

# Preparation and optimization of calcium fluoride particles for dental applications

Joachim Koeser · Thiago Saads Carvalho ·  
Uwe Pieleles · Adrian Lussi

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**Abstract** Fluorides are used in dental care due to their beneficial effect in tooth enamel de-/remineralization cycles. To achieve a desired constant supply of soluble fluorides in the oral cavity, different approaches have been followed. Here we present results on the preparation of  $\text{CaF}_2$  particles and their characterization with respect to a potential application as enamel associated fluoride releasing reservoirs.  $\text{CaF}_2$  particles were synthesized by precipitation from soluble  $\text{NaF}$  and  $\text{CaCl}_2$  salt solutions of defined concentrations and their morphology analyzed by scanning electron microscopy.  $\text{CaF}_2$  particles with defined sizes and shapes could be synthesized by adjusting the concentrations of the precursor salt solutions. Such particles interacted with enamel surfaces when applied at fluoride concentrations correlating to typical dental care products. Fluoride release from the synthesized  $\text{CaF}_2$  particles was observed to be largely influenced by the concentration of phosphate in the solution. Physiological solutions with phosphate concentration similar to saliva (3.5 mM) reduced the fluoride release from pure  $\text{CaF}_2$  particles by a factor of 10–20  $\times$  as compared to phosphate free buffer solutions. Fluoride release was even lower in human saliva. The fluoride release could be increased by the addition of phosphate in substoichiometric amounts during  $\text{CaF}_2$  particle synthesis. The presented results

demonstrate that the morphology and fluoride release characteristics of  $\text{CaF}_2$  particles can be tuned and provide evidence of the suitability of synthetic  $\text{CaF}_2$  particles as enamel associated fluoride reservoirs.

## 1 Introduction

Fluoride is well known for its beneficial effect on tooth de-/remineralization cycles. To keep the levels of fluoride on the enamel surface sufficiently high during the intervals between the application cycles of fluoride containing dentifrice much research has been dedicated to the investigation of continuously releasing fluoride reservoirs. Such reservoirs can be either synthetic fluoride-containing matrices [1–3] or the  $\text{CaF}_2$ -like deposits observed on the enamel surface following the topical application of fluoride [4–6]. The formation of  $\text{CaF}_2$  particles however is not well understood and little is known about their stability and fluoride release under the conditions of the oral environment and how these parameters might be optimized. While methods have been developed to synthesize defined and fast releasing calcium fluoride particles in vitro [7, 8] the factors influencing the formation and morphology of calcium fluoride particles are largely unknown. Recent research on the formation of  $\text{MgF}_2$  particles by precipitation from soluble magnesium- and fluoride-containing salt solutions has demonstrated that the morphology of such particles with respect to size and shape can be varied by tuning the concentrations of  $\text{Mg}^{2+}$  and  $\text{F}^-$  in the starting solutions [9, 10]. Here we present data on defined morphological variations of  $\text{CaF}_2$  particles prepared by precipitation from soluble  $\text{NaF}$  and  $\text{CaCl}_2$  precursor salt solutions and their optimization as fluoride releasing reservoirs on the enamel surface.

J. Koeser (✉) · U. Pieleles  
Institute for Chemistry and Bioanalytics, University of Applied  
Sciences and Arts Northwestern Switzerland, Gruendenstrasse  
40, 4132 Muttenz, Switzerland  
e-mail: joachim.koeser@fhnw.ch

J. Koeser · T. S. Carvalho · A. Lussi  
Department of Preventive, Restorative and Pediatric Dentistry,  
University of Bern, 3010 Bern, Switzerland

## 2 Materials and methods

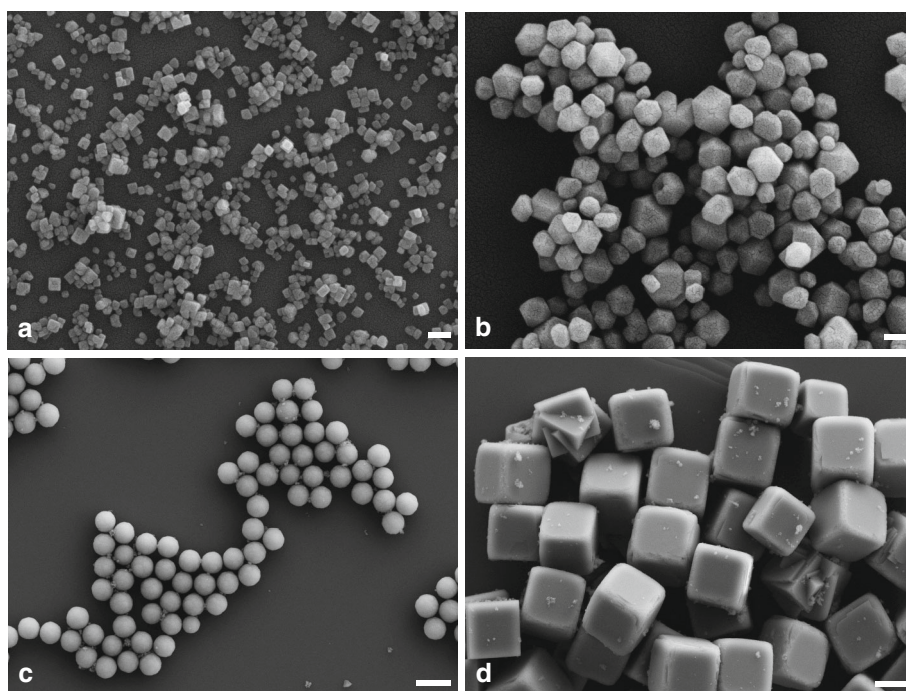
### 2.1 CaF<sub>2</sub> particle assembly and purification

CaF<sub>2</sub> particles were prepared by rapid mixing of aqueous CaCl<sub>2</sub> and NaF solutions in a 1:1 volume ratio. Precipitates formed within a short time as was evident from the changing turbidity of the samples. The particles were purified after at least 16 h of assembly by centrifugation at 5,000×g to 20,000×g, depending on the particle size, washed by resuspension in saturated CaF<sub>2</sub> solution, and then followed by a second centrifugation and vacuum drying. For the scanning electron microscopical (SEM) examination CaF<sub>2</sub> particles were assessed either directly after the resuspension step or upon resuspension of the dried pellet. A drop of the particle suspension was transferred onto freshly cleaved mica, dried, gold sputter coated and examined in a Zeiss Supra SEM. The CaF<sub>2</sub> particles used in this study were produced by a 1:1 volumetric mixing of (a) 50 mM NaF and 250 mM CaCl<sub>2</sub> (“CaF<sub>2</sub>-1” particles), (b) 50 mM NaF and 40 mM CaCl<sub>2</sub>, (c) 10 mM NaF and 1 M CaCl<sub>2</sub> and (d) 8 mM NaF and 40 mM CaCl<sub>2</sub> (“CaF<sub>2</sub>-2” particles) (Fig. 1). For experiments investigating the effect of phosphate during CaF<sub>2</sub> particle assembly, 1 M phosphate buffer pH 7 was added to the NaF and CaCl<sub>2</sub> solutions to reach final concentrations of either 0.01 mM phosphate, 0.1 mM phosphate or 1 mM phosphate.

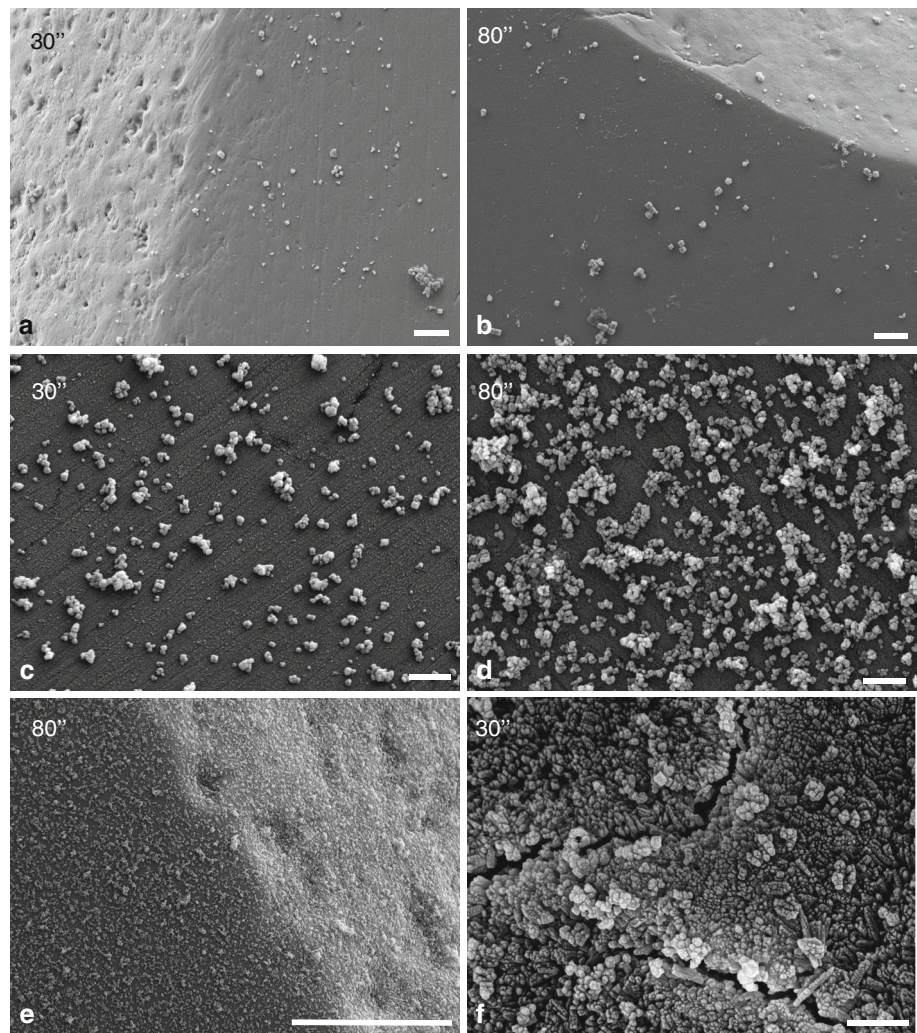
### 2.2 Preparation of tooth enamel specimens and CaF<sub>2</sub> particle adhesion assay

All enamel specimens were prepared from caries-free human molar teeth extracted by dental practitioners in Switzerland. Before the extraction, the patients were informed about the use of their teeth for research purposes and consent was obtained. All teeth were stored in 1 % chloramine T trihydrate solution after the extraction. Human molars were cut using an Isomet<sup>®</sup> Low Speed Saw (Buehler, Düsseldorf, Germany), separating the crowns from the roots. Subsequently the top of the crown was polished while the other sides were left untreated to produce both a native and a polished tooth enamel surface area. For the enamel polishing the crowns were serially abraded under constant tap water cooling using a Knuth Rotor machine (LabPol 21, Struers, Copenhagen, Denmark) with silicon carbide paper discs of grain size 18, 8, and 5 μm, for 60 s each. As a final step the crowns were polished for 60 s with 3 μm diamond abrasive on Struers polishing cloth under constant cooling (LaboPol-6, DP-Mol Polishing, DP-Stick HQ, Struers, Copenhagen, Denmark). Between two polishing steps and after the final polishing, all slabs were ultrasonicated for 1 min in tap water and rinsed. Thus, all prepared specimens had a flat ground enamel area with a 200 μm cut off layer. Samples were stored in a mineral solution (1.5 mM CaCl<sub>2</sub>, 1 mM KH<sub>2</sub>PO<sub>4</sub>, 50 mM NaCl, pH 7.0) and underwent further

**Fig. 1** SEM images of calcium fluoride particles prepared by precipitation from NaF and CaCl<sub>2</sub> solutions. Calcium fluoride particles were precipitated by 1:1 mixing of soluble NaF and CaCl<sub>2</sub> precursor solutions with molar concentrations of: **a** 50 mM NaF + 250 mM CaCl<sub>2</sub> (“CaF<sub>2</sub>-1”), **b** 50 mM NaF + 40 mM CaCl<sub>2</sub>, **c** 10 mM NaF + 1,000 mM CaCl<sub>2</sub>, **d** 8 mM NaF + 40 mM CaCl<sub>2</sub> (“CaF<sub>2</sub>-2”). Scale bars are 200 nm (**a**, **b**) and 2 μm (**c**, **d**)



**Fig. 2** SEM images of enamel surfaces following incubation with calcium fluoride particles. Enamel surfaces were incubated with calcium fluoride particles in solution, corresponding to 450 ppm fluoride (**a**, **c**, **f**) or 1,450 ppm fluoride (**b**, **d**, **e**), and for 30 s (**a**, **c**, **f**) or 80 s (**b**, **d**, **e**). All surface were then washed and examined by SEM. **a** and **b** “CaF<sub>2</sub>-2” particles, **c**–**f** “CaF<sub>2</sub>-1” particles. **e** Shows particle adhesion on polished (left side of the image) and native enamel (right side of the image), **f** is a larger magnification of particles adhering to an area of eroded enamel. Scale bars are 10 μm (**a**, **b**, **e**) and 500 nm (**c**, **d**, **f**)



polishing with a 1 μm diamond abrasive (60 s, LaboPol-6, DP-Mol Polishing, DP-Stick HQ, Struers, Copenhagen, Denmark) immediately before the experiment.

CaF<sub>2</sub>-1 and CaF<sub>2</sub>-2 particles were resuspended in saturated CaF<sub>2</sub> solution at amounts corresponding to either 450 ppm or 1,450 ppm fluoride and incubated with the prepared enamel specimens under gentle agitation. Subsequently excess particles were removed by dipping the samples in water several times. The samples they were analyzed by SEM after drying.

### 2.3 Image processing and analysis

For the analysis of the CaF<sub>2</sub> particle coating densities on enamel surfaces the Fiji/ImageJ software package was employed [11]. The particle analysis mode was used and the values for the threshold adjustment, particles pixel size and particle circularity were optimized with respect to the brightness, magnification and resolution of the analyzed

SEM images. Images with a large z range (Fig. 2a, e) were focus stacked using the combineZP software [12].

### 2.4 Fluoride release from CaF<sub>2</sub> particles

The fluoride release from different CaF<sub>2</sub> particle types was followed over 90 min with a fluoride sensitive electrode (perfectION™, Mettler Toledo, Greifensee, Switzerland). A total of 10 mg of dried CaF<sub>2</sub> particles was mixed with 100 ml of buffer (20 mM Hepes, 130 mM KCl, 1.5 mM CaCl<sub>2</sub> either with or without 3.5 mM sodium phosphate, pH 7.05) and stirred at 200 rpm with the fluoride sensitive electrode inserted in the solution. Soluble fluoride was measured again at least 24 h later and the values did not exceed 150 % of the soluble amount detected after 90 min. For the analysis of particle dissolution in saliva, stimulated human saliva was collected in chilled vials, from 30 healthy individuals, 2 h after their last meal or oral hygiene. The saliva was pooled, centrifuged from 20 min

at 4 °C (3,000 x g) and separated into 6 ml aliquots. Then, 0.6 mg of dried particles were incubated with 6 ml of pooled stimulated human saliva, stirred at 200 rpm and the fluoride-sensitive electrode inserted in this solution.

### 3 Results

#### 3.1 CaF<sub>2</sub> particle assembly

CaF<sub>2</sub> particles could be produced by precipitation following the mixing of soluble fluoride and calcium salt solutions. For the experiments described here, aqueous NaF and CaCl<sub>2</sub> solutions were used. One to one volumetric mixing of solutions with concentrations of F<sup>-</sup> ranging from 5 mM to 75 mM and of Ca<sup>2+</sup> ranging from 8 mM to 2 M resulted in the precipitation of particles with different morphologies. Some of the different shapes we observed were round, cubic, hexagonal, and irregular formed particles in the size range of approximately 50 nm to 2 μm (see Fig. 1 and [13]). Although the particle shapes could not be predicted in advance a general trend towards larger assemblies at lower parent ion concentrations was observed.

#### 3.2 CaF<sub>2</sub> particle interaction with tooth enamel surfaces

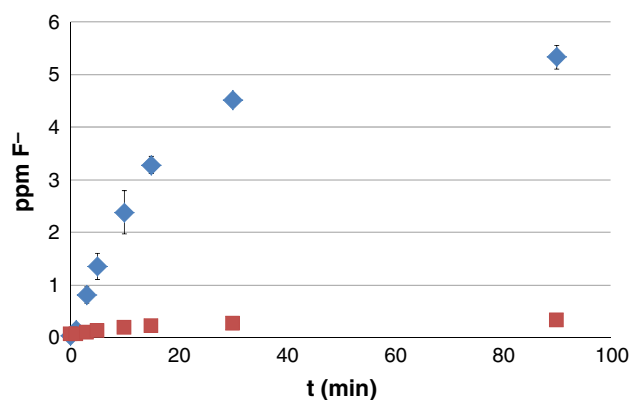
As a first step in the investigation of the suitability of the prepared CaF<sub>2</sub> particles as fluoride reservoirs, their interaction with tooth enamel surfaces was analyzed. Suspensions from two particle types of different size, smaller “CaF<sub>2</sub>-1” and larger “CaF<sub>2</sub>-2” particles (see Fig. 1), were incubated with human enamel samples at concentrations corresponding to typical dental care product fluoride levels. One surface of each tooth sample had been polished while the rest was left untreated in order to compare the interaction of the particles with ‘native’ and polished enamel surfaces. Figure 2 shows the results of these experiments. The coating density of the particles on enamel increased with the particle concentration and incubation times. No obvious differences were observed in the interaction of CaF<sub>2</sub> particles with polished and unpolished tooth surfaces. The smaller “CaF<sub>2</sub>-1” particles covered a larger area as compared to the larger “CaF<sub>2</sub>-2” particles. SEM images were analyzed to compare the typical enamel surface coverage with the two different CaF<sub>2</sub> particle types. Upon incubation with “CaF<sub>2</sub>-1” particles, a coverage of 13 % was reached after 30 s incubation with 450 ppm fluoride particles and 40 % after 80 s incubation with 1,450 ppm fluoride particles. The coverage of the enamel surfaces with the larger “CaF<sub>2</sub>-2” particles were considerably lower, corresponding to <1 % (30 s, 450 ppm) and 3 % (80 s, 1,450 ppm).

#### 3.3 Fluoride release from CaF<sub>2</sub> particles

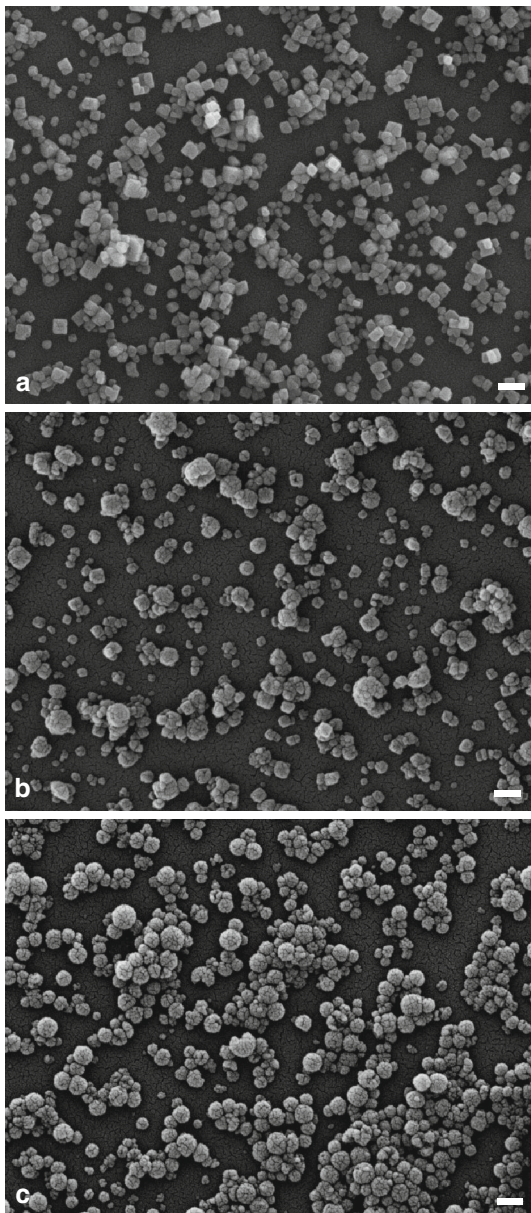
To further investigate the suitability of the synthesized CaF<sub>2</sub> particles as fluoride storage reservoirs, their dissolution in physiological buffers was analyzed. These experiments were performed with “CaF<sub>2</sub>-1” particles. It turned out that the fluoride release was strongly influenced by the presence of phosphate in the buffer solution which prompted the comparison of the release in buffers without phosphate and with 3.5 mM phosphate, the latter one corresponding approximately to saliva levels [14, 15] (Fig. 3). Under the experimental conditions chosen the levels of soluble fluoride after 90 min reached  $5.33 \pm 0.22$  ppm in buffers without phosphate and  $0.32 \pm 0.02$  ppm in buffers with 3.5 mM phosphate. When incubated with pooled stimulated human saliva the same concentration of particles released approximately 0.1–0.2 ppm fluoride.

#### 3.4 Enhanced fluoride release from CaF<sub>2</sub> particles

Next, it was investigated if the fluoride release from the CaF<sub>2</sub> particles could be influenced by the addition of excipients during their synthesis. The literature described a destabilizing effect of phosphate during the formation of CaF<sub>2</sub> particles [16]. Thus, a modified protocol for the preparation of CaF<sub>2</sub> particles was developed that applied the same CaCl<sub>2</sub> and NaF concentrations as during the preparation of “CaF<sub>2</sub>-1” particles, however in the presence of substoichiometric amounts of phosphate. When CaF<sub>2</sub> particles were assembled from final concentrations of 25 mM fluoride ions and 125 mM calcium ions, already the addition of as little as 0.01 mM phosphate during

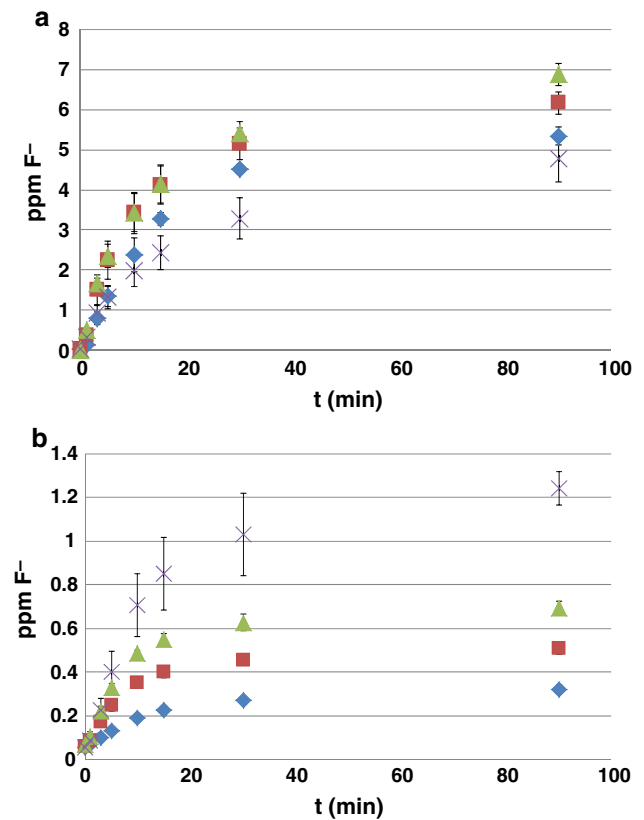


**Fig. 3** Time-dependent fluoride release from “CaF<sub>2</sub>-1” particles upon incubation in buffers with different phosphate concentrations. Fluoride release (in ppm) from 100 mg/l “CaF<sub>2</sub>-1” particles in buffers with 3.5 mM phosphate (diamonds) or without phosphate (squares) under constant stirring. Error bars for the release experiments in buffers without phosphate are smaller than the size of the symbol



**Fig. 4** SEM images of calcium fluoride particles prepared in the presence of different concentrations of phosphate. Calcium fluoride particles were precipitated by 1:1 mixing of soluble NaF and CaCl<sub>2</sub> precursor solutions with molar concentrations of 50 mM NaF and 250 mM CaCl<sub>2</sub> in the presence of **a** 0 mM phosphate, **b** 0.01 mM phosphate and **c** 1 mM phosphate. Scale bars are 200 nm

assembly had a pronounced effect on the morphology of the resulting particles (Fig. 4). Particles assembled in the presence of phosphate had a more globular appearance and exhibited a rough surface structure. The fluoride release from such synthesized particles in physiological buffers with 3.5 mM phosphate showed marked differences when compared to the release from “CaF<sub>2</sub>-1” particles synthesized in the absence of phosphate (Fig. 5). The presence of 0.01 mM phosphate during assembly nearly doubled the solubility of fluoride from the particles. A maximum of



**Fig. 5** Influence of phosphate on the fluoride release kinetics from calcium fluoride particles. Fluoride release from calcium fluoride particles synthesized in the presence of 0 mM phosphate (*diamonds*), 0.01 mM phosphate (*squares*), 0.1 mM phosphate (*triangles*) and 1 mM phosphate (*crosses*) upon resuspension in buffer with 0 mM phosphate (**a**) or 3.5 mM phosphate (**b**) at particle concentrations of 100 mg/l. The fluoride release was monitored under constant stirring with a fluoride-sensitive electrode

$1.24 \pm 0.08$  ppm soluble fluoride was released from particles assembled in the presence of 1 mM phosphate within 90 min in 3.5 mM phosphate-containing buffer.

#### 4 Discussion

The present study reports results on the preparation of tailor-made CaF<sub>2</sub> particles and their suitability as enamel bound fluoride reservoirs for dental care applications.

Generally the formation of CaF<sub>2</sub> and CaF<sub>2</sub>-like material during dental care is limited by the low availability of calcium ions [17]. This problem can be circumvented by the addition of in vitro synthesized CaF<sub>2</sub> particles to dental care products.

The results demonstrate that the size and shape of in vitro assembled CaF<sub>2</sub> particles can be adjusted in a range between 50 nm and several micrometers. Globular, cubic, hexagonal or irregular-shaped particles can be generated (see also [13]). For the experiments presented here, cubic

rather than round particles were chosen since they were expected to have larger and thus stronger interaction sites with the enamel surface. When such particles were applied to tooth enamel samples in concentrations corresponding to the fluoride content of typical mouth rinses or toothpastes, they interacted with enamel surfaces leading to coverage levels of up to 40 %.

Fluoride levels in oral fluids play an important role in the de-/remineralization cycles of tooth enamel. While enamel demineralization is best reduced in the presence of relatively high concentrations of fluoride [18], remineralization benefits from considerably lower concentrations of salivary or plaque fluoride. Epidemiologic studies found a positive correlation between fluoride levels of 0.02 ppm and 0.04 ppm in children and their incidence rate for caries [19, 20]. Furthermore in vitro studies on remineralization models demonstrated significant enhancement of enamel remineralization in the presence of lower sub ppm levels of fluoride [20]. Generally salivary levels of 0.1 ppm are expected to largely reduce caries even in high risk individuals [21]. Under the experimental condition chosen in the present report, the fluoride release characteristics of the calcium fluoride particles resulted in soluble fluoride levels well above 0.3 ppm if particles were synthesized in the absence of soluble phosphate and above 1 ppm if prepared in the presence of 1 mM phosphate.

The results presented here represent data from in vitro studies and the conditions on the enamel surface in the oral cavity most likely will influence the fluoride release in vivo. An in situ study conducted by Tenuta et al. [22] investigated plaque fluid fluoride levels in *Streptococcus mutans* test plaques in contact with in vitro generated CaF<sub>2</sub> deposits in the oral cavity and their correlation with surface microhardness changes upon a cariogenic challenge. Their results showed a clear positive correlation between the presence of CaF<sub>2</sub> deposits, fluoride levels in the plaque fluid and reduced changes in surface microhardness after a cariogenic challenge. The protective effect was strongest with freshly deposited CaF<sub>2</sub>, however even after 48 h of aging in artificial saliva, surface microhardness losses were reduced by more than a factor of two as compared to CaF<sub>2</sub>-free samples. The amounts of CaF<sub>2</sub> deposited in these experiments were approximately 20 µg fluoride per cm<sup>2</sup>, which is comparable to the amounts deposited when the present “CaF<sub>2</sub>-1” particles interacted with enamel surfaces. More specifically, assuming an average particle size of 100 nm, the amount deposited can be calculated to be in the range of 12 µg fluoride per cm<sup>2</sup> for the 40 % coverage rate observed after 80 s of incubation (Fig. 2). Future work will have to focus on the long-term fluoride release from the CaF<sub>2</sub> particles presented here under the conditions of the oral cavity. The literature describes the formation of a pH-dependent protective phosphate layer on the surface of

CaF<sub>2</sub> particles in physiological solutions containing soluble phosphate and which influences the release of fluoride from such particles over time and during cariogenic challenges [23–25]. The formation of such a phosphate layer can explain the results of the present study, where the levels of soluble fluoride were lower in buffers containing phosphate in relation to phosphate-free buffers. Furthermore, the formation of a layer could also explain the reduced fluoride release observed after 48 h of aging of the CaF<sub>2</sub> deposits in the publication by Tenuta et al. [22]. However, there is still a sufficient amount of fluoride released to have a beneficial effect during a cariogenic challenge, and several authors report an increased fluoride release from phosphate coated CaF<sub>2</sub> under the acidic conditions of a cariogenic challenge [17, 26, 27]. The latter scenario provides support for the potential of synthesized CaF<sub>2</sub> particles serving as caries reducing enamel associated fluoride reservoirs.

The formation of the phosphate layer has been regarded as the major mechanism influencing CaF<sub>2</sub> solubility in saliva [24]. When investigating the solubility of CaF<sub>2</sub> in water, 2 mmol/l phosphate solution or saliva, Saxegaard et al. [24] observed that the rate of dissolution of CaF<sub>2</sub> was comparably lower in both saliva and in the phosphate solution. In our experimental model, the solubility of CaF<sub>2</sub> did decrease in the phosphate solution (3.5 mM), possibly due to the phosphate layer, but, strikingly, the reduction in CaF<sub>2</sub> solubility was even more pronounced in the presence of saliva. In that case, the presence of other compounds in saliva, such as proteins from the salivary pellicle, also affect the dissolution of CaF<sub>2</sub> [28]. When the tooth is exposed to saliva, pellicle precursor proteins, such as statherin, almost immediately interact with the tooth surface, thus triggering the initial stages of a perm-selective pellicle formation [29]. The salivary pellicle is then able to modify the transport of calcium ions to and from the tooth surface, which also explains the lower solubility of CaF<sub>2</sub> in saliva.

In vivo, the precipitation of CaF<sub>2</sub> particles onto the tooth surface usually occurs in the presence of the salivary pellicle. These CaF<sub>2</sub> particles are often described as having a globular spherical shape with a nodular surface [4]. Remarkably, our results showed that such structures were often found when CaF<sub>2</sub> particles were formed in the presence of phosphate. Furthermore, the presence of phosphate during particle formation also influenced the CaF<sub>2</sub> solubility, where the addition of substoichiometric amounts of phosphate during CaF<sub>2</sub> particle synthesis was able to considerably increase fluoride solubility. Consequently, the presence of saliva in vivo can serve as a source for phosphate during particle assembly, thus leading to globular spherical shaped, albeit more soluble, CaF<sub>2</sub> particles. On the other hand, the presence of a salivary film may lead to greater amounts of CaF<sub>2</sub> formed on the tooth surface [30],

as well as to the formation of the phosphate layer and salivary pellicle over the particles, which is related to lower solubility. Nevertheless, more studies are still necessary to further elucidate the effect of the interaction of phosphate during and after particle assembly, as well as the effect of salivary proteins on the solubility of the CaF<sub>2</sub> particles *in vivo*.

Future experiments could combine the particle coverage rates of enamel described here with typical saliva dynamics and ion diffusion rates in the dental plaque to give information on the *in vivo* fluoride reservoir properties of the present CaF<sub>2</sub> particles described here. Of special interest in this respect will be the examination of calcium fluoride particles in caries models working with bacterial biofilms.

## 5 Conclusion

The main finding of the presented study is that CaF<sub>2</sub> particle assembly is influenced by the concentrations of Ca<sup>2+</sup> and F<sup>-</sup> ions in the parent salt solutions and the presence of modifying compounds. This offers the possibility to tune the morphology and fluoride release kinetics of such particles to suit the specific requirements of different topical applications of fluoride containing dental care products.

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