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A Turn-on Fluorescent Chemosensor for Zn²⁺ Based on Quinoline in Aqueous Media

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Abstract A simple "off-on fluorescence type" chemosensor 1 3-((2-(dimethylamino)ethyl)amino)-N-(quinolin-8yl)propanamide has been synthesized for Zn²⁺. The receptor 1 comprises the quinoline moiety as fluorophore and the N,N'dimethylethane-1,2-diamine as a binding site. 1 showed a remarkable fluorescence enhancement in the presence of Zn²⁺ in aqueous solution. Importantly, the chemosensor 1 could be used to detect and quantify Zn²⁺ in water samples. In particular, this chemosensor could clearly distinguish Zn²⁺ from Cd²⁺. The binding properties of 1 with Zn²⁺ ions were investigated by UV-vis, fluorescence, electrospray ionization mass spectroscopy and ¹H NMR titration.

Keywords Fluorescence enhancement \cdot Determination of Zn ion \cdot Chemosensor \cdot Quinoline

Introduction

Zinc is the second most abundant transition metal ion in human body [1-7]. Zinc has attracted a great deal of attention [8-13], because it plays very important role in variety of

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² School of Ecology & Environmental Systems, Kyungpook National University, Sangju 37224, South Korea physiological and pathological processes such as apoptosis, catalytic function of protein, enzyme regulation and so on [14-20]. Especially, labile Zn^{2+} has been implicated in signaling processes in the brain, immunological function and gene transcription [21-24]. Its deficiency generates unbalanced metabolism, which in turn can induce retarded growth in children, brain disorders and high blood cholesterol, and also be implicated in various neurodegenerative disorders such as Alzheimer's disease, epilepsy, ischemic stroke, and infantile diarrhea. Excess zinc may also cause serious neurological disorders such as Alzheimer's and Parkinson's diseases [25-28]. Thus, a technique to detect and visualize free zinc ions would be highly demanded [29-34].

To date, many chemosensors have been reported to detect trace amount of Zn^{2+} . Many of them, however, have disadvantages such as insufficient sensitivity or selectivity, and inhibition problems from other transition metal ions, especially Cd^{2+} , which is in the same group of the periodic table and shows similar properties to Zn^{2+} [35–38]. Thus, low cost and easily prepared Zn^{2+} selective fluorescence chemosensors are needed for convenience [39–45].

In view of this necessity and as part of our effort devoted to zinc ion recognition, we have considered the combination of a quinoline moiety known as having desirable photo-physical properties as a fluorophore group and a N, N'-dimethyl ethylene amine as a binding site (Scheme 1) [46–48]. Especially, we expected that the N, N'-dimethyl ethylene amine group, being hydrophilic in nature, would increase water-solubility of the chemosensor.

Herein, we report a new chemosensor 1 for Zn^{2+} , composed of the quinoline and N, N'-dimethyl ethylene amine moieties. We have observed its prominent fluorescence enhancement in the presence of zinc ion, while there was no enhancement in the presence of other metal ions. In particular, it was able to distinguish Zn^{2+} from Cd^{2+} .

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Experiments

Reagents and Instrument

All the solvents and reagents (analytical grade and spectroscopic grade) were obtained commercially and used as received. ¹H and ¹³C NMR spectra were recorded on a Varian 400 MHz and 100 MHz spectrometer, respectively and chemical shifts were reported in ppm, relative to tetramethylsilane Si(CH₃)₄. Absorption spectra were recorded at 25 °C using a Perkin Elmer model Lambda 25 UV/Vis spectrometer. The emission spectra were recorded on a Perkin-Elmer LS45 fluorescence spectrometer. Electrospray ionization mass spectra (ESI-MS) were collected on a Thermo Finnigan (San Jose, CA, USA) LCQTM Advantage MAX quadrupole ion trap instrument. Elemental analysis for carbon, nitrogen, and hydrogen was carried out by using a Flash EA 1112 elemental analyzer (thermo) in Organic Chemistry Research Center of Sogang University, Korea.

Synthesis of Receptor 1

3-Chloro-N-(quinolin-8-yl)propanamide (1.17 g, 5 mmol) and potassium iodide (8 mmol, 1.33 g) were dissolved in MeCN (20 mL) and stirred for 1 h. Then, N,N'-dimethylethane-1,2diamine (0.44 mL, 5 mmol) and sodium hydroxide (0.24 g, 6 mmol) were added in the resulting solution. It was stirred for 12 h at room temperature. The solvent was removed under reduced pressure to obtain bright yellow oil, which was dissolved in methylene chloride and washed twice with water. Then, the solution was purified by silica gel column chromatography (10:1 v/v CH₂Cl₂-CH₃OH) (Scheme 1). The solvent was evaporated under vacuo. Yield: 0.97 g (68 %). ¹H NMR (400 MHz, DMSO- d_6 , ppm): $\delta = 10.59$ (s, 1 H), 8.85 (d, J =4 Hz, 1 H), 8.51 (d, J = 8 Hz, 1 H), 8.27 (d, J = 8 Hz, 1 H), 7.52 (d, J = 4 Hz, 1 H), 7.50 (t, J = 4 Hz, 1 H), 7.38 (t, J = 8 Hz)1 H), 2.97 (t, J = 6.4 Hz, 4 H), 2.79 (t, J = 6.4 Hz, 4 H), 2.41 (s, 6 H); ¹³C NMR (100 MHz, CD₃CN, ppm): 163.09, 162.82, 162.52, 149.15, 137.13, 136.03, 127.05, 122.31, 122.12, 116.65, 55.77, 49.11, 48.73, 43.54, 35.09, 27.59. LRMS (ESI): m/z calcd for $C_{16}H_{22}N_4O-H^++Zn^{2+}$: 547.23; found 547.20. Elemental analysis calcd (%) for C₁₆H₂₃N₄: C, 67.11; H, 7.74; N, 19.56; found: C, 66.87; H, 7.92; N, 19.83.

Fluorescence Titration of 1 Toward Zn²⁺

The receptor **1** (1.72 mg, 0.006 mmol) was dissolved in MeCN (2 mL) and 20 μ L of the receptor **1** (3 mM) was diluted to 2.98 mL MeCN/bis-tris buffer solution (3:7, *v*/v) to make the final concentration of 20 μ M. Zn(NO₃)₂ 6H₂O (11.9 mg, 0.04 mmol) was dissolved in MeCN (2 mL) and 3–36 μ L of the Zn²⁺ solution (20 mM) was transferred to each receptor



Fig. 1 Fluorescence spectral changes of 1 (20 μ M) in the presence of different metal ions (12 equiv) such as Al³⁺, Zn²⁺, Cd²⁺, Cu²⁺, Fe³⁺, Mg²⁺, Cr³⁺, Hg²⁺, Co²⁺, Ni²⁺, Na⁺, K⁺, Ca²⁺, Mn²⁺ and Pb²⁺ with an excitation of 523 nm in a mixture of MeCN/bis-tris buffer solution (3:7, ν/ν)



Fig. 2 Fluorescence spectral changes of 1 (20 μ M) in the presence of different concentrations of Zn²⁺ ions in a mixture of MeCN/bis-tris buffer solution (3:7, ν /v). Inset: Fluorescence intensity at 523 nm versus the number of equiv. of Zn²⁺ added

solutions prepared above. After mixing them for a few seconds, fluorescence spectra were taken at room temperature.

UV-vis Titration of 1 Toward Zn²⁺

The receptor **1** (1.72 mg, 0.006 mmol) was dissolved in MeCN (2 mL) and 30 μ L of the receptor **1** (3 mM) were diluted to 2.97 mL MeCN/bis-tris buffer solution (3:7, *v*/v) to make the final concentration of 30 μ M. Zn(NO₃)₂ 6H₂O (11.9 mg, 0.04 mmol) was dissolved in MeCN (2 mL) and 0.9–9 μ L of the Zn²⁺ solution (20 mM) were added to the receptor **1** solution prepared above. After mixing them for a few seconds, UV-vis spectra were obtained at room temperature.

Fig. 3 UV-vis titration of 1 (30 μ M) with Zn²⁺ (0–2 equiv). Inset: Absorption titration profile of 1 with Zn²⁺ at 322 nm

Job Plot Measurements

The receptor **1** (1.72 mg, 0.006 mmol) was dissolved in MeCN (2 mL). 500 μ L of the receptor solution was taken and diluted with MeCN/bis-tris buffer solution (3:7, *v*/v) to make the final concentration of 50 μ M. The total volume of the receptor solution was 30 mL. Zn(NO₃)₂ 6H₂O (11.9 mg, 0.04 mmol) was dissolved in MeCN (2 mL). 75 μ L of the zinc solution (3:7, *v*/v). The total volume of zinc solution was 30 mL. 0.3, 0.6, 0.9, 1.2, 1.5, 1.8, 2.1, 2.4 and 2.7 mL of the **1** solution were taken and transferred to vials. 2.7, 2.4, 2.1, 1.8, 1.5, 1.2, 0.9, 0.6 and 0.3 mL of the zinc solution were added to each diluted **1** solution. Each vial had a total volume of 3 mL. After reacting them for a few seconds, fluorescence spectra were taken at room temperature.

Competition with Other Metal Ions

The receptor **1** (1.72 mg, 0.006 mmol) was dissolved in MeCN (2 mL) and 20 μ L of this solution (3 mM) was diluted with 2.98 mL of MeCN/bis-tris buffer solution (3:7, ν/ν) to make the final concentration of 20 μ M. MNO₃ (M = Na, K, 0.04 mmol) or M(NO₃)₂ (M = Zn, Cd, Cu, Mg, Co, Ni, Ca, Mn and Pb, 0.04 mmol) or M(NO₃)₃ (M = Al, Fe and Cr, 0.04 mmol) or M(ClO₄)₂ (M = Fe, 0.04 mmol) was separately dissolved in MeCN (2 mL). 36 μ L of Zn²⁺ solution and each metal solution were taken, respectively, and added to receptor **1** prepared above to give 12 equiv. After mixing them for a few seconds, fluorescence spectra were obtained at room temperature.



Fig. 4 Positive-ion electrospray ionization mass spectrum of 1 (100 μ M) upon addition of Zn(NO₃)₂ (1 equiv)



pH Effect Test of 1 Toward Zn²⁺

A series of buffers with pH values ranging from 2 to 12 was prepared by mixing sodium hydroxide solution and hydrochloric acid in bis-tris buffer. After the solution with a desired pH was achieved, receptor 1 (1.72 mg, 0.006 mmol) was dissolved in MeCN (2 mL), and then 20 μ L of the receptor 1 (3 mM) were diluted with 2.98 mL MeCN/bis-tris buffer solution (3:7, *v*/v) to make the final concentration of 20 μ M. Zn(NO₃)₂ 6H₂O (11.9 mg, 0.04 mmol) was dissolved in MeCN (2 mL). 36 μ L of the Zn2+ solution (20 mM) were transferred to each receptor solution (20 μ M) prepared above. After mixing them for a few seconds, fluorescence spectra were obtained at room temperature.



Fig. 5 Competitive selectivity of **1** (20 μ M) toward Zn²⁺ (12 equiv) in the presence of other metal ions (12 equiv) with an excitation of 370 nm in a mixture of MeCN/bis-tris buffer solution (3:7, ν/v)

NMR Titration of 1 Toward Zn²⁺

Four NMR tubes of **1** (0.28 mg, 0.01 mmol) dissolved in CD_3CN (0.7 mL) were prepared, and four different equiv. (0, 0.5, 0.8 and 1 equiv) of zinc nitrate dissolved in CD_3CN (0.3 mL) were added separately to the solutions of **1**. After shaking them for a few seconds, the ¹H NMR spectra were taken.

Determination of Zn²⁺ in Water Samples

Fluorescence spectral measurements of water samples containing Zn^{2+} were performed by adding 20 μ L of 3 mmol/L stock solution of **1** and 0.60 mL of 50 mmol/L bis-tris buffer



Fig. 6 Fluorescence intensity (at 523 nm) of **1** (20 μ M) in the presence of Zn²⁺ at different pH values (2–12) in a mixture of MeCN/bis-tris buffer solution (3:7, ν /v)





Fig. 7 ¹H NMR titration of 1 with Zn(NO₃)₂ 6H₂O

solution to 2.38 mL sample solutions. After well mixed, the solutions were allowed to stand at 25 $^{\circ}$ C for 2 min before the test.

Theoretical Calculation Methods

All DFT/TDDFT calculations based on the hybrid exchange-correlation functional B3LYP [49, 50] were carried out using Gaussian 03 program [51]. The 6-31G** basis set [52, 53] was used for the main group elements, whereas the Lanl2DZ effective core potential (ECP) [54, 55] was employed for Zn. In vibrational

frequency calculations, there was no imaginary frequency for the optimized geometries of **1** and $1-Zn^{2+}$, suggesting that these geometries represented local minima. For all calculations, the solvent effect of water was considered by using the Cossi and Barone's CPCM (conductor-like polarizable continuum model) [56, 57]. To investigate the electronic properties of singlet excited states, time-dependent DFT (TDDFT) was performed in the ground state geometries of **1** and $1-Zn^{2+}$. Thirty lowest singlet states were calculated and analyzed. The GaussSum 2.1 [58] was used to calculate the contributions of molecular orbital in electronic transitions.







Fig. 8 Fluorescence intensity (at 523 nm) of **1** as a function of Zn(II) concentration. [**1**] = 20 µmol/L, [Zn(II)] = 0–120 µmol/L. Conditions: all samples were conducted in a mixture of MeCN/bis-tris buffer solution (3:7, ν/ν). λ_{ex} and λ_{em} were 370 and 523 nm, respectively

Results and Discussion

Synthesis of 1

The compound **1** 3-((2-(dimethylamino)ethyl)amino)-N-(quinolin-8-yl)propanamide was synthesized by substitution reaction of 3-chloro-N-(quinolin-8-yl)propanamide and N,N'-dimethyl ethylene amine in acetonitrile (Scheme 1), and characterized by ¹H NMR, ¹³C NMR, elemental analysis and ESI-mass spectrometry.

Fluorescence and Absorption Spectroscopic Studies of 1 Toward Zn²⁺

The fluorometric behavior of the receptor **1** toward various metal ions was studied in a mixture of MeCN/bis-tris buffer solution (3:7, ν/ν). When excited at 370 nm, receptor **1** exhibited a weak fluorescence emission ($\lambda_{max} = 523$ nm) compared to that (424 folds) in the presence of Zn²⁺ (Fig. 1). By contrast, upon addition of other metal ions such as Al³⁺, Cd²⁺, Cu²⁺,

 Table 1
 Determination of Zn(II) in water samples

Sample	Zn(II) added (µmol/L)	Zn(II) found (µmol/L)	Recovery (%)	R.S.D. (<i>n</i> = 3) (%)
Tap water	0.00	0.00		
	4.00	4.12	103.0	0.37
Water sample	0.00	0.00		
	4.00 ^[a]	9.82	98.2	1.88

[a] Synthesized by deionized water, 6.00 μ mol/L Zn(II), 10 μ mol/L Cd(II), Pb(II), Na(I), K(I), Ca(II), Mg(II). Conditions: [1] = 20 μ mol/L in 10 mM MeCN:bis-tris buffer solution (3:7, pH 7.0)

Fe²⁺, Fe³⁺, Mg²⁺, Cr³⁺, Hg²⁺, Co²⁺, Ni²⁺, Na⁺, K⁺, Ca²⁺, Mn²⁺ and Pb²⁺, either no or slight increase in intensity was observed. These results indicated that the receptor 1 could be used as a fluorescence chemosensor for Zn²⁺ and discriminate Zn²⁺ from Cd²⁺ [59–63]. Moreover, we examined the fluorometric properties of 1 with Zn²⁺ in polar and non-polar solvents such as chloroform, methanol (MeOH), acetonitrile (MeCN) and N,N-dimethylformamide (DMF) (Fig. S1). 1 displayed strong fluorescence with Zn²⁺, which featured a red-shift with increase of the solvent polarity.

To further investigate the chemosensing properties of 1, fluorescence titration of the receptor 1 with Zn^{2+} ion was carried out. As shown in Fig. 2, the emission intensity of 1 at 523 nm gradually increased until the amount of Zn^{2+} reached 12 equiv. The binding properties of 1 with Zn^{2+} were further studied by UV-vis titration experiments (Fig. 3). UV-vis absorption spectrum of 1 showed two absorption bands at 240 nm and 310 nm. Upon the addition of Zn^{2+} ion to the solution of 1, the two bands have red-shifted to 257 and 367 nm, respectively. Meanwhile, three clear isosbestic points were observed at 246 nm, 283 and 336 nm, implying the undoubted conversion of free 1 to a zinc complex.

The Job plot showed a 1:1 complexation stoichiometry between 1 and Zn²⁺ (Fig. S2) [64], which was further confirmed by ESI-mass spectrometry analysis (Fig. 4). The positive-ion mass spectrum of 1 upon addition of 1 equiv. of Zn²⁺ showed the formation of the $[1-H^++Zn^{2+}+2IPA+DMSO]^+$ [m/z: 547.20; calcd, 547.23]. From the fluorescence titration data, the association constant for 1 with Zn²⁺ was determined as 1.4 x 10⁴ M⁻¹ using Benesi-Hildebrand method (Fig. S3) [65]. This value is within the range of those (1.0 ~ 1.0 x 10¹²) reported for Zn²⁺ sensing chemosensors [66–68]. The limit of detection was estimated to check the efficiency of the probe, which was based on the 3 σ /slope (Fig. S4) [69, 70]. The detection limit for Zn²⁺ was determined as 7.1 μ M, which was much lower than the WHO guideline (76 μ M) for Zn²⁺ ions in drinking water [71, 72].

To explore the ability of 1 as a fluorescence receptor for Zn^{2+} , interference experiments were performed in the presence of Zn^{2+} (12 equiv) mixed with various metal ions (12 equiv) (Fig. 5). There was no interruption for the detection of Zn^{2+} in the presence of Mg^{2+} , Hg^{2+} , Ni^{2+} , Na^+ , K^+ , Ca^{2+} , Mn^{2+} and Pb^{2+} , while relatively low detectable responses were observed in the presence of Al^{3+} , Cu^{2+} , Fe^{2+} , Fe^{3+} , Cr^{3+} and Co^{2+} . On the other hand, Cd^{2+} ion hardly inhibited the fluorescence intensity of $1-Zn^{2+}$. These results suggest that 1 could be a good sensor for Zn^{2+} and, indeed, distinguish Zn^{2+} from Cd^{2+} commonly having similar properties in the same group of the periodic table.

The pH dependence of the $1-Zn^{2+}$ complex was examined. Over the pH range tested, the fluorescence



Fig. 9 Energy minimized structures of (a) 1 and (b) 1-Zn²⁺

intensity of $1-Zn^{2+}$ displayed strong pH dependence (Fig. 6). An intense and stable fluorescence of $1-Zn^{2+}$ found in the pH range of 7.0–12.0 warrants its application under physiological conditions, without any change in detection results.

¹H NMR Spectroscopic Studies of 1 Toward Zn²⁺

The ¹H NMR titration experiments were studied to further examine the binding mode between **1** and Zn^{2+} ion (Fig. 7). Upon addition of Zn^{2+} to receptor **1**, H_{10} disappeared at 0.5 equiv. H_8 , H_9 , H_{11} , H_{12} and H_{13} showed significantly downfield shift, while H_7 shifted upfield and the protons in quinoline slightly shifted downfield or upfield. There was no shift in the position of proton signals on further addition of Zn^{2+} (>1.0 equiv). These results suggest that two nitrogen atoms in dimethyl ethylene amine might coordinate to Zn^{2+} ion (Scheme 2). Based on these results, we proposed that the low fluorescence of **1** could be due to photoinduced electron transfer (PET) from lone-pair electrons of receptor (dimethyl ethylene amine) to fluorophore (quinolone). Thus, 'off-on'

fluorescence of 1 caused by Zn^{2+} might be attributed to the inhibition of PET (Scheme 2).

Determination of Zinc ion in Water Samples

We constructed a calibration curve for the determination of Zn^{2+} by 1 (Fig. 8). Receptor 1 showed a good linear relationship between the fluorescence intensity of 1 and Zn^{2+} concentration (0–120 μ M) with a correlation coefficient of R² = 0.9821 (n = 3). This result indicates that **1** is suitable for quantitative detection of Zn^{2+} . In order to examine the applicability of the receptor 1 in environmental samples, the chemosensor was applied for the determination of Zn^{2+} in water samples. First, tap water samples were chosen. As shown in Table 1, one can see a satisfactory recovery and R.S.D. values of water samples. Also, we prepared artificial polluted water samples by adding various metal ions known as being involved in industrial processes into deionized water. The results were also summarized in Table 1, which exhibited a satisfactory recovery and R.S.D. values for the artificial water samples.

Theoretical Calculations

To gain an insight into fluorescent sensing mechanism for 1- Zn^{2+} , time-dependent density functional theory (TD-DFT) calculations were performed at the optimized geometries (S_0) of 1 and 1-Zn²⁺ complex (Fig. 9). In case of 1, the main molecular orbital (MO) contributions of the first lowest excited states were determined for HOMO \rightarrow LUMO and HOMO - 1 \rightarrow LUMO transition (332.82 nm, Fig. S5). As shown in Fig. S6, HOMO - 1 \rightarrow LUMO of 1 indicates $\pi \rightarrow \pi^*$ transition in quinoline moiety, which means radiative transition. HOMO \rightarrow LUMO of 1 indicates PET from dimethyl ethylene amine to quionline, which could explain the non-radiative process of 1. For $1-Zn^{2+}$ complex (Fig. S7), the third lowest excited state was considered as main transition of 1-Zn²⁺ complex (oscillator strength = 0.1317), while its first lowest excited state showed minor transition (oscillator strength = 0.0284). The main molecular orbital (MO) contribution of the third lowest excited state was determined for HOMO - 1 \rightarrow LUMO (330.48 nm). As shown in Fig. S5, it shows $\pi \rightarrow$ π^* transition in quinoline moiety, which indicates radiative transition. Thus, these results suggested that the sensing mechanism of 1 toward Zn²⁺ was originated by inhibition of PET process [73].

Conclusion

We have synthesized a new fluorescent chemosensor 1, which displays high sensitivity and selectivity toward zinc in aqueous media. The complexation of 1 with Zn^{2+} exhibited a pronounced enhancement in the fluorescence emission. Moreover, the detection limit (7.1 μ M) is much lower than the WHO detection level (76 μ M) for Zn^{2+} ions in drinking water. Most importantly, recovery studies of the water samples added with Zn^{2+} demonstrated its value in the practical application. Therefore, we believe that receptor 1 will be a prototype for the practicable system for detecting Zn^{2+} concentrations in environmental systems.

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