

An updated Alzheimer's disease progression model: incorporating non-linearity, beta regression, and a third-level random effect in NONMEM

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Abstract Our objective was to expand our understanding of the predictors of Alzheimer's disease (AD) progression to help design a clinical trial on a novel AD medication. We utilized the Coalition Against Major Diseases AD dataset consisting of control-arm data (both placebo and stable background AD medication) from 15 randomized double-blind clinical trials in mild-to-moderate AD patients (4,495 patients; July 2013). Our ADAS-cog longitudinal model incorporates a beta-regression with between-study, -subject, and -residual variability in NONMEM; it suggests that faster AD progression is associated with younger age and higher number of apolipoprotein E type 4 alleles (APOE*4), after accounting for baseline disease severity. APOE*4, in particular, seems to be implicated in the AD pathogenesis. In addition, patients who are already on stable background AD medications appear to have a faster progression relative to those who are not receiving AD medication. The current knowledge

does not support a causality relationship between use of background AD medications and higher rate of disease progression, and the correlation is potentially due to confounding covariates. Although causality has not necessarily been demonstrated, this model can inform inclusion criteria and stratification, sample size, and trial duration.

Keywords Alzheimer's disease · Disease progression · Patient-level model-based meta-analysis · Coalition Against Major Diseases (CAMD) · ADAS-cog · NONMEM

Introduction

Alzheimer's disease (AD) remains the largest unmet medical need in neurology. AD is a progressive and lethal disease occurring in the brain, and is the 6th most common cause of death in the United States (US). More than 5 million Americans are living with AD. This number is estimated to virtually triple by 2050 with the aging of the "Baby Boomer" generation; as a consequence, the direct costs for care is expected to increase from \$203 billion in 2013 to \$1.2 trillion [1].

There is no available treatment to prevent, stop or slow the course of AD (i.e., disease-modifying therapies). Currently, four prescription drugs are available on the US market for AD symptomatic relief: donepezil, rivastigmine, galantamine (acetylcholinesterase inhibitors, AChEIs), and memantine (an *N*-methyl-D-aspartate receptor antagonist). Multiple AD-modifying therapies have failed in development due to several reasons including insufficient understanding of pathogenesis, diagnosis uncertainty, difficult selection of therapeutic targets, and non-optimal clinical trial design [2].

The Coalition Against Major Diseases: Data used in preparation of this article were obtained from the Coalition Against Major Diseases (CAMD) database (<http://codr.c-path.org>). A complete listing of CAMD members can be found at: <http://c-path.org/programs/camd/>.

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This analysis presents a potential tool to enhance clinical trial design from proof-of-concept through registration studies. A disease-drug-trial model is a useful tool to aid clinical study design by determining (1) the most responsive patient population to treatment, (2) the number of patients to be enrolled in the trial, (3) the trial duration and the optimal timing of response measurement, (4) the optimal outcome measure (e.g., different endpoints measuring different aspects of the disease, combinations of sub-items or composite endpoints), among others [3].

The Alzheimer's disease assessment scale-cognitive subscale (ADAS-cog) is a widely used, validated measure of cognitive deterioration in clinical trials of patients with mild-to-moderate AD dementia [4], and several models have been developed to describe ADAS-cog. Holford et al. [5–7] in early 1990 s' described the effect of tacrine in the cognitive function of approximately 2,500 patients with AD with a linear disease progression rate. In 2010, Ito et al. [8] conducted a summary-level model-based meta-analysis of 52 trials and 19,972 patients with mild-to-moderate AD; this analysis employed a linear model for disease progression and rate of progression was a function of the disease severity. Subsequently, Ito et al. [9] performed a patient-level model-based meta-analysis of 817 patients from the Alzheimer's Disease Neuroimaging Initiative (ADNI); that analysis identified individual covariates influencing changes in the ADAS-cog score predicting curvilinear disease progression rate: age, sex, apolipoprotein E type 4 allele (APOE*4) carrier/non-carrier [10], and baseline disease severity. In 2011, Yang et al. [11] used an exponential model to describe the rate of cognitive decline in patients from the ADNI database (193 patients with AD and 397 patients with mild cognitive impairment); where synchronization method was developed to predict the disease progression timeline for each patient (i.e., the initial time represents the start of the cognitive decline instead of the time of enrollment in the ADNI trial) incorporating changes in cerebrospinal fluid and imaging biomarkers. In 2012, Samtani et al. [12] utilized a generalized logistic model to describe the non-linear rate of disease progression in 191 patients with AD from the ADNI database; this analysis evaluated wide range of covariates (e.g., imaging volumetric brain measures, serum biomarkers, and other baseline cognitive tests) and identified that the disease progression rate was influenced by age, total serum cholesterol, APOE*4 carrier/non-carrier, trail making test (part B) score, and ADAS-cog at each time point. In the same year, Rogers et al. [13] published a model-based meta-analysis incorporating beta regression (restricting predictions to the test range of 0–70) to describe the longitudinal progression of ADAS-cog with summary- and patient-level data; summary-level data was obtained from 73 published trials (representing 17,235 patients), and patient-level data

was obtained from the ADNI (186 AD patients) and the Coalition Against Major Diseases (CAMD; 3,223 AD patients, data cut-off September 2010) databases. They improved prediction performance by using a generalized logistic function (via a logit transformation) along with beta-distributed residual variability and included an evaluation of APOE*4 with allele count.

The goal of the present analysis is to integrate aspects from the previously published mathematical models and expand our understanding of AD progression. To this end, we utilized an updated CAMD dataset consisting of control-arm data (both placebo and stable background AD medication) from randomized double-blind clinical trials in mild-to-moderate AD patients (4,495 patients; July 2013). We address three critical and relevant questions to the research community working on AD drug development:

1. whether stable background AD medications influence disease progression,
2. whether disease severity of enrolled patients has changed in recent years,
3. whether the number of APOE*4 alleles have different magnitude of effect on disease progression.

For the first question, an exploratory analysis of the longitudinal ADAS-cog mean profile stratified by patients on placebo only versus patients on background AD medication suggests the influence of medication in ADAS-cog trajectory. In spite of all innovations implemented by the most recent publication on the CAMD data [13], the apparent effect of AD medication remains to be investigated. This is important given that 67 % of the patients in the CAMD database were taking at least one AD medication as background therapy. For the second question, we conducted an exploratory analysis on whether baseline disease severity has changed by trial year possibly due to changes in life style, awareness of the disease, diagnostics and patient care. For the third question, the current larger number of genotyped patients in the CAMD database allows for a reevaluation of the effect of the APOE*4 variant on rate of disease progression.

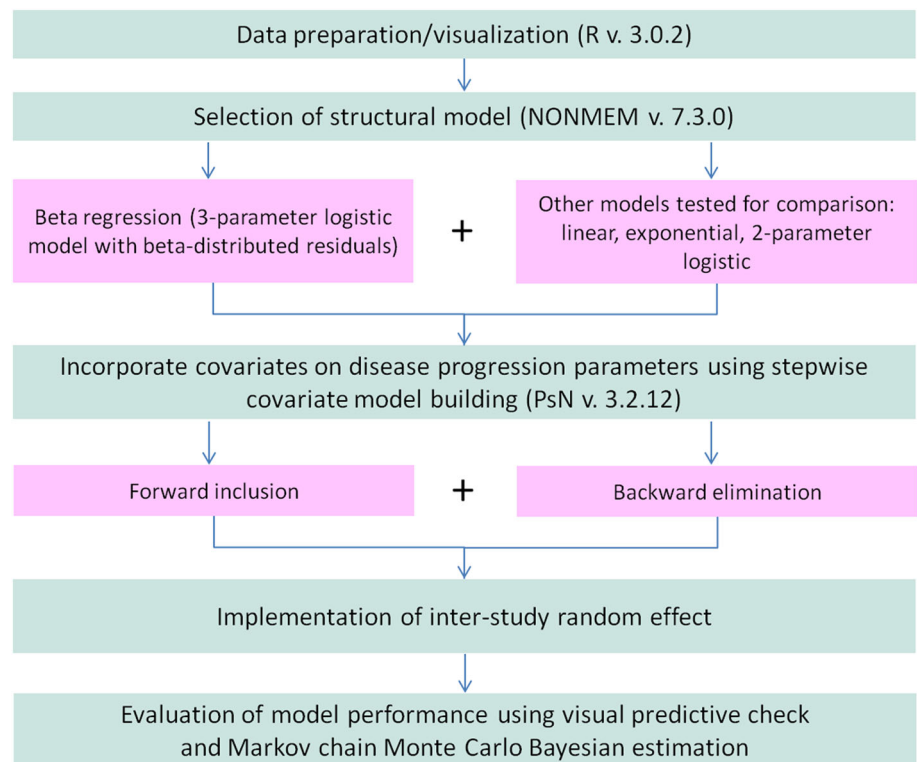
To our knowledge, this is the first time in the context of a non-linear mixed effect modeling of ADAS-cog meta-analysis that a beta-regression with between-study, -subject, and -residual variability has been implemented in NONMEM.

Methods

Data

Data from the CAMD database (with a cutoff in July 2013) was utilized in this analysis. CAMD is a public-private

Fig. 1 Schematic representation of the model building process



consortium led by the Critical Path Institute, which brings together government, regulatory agencies, academic institutions, pharmaceutical industry and non-profit research organizations. CAMD, launched in September of 2008, aims to advance new tools and methods to enhance the development of new AD and Parkinson’s disease therapies [14]. The consortium focuses on precompetitive sharing of patient-level data from control arms (both placebo and stable background AD medication) from randomized-controlled-trials in AD, conducted by pharmaceutical companies that are members of CAMD. The dataset included 4,495 patients with mild-to-moderate AD from 15 studies. The 11-item ADAS-cog (ADAS-cog11) used in this analysis has been widely used as a primary endpoint in mild to moderate AD clinical trials, and the same algorithm has been used to derive ADAS-cog11 across clinical studies in the CAMD database. Missing ADAS-cog records or scores before placebo administration were excluded for this analysis. Available patient characteristics include age, sex, enrollment year in the trial, baseline mini-mental state examination (MMSE [15]), APOE*4 genotype (allele combinations), and background use of AD medication (donepezil, galantamine, rivastigmine, or memantine).

Model building process

The model building followed the steps outlined in Fig. 1. Data preparation and visualization were completed using R (version 3.0.2; Vienna, Austria [16]). Selection of the base

model structure with incorporation of two levels of random effects (inter-individual and residual variability) preceded covariate model building and incorporation of a third level of random effects for study; finally, model performance was evaluated with goodness-of-fit and visual predictive checks.

Selection of the base model structure

Selection of the base model structure (summarized in Table 1) acknowledged the previously published ADAS-cog longitudinal models [5–9, 11–13] and used NONMEM® (version 7.3.0; Ellicott City, Maryland, USA [17]). The most recently published ADAS-cog longitudinal models utilized generalized logistic functions to describe the non-linear AD progression rate [12, 13]; as such, the Richard’s function with its analytical solution was tested [18] (Eq. 1).

$$ADAScog(t) = \frac{ADAScog_0 \cdot 70}{\left[ADAScog_0^\beta + (70^\beta - ADAScog_0^\beta) \cdot e^{-\beta \cdot r \cdot t} \right]^{1/\beta}} \tag{1}$$

where ADAScog₀ is the population baseline ADAS-cog, r is the intrinsic rate of progression, t is time in years, β is the shape factor of the Richard’s function allowing for non-centrality of the inflection of disease progression rate, 70 is the maximum possible observed ADAS-cog score. Other literature models—linear, exponential and 2-parameter logistic models—were also tested for comparison.

Table 1 Selection of the base model structure

Base models	Disease progression rate	OFV	Number of parameters	AIC
<p><i>Richards function (generalized logistic model)</i></p> $\frac{dADAScog}{dt} = r \cdot ADAScog \cdot \left[1 - \left(\frac{ADAScog}{70} \right)^\beta \right]$ <p>where r is the intrinsic rate of progression, t is time in years, β is the shape factor of the Richards function, 70 is the maximum observed ADAS-cog score</p>	Non-linear	-60,776	3	-60,756
<p><i>Exponential model</i></p> $\frac{dADAScog}{dt} = r \cdot ADAScog$	Non-linear	-60,687	2	-60,674
<p><i>Logistic model (symmetric)</i></p> $\frac{dADAScog}{dt} = r \cdot ADAScog \cdot \left[1 - \frac{ADAScog}{70} \right]$	Non-linear	-60,262	2	-60,249
<p><i>Linear model with rate of progression dependent on MMSE (fractional change in slope per baseline MMSE different from 20)</i></p> $\frac{dADAScog}{dt} = r$ $r = \theta_r \cdot (1 + (MMSE - 20) \cdot \theta_{MMSE})$ <p>where θ_r is the population estimate for the intrinsic rate of progression r, MMSE is the baseline mini-mental state examination score, 20 is median baseline MMSE across all subjects, θ_{MMSE} is the MMSE fitted coefficient.</p>	Non-linear in a population level (non-linearity introduced by the effect of MMSE in slope)	-60,143	3	-60,123
<p><i>Linear model</i></p> $\frac{dADAScog}{dt} = r$	Linear	-60,008	2	-59,994
<p><i>Linear model with rate of progression dependent on MMSE (exponential relationship)</i></p> $\frac{dADAScog}{dt} = r$ $r = \theta_r \cdot e^{((MMSE-20) \cdot \theta_{MMSE})}$	Non-linear in a population level (non-linearity introduced by the effect of MMSE in slope)	-60,008	3	-59,988

ADAS-cog is the cognition portion of the Alzheimer's Disease Assessment Scale; MMSE is mini-mental state examination; OFV is the objective function value. AIC is the Akaike information criterion with a per parameter penalty of 6.635

Incorporation of three levels of random effects

Three levels of random variability were included in the final model: inter-study, inter-individual (patient) and residual variability. Inter-individual random effects were tested for baseline ADAS-cog ($ADAScog_0$) and intrinsic rate of progression (r) parameters. Normally distributed inter-subject variability of the intrinsic rate of progression was estimated on the linear scale. This structure permitted that, at the patient-level, the cognitive function could capture improvement, decline or lack of change during the study. Log-normally distributed inter-subject variability was estimated for baseline ADAS-cog. This structure prevented the model from predicting nonsensical negative baseline scores at the patient-level. The same error

structures were also utilized to account for inter-study variability in the intrinsic rate of progression and baseline ADAS-cog. The incorporation of inter-study variability, though, was only conducted in the final model building process (i.e., after the covariate model building) to minimize computational time.

$$ADAScog_{0ij} = ADAScog_0 \cdot \exp\left(\eta_{baseline_i}^{study} + \eta_{baseline_{ij}}^{patient}\right) \quad (2)$$

$$r_{ij} = r + \eta_{intrinsic\ rate_i}^{study} + \eta_{intrinsic\ rate_{ij}}^{patient} \quad (3)$$

where $ADAScog_0$ is the population baseline ADAS-cog; $ADAScog_{0ij}$ is the individual baseline ADAS-cog; r is the population intrinsic rate of progression; r_{ij} is the individual intrinsic rate of progression; η^{study} is the inter-study

random effect, $\eta_{\text{baseline},i}^{\text{study}} \sim N(0, \omega_{\text{baseline},\text{study}}^2)$, $\eta_{\text{intrinsic rate},i}^{\text{study}} \sim N(0, \omega_{\text{intrinsic rate},\text{study}}^2)$; η^{patient} is the inter-individual random effect, $\eta_{\text{baseline},ij}^{\text{patient}} \sim N(0, \omega_{\text{baseline},\text{patient}}^2)$, $\eta_{\text{intrinsic rate},ij}^{\text{patient}} \sim N(0, \omega_{\text{intrinsic rate},\text{patient}}^2)$.

Residual variability was assumed to be beta-distributed [13, 19, 20]. Because ADAS-cog is bounded between 0 and 70, the expectation for progression should be non-linear given the ceiling and floor effects; in addition, the error distribution should be heteroscedastic since the variance must approach zero as their mean approach the boundaries [19]. The combination of a logistic model with beta-distributed residuals, also known as beta regression, constrains the model predictions to the 0–70 range.

The probability density function of the beta distribution [20], with ADAS-cog score as the dependent variable is:

$$f(\text{ADAScog}; \alpha, \beta) = \frac{\Gamma(\alpha + \beta)}{\Gamma(\alpha) \Gamma(\beta)} \cdot \text{ADAScog}^{\alpha-1} \cdot (1 - \text{ADAScog})^{\beta-1} \quad (4)$$

where $\text{ADAScog} \in (0, 1)$; α and β are shape parameters with α pulling the density toward 0 and β pulling the density toward 1, $\alpha > 0$, $\beta > 0$; $\Gamma(\dots)$ denotes the gamma function.

According to Eq. 4, the probability density function goes to zero if ADAS-cog equals 0 or 1 which explains why ADAS-cog must belong to the open unit interval (0,1). As a consequence the following transformation [20] of ADAS-cog was performed:

$$\text{ADAScog}' = \frac{\text{ADAScog} - 0}{70 - 0} \quad (5)$$

$$\text{ADAScog}'' = \frac{[\text{ADAScog}' \cdot (n - 1) + 0.5]}{n}$$

where n is the total sample size ($n = 4,495$); 0 is the lowest and 70 is the highest possible ADAS-cog score. The rationale for the above transformation is discussed in the supplemental material of Smithson and Verkuilen [20]. Assuming $\text{ADAS-cog} \sim \text{beta}(\alpha, \beta)$, then:

$$\mu = \frac{\alpha}{\alpha + \beta} \quad (6)$$

$$\sigma^2 = \frac{\mu \cdot (1 - \mu)}{(\tau + 1)} \quad (7)$$

where μ is the expected value of the beta distribution; σ^2 is the variance of the beta distribution; $\tau = \alpha + \beta$. Therefore, the dispersion of the distribution is determined by μ and τ .

Within NONMEM, τ was estimated as a fixed effect parameter and μ corresponded to the individual ADAS-cog prediction. The shape parameters α and β were then calculated as below and substituted into Eq. 4:

$$\alpha = \mu \cdot \tau \quad (8)$$

$$\beta = (1 - \mu) \cdot \tau \quad (9)$$

Nemes' approximation to the gamma function was incorporated in the model [19]:

$$\Gamma(X) \sim \left(\frac{X}{e}\right)^X \cdot \sqrt{\frac{2\pi}{X}} \cdot \left(1 + \frac{1}{15 \cdot X^2}\right)^{\frac{5}{4}X} \quad (10)$$

where X represents α , β , or $(\alpha + \beta)$ in the probability density function (Eq. 4).

The NONMEM Laplacian estimation method was used to take into account the second derivative of the inter-individual variability parameters.

Covariate model building

Although age and MMSE may have undergone variation over time, we have considered only baseline values for modeling purpose. Only 6 of 15 studies had APOE*4 genotype information; for other studies, genotype was not performed or genetic data was not provided. APOE*4 was categorized as 0 (non-carrier), 1 (heterozygous), 2 (homozygous), and 3 (unknown). There were no missing values for the other covariates.

The selected base model structure (Richard's function) intrinsically accounts for disease severity on the rate of progression (i.e., ADAS-cog score at a specific time point). MMSE was not included in the covariate model building because it is an alternative measure of cognitive impairment or disease severity. Similar to what was demonstrated

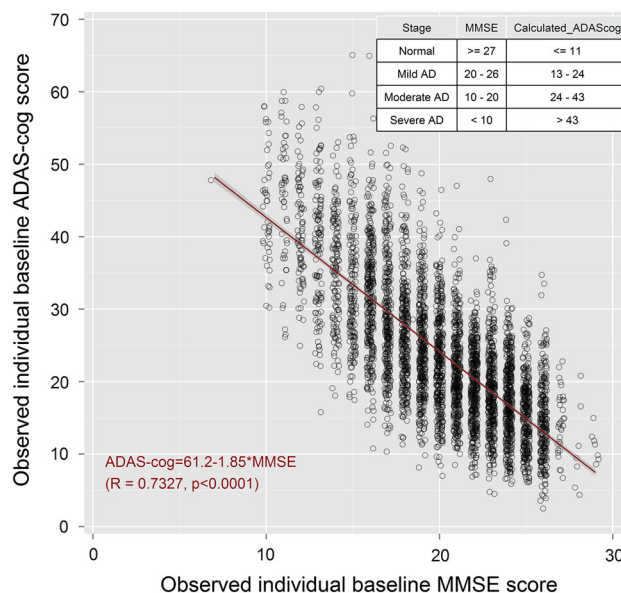


Fig. 2 Linear regression between observed baseline ADAS-cog and MMSE score. ADAS-cog scores presented in the table were calculated based on the linear regression equation. Observations were slightly jittered to aid interpretation

by Ito et al. [8], baseline MMSE scores were highly correlated with baseline ADAS-cog scores (Fig. 2). In addition to the covariates reported in the literature, we investigated the effect of stable background AD medication and patient enrollment year in the trial.

Before performing the covariate model building, we examined the presence of correlation among the covariates of interest. Figure 3 shows that there was a correlation between the patient enrollment year in the trial (START) and APOE*4 genotype (APOE4) as well as use of AD medication (COMED). In trials before ~2001, APOE*4 genotyping was not performed and controls were not taking AD medication. For this reason, the patient enrollment year in the trial was not included in the covariate model building. Each covariate was tested as predictor of baseline ADAS-cog and intrinsic rate of progression using a step-wise covariate model building in Perl-speaks-NONMEM (PsN, version 3.5.4; Uppsala, Sweden [21]).

An exploratory analysis on how baseline disease severity changed from 1994 to 2008 was conducted and compared to literature data. The literature data (summary-level) from 1990 to 2008 was obtained from Ito et al. [8] and updated with studies through 2010. This updated dataset was used in the disease progression model by Rogers et al. [13].

Model selection criteria

Overall model selection was guided by objective function value (OFV, NONMEM version 7.3.0; Ellicott City,

Maryland, USA [22]) and Akaike information criterion (AIC [23]) with a per-parameter (k) penalty of 6.635 ($AIC = 6.635k + OFV$, equivalent to χ^2 with 1 *df*, $P = 0.01$). In addition, goodness-of-fit plots were generated. Ill-conditioning of the parameter space was assessed by inspecting the eigenvalues of the covariance matrix. The ratio of the largest to the smallest eigenvalue (i.e., condition number) was not to exceed 1,000.

Once the final model was identified, 500 datasets identical in structure and covariate values to the original dataset were simulated, using the parameter estimates from the final model to evaluate the model performance. The longitudinal ADAS-cog scores over time stratified by covariate were generated and visually compared with the observed data (visual predictive check, VPC) at each selected percentile (5th, 50th, 95th). The model was also run using the Markov chain Monte Carlo Bayesian estimation method to obtain confidence intervals for the parameter estimates.

Results

Data summary

Table 2 describes the patient characteristics stratified by study. Patients with mild and moderate stage of the disease were approximately equally distributed for the combined studies. From the 15 studies, 8 studies had control arms on

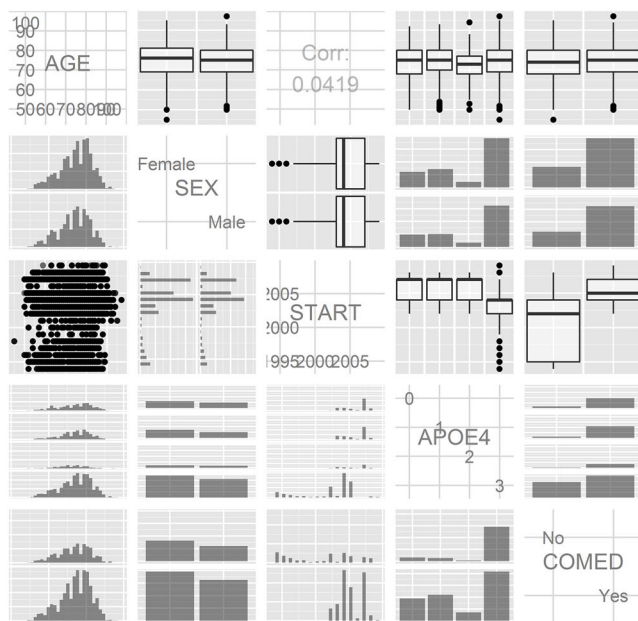


Fig. 3 Correlation among covariates and histograms of available covariates over year. AGE: age in years; START: patient enrollment year in the trial; APOE4: 0 is APOE*4 non-carrier, 1 is APOE*4



heterozygous, 2 is APOE*4 homozygous, 3 is unknown genotype; COMED: use of background AD medication

Table 2 Patient characteristics by study in the CAMD database

Study	Duration (weeks)	Sample size	Age (mean ± SD)	Female (%)	APOE*4 alleles (%)				Baseline MMSE (mean ± SD)	Baseline ADAS-cog (mean ± SD)	Background use of AD medication (%)
					0	1	2	NA			
1000	12	66	74 ± 9	54.5	–	–	–	100.0	21 ± 4	20 ± 7	100.0
1009	12	164	74 ± 6	55.5	53.0	36.6	9.8	0.6	21 ± 4	24 ± 12	0.0
1013	78	718	74 ± 8	50.1	–	–	–	100.0	21 ± 3	24 ± 9	80.5
1014	78	642	75 ± 8	56.2	–	–	–	100.0	21 ± 3	21 ± 8	75.5
1055	52	139	73 ± 8	58.3	–	–	–	100.0	19 ± 4	25 ± 10	0.0
1056	54	480	73 ± 8	56.5	41.0	41.7	17.3	0.0	20 ± 4	24 ± 10	99.8
1057	54	488	74 ± 8	61.3	43.9	46.9	9.2	0.0	19 ± 4	25 ± 11	99.6
1058	24	160	72 ± 9	59.4	51.9	38.8	9.4	0.0	20 ± 4	25 ± 10	0.0
1105	80	323	73 ± 9	51.1	32.8	38.1	11.1	18.0	21 ± 4	22 ± 10	99.1
1131	24	57	75 ± 9	59.6	–	–	–	100.0	24 ± 1	21 ± 4	0.0
1133	30	162	73 ± 7	61.1	–	–	–	100.0	19 ± 5	28 ± 12	0.0
1135	30	272	71 ± 9	55.5	–	–	–	100.0	20 ± 4	25 ± 10	0.0
1137	24	216	76 ± 8	50.5	–	–	–	100.0	17 ± 4	28 ± 10	100.0
1138	24	202	77 ± 8	57.4	–	–	–	100.0	17 ± 3	28 ± 10	0.0
1142	78	406	76 ± 8	55.9	30.0	42.9	17.2	9.9	21 ± 3	23 ± 9	91.1
Total	–	4,495	74 ± 8	55.5	18.0	18.9	5.9	57.2	20 ± 4	24 ± 10	66.7
Range	[12, 80]	[57, 718]	–	[50.1, 61.3]	–	–	–	–	–	–	[0.0, 100.0]

The study number is a unique study identifier in the CAMD database

APOE*4 apolipoprotein E type 4 allele, AD Alzheimer’s disease, SD standard deviation, NA not available

stable AD medication (66.7 %). From the 42.8 % of the patients with known APOE*4 genotype across all studies, 42 % were non-carriers (0 alleles), 44 % were heterozygous (1 allele), and 14 % were homozygous (2 alleles) for this genetic variant which aligns with previous reports [24].

Base model selection

Base models were tested for each of the literature-reported model structures as described in the methods section with model forms, characteristics, and AIC presented in Table 1. Richard’s function, a 3-parameter or generalized logistic model (Eq. 1), was the most appropriate to describe AD progression over time as assessed by AIC. Richard’s function allowed for an asymmetric, inverted, concave relationship between disease progression rate and severity (Fig. 4). The parameter β allowed for asymmetry with respect to the inflection point which was estimated to occur at an ADAS-cog of ~52.

Final model

Richard’s function was utilized as the base model for the covariate model building. The final model parameter

estimates are presented in Table 3. NONMEM code is presented in the Appendix. The identified covariates of baseline ADAS-cog were sex and APOE*4 genotype. Baseline ADAS-cog was lower for males when compared to females and increased with the number of APOE*4 alleles. The identified covariates of rate of progression were age, APOE*4 genotype and AD medication status. Patients who were younger, on stable background AD medication, and with more APOE*4 alleles had a higher rate of progression. The count of APOE*4 alleles (when known) showed a linear allelic effect on disease progression. In addition, the estimated average number of APOE*4 copies in patients with unknown genotype was very similar to the observed average number of copies in patients with known genotype (0.78 vs. 0.72, respectively). As consequence, we considered APOE*4 genotype as a continuous variable in the final model and imputed missing allele count as 0.72. Importantly, similar estimates were obtained for APOE*4 effect on both baseline and rate of progression when the model was run excluding patients with imputed genotype. The model-predicted average ADAS-cog progression stratified by covariates is presented in Fig. 5. Although patient enrollment year in the trial was not included in the covariate model building, Fig. 6a depicts patient enrollment year in the trial versus the baseline MMSE and shows that the

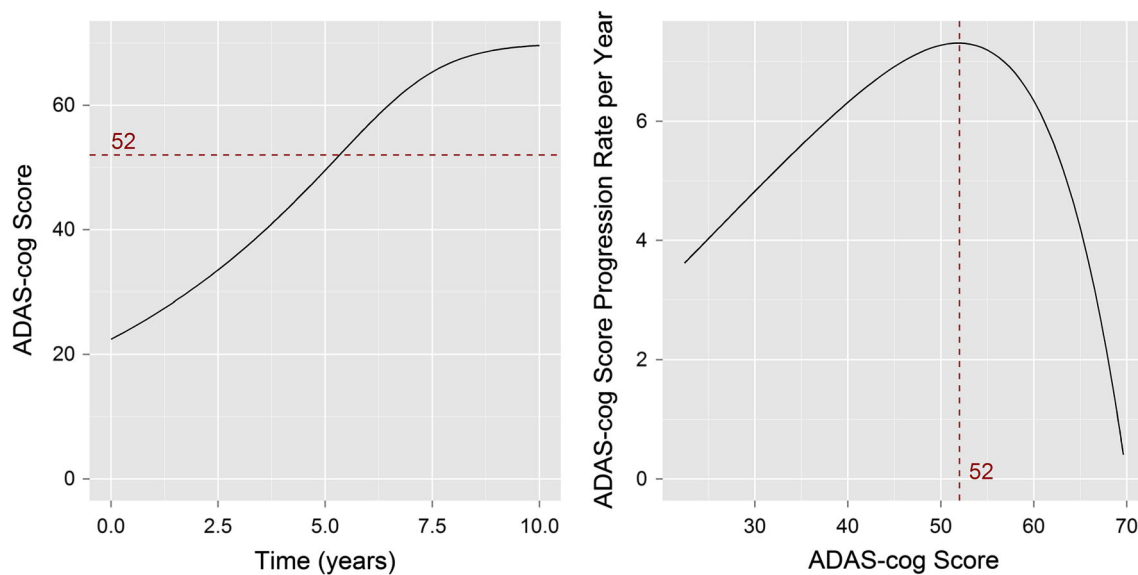


Fig. 4 Non-linearity of rate of disease progression estimated with the Richard's function. Dashed line is the estimated inflection point of 52

Table 3 Base and final model parameter estimates

Parameter	Base model parameter estimates	Final model parameter estimates	Final model parameter estimates (95 % CI lower, upper) from Markov chain Monte Carlo Bayesian
θ_{baseline}	21.7	22.2	22.3 (22.0, 22.6)
$\theta_{\text{sex (male)}}$	–	0.953	0.955 (0.934, 0.976)
$\theta_{\text{apoe4 (centered at 0.72)}}$	–	0.0372	0.0379 (0.013, 0.063)
$\theta_{\text{intrinsic rate (year}^{-1}\text{)}}$	0.147	0.153	0.153 (0.143, 0.163)
$\theta_{\text{age (centered at 75)}}$	–	–0.024	–0.024 (–0.031, –0.017)
$\theta_{\text{apoe4 (centered at 0.72)}}$	–	0.195	0.199 (0.078, 0.320)
$\theta_{\text{comed (no AD medication)}}$	–	–0.302	–0.303 (–0.446, –0.160)
β	7.15	6.91	6.97 (5.58, 8.36)
SD of $\eta_{\text{baseline}}^{\text{patient}}$	0.405	0.395	0.399 (0.389, 0.408)
SD of $\eta_{\text{intrinsic rate}}^{\text{patient}}$	0.206	0.203	0.206 (0.197, 0.215)
SD of $\eta_{\text{baseline}}^{\text{study}}$	–	0.099	0.094 (0.034, 0.129)
SD of $\eta_{\text{intrinsic rate}}^{\text{study}}$	–	0.026	0.024 (0.003, 0.034)
τ	87.8	87.5	87.3 (85.4, 89.2)

SD is standard deviation; η^{patient} is the inter-individual variability; η^{study} is the inter-study variability. Inter-study variability was not introduced in the base model. The covariate relationships were as following: $ADAS_{\text{cog}0} = \theta_{\text{ADAS}_{\text{cog}0}} \cdot \text{SEX} \cdot \text{APOE4}'$ and $r = \theta_r \cdot \text{AGE} \cdot \text{APOE4}'' \cdot \text{COMED}$. Where: $\text{SEX} = \theta_{\text{sex(male)}}$; $\text{APOE4}' = 1 + \theta'_{\text{apoe4}} \cdot (\text{APOE4} - 0.72)$, with 0.72 corresponding to the median value; $\text{AGE} = 1 + \theta_{\text{age}} \cdot (\text{AGE} - 75)$, with 75 corresponding to the median value; $\text{APOE4}'' = 1 + \theta''_{\text{apoe4}} \cdot (\text{APOE4} - 0.72)$, with 0.72 corresponding to the median value; $\text{COMED} = 1 + \theta_{\text{comed(no AD medication)}}$, with 1 corresponding to patients on AD medication

baseline disease severity has only slightly changed from 1994 to 2008 (from approximately 19.5–20.7, $P < 0.0001$; based on the linear regression analysis in the individuals who were not on background AD medication). This finding is in agreement with the literature (summary-level) data presented in Fig. 6b.

Inter-individual variability was larger than inter-study variability. The inter-individual variability for baseline ADAS-cog was about twofold higher than that of the rate of progression (0.395 vs. 0.203, respectively). The inter-study variability was lower for both baseline ADAS-cog and rate of progression (0.099 vs. 0.026).

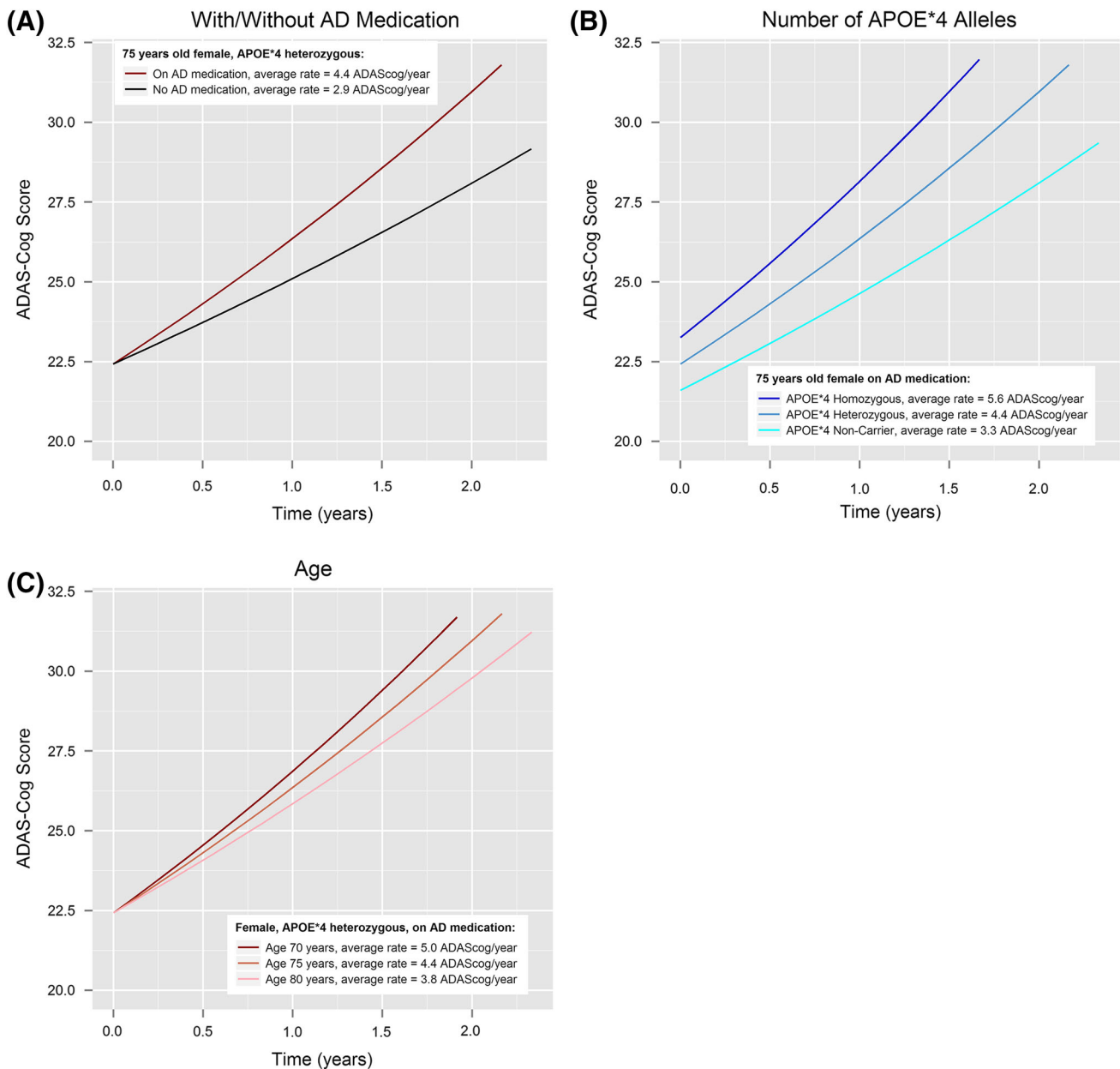


Fig. 5 Model predicted average ADAS-cog progression

Evaluation of model performance

Model selection criteria suggest the appropriateness of the model in describing the longitudinal changes in ADAS-cog score. The residuals plots (Supplementary Figure S1) did not indicate a trend of the residuals versus the population predictions and time. The observed ADAS-cog scores versus individual predictions approximated the identity line; the same was not quite observed for population predictions, likely due to the skewness in ADAS-cog scores. Figure 7 shows the VPCs stratified by

study, number of APOE*4 alleles and use of background AD medication. Most of the observed data lie within the 5th and 95th percentiles of the simulations, and the observed and predicted percentiles are similar indicating reasonable estimation. In addition, the condition number from the Laplacian estimation method was 473 and the final parameter values were estimated with good precision when utilizing the Markov chain Monte Carlo Bayesian estimation method, and shrinkage was minimal for inter-study, inter-individual, and residual variability (Table 3).

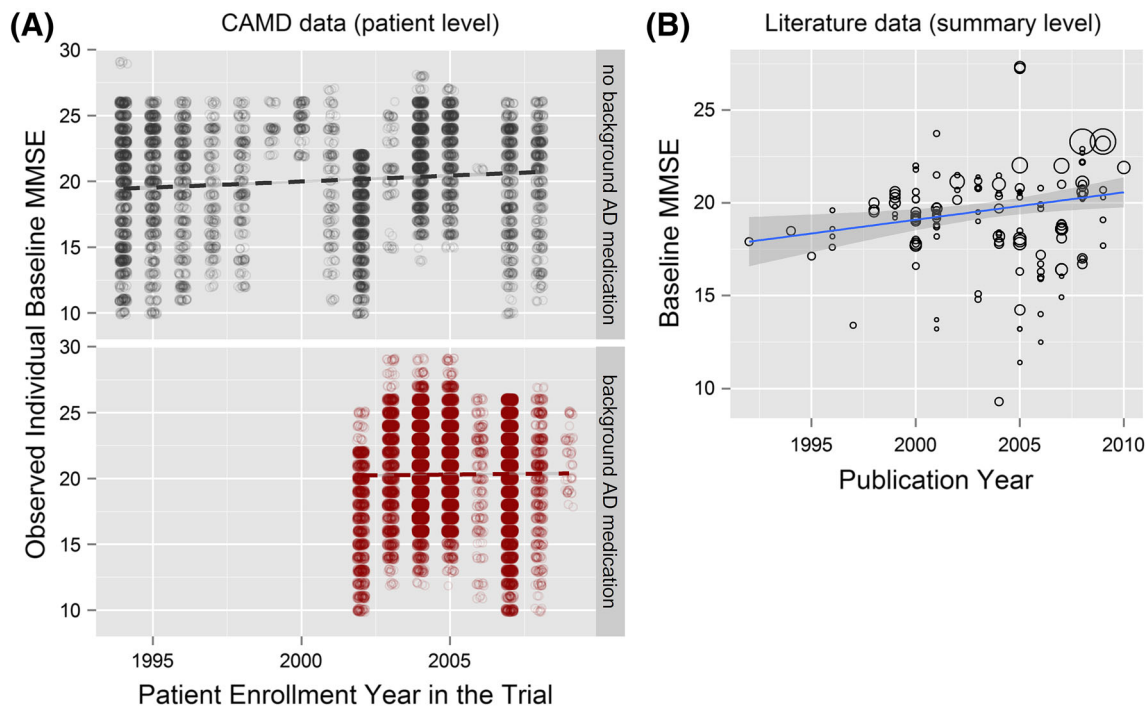


Fig. 6 Observed baseline disease severity over year. **a** Circles indicate each individual patient data, and *line* is the weighted linear regression. **b** Circles indicate mean values from each published trial,

size of the *circle* is proportional to the number of patients in the trial, *line* is the weighted linear regression by number of patients in the trial, and *shaded area* is the standard error

Discussion

This work utilized the accumulated knowledge on AD progression through the ADAS-cog longitudinal models available in the literature and expanded our understanding of the predictors of AD progression rate. In this analysis, we tested all model structures published previously [5–9, 11–13] and chose a generalized logistic function (i.e., Richard's function) to describe changes in ADAS-cog over time for patients with mild-to-moderate AD dementia. This analysis also incorporated a mixed-effects beta regression along with a third level of random effect for study in NONMEM.

Our analysis shows that an average (i.e., typical) patient with mild-to-moderate AD in the CAMD database is represented by a 75-year-old female on standard-of-care AD therapy and APOE*4 status unknown (i.e., genotype information was not available). We considered APOE*4 genotype as a continuous variable in the final model and imputed missing allele count as 0.72 (i.e., the observed average number of alleles in patients with known genotype) as described in the results section. This patient would enter a study with a baseline ADAS-cog score of 22.2 and an initial rate of progression of 3.4 ADAS-cog points/year. The estimated baseline ADAS-cog agrees with the observed average baseline ADAS-cog score of 24 reported in the Table 2. The rate of progression aligns with the work

conducted by Rogers et al. [13] who reported a rate of natural disease progression of 3.97 ADAS-cog score/year for an average 75-year-old female with MMSE of 21 and APOE*4 heterozygous. According to the mapping function proposed in Fig. 2, an MMSE of 21 corresponds to an ADAS-cog score of 22.4. Compared to the baseline ADAS-cog of an average patient, males had a ~5 % lower baseline value; APOE*4 non-carriers and homozygous, a ~2.7 % lower and ~4.8 % higher baseline value, respectively (Fig. 6). Compared to the initial rate of progression for such an average patient, individuals who are not on AD medication had a ~30 % lower progression rate. Additionally, APOE*4 non-carriers had a ~14 % lower progression rate, while APOE*4 homozygous have a ~25 % higher rate (Fig. 5). In addition, younger baseline age was associated with higher rate of progression (i.e., a 70-year-old had a ~12 % higher, and an 80-year-old, a 12 % lower initial rate when compared to a 75-year-old patient).

The asymmetric, inverted, concave relationship between AD progression rate and severity was appropriately described by the Richard's function. This function describes an increase in the rate of change when ADAS-cog is below the inflection point, and a decrease in the rate of change when ADAS-cog is above the inflection point (Fig. 4). The restriction in the rate of increase in ADAS-cog score is imposed by the second term of the Richard's

function (Eq. 1): as ADAS-cog scores approach 70, the rate of change tends to zero, reflecting a vertical asymptote in rate or plateau in score (Fig. 4). This is coherent with what is seen in clinical practice: patients with very severe AD dementia tend to experience loss of speech which compromises their ability to perform the tasks that compose the ADAS-cog scale [4]. As a consequence, ADAS-cog scores mathematically plateau, and the worsening in cognitive impairment can no longer be captured by the scale (i.e., ceiling effect). In severe AD dementia, other clinical endpoints are recommended, for instance, the Severe Impairment Battery [25]. Another aspect is that the shape parameter of the Richard's function allows for an inflection point that is not restricted to the midpoint of the scale. In this analysis, the estimated inflection point of 52 is closer to the top of the scale, and similar to the inflection point of 46 estimated in the previous work utilizing CAMD data [13].

The incorporation of beta-distributed residual variability along with a logistic model constrained the model prediction within the realistic range of ADAS-cog scores of 0–70. The variance of the beta distribution depends on the expected value and τ as shown in Eqs. 6 and 7. This accommodates the heteroscedasticity of the residual variability because the variance is lower when ADAS-cog scores are close to the boundaries. In our analysis, we estimated a τ of 87.5. For example, for an expected ADAS-cog score of 22.2, the calculated standard deviation of the beta distribution is ~ 0.05 .

The predictors of baseline ADAS-cog were sex and APOE*4 genotype; whereas the predictors of rate of progression were baseline age, APOE*4 genotype and AD medication status. Their effects were statistically significant based on the 95 % confidence intervals which did not include zero (Table 3). Previous AD models showed a faster disease progression rate related to increase in disease severity [8, 9, 12], younger baseline age [9, 12, 13], APOE*4 carrier status [9, 12, 13], and female [9, 13]. The Richard's function intrinsically accounts for the effect of disease severity in the rate of progression by considering the ADAS-cog score at any time point (e.g., rate of progression increases with ADAS-cog score below the inflection point); hence, all covariates associated with a higher rate of progression have additional effects given a certain disease severity.

The effects of APOE*4 genotype on the rate of AD progression is noteworthy. Our analysis indicates an additive allelic effect on disease progression (Fig. 5b). This is a novel finding for the AD research community. Although the APOE*4 has been associated with a higher rate of disease progression in previous publications, Ito et al. and Samtani et al. modeled the APOE*4 genotypes as two categories (i.e., non-carriers vs. carriers) [9, 12]. Rogers

et al. [13] modeled APOE*4 genotype as three categories (i.e., non-carriers, heterozygous and homozygous) but the estimated rate of disease progression in heterozygous did not statistically differ from that in non-carriers. APOE*4 has been implicated as the strongest risk factor for developing late onset AD but the mechanism remains unclear [10, 26]. It has been hypothesized that the APOE*4 variant is associated with build-up of amyloid plaques in the brain, a hallmark of AD pathology believed to contribute to neuronal death [10]. For example, Corder et al. [27] reported that the risk for AD increased from 20 to 90 % and the average age of disease onset decreased from 84 to 68 years with increasing number of APOE*4 alleles in 42 families. Also, Xu et al. [28] reported that one APOE*4 allele has been associated with increased risk of mild cognitive impairment, whereas two APOE*4 alleles have been associated with increased risk of dementia suggesting that APOE*4 homozygous patients have a very short transition through the mild cognitive impairment stage. Therefore, our findings are coherent with prior findings of a significant allelic effect on disease progression.

The effect of AD medication status on the rate of progression should be interpreted with caution. In this analysis, we have observed a faster progression in patients on stable background AD medications compared to those who are not receiving AD medication (Fig. 5a). First, the number of patients who are not receiving AD medication in studies longer than 30 weeks represents a minor sample size ($N = 475$, ~ 15 %) as compared to the total number of patients in such studies ($N = 3,196$). Second, the current standard-of-care medications for AD are AChEIs or/and memantine, which are all approved as symptomatic drugs (i.e., not disease modifying). As a consequence, these drugs are expected to have an effect equivalent to a reduction in the baseline ADAS-cog (i.e., offset effect), but not to affect the rate of disease progression. Therefore, the higher rate of disease progression in patients on stable background medication is unexpected. Third, if patients who are on AD medication truly had a higher baseline ADAS-cog, their disease should progress at a higher rate according to the Richard's function (the rate of change in ADAS-cog increases with ADAS-cog score below the inflection point; Fig. 4). However, this higher "true" baseline hypothesis would not fully explain the >20 % higher rate of progression with patient on stable background medication considering the relatively small symptomatic effect of AChEIs or memantine (e.g., the effect size of AChEIs is 2–4 points decrease in ADAS-cog [8]). One possible explanation is that patients who are already on stable background AD medication are in a more severe stage of the disease than patients who are not, and this is not adequately captured by baseline MMSE and ADAS-cog; in this case, the underlying disease progression is faster in

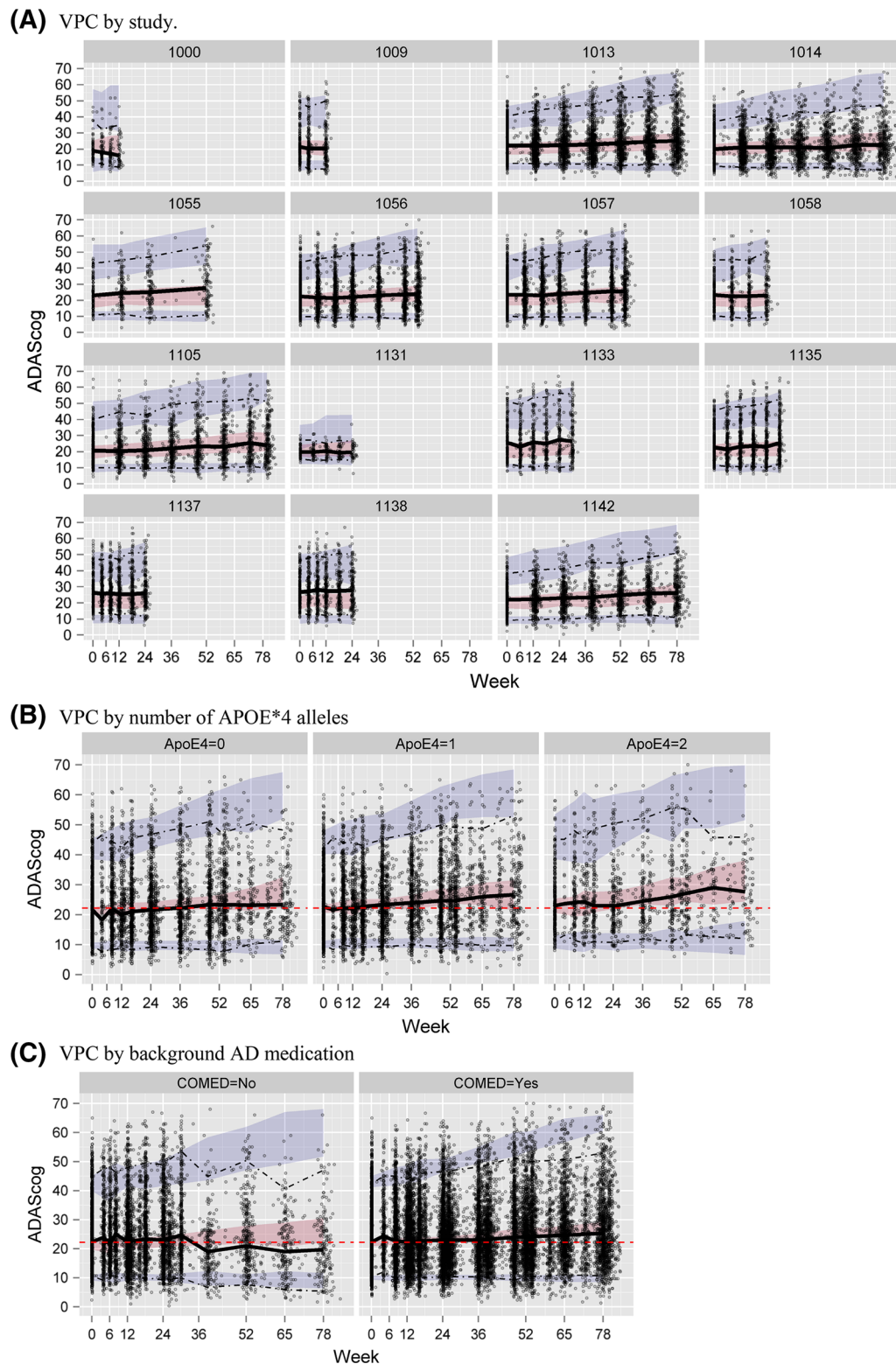


Fig. 7 Visual Predictive Check (VPC) from $N = 500$ Simulations. *Solid and dashed black lines* are observed median, 5th and 95th percentiles, and shaded areas are corresponding predicted intervals from the final model. *Horizontal red dashed line* is ADAS-cog

baseline (22.2) for an average patient with mild-to-moderate AD in the CAMD database. A difference variance structure was used for Study 1131 in which patients with milder AD were enrolled (Color figure online)

patients on AD medication, which would not be modified by purely symptomatic agents, like the ones received by this population. In studies 1013 (81 % of the patients on AD medication) and 1014 (76 % of the patients on AD medication), patients on AD medication had an average higher baseline ADAS-cog and faster disease progression (Fig. 8). A possible more convincing explanation is that potential confounders are not available in the CAMD database; for instance, other aspects besides cognition (i.e., impaired ability to function and perform in daily life, behavioral problems and co-morbidities) that lead clinicians to diagnose AD and prescribe medications [29, 30]. No single assessment tool can capture the multifaceted aspect of AD progression; hence, decisions as to treatment strategies were made not only on the basis of a single outcome measure. Therefore, the current knowledge does not support a causality relationship between use of standard-of-care medications and higher rate of disease progression. This observed correlation is likely due to underlying information currently unavailable in the trials integrated within the CAMD database, which can help explain several additional aspects of AD progression. It is also noted that AD medication is a significant covariate only for the rate of progression and not for the baseline ADAS-cog. There are two possible explanations for this: (1) in studies 1013 and 1014, the number of patients who are not on AD medication is much smaller when compared to the number of patients who are on AD medication (Fig. 8); (2) except studies 1013 and 1014, the other studies had only patients who either used or did not use background AD medication; hence, large inter-individual and

inter-study variability may have masked the AD medication effect for baseline ADAS-cog.

Limitations of this analysis include the need to develop a dropout model utilizing this updated CAMD database. The average dropout was estimated as 14 % (95 % CI 10–17 %) using a random effects meta-analysis model (metafor package in R), which accounts for between-study variability and heterogeneity. Thus, accounting for missing data becomes important when conducting simulation-based diagnostics or clinical trial simulations. Although the VPC presented in Fig. 7 does not seem greatly compromised, dropout should be accounted for when utilizing this model as a clinical trial simulation tool. An average higher baseline ADAS-cog—26.1 (95 % CI 23.9–28.3) versus 23.8 (95 % CI 22.5–25.0)—and an average older age—75.8 (95 % CI 74.7–76.9) versus 73.9 (95 % CI 73.0–74.7)—was estimated for patients who dropped out using a random effects meta-analysis model. This is in agreement with the dropout model developed by Rogers et al. [13] Second, we have only utilized data from the CAMD database, hence, control-arm data. For this reason, we were not able to differentiate between underlying natural disease progression and placebo effect in the clinical trials. Based on a summary-level meta-analysis of 52 trials, Ito et al. [8] estimated that the maximum placebo effect occurred at around 11 weeks, and disappeared within a year. Third, because year of enrollment was found to be correlated with availability of APOE*4 genotype information and AD medication status, we have not systematically tested its effect on ADAS-cog baseline and rate of progression. However, year of enrollment versus the

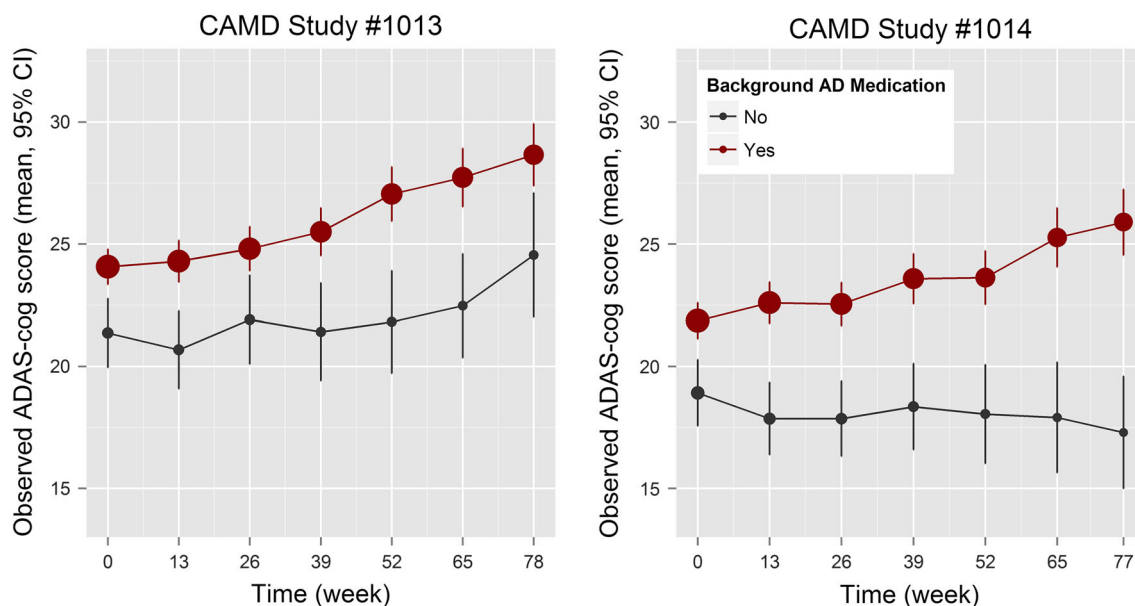


Fig. 8 Mean observed ADAS-cog score for CAMD studies 1013 and 1014. Area of the points is proportional to the number of patients at each time point

baseline MMSE indicates that the baseline disease severity has only slightly changed from 1994 to 2008 (Fig. 6a). Fourth, other biomarkers such as cerebrospinal fluid and imaging volumetric brain measures could not be tested as predictors of disease severity or progression due to the unavailable biomarker information in the current CAMD database.

In summary, to our knowledge this is the first work that evaluates the effect of stable background AD medication (vs. placebo only) and the effect of APOE*4 genotypes across 15 clinical studies using patient-level data from the CAMD database on baseline ADAS-cog and its rate of progression. We also investigated whether baseline disease severity had changed between older and more recent trials, possibly due to changes in life style, awareness of the disease, diagnostics and patient care. Our model suggests that faster AD progression is associated with younger baseline age and higher number of APOE*4 alleles, after accounting for baseline disease severity. APOE*4, in particular, seems to be implicated in the AD pathogenesis, and genotyping is especially important for reducing variability in trial results. To be interpreted with caution, patients

entering studies on stable background AD medications appear to have a faster progression than those without background AD medication. The current knowledge does not support or implicate a causality relationship between use of background AD medications and higher rate of disease progression, and the association, if validated, might be due to confounding covariates. Although causality has not necessarily been demonstrated, these factors can inform inclusion criteria and stratification, sample size, and trial duration. Finally, this work expands our understanding of the predictors of AD progression and our model informs relevant aspects to help design a clinical trial on a novel AD medication.

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Appendix: NONMEM 7.3 code for final model

```

$PROBLEM CAMD ADAS-COG TOTAL

$INPUT

ID                ; Patient identification
STUDY             ; Study identification
TIME              ; Time (days)
AGE               ; Patient age (years)
SEX               ; Patient sex (SEX=1, if patient is female SEX=2, if patient is male)
MDV               ; Missing dependent variable
ADASTRANS=DV     ; Dependent variable, this is the transformed ADAS-cog score to the unit
                  ; interval (0,1)
COMED2            ; AD medication status (COMED2=1, if patient is on stable background AD
                  ; Medication; COMED2=0, if patient is on placebo)
APOE4C            ; APOE*4 number of alleles (as a continuous variable)

$DATA nonmemdata022614.csv IGNORE = @ IGNORE = (MDV.EQ.1, ID.EQ.528)

$PRED

```


; Defining Covariate Relationships

```

;;; BLAPOE4C-DEFINITION START
BLAPOE4C = ( 1 + THETA(9)*(APOE4C - 0.72))
;;; BLAPOE4C-DEFINITION END

;;; SLCOMED2-DEFINITION START
IF(COMED2.EQ.1) SLCOMED2 = 1 ; Most common
IF(COMED2.EQ.0) SLCOMED2 = 1+THETA(8)
;;; SLCOMED2-DEFINITION END

;;; SLAPOE4C-DEFINITION START
SLAPOE4C = ( 1 + THETA(7)*(APOE4C - 0.72))
;;; SLAPOE4C-DEFINITION END

;;; SLAGE-DEFINITION START
SLAGE = ( 1 + THETA(6)*(AGE - 75.00))
;;; SLAGE-DEFINITION END

;;; SL-RELATION START
SLCOV=SLAGE*SLAPOE4C*SLCOMED2
;;; SL-RELATION END

;;; BLSEX-DEFINITION START
IF(SEX.EQ.1) BLSEX = 1 ; Most common
IF(SEX.EQ.2) BLSEX = THETA(5)
;;; BLSEX-DEFINITION END

;;; BL-RELATION START
BLCOV=BLSEX*BLAPOE4C
;;; BL-RELATION END

```

; Time Unit Conversion

```
YTIME = TIME/365.25 ; Time in years
```

; Defining Thetas

```

TVBL = THETA(1) ; Population baseline ADAS-Cog
TVBL = BLCOV*TVBL
TVSL = THETA(2) ; Population intrinsic rate of progression
TVSL = SLCOV*TVSL

STEWSSV = 1 ; Within-individual variability for study 1131
STEBSSV = 1 ; Inter-individual variability for study 1131
IF(STUDY.EQ.1131) THEN
  STEWSSV = EXP(THETA(10))
  STEBSSV = EXP(THETA(11))
ENDIF

BL = TVBL*EXP(ETA(1)*STEBSSV + ETA(3))
SL = TVSL + ETA(2) + ETA(4)

TVSHAPE = THETA(3) ; Population shape factor of the Richard's function
TAU = THETA(4)*STEWSSV ; Tau is the precision parameter of the beta distribution (=ALPHA+BETA)
; Variance of the beta distribution is MUR*(1-MUR)/(TAU+1),
; meaning that the variance increases as TAU decreases;
; although, it is not the sole determinant of dispersion,
; since dispersion also depends on MUR

```

```

; Structural Model to Describe the Rate of Disease Progression
; Using Richards logistic growth model: three-parameter logistic model
; Tsoularis & Wallace. Mathematical Biosciences, 179:21-55, 2002.

; GG=1.0E-30

; IF (BL<GG) BL=GG
DEN1 = BL**TVSHAPE
DEN2 = (70**TVSHAPE) - (BL**TVSHAPE)
DEN3 = EXP(-TVSHAPE*SL*YTIME)
DENN = DEN1 + DEN2*DEN3
; IF (DENN<GG) DENN=GG
MUR = BL/((DENN)**(1/TVSHAPE)) ; Individual ADAS-cog response

; IF (TVBL<GG) TVBL=GG
TVDEN1 = TVBL**TVSHAPE
TVDEN2 = (70**TVSHAPE) - (TVBL**TVSHAPE)
TVDEN3 = EXP(-TVSHAPE*TVSL*YTIME)
TVDENN = TVDEN1 + TVDEN2*TVDEN3
; IF (TVDENN<GG) TVDENN=GG
TVMUR = TVBL/((TVDENN)**(1/TVSHAPE)) ; Population ADAS-Cog response

F = MUR
IPRED = F

; Beta Regression
; Reference: Smithson & Verkuilen. Psychological Methods, 11: 54-71, 2006
; MUR is the location parameter of the beta distribution
; TAU is the precision parameter of the beta distribution
; MUR and TAU have no restriction on each other, so that they can be modeled
; separately

ALPHA = MUR*TAU ; Beta distribution shape parameter pulling density toward 0 (alpha>0)
BETA = (1-MUR)*TAU ; Beta distribution shape parameter pulling density toward 1 (beta >0)

X1 = ALPHA+BETA ; It is really TAU
X2 = ALPHA
X3 = BETA

; Nemes Approximation of the LN(Gamma) Distribution

; Bound the result to assist with convergence
; GG1=1.0E-150
; IF (X1<GG1) X1=GG1
; IF (X2<GG1) X2=GG1
; IF (X3<GG1) X3=GG1

LGAMMAX1 = X1*(LOG(X1)-1) + 0.5*(LOG(2*3.1415)-LOG(X1)) + (5/4)*X1*LOG(1+(1/(15*X1**2)))
LGAMMAX2 = X2*(LOG(X2)-1) + 0.5*(LOG(2*3.1415)-LOG(X2)) + (5/4)*X2*LOG(1+(1/(15*X2**2)))
LGAMMAX3 = X3*(LOG(X3)-1) + 0.5*(LOG(2*3.1415)-LOG(X3)) + (5/4)*X3*LOG(1+(1/(15*X3**2)))

; Log-likelihood of the beta distribution (LN of the probability density function of the beta
; distribution)

WDV=DV
IF (WDV<0.001) THEN
  WDV=0.001
ENDIF
IF (WDV>0.999) THEN

```

```

WDV=0.999
ENDIF

LLBETA = LGAMMAX1 - LGAMMAX2 - LGAMMAX3 + (ALPHA-1)*LOG(WDV) + (BETA-1)*LOG(1-WDV)

; Defining Residuals and Y

SOR = (WDV-IPRED)/SQRT(IPRED*(1-IPRED)/(1+TAU)) ;Standardized ordinary residuals

LLB2=-2.0*LLBETA

; IF (LLB2.GT.300.0) LLB2=300.0
; IF (LLB2.LT.-300.0) LLB2=-300.0

Y = LLB2

; Initial Estimates

$THETA
(0, 22.3, 100) ; 1 TVBL
(-100, 0.151, 50) ; 2 TVSL
(0, 6.98, 100) ; 3 TVSHAPE
(-300, 87.8, 300) ; 4 TAU
(0,1) ; 5 BLSEX1
(0.1) ; 6 SLAGE1
(0.2) ; 7 SLAPOE4C1
(-0.5, -0.275, 0.1) ; 8 SLCOMED21
(0.1) ; 9 BLAPOE4C1
(0.1) ; 10 STE WIV EFFECT
(0.1) ; 11 STE IIV EFFECT

$OMEGA BLOCK(2) ; Inter-individual variability block
0.156 ; IIV_BL
0.0224 ; CORR IIV_BL:IIV_SL
0.0424 ; IIV_SL

$OMEGA BLOCK(2) ; Inter-study variability block
0.0084 ; ISV_BL
0.0014 ; CORR ISV_BL:ISV_SL
0.0004 ; ISV_SL

$LEVEL ; Defines that ETAs for individual are nested within study
STUDY=(3[1],4[2]) ; Means that ETA1 is nested within ETA3, and ETA2 is nested within ETA4

$ESTIMATION MAXEVAL=9999 PRINT=1 METHOD=COND -2LL LAPLACIAN NOHABORT FNLETA=0 NSIG=2 SIGL=7 SLOW
MCETA=10 NONINFETA=1 FILE=run4.ext

$COV MATRIX=R UNCONDITIONAL SIGL=9 SLOW PRINT=E

```

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