

# Cancer-Related Inflammation

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**Abstract** Solid tumors consist of neoplastic cells, non-malignant stromal cells, and migratory hematopoietic cells. Complex interactions between the cell types in this microenvironment regulate tumor growth, progression, metastasis, and angiogenesis. The cells and mediators of inflammation form a major part of the epithelial tumor microenvironment. In some cancers, inflammatory conditions precede development of malignancy; in others, oncogenic change drives a tumor-promoting inflammatory milieu. Whatever its origin, this “smoldering” inflammation aids proliferation and survival of malignant cells, stimulates angiogenesis and metastasis, subverts adaptive immunity, and alters response to hormones and chemotherapy. Cytokines are major mediators of communication between cells in the inflammatory tumor microenvironment. It is known that neoplastic cells often over-express proinflammatory mediators including proteases, eicosanoids, cytokines, and chemokines. Several cytokines such as macrophage migratory inhibitory factor (MIF), TNF- $\alpha$ , IL-6, IL-17, IL-12, IL-23, IL-10, and TGF- $\beta$  have been linked with both experimental and human cancers and can either promote or inhibit tumor development. MIF is a major cytokine in many cancers and there is evidence that the cytokine is produced by both malignant cells and infiltrating leukocytes. In this article we will discuss the role of cancer-associated inflammation and the particular role of MIF in malignant disease.

**Keywords** Cytokines · cancer · malignant disease · inflammation · MIF · tumorigenesis

## Introduction

Cytokines are mediators that regulate a broad range of processes involved in the pathogenesis of cancer. Inflammation is key to the integrity and survival of multicellular organisms, but deregulation of this powerful component of the immune system is a characteristic of much chronic pathology, including cancer [1, 2]. Inflammatory processes can promote, or maybe even initiate, malignant disease [3–6].

## Cancer and Inflammation

Cancer-related inflammation is an essential process in malignant disease, with common and defined players at different stages of progression [3, 7]. Until recently, the field has been driven by the hypothesis that extrinsic inflammatory pathways promote or, in some cases, initiate cancer—i.e., that inflammation causes or promotes cancer [1] (Table 1). However there is now evidence that there is an intrinsic inflammation pathway activated by genetic events that cause neoplasia; i.e., cancer causes inflammation [7–13]. Activation of oncogenes such as *myc*, *ras*, and *ret*, or inactivation of tumor suppressors such as pVHL, leads to constitutive production of inflammatory cytokines by the initiated cell. Oncogene and tumor suppressor pathways are proven intracellular targets for therapies, but these recent data mean that inflammatory cytokines and their receptors are druggable extracellular targets of the genetic changes in malignant disease. The cells and mediators of inflammation also form a major part of the tumor microenvironment (Fig. 1). In some cancers, inflammatory conditions precede development of malignancy; in others, oncogenic changes drive a tumor-promoting inflammatory milieu. Whatever its origin, this “smoldering” inflammation aids proliferation and survival of malignant cells, angiogenesis, and metastasis; subverts

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**Table I** The association between inflammation and cancer in different organ systems

System	Inflammation
Respiratory	Asbestos and mesothelioma Silica, cigarette smoke and bronchial cancer Chronic asthma and bronchial cancer
Genitourinary	Human papilloma virus and penile carcinoma Schistosomiasis and bladder cancer
Reproductive	Pelvic inflammatory disease, ovarian epithelial inflammation and ovarian cancer
Gastrointestinal	Epstein-Barr virus and nasopharyngeal cancer Barrett's metaplasia and esophageal cancer <i>H. pylori</i> gastritis and gastric cancer Chronic pancreatitis and pancreatic cancer Chronic cholecystitis and gallbladder cancer Hepatitis and hepatocellular cancer Inflammatory bowel disease and colorectal cancer Human papilloma virus and anogenital cancer
Hematopoietic	Epstein-Barr virus and Burkitt's lymphoma HTLV1 and T cell leukemia/lymphoma

adaptive immunity; and alters response to hormones and chemotherapeutic agents [14, 15]. The cytokine network is of great importance in the processes of cancer-related inflammation, regulating both host and malignant cells in the tumor microenvironment [16]. The epidemiological data available are impressive and show a clear association between chronic inflammatory conditions and subsequent malignant transformation in the inflamed tissue.

### Cytokines in Inflammation and Cancer

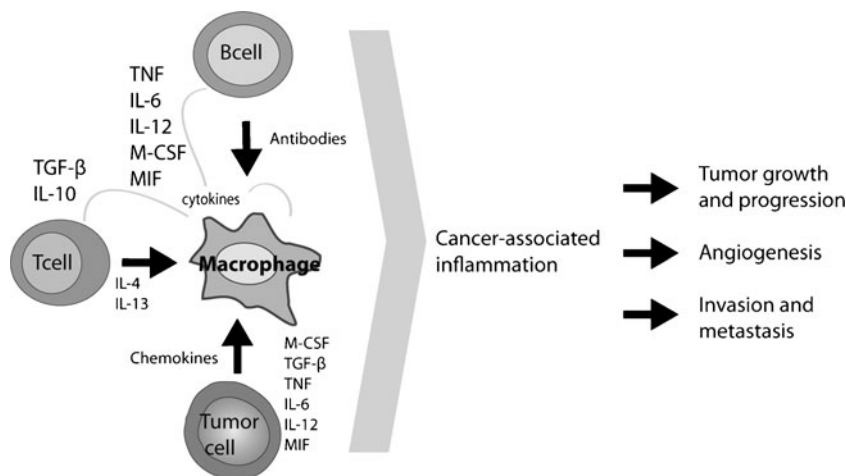
It is known that neoplastic cells often over-express proinflammatory mediators, including proteases, eicosanoids, cytokines,

and chemokines. Several cytokines, such as tumor necrosis factor (TNF)- $\alpha$ , interleukin (IL)-6, IL-17, IL-12, IL-23, IL-10, transforming growth factor (TGF)- $\beta$ , and macrophage migration inhibitory factor (MIF) have been linked with both experimental and human cancers and can either promote or inhibit tumor development (Table II). Here we will describe some of the cytokines that may be potential targets for cancer therapy.

TNF- $\alpha$  is the prototypical proinflammatory cytokine. The critical role of TNF- $\alpha$  in chronic inflammatory diseases is well known [36]. Although originally shown to be toxic to tumor cells in high doses, the tumor-promoting function of TNF- $\alpha$  has been clearly demonstrated in mice [37]. Malignant cells or inflammatory cells in the tumor microenvironment can produce TNF- $\alpha$ , and TNF- $\alpha$  signaling can promote cell survival, invasion, and angiogenesis [38]. In mesothelioma, phagocytosis of asbestos fibers by myeloid cells leads to TNF- $\alpha$  secretion that promotes cell survival and thereby reduces asbestos-induced cytotoxicity [13]. TNF- $\alpha$  has also been shown to contribute to tumor initiation by stimulation of genotoxic reactive nitrogen (RNS) and oxygen (ROS) species [39]. Genetic polymorphisms that enhance TNF- $\alpha$  production are associated with increased risk of multiple myeloma (MM), bladder cancer, hepatocellular carcinoma (HCC), gastric cancer, and breast cancer, as well as poor prognosis in various hematological malignancies [40]. Other actions of TNF- $\alpha$  that may promote tumorigenesis include angiogenesis, impairment of immune surveillance through T-cell suppression, and inhibition of the cytotoxic activity of activated macrophages [41].

TNF- $\alpha$  binds to 2 receptors—the ubiquitously expressed TNFR1 and TNFR2, which is restricted to expression on hematopoietic cells. Previous studies show that TNFR1 is important in tumor promotion [42]. TNFR1 knockout mice are resistant to skin carcinogenesis [44] and experimental lung metastases in the renal cancer (RENCA) model [43]. Experimental liver metastases were attenuated in TNFR1-deficient mice [44]. In chimeric mice whose bone marrow

**Fig. 1** Simplified overview of the interaction of cytokines in the tumour microenvironment. Several cell types contribute by producing these cytokines and their action will unfold on the surrounding cells. As a net result the cytosine milieu contributes to promotion of several key hallmarks of cancer, tumour growth and progression, angiogenesis and invasion/metastasis



**Table II** Cancer-associated cytokines

Target	Disease
TNF- $\alpha$	Ovarian cancer [17], pancreatic cancer [18]
IL-4	Lung cancer [19] Breast cancer [20]
IL-6	Pancreatic cancer [21, 22] Breast cancer [23]
IL-11	Gastric cancer [24] Breast cancer [25] Pancreatic cancer
IL-13	Ovarian cancer [26] Head and neck [27] Glioma [28]
IL-17	Ovarian cancer [17] [29] Bladder cancer [30] Melanoma [30] Colon Cancer [31]
MIF	CLL [32] Breast cancer [33] Colon cancer [34] Prostate cancer [35]

was repopulated with TNFR1 $^{-/-}$  cells, development of colitis and colon cancer was attenuated [45]. Several studies have demonstrated that stromal cell TNF- $\alpha$  is tumor promoting. In a genetic model, bone marrow cell TNF- $\alpha$  was implicated in promotion of inflammation-associated liver tumors [6]. In a model where chemical damage led to liver cancer, Kupffer cell TNF- $\alpha$  was one of the mitogens driving proliferation of genetically damaged hepatocytes [46]. In a chemically induced model of colitis and colorectal cancer, mononuclear cell TNF- $\alpha$  was implicated in inflammation and subsequent tumor development [45]. Several studies have reported therapeutic activity of anti-TNF- $\alpha$  antibodies, or a TNF receptor fusion molecule, in genetic models of liver and colorectal cancer [6, 47]; a carcinogen-induced model of colorectal cancer [45]; and pancreatic cancer xenografts [48]—although the exact mechanisms of action are not understood.

In many different experimental and human cancers, malignant cells produce TNF- $\alpha$  during tumor growth and spread [10, 37, 38, 48–53]. Preclinical experiments with TNF- $\alpha$  antagonists and early-phase clinical trials of TNF- $\alpha$  antagonists in patients with advanced cancer suggest that this inflammatory cytokine may be a useful target [49, 54–56].

IL-6, a pleiotropic inflammatory cytokine, is considered a key growth factor for both malignant and immune cells. Most IL-6 target genes are involved in cell cycle progression and suppression of apoptosis, which underscores the importance of IL-6 in cell survival and tumorigenesis. IL-6 is suggested to have a pivotal role in the pathogenesis of Castleman’s disease [57] and MM [58]. Clear evidence that IL-6 governs the

growth of MM, a malignant disorder of plasma cells, has come from studies using IL-6 knockout mice, which were found to be resistant to plasmacytoma induction [58]. In MM, stromal cells in the bone marrow produce IL-6. Its synthesis by these cells can be further enhanced by their interaction with malignant plasma cells. New IL-6 antagonists such as small molecules and monoclonal antibodies are being evaluated for treatment of MM. IL-6 is also a key mediator of inflammatory disease. IBD and colitis are associated with high concentrations of IL-6 [59]. In experimental models of colitis-associated colon cancer (CAC), HCC IL-6 production by myeloid cells is also critical for carcinogenesis [4, 60, 61].

IL-10 is an immunosuppressive and anti-inflammatory cytokine also linked with inflammation-associated cancer [7]. IL-10-deficient mice develop spontaneous colitis due to hyperactivation of immune cells, and eventually they develop CAC. Expression of IL-10 by tumor cells and macrophages is thought to promote the development of Burkitt’s lymphoma through the production of the TNF family member B-cell activating factor (BAFF), which promotes B-cell and lymphoma survival [62]. An elevated amount of IL-10 in the plasma has been correlated with poor prognosis in diffuse large B-cell lymphoma patients [63]. In addition to direct growth modulation of malignant cells, the ability of IL-10 to suppress adaptive immune responses has been suggested to favor tumor escape from immune surveillance [64]. Therefore, IL-10 has complex effects on tumor development. In some experimental systems, IL-10 is found to exert antitumor activity, while in other cases it can promote tumorigenesis.

MIF was one of the first cytokine activities to be described [65, 66], originally being identified as a product of activated T lymphocytes that inhibited the random migration of cultured macrophages. It is now clear that MIF is a key regulator of immune and inflammatory responses and is produced by a range of cells and tissues. Subsequent studies of MIF

**Table III** Mode of action for MIF in tumor development and progression

Function	Role for development and progression
P53 inhibition	Tumour heterogeneity [85] Inhibition of apoptosis [78] Promotion of proliferation [86]
MAPK activation	Invasion [79] Block of apoptosis [79] Chemoresistance [86] Pro-angiogenic [34]
COX2 pathway	Direct tumor growth promotion Invasion and metastasis [79] Pro-angiogenic [34]
Immune escape	MDSC maintenance [86] T cell inhibition [87]

expression *in vivo* have established an important role in host response to endotoxic shock [67] and the inflammatory pathologies responsible for arthritis [68]. MIF is a key inducer of inflammatory cytokines such as TNF- $\alpha$  and IL-1 [67], and macrophages from MIF knockout mice have a severely diminished TNF- $\alpha$  response to bacterial endotoxin both *in vitro* and *in vivo* [69]. Recent studies describe the existence of a MIF-glucocorticoid counter-regulatory system that controls inflammation and immune response [70], and released MIF can supersede the glucocorticoid immunosuppressive effects [71]. MIF binds to the extracellular domain of CD74—the cell-surface form of the MHC class-II-associated invariant chain [72]—but needs CD44 as an integral member of the CD74 receptor complex leading to MIF signal transduction [73].

MIF expression is increased during the evolution of several malignancies [74]. MIF also has been implicated in the angiogenic switch of early cancer [75]. We reported earlier a role for MIF in macrophage-induced ovarian cancer cell invasiveness [76] *in vitro* and demonstrated that the inhibition of ovarian cancer-cell-derived MIF expression by MIF siRNA inhibits tumor growth, progression, and neo-angiogenesis, while increasing survival rates [77]. MIF also has the ability to protect tumor cells from apoptosis [77, 78]. Recently, Bernhagen et al. [79] showed for the first time that MIF is a potential ligand for the chemokine receptors CXCR2 and CXCR4, both of which have been shown to play a major role in ovarian cancer [38, 80–84]. Table III provides an overview of the mode of action for MIF.

The role of inflammation in tumor development is likely to depend on the nature of the tumor and inflammatory cell interaction. While inflammatory cells may produce growth factors for tumor cells, in turn the tumor microenvironment may provide factors that suppress antitumor immune responses. Identifying the mechanisms by which inflammation is deregulated in cancer and provides tumor-promoting signals may offer new therapeutic opportunities in cancer therapy. One caveat for cytokines as targets in cancer therapy: although inflammation is clearly linked with tumorigenesis, both the innate and adaptive immune systems have the capacity to recognize and eliminate malignant cells [88, 89]. Many proinflammatory cytokines may function in tumor immune surveillance, and it is vital to determine whether this mechanism offers potential as a therapeutic target.

**Conflicts of interest** The authors declare that they have no conflict of interest.

## References

- Balkwill F, Mantovani A. Inflammation and cancer: back to Virchow? *Lancet*. 2001;357(9255):539–45.
- Coussens LM, Werb Z. Inflammatory cells and cancer: think different! *J Exp Med*. 2001;193(6):F23–6.
- Balkwill F, Charles KA, Mantovani A. Smoldering and polarized inflammation in the initiation and promotion of malignant disease. *Cancer Cell*. 2005;7(3):211–7.
- Greten FR, Eckmann L, Greten TF, Park JM, Li ZW, Egan LJ, et al. IKKbeta links inflammation and tumorigenesis in a mouse model of colitis-associated cancer. *Cell*. 2004;118(3):285–96.
- Houghton J, Stoicov C, Nomura S, Rogers AB, Carlson J, Li H, et al. Gastric cancer originating from bone marrow-derived cells. *Science*. 2004;306(5701):1568–71.
- Pikarsky E, Porat RM, Stein I, Abramovitch R, Amit S, Kasem S, et al. NF-kappaB functions as a tumour promoter in inflammation-associated cancer. *Nature*. 2004;431(7007):461–6.
- Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. *Nature*. 2008;454(7203):436–44.
- Ancrile B, Lim KH, Counter CM. Oncogenic Ras-induced secretion of IL6 is required for tumorigenesis. *Genes Dev*. 2007;21(14):1714–9.
- Borrello MG, Alberti L, Fischer A, Degl'innocenti D, Ferrario C, Gariboldi M, et al. Induction of a proinflammatory program in normal human thyrocytes by the RET/PTC1 oncogene. *Proc Natl Acad Sci U S A*. 2005;102(41):14825–30.
- Galban S, Fan J, Martindale JL, Cheadle C, Hoffman B, Woods MP, et al. von Hippel-Lindau protein-mediated repression of tumor necrosis factor alpha translation revealed through use of cDNA arrays. *Mol Cell Biol*. 2003;23(7):2316–28.
- Mantovani A, Schioppa T, Porta C, Allavena P, Sica A. Role of tumor-associated macrophages in tumor progression and invasion. *Cancer Metastasis Rev*. 2006;25(3):315–22.
- Soucek L, Lawlor ER, Soto D, Shchors K, Swigart LB, Evan GI. Mast cells are required for angiogenesis and macroscopic expansion of Myc-induced pancreatic islet tumors. *Nat Med*. 2007;13(10):1211–8.
- Yang H, Bocchetta M, Kroczyńska B, Elmishad AG, Chen Y, Liu Z, et al. TNF-alpha inhibits asbestos-induced cytotoxicity via a NF-kappaB-dependent pathway, a possible mechanism for asbestos-induced oncogenesis. *Proc Natl Acad Sci U S A*. 2006;103(27):10397–402.
- Balkwill F. Immunology for the next generation. *Nature Rev Immunol*. 2005;5(6):509–12.
- Bonecchi R, Borroni EM, Anselmo A, Doni A, Savino B, Mirolo M, et al. Regulation of D6 chemokine scavenging activity by ligand- and Rab11-dependent surface up-regulation. *Blood*. 2008;112(3):493–503.
- Balkwill F, Coussens LM. Cancer: an inflammatory link. *Nature*. 2004;431(7007):405–6.
- Charles KA, Kulbe H, Soper R, Escorcio-Correia M, Lawrence T, Schultheis A, et al. The tumor-promoting actions of TNF-alpha involve TNFR1 and IL-17 in ovarian cancer in mice and humans. *J Clin Invest*. 2009;119:3011–23.
- Maniati E, Bossard M, Cook N, Candido JB, Emami-Shahri N, Nedospasov SA, et al. Crosstalk between the canonical NF-kappaB and Notch signaling pathways inhibits Ppargamma expression and promotes pancreatic cancer progression in mice. *J Clin Invest*. 2011;121:4685–99.
- Michels CE, Scales HE, Saunders KA, McGowan S, Brombracher F, Alexander J, et al. Neither interleukin-4 receptor alpha expression on CD4+ T cells, or macrophages and neutrophils is required for protective immunity to *Trichinella spiralis*. *Immunology*. 2009;128:e385–94.
- DeNardo DG, Barreto JB, Andreu P, Vasquez L, Tawfik D, Kolhatkar N, et al. CD4(+) T cells regulate pulmonary metastasis of mammary carcinomas by enhancing protumor properties of macrophages. *Cancer Cell*. 2009;16:91–102.
- Lesina M, Kurkowski MU, Ludes K, Rose-John S, Treiber M, Kloppel G, et al. Stat3/Socs3 activation by IL-6 transsignaling

- promotes progression of pancreatic intraepithelial neoplasia and development of pancreatic cancer. *Cancer Cell*. 2011;19:456–69.
22. Bellone G, Smirne C, Mauri FA, Tonel E, Carbone A, Buffolino A, et al. Cytokine expression profile in human pancreatic carcinoma cells and in surgical specimens: implications for survival. *Cancer Immunol Immunother*. 2006;55:684–98.
  23. Korkaya H, Kim GI, Davis A, Malik F, Henry NL, Ithimakin S, et al. Activation of an IL6 inflammatory loop mediates trastuzumab resistance in HER2+ breast cancer by expanding the cancer stem cell population. *Mol Cell*. 2012;47:570–84.
  24. Howlett M, Giraud AS, Lescesen H, Jackson CB, Kalantzis A, Van Driel IR, et al. The interleukin-6 family cytokine interleukin-11 regulates homeostatic epithelial cell turnover and promotes gastric tumor development. *Gastroenterology*. 2009;136:967–77.
  25. Hanavadi S, Martin TA, Watkins G, Mansel RE, Jiang WH. Expression of interleukin 11 and its receptor and their prognostic value in human breast cancer. *Ann Surg Oncol*. 2006;13:802–8.
  26. Fujisawa T, Joshi BH, Puri RK. IL-13 regulates cancer invasion and metastasis through IL-13Ralpha2 via ERK/AP-1 pathway in mouse model of human ovarian cancer. *Int J Cancer*. 2012;131:344–56.
  27. Kawakami M, Kawakami K, Kasperbauer JL, Hinkley LL, Tsukuda M, Strome SE, et al. Interleukin-13 receptor alpha2 chain in human head and neck cancer serves as a unique diagnostic marker. *Clin Cancer Res*. 2003;9:6381–8.
  28. Kioi M, Kawakami M, Shimamura T, Husain SR, Puri RK. Interleukin-13 receptor alpha2 chain: a potential biomarker and molecular target for ovarian cancer therapy. *Cancer*. 2006;107:1407–18.
  29. Kryczek I, Wei S, Szeliga W, Vatan L, Zou W. Endogenous IL-17 contributes to reduced tumor growth and metastasis. *Blood*. 2009;114:357–9.
  30. Wang L, Yi T, Kortylewski M, Pardoll DM, Zeng D, Yu H. IL-17 can promote tumor growth through an IL-6-Stat3 signaling pathway. *J Exp Med*. 2009;206:1457–64.
  31. Grivennikov SI, Wang K, Mucida D, Stewart CA, Schnabl B, Jauch D, et al. Adenoma-linked barrier defects and microbial products drive IL-23/IL-17-mediated tumour growth. *Nature*. 2012;491:254–8.
  32. Reinart N, Nguyen PH, Boucas J, Rosen N, Kvasnicka HM, Heukamp L, et al. Delayed development of chronic lymphocytic leukemia in the absence of macrophage migration inhibitory factor. *Blood*. 2012 [epub ahead of print].
  33. Verjans E, Noetzel E, Bektas N, Schutz AK, Lue H, Lennartz B, et al. Dual role of macrophage migration inhibitory factor (MIF) in human breast cancer. *BMC Cancer*. 2009;9:230.
  34. Wilson JM, Coletta PL, Cuthbert RJ, Scott N, MacLennan K, Hawcroft G, et al. Macrophage migration inhibitory factor promotes intestinal tumorigenesis. *Gastroenterology*. 2005;129:1485–503.
  35. Meyer-Siegler KL, Iczkowski KA, Leng L, Bucala R, Vera PL. Inhibition of macrophage migration inhibitory factor or its receptor (CD74) attenuates growth and invasion of DU-145 prostate cancer cells. *J Immunol*. 2006;177:8730–9.
  36. Feldmann M, Maini SR. Role of cytokines in rheumatoid arthritis: an education in pathophysiology and therapeutics. *Immunol Rev*. 2008;223(1):7–19.
  37. Moore RJ, Owens DM, Stamp G, Arnott C, Burke F, East N, et al. Mice deficient in tumor necrosis factor-alpha are resistant to skin carcinogenesis. *Nat Med*. 1999;5(7):828–31.
  38. Kulbe H, Thompson R, Wilson JL, Robinson S, Hagemann T, Fatah R, et al. The inflammatory cytokine tumor necrosis factor-alpha generates an autocrine tumor-promoting network in epithelial ovarian cancer cells. *Cancer Res*. 2007;67(2):585–92.
  39. Hussain SP, Hofseth LJ, Harris CC. Radical causes of cancer. *Nat Rev Cancer*. 2003;3(4):276–85.
  40. Mocellin S, Rossi CR, Pilati P, Nitti D. Tumor necrosis factor, cancer and anticancer therapy. *Cytokine Growth Factor Rev*. 2005;16(1):35–53.
  41. Elgert KD, Alleva DG, Mullins DW. Tumor-induced immune dysfunction: the macrophage connection. *J Leukoc Biol*. 1998;64(3):275–90.
  42. Arnott CH, Scott KA, Moore RJ, Robinson SC, Thompson RG, Balkwill FR. Expression of both TNF-alpha receptor subtypes is essential for optimal skin tumour development. *Oncogene*. 2004;23(10):1902–10.
  43. Tomita Y, Yang X, Ishida Y, Nemoto-Sasaki Y, Kondo T, Oda M, et al. *Int J Cancer*. 2004;112(6):927–33.
  44. Kitakata H, Nemoto-Sasaki Y, Takahashi Y, Kondo T, Mai M, Mukaida N. Essential roles of tumor necrosis factor receptor p55 in liver metastasis of intrasplenic administration of colon 26 cells. *Cancer Res*. 2002;62(22):6682–7.
  45. Popivanova BK, Kitamura K, Wu Y, Kondo T, Kagaya T, Kaneko S, et al. Blocking TNF-alpha in mice reduces colorectal carcinogenesis associated with chronic colitis. *J Clin Invest*. 2008;118(2):560–70.
  46. Maeda S, Kamata H, Luo JL, Leffert H, Karin M. IKKbeta couples hepatocyte death to cytokine-driven compensatory proliferation that promotes chemical hepatocarcinogenesis. *Cell*. 2005;121(7):977–90.
  47. Rao VP, Poutahidis T, Ge Z, Nambiar PR, Boussahmain C, Wang YY, et al. Innate immune inflammatory response against enteric bacteria *Helicobacter hepaticus* induces mammary adenocarcinoma in mice. *Cancer Res*. 2006;66(15):7395–400.
  48. Egberts JH, Cloosters V, Noack A, Schniewind B, Thon L, Klose S, et al. Anti-tumor necrosis factor therapy inhibits pancreatic tumor growth and metastasis. *Cancer Res*. 2008;68(5):1443–50.
  49. Harrison ML, Obermueller E, Maisey NR, Hoare S, Edmonds K, Li NF, et al. Tumor necrosis factor alpha as a new target for renal cell carcinoma: two sequential phase II trials of infliximab at standard and high dose. *J Clin Oncol*. 2007;25(29):4542–9.
  50. Naylor MS, Stamp GWH, Foulkes WD, Eccles D, Balkwill FR. Tumor necrosis factor and its receptors in human ovarian cancer. *J Clin Invest*. 1993;91(5):2194–206.
  51. Petersen SL, Wang L, Yalcin-Chin A, Li L, Peyton M, Minna J, et al. Autocrine TNFalpha signaling renders human cancer cells susceptible to Smac-mimetic-induced apoptosis. *Cancer Cell*. 2007;12(5):445–56.
  52. Stathopoulos GT, Kollintza A, Moschos C, Psallidas I, Sherrill TP, Pitsinos EN, et al. Tumor necrosis factor-alpha promotes malignant pleural effusion. *Cancer Res*. 2007;67(20):9825–34.
  53. Zins K, Abraham D, Sioud M, Aharinejad S. Colon cancer cell-derived tumor necrosis factor-alpha mediates the tumor growth-promoting response in macrophages by up-regulating the colony-stimulating factor-1 pathway. *Cancer Res*. 2007;67(3):1038–45.
  54. Brown ER, Charles KA, Hoare SA, Rye RL, Jodrell DI, Aird RE, et al. A clinical study assessing the tolerability and biological effects of infliximab, a TNF-alpha inhibitor, in patients with advanced cancer. *Ann Oncol*. 2008;19(7):1340–6.
  55. Madhusudan S, Foster M, Braybrooke J, Muthuramalingam SR, Wilner S, Kaur K, et al. A phase II study of Etanercept (Enbrel), a tumor necrosis factor-alpha inhibitor in patients with metastatic breast cancer. *Clin Cancer Res*. 2004;10(19):6528–34.
  56. Madhusudan S, Muthuramalingam SR, Braybrooke JP, Wilner S, Kaur K, Han C, et al. Study of etanercept, a tumor necrosis factor-alpha inhibitor, in recurrent ovarian cancer. *J Clin Oncol*. 2005;23(25):5950–9.
  57. Screpanti I, Musiani P, Bellavia D, Cappelletti M, Aiello FB, Maroder M, et al. Inactivation of the IL-6 gene prevents development of multicentric Castleman's disease in C/EBP beta-deficient mice. *J Exp Med*. 1996;184(4):1561–6.

58. Bommert K, Bargou RC, Stuhmer T. Signalling and survival pathways in multiple myeloma. *Eur J Cancer*. 2006;42(11):1574–80.
59. Mudter J, Amoussina L, Schenk M, Yu J, Brustle A, Weigmann B, et al. The transcription factor IFN regulatory factor-4 controls experimental colitis in mice via T cell-derived IL-6. *J Clin Invest*. 2008;118(7):2415–26.
60. Naugler WE, Sakurai T, Kim S, Maeda S, Kim K, Elsharkawy AM, et al. Gender disparity in liver cancer due to sex differences in MyD88-dependent IL-6 production. *Science*. 2007;317(5834):121–4.
61. Weigmann B, Lehr HA, Yancopoulos G, Valenzuela D, Murphy A, Stevens S, et al. The transcription factor NFATc2 controls IL-6-dependent T cell activation in experimental colitis. *J Exp Med*. 2008;205(9):2099–110.
62. Ogden CA, Pound JD, Bath BK, Owens S, Johannessen I, Wood K, et al. Enhanced apoptotic cell clearance capacity and B cell survival factor production by IL-10-activated macrophages: implications for Burkitt's lymphoma. *J Immunol*. 2005;174:3015–23.
63. Lech-Maranda E, Bienvenu J, Michallet AS, Houot R, Robak T, Coiffier B, et al. Elevated IL-10 plasma levels correlate with poor prognosis in diffuse large B-cell lymphoma. *Eur Cytokine Netw*. 2006;17(1):60–6.
64. Mocellin S, Marincola FM, Young HA. Interleukin-10 and the immune response against cancer: a counterpoint. *J Leukoc Biol*. 2005;78(5):1043–51.
65. Bloom BR, Bennett B. Mechanism of a reaction in vitro associated with delayed-type hypersensitivity. *Science*. 1966;153(3731):80–2.
66. David JR. Delayed hypersensitivity in vitro: its mediation by cell-free substances formed by lymphoid cell-antigen interaction. *Proc Natl Acad Sci U S A*. 1966;56(1):72–7.
67. Calandra T, Echtenacher B, Roy DL, Pugin J, Metz CN, Hultner L, et al. Protection from septic shock by neutralization of macrophage migration inhibitory factor. *Nat Med*. 2000;6(2):164–70.
68. Leech MC, Metz C, Santos L, Peng T, Holdsworth SR, Bucala R, et al. Involvement of macrophage migration inhibitory factor in the evolution of rat adjuvant arthritis. *Arthritis Rheum*. 1998;41(5):910–7.
69. Calandra T, Roger T. Macrophage migration inhibitory factor: a regulator of innate immunity. *Nat Rev Immunol*. 2003;3(10):791–800.
70. Calandra T, Bernhagen J, Metz CN, Spiegel LA, Bacher M, Donnelly T, et al. MIF as a glucocorticoid-induced modulator of cytokine production. *Nature*. 1995;377(6544):68–71.
71. Bacher M, Metz CN, Calandra T, Mayer K, Chesney J, Lohoff M, et al. An essential regulatory role for macrophage migration inhibitory factor in T-cell activation. *Proc Natl Acad Sci U S A*. 1996;93(15):7849–54.
72. Leng L, Metz CN, Fang Y, Xu J, Donnelly S, Baugh J, et al. MIF signal transduction initiated by binding to CD74. *J Exp Med*. 2003;197(11):1467–76.
73. Shi X, Leng L, Wang T, Wang W, Du X, Li J, et al. CD44 is the signaling component of the macrophage migration inhibitory factor-CD74 receptor complex. *Immunity*. 2006;25(4):595–606.
74. Mitchell RA. Mechanisms and effectors of MIF-dependent promotion of tumorigenesis. *Cell Signal*. 2004;16(1):13–9.
75. Wilson JM, Coletta PL, Cuthbert RJ, Scott N, MacLennan K, Hawcroft G, et al. Macrophage migration inhibitory factor promotes intestinal tumorigenesis. *Gastroenterology*. 2005;129(5):1485–503.
76. Hagemann T, Wilson J, Kulbe H, Li NFF, Leinster DA, Charles K, et al. Macrophages induce invasiveness of epithelial cancer cells via NF-kappa B and JNK. *J Immunol*. 2005;175(2):1197–205.
77. Hagemann T, Robinson SC, Thompson R, Charles KA, Kulbe H, Balkwill FR. Ovarian cancer cell-derived MIF enhances tumor growth, progression and angiogenesis. *Mol Cancer Ther*. 2007;6(7):1–10.
78. Hudson JD, Shoaibi MA, Maestro R, Carnero A, Hannon GJ, Beach DH. A proinflammatory cytokine inhibits p53 tumor suppressor activity. *J Exp Med*. 1999;190:1375–82.
79. Bernhagen J, Krohn R, Lue H, Gregory JL, Zerneck A, Koenen RR, et al. MIF is a noncognate ligand of CXC chemokine receptors in inflammatory and atherogenic cell recruitment. *Nat Med*. 2007;13:587–96.
80. Kulbe H, Hagemann T, Szlosarek PW, Balkwill FR, Wilson JL. The inflammatory cytokine tumor necrosis factor-alpha regulates chemokine receptor expression on ovarian cancer cells. *Cancer Res*. 2005;65(22):10355–62.
81. Scotton C, Milliken D, Wilson J, Raju S, Balkwill F. Analysis of CC chemokine and chemokine receptor expression in solid ovarian tumours. *Br J Cancer*. 2001;85(6):891–7.
82. Scotton CJ, Wilson JL, Milliken D, Stamp G, Balkwill FR. Epithelial cancer cell migration: a role for chemokine receptors? *Cancer Res*. 2001;61(13):4961–5.
83. Scotton CJ, Wilson JL, Scott K, Stamp G, Wilbanks GD, Fricker S, et al. Multiple actions of the chemokine CXCL12 on epithelial tumor cells in human ovarian cancer. *Cancer Res*. 2002;62(20):5930–8.
84. Venkatakrisnan G, Salgia R, Groopman JE. Chemokine receptors CXCR-1/2 activate mitogen-activated protein kinase via the epidermal growth factor receptor in ovarian cancer cells. *J Biol Chem*. 2000;275(10):6868–75.
85. Fingerle-Rowson G, Petrenko O, Metz CN, Forsthuber TG, Mitchell R, Huss R, et al. The p53-dependent effects of macrophage migration inhibitory factor revealed by gene targeting. *Proc Natl Acad Sci U S A*. 2003;100:9354–9.
86. Simpson KD, Templeton DJ, Cross JV. Macrophage migration inhibitory factor promotes tumor growth and metastasis by inducing myeloid-derived suppressor cells in the tumor microenvironment. *J Immunol*. 2012 [Epub ahead of print].
87. Gabbitass RF, Annels NE, Stocken DD, Pandha HA, Middleton GW. Elevated myeloid-derived suppressor cells in pancreatic, esophageal and gastric cancer are an independent prognostic factor and are associated with significant elevation of the Th2 cytokine interleukin-13. *Cancer Immunol Immunother*. 2011;60:1419–30.
88. Dranoff G. Cytokines in cancer pathogenesis and cancer therapy. *Nat Rev Cancer*. 2004;4(1):11–22.
89. Lawrence T, Hagemann T, Balkwill F. Cancer. Sex, cytokines, and cancer. *Science*. 2007;317(5834):51–2.