

Olfactory Response of the Spotted Asparagus Beetle, *Crioceris duodecimpunctata* **(L.) to Host Plant Volatiles**

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Abstract

The spotted asparagus beetle, *Crioceris duodecimpunctata* (L.) is an invasive host-specifc pest of asparagus cultivations. To contribute to the understanding of the role of plant volatiles in host-fnding by this species, behavioural and electrophysiological tests were carried out. Y-tube olfactometer bioassays, testing intact or mechanically-damaged cladophylls vs. clean air, revealed sexually-dimorphic responses with males being the only sex attracted to both plant materials. Electroantennographic (EAG) assays showed that antennae of both sexes can perceive a wide range of asparagus volatiles. Male and female EAG profles were almost similar and (*Z*)-3-hexen-1-ol was by far the most EAG-active compound. (*E*)-2-hexenal, (\pm) -linalool, and 3-heptanone elicited the strongest EAG amplitude within the corresponding chemical groups. Eight of the most EAG-active compounds elicited dose-dependent responses indicating the sensitivity of male and female olfactory systems to changes in stimulus concentration. In a Y-tube olfactometer bioassay, (*Z*)-3-hexen-1-ol at the doses of 1, 10, and 50 μg did not elicit female attraction whereas a signifcant attraction at the 10 μg dose and a repellent efect at the 50 μg dose was induced in males. Sexual dimorphism of male behavioural response to host plant volatiles is discussed. This study provides a basis for future investigations that could contribute to the development of semiochemical-based monitoring and management strategies for this pest.

Keywords Chrysomelidae · *Asparagus officinalis* · EAG · Olfactometer bioassays · VOCs · Kairomone

Introduction

Host plant utilization by phytophagous insects depends on coordinated insect-plant interaction (Thorpe et al. [1947](#page-9-0); Dethier [1947\)](#page-8-0). Host plant location by phytophagous insects is mediated by numerous sensory inputs, including olfactory and gustatory cues as well as physical and visual information (Visser [1986;](#page-9-1) Pickett et al. [1998;](#page-9-2) Bruce et al. [2005](#page-8-1); Van den Berg et al. [2008\)](#page-9-3). In the Chrysomelidae family, orientation to host plant cues has been investigated intensively for the Colorado potato beetle, *Leptinotarsa decemlineata* (Say). Adult *L. decemlineata* attraction to volatiles emitted by undamaged potato plants was observed about one century ago (McIndoo [1926\)](#page-8-2) and was even the frst evidence that host plant odours attract insects (Dickens [2000a](#page-8-3)). Later on,

the olfactory sensitivity of male and female Colorado potato beetle antennal receptors to plant volatiles was carefully investigated by electrophysiological experiments (Visser [1979;](#page-9-4) Ma and Visser [1978;](#page-8-4) Weissbecker et al. [1997](#page-9-5); Dickens [1999](#page-8-5)) and adults' attraction to a fve-component blend comprising of (E) -2-hexen-1-ol, (Z) -3-hexen-1-ol, (\pm) -linalool, nonanal, and methyl salicylate was demonstrated (Dickens [2000b](#page-8-6)).

The spotted asparagus beetle, *Crioceris duodecimpunctata* (L.) (Coleoptera, Chrysomelidae), is a major monophagous pest of commercially grown *Asparagus officinalis* L. in most areas of production (LeSage et al. [2008](#page-8-7); Morrison III and Szendrei [2014](#page-8-8)). Native to the Palearctic region (Drake and Harris [1932\)](#page-8-9), mostly around the Mediterranean Sea (FGP Consortium 2014), *C. duodecimpuntata* was frst detected outside its native range in 1881 in the United States near Baltimore, MD (Chittenden [1917](#page-8-10)). Subsequently, the pest spread north and westward across Canada and the United States (LeSage et al. [2008](#page-8-7)) where it can now be found wherever asparagus is commonly grown (Capinera 2001 ; Morrison III and Szendrei [2014](#page-8-8)).

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C. duodecimpunctata overwinters as an adult and undergoes two generations per year (Pollini [1998](#page-9-6); Capinera [2001](#page-8-11)). In the spring, adults emerge from overwintering shelters such us hollow asparagus stems and other crop debris and feed on asparagus spears. Subsequently, they deposit eggs singly on cladophylls and ramifcations of thin asparagus branches. Larvae complete their development feeding on 2-5 asparagus berries (Fink [1913;](#page-8-12) Dingler 1935). Mature larvae fall off the plant and pupate in the soil. The newly emerged adults repeat this cycle from which overwintering adults will develop.

In the spring, adult feeding on emerging asparagus spears creates small pits in the epidermis (Capinera [2001](#page-8-11); Morrison III and Szendrei [2014\)](#page-8-8) resulting in severe direct and aesthetic damage to commercial production. Later in the season, the feeding activity of *C. duodecimpunctata* adults and larvae on leaves causes reduction of plant photosynthetic capability reducing crop production in the subsequent year (Capinera [2001\)](#page-8-11). Moreover, larval feeding on berries prevents seed formation (Capinera [2001\)](#page-8-11) and therefore could be a problem for breeders and seed producers (Morrison and Szendrei 2014).

The life cycle of *C. duodecimpunctata* is similar to that of the sympatric *Crioceris asparagi* (L.), but damage to asparagus spears by *C. duodecimpunctata* is considered less important due to its later emergence in the season (Morrison III and Szendrei [2014\)](#page-8-8). However, the adult fight periods, as well as the relative abundance of the two species in diferent areas, remain to be fully defned. For example, in the Foggia Province (Apulia, southern Italy), the most important area of green asparagus cultivation in Italy, only the presence of *C. duodecimpunctata* was noticed from the end of April to June, corresponding to the majority of the harvesting period, as well as a possible overlap of adult fight periods of the two *C. duodecimpunctata* generations (personal observations).

In this context, the identifcation of host-plant compounds to attract *C. duodecimpunctata* adults could greatly contribute to the development of semiochemical-based monitoring tools useful to improve the timing of control measures and to develop low-impact direct control means such as mass trapping and/or attract and kill methods.

To date, no studies have been carried out to investigate the capability of male and female *C. duodecimpunctata* to perceive and orient to host-plant odours. To contribute to the knowledge on the role of chemical signals in host location by *C. duodecimpunctata* adults, in this study Y-tube olfactometer bioassays and electroantennographic (EAG) tests were performed. Experiments were designed to investigate: 1) the behavioural response of males and females to odours emitted by intact or mechanically damaged host plant cladophylls; 2) the antennal capability of both sexes to perceive a range of asparagus volatile organic compounds (VOCs); and, 3) the behavioural response of males and females to the most EAG-active compound (*Z*)-3-hexen-1-ol.

Methods and Materials

Insects During April to June 2018, adults of *C. duodecimpunctata* were collected from infested asparagus plants in privately owned lands near Borgo Mezzanone and Lesina (Foggia, Apulia Region, Italy). Permission to collect insects was obtained from the owners. The feld-collected adults were transferred to rearing cages $(40 \times 40 \times 60 \text{ cm})$ and maintained at 25 ± 2 °C, $60 \pm 5\%$ relative humidity (r.h.), and L16:D8 photoperiod. Insects were fed with *A. officinalis* stems and spears placed with the base into a glass beaker (500 mL) containing tap water (300 mL) and replaced every 2 days.

All individuals used in the experiments were actively feeding on plant materials and exhibiting mating activity. A small group of insects was also dissected and their mature reproductive status was confrmed by the presence of welldeveloped ovaries, ovarioles, and accessory glands (Bean et al. [2007](#page-8-14); Gafke et al. [2020](#page-8-15)).

Prior to EAG and behavioural tests, adult beetles were kept individually in transparent plastic containers (6 cm i.d. \times 8 cm) covered with a fine mesh net (1 mm) without food supply and in the absence of asparagus odours for at least 30 min. Experimental insects were used only once. At the end of each experiment, the specimen was dissected and the sex determined by observing the genitalia with a stereomicroscope (SPZ series, Optika, Ponteranica, Italy).

Plant Materials *A. officinalis* (cultivar Sunlim F1) stems (approximately 50 cm long) were collected in the feld from the end of April to June and placed with the cut ends into a glass beaker containing tap water. Plant materials were used for behavioural bioassays no later than 24 h after cutting.

Chemicals Twenty-three VOCs were selected among the most abundant identifed from intact or mechanically damaged *A. officinalis* spears and cladophylls (Sun et al. [2001](#page-9-7); Thibout et al. [2005;](#page-9-8) Morrison III et al. [2016](#page-8-16)) and to represent diferent chemical classes (Table [1](#page-2-0)). Compounds were purchased from Sigma-Aldrich (Milan, Italy). For EAG experiments, all compounds were dissolved in mineral oil (Sigma-Aldrich) and stored at −20 °C until needed.

Olfactometer Bioassays The behavioural response of *C. duodecimpunctata* males and females to odours of host plant material was assessed in a glass Y-tube olfactometer (each arm 26 cm long at 75 °C angle, stem 30 cm long, 6.0 cm i.d.) similar to that previously described (Germinara et al. [2011](#page-8-17)). Each arm of the Y-tube was connected to a glass cylinder **Table 1** Absolute EAG responses of male and female *Crioceris duodecimpunctata* antennae $(n=6)$ to the 1 mg dose (10 μL of a 100 μg/μL) of volatile organic compounds (VOCs) identifed from Asparagus officinalis

^aFor each compound, mean male and female EAG responses were compared by Student's *t* test for independent samples $(df=10; P=0.05)$

(9 cm long and 6.0 cm i.d.; 1 cm high, 2 cm i.d. screw-cup central opening) as an odour source container. The apparatus was put into an observation chamber $(80 \times 80 \times 70 \text{ cm})$ and illuminated from above by two 28-W cool white fuorescent lamps providing uniform lighting (2500 lx) inside the tube. A purifed (activated charcoal) and humidifed airfow maintained at 60 mL/min by a fowmeter was pumped through each arm.

Two choice tests were conducted: (1) intact cladophylls versus clean air; (2) mechanically damaged cladophylls versus clean air. Intact test material (1.25 g) consisted of cladophylls (4-5 cm long) detached from the stems 30 min before the experiment and placed with the cut end in a glass vial containing water to maintain the physiological water content (Tasin et al. [2005\)](#page-9-9). Mechanically damaged test material (1.25 g) consisted of cladophylls detached from plant stems and cut into pieces (1 cm long) using scissors 30 min before the experiment.

The behavioural response of adult beetles to diferent concentrations of the strongest antennal stimulant (*Z*)- 3-hexen-l-ol was also tested. In this case, the odour chamber contained a flter paper (Whatman, cat. No 1001–110,

Buckinghamshire, UK) disk (2 cm i.d.) (loaded with 1, 10, or 50 μg (10 μL of a 0.1 μg, 1 μg, or 5 μg/μL mineral oil solution, respectively) of (*Z*)-3-hexen-l-ol while the other chamber contained a similar flter paper disk loaded with 10 μL of mineral oil. A white cotton thread (8 cm long; 0.5 mm i.d) was passed through the centre of a disk using a needle. The disk was then inserted into the odor chamber through the central opening and suspended in the centre of the cross section by properly fxing the cotton thread ends between the screw cap and the central opening of the chamber.

Bioassays were run between 10.00 and 14.00 h at 25 ± 2 °C and $60 \pm 5\%$ r.h. Each experiment lasted 10 min. Individual insects, within 3 days of feld collection, were released at the open end of the Y-tube stem. A choice was recorded when the insect moved 3 cm up an arm of the Y-tube and remained beyond the decision line (marked on both arms) for more than 30 s. The time spent by test insects in each arm was also recorded. After 3 individuals were tested, the olfactometer was cleaned with distilled water and acetone, dried (200 °C for 30 min) and test and control stimuli renewed. The position of the treatments in the arms

were switched to avoid positional bias. For each test stimulus, at least 30 beetles of each sex, used once, were tested. Individuals used in these experiments were from cohorts of 40–50 adults collected every 3-4 days from the end of April to June.

Electroantennography (EAG) The EAG technique was used to assess the antennal selectively and sensitivity of *C. duodecimpunctata* males and females to the selected 23 volatile organic compounds (VOCs).

An antenna was excised at the base and mounted between glass electrodes flled with Kaissling's saline (Kaissling and Thorson [1980\)](#page-8-18). The electrical continuity between the antennal preparation and an IDAC-4 amplifer (Syntech Laboratories, Hilversum, The Netherlands) connected to a PC equipped with the Software EAG Pro (Syntech Laboratories, Hilversum, The Netherlands) was maintained using AgClcoated silver wires.

Just before EAG experiments, 10 μL of each test solution (100 μ g/ μ L) was adsorbed onto a filter paper strip (1 cm, Whatman No. 1) placed in a Pasteur pipette (15 cm long), which served as an odour cartridge. Vapour stimuli (2.5 cm^3) were blown by a disposable syringe for 1 s into a constant stream of charcoal-fltered humidifed air (500 mL/min) fowing in a glass delivery tube (i. d. 8 mm) with the outlet positioned at approximately 1 cm from the antenna.

For each sex, antennal selectivity was assessed based on the EAG response to the 1 mg dose (10 μL of a 100 μg/μL mineral oil solution) of each test VOC. To evaluate antennal sensitivity, EAG dose-response curves were calculated on stimulation with 0.01, 0.1, 1, 10, and 100 μg doses (10 μ L of 0.001, 0.01, 0.1, 1, 10, 100 μg/μL mineral oil solutions, respectively) of 8 compounds selected among the most EAG-active in diferent chemical classes.

Control (10 μL mineral oil) and standard (10 μL of 10 μg/ μL (*Z*)-3-hexen-l-ol mineral oil solution) stimuli were applied at the beginning of the experiment and after each group of 6 test stimuli. To allow the recovery of the antennal responsiveness, stimuli were presented at 1-min intervals.

In antennal selectivity experiments, each compound was tested on 6 antennae of diferent males and females. In antennal sensitivity experiments, test compounds were assessed on 3 antennae of diferent specimens of each sex.

Data Analyses The absolute EAG response (mV) to each stimulus was subtracted by the mean response to the two nearest controls (mineral oil) to compensate for solvent and mechanosensory artifacts (Raguso and Light [1998](#page-9-10)). The subsequent EAG value was corrected according to the reduction of the EAG amplitude to the standard stimulus to compensate for the decrease of the antennal responsiveness during the experiment (Otter et al. [1991\)](#page-9-11).

The corrected male and female EAG responses to the 1 mg dose of each compound were compared to a "0" value using the Wilcoxon rank sum test and considered measurable if significant at $P = 0.05$. The mean EAG responses of males and females to diferent test compounds of each chemical group were subjected to analysis of variance (ANOVA) followed by the Tukey's HSD (Honestly Signifcant Diference) test ($P = 0.05$) or to Student's *t* test for mean comparison. Prior to these analyses, values were \sqrt{x} -transformed and tested for homogeneity of variance using Levene's test. The mean male and female EAG responses to each test stimulus were compared using the Student's *t* test ($P = 0.05$) for independent samples. In EAG dose-response curves, the activation threshold was taken as the frst dose at which the mean response was higher than a "0" value using the Shapiro-Wilk test for normality followed by the one-sample Student's *t* test $(P=0.05)$ (Germinara et al. [2017\)](#page-8-19); the saturation level was considered to be the lowest dose at which the mean response was equal to or less than the previous dose (Germinara et al. [2009\)](#page-8-20). Signifcant diferences between the number of beetles choosing the treatment or control arm of the olfactometer were compared using χ^2 tests. The differences between the time spent by beetles in each arm were analyzed by paired sample *t*-tests. Statistical analyses were performed using SPSS (Statistical Package for the Social Sciences) version 10.0.7 for Windows (SPSS Inc., Chicago, IL, USA).

Results

Behavioural Response to Plant Material In Y-tube behavioural bioassays, males presented with intact cladophylls vs. clean-air control exhibited a signifcant preference for the odour stimulus (χ 2 = 13.37, *df* = 1, *P* < 0.001) and spent significantly $(t=3.97, df=32, P<0.001)$ more time in the treatment arm (Table [2](#page-4-0)). When males were given a choice between mechanically damaged cladophylls and clean air, there was no preference $(\chi^2 = 2.13, df = 1, P = 0.14)$ but they spent significantly $(t=3.67, df=33, P<0.001)$ more time in the treatment arm. Odours from intact and mechanically damaged cladophylls did not elicit a signifcant attraction in female beetles measured either as frst choice or as time spent in the treatment arm when clean air was the alternative.

Antennal Selectivity The mean EAG responses of *C. duodecimpunctata* males and females to diferent test stimuli are shown in (Table [1\)](#page-2-0). At the 1 mg dose, all test compounds elicited measurable EAG responses (*P*<0.05 in all Wilcoxon rank sum test) ranging from 0.12 ± 0.05 mV for α-pinene to 4.22 ± 0.16 mV for (*Z*)-3-hexen-1-ol in males, and from 0.06 ± 0.03 mV for hexadecene to 3.52 ± 0.24 mV for (*Z*)-3-hexen-1-ol in females. Among all compounds tested, the

Female $32(22)$ 59.09 0.73 0.39 1.97 \pm 0.51 1.02 \pm 0.35 1.39 0.18

Table 2 Response of *C. duodecimpunctata* males and females in a Y-tube olfactometer to different odour sources of *A. officinalis* (cultivar Sunlim F1)

a Total sample size (N, number of individuals that made a choice in parentheses)

^bProportion of individuals (of those that made a choice) that chose the treated arm first

highest EAG responses were evoked by (*Z*)-3-hexen-1-ol, 1-octen-3-ol, 3-heptanone, (*E*)-2-hexenal, heptanal, nonanal, hexanal, and (\pm) -linalool in both sexes. The weakest stimulants were α-pinene and β-pinene in males and hexadecene and *p*-cymene in females. The mean male EAG response to (Z) -3-hexen-1-ol was significantly $(t=2.39, df=10,$ $P=0.038$) higher than that of females. For the remaining compounds, male and female EAG responses did not difer significantly $(t=0.03-1.60, df=10, P=0.14-0.99)$.

In both sexes, signifcant diferences were found among the EAG responses to test compounds in the chemical groups of aliphatic alcohols (male, $t = 4.625$, $df = 10$, $P = 0.001$; female, $t = 6.625$, $df = 10$, $P < 0.001$), terpenes (male, *F* = 17.238; *df* = 9, *P* < 0.001; female, *F* = 8.013; *df* = 9, *P*<0.001), and others (male, *F*=8.589; *df*=5, *P*<0.001; female, $F = 16.886$; $df = 5$, $P < 0.001$). As regards aliphatic aldehydes, there were signifcant diferences among the EAG responses of males ($F = 2.787$; $df = 4$, $P = 0.048$) but not of females (*F*=1.809; *df*=4, *P*=0.159). In both sexes, (*E*)-2-hexenal, (*Z*)-3-hexenol, (±)-linalool, and 3-heptanone elicited the strongest EAG amplitude within the corresponding chemical groups (Fig. [1](#page-5-0)).

Antennal Sensitivity The antennal sensitivity of adult *C. duodecimpunctata* antennae toward increasing concentrations of 8 EAG-active compounds is reported in Fig. [2.](#page-6-0) For all compounds, the amplitude of the EAG response increased with dose. For both sexes, the activation threshold was recorded at the 0.1 μg dose for (*Z*)-3-hexen-1-ol, (E) -2-hexenal, heptanal, nonanal, and (\pm) -linalool and at the 1 μg dose for the remaining compounds. Male and female EAG responses increased from 100 to 1000 μg doses of all compounds, indicating that no saturation of olfactory receptors occurred at the 100 μg dose. Mean male EAG response was signifcantly higher than that of female to 0.1 (*t*=4.756; *df*=4; *P*=0.009) 10 (*t*=3.087; *df*=4; *P*=0.037) and 1000 μg (*t*=3.401; *df*=4; *P*=0.027) of (*Z*)-3-hexen-1-ol, 100 μg (*t*=11.420; *df*=4; *P*<0.001) of (*E*)-2-hexenal, 10 μg (*t*=4253; *df*=4; *P*=0.013) of heptanal, and 100 μg of (\pm)-linalool ($t = 3.101$; $df = 4$; $P = 0.036$).

Behavioural Response to (Z)‑3‑Hexen‑1‑Ol The behavioural responses of adult beetles to increasing doses of the most EAG-active compound (*Z*)-3-hexen-1-ol vs. mineral oil are reported in Table [3](#page-7-0). The 1 μg dose did not elicit signifcant responses in both sexes. At the 10 μg dose, males exhibited a significant preference for the test compound (χ ² = 7.53, $df = 1$, $P < 0.01$) and spent significantly more time in the treatment arm ($t = 3.96$, $df = 28$, $P < 0.001$). At the 50 µg dose, males did not show any preference but they spent significantly more time in the control arm $(t=2.63, df=17)$, *P*=0.013). The 10 and 50 μg doses of (*Z*)-3-hexen-1-ol did not elicit a signifcant response in females measured either as frst choice or time spent in the treatment arm.

Discussion

In Y-tube olfactometer bioassays, adults of *C. duodecimpunctata* exhibited sexually-dimorphic behavioural responses to the odours emitted by cladophylls of the host plant *A. officinalis*. In the absence of visual stimuli, males were signifcantly attracted by odours of cladophylls while females were not. In more detail, male preference towards intact cladophylls was signifcant both in terms of frst choice and time spent in the treatment arm while towards mechanically damaged cladophylls males only spent signifcantly more time in the treatment arm. In these latter experiments, a higher odour concentration in the olfactometer due to tissue breakdown may have initially interfered with the insect orientation towards the host odour source making the frst choice not signifcant. Similar sexually-dimorphic behavioural responses to plant volatiles have been reported for *L. decemlineata* whose males oriented preferentially to specifc blends of host plant volatiles whilst females showed little preference (Dickens [2000a](#page-8-3); Dickens [2006](#page-8-21)). This behaviour in *L. decemlineata* was considered consistent with the presence of a male-produced aggregation pheromone (Dickens et al. [2002](#page-8-22)) and it was suggested that pioneer males initiate colonisation, locating host plants using odour signals and **Fig. 1** EAG (mean ± S.E.) response profle of adult *C. duodecimpunctata* to a range of VOCs. Within each chemical group, compounds are listed according to the mean amplitude of the EAG response $(n=6)$ elicited at the 1 mg (10 μL of a 100 μg/μL mineral oil solution) dose

Absolute EAG response in mV (mean \pm S.E.)

then produce an aggregation pheromone to which colonising beetles of both sexes respond (Dickens [2006](#page-8-21); Landolt and Phillips [1997\)](#page-8-23).

Interestingly, a similar host-fnding behaviour has been proposed for the asparagus fy, *Plioreocepta poeciloptera* (Schrank) (Diptera: Tephritidae). In this monophagous pest, males are attracted to the host plant in the frst day after emergence and begin to release sex pheromone the following day when the combination of male pheromone and the host plant volatiles, mainly green asparagus volatiles, attract inexperienced virgin females which respond to the male pheromone only in the presence of plant volatiles (Thibout et al. [2005\)](#page-9-8). To date, the use of intraspecifc semiochemicals by *C. duodecimpunctata* adults has not been investigated; however, male-produced aggregation pheromones that attract both sexes over long distances have been identifed in diferent chrysomelids including the closely-related species *Oulema melanopus* L. in the Criocerinae subfamily (Rao et al. [2002](#page-9-12)).

Despite diferences between sexes in the behavioural response to host plant odours, EAG selectivity assays revealed a high similarity between the male and female response profles to compounds tested in this study. The EAG responses of males and females to diferent compounds did not difer signifcantly, except for (*Z*)-3-hexen-1-ol which elicited higher EAG responses in males than in females. The similarity in antennal responses of male and female insects to plant odours has been reported for several phytophagous pests and could arise from the use of the same chemical stimuli present in the habitat they share (Li et al. [1992](#page-8-24)). However, sexually dimorphic EAG responses are more likely due to a diferent number of receptor neurons tuned to individual compounds in male and female antennae as a result of diferent roles played by the same compound in the ecology of each sex (Germinara et al. [2009](#page-8-20)). Among all compounds tested, (*Z*)-3-hexen-1-ol was by far the most EAG-active. In both sexes, the response to this compound was about 2-fold higher than that recorded for the second most EAG-active compound, 1-octen-3-ol.

All 8 compounds tested in EAG sensitivity assays elicited dose-dependent responses in males and females with only a few signifcant diferences, at certain doses, between sexes proving the insect olfactory system to be sensitive to change in odour concentration. In both sexes, the lowest activation threshold was recorded for (*Z*)-3-hexen-1-ol, (*E*)-2-hexenal, heptanal, nonanal, and (\pm) -linalool suggesting the insects'

Fig. 2 EAG (mean \pm S.E.) dose-response curves of adult *C. duodecimpunctata* (*n*=3) to some VOCs identifed from *A. officinalis*. Arrows indicate the activation thresholds. Asterisks indicate signifcant diferent between sexes (* $P = 0.05$, ** *P*=0.01: Student's *t* test)

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Table 3 Response of *C. duodecimpunctata* males and females in a Y-tube olfactometer to increasing doses of a (*Z*)-3-hexen-1-ol (10 μL of 0.1, 1,5 μg/μL mineral oil solution) versus mineral oil (10 μl) control

Sex	Odour sources	Dose (μg) of (Z) -3-hexen- 1-ol	N^a	First choice			Number of minutes spent in arm (mean \pm S.E.)			
				$\%$	χ^2	P value	Treated	Control	t-value	P value
Male	(Z) -hexen-1-ol vs. mineral oil	-1	40 (17)	52.94	0.06	0.81	1.03 ± 0.36	0.78 ± 0.28	0.53	0.60
		10	34(29)	71.43	7.76	${<}0.01$	$3.23 + 0.47$	0.87 ± 0.27	3.96	< 0.001
		50	31 (18)	27.78	3.56	0.06	0.62 ± 0.30	2.49 ± 0.59	-2.63	0.013
Female	(Z) -hexen-1-ol vs. mineral oil	$\overline{1}$	30(16)	31.25	2.25	0.13	$0.71 + 0.31$	1.87 ± 0.59	-1.60	0.12
		10	30(25)	64.00	1.96	0.16	$3.14 + 0.60$	2.33 ± 0.57	-0.77	0.45
		50	30(20)	35.00	1.80	0.18	$1.16 + 0.41$	2.05 ± 0.56	-1.15	0.26

^aTotal sample size (N, number of individuals that made a choice in parentheses)

^bProportion of individuals (of those that made a choice) that chose the treated arm first

capability to detect them from a longer distance. Overall, EAG experiments clearly showed that both *C. duodecimpunctata* sexes were able to perceive a wide range of plant volatiles strongly suggesting a role of these chemical cues not only in male but also in the female ecology. Future experiments should address whether EAG-active plant volatiles could enhance or synergy female long distance attraction to a putative male-produced aggregation pheromone (Landolt and Phillips [1997;](#page-8-23) Reddy and Guerrero [2004](#page-9-13); Dickens [2006](#page-8-21)).

According to the results of electrophysiological tests, the behavioural response of *C. duodecimpunctata* males and females to the most EAG-active compound, (*Z*)-3-hexeno-1-ol was evaluated in further Y-tube olfactometer bioassays. Sexual-dimorphic responses similar to those seen with host plant materials were observed. In the experimental conditions adopted, none of the three (*Z*)-3-hexen-1-ol doses tested elicited a signifcant preference in females. On the contrary, males were signifcantly attracted, both at frst choice and time spent in the treatment arm, by the 10 μg dose of (*Z*)-3-hexen-1-ol but they were repelled by the 50 μg. (*Z*)-3-hexen-1-ol is a general green leaf volatile (GLV) (Paré and Tumlinson [1999](#page-9-14)) contributing to the "green odour" of the leaves of numerous plant species that plays diferent roles in insect-plant interactions (Wei and Kang [2011](#page-9-15)). The GLVs may be produced constitutively or induced by leaf damage and may be caused by abiotic or biotic elicitation (Visser [1986;](#page-9-1) Loughrin et al. [1996](#page-8-25); Turlings et al. [1998](#page-9-16); Ruther et al. [2002;](#page-9-17) Graus et al. [2004](#page-8-26); Chamberlain et al. [2006](#page-8-27)). GLVs have been identifed as major components of *A. officinalis* including (*Z*)-3-hexen-1-ol, which was identifed from healthy plants and shown to be attractive to the asparagus miner, *Ophiomyia simplex* Loew (Diptera: Agromyzidae) (Morrison III et al. [2016](#page-8-16)). Among chrysomelids, diferent behavioural activities have been reported for (*Z*)-3-hexen-1-ol, including diferential responses to variation in concentrations as in this study. In *Cassida denticollis* Sufrian (Coleoptera: Chrysomelidae),

(*Z*)-3-hexen-1-ol enhanced the ability of larvae to distinguish stems of host plants (Müller and Hilker [2000\)](#page-8-28). Adults of *L. decemlineata* were attracted to a five-component blend comprising low doses of the green leaf volatiles (*E*)-2-hexen-1-ol and (*Z*)-3-hexen-1-ol (Dickens [1999\)](#page-8-5), but volatile blends containing relatively high amounts of the same green leaf volatiles were unattractive or repellent (Dickens [2000a](#page-8-3)).

It is interesting to note that while the response of *L. decemlineata* adults is due to ratio of GLVs in the blend rather than individual components, which were individually inactive (Ma and Visser [1978](#page-8-4); Visser [1979;](#page-9-4) Dickens [2000a](#page-8-3)), in *C. duodecimpunctata,* (*Z*)-3-hexen-1-ol alone elicited signifcant behavioural responses in males. This attraction, to be confrmed in further feld studies, along with the observation that (*Z*)-3-hexen-1-ol is released by healthy asparagus plants (Morrison III et al. [2016](#page-8-16)) strengthens the idea that this compound plays a key role at least in the male hostsearching behaviour.

In conclusion, this study demonstrated that *C. duodecimpunctata* adults can detect a variety of *A. officinalis* volatiles with a particular antennal sensitivity to (*Z*)-3-hexen-1-ol in both sexes. However, behavioural bioassays also revealed sexually-dimorphic responses to odours of plant materials and to (*Z*)-3-hexen-1-ol with males being the only responsive sex. Further investigations should test the behavioural activity of combinations of GLVs with other asparagus volatiles towards females to fnd out possible attractive blends and identify a putative male-produced aggregation pheromone in *C. duodecimpunctata*. The kairomonal activity of (*Z*)-3-hexen-1-ol has potential practical interest mainly to develop efective monitoring and control tools of this pest.

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References

- Bean DW, Wang T, Bartelt RJ, Zilkowski BW (2007) Diapause in the leaf beetle *Diorhabda elongata* (Coleoptera: Chrysomelidae), a biological control agent for tamarisk (*Tamarix* spp.). Environ Entomol 36:531–540. [https://doi.org/10.1603/0046-225X\(2007\)](https://doi.org/10.1603/0046-225X(2007)36[531:DITLBD]2.0.CO;2) [36\[531:DITLBD\]2.0.CO;2](https://doi.org/10.1603/0046-225X(2007)36[531:DITLBD]2.0.CO;2)
- Bruce TJ, Wadhams LJ, Woodcock CM (2005) Insect host location: a volatile situation. Trends Plant Sci 10(6):269–274. [https://doi.org/](https://doi.org/10.1016/j.tplants.2005.04.003) [10.1016/j.tplants.2005.04.003](https://doi.org/10.1016/j.tplants.2005.04.003)
- Capinera JL (2001) Order Coleoptera-beetles, weevils. White Grubs and Wireworms. Handbook of vegetable pests, Academic, San Diego
- Chamberlain K, Khan ZR, Pickett JA, Toshova T, Wadhams LJ (2006) Diel periodicity in the production of green leaf volatiles by wild and cultivated host plants of stemborer moths, *Chilo partellus* and *Busseola fusca*. J Chem Ecol 32:565–577. [https://doi.org/10.](https://doi.org/10.1007/s10886-005-9016-5) [1007/s10886-005-9016-5](https://doi.org/10.1007/s10886-005-9016-5)
- Chittenden FH (1917) The asparagus beetles and their control. United States Department of Agriculture, Washington D.C
- Dethier VG (1947) Chemical insect attractants and repellents. Blakiston, Philadelphia
- Dickens JC (1999) Predator-prey interactions: olfactory adaptations of generalist and specialist predators. Agr Forest Entomol 1(1):47– 54. <https://doi.org/10.1046/j.1461-9563.1999.00007.x>
- Dickens JC (2000a) Orientation of Colorado potato beetle to natural and synthetic blends of volatiles emitted by potato plants. Agri Forest Entomol 2(3):167–172. [https://doi.org/10.1046/j.1461-](https://doi.org/10.1046/j.1461-9563.2000.00065.x) [9563.2000.00065.x](https://doi.org/10.1046/j.1461-9563.2000.00065.x)
- Dickens JC (2000b) Sexual maturation and temporal variation of neural responses in adult Colorado potato beetles to volatiles emitted by potato plants. J Chem Ecol 26:1265–1279. [https://doi.org/10.](https://doi.org/10.1023/A:1005492229377) [1023/A:1005492229377](https://doi.org/10.1023/A:1005492229377)
- Dickens JC, Oliver JE, Hollister B, Davis JC, Klun JA (2002) Breaking a paradigm: male-produced aggregation pheromone for Colorado potato beetle. J Exp Biol 205:1925–1933
- Dickens JC (2006) Plant volatiles moderate response to aggregation pheromone in Colorado potato beetle. J App Entomol 130:26–31. <https://doi.org/10.1111/j.1439-0418.2005.01014.x>
- Dingler M (1935) Über unsere beiden Spargelkäter (*Crioceris duodecimpunctata* L. und *Crioceris asparagi* L.). Z Angew Entomol 21(3):415–442
- Drake CJ, Harris HM (1932) Asparagus insects in Iowa. Circular, Ames
- Fink DE (1913) The asparagus miner and the twelve-spotted asparagus beetle. Cornell University Agricultural Experiment Station, Ithaca
- Gafke MA, Sing SE, Millar JG, Dudley TL, Bean WD, Peterson RKD, Weaver DK (2020) An herbivore-induced plant volatile from saltcedar (*Tamarix* spp.) is repellent to *Diorhabda carinulata* (Coleoptera: Chrysomelidae). Environ Entomol 49:1063–1070. <https://doi.org/10.1093/ee/nvaa079>
- Germinara GS, De Cristofaro A, Rotundo G (2009) Antennal olfactory responses to individual cereal volatiles in *Theocolax elegans* (Westwood) (Hymenoptera, Pteromalidae). J Stored Prod Res 45(3):195–200.<https://doi.org/10.1016/j.jspr.2009.02.002>
- Germinara GS, De Cristofaro A, Rotundo G (2011) Chemical cues for host location by the chestnut gall wasp, *Dryocosmus kuriphilus*. J Chem Ecol 37(1):49–56. [https://doi.org/10.1007/](https://doi.org/10.1007/s10886-010-9893-0) [s10886-010-9893-0](https://doi.org/10.1007/s10886-010-9893-0)
- Germinara GS, Ganassi S, Pistillo MO, Di Domenico C, De Cristofaro A, Di Palma AM (2017) Antennal olfactory responses of adult meadow spittlebug, *Philaenus spumarius*, to volatile organic compounds (VOCs). PLoS One 12(12). [https://doi.org/10.1371/](https://doi.org/10.1371/journal.pone.0190454) [journal.pone.0190454](https://doi.org/10.1371/journal.pone.0190454)
- Graus M, Schnitzler JP, Hansel A, Cojocariu C, Rennenberg H, Wisthaler A, Kreuzwieser J (2004) Transient release of oxygenated volatile organic compounds during light-dark transitions in grey poplar leaves. Plant Physiol 135(4):1967–1975. [https://doi.](https://doi.org/10.1104/pp.104.043240) [org/10.1104/pp.104.043240](https://doi.org/10.1104/pp.104.043240)
- Kaissling KE, Thorson J (1980) Insect olfactory sensilla: structural, chemical and electrical aspects of the functional organisation. In: Satelle DB et al (eds) Receptors for neurotransmitters, hormones and pheromones in insects. Elsevier/North-Holland Biomedical Press, New York, pp 261–282
- Landolt PJ, Phillips TW (1997) Host plant infuences on sex pheromone behavior of phytophagous insects. Annu Rev Entomol 42:371–391.<https://doi.org/10.1146/annurev.ento.42.1.371>
- Li Y, Dickens JC, Steiner WWM (1992) Antennal olfactory responsiveness of *Microplitis croceipes* (Hymenoptera: Braconidae) to cotton plant volatiles. J Chem Ecol 18:1761–1773. [https://doi.org/](https://doi.org/10.1007/BF02751101) [10.1007/BF02751101](https://doi.org/10.1007/BF02751101)
- LeSage L, Dobesberger EJ, Majka CG (2008) Introduced leaf beetles of the maritime provinces, 6: the common asparagus beetle, *Crioceris asparagi* (L.), and the twelve-spotted asparagus beetle, *Crioceris duodecimpunctata* (L.) (Coleoptera: Chrysomelidae). P Entomol Soc Wash 110(3):602–621. [https://doi.org/10.4289/](https://doi.org/10.4289/07-075.1) [07-075.1](https://doi.org/10.4289/07-075.1)
- Loughrin JH, Potter DA, Hamilton-Kemp TR, Byers ME (1996) Role of feeding-induced plant volatiles in aggregative behavior of the japanese beetle (Coleoptera: Scarabaeidae). Environ Entomol 25(5):1188–1191.<https://doi.org/10.1093/ee/25.5.1188>
- Ma WC, Visser JH (1978) Single unit analysis of odour quality coding by the olfactory antennal receptor system of the Colorado beetle. Entomol Exp App 24(3):520–533. [https://doi.org/10.1111/j.1570-](https://doi.org/10.1111/j.1570-7458.1978.tb02813.x) [7458.1978.tb02813.x](https://doi.org/10.1111/j.1570-7458.1978.tb02813.x)
- McIndoo NE (1926) An insect olfactometer. J Econ Entomol 19(3):545–571.<https://doi.org/10.1093/jee/19.3.545>
- Morrison WR III, Szendrei Z (2014) The common asparagus beetle and spotted asparagus beetle (Coleoptera: Chrysomelidae): identifcation, ecology, and management. J Integr Pest Manag 5(3):B1–B6. <https://doi.org/10.1603/IPM14004>
- Morrison WR III, Ingrao A, Ali J, Szendrei Z (2016) Identifcation of plant semiochemicals and evaluation of their interactions with early spring insect pests of asparagus. J Plant Interact 11(1):11-19.<https://doi.org/10.1080/17429145.2015.1133848>
- Müller C, Hilker M (2000) The effect of a green leaf volatile on host plant fnding by larvae of a herbivorous insect. Naturwissenschaften 87(5):216–219.<https://doi.org/10.1007/s001140050706>
- Otter CD, Tchicaya T, Schutte AM (1991) Efects of age, sex and hunger on the antennal olfactory sensitivity of tsetse fies. Physiol Entomol 16(2):173–182. [https://doi.org/10.1111/j.1365-3032.](https://doi.org/10.1111/j.1365-3032.1991.tb00554.x) [1991.tb00554.x](https://doi.org/10.1111/j.1365-3032.1991.tb00554.x)
- Paré PW, Tumlinson JH (1999) Plant volatiles as a defense against insect herbivores. Plant Physiol 121(2):325–332. [https://doi.org/](https://doi.org/10.1104/pp.121.2.325) [10.1104/pp.121.2.325](https://doi.org/10.1104/pp.121.2.325)
- Pickett JA, Wadhams LJ, Woodcock CM (1998) Insect supersense: mate and host location by insect model systems for exploiting olfactory interactions. Biochemist 20:8–13
- Pollini A (1998) Manuale di entomologia applicata. Edagricole-Edizioni Agricole, Bologna
- Raguso RA, Light DM (1998) Electroantennogram responses of male Sphinx perelegans hawkmoths to floral and "green-leaf volatiles". Entomol Exp App 86(3):287–293. [https://doi.org/10.1046/j.1570-](https://doi.org/10.1046/j.1570-7458.1998.00291.x) [7458.1998.00291.x](https://doi.org/10.1046/j.1570-7458.1998.00291.x)
- Rao S, Cossé AA, Bartelt RJ, Zilkowski BW (2002) Field responses of cereal leaf beetle, *Oulema melanopus* (Coleoptera: Chrysomelidae) to its aggregation pheromone. In: Entomological Society of America annual meeting, Fort Lauderdale, 17-20 November 2002
- Reddy GV, Guerrero A (2004) Interactions of insect pheromones and plant semiochemicals. Trends Plant Sci 9(5):253–261. [https://doi.](https://doi.org/10.1016/j.tplants.2004.03.009) [org/10.1016/j.tplants.2004.03.009](https://doi.org/10.1016/j.tplants.2004.03.009)
- Ruther J, Reinecke A, Hilker M (2002) Plant volatiles in the sexual communication of *Melolontha hippocastani*: response towards time-dependent bouquets and novel function of (*Z*)-3-hexen-1-ol as a sexual kairomone. Ecol Entomol 27(1):76–83. [https://doi.org/](https://doi.org/10.1046/j.1365-2311.2002.0373a.x) [10.1046/j.1365-2311.2002.0373a.x](https://doi.org/10.1046/j.1365-2311.2002.0373a.x)
- Sun R, Wang Y, Chin CK, Garrison SA (2001) Volatile compounds in *Asparagus officinalis* L. in: X international Asparagus symposium, A. Uragami, Niigata, 589 pp 257–266. [https://doi.org/10.](https://doi.org/10.17660/ActaHortic.2002.589.35) [17660/ActaHortic.2002.589.35](https://doi.org/10.17660/ActaHortic.2002.589.35)
- Tasin M, Anfora G, Ioriatti C, Carlin S, De Cristofaro A, Schmidt S, Bengtsson M, Versini G, Witzgall P (2005) Antennal and

behavioral responses of grapevine moth *Lobesia botrana* females to volatiles from grapevine. J Chem Ecol 31(1):77–87. [https://doi.](https://doi.org/10.1007/s10886-005-0975-3) [org/10.1007/s10886-005-0975-3](https://doi.org/10.1007/s10886-005-0975-3)

- Thibout E, Pierre D, Mondy N, Lecomte C, Biemont JC, Auger J (2005) Host-plant fnding by the asparagus fy, *Plioreocepta poeciloptera* (Diptera: Tephritidae), a monophagous, monovoltine tephritid. B Entomol Res 95:393–399. [https://doi.org/10.1079/](https://doi.org/10.1079/BER2005370) [BER2005370](https://doi.org/10.1079/BER2005370)
- Thorpe WH, Crombie AC, Hill R, Darrah JH (1947) The behaviour of wireworms in response to chemical stimulation. J Exp Biol 23(3-4):234–266
- Turlings TC, Lengwiler UB, Bernasconi ML, Wechsler D (1998) Timing of induced volatile emissions in maize seedlings. Planta 207(1):146–152. <https://doi.org/10.1007/s004250050466>
- Van den Berg J, Torto B, Pickett JA, Smart LE, Wadhams LJ, Woodcock CM (2008) Infuence of visual and olfactory cues on feld trapping of the pollen beetle, *Astylus atromaculatus* (Coleoptera: Melyridae). J Appl Entomol 132(6):490–496. [https://doi.org/10.](https://doi.org/10.1111/j.1439-0418.2007.01259.x) [1111/j.1439-0418.2007.01259.x](https://doi.org/10.1111/j.1439-0418.2007.01259.x)
- Visser JH (1979) Electroantennogram responses of the Colorado beetle, *Leptinotarsa decemlineata*, to plant volatiles. Entomol Exp App 25(1):86–97.<https://doi.org/10.1111/j.1570-7458.1979.tb02851.x>
- Visser JH (1986) Host odour reception in phytophagous insects. Annu Rev Entomol 31:121–144. [https://doi.org/10.1146/annurev.en.31.](https://doi.org/10.1146/annurev.en.31.010186.001005) [010186.001005](https://doi.org/10.1146/annurev.en.31.010186.001005)
- Wei J, Kang L (2011) Roles of (*Z*)-3-hexenol in plant-insect interactions. Plant Signal Behav 6(3):369–371. [https://doi.org/10.4161/](https://doi.org/10.4161/psb.6.3.14452) [psb.6.3.14452](https://doi.org/10.4161/psb.6.3.14452)
- Weissbecker B, Schütz S, Klein A, Hummel HE (1997) Analysis of volatiles emitted by potato plants by means of a Colorado beetle electroantennographic detector. Talanta 44(12):2217–2224. [https://doi.org/10.1016/S0039-9140\(97\)00037-4](https://doi.org/10.1016/S0039-9140(97)00037-4)