

# Life in Burrows Channelled the Morphological Evolution of the Skull in Rodents: the Case of African Mole-Rats (Bathyergidae, Rodentia)

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**Abstract** African mole-rats are fossorial rodents that consist of five chisel-tooth digging genera (*Heterocephalus*, *Heliophobius*, *Georychus*, *Fukomys*, and *Cryptomys*) and one scratch digger (*Bathyergus*). They are characterized by striking physiological, morphological, and behavioral adaptations intimately related to their subterranean life. The influence of their mode of life in shaping the cranial morphology has yet to be evaluated in comparison to other Ctenohystrica, especially fossorial genera, which include the subterranean genera *Spalacopus* and *Ctenomys*. In our study, we seek to determine to what extent subterranean life affects the morpho-functional properties of the skull among fossorial ctenohystricans. 3D geometric morphometric analyses were performed on 277 skulls, encompassing 63 genera of Ctenohystrica, and complemented by biomechanical studies. African mole-rats and other subterranean Ctenohystrica, especially chisel-tooth diggers, have a short snout, a wide cranium with enlarged zygomatic arches, and a strongly hystricognathous mandible. Even if convergences are also manifest between most

fossorial Ctenohystrica, subterranean rodents departed from the main ctenohystrican allometric trends in having a skull shape less size-dependent, but under stronger directional selection with intense digging activity as a major constraint. African mole-rats, notably chisel-tooth diggers, show important mechanical advantage for the temporalis muscles favoring higher forces at the bite point, while mechanical advantage of the superficial masseter muscles is lower compared to other Ctenohystrica. If subterranean species can be clearly discriminated based on their skull morphology, the intrinsic mosaic of anatomical characters of each genus (e.g., skull, teeth, and muscles) can be understood only in the light of their ecology and evolutionary history.

**Keywords** Ctenohystrica · Masticatory apparatus · Fossorial · Subterranean · Geometric morphometrics · Biomechanics

## Introduction

Among placental mammals, rodents show a very wide range of modes of life (e.g., cursorial, scansorial, jumping, flying, aquatic, fossorial), and this ecological diversity partly explains their blooming evolutionary history. As fossorial rodents do not have the appeal of various more charismatic mammals, in part due to their cryptic lifestyle, they did not attract the interest of researchers until the end of the 1970's (e.g., Nevo 1979). However, recent advances in the last decades have enabled a better understanding of the behavior and the striking biological characteristics of these fascinating rodents (Nevo 1999; Lacey et al. 2000; Begall et al. 2007). Rodents are defined as fossorial when they show specializations for digging (e.g., Lacey et al. 2000). Fossorial rodents are widespread nowadays, and they are present in diverse extant groups, phylogenetically distant, such as the Aplodontidae, Sciuridae,

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Bathyergidae, Ctenomyidae, Octodontidae, Echimyidae, Geomyidae, Spalacidae, and Cricetidae (Arvicolinae and Sigmodontinae). The fossorial lifestyle is more or less developed in rodents. Less specialized fossorial species spend a significant part of their lifetime aboveground, especially for foraging (e.g., some Echimyidae and Cricetidae). Conversely, some rodents show a high fossorial activity (e.g., Bathyergidae, Geomyidae, and some Spalacidae); they spend most of their lifetime underground, including when looking for food and sexual partners, but also to disperse. These species are generally referred to as subterranean (Nevo 1979). In general, fossorial rodents use two main types of digging tools: their strong forearms bearing robust claws for scratching, and/or their prominent and procumbent incisors as a chisel to break up, loosen, and remove the soil. These adaptive characters repeatedly appeared during the rodent evolutionary history.

The first known fossorial rodents probably appeared during the end of the Eocene (~40 Mya, Matthew and Granger 1923; Nevo 1999; Jardine et al. 2012). This lifestyle became more common during the Oligocene, probably because of new types of food resources available and the need of finding alternative shelters in relation to increasing aridity combined with gradual disappearance of forested environments (Nevo 1999; Jardine et al. 2012). Apart from pocket gophers (Geomyidae) and relatives of the mountain beaver (Aplodontioidea), which are among a few extant fossorial groups diversified during the Oligocene epoch (Jardine et al. 2012), the diversity of most of extant groups is the result of recent adaptive radiations (15–5 Mya), such as African mole-rats (Bathyergidae; Cook et al. 2000).

African mole-rats belong to the Ctenohystrica, and comprise species using both main modes of digging (Stein 2000) with five chisel-tooth digging genera (*Heterocephalus*, *Heliophobius*, *Georychus*, *Fukomys*, and *Cryptomys*) and one scratch digging genus (*Bathyergus*). These mole-rats acquired many peculiar morphological, physiological, and behavioral adaptations as a consequence of strict life in subterranean environment (Bennett and Faulkes 2000). Among these adaptations, the most striking ones remain the low basal metabolic rate that likely helped them to compensate high energetic costs of foraging (Zelová et al. 2007), and the high thermal conductance that limits overheating due to activity in humid subterranean environments. Bathyergid rodents also display morphological adaptations related to their strict subterranean life, the most remarkable being the procumbent incisors, and reduced eyes and pinnae (Stein 2000). Interestingly, *Heliophobius* is one of the rare examples of mammals that shows a continuous dental replacement characterized by the constant development of molars at the back of the jaw, then migrating toward the front (Gomes Rodrigues et al. 2011). This adaptation is likely to be linked to strong dental wear partly due to the impossibility to use cooperative digging strategies combined with a high chisel-tooth digging activity in

different soils, which could be extremely hard during the dry season (Gomes Rodrigues et al. 2011). As most of the physiological and anatomical characteristics of bathyergids are deeply influenced by their subterranean lifestyle, we seek to determine to what extent the cranial morphology and the associated mechanics of masticatory muscles were impacted in these rodents compared to their relatives.

The masticatory apparatus of rodents is a highly plastic region of the skull, and some studies have already shown that its shaping shows a significant ecological component driven by factors such as habitat and feeding habit (Michaux et al. 2008; Hautier et al. 2012; Casanovas-Vilar and Van Dam 2013; Renaud et al. 2015). Our aim is to evaluate cranial and mandibular variations between African mole-rats and other Ctenohystrica in order to assess the influence of phylogenetic and size components on skull morphology. Variations have also been measured within African mole-rats in relation to their digging skills. Finally, we focus on the morpho-functional properties of the masticatory apparatus to draw comparisons between fossorial Ctenohystrica based on their mode of digging and their foraging activity (see Table 1).

## Material and Methods

### Sample Composition

The database compiled by Hautier et al. (2012) has been complemented by new material of Ctenohystrica from the Natural History Museum in London (NHM). We analyzed 246 mandibles and 277 crania belonging to rodents of both sexes (but data are frequently missing), representing 63 genera and 17 families of Ctenohystrica (Table 1, Online resource 1), and most bathyergid genera were added. Bathyergids were not investigated in Hautier et al. (2011, 2012) because their peculiar cranial morphology precludes the retrieval of some of the landmarks defined on the crania and mandibles. *Cryptomys* was not considered here; however, this genus is very close to *Fukomys* in shape (in fact, *Fukomys* was recently split from *Cryptomys* sensu lato based mainly on genetic data; see Kock et al. 2006 for further details). In our study, African mole-rats (Bathyergidae), *Spalacopus* (Octodontidae), and *Ctenomys* (Ctenomyidae), which have a prevailing subterranean activity (see Table 1), were designated as subterranean rodents (sensu Nevo 1979).

### Geometric Morphometric Methods

The mandibular and cranial forms were quantified with 19 and 59 anatomical landmarks, respectively (Fig. 1 and Online resource 2). Digital data of all specimens were acquired using a Microscribe 3-D digitizer and using X-ray micro-computed tomography ( $\mu$ CT). One skull of *Heliophobius* (ID 13) was

**Table 1** List of analyzed taxa, material, mode of digging and foraging activity of fossorial species (in bold)

| Family           | Genus  | Cranium      | Mandible   | Mode of digging / foraging activity |
|------------------|--|--------------|------------|-------------------------------------|
| Abrocomiidae     | <i>Abrocoma</i>  | 4            | 5          |                                     |
| Bathyergidae     | <b><i>Bathyergus</i>*</b>  | <b>12</b>    | <b>7</b>   | <b>Scratch/U(A)</b>                 |
|                  | <b><i>Fukomys vandewoestijneae</i>* / <i>Fukomys mechowii</i>*</b> | <b>10/10</b> | <b>5/7</b> | <b>Chisel-tooth/U</b>               |
|                  | <b><i>Georchus</i>*</b>  | <b>3</b>     | <b>3</b>   | <b>Chisel-tooth/U</b>               |
|                  | <b><i>Heliophobius</i>*</b>  | <b>13</b>    | <b>8</b>   | <b>Chisel-tooth/U</b>               |
|                  | <b><i>Heterocephalus</i>*</b>                                      | <b>10</b>    | <b>10</b>  | <b>Chisel-tooth/U</b>               |
| Capromyidae      | <i>Capromys</i>  | 4            | 4          |                                     |
| Caviidae         | <i>Cavia</i>   | 5            | 5          |                                     |
|                  | <i>Caviella</i>  | 2            | 2          |                                     |
|                  | <i>Dolichotis</i>  | 5            | 5          |                                     |
|                  | <i>Galea</i>   | 2            | 2          |                                     |
|                  | <i>Hydrochaerus</i>  | 6            | 6          |                                     |
|                  | <i>Kerodon</i>   | 1            | 1          |                                     |
|                  | <i>Monticavia</i>  | 2            | 2          |                                     |
|                  | <i>Nanocavia</i>   | 2            | 2          |                                     |
| Chinchillidae    | <i>Chinchilla</i>  | 3            | 3          |                                     |
|                  | <i>Lagidium</i>  | 5            | 5          |                                     |
|                  | <b><i>Lagostomus</i></b>   | <b>6</b>     | <b>6</b>   | <b>Scratch/A</b>                    |
| Ctenodactylidae  | <i>Ctenodactylus</i>   | 4            | 2          |                                     |
|                  | <i>Felovia</i>   | 1            | 1          |                                     |
|                  | <i>Massoutiera</i>   | 3            | 3          |                                     |
| Ctenomyidae      | <b><i>Ctenomys</i>*</b>  | <b>7</b>     | <b>7</b>   | <b>Scratch, Chisel-tooth/U-A</b>    |
| Cuniculidae      | <i>Cuniculus</i>   | 9            | 9          |                                     |
| Dasyproctidae    | <i>Dasyprocta</i>  | 10           | 10         |                                     |
|                  | <i>Myoprocta</i>   | 10           | 9          |                                     |
| Diatomyidae      | <i>Laonastes</i>   | 3            | 3          |                                     |
| Dinomyidae       | <i>Dinomys</i>   | 3            | 3          |                                     |
| Echimyidae       | <i>Callistomys</i>   | 1            | 0          |                                     |
|                  | <b><i>Carterodon</i></b>   | <b>2</b>     | <b>2</b>   | <b>Scratch/A</b>                    |
|                  | <b><i>Clyomys</i></b>  | <b>2</b>     | <b>2</b>   | <b>Scratch/A</b>                    |
|                  | <i>Dactylomys</i>  | 2            | 2          |                                     |
|                  | <i>Diplomys</i>  | 1            | 1          |                                     |
|                  | <i>Echimys</i>   | 6            | 6          |                                     |
|                  | <b><i>Euryzygomatomys</i></b>                                      | <b>2</b>     | <b>2</b>   | <b>Scratch/A</b>                    |
|                  | <i>Geocapromys</i>   | 2            | 2          |                                     |
|                  | <i>Hoplomys</i>  | 2            | 2          |                                     |
|                  | <i>Isothrix</i>  | 2            | 2          |                                     |
|                  | <i>Kannabateomys</i>   | 2            | 2          |                                     |
|                  | <i>Lonchothrix</i>   | 1            | 1          |                                     |
|                  | <i>Makalata</i>  | 2            | 2          |                                     |
|                  | <i>Mesocapromys</i>  | 2            | 2          |                                     |
|                  | <i>Mesomys</i>   | 7            | 7          |                                     |
|                  | <b><i>Myocastor</i></b>  | <b>6</b>     | <b>6</b>   | <b>Scratch/A</b>                    |
|                  | <i>Ollalamys</i>   | 0            | 1          |                                     |
|                  | <i>Phyllomys</i>   | 2            | 2          |                                     |
|                  | <i>Plagiodontia</i>  | 1            | 1          |                                     |
|                  | <i>Proechimys</i>  | 18           | 15         |                                     |
| <i>Trichomys</i> | 4  | 3            |            |                                     |
| <i>Trinomys</i>  | 4  | 4            |            |                                     |

**Table 1** (continued)

| Family              | Genus                | Cranium | Mandible | Mode of digging / foraging activity |
|---------------------|----------------------|---------|----------|-------------------------------------|
| Erethizontidae      | <i>Chaetomys</i>     | 1       | 1        |                                     |
|                     | <i>Coendou</i>       | 4       | 3        |                                     |
|                     | <i>Echinoprocta</i>  | 2       | 2        |                                     |
|                     | <i>Erethizon</i>     | 3       | 3        |                                     |
|                     | <i>Shiggurus</i>     | 2       | 2        |                                     |
| Hystriidae          | <i>Atherurus</i>     | 5       | 5        |                                     |
|                     | <i>Hystrix</i>       | 10      | 8        |                                     |
|                     | <i>Trichys</i>       | 3       | 3        |                                     |
| Octodontidae        | <i>Octodon</i>       | 6       | 4        | Scratch/A                           |
|                     | <i>Octodontomys</i>  | 2       | 2        |                                     |
|                     | <i>Spalacopus*</i>   | 3       | 2        | Chisel-tooth/U-A                    |
|                     | <i>Tympanoctomys</i> | 1       | 1        |                                     |
| Petromuridae        | <i>Petromus</i>      | 4       | 3        |                                     |
| Thryonomyidae       | <i>Thryonomys</i>    | 5       | 5        |                                     |
| Number of specimens |                      | 277     | 246      |                                     |

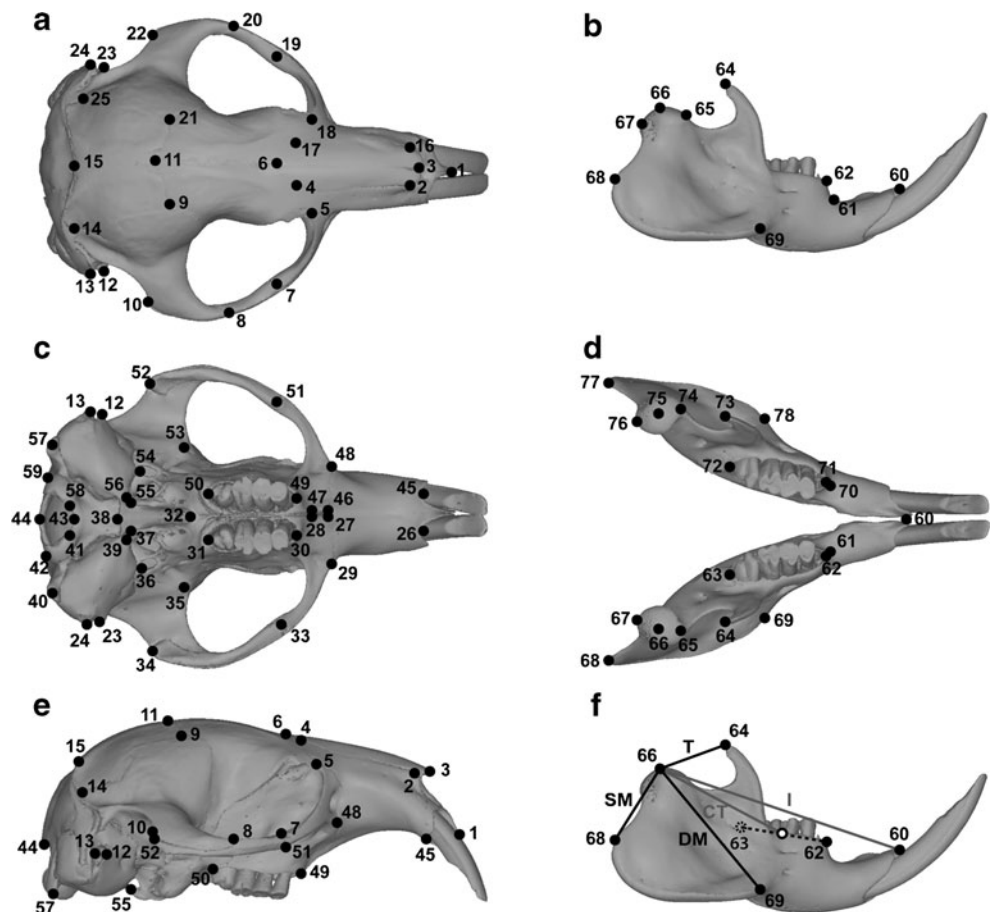
\*:subterranean (sensu Nevo 1979), *A* Aboveground, *U* Underground

“–” means both, “( )” means occasionally. Data derived mainly from Nowak (1999) and Stein (2000)

scanned using a GE phoenix nanotom 180 at energy of 100 keV with a cubic voxel of 22.727  $\mu\text{m}$ , in order to visualize

virtual deformations within the bathyergid dataset, while a scan of *Proechimys* (see Hautier et al. 2011, 2012) was used

**Fig. 1** Landmarks digitized on the cranium (**a**, dorsal view, **c**, ventral view, **e**, lateral view) and the mandible (**b**, lateral view, **d**, occlusal view) of *Heliophobius* (ID13) imaged by using X-ray conventional microtomographic 3D renderings. The landmarks 36 and 54, and 40 and 57 were only used for the analyses of Bathyergidae, because they were not included when the Ctenohystrica database was realized (see Hautier et al. 2012). (**f**) Measure of inlevers (*black lines*), and outlevers (*grey lines*) between landmarks digitized on the mandible of *Heliophobius*. Outlevers of cheek teeth correspond to the distance between the landmark 66 (condyle), and an approximation of the middle of the tooth row using the middle point of the distance between the landmarks 62 and 63. *T* Temporalis, *SM* Superficial Masseter, *DM* Deep Masseter, *I* Incisor, *CT* Cheek Teeth



for *Ctenohystrica*. Because the mandible of rodents is constituted of a unique dentary bone of relatively simple shape, most of the landmarks taken on the dentary were of type 2 (e.g., maxima of curvature – Fig. 1; Bookstein 1991). All configurations (sets of landmarks) were superimposed using the Procrustes method of generalized least squares superimposition (GLS scaled, translated, and rotated configurations so that the intralandmark distances were minimized) following the method used by Rohlf (1999) and Bookstein (1991). Subsequently, mandibular and cranial forms of each specimen were represented by centroid size  $S$ , and by multidimensional shape vector  $v$  in linearized Procrustes shape space. Shape variability of the cranium and mandible was analyzed by principal components analyses (PCA) of shape (Dryden and Mardia 1998). Analysis and visualization of patterns of shape variation were performed with the interactive software package MORPHOTOOLS (Specht et al. 2007; Lebrun et al. 2010). A public version is currently being developed (contact renaud.lebrun@univ-montp2.fr for further information). A multivariate regression of the first 35 PCs (95 % of the variance) on size, estimated by the logarithm of the centroid size, allowed us to take into account the potentially confounding effects of size allometry on shape. Additionally, univariate regressions were realized using the first two principal components and the logarithm of the centroid size of each specimen. We conducted separate regression analyses for both bathyergids and non-bathyergid *Ctenohystrica* in order to search for size trends within these groups; differences in slope were tested using ANCOVA. Multivariate regressions and ANCOVAs were performed with Past 2.06 (Hammer et al. 2001). To account for the influence of phylogeny on our result, we regressed phylogenetically independent contrasts (PICs; Felsenstein 1985) to test whether the size (log centroid size) and shape components (i.e., PC1 to 3) show evidence of correlated evolution. The phylogeny used in the PIC analysis derived from Fabre et al. (2012). PIC analysis was implemented using the PDAP:PDTREE module, version 1.14 (Midford et al. 2008) of Mesquite, version 2.71 (Maddison and Maddison 2009).

Furthermore, covariation patterns between the crania and the mandibles were studied using 2-blocks partial least square analysis, as described by Bookstein et al. (2003), only adapted to allow for the use of 3D landmarks. For the  $N=237$  specimens for which both cranial ( $k=59$ ) and mandibular ( $l=19$ ) landmarks had been digitized, cranial and mandibular landmark configurations were aligned separately using GLS, yielding a cranial matrix of  $N*3k$  shape coordinates and a mandibular matrix of  $N*3l$  shape coordinates. The PLS analysis computed a series of pairs of unit vectors, the singular cranial and mandibular warps ( $U_c$  and  $U_m$ ), each being

of length  $3k$  and  $3l$ , respectively. These pairs of singular warps maximize the covariance between the two sets of shape coordinates. Cranial and mandibular projection scores of the specimens on the singular warps were subsequently computed.

### Biomechanical Analyses

They allow the quantification of the mechanical advantage of each adductor muscle (i.e., temporalis, superficial masseter, and deep masseter) by measuring the force transmitted from the muscle to the bite point (i.e., incisor and cheek teeth). It can be estimated as the ratio of the inlever (distance from the condyle or fulcrum to the point of muscle attachment) and the outlever (distance from the condyle to the bite point; Hiiemae 1971). Three adductor muscles have been considered for inlevers: the temporalis that attaches on the coronoid process (landmarks 64 and 73), the superficialis masseter that attaches along the angular process (landmarks 68 and 77), and the deep masseter that attaches on the masseteric crest (landmarks 69 and 78) (Cox and Jeffery 2011; Baverstock et al. 2013; Renaud et al. 2015). Rodents are characterized by two dental regions that were alternatively considered for outlevers: the incisors, used for gnawing isolated from the cheek teeth, used for chewing. Unfortunately, the bite point at the tip of the incisor could not be used, because of large intraspecific variation due to wear (see Casanovas-Vilar and Van Dam 2013). Thus, no interpretation could be made on the incisor procumbency. Consequently, the landmark (60), located at the posterior part of the alveolar margin of incisor, was used for the outlever. Six estimates of biomechanical advantage (inlever/outlever) were thus considered for a given mandible: T/I (temporalis/incisor), SM/I (superficial masseter/incisor), DM/I (deep masseter/incisor), T/CT (temporalis/cheek teeth), SM/CT (superficial masseter/cheek teeth), and DM/CT (deep masseter/cheek teeth). Differences between bathyergids in the inlever/outlever ratios were tested using post-hoc multiple mean comparison tests: the Fischer's Least Significant Difference test (LSD) and the Tukey's Honestly Significant Difference test (HSD), the latter being less sensitive but more conservative than the former (Online resource 3).

## Results

### Comparison of Skull Shape Among *Ctenohystrica*

According to both PCAs performed on the whole dataset, bathyergids can be clearly discriminated from other non-fossorial or fossorial ctenohystricans (Fig. 2). The crania of fossorial rodents plot on the positive side of PC1, with bathyergid rodents on the most positive values. Among non-bathyergid subterranean *Ctenohystrica*, *Spalacopus*



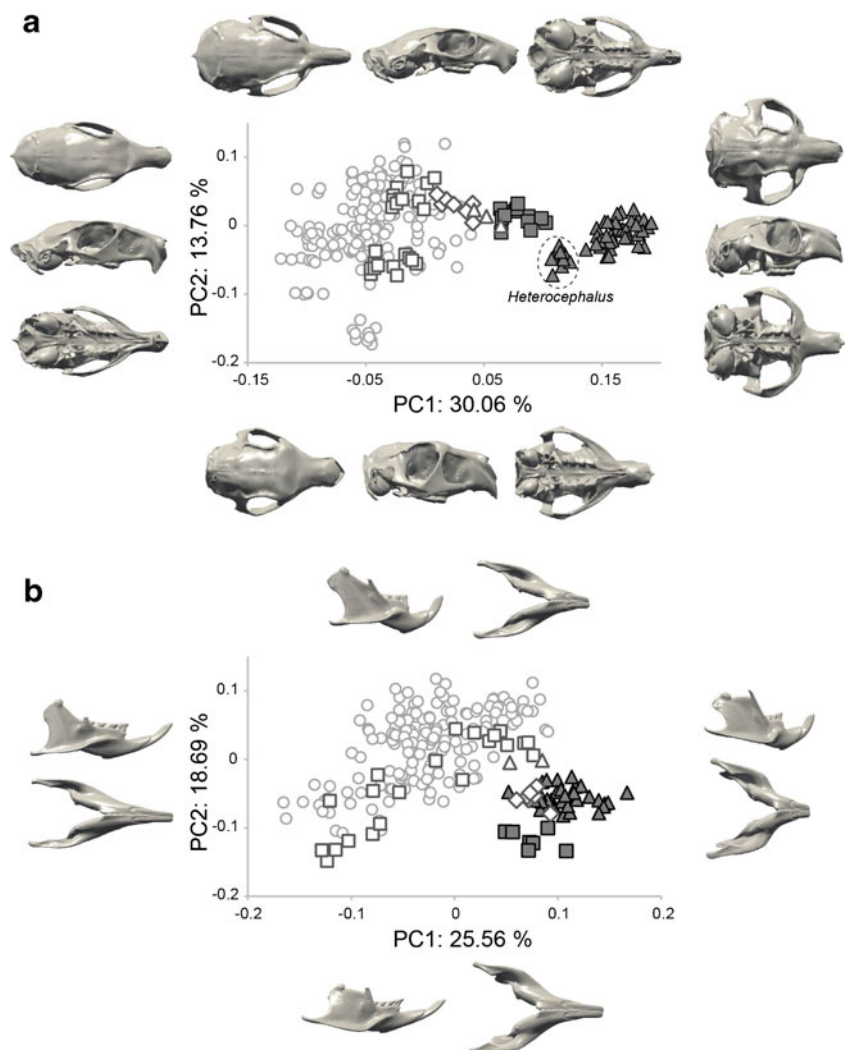
and *Ctenomys* plot in the vicinity of bathyergids, and more generally, chisel-tooth diggers show the most distinctive shape. Fossorial rodents, especially subterranean forms, are characterized by a short and wide cranium having a wide snout, but large zygomatic arches, small incisive foramina (landmarks 27–28 and 46–47), a narrow frontal area, a dorsal part of squamosal bone more developed, and an elevated and laterally extended occipital region. This shape clearly departs from the cranial morphology of non-fossorial rodents, which have a gracile and elongated cranium with an extended fronto-parietal area, a low cranial height, and an occipital part protruding backward (Fig. 2a). PC2 discriminates species with enlarged snout and a narrow braincase from species with a short and gracile snout and a large braincase (Fig. 2a).

For mandibles, *Ctenomys* plots with bathyergids on the positive values of PC1 (Fig. 2b). All these species are characterized by a short mandible, a strong hystricognathous condition, with enlarged coronoid and

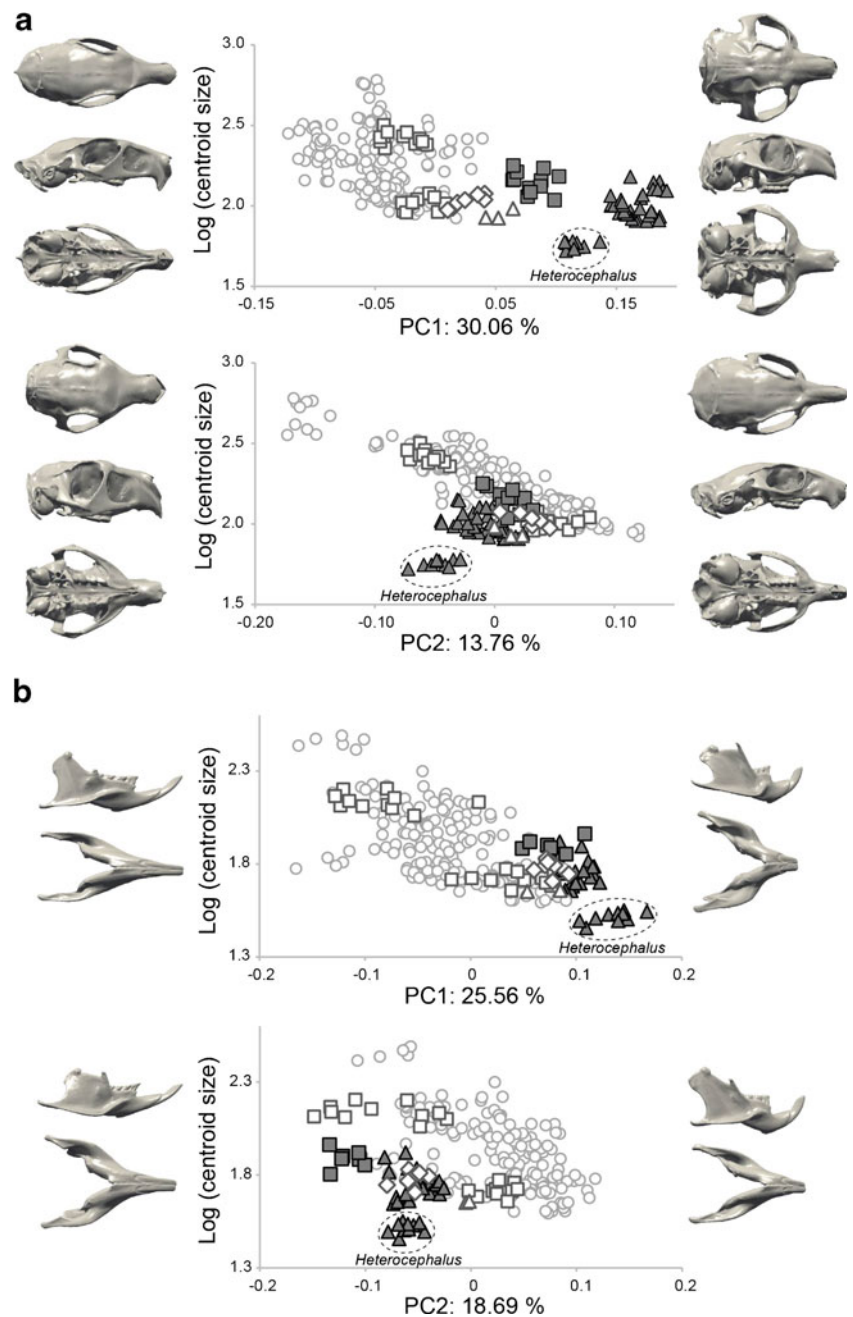
reduced condylar processes. These morphological trends are here again more pronounced in the chisel-tooth diggers. On the negative values of PC2, some taxa (e.g., *Bathyergus*) show an elongated mandible with elongated angular processes and reduced condyle, compared to other scratch diggers, which have more developed condylar and reduced angular processes.

A multivariate regression of the shape component on size, estimated by the logarithm of centroid size, was highly significant (mandible:  $F=28.16$ ,  $p<0.001$ ,  $df=213$ ; cranium:  $F=86.61$ ,  $p<0.001$ ,  $df=245$ ). As such, allometry therefore explains a substantial part of shape variation, and plays an important role in determining the pattern of morphological differentiation of the skull. The Phylogenetic Independent Contrasts (PIC) analysis is congruent with these results, and indicates that after phylogenetic correction, allometry plays a crucial role in determining the pattern of morphological differentiation of the cranium (PC1,  $r^2=0.08$ ;  $p=0.03$ ; PC2,  $r^2=0.52$ ;  $p=1.4\times 10^{-10}$ ) and the

**Fig. 2** Principal component analyses realized on (a) crania and (b) mandibles of Ctenohystrica and associated virtual deformations on the extreme sides of each axis. Symbols used: (○) non-fossorial; (□) scratch digger; (△) chisel-tooth digger (*Spalacopus*); (◇) both modes of digging (*Ctenomys*); (■) scratch digging Bathyergidae (*Bathyergus*); (▲) chisel-tooth digging Bathyergidae



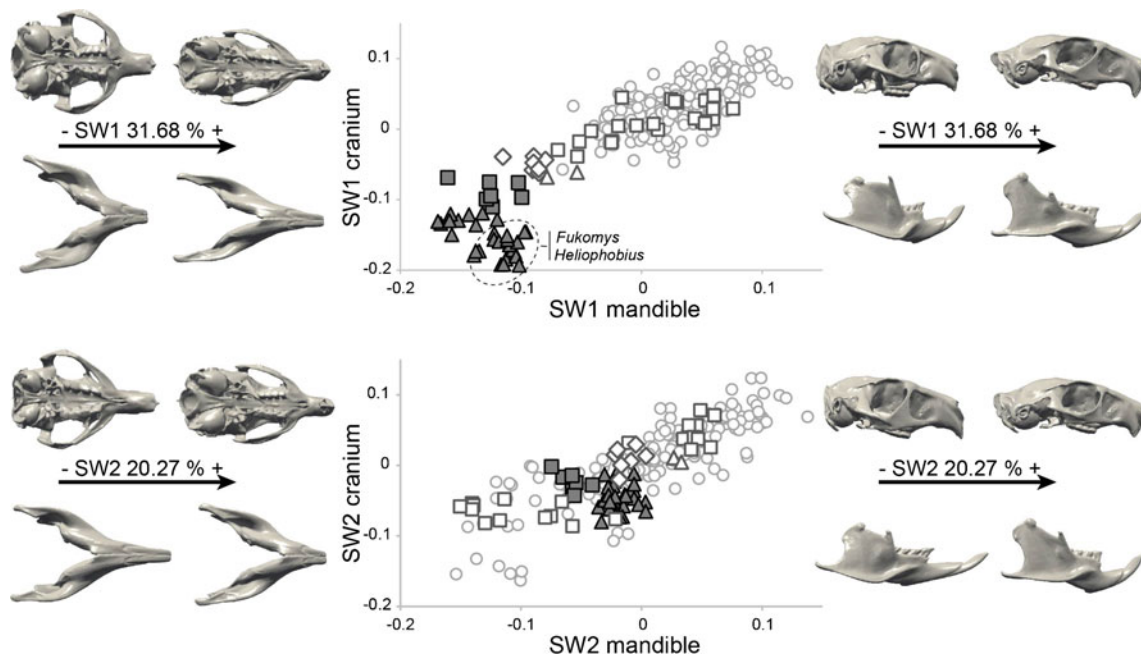
**Fig. 3** Regression of the first and second principal component on the centroid size of (a) crania and (b) mandibles of Ctenohystrica and associated virtual deformations. Symbols used: (○) non-fossorial; (□) scratch digger; (△) chisel-tooth digger (*Spalacopus*); (◇) both modes of digging (*Ctenomys*); (■) scratch digging Bathyergidae (*Bathyergus*); (▲) chisel-tooth digging Bathyergidae



mandible (PC1,  $r^2=0.31$ ;  $p=4.4 \times 10^{-6}$ ; PC2,  $r^2=0.18$ ;  $p=7.5 \times 10^{-4}$ ). The regression of the first mandibular and cranial PCs on centroid size (Fig. 3a and b) shows that most fossorial Ctenohystrica are of very small size (e.g., *Heterocephalus*). However, some of the less specialized fossorial species (*Lagostomus*, *Myocastor*) have larger skulls. When PC2 are regressed against the centroid size, bathyergid rodents (especially chisel-tooth diggers like *Heterocephalus*) and other subterranean genera (*Spalacopus* and *Ctenomys*) clearly depart from the regression lines, which corresponds to the main trends observed

in Ctenohystrica (see Hautier et al. 2012). ANCOVAs indicate significant differences in the slope between the crania of bathyergid and non-bathyergid Ctenohystrica (PC1,  $F=6.85$ ,  $p=0.009$ ; PC2,  $F=6113.7$ ,  $p=1.8 \times 10^{-22}$ ). In contrast, there are no significant differences in the slope when considering their mandibles (PC1,  $F=2.144$ ,  $p=0.14$ ; PC2,  $F=18.69$ ,  $p=0.67$ ).

Covariation evidently exists between cranium and mandible in Ctenohystrica, and underscores the differences observed between the skull shape of non-fossorial species and fossorial rodents, bathyergids in



**Fig. 4** First two singular warp (SW) mandibular and cranial scores and associated mandibular and cranial covariation patterns for the subset of specimens for which the crania and the mandibles had been digitized.

**Symbols used:** (○) non-fossorial; (□) scratch digger; (△) chisel-tooth digger (*Spalacopus*); (◇) both modes of digging (*Ctenomys*); (■) scratch digging Bathyergidae (*Bathyergus*); (▲) chisel-tooth digging Bathyergidae

particular ( $r^2=0.78$ ,  $p=4.3 \times 10^{-80}$ ; Fig. 4). Covariation patterns are different between chisel-tooth diggers and scratch diggers, in both bathyergids and other fossorial Ctenohystrica. Here again, a few bathyergid genera (*Heliophobius*, and both *Fukomys* species) set apart from the general regression on SW1, and an ANCOVA shows significant differences in the slope between the skull of bathyergids and non-bathyergid Ctenohystrica on SW1 ( $F=45.27$ ,  $p=1.29 \times 10^{-10}$ ). A relation is also present on SW2 ( $r^2=0.66$ ;  $p=5 \times 10^{-59}$ ), but it is weaker and poorly informative regarding the comparison of fossorial and non-fossorial ctenohystricans.

#### Comparison of the Lever Arm Lengths Between Fossorial and Non-fossorial Ctenohystrica

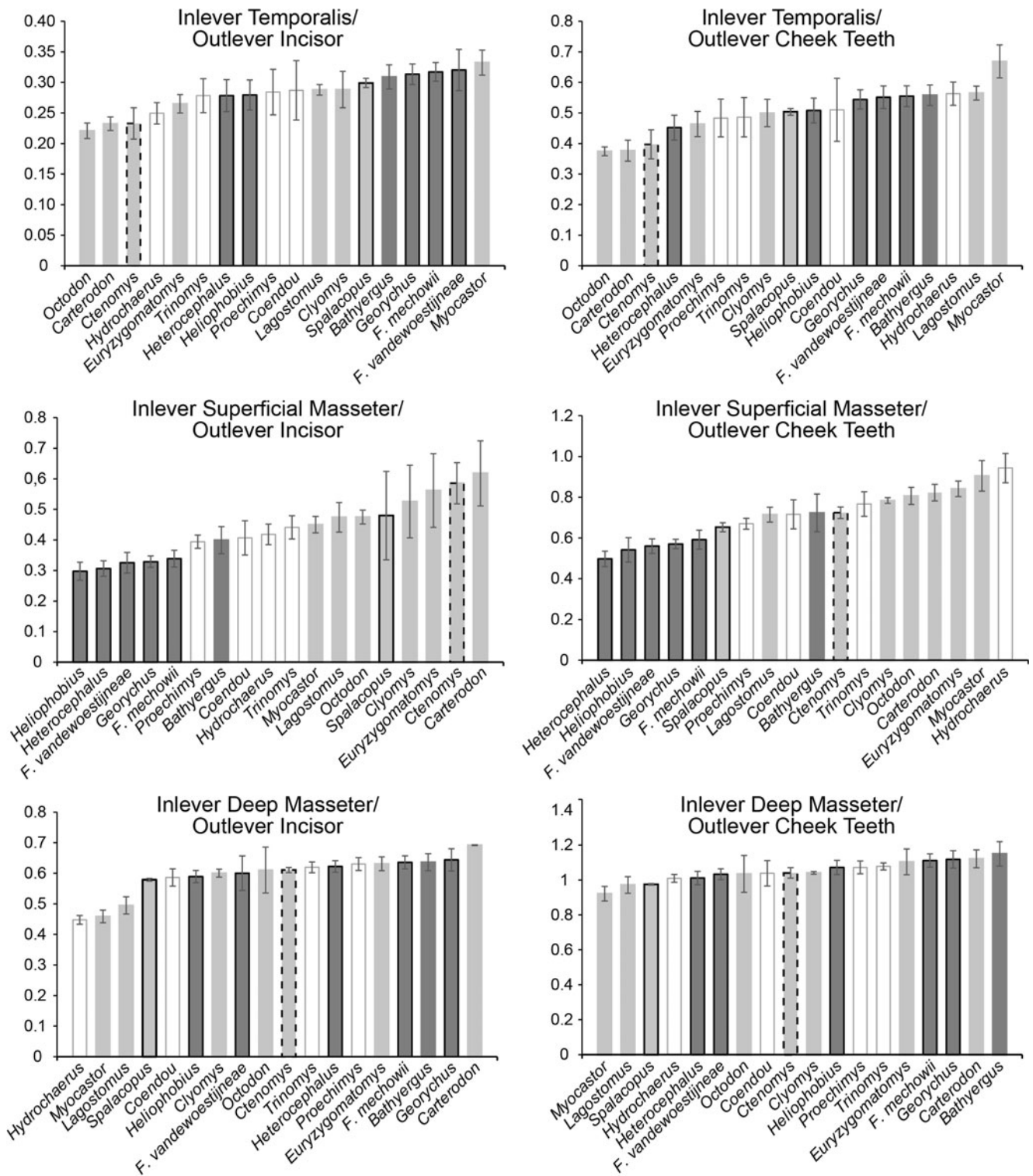
As expected, the results are slightly different whether we considered the incisors or the cheek teeth as the outlever. Except for *Heterocephalus* and *Heliophobius*, chisel-tooth diggers tend to show high values of lever arm ratios for the temporalis muscle (Fig. 5; incisor and cheek teeth). In contrast, chisel-tooth digging bathyergids are characterized by low values of ratios for the superficial masseter (Fig. 5; incisor and cheek teeth). *Bathyergus* always shows higher values than other bathyergids, and so do *Spalacopus* and *Ctenomys*. Whatever the outlever considered for the superficial masseter, scratch diggers are always characterized by high values of ratios, while non-fossorial rodents mostly

sit in the middle of the graph. The results for the deep masseter are more difficult to interpret because of the really low variation range of the arm ratios for this muscle (Fig. 5), and no clear pattern of differentiation emerges in relation to the different digging modes. When focusing on mole-rats, statistical tests reveal that African mole-rats present significant differences regarding most inlever/outlever ratios (Online resource 3). In contrast, *F. mechowii* and *Georychus* do not show any significant differences. Interestingly, both taxa show highly significant differences with *F. vandewoestijneae* only for the deep masseter, and with *Bathyergus* only for the superficial masseter. *Heliophobius* and *Heterocephalus* only display slight significant differences when the outlever arm of the molars is considered, with higher ratios for *Heliophobius*. However, both genera show strong differences with most of other bathyergids.

#### Comparison of Skull Shape Among the Bathyergidae

The principal component analyses clearly demonstrate that chisel-tooth diggers and scratch diggers (i.e., *Bathyergus*) show distinct crania and mandibles (Fig. 6a). On PC1, *Heterocephalus* sets apart from other mole-rats on the negative values in having a larger braincase, a shorter snout, and reduced jugal bones, while the maxillaries are enlarged at the level of the zygomatic arches. The second component clearly separates scratch diggers from chisel-





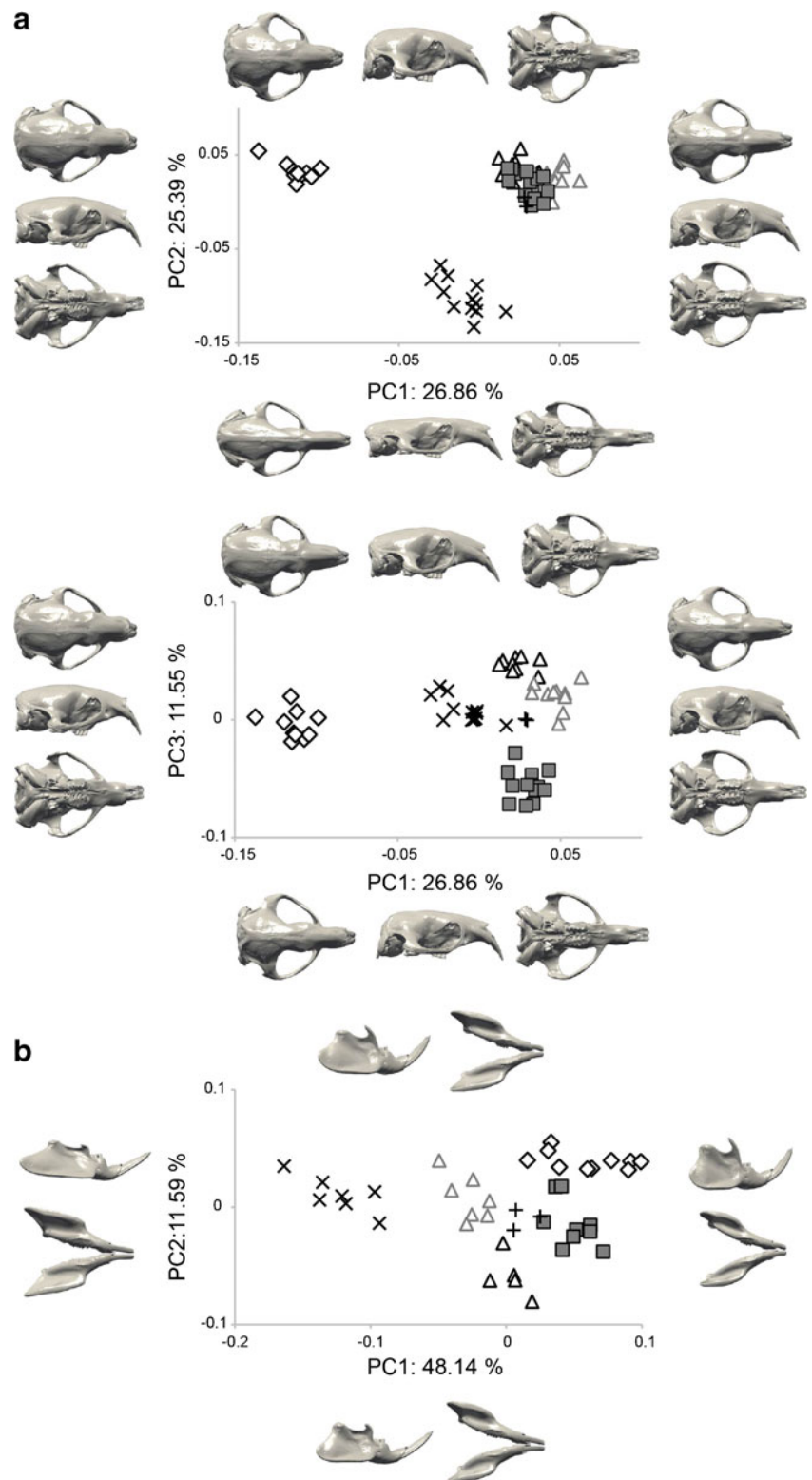
**Fig. 5** Histograms representing the mean inlever/outlever ratio with corresponding standard deviations for fossorial taxa and a few other Ctenohystrica. Symbols used: (□) non-fossorial; (■) scratch digger;

(■) chisel tooth digger; (■) both modes of digging; (■) scratch digging Batherygidae; (■) chisel tooth digging Batherygidae

tooth diggers. *Bathyergus* shows a more slender cranium with an elongated snout, and a lower cranial height. On

the third principal component, *Heliophobius* plots on the negative values and differs from other mole-rats in having

**Fig. 6** Principal component analyses realized on crania and mandibles of Bathyergidae, and associated virtual deformations on the extreme sides of each axis. Symbols used: (x) *Bathyergus*; ( $\Delta$ ) *Fukomys vandewoestijneae*; ( $\triangle$ ) *Fukomys mechowii*; (+) *Georychus*; ( $\blacksquare$ ) *Heliophobius*; ( $\diamond$ ) *Heterocephalus*



a smaller braincase, a reduced snout, but wider zygomatic arches and a shorter tooth row.

Mandible shapes of scratch and chisel-tooth diggers are clearly separated on PC1 (Fig. 6b). *Bathyergus*

shows an elongated mandible with enlarged angular and reduced coronoid processes, while other mole-rats have a shorter mandible with more developed coronoid and reduced angular processes.

## Discussion

### Convergent Morphological Trends in Fossorial Ctenohystrica

Clear morphological trends have been observed between fossorial and non-fossorial Ctenohystrica. Both the mandible and cranium showed a strong imprint of the different degrees of fossoriality observed among Ctenohystrica. However, the mandible, with its simple morphology, appears to be more prone to convergent evolution because non-related groups with similar ecology sat closer to each other in the mandibular morphospace (e.g., chisel-tooth diggers). Subterranean rodents have a short and wide skull, especially chisel-tooth diggers, and this morphology is frequently associated with change in body size. A small size favors life underground because the costs of burrowing generally increase with body size (Vleck 1981). We showed that the more fossorial the species are (e.g., subterranean forms), the more these morphological characters are pronounced. Conversely, less specialized fossorial species (e.g., *Lagostomus*, *Myocastor*) are usually bigger precluding an exclusive life underground. The skull is generally less constrained in scratch diggers, which have larger skull and body size. In subterranean species such as *Ctenomys*, *Spalacopus*, and bathyergids, the convergent acquisition of enlarged zygomatic arches and squamosal bones, where the adductor muscles attach, is clearly linked to strong fossorial adaptations. Similarities are also observed at the level of the mandibular ramus of these species, in which the articular condyle is generally less protruding than the coronoid and/or angular processes, allowing large insertions for adductor muscles. This kind of configuration is frequently present in other fossorial rodents, especially in some Spalacidae (Stein 2000), and already occurred in one of the first known fossorial Ctenohystrica, *Tsaganomys altaicus* from the Oligocene of Mongolia (Bryant and McKenna 1995). But in spite of appearances, the rough skull similarities observed among the investigated fossorial rodents should not conceal the fact that different associations of morphological features exist for a given digging mode (e.g., *Ctenomys*, *Spalacopus*, and African mole-rats).

### The Special Case of African-Mole Rats

Evolutionary trends associated with cranial shape in Bathyergidae do not match those observed in most fossorial and non-fossorial Ctenohystrica. Even if a size reduction appears to be a prerequisite for living underground, it did not totally constrain the shaping of the skull in bathyergids as it did in most Ctenohystrica, except for subterranean rodents (*Spalacopus* and *Ctenomys*). African mole-rats clearly depart from most Ctenohystrica in showing highly derived cranial and mandibular morphologies. This is notably shown by

chisel-tooth diggers (e.g., *Heterocephalus*, Fig. 3), which have reduced and rounded angular processes, as well as pronounced pterygoid processes associated with a very backward origin of the alveolus of the upper incisors, which are much more procumbent. The relative morphological isolation of African mole-rats explains why their phylogenetical relationships have been the subject of a longstanding debate over the past century (Tullberg 1899; Ellerman 1940; Landry 1957; de Graaff 1979). Bathyergid skulls show more resemblances with other subterranean rodents, such as *Spalacopus* and *Ctenomys*, than with their sister taxa *Thryonomys* and *Petromus*. Their unique morphology highlights a deep ecological component, which is due to strict life underground associated with a prolonged use of incisor for digging. As a result, the skull of bathyergids underwent strong directional selection related to their singular ecology, which allows them to depart from the morphological pathways that ruled the evolution of most Ctenohystrica and were more tightly coupled to allometric variations (Hautier et al. 2012). These morphological modifications might have partly been favored by the miniaturization of the eyes, as it is usually the case in strict subterranean species (e.g., some mole-rats), or to changes in their positioning on the skull roof in subterranean species having also aboveground activity (e.g., *Ctenomys* and *Spalacopus*; Stein 2000). Because the position of the eye was suggested as one of the limiting factor determining the morphological evolution of the masticatory apparatus in Ctenohystrica (Hautier et al. 2012), a relaxed selection on this character might have released some room for the development of adductor muscles (Cox and Faulkes 2014).

### Masticatory Mechanics of the Main Digging Groups

All chisel-tooth digging bathyergids except *Bathyergus* are characterized by shorter mandibles with elongated coronoid, rounded angular processes, and masseteric crests placed anteriorly, and this will have a major impact on the biomechanics of the jaw. The mandibular morphology in *Bathyergus* rather resembles the association of features found in other digging hystricognaths that show elongated angular and reduced coronoid processes. This is probably related to its scratch digging behavior associated with life in soft sandy soils (Bennett et al. 2009), relaxing constraints on the skull as in most non-bathyergid fossorial species compared to chisel-tooth diggers that generally dig in harder soils. Mole-rats have been shown to be forceful biters that can exert bite forces higher than expected for a mammal of comparable body size, especially at the level of molars (Van Daele et al. 2009). This is partly favored by the “most freely moveable jaws of any rodent” (Landry 1957) allowed by unfused symphysis between the two halves of the mandible, and enlarged and flat glenoid fossa (de Graaff 1979). Only a few descriptions of the masticatory musculature of bathyergids can be found in the

literature, most of them just briefly mentioned the specific nature of their myology (Tullberg 1899; Morlok 1983; Van Daele et al. 2009; Cox and Faulkes 2014). Nonetheless, interesting comparisons of masticatory muscles have recently been drawn between subterranean, less fossorial, and non-fossorial ctenohystricans (*Ctenomys*, *Octodon*, and *Chinchilla*, respectively; Becerra et al. 2014). The mechanics of the masticatory apparatus are complex because each muscle acts with a different lever advantage; some muscles are specialized for large speed and some for large force (Kardong 2006). The mechanics of lever imply that output force and output speed work in opposite directions: short output lever arms relative to input lever arms favor force while long output lever arms favor speed (Kardong 2006).

In *Heterocephalus*, the temporalis represents the largest jaw-closing muscle (Cox and Faulkes 2014), and this muscle seems to dominate the masticatory musculature in all other bathyergids (Morlok 1983; Tullberg 1899; Van Daele et al. 2009). Druzinsky (2010) suggested that the enlargement of the temporalis may result from the widening of the skull commonly observed in fossorial species. A large temporalis is likely to produce substantial forces at the teeth (Hiimae 1971). Mechanical advantages of the input lever arms of the temporalis were rather variable across the different digging types (Fig. 5) and no clear pattern emerged to distinguish chisel-tooth diggers from scratch diggers. Yet, most of the bathyergids and fossorial caviomorphs showed high value of the ratio when the incisors were considered as the outlever. Such a result can be partly explained by a combination of a shortening of the mandible, especially the diastema, and an elongation of the coronoid process, which produces the reduction of the output lever arm of the jaw-closing muscles and, in turn, an increase of the mechanical advantage for the temporalis (i.e., inlever T larger relative to outlever I). By moving the output closer to the fulcrum, the temporalis muscle can create more output force at the level of the incisors. However, the fact that we measured the outlever at the anterodorsal edge of the incisor alveolus instead of its tip surely had an impact on the results as the output is displaced closer to the fulcrum.

In the naked mole-rat, the superficial and deep masseters together represent almost 50 % of the masticatory musculature and are likely to deliver high bite force especially at the level of the cheek teeth (Cox and Faulkes 2014). The superficial masseter is reduced laterally and wraps around the ventral margin of the mandible; it shows a really large pars reflexa. Its strong horizontal component implies that it probably acts as the main protractor of the mandible (Cox and Faulkes 2014). Due the high position of the landmarks defining the angular process (Landmarks 68 and 77, Fig. 1b and d), which is close to the condyle (i.e., inlever SM smaller relative to outlever I), the tooth digging bathyergids always showed the lowest values of ratios. Lever arms of the superficial masseter

are less favorable to force output in the mandible of the bathyergids but more favorable to speed. Compared to the small input lever arm defined between the condyle and the angular process, the relatively greater output lever arm increases the speed at the expense of the output force. By moving the input force closer to the fulcrum, the superficial muscle can generate higher output speed. In contrast to the clear-cut results obtained for the superficial masseter, the lever arm ratios of the deep masseter were not significantly different between rodents of different modes of digging (Fig. 5).

The temporalis and the superficial masseter show opposite mechanical advantages and make different contribution to force and speed output. The temporalis ratio favors force, which is important during chisel-tooth digging, while the superficial masseter has a mechanical advantage favoring velocity.

### Cranial Evolution in African Mole-Rats

In addition to the main morphological characters related to fossorial life that best characterize African mole-rats, significant divergences can be observed among them. The skull of *Bathyergus* strongly differs from other mole-rats in being more elongated. The enlarged snout associated with a gracile skull can be related to the fact that *Bathyergus* mainly uses its foreclaws for digging in soft sandy soils, contrary to most mole-rats living in hard soils that use their incisors and have strongest masticatory muscles. However, *Bathyergus* still shows a high masticatory effectiveness, notably at the level of the cheek teeth. This is in line with the fact *Bathyergus* has high-crowned cheek teeth. Despite the fact that they show obvious differences in cranial shape, *Bathyergus* and its sister genus *Georychus* display close biomechanical affinities, especially regarding the deep masseter and the temporalis. *Bathyergus* also shows the same affinities with *F. mechowii*, the largest *Fukomys* species, which is able to construct very large burrow systems (Šumbera et al. 2012). More generally, the mosaic of cranial and mandibular characters of *Bathyergus* can be regarded as intermediate between the Ctenohystrica and Bathyergidae. However, this does not imply that *Bathyergus* can be considered as a putative ancestral bathyergid skull shape. The oldest bathyergid skulls known in the fossil record (Lavocat 1973; Mein and Pickford 2008) are more similar to those of *Heterocephalus* and *Heliophobius*, which belong to ancient lineages (Faulkes et al. 2004; Van Daele et al. 2007; Patterson and Upham 2014). Even if *Bathyergus* presents primitive characters, such as the connection between the pterygoid fossa and the orbit as in most Ctenohystrica (Landry 1957), this connection is already present in ancestral forms with short and wide cranium, such as *Renefossor*<sup>†</sup> and *Proheliophobius*<sup>†</sup> (Lavocat 1973). Moreover, in most Miocene taxa (e.g., *Bathyergoides*<sup>†</sup>, *Renefossor*<sup>†</sup>, *Proheliophobius*<sup>†</sup>, *Efeldomys*<sup>†</sup>), the infraorbital



foramina are not reduced like in extant species. In *Efeldomys*<sup>†</sup>, the incisive foramina also are longer than in all extant bathyergids, including *Bathyergus* (Lavocat 1973; Mein and Pickford 2008). Consequently, the skull shape of *Bathyergus* by no means should be seen as ancestral. It presents an association of derived features, some of which evolved convergently in *Ctenomys* and *Spalacopus*. *Bathyergus* probably derived from a basal stock of mole-rats that was already strongly adapted to fossorial life, and for which putative active chisel-tooth diggers have been recognized based on the presence of procumbent incisors (e.g., *Renefossor*<sup>†</sup> and *Proheliophobius*<sup>†</sup>; Lavocat 1973).

*Heterocephalus* belongs to an early diverging lineage of Ctenohystrica (Patterson and Upham 2014), and is a highly derived genus, in terms of morphological and biological characters. However, it is difficult to explain why *Heterocephalus* has an enlarged braincase. This braincase enlargement might simply be due to allometric constraints because most small ctenohystricans tend to show relatively larger braincase (Hautier et al. 2012). Whatever the cause, this character is probably not related to the social behavior of *Heterocephalus*, because other social mole-rats, such as *Fukomys*, do not have an enlarged skull. Contrary to *Heterocephalus*, *Heliophobius* only slightly differs from other mole-rats, and has a small braincase and a wider skull, which permit optimizing the area of attachment and the strength of adductor muscles. The digging activity of this solitary species, which is capable of removing hard soil and constructing extensive burrows (Barčiová et al. 2009), surely had a major impact on the evolution of its cranial morphology, including their unique continuous dental replacement. Even if *Heliophobius* presents a mechanical advantage for its adductor muscles lower than most mole-rats, it comparatively and relatively shows higher output forces at the level of the molars, meaning that a very strong resulting pressure can be exerted at this level. High output force on the molars can also contribute to the high wear affecting the dentition due to high abrasion during eating and also due to high attrition during chisel-tooth digging in hard soils. We showed that the dental length in *Heliophobius* is also reduced compared to most mole-rats, so a continuous dental replacement constitutes a suitable means to maintain a functional dentition. In *Heterocephalus*, the reduced output force on the molars is compatible with the observed reduction of their dentition, which does not need a superimposed adaptation to fight wear, not even very high-crowned teeth.

We have demonstrated that the skull of mole-rats, and more generally, of subterranean rodents, is strongly and historically influenced by the nature of their fossorial activity, which in turn minimizes the size effect. It cannot be assumed that the evolution of their skulls was shaped by the intensity of digging activity, inasmuch as records of daily energetic expenditure in social and solitary mole-rats are quite similar (Zelová et al. 2011). It cannot be proposed either that their skull morphology

is strongly influenced by their social or solitary behavior, because the solitary *Georychus* and the social *Fukomys* species display close cranial morphologies, while they probably diverged about 10–15 Mya (Patterson and Upham 2014). Nonetheless, social polymorphism and sexual dimorphism (e.g., *Bathyergus suillus*, Hart et al. 2007; *Fukomys mechowii*, Chimimba et al. 2010) could explain some parts of the intra-specific variation, alongside the variations observed between populations of African mole-rat species living in different environments. Previous studies showed that skull shape of Ctenohystrica is indeed plastic and highly sensitive to the environments, and that the soil hardness and compactness can have a significant impact on its evolution (Vassallo 1998; Mora et al. 2003; Barčiová et al. 2009). Consequently, the different digging strategies in African mole-rats will be more accurately understood when placed in the climatic and environmental context of their origins, because their local adaptations and their current habits only reflect shaded parts of their evolutionary history.

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