of heat-induced whey protein gels to create a hybrid gel with a compressed and stable structure. By

using the heat-induced approach, Liu et al. ([2020a\)](#page-18-12) investigated the gel characteristics and microstructure by comparing thick egg whites (TKEW) and thin egg whites (TNEW). The results showed that the heat denaturation temperature of TKEW was greater than that of TNEW (72.51 °C). In addition, the heat-induced approach was used to examine the rheological characteristics of hydrogels made from pea protein isolate (PPI) using microbial transglutaminase (MTGase) as a crosslinking agent (Shand et al. [2008\)](#page-19-13). After heating the PPI hydrogel at 92 °C, MTGase was added and incubated at 4 and 50 °C, respectively. The results demonstrated that PPI gels incubated at 4 °C had higher shear stress and strain than those at 50 °C. To sum up, the temperature of heating and the heating time frame are two crucial factors that afect the structure, characteristics, uses, and applications of hydrogels in heat-induced hydrogels.

#### **Cold‑Set Induced Hydrogels**

Whenever the protein-containing liquid undergoes heating at a neutral pH level (above the isoelectric point), beneath the protein gelling extent, and with low ionic strength, the initial step of cold-set gel formation starts in globular food proteins, leading to denaturation and partial unfolding of proteins (Abaee et al. [2017\)](#page-15-0). The next step is either salt supplementation (mono, di, and multivalent cations) (saltinduced cold gelation) or acidifcation to reach the protein's isoelectric pH to reduce inter-protein repulsion and create cross-linkages (divalent and multivalent cations) between clusters of proteins (Kuhn et al. [2010](#page-18-20)). A study showed how globular proteins can be cold-gelled using calcium chloride and sodium chloride (Kuhn et al. [2011](#page-18-21)). To create whey protein-based cold-set gels with increased nutritional value and functioning, other cations, such as iron and magnesium (divalent), have also been used (da Silva et al. [2010](#page-16-13)). One of the primary benefts of cold-induced gel formation over other techniques is the ability to interact bioactive ingredients with polypeptide chains through a variety of interactions that include hydrogen bonding, electrostatic force, and hydrophobic interactions due to exposure to various functional groups inside protein molecules. According to Abaee et al. [\(2017\)](#page-15-0), targeted delivery systems can be developed through these newly established connections between bioactive chemicals and proteins. Additionally, the cold-induced method gives the fnal shape, structure, and texture of the generated gel (Egan et al. [2014\)](#page-17-15).

#### **Pressure‑Induced Hydrogels**

Food proteins can also be gelated via high-pressure processing, a unique method demonstrated by several workers. Proteins are denatured by high-pressure processing by rupturing electrostatic and hydrophobic interactions. In comparison to

the heat-induced gelation approach, it has been suggested to ofer fewer advantages because it solely afects the breaking of hydrogen bonds (Ngarize et al. [2005](#page-18-22)). These advantages include the production of a softer and smoother texture as well as the prevention of browning reactions (Apichartsrangkoon [2003\)](#page-16-25). In a study by Semasaka et al. ([2019\)](#page-19-15), bovine serum albumin, also known as BSA, and gelatin were combined to form a composite hydrogel at 10 and 80 °C for 15 min, respectively. The fndings showed that at 10 °C, a continuous gelatin network structure with dispersed BSA inclusions existed, whereas, at 80 °C, an uninterrupted BSA reversed dispersion phase with supportive gelatin inclusions existed. The unique characteristic of the gelatinization of starch procedure carried on alternatively by extreme pres-sure or heat was also revealed by Liu et al. ([2020a\)](#page-18-12). They discovered that starch was gelatinized in diferent ways as a result of high pressure and heat. High temperature-induced the gelatinization process selectively disrupted unstable crystallites whereas high pressure-induced the gelatinization process disturbed crystallites of all stabilities. Hydrogel structure and gelation properties were mainly studied using high-pressure processing. However, for use as a delivery mechanism for bioactive components, this hydrogel production process will need to be improved signifcantly in the future.

#### **Enzyme‑Induced Hydrogels**

Enzymatically crosslinked hydrogels have been receiving a growing amount of attention for a few decades. The majority of enzymes are capable of catalyzing reactions that are enzymatic at neutral pH, in an aqueous environment, and at mild temperatures, suggesting that they can be used to create in situ hydrogels (Teixeira et al. [2012\)](#page-19-27). Due to one of the best qualities of this sort of reaction, undesirable side reactions or toxicity could happen while employing photoinitiator compounds or organic solvent compounds. Also, enzymes specifcally target a substrate (Davachi et al. [2022](#page-16-26)). Because of the relatively mild form of the enzyme-mediated processes that occur under typical physiological settings, this emphasises the beneft of this technique in interconnecting organic polymers, which are unable to endure strong chemical conditions. Therefore, any potential loss of bioactivity is avoided as a result. Sofa et al. ([2002](#page-19-28)) found that modulation of enzyme activity can directly regulate the polymerization reaction. Thiol enzymes called transglutaminases to catalyse post-translational protein modifcation, mostly by causing the formation of isopeptide bonds, but also by covalently conjugating polyamines, esterifying lipids, and deaminating glutamine residues (Krishna and Wold [1998](#page-17-16)). Transglutaminases catalyse the production of covalent connections between the amino acid glutamine (peptide-bound) or carboxylic group and the free amine group of the protein,

<span id="page-11-0"></span>**Fig. 2** Layout of potential applications of hydrogels in various felds



serving as a substitute for chemical crosslinking. These connections are highly resistant to proteolytic breakdown once they have been established. As a result, co-factors are not required to create stable polymeric networks (Teixeira et al. [2012](#page-19-27)). Fish protein (Li et al. [2019\)](#page-18-23), whey protein (Agyare and Damodaran [2010\)](#page-16-16), casein (Duerasch et al. [2018](#page-16-27)), gelatin (Pilipenko et al. [2019](#page-19-19)), soy protein (Zhang et al. [2020](#page-20-0)), and chitosan have all been used as a result of transglutaminase in recent years (Jiang et al. [2016](#page-17-17)).

## **Applications of Protein Hydrogels**

Hydrogel has several applications in various felds, (Fig. [2](#page-11-0); Table [2](#page-12-0)) and under this section, applications in biomedical sectors, food sectors and agriculture sectors are covered.

## **Biomedical Sector**

Hydrogels, nanofibers, hydrogel nanofibers, foam and spongy, bilayered, and trilayered scafolds are some of the biomaterials being created and studied to hasten the healing of chronic wounds (Ijaola et al. [2022](#page-17-18)). Hydrogels composed of gelatin and hyaluronic acid (HA) which have been cross-linked in diferent ratios have been studied for wound healing using both vivo and in vitro wound repair models to evaluate their efectiveness. environmentally friendly gelatin/hyaluronic acid hydrogels may be useful as wound dressings due to enhanced cell growth in vitro compared to controls and wound healing in vivo (Wu et al. [2017](#page-20-8)). In situforming gelatin and oxidised alginate, hydrogels were the subject of another study as potential wound-healing material. The fndings showed that a modest amount of borax caused periodate oxidised alginate to quickly cross-link the hydrogel's proteins, causing gelatin to cross-link in the wound. (Balakrishnan et al. [2005\)](#page-16-28). Additionally, keratin from discarded human hair or wool was employed to create hydrogels that were then cross-linked using an electron beam in a mixture of polyvinyl alcohol (PVA) and polyethylene imine. Compared to the control group (blank or hydrocolloid wound dressings), injuries that received keratin-based hydrogel dressings closed more quickly, most likely as a result of the creation of new collagen (Park et al. [2015](#page-18-24)). Electro-spun nanofbrous mats made of casein and PVA successfully promoted haemostasis by generating thrombin whereas increased epithelial proliferation and restoration were achieved by adding casein (Biranje et al. [2019\)](#page-16-29). Additionally, wound healing has been aided by developing casein/

## <span id="page-12-0"></span>**Table 2** Applications of protein-based hydrogel



Applications	<b>Materials</b>	<b>Key Findings</b>	References
	Carotenoids delivery Rice starch -whey protein isolate	Beta-carotene bioaccessibility was compared between emulsions, hydrogels and filled hydrogels and was found to be more in filled hydrogels.	Mun et al. $2015$
	Zein-oligochitosan	Encapsulation of astaxanthin was done in the zein- oligochitosan complex having improved UV-light and storage stabilities.	Jiang and Zhu 2019
Fat replacers	Protein and dietary fibers	Hydrogel particles were formed using gelatin and pectin to mimic starch granules.	Wu et al. 2014
Agriculture	Whey protein (WP) and alginic acid (AA)	By utilizing WP and AA hydrogels were created by calcium chloride as a crosslinking agent for the sustained release of urea fertilizer in the crop.	Di Martino et al. 2021

**Table 2** (continued)

cellulose/chitosan scafolds. Nucleation sites for blood clotting ions were revealed by the presence of casein, which was needed for haemostasis (Biranje et al. [2022\)](#page-16-30). Hydrogels made from casein and alginates are being investigated for use in intravenous injection and can be used in wound treatment owing to their porous nature, helpful in improving cell survival (Patwa et al. [2020](#page-19-30)).

Hydrogels featuring a three-dimensional could be employed in tissue engineering, and have proven to be very promising materials. They are qualifed as potential candidates for implementation in tissue engineering techniques because they can mimic the physico-chemical characteristics of the original ECM, or extracellular matrix, (Lee and Mooney [2001\)](#page-18-26). Gelatin and alginate hydrogel nanofibers discussed by Majidi et al. ([2018](#page-18-25)) showed increased adhesion of cells, mobility, growth, and development, suggesting they can be useful in the treatment of stem cells and regeneration of tissues. An anisotropic collagen hydrogel nanofber was created by Wakuda et al. ([2018](#page-19-18)) wherein researchers employed a core-shell electrospinning process to meet the objective. It is vital to have anisotropy to closely approximate ECM, as a result of which the hydrogel nanofber system fnds applicability in tissue engineering scafolds.

Numerous ways to deliver drugs (DDS) have been developed to meet the challenge of polypeptide and protein drug delivery (McClements [2017\)](#page-18-2). Three-dimensional water-swollen polymeric structures are known as hydrogels. These types of DDS are created when polymeric chains interact physically or chemically. Due to their signifcant amount of water, they are utilised to administer a variety of drugs and biologically active substances (Khodaverdi et al.  $2017$ ). They offer several characteristics that make them ideal for medication delivery: (a) the ability to manufacture microparticles, nanoparticles, and flms in various physical shapes. (b) Their permeability facilitates the controlled release of drugs for smaller molecules or macromolecules across the hydrogel network and provides an ideal atmosphere for the incorporation of drugs

into the gel structure. (c) These include basic formulations, which allow for high local concentrations of encapsulated medications to stay in the tissues over time. (d) These are also quite biocompatible, which makes them perfect for application in vivo (Hoare and Kohane [2008](#page-17-23)). Natural hydrogels based on proteins, including gelatin, collagen, casein, and albumin, have been extensively studied and are regarded as an important DDS class (Elzoghby et al. [2011\)](#page-17-11). Bovine milk's primary ingredient, casein, is widely available, reasonably priced, and possesses several fascinating fundamental and physicochemical characteristics, such as hydrophilicity and the availability of reaction sites for later modifcation, which make it an appealing option for novel DDS (Song et al. [2009\)](#page-19-20). Various innovative DDS based on casein have recently been produced and successfully used, including self-assembled nanostructures, nanocomposite-based flms, crosslinked casein micellar compounds, and hydrogels. Another signifcant attribute of a hydrogel is its ability to store water, and it has been discovered that casein hydrogel has a lower tendency to swell (Kruif et al. [2015](#page-16-17)). Based on such findings, casein appears to be a suitable material for building biocompatible polymeric hydrogels for delivering labile molecules like peptides and proteins. Chitosan-based freeze-dried hydrogels for the delivery of drugs including cafeine, ascorbic acid, and 5-fuorouracil (5-FU) were developed. Results depicted that chitosan can be used for in-situ target therapy (Damiri et al. [2020\)](#page-16-31). Also, a similar study using chitosan and magnetite nanoparticles was conducted by Fouad et al. ([2020\)](#page-17-24). Hydrogel-forming microneedles are another smart approach for the transdermal delivery of drugs. The drug is delivered via a topical route using the microneedle delivery system by momentarily disturbing the skin's surface layer, which follows the difusion mechanism. On a tiny patch, an array of hundreds of microneedles is organized to help distribute enough medication to have a therapeutic impact.

#### **Food Sector**

There are several uses for hydrogels in the food business, including food packaging and acting as transporters for bioactive components. Hydrogels can offer antimicrobial benefts and control the humidity produced by the food when used in packaging materials. This makes it possible to create smart packaging that enhances the quality, protection, and potential of food to be preserved for a long time (Shewan and Stokes [2013\)](#page-19-31). To prepare packaging flms for fresh potatoes, flms containing a mixture of agar, collagen and alginate as well as antimicrobial ingredients like nanoparticles of silver and extract of seeds from grapefruit were mixed (Wang and Rhim [2015](#page-20-9)). The water absorption capacity of the fnished film was found to be 23.6 per cent higher than its weight. In addition, the flm demonstrated potent antibacterial action against the strain of Listeria monocytogenes (gram-positive) and the strain of Escherichia coli (gram-negative), both species are known to cause illness in humans. Caseinate and methoxyl pectin were complexed by electrostatic force by Zhang et al. ([2014](#page-20-1)) to produce hydrogel microspheres that contained fsh oil. First, a caseinate-based emulsion made from oil in water was created at pH 7 utilising caseinate as the emulsifer. Pectin was then added after the solution had been pH-adjusted to 4.5. Finally, to increase the longterm stability of the hydrogel matrix, the transglutaminase enzyme was employed as a cross-linking agent. Due to the encapsulation, which prohibited the polyunsaturated fatty acids (PUFA) from becoming oxidised, this showed the possibility of such hydrogel particles for delivering PUFA lipids in foods and beverages. At the same time, it does not afect their digestion in a small intestine model.

In addition, another important variable determining the quality of food products is how food is perceived to have been prepared. Hydrogel, a soft material, has many textural qualities (elasticity, hardness, chewiness, etc.). As a result, hydrogels can be utilised to enhance the texture and mouthfeel of foods in addition to substituting food components to reduce calorie consumption. For instance, emulsion hydrogels can change the textural properties of food (Devezeaux et al. [2016](#page-17-27)). It is interesting to note that a successful strategy for cutting calories in food is to substitute calorie-dense ingredients like meat or grain with calorie-light alternatives like hydrogels with excellent textural qualities or low oil content (Guo et al. [2013\)](#page-17-28).

Due to the three-dimensional porous networks, the hydrogel is the perfect encapsulating option for water-soluble components, which may retain as much as several thousand times their dry mass. Hydrogels have been extensively used in the food industry as encapsulation solutions throughout the past few decades (Zhang et al. [2016\)](#page-20-2). In contrast to the situation in the food business, the medical industry has conducted a large number of studies on the usage of hydrogels in encapsulation. Structurally, nanogels, hydrogels with core-shell structures, emulsifed hydrogels, microgels and hydrogel nanomaterials are all structurally suitable for use as encapsulation platforms. Both the research on utilizing hydrogels for encapsulation and the research on using hydrogels for immobilization (of enzymes, cells, microorganisms, and so on) share certain parallels, although, some diferences exist (Facin et al. [2015\)](#page-17-29).

On the one hand, while immobilisation can occur within the hydrogel matrix or on its surface, encapsulation almost typically occurs exclusively within the network of the hydrogel. Additionally, since hydrogels are a three-dimensional network of interconnected pores, both the exterior and interior structural characteristics of hydrogels have a signifcant impact on their ability to immobilise (Lim et al. [2019](#page-18-28)). An extremely permeable hydrogel with a high or even super high absorption capacity may attract more attention from the general population, visualizing an immediate demand for developing products with decreased amounts of fat or starch. Hydrogels can be an efective tool in calorie reduction, either by increasing satiety or decreasing intake (Cao and Mezzenga [2020\)](#page-16-32). Using proteins and fbre from food as light material approaches, Wu et al. ([2014\)](#page-20-10) produced hydrogel particles that had an excellent texture, that might serve as a more nutritious substitute for starchy fakes.

## **Enhancement of Nutrient Absorption and Digestibility of Hydrogels**

Proteins are commonly employed in producing hydrogels due to their technological properties, which allow them to be used in various culinary products. Dietary matrices and digestive environments infuence proteins because they are altered by pH, ionic strength, and proteolysis (Ozel et al. [2018](#page-18-29)). In the passage through the GIT (gastrointestinal tract), protein hydrogels can swell signifcantly under stomach conditions (Hashemi et al. [2017](#page-17-30)). Proteins with a pKa of 5 or below, may be associated with swelling during the gastric phase owing to the positive charges under the stomach's conditions, allowing the easy entry of water. However, it might be challenging to identify the cause of protein-gel disintegration, since it involves enzymatic activity and structural modifcations that cause gel collapse. The gel protein in GIT's gel can have its swelling and breaking properties adjusted by chemical cross-linking or heat-induced aggregation (Okuro et al. [2021](#page-18-30)).

Additionally, since chemicals used to make polysaccharide hydrogels primarily do not react to enzymatic action, other chemical and physical changes within the GIT specifcally stimulate modifcation (e.g. ionic strength, pH, shrinking and mechanical properties, swelling,). Gellan gum, alginate, pectin, and carrageenan are examples of anionic biopolymers that include carboxylic or sulphate groups,

making them particularly susceptible to the medium's pH and ionic strength (Okuro et al. [2021](#page-18-30)). Thus, the change in molecular conformation of these polysaccharides (which takes place during the passage through the GIT) can impact the macroscale and features associated with gels (Santos and Cunha [2018](#page-19-32)).

#### **Agricultural Sector**

Hydrogels have some potential applications in agriculture, which include the transport and precise distribution of minerals or agricultural fertilizing agents, the efficient use of wait and soil minerals, and the ability to provide desirable characteristics to agronomic goods without jeopardizing the ecosystem or its natural assets (Guilherme et al. [2015](#page-17-31)). The availability of moisture in the grounds allows for the supply of nutrients to vegetation that is essential for their growth. Even in dry soils, improving the environment for plant growth by increasing the amount of lost water is a benefactor. Hydrogels with superabsorbent properties can take in a substantial amount of water while maintaining their original size and weight. By decreasing the need for irrigation and the soil's propensity towards compaction, putting a stop to erosion and water run-of, boosting soil aeration and microbial activity, and so on, the use of these materials in agriculture could help to alleviate some of the issues that plague the agricultural industry (Rudzinski et al. [2002](#page-19-33)). Numerous potential uses for hydrogels exist in the area of water treatment, including the elimination of hazardous metals to be used further for irrigation purposes. Functional groups including sulfonic acid, carboxylic acid, amidoxime, amines, the hydroxyl group, and many more may function as chelating compounds for association with metallic ions present in these materials (Ekebafe et al. [2011\)](#page-17-32). Protein hydrogels can play several signifcant roles in agriculture that improve crop productivity, soil health, and sustainability as a whole. The study that follows shows how protein hydrogels are used in agriculture. Da Martino et al. [\(2021](#page-17-26)) utilised whey protein and diferent concentrations of alginic acid for hydrogel preparation to retain soil moisture and efficient release of urea.

#### **Conclusion**

According to the fndings of this review, proteins, and more specifcally food proteins are a great candidate for smart as well as composite hydrogels that are capable of releasing biological molecules in a variety of circumstances due to their intrinsic properties. Bioactive substances can be made more stable and bioavailable with the help of a food hydrogel based on a protein which could be an excellent candidate to cater to the requirement. Hydrogels based on proteins have numerous uses in the biomaterials, drug, and food industries. Even though food protein hydrogels have already been widely employed and researched as bioactive ingredient delivery methods, some major scientifc issues still need to be addressed for wider practical applications. Due to their water absorption and swelling properties, these hydrogels are ideal for encapsulating specifc molecules and releasing them under controlled conditions. A growing number of studies have been published in scholarly journals regarding the structure of hydrogel prepared from milk proteins. Food hydrogel's interactions with the body are still unresolved and understudied scientifc questions. This shows the wide range of possibilities that can be found in this subject of study. More inputs are required to further understand the structural features of protein-based hydrogels and discover industrial applications for these soft materials.

On the other hand, in terms of protein delivery approaches, hydrogels have several limitations, including irregular tissue formation, lack of spatial dissemination, cell dissociation and breakdown of proteins. Researchers should address the challenges associated with obtaining tailor-made drug releases and close the discrepancy between preliminary trials and results from clinical investigations. In the near times, studies must take into consideration the particular characteristics of the target tissues the attributes of Proteinbased Hydrogels (PBHs) and their level of sensitivity in the target tissue. It carries immense importance in the feld of protein characterisation leading to the importance of understanding the protein's structure, the advantageous associations among protein molecules and the matrix of polymers inside hydrogels, distribution strategies, and taking into account the importance of essential properties like biological compatibility. The potential to address complicated problems in healthcare, biotechnology, and other areas becomes more and more clear as research in this area develops. The scientifc community may further unlock the incredible potential of protein-based hydrogels and lead humanity into an era of transformative applications by accepting the recommendations for future research areas.

**Author contributions** Conceptualization – BM, RC Methodology- BM, RC, SK Original Draft Writing- BM Supervision- RC Editing- BM Review and Editing- RC, SK Resources- BM, SK.

#### **Declarations**

**Conflict of interest** The authors declare no competing interests.

## **References**

<span id="page-15-0"></span>Abaee A, Mohammadian M, Jafari SM (2017) Whey and soy protein-based hydrogels and nano-hydrogels as bioactive delivery systems. Trends Food Sci Technol 70:69–81. [https://doi.org/10.](https://doi.org/10.1016/j.tifs.2017.10.011) [1016/j.tifs.2017.10.011](https://doi.org/10.1016/j.tifs.2017.10.011)

- <span id="page-16-22"></span>Afewerki S, Sheikhi A, Kannan S, Ahadian S, Khademhosseini A (2019) Gelatin polysaccharide composite scafolds for 3D cell culture and tissue engineering: towards natural therapeutics. Bioeng Transl Med 4(1):96–11.<https://doi.org/10.1002/btm2.10124>
- <span id="page-16-16"></span>Agyare KK, Damodaran S (2010) pH-stability and thermal properties of microbial transglutaminase-treated whey protein isolate. J Agric Food Chem 58(3):1946–1953. [https://doi.org/10.1021/](https://doi.org/10.1021/jf903530d) [jf903530d](https://doi.org/10.1021/jf903530d)
- <span id="page-16-1"></span>Ahmed EM (2015) Hydrogel: preparation, characterization, and applications: a review. J Adv Res 6(2):105–121. [https://doi.org/10.](https://doi.org/10.1016/j.jare.2013.07.006) [1016/j.jare.2013.07.006](https://doi.org/10.1016/j.jare.2013.07.006)
- <span id="page-16-12"></span>Aigner T, Stöve J (2003) Collagens—major component of the physiological cartilage matrix, a major target of cartilage degeneration, major tool in cartilage repair. Adv Drug Deliv Rev 55(12):1569– 1593. <https://doi.org/10.1016/j.addr.2003.08.009>
- <span id="page-16-20"></span>Alavi F, Emam-Djomeh Z, Yarmand MS, Salami M, Momen S, Moosavi-Movahedi AA (2018) Cold gelation of curcumin loaded whey protein aggregates mixed with k-carrageenan: impact of gel microstructure on the gastrointestinal fate of curcumin. Food Hydrocoll 85:267–280. [https://doi.org/10.1016/j.foodhyd.2018.](https://doi.org/10.1016/j.foodhyd.2018.07.012) [07.012](https://doi.org/10.1016/j.foodhyd.2018.07.012)
- <span id="page-16-2"></span>Ali A, Ahmed S (2018) Recent advances in edible polymer based hydrogels as a sustainable alternative to conventional polymers. J Agric Food Chem 66(27):6940–6967. [https://doi.org/10.1021/](https://doi.org/10.1021/acs.jafc.8b01052) [acs.jafc.8b01052](https://doi.org/10.1021/acs.jafc.8b01052)
- <span id="page-16-21"></span>Alting AC, de Jongh HH, Visschers RW, Simons JWF (2002) Physical and chemical interactions in cold gelation of food proteins. J Agric Food Chem 50(16):4682–4689. [https://doi.org/10.1021/](https://doi.org/10.1021/jf011657m) [jf011657m](https://doi.org/10.1021/jf011657m)
- <span id="page-16-14"></span>Antoine EE, Vlachos PP, Rylander MN (2014) Review of collagen I hydrogels for bioengineered tissue microenvironments: characterization of mechanics, structure, and transport. Tissue Eng Part B: Reviews 20(6):683–696. [https://doi.org/10.1089/ten.teb.](https://doi.org/10.1089/ten.teb.2014.0086) [2014.0086](https://doi.org/10.1089/ten.teb.2014.0086)
- <span id="page-16-25"></span>Apichartsrangkoon A (2003) Efects of high pressure on rheological properties of soy protein gels. Food Chem 80(1):55–60. [https://](https://doi.org/10.1016/S0308-8146(02)00235-2) [doi.org/10.1016/S0308-8146\(02\)00235-2](https://doi.org/10.1016/S0308-8146(02)00235-2)
- <span id="page-16-10"></span>Ashfaq A, Jahan K, Islam RU, Younis K (2022) Protein-based functional colloids and their potential applications in food: a review. LWT 154:112667.<https://doi.org/10.1016/j.lwt.2021.112667>
- <span id="page-16-4"></span>Bae KH, Kurisawa M (2016) Emerging hydrogel designs for controlled protein delivery. Biomater Sci 4(8):1184–1192. [https://doi.org/](https://doi.org/10.1039/C6BM00330C) [10.1039/C6BM00330C](https://doi.org/10.1039/C6BM00330C)
- <span id="page-16-28"></span>Balakrishnan B, Mohanty M, Umashankar PR, Jayakrishnan A (2005) Evaluation of an in situ forming hydrogel wound dressing based on oxidized alginate and gelatin. Biomaterials 26(32):6335– 6342. <https://doi.org/10.1016/j.biomaterials.2005.04.012>
- <span id="page-16-29"></span>Biranje S, Madiwale P, Adivarekar RV (2019) Porous electrospun Casein/PVA nanofbrous mat for its potential application as wound dressing material. J Porous Mater 26(1):29–40. [https://](https://doi.org/10.1007/s10934-018-0602-7) [doi.org/10.1007/s10934-018-0602-7](https://doi.org/10.1007/s10934-018-0602-7)
- <span id="page-16-30"></span>Biranje SS, Sun J, Cheng L, Cheng Y, Shi Y, Yu S, …, Liu J (2022) Development of cellulose nanofbril/casein-based 3D composite hemostasis scafold for potential wound-healing application. ACS Appl Mater Interfaces 14(3):3792–3808. [https://doi.org/10.](https://doi.org/10.1021/acsami.1c21039) [1021/acsami.1c21039](https://doi.org/10.1021/acsami.1c21039)
- <span id="page-16-19"></span>Brodkorb A, Croguennec T, Bouhallab S, Kehoe JJ (2016) Heatinduced denaturation, aggregation and gelation of whey proteins. Advanced dairy Chemistry. Springer, New York, NY, pp 155–178. [https://doi.org/10.1007/978-1-4939-2800-2\\_6.](https://doi.org/10.1007/978-1-4939-2800-2_6)
- <span id="page-16-5"></span>Cai Y, Shen H, Zhan J, Lin M, Dai L, Ren C et al (2017) Supramolecular trojan horse for nuclear delivery of dual anticancer drugs. J Am Chem Soc 139:2876–2879. [https://doi.org/10.1021/jacs.](https://doi.org/10.1021/jacs.6b12322) [6b12322](https://doi.org/10.1021/jacs.6b12322)
- <span id="page-16-3"></span>Caló E, Khutoryanskiy VV (2015) Biomedical applications of hydrogels: a review of patents and commercial products. Eur Polymer J 65:252–267.<https://doi.org/10.1016/j.eurpolymj.2014.11.024>
- <span id="page-16-32"></span>Cao Y, Mezzenga R (2020) Design principles of food gels. Nat Food 1(2):106–118. <https://doi.org/10.1038/s43016-019-0009-x>
- <span id="page-16-6"></span>Catoira MC, Fusaro L, Di Francesco D, Ramella M, Boccafoschi F (2019) Overview of natural hydrogels for regenerative medicine applications. J Mater Sci: Mater Med 30(10):1–10. [https://doi.](https://doi.org/10.1007/s10856-019-6318-7) [org/10.1007/s10856-019-6318-7](https://doi.org/10.1007/s10856-019-6318-7)
- <span id="page-16-11"></span>Cen L, Liu WEI, Cui LEI, Zhang W, Cao Y (2008) Collagen tissue engineering: development of novel biomaterials and applications. Pediatr Res 63(5):492–496. [https://doi.org/10.1203/PDR.0b013](https://doi.org/10.1203/PDR.0b013e31816c5bc3) [e31816c5bc3](https://doi.org/10.1203/PDR.0b013e31816c5bc3)
- <span id="page-16-8"></span>Censi R, Di Martino P, Vermonden T, Hennink WE (2012) Hydrogels for protein delivery in tissue engineering. J Controlled Release 161(2):680–692.<https://doi.org/10.1016/j.jconrel.2012.03.002>
- <span id="page-16-23"></span>Chen Q, Zhu L, Chen H, Yan H, Huang L, Yang J, Zheng J (2015) A novel design strategy for fully physically linked double network hydrogels with tough, fatigue resistant, and self-healing properties. Adv Funct Mater 25(10):1598–1607. [https://doi.org/10.](https://doi.org/10.1002/adfm.201404357) [1002/adfm.201404357](https://doi.org/10.1002/adfm.201404357)
- <span id="page-16-15"></span>Chen Z, Du T, Tang X, Liu C, Li R, Xu C, …, Wu J (2016) Comparison of the properties of collagen–chitosan scafolds after γ-ray irradiation and carbodiimide cross-linking. J Biomater Sci Polym Ed 27(10):937–953. [https://doi.org/10.1080/09205063.](https://doi.org/10.1080/09205063.2016.1169478) [2016.1169478](https://doi.org/10.1080/09205063.2016.1169478)
- <span id="page-16-0"></span>Chirani N, Yahia LH, Gritsch L, Motta FL, Chirani S, Farè S (2015) History and applications of hydrogels. J Biomedical Sci 4(02):1–23
- <span id="page-16-24"></span>Chung C, Degner B, Decker EA, McClements DJ (2013) Oil-flled hydrogel particles for reduced-fat food applications: fabrication, characterization, and properties. Innov Food Sci Emerg Technol 20:324–334. <https://doi.org/10.1016/j.ifset.2013.08.006>
- <span id="page-16-13"></span>da Silva MV, Delgado JMPQ, Gonçalves MP (2010) Impact of Mg2 + and tara gum concentrations on fow and textural properties of WPI solutions and cold-set gels. Int J Food Prop 13(5):972–982. <https://doi.org/10.1080/10942910902927128>
- <span id="page-16-31"></span>Damiri F, Bachra Y, Bounacir C, Laaraibi A, Berrada M (2020) Synthesis and characterization of lyophilized chitosan-based hydrogels cross-linked with benzaldehyde for controlled drug release. J Chem. <https://doi.org/10.1155/2020/8747639>
- Damiri F, Kommineni N, Ebhodaghe SO, Bulusu R, Jyothi VGS, Sayed AA, …, Berrada M (2022b) Microneedle-based natural polysaccharide for drug delivery systems (DDS): progress and challenges. Pharmaceuticals 15(2):190
- <span id="page-16-7"></span>Dasgupta A, Mondal JH, Das D (2013) Peptide hydrogels. RSC Adv 3(24):9117–9149. <https://doi.org/10.1039/C3RA40234G>
- <span id="page-16-9"></span>Davachi SM, Haramshahi SMA, Akhavirad SA, Bahrami N, Hassanzadeh S, Ezzatpour S, …, Bagher Z (2022) Development of chitosan/hyaluronic acid hydrogel scafolds via enzymatic reaction for cartilage tissue engineering. Mater Today Commun 30:103230. <https://doi.org/10.1016/j.mtcomm.2022.103230>
- <span id="page-16-27"></span>Duerasch A, Wissel J, Henle T (2018) Reassembling of alkali-treated casein micelles by microbial transglutaminase. J Agr Food Chem 66(44):11748–11756
- <span id="page-16-26"></span>de Kruif CK, Anema SG, Zhu C, Havea P, Coker C (2015) Water holding capacity and swelling of casein hydrogels. Food Hydrocolloids 44:372–379.<https://doi.org/10.1016/j.foodhyd.2014.10.007>
- <span id="page-16-17"></span>Deeth H, Bansal N (2019) Whey proteins: an overview. Whey Proteins. <https://doi.org/10.1016/B978-0-12-812124-5.00001-1>
- <span id="page-16-18"></span>Devezeaux de Lavergne M, Strijbosch VM, Van den Broek AW, Van de Velde F, Stieger M (2016) Uncoupling the impact of fracture properties and composition on sensory perception of emulsionflled gels. J Texture Stud 47(2):92–111. [https://doi.org/10.1111/](https://doi.org/10.1111/jtxs.12164) [jtxs.12164](https://doi.org/10.1111/jtxs.12164)
- <span id="page-17-27"></span>Di Martino A, Khan YA, Durpekova S, Sedlarik V, Elich O, Cechmankova J (2021) Ecofriendly renewable hydrogels based on whey protein and for slow release of fertilizers and soil conditioning. J Clean Prod 285:124848
- <span id="page-17-26"></span>Dickinson E (2006) Structure formation in casein-based gels, foams, and emulsions. Colloids Surf A: Physicochem Eng Aspects 288(1–3):3–11. <https://doi.org/10.1016/j.colsurfa.2006.01.012>
- <span id="page-17-10"></span>Ding X, Yao P (2013) Soy protein/soy polysaccharide complex nanogels: folic acid loading, protection, and controlled delivery. Langmuir 29(27):8636–8644. [https://doi.org/10.1021/la401](https://doi.org/10.1021/la401664y) [664y](https://doi.org/10.1021/la401664y)
- <span id="page-17-9"></span>Doillon CJ, Drouin R, Côte MF, Dallaire N, Pageau JF, Laroche G (1997) Chemical inactivators as sterilization agents for bovine collagen materials. J Biomed Mater Res: Of J Soc Biomater Jpn Soc Biomater 37(2):212–221. [https://doi.org/10.1002/](https://doi.org/10.1002/(SICI)1097-4636) [\(SICI\)1097-4636](https://doi.org/10.1002/(SICI)1097-4636)
- <span id="page-17-7"></span>Dong F, Padua GW, Wang Y (2013) Controlled formation of hydrophobic surfaces by self-assembly of an amphiphilic natural protein from aqueous solutions. Soft Matter 9(25):5933–5941. <https://doi.org/10.1039/C3SM50667C>
- <span id="page-17-13"></span>Egan T, O'Riordan D, O'Sullivan M, Jacquier JC (2014) Cold-set whey protein microgels as pH modulated immobilisation matrices for charged bioactives. Food Chem 156:197–203. [https://](https://doi.org/10.1016/j.foodchem.2014.01.109) [doi.org/10.1016/j.foodchem.2014.01.109](https://doi.org/10.1016/j.foodchem.2014.01.109)
- <span id="page-17-15"></span>Ekebafe LO, Ogbeifun DE, Okieimen FE (2011) Polymer applications in agriculture. Biokemistri, 23(2)
- <span id="page-17-32"></span>Elzoghby AO, El-Fotoh WSA, Elgindy NA (2011) Casein-based formulations as promising controlled release drug delivery systems. J Controlled Release 153(3):206–216. [https://doi.org/10.](https://doi.org/10.1016/j.jconrel.2011.02.010) [1016/j.jconrel.2011.02.010](https://doi.org/10.1016/j.jconrel.2011.02.010)
- <span id="page-17-11"></span>Facin BR, Moret B, Baretta D, Belfore LA, Paulino AT (2015) Immobilization and controlled release of β-galactosidase from chitosan-grafted hydrogels. Food Chem 179:44–51. [https://doi.](https://doi.org/10.1016/j.foodchem.2015.01.088) [org/10.1016/j.foodchem.2015.01.088](https://doi.org/10.1016/j.foodchem.2015.01.088)
- <span id="page-17-29"></span>Fan Y, Zeng X, Yi J, Zhang Y (2020) Fabrication of pea protein nanoparticles with calcium-induced cross-linking for the stabilization and delivery of antioxidative resveratrol. Int J Biol Macromol 152:189–198. [https://doi.org/10.1016/j.ijbiomac.](https://doi.org/10.1016/j.ijbiomac.2020.02.248) [2020.02.248](https://doi.org/10.1016/j.ijbiomac.2020.02.248)
- <span id="page-17-8"></span>Fouad D, Bachra Y, Ayoub G, Ouaket A, Bennamara A, Knouzi N, Berrada M (2020) A novel drug delivery system based on nanoparticles of magnetite Fe 3 O 4 embedded in an auto cross-linked chitosan. Chitin and chitosan-physicochemical properties and industrial applications. IntechOpen, London
- <span id="page-17-24"></span>Gil ES, Hudson SM (2004) Stimuli-reponsive polymers and their bioconjugates. Prog Polym Sci 29(12):1173–1222. [https://doi.org/](https://doi.org/10.1016/j.progpolymsci.2004.08.003) [10.1016/j.progpolymsci.2004.08.003](https://doi.org/10.1016/j.progpolymsci.2004.08.003)
- <span id="page-17-5"></span>Gong CY, Shi S, Dong PW, Zheng XL, Fu SZ, Guo G, …, Qian ZY (2009) In vitro drug release behavior from a novel thermosensitive composite hydrogel based on Pluronic f127 and poly (ethylene glycol)-poly (ε-caprolactone)-poly (ethylene glycol) copolymer. BMC Biotechnol 9(1):1–13. [https://doi.org/10.1186/](https://doi.org/10.1186/1472-6750-9-8) [1472-6750-9-8](https://doi.org/10.1186/1472-6750-9-8)
- <span id="page-17-4"></span>González-Ferrero C, Irache JM, González-Navarro CJ (2018) Soybean protein-based microparticles for oral delivery of probiotics with improved stability during storage and gut resistance. Food Chem 239:879–888.<https://doi.org/10.1016/j.foodchem.2017.07.022>
- <span id="page-17-20"></span>Gorji SG, Gorji EG, Mohammadifar MA, Zargaraan A (2014) Complexation of sodium caseinate with gum tragacanth: effect of various species and rheology of coacervates. Int J Biol Macromol 67:503–511. <https://doi.org/10.1016/j.ijbiomac.2014.02.037>
- <span id="page-17-1"></span>Guilherme MR, Aouada FA, Fajardo AR, Martins AF, Paulino AT, Davi MF, …, Muniz EC (2015) Superabsorbent hydrogels based on polysaccharides for application in agriculture as soil conditioner and nutrient carrier: a review. Eur Polymer J 72:365–385. <https://doi.org/10.1016/j.eurpolymj.2015.04.017>
- <span id="page-17-31"></span>Gun'ko VM, Savina IN, Mikhalovsky SV (2017) Properties of water bound in hydrogels. Gels 3(4):37. [https://doi.org/10.3390/gels3](https://doi.org/10.3390/gels3040037) [040037](https://doi.org/10.3390/gels3040037)
- <span id="page-17-0"></span>Guo Q, Ye A, Lad M, Dalgleish D, Singh H (2013) The breakdown properties of heat-set whey protein emulsion gels in the human mouth. Food Hydrocoll 33(2):215–224. [https://doi.org/10.](https://doi.org/10.1016/j.foodhyd.2013.03.008) [1016/j.foodhyd.2013.03.008](https://doi.org/10.1016/j.foodhyd.2013.03.008)
- <span id="page-17-28"></span>Gupta C, Arora S, Syama MA, Sharma A (2017) Preparation of milk protein-vitamin a complexes and their evaluation for vitamin A binding ability. Food Chem 237:141–149
- <span id="page-17-21"></span>Gyles DA, Castro LD, Silva Jr JOC, Ribeiro-Costa RM (2017) A review of the designs and prominent biomedical advances of natural and synthetic hydrogel formulations. Eur Polymer J 88:373–392.<https://doi.org/10.1016/j.eurpolymj.2017.01.027>
- <span id="page-17-2"></span>Hashemi B, Madadlou A, Salami M (2017) Functional and in vitro gastric digestibility of the whey protein hydrogel loaded with nanostructured lipid carriers and gelled via citric acid-mediated crosslinking. Food Chem 237:23–29. [https://doi.org/10.](https://doi.org/10.1016/j.foodchem.2017.05.077) [1016/j.foodchem.2017.05.077](https://doi.org/10.1016/j.foodchem.2017.05.077)
- <span id="page-17-30"></span>Hertzler SR, Lieblein-Boff JC, Weiler M, Allgeier C (2020) Plant proteins: assessing their nutritional quality and efects on health and physical function. Nutrients 12(12):3704. [https://](https://doi.org/10.3390/nu12123704) [doi.org/10.3390/nu12123704](https://doi.org/10.3390/nu12123704)
- <span id="page-17-12"></span>Hoare TR, Kohane DS (2008) Hydrogels in drug delivery: progress and challenges. Polymer 49(8):1993–2007. [https://doi.org/10.](https://doi.org/10.1016/j.polymer.2008.01.027) [1016/j.polymer.2008.01.027](https://doi.org/10.1016/j.polymer.2008.01.027)
- <span id="page-17-23"></span>Iizawa T, Taketa H, Maruta M, Ishido T, Gotoh T, Sakohara S (2007) Synthesis of porous poly (N-isopropylacrylamide) gel beads by sedimentation polymerization and their morphology. J Appl Polym Sci 104(2):842–850.<https://doi.org/10.1002/app.25605>
- <span id="page-17-3"></span>Ijaola AO, Akamo DO, Damiri F, Akisin CJ, Bamidele EA, Ajiboye EG, …, Asmatulu E (2022) Polymeric biomaterials for wound healing applications: a comprehensive review. J Biomater Sci Polym Ed 33(15):1998–2050
- <span id="page-17-18"></span>Jiang GL, Zhu MJ (2019) Preparation of astaxanthin-encapsulated complex with zein and oligochitosan and its application in food processing. Lwt – Food Sci Technol 106:179–185. [https://doi.](https://doi.org/10.1016/j.lwt.2019.02.055) [org/10.1016/j.lwt.2019.02.055](https://doi.org/10.1016/j.lwt.2019.02.055)
- <span id="page-17-25"></span>Jiang SJ, Zhang X, Ma Y, Tuo Y, Qian F, Fu W, Mu G (2016) Characterization of whey protein-carboxymethylated chitosan composite flms with and without transglutaminase treatment. Carbohydr Polym 153:153–159. [https://doi.org/10.1016/j.carbp](https://doi.org/10.1016/j.carbpol.2016.07.094) [ol.2016.07.094](https://doi.org/10.1016/j.carbpol.2016.07.094)
- <span id="page-17-17"></span>Jonker AM, Löwik DW, Van Hest JC (2012) Peptide-and proteinbased hydrogels. Chem Mater 24(5):759–773. [https://doi.org/](https://doi.org/10.1021/cm202640w) [10.1021/cm202640w](https://doi.org/10.1021/cm202640w)
- <span id="page-17-6"></span>Kang HG, Lee SB, Lee YM (2005) Novel preparative method for porous hydrogels using overrun process. Polym Int 54(3):537– 543.<https://doi.org/10.1002/pi.1719>
- <span id="page-17-14"></span>Khodaverdi E, Javan M, Tabassi SAS, Khameneh B, Kamali H, Hadizadeh F (2017) Sustained drug delivery system for insulin using supramolecular hydrogels composed of tri-block copolymers. J Pharm Invest 47(3):263–273. [https://doi.org/10.1007/](https://doi.org/10.1007/s40005-016-0290-8) [s40005-016-0290-8](https://doi.org/10.1007/s40005-016-0290-8)
- <span id="page-17-22"></span>Krishna RG, Wold F (1998) Posttranslational modifcations. In Proteins (pp. 121–206). Academic Press. [https://doi.org/10.1016/](https://doi.org/10.1016/B978-012058785-8/50004-9) [B978-012058785-8/50004-9](https://doi.org/10.1016/B978-012058785-8/50004-9)
- <span id="page-17-16"></span>Krunić T, Obradović NS, Rakin MB (2019) Application of whey protein and whey protein hydrolysate as protein based carrier for probiotic starter culture. Food Chem 293:74–82. [https://](https://doi.org/10.1016/j.foodchem.2019.04.062) [doi.org/10.1016/j.foodchem.2019.04.062](https://doi.org/10.1016/j.foodchem.2019.04.062)
- <span id="page-17-19"></span>Kuhn KR, Cavallieri ÂLF, Da Cunha RL (2010) Cold-set whey protein gels induced by calcium or sodium salt addition. Int J Food Sci Technol 45(2):348–357. [https://doi.org/10.1111/j.](https://doi.org/10.1111/j.1365-2621.2009.02145.x) [1365-2621.2009.02145.x](https://doi.org/10.1111/j.1365-2621.2009.02145.x)
- <span id="page-18-20"></span>Kuhn KR, Cavallieri ÂLF, Da Cunha RL (2011) Cold-set whey protein–faxseed gum gels induced by mono or divalent salt addition. Food Hydrocoll 25(5):1302–1310. [https://doi.org/10.](https://doi.org/10.1016/j.foodhyd.2010.12.005) [1016/j.foodhyd.2010.12.005](https://doi.org/10.1016/j.foodhyd.2010.12.005)
- <span id="page-18-21"></span>Kuijpers AJ, Engbers GH, Krijgsveld J, Zaat SA, Dankert J, Feijen J (2000) Cross-linking and characterisation of gelatin matrices for biomedical applications. J Biomater Sci Polym Ed 11(3):225–243. <https://doi.org/10.1163/156856200743670>
- <span id="page-18-10"></span>Le XT, Rioux LE, Turgeon SL (2017) Formation and functional properties of protein–polysaccharide electrostatic hydrogels in comparison to protein or polysaccharide hydrogels. Adv Colloid Interface Sci 239:127–135. [https://doi.org/10.1016/j.cis.](https://doi.org/10.1016/j.cis.2016.04.006) [2016.04.006](https://doi.org/10.1016/j.cis.2016.04.006)
- <span id="page-18-9"></span>Lee KY, Mooney DJ (2001) Hydrogels for tissue engineering. Chem Rev 101(7):1869–1880. <https://doi.org/10.1021/cr000108x>
- <span id="page-18-26"></span>Li F, Li S, El Ghzaoui A, Nouailhas H, Zhuo R (2007) Synthesis and gelation properties of PEG – PLA – PEG triblock copolymers obtained by coupling monohydroxylated PEG – PLA with adipoyl chloride. Langmuir 23(5):2778–2783. [https://doi.org/10.](https://doi.org/10.1021/la0629025) [1021/la0629025](https://doi.org/10.1021/la0629025)
- <span id="page-18-18"></span>Li A., Guo C, Li X, Li P, Yang X, Guo Y (2021) Gelation mechanism and physical properties of glucono-δ-lactone induced alginate sodium/casein composite gels. Food Hydrocolloid 118:106775
- <span id="page-18-3"></span>Li J, Mo L, Lu CH, Fu T, Yang HH, Tan W (2016) Functional nucleic acid-based hydrogels for bioanalytical and biomedical applications. Chem Soc Rev 45(5):1410–1431. [https://doi.org/10.](https://doi.org/10.1039/C5CS00586H) [1039/C5CS00586H](https://doi.org/10.1039/C5CS00586H)
- <span id="page-18-0"></span>Li Y, Xiong S, Yin T, Hu Y, You J (2019) The gastric digestion kinetics of silver carp (Hypophthalmichthys molitrix) surimi gels induced by transglutaminase. Food Chem 283:148–154. <https://doi.org/10.1016/j.foodchem.2019.01.032>
- <span id="page-18-23"></span>Lim S, Jung GA, Muckom RJ, Glover DJ, Clark DS (2019) Engineering bioorthogonal protein– polymer hybrid hydrogel as a functional protein immobilization platform. Chem Commun 55(6):806–809. <https://doi.org/10.1039/C8CC08720B>
- <span id="page-18-28"></span>Liu K, Hettiarachchy N, Kalapathy U (1997) Soybean protein products. Soybeans: Chem, Technol Util. [https://doi.org/10.1007/](https://doi.org/10.1007/978-1-4615-1763-4_8) [978-1-4615-1763-4\\_8](https://doi.org/10.1007/978-1-4615-1763-4_8)
- <span id="page-18-15"></span>Liu K, Li QM, Pan LH, Qian XP, Zhang HL, Zha XQ, Luo JP (2017) The effects of lotus root amylopectin on the formation of whey protein isolate gels. Carbohydr Polym 175:721–727. [https://](https://doi.org/10.1016/j.carbpol.2017.08.041) [doi.org/10.1016/j.carbpol.2017.08.041](https://doi.org/10.1016/j.carbpol.2017.08.041)
- <span id="page-18-11"></span>Liu X, Wang J, Huang Q, Cheng L, Gan R, Liu L, Geng F (2020a) Underlying mechanism for the diferences in heat-induced gel properties between thick egg whites and thin egg whites: gel properties, structure and quantitative proteome analysis. Food Hydrocoll 106:105873. [https://doi.org/10.1016/j.foodh](https://doi.org/10.1016/j.foodhyd.2020.105873) [yd.2020.105873](https://doi.org/10.1016/j.foodhyd.2020.105873)
- Majidi SS, Slemming-Adamsen P, Hanif M, Zhang Z, Wang Z, Chen M (2018) Wet electrospun alginate/gelatin hydrogel nanofbers for 3D cell culture. Int J Biol Macromol 118:1648–1654. <https://doi.org/10.1016/j.ijbiomac.2018.07.005>
- <span id="page-18-12"></span>Maltais A, Remondetto GE, Gonzalez R, Subirade M (2005) Formation of soy protein isolate cold-set gels: protein and salt efects. J Food Sci 70(1):C67–C73. [https://doi.org/10.1111/j.](https://doi.org/10.1111/j.1365-2621.2005.tb09023.x) [1365-2621.2005.tb09023.x](https://doi.org/10.1111/j.1365-2621.2005.tb09023.x)
- <span id="page-18-25"></span>Maolin Z, Jun L, Min Y, Hongfei H (2000) The swelling behavior of radiation prepared semi-interpenetrating polymer networks composed of polyNIPAAm and hydrophilic polymers. Radiat Phys Chem 58(4):397–400. [https://doi.org/10.1016/S0969-](https://doi.org/10.1016/S0969-806X(99)00491-0) [806X\(99\)00491-0](https://doi.org/10.1016/S0969-806X(99)00491-0)
- <span id="page-18-2"></span>McClements DJ (2017) Recent progress in hydrogel delivery systems for improving nutraceutical bioavailability. Food Hydrocoll 68:238–245.<https://doi.org/10.1016/j.foodhyd.2016.05.037>
- <span id="page-18-5"></span>Mishra S, Rani P, Sen G, Dey KP (2018) Preparation, properties and application of hydrogels: a review. Hydrogels. [https://doi.org/](https://doi.org/10.1007/978-981-10-6077-9_6) [10.1007/978-981-10-6077-9\\_6](https://doi.org/10.1007/978-981-10-6077-9_6)
- Mohanty AK, Vivekanandhan S, Pin JM, Misra M (2018) Composites from renewable and sustainable resources: challenges and innovations. Science 362(6414):536–542. [https://doi.org/10.](https://doi.org/10.1126/science.aat9072) [1126/science.aat9072](https://doi.org/10.1126/science.aat9072)
- <span id="page-18-7"></span>Mun S, Kim YR, McClements DJ (2015) Control of β-carotene bioaccessibility using starch-based flled hydrogels. Food Chem 173:454–461.<https://doi.org/10.1016/j.foodchem.2014.10.053>
- <span id="page-18-17"></span>Nascimento LGL, Casanova F, Silva NFN, de Carvalho Teixeira AVN, de Carvalho AF (2020) Casein-based hydrogels: a mini-review. Food Chem 314:126063. [https://doi.org/10.1016/j.foodchem.](https://doi.org/10.1016/j.foodchem.2019.126063) [2019.126063](https://doi.org/10.1016/j.foodchem.2019.126063)
- <span id="page-18-27"></span>Ngarize S, Adams A, Howell N (2005) A comparative study of heat and high pressure induced gels of whey and egg albumen proteins and their binary mixtures. Food Hydrocoll 19(6):984–996. <https://doi.org/10.1016/j.foodhyd.2004.12.008>
- <span id="page-18-14"></span>Ni N, Dumont MJ (2017) Protein-based hydrogels derived from industrial byproducts containing collagen, keratin, zein and soy. Waste Biomass Valoriz 8(2):285–300. [https://doi.org/10.1007/](https://doi.org/10.1007/s12649-016-9684-0) [s12649-016-9684-0](https://doi.org/10.1007/s12649-016-9684-0)
- <span id="page-18-22"></span>Nonoyama T, Wada S, Kiyama R, Kitamura N, Mredha MTI, Zhang X, …, Gong JP (2016) Double-network hydrogels strongly bondable to bones by spontaneous osteogenesis penetration. Adv Mater 28(31):6740–6745. <https://doi.org/10.1002/adma.201601030>
- <span id="page-18-1"></span>Okuro PK, Santos TP, Cunha RL (2021) Compositional and structural aspects of hydro-and oleogels: similarities and specifcities from the perspective of digestibility. Trends Food Sci Technol 111:55–67. <https://doi.org/10.1016/j.tifs.2021.02.053>
- <span id="page-18-16"></span>Ozel B, Aydin O, Grunin L, Oztop MH (2018) Physico-chemical changes of composite whey protein hydrogels in simulated gastric fuid conditions. J Agric Food Chem 66(36):9542–9555. <https://doi.org/10.1021/acs.jafc.8b02829>
- <span id="page-18-30"></span>Oztop MH, McCarthy KL, McCarthy MJ, Rosenberg M (2014) Monitoring the effects of divalent ions  $(mn + 2$  and  $ca + 2)$  in heat-set whey protein gels. LWT-Food Sci Technol 56(1):93-100. [https://](https://doi.org/10.1016/j.lwt.2013.10.043) [doi.org/10.1016/j.lwt.2013.10.043](https://doi.org/10.1016/j.lwt.2013.10.043)
- <span id="page-18-29"></span>Palmese LL, Thapa RK, Sullivan MO, Kiick KL (2019) Hybrid hydrogels for biomedical applications. Curr Opin Chem Eng 24:143– 157. <https://doi.org/10.1016/j.coche.2019.02.010>
- <span id="page-18-19"></span>Pan K, Zhong Q (2016) Low energy, organic solvent-free co-assembly of zein and caseinate to prepare stable dispersions. Food Hydrocolloids 52:600–606. [https://doi.org/10.1016/j.foodhyd.2015.](https://doi.org/10.1016/j.foodhyd.2015.08.014) [08.014](https://doi.org/10.1016/j.foodhyd.2015.08.014)
- <span id="page-18-4"></span>Panahi R, Baghban-Salehi M (2019) Protein-based hydrogels. Cellulose-based superabsorbent hydrogels. Springer, Cham., pp 1561–1600. [https://doi.org/10.1007/978-3-319-77830-3\\_52.](https://doi.org/10.1007/978-3-319-77830-3_52)
- <span id="page-18-13"></span>Pardeshi S, Damiri F, Zehravi M, Joshi R, Kapare H, Prajapati MK, …, Barai HR (2022) Functional thermoresponsive hydrogel molecule to material design for biomedical applications. Polymers 14(15):3126
- <span id="page-18-8"></span>Park M, Shin HK, Kim BS, Kim MJ, Kim IS, Park BY, Kim HY (2015) Efect of discarded keratin-based biocomposite hydrogels on the wound healing process in vivo. Mater Sci Eng: C 55:88–94. <https://doi.org/10.1016/j.msec.2015.03.033>
- <span id="page-18-6"></span>Patwa R, Zandraa O, Capáková Z, Saha N, Sáha P (2020) Efect of ironoxide nanoparticles impregnated bacterial cellulose on overall properties of alginate/casein hydrogels: potential injectable biomaterial for wound healing applications. Polymers 12(11):2690. <https://doi.org/10.3390/polym12112690>
- <span id="page-18-24"></span>Pilipenko N, Goncalves OH, Bona E, Fernandes IP, Pinto JA, Sorita GD, …, Barreiro MF (2019) Tailoring swelling of alginate-gelatin hydrogel microspheres by crosslinking with calcium chloride combined with transglutaminase. Carbohydr Polym 223:115035. <https://doi.org/10.1016/j.carbpol.2019.115035>
- <span id="page-19-30"></span>Pradhan S, Hassani I, Clary JM, Lipke EA (2016) Polymeric biomaterials for in vitro cancer tissue engineering and drug testing applications. Tissue Eng Part B: Rev 22(6):470–484. [https://doi.](https://doi.org/10.1089/ten.teb.2015.0567) [org/10.1089/ten.teb.2015.0567](https://doi.org/10.1089/ten.teb.2015.0567)
- <span id="page-19-19"></span>Puppo MC, Anon MC (1998) Structural properties of heat-induced soy protein gels as afected by ionic strength and pH. J Agric Food Chem 46(9):3583–3589. <https://doi.org/10.1021/jf980006w>
- <span id="page-19-25"></span>Pyarasani RD, Jayaramudu T, John A (2019) Polyaniline-based conducting hydrogels. J Mater Sci 54(2):974–996. [https://doi.org/](https://doi.org/10.1007/s10853-018-2977-x) [10.1007/s10853-018-2977-x](https://doi.org/10.1007/s10853-018-2977-x)
- <span id="page-19-24"></span>Ramos AM, Pereira S, Cidade MT, Pereira G, Branquinho R, Pereira L, …, Fortunato E (2013) Preparation and characterization of cellulose nanocomposite hydrogels as functional electrolytes. Solid State Ionics 242:26–32. <https://doi.org/10.1016/j.ssi.2013.03.028>
- <span id="page-19-3"></span>Ren C, Chu L, Huang F, Yang L, Fan H, Liu J et al (2017) A novel H2O2responsive supramolecular hydrogel for controllable drug release. RSC Adv 7:1313–1317. [https://doi.org/10.1039/C6RA2](https://doi.org/10.1039/C6RA26536G) [6536G](https://doi.org/10.1039/C6RA26536G)
- <span id="page-19-8"></span>Rouse JG, Van Dyke ME (2010) A review of keratin-based biomaterials for biomedical applications. Materials 3(2):999–1014. [https://](https://doi.org/10.3390/ma3020999) [doi.org/10.3390/ma3020999](https://doi.org/10.3390/ma3020999)
- <span id="page-19-6"></span>Rudzinski WE, Dave AM, Vaishnav UH, Kumbar SG, Kulkarni AR, Aminabhavi TM (2002) Hydrogels as controlled release devices in agriculture. Des Monomers Polym 5(1):39–65. [https://doi.org/](https://doi.org/10.1163/156855502760151580) [10.1163/156855502760151580](https://doi.org/10.1163/156855502760151580)
- <span id="page-19-4"></span>Sabbagh F, Muhamad II, Pa'e N, Hashim Z (2019) Strategies in improving properties of cellulose-based hydrogels for smart applications. Cellulose-based superabsorbent hydrogels. Springer, Cham, pp 887–908. [https://doi.org/10.1007/978-3-](https://doi.org/10.1007/978-3-319-77830-3_30) [319-77830-3\\_30.](https://doi.org/10.1007/978-3-319-77830-3_30)
- <span id="page-19-33"></span>Sahiner M, Alpaslan D, Bitlisli BO (2014) Collagen-based hydrogel flms as drug-delivery devices with antimicrobial properties. Polym Bull 71(11):3017–3033. [https://doi.org/10.1007/](https://doi.org/10.1007/s00289-014-1235-x) [s00289-014-1235-x](https://doi.org/10.1007/s00289-014-1235-x)
- <span id="page-19-5"></span>Saini K (2017) Preparation method, Properties and Crosslinking of hydrogel: a review. PharmaTutor 5(1):27–36
- <span id="page-19-16"></span>Santin M, Ambrosio L (2008) Soybean-based biomaterials: preparation, properties and tissue regeneration potential. Expert Rev Med Dev 5(3):349–358. [https://doi.org/10.1586/17434440.5.3.](https://doi.org/10.1586/17434440.5.3.349) [349](https://doi.org/10.1586/17434440.5.3.349)
- Santos TP, Cunha RL (2018) Role of process variables on the formation and in vitro digestion of gellan gels. Carbohydr Polym 192:111– 117. <https://doi.org/10.1016/j.carbpol.2018.03.062>
- <span id="page-19-23"></span>Seliktar D (2012) Designing cell-compatible hydrogels for biomedical applications. Science 336(6085):1124–1128. [https://doi.org/10.](https://doi.org/10.1126/science.1214804) [1126/science.1214804](https://doi.org/10.1126/science.1214804)
- <span id="page-19-32"></span>Semasaka C, Dekiwadia C, Buckow R, Kasapis S (2019) Modeling counterion partition in composite gels of BSA with gelatin following high pressure treatment. Food Chem 285:104–110. <https://doi.org/10.1016/j.foodchem.2019.01.125>
- Shand PJ, Ya H, Pietrasik Z, Wanasundara PKJPD (2008) Transglutaminase treatment of pea proteins: Efect on physicochemical and rheological properties of heat-induced protein gels. Food Chem 107(2):692–699. [https://doi.org/10.1016/j.foodchem.2007.](https://doi.org/10.1016/j.foodchem.2007.08.095) [08.095](https://doi.org/10.1016/j.foodchem.2007.08.095)
- <span id="page-19-0"></span>Shavandi A, Silva TH, Bekhit AA, Bekhit AEDA (2017) Keratin: dissolution, extraction and biomedical application. Biomaterials Sci 5(9):1699–1735
- <span id="page-19-15"></span>Shewan HM, Stokes JR (2013) Review of techniques to manufacture micro-hydrogel particles for the food industry and their applications. J Food Eng 119(4):781–792. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jfoodeng.2013.06.046) [jfoodeng.2013.06.046](https://doi.org/10.1016/j.jfoodeng.2013.06.046)
- <span id="page-19-13"></span>Shi W, Dumont MJ, Ly EB (2014) Synthesis and properties of canola protein-based superabsorbent hydrogels. Eur Polymer J 54:172– 180. <https://doi.org/10.1016/j.eurpolymj.2014.03.007>
- <span id="page-19-17"></span>Shibayama M (2012) Structure-mechanical property relationship of tough hydrogels. Soft Matter 8(31):8030–8038. [https://doi.org/](https://doi.org/10.1039/C2SM25325A) [10.1039/C2SM25325A](https://doi.org/10.1039/C2SM25325A)
- <span id="page-19-31"></span>Shin S, Ikram M, Subhan F, Kang HY, Lim Y, Lee R, …, Yoon S (2016) Alginate–marine collagen–agarose composite hydrogels as matrices for biomimetic 3D cell spheroid formation. RSC Adv 6(52):46952–46965.<https://doi.org/10.1039/C6RA01937D>
- <span id="page-19-11"></span>Shukla R, Cheryan M (2001) Zein: the industrial protein from corn. Ind Crops Prod 13(3):171–192. [https://doi.org/10.1016/S0926-](https://doi.org/10.1016/S0926-6690(00)00064-9) [6690\(00\)00064-9](https://doi.org/10.1016/S0926-6690(00)00064-9)
- <span id="page-19-10"></span>Singhal R, Gupta K (2016) A review: tailor-made hydrogel structures (classifcations and synthesis parameters). Polym-Plast Technol Eng 55(1):54–70. [https://doi.org/10.1080/03602559.2015.10505](https://doi.org/10.1080/03602559.2015.1050520) [20](https://doi.org/10.1080/03602559.2015.1050520)
- <span id="page-19-26"></span>Slaughter BV, Khurshid SS, Fisher OZ, Khademhosseini A, Peppas NA (2009) Hydrogels in regenerative medicine. Adv Mater 21(32–33):3307–3329.<https://doi.org/10.1002/adma.200802106>
- <span id="page-19-21"></span>Sofa SJ, Singh A, Kaplan DL (2002) Peroxidase-catalyzed crosslinking of functionalized polyaspartic acid polymers. J Macromol Sci Part A 39(10):1151–1181
- <span id="page-19-7"></span>Song F, Zhang LM, Yang C, Yan L (2009) Genipin-crosslinked casein hydrogels for controlled drug delivery. Int J Pharm 373(1–2):41– 47.<https://doi.org/10.1016/j.ijpharm.2009.02.005>
- <span id="page-19-9"></span>Sousa FFO, Luzardo-Álvarez A, Blanco-Méndez J, Martín-Pastor M (2012) NMR techniques in drug delivery: application to zein protein complexes. Int J Pharm 439(1–2):41–48. [https://doi.org/](https://doi.org/10.1016/j.ijpharm.2012.09.046) [10.1016/j.ijpharm.2012.09.046](https://doi.org/10.1016/j.ijpharm.2012.09.046)
- <span id="page-19-28"></span>Tang MX, Zhu YD, Li D, Adhikari B, Wang LJ (2019) Rheological, thermal and microstructural properties of casein/κ-carrageenan mixed systems. Lwt-Food Sci Technol 113:108296. [https://doi.](https://doi.org/10.1016/j.lwt.2019.108296) [org/10.1016/j.lwt.2019.108296](https://doi.org/10.1016/j.lwt.2019.108296)
- <span id="page-19-20"></span>Teimouri S, Morrish C, Panyoyai N, Small DM, Kasapis S (2019) Diffusion and relaxation contributions in the release of vitamin B6 from a moving boundary of genipin crosslinked gelatin matrices. Food Hydrocolloids 87:839–846. [https://doi.org/10.1016/j.foodh](https://doi.org/10.1016/j.foodhyd.2018.09.015) [yd.2018.09.015](https://doi.org/10.1016/j.foodhyd.2018.09.015)
- <span id="page-19-22"></span>Teixeira LSM, Feijen J, van Blitterswijk CA, Dijkstra PJ, Karperien M (2012) Enzyme-catalyzed crosslinkable hydrogels: emerging strategies for tissue engineering. Biomaterials 33(5):1281–1290. <https://doi.org/10.1016/j.biomaterials.2011.10.067>
- <span id="page-19-14"></span>Totosaus A, Montejano JG, Salazar JA, Guerrero I (2002) A review of physical and chemical protein-gel induction. Int J Food Sci Technol 37(6):589–601. [https://doi.org/10.1046/j.1365-2621.](https://doi.org/10.1046/j.1365-2621.2002.00623.x) [2002.00623.x](https://doi.org/10.1046/j.1365-2621.2002.00623.x)
- <span id="page-19-29"></span>Ullah F, Othman MBH, Javed F, Ahmad Z, Akil HM (2015) Classifcation, processing and application of hydrogels: a review. Mater Sci Eng: C 57:414–433.<https://doi.org/10.1016/j.msec.2015.07.053>
- <span id="page-19-27"></span>Varaprasad K, Raghavendra GM, Jayaramudu T, Yallapu MM, Sadiku R (2017) A mini review on hydrogels classifcation and recent developments in miscellaneous applications. Mater Sci Eng: C 79:958–971. <https://doi.org/10.1016/j.msec.2017.05.096>
- <span id="page-19-12"></span>Wakuda Y, Nishimoto S, Suye SI, Fujita S (2018) Native collagen hydrogel nanofbres with anisotropic structure using core-shell electrospinning. Sci Rep 8(1):1–10
- <span id="page-19-1"></span>Wang LF, Rhim JW (2015) Preparation and application of agar/alginate/collagen ternary blend functional food packaging flms. Int J Biol Macromol 80:460–468. [https://doi.org/10.1016/j.ijbiomac.](https://doi.org/10.1016/j.ijbiomac.2015.07.007) [2015.07.007](https://doi.org/10.1016/j.ijbiomac.2015.07.007)
- <span id="page-19-2"></span>Wang X, Zeng M, Qin F, Adhikari B, He Z, Chen J (2018) Enhanced CaSO4-induced gelation properties of soy protein isolate emulsion by pre-aggregation. Food Chem 242:459–465. [https://doi.](https://doi.org/10.1016/j.foodchem.2017.09.044) [org/10.1016/j.foodchem.2017.09.044](https://doi.org/10.1016/j.foodchem.2017.09.044)
- <span id="page-19-18"></span>Wattie B, Dumont MJ, Lefsrud M (2018) Synthesis and properties of feather keratin-based superabsorbent hydrogels. Waste

Biomass Valoriz 9(3):391–400. [https://doi.org/10.1007/](https://doi.org/10.1007/s12649-016-9773-0) [s12649-016-9773-0](https://doi.org/10.1007/s12649-016-9773-0)

- <span id="page-20-9"></span>Wei Y, Xie R, Lin Y, Xu Y, Wang F, Liang W, Zhang J (2016) Structure formation in pH-sensitive hydrogels composed of sodium caseinate and N, O-carboxymethyl chitosan. Int J Biol Macromol 89:353–359
- <span id="page-20-5"></span>Whitford D (2005) Proteins: structure and function. Wiley, New York
- Wu BC, Degner B, McClements DJ (2014) Soft matter strategies for controlling food texture: formation of hydrogel particles by biopolymer complex coacervation. J Phys: Condens Matter 26(46):464104
- <span id="page-20-7"></span>Wu S, Deng L, Hsia H, Xu K, He Y, Huang Q, …, Peng C (2017) Evaluation of gelatin-hyaluronic acid composite hydrogels for accelerating wound healing. J Biomater Appl 31(10):1380–1390. <https://doi.org/10.1177/0885328217702526>
- <span id="page-20-3"></span>Xiao Y, Li J, Liu Y, Peng F, Wang X, Wang C, …, Xu H (2020) Gel properties and formation mechanism of soy protein isolate gels improved by wheat bran cellulose. Food Chem 324:126876. <https://doi.org/10.1016/j.foodchem.2020.126876>
- <span id="page-20-10"></span>Zhang JT, Bhat R, Jandt KD (2009) Temperature-sensitive PVA/PNI-PAAm semi-IPN hydrogels with enhanced responsive properties. Acta Biomater 5(1):488–497
- <span id="page-20-8"></span>Zhang P, Cheetham AG, Lock LL, Cui H (2013) Cellular uptake and cytotoxicity of drug-peptide conjugates regulated by conjugation site. Bioconjug Chem 24:604–613. [https://doi.org/10.1021/](https://doi.org/10.1021/bc300585h) [bc300585h](https://doi.org/10.1021/bc300585h)
- <span id="page-20-4"></span>Zhang Z, Decker EA, McClements DJ (2014) Encapsulation, protection, and release of polyunsaturated lipids using biopolymerbased hydrogel particles. Food Res Int 64:520–526. [https://doi.](https://doi.org/10.1016/j.foodres.2014.07.020) [org/10.1016/j.foodres.2014.07.020](https://doi.org/10.1016/j.foodres.2014.07.020)
- <span id="page-20-2"></span>Zhang HJ, Sun TL, Zhang AK, Ikura Y, Nakajima T, Nonoyama T, …, Gong JP (2016a) Tough physical double-network hydrogels based on amphiphilic triblock copolymers. Adv Mater 28(24):4884–4890. <https://doi.org/10.1002/adma.201600466>
- <span id="page-20-0"></span>Zhang M, Yang Y, Acevedo NC (2020) Effect of oil content and composition on the gelling properties of Egg-SPI proteins stabilized emulsion gels. Food Biophys 15(4):473–481. [https://doi.org/10.](https://doi.org/10.1007/s11483-020-09646-8) [1007/s11483-020-09646-8](https://doi.org/10.1007/s11483-020-09646-8)
- <span id="page-20-1"></span>Zhu J, Li F, Wang X, Yu J, Wu D (2018) Hyaluronic acid and polyethylene glycol hybrid hydrogel encapsulating nanogel with hemostasis and sustainable antibacterial property for wound healing. ACS Appl Mater Interfaces 10(16):13304–13316. [https://doi.org/](https://doi.org/10.1021/acsami.7b18927) [10.1021/acsami.7b18927](https://doi.org/10.1021/acsami.7b18927)
- <span id="page-20-6"></span>Zohuriaan-Mehr MJ, Pourjavadi A, Salimi H, Kurdtabar M (2009) Protein‐and homo poly (amino acid) based hydrogels with super swelling properties. Polym Adv Technol 20(8):655–671. [https://](https://doi.org/10.1002/pat.1395) [doi.org/10.1002/pat.1395](https://doi.org/10.1002/pat.1395)

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.