Materials for life sciences



Drug self-gating fluorescent nanoparticles for pH-responsive doxorubicin delivery

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ABSTRACT

Intelligent drug delivery systems have attracted great attention in the field of biomedicine and cancer diagnosis. In this work, a drug delivery system that can be gated by doxorubicin itself and together with pH-responsive ability has been designed and prepared based on the upconversion nanoparticles. The drug delivery system is a special core–shell structure, consisting of upconversion nanoparticle core and mesoporous silica shell. The new system tactfully bypasses the use of auxiliary capping agents and exhibits desirable drug release at pH = 5, enhancing HeLa cells inhibition. The introduction of Schiff base plays a key role in the process of achieving pH-responsive drug release. Moreover, upconversion nanoparticles could emit bright yellow-green fluorescence (540 nm) under the irradiation of near-infrared light (980 nm) for in vivo bioimaging. This characteristic provides the possibility of locating tumor tissues and real-time tracing drug delivery.

Introduction

Research on cancer is still a crucial issue worldwide; precise diagnosis and fundamental therapy of cancer are still important challenges. Stimuli-responsive drug delivery systems that could specifically transport antineoplastic drugs into tumor tissues with minimal premature release are highly expected [1, 2]. In this respect, nanostructured materials perform a very important role because of their intrinsic small size that allows their easy incorporation as probes into living cells or organisms [3, 4]. Multifunctional response nanocomposites for stimuli-responsive drug delivery have been developed, such as pH,

temperature, light, ionic strength and magnetic field [5–9].

Among the various multifunctional nanoparticles, due to the special characteristics of tunable multicolor sharp emission, large anti-Stokes shifts, great photostability, superior light penetration depth and autofluorescence-free nature, rare-earth-doped upconversion nanoparticles (UCNPs) have been demonstrated as one of the most promising biomedicine nanomaterials [10–13]. In the field of biomedicine, UCNPs can be used as drug carriers and luminescence labels to track the drug release process in living system [14–17]. However, the poor biocompatibility and low specific surface area of UCNP nanoparticles restrict their direct applications [18, 19].

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Mesoporous silica nanomaterial (MSN) is an outstanding drug carrier with excellent biocompatibility and the ease of surface modification [20, 21]. UCNPs composites with MSN as drug loading materials have attracted considerable attention [22]. To realize the stimuli-responsiveness, organic polymer molecules [23], DNA [24], peptides [25] and inorganic nanoparticles [26] have been usually used as capping agents coated outside the UCNP@MSN composites. Jun Lin et al. reported a controlled release system based on upconversion luminescent microspheres of NaYF4:Yb/Er@SiO2 coated with the smart hydrogel poly (N-isopropylacrylamide-comethacrylic acid) shell [27]. Boshi Tian et al. prepared the functional UCNPs@mSiO2-PAA nanocomposite with UCNPs as core and poly (acrylic acid) (PAA)-gated mesoporous silica as shell for oral drug delivery [23]. A facile methodology UCNPs@mSiO2-PEG was developed to construct as a multifunctional anticancer drug nanocarrier by combining the special advantages of upconversion nanoparticles and mesoporous silica, which can be used as an anticancer drug doxorubicin (DOX) delivery carrier and applied for magnetic resonance imaging (MRI) imaging [28]. Drug delivery system with the features of multifunctional synergy has become a hot research topic. Despite the excellent performance, the delivery systems still have critical problems such as unpredictable toxicity of the capping agents as well as the scarcity of bioresponsiveness [29, 30].

In the present study, a drug delivery system that can be gated by DOX itself and together with pHresponsive ability has been designed and prepared based on the upconversion nanoparticles (UCNPs), shown in Fig. 1a. MSN was used as the intermediate layer covered on the UCNPs for loading with doxorubicin DOX. The drug DOX itself could act as a pHsensitive "gatekeeper" to minimize the potential risks of auxiliary capping agents. This system has dual functions with responsive drug delivery and fluorescence imaging. The fluorescence property of UCNPs provides the possibility of locating tumor tissues and real-time tracing drug delivery.

Experimental

Preparation of UCNP@nSiO₂@mSiO₂ nanoparticles

According to the literature protocols with modifications, UCNPs were synthesized via hydrothermal reaction. In a typical experiment, $YCl_3 \cdot 6H_2O$ (0.78 mmol), $YbCl_3 \cdot 6H_2O$ (0.2 mmol) and $ErCl_3 \cdot 6H_2O$ (0.02 mmol) were added to a three-neck round-bottom flask containing 15 ml H₂O. Then, 15 ml of disodium EDTA solution (1 mmol) and 15 ml of NaF solution (6 mmol) were sequentially added with magnetic stirring. After stirring for 1 h, the mixture was transferred to a Teflon autoclave, and the reaction was heated at 180 °C for 2 h. After the temperature was collected by centrifugation, washed with water and ethanol, and finally dried in a vacuum oven.

The as-prepared UCNPs were coated with dense silica and mesoporous silica sequentially by means of modified sol–gel reaction. Firstly, a quantity of UCNPs was dispersed into a mixture consisting of 1 ml ammonia, 15 ml distilled water and 60 ml ethanol, and then, 50 µl TEOS was slowly add. After hydrolysis reaction at room temperature for 4 h, the UCNP@nSiO₂ nanoparticles were collected by centrifugation and washed. Afterward, the above coating process was repeated to get UCNP@nSiO₂@C-SiO₂ except that CTAB should be added as pore template and the amount of TEOS was increased to 100 µl.

Surface functionalization of UCNP@nSiO₂@mSiO₂

An amount of UCNP@nSiO₂@C-SiO₂ nanoparticles were dispersed in 30 ml toluene containing 50 μ l APTES and 10 μ l TEA. After the solution was uniformly dispersed by ultrasound, the reaction was refluxed at 110 °C oil bath for 12 h under the protection of argon, receiving UCNP@nSiO₂@C-SiO₂-NH₂. The product was collected by centrifugation and washed with toluene and ethanol several times.

In order to modify the external surface with aldehyde groups, the UCNP@nSiO₂@C-SiO₂-NH₂ was put into 30 ml deionized water/DMSO mixture solution (4:1, v/v), containing 4-formylbenzoic acid (15.0 mg, 0.10 mmol), EDC (23.0 mg, 0.12 mmol) and NHS (13.8 mg, 0.12 mmol). After stirring at room temperature for 24 h, aldehyde groups were successfully introduced onto the surface by amidation reaction between carboxyl groups and amino groups for subsequent drug delivery. Finally, the nanoparticles were dispersed in 100 ml acetone and refluxed at 80 °C overnight to remove the surfactant template.



Figure 1 Schematic of the UCNP-based drug delivery system.

In this way, the UCNP@nSiO₂@mSiO₂-CHO nanoparticle was obtained, as shown in Fig. 1b.

Drug loading and release

DOX was chosen as a model drug to investigate the drug delivery behavior from the UCNP@nSiO₂@-mSiO₂-CHO nanocomposite. UCNP@nSiO₂@mSiO₂-CHO (10 mg) was soaked in 10 mL deionized water containing 50 μ g DOX at pH 6.0 and room temperature for 24 h in the dark. Adjusting the solution to pH = 8.0 and stirring for 2 h, the outlets of mesoporous nanoparticles were capped with DOX through reversible covalent benzoic–imine linkage between the aldehyde group and amine group in DOX. After centrifugation and washing, the DOX-loaded and DOX-gated nanoparticles were prepared. The schematic diagram is shown in Fig. 1c.

To study the pH-responsive release properties, the DOX loading samples were immersed in 10 mL of pH = 7.4, 6.8 and 5.0 PBS buffer solutions at room

temperature with gentle shaking. The amounts of released DOX were analyzed by UV–vis spectrophotometer. The details are in the support information.

In vitro cytotoxicity assay

The in vitro cytotoxicity was measured by MTT assay on HeLa cells. Cells with a density of 20000 cells per well were cultured in a 96-well plate at 37 °C for 24 h. Then, the cells were incubated with different concentrations of UCNP@n-SiO₂@mSiO₂, DOX self-gated nanoparticles and free UCNP for 24 h. Then, after cleaning with PBS, 5.0 mg MTT (20 μ l of 3-(4, 5-dimethyl-2-thiazolyl)-2, 5-diphenyl-2-H-tetrazolium bromide solution) was added to 96-well plates. After 4 h, the solution was removed and cleaned, and then, 100 μ l of dimethyl sulfoxide (DMSO) was added to each well and shaken slowly for 10 min at room temperature.

Characterization

Transmission electron microscopy (TEM) was performed on JEM-2100F to characterize the change of sample morphology. The crystal structures were characterized by X-ray diffraction (XRD, Rigaku D/max 200 diffractometer, Cu K α) at a scan rate of 2°/min. Nitrogen adsorption–desorption isotherm was measured on a COULTER SA 3100 apparatus at a liquid nitrogen temperature (77 K). The surface modification and functional group modification of the material can be tested by Fourier transformation infrared spectrometer (FT-IR). IVIS Lumina XRMS series III (PerkinElmer) was used for the in vivo imaging. The UC emission spectra were obtained on an FLS1000 Edinburgh Analytical Instrument equipped using a 980 nm laser as the excitation source.

Results and discussion

Preparation and characterization of UCNP@nSiO₂@mSiO₂ nanoparticles

UCNPs were synthesized using modified hydrothermal method. The morphology and structures of samples were tested by TEM. In Fig. 2a, it can be observed that the NaFY₄:Yb, Er nanocrystals are well monodispersed in water and display uniform morphology with a size of around 100 nm. In the silica coating process, for no fluorescence quenching of the nanoparticles, a thin and dense silica layer was first coated on the surface of UCNPs. Figure 2b shows the UCNP@nSiO₂ nanoparticles; the thickness of solid silica is about 8 nm. The resulting particles were then coated with a layer of mesoporous silica; Fig. 2c shows the UCNP@nSiO₂@mSiO₂ structure. The XRD patterns of the UCNPs, UCNP@nSiO₂ and UCNP@nSiO₂ @mSiO₂ are separately displayed in Fig. 3. Compared with the cubic phase of β -NaYF4 (JCPDS: 06-0342), they all show face-centered cubic phase structures, suggesting that the samples possess intact crystal structure. In the patterns of UCNPs@nSiO₂@mSiO₂, the peak at $2\theta = 22^{\circ}$ is a broad diffraction peak of amorphous silica, which suggests that the mesoporous silica shell was coated on the UCNPs surface.

Figure 4 shows the N₂ adsorption–desorption isotherms of UCNPs@nSiO₂@mSiO₂ nanocomposites. The sample exhibits typical type-IV isotherms, which further confirms the existence of mesoporous structure. The BET surface area and total pore volume of UCNPs@@nSiO₂@mSiO₂ were 88.76 m² g⁻¹ and 0.096 cm³ g⁻¹, respectively. The BJH pore sizes of UCNPs@nSiO₂@mSiO₂ were estimated to be 4.3 nm. Large specific surface area and moderate pore size provide satisfactory preconditions for the loading of drugs.

The upconversion luminescence spectra and the following photographs of UCNPs, UCNPs@nSiO₂ and UCNPs@nSiO₂@mSiO₂ under 980-nm excitation are shown in Fig. 5. It can be seen from the graph that when excited by 980 nm laser, all samples emit bright yellow-green fluorescence, and the fluorescence spectra show obvious two emission peaks at 540 nm and 655 nm, resulting in the energy transition from ${}^{4}S_{3/2} \rightarrow {}^{4}I_{15/2}$ and ${}^{4}F_{9/2} \rightarrow {}^{4}I_{15/2}$ of Er^{3+} ions, respectively. The intensity of emission peaks



Figure 2 TEM images of a UCNPs, b UCNP@nSiO₂ and c UCNP@nSiO₂@mSiO₂.



Figure 3 XRD patterns of UCNPs, UCNP@nSiO₂, UCNPs@nSiO₂@mSiO₂ and the standard card of β -NaYF4 (JCPDS: 06-0342).



Figure 4 N_2 adsorption-desorption isotherm and pore size distribution (inset) of UCNP@nSiO₂@mSiO₂ nanoparticles.

decreases to a certain extent after coated with SiO_2 shell layer by layer on the particle surface, but still retains strong yellow-green emission intensity.

Surface functionalization and characterization of UCNP@nSiO₂@mSiO₂

Schiff base reaction is a reversible condensation reaction sensitive to acid and base. The formed imine group is very active and can be broken in acidic environment. Amino groups can be first grafted onto UCNP@nSiO₂@mSiO₂ surface with silane coupling



Figure 5 Fluorescence spectra and digital photographs of (a) UCNPs, (b) UCNP@nSiO₂ and (c) UCNP@nSiO₂@mSiO₂.

agent APTES. Then, 4-carboxybenzaldehyde is introduced onto the surface of particles to complete the aldehyde modification. Schiff base reaction between amino groups inherent in DOX and aldehyde groups on nanoparticles comes to achieve DOX self-capped mesoporous and pH-responsive drug release.

Figure 6a shows the chromogenic reaction of ninhydrin; the UCNP@nSiO2@mSiO2-NH2 solution turned blue, while the supernatant does not change in color. The comparison results show that amino groups have been successfully grafted onto the surface of the particles without the existence of free APTES. Figure 6b displays the FT-IR spectra of UCNP@nSiO₂@mSiO₂, UCNP@nSiO₂@mSiO₂-NH₂ and UCNP@nSiO2@mSiO2-CHO. For UCNP@n SiO₂@mSiO₂ FT-IR spectroscopy, absorption bands at 1080 cm⁻¹ (Si–O–Si antisymmetric stretching vibration), 800 cm⁻¹ (Si–O–Si symmetric stretching vibration) and 960 cm⁻¹(Si–OH bending vibration) are presented. After modification with APTES, the stretching vibration of C-N and N-H increases the peak at 1030–1230 cm⁻¹ and 3400–3500 cm⁻¹, which is shown in curve (II). For the UCNP@nSiO₂@mSiO₂-CHO shown in curve (III), the characteristics of amide bond (1643 cm^{-1}) and aldehyde group (1551 cm⁻¹) were clearly observed, confirming that UCNP@nSiO2@mSiO2 was functionalized successfully with -CHO groups.

Nanocomposites modified with -CHO groups have high fluorescence properties, as shown in

Figure 6 a Chromogenic reaction of ninhydrin, b FT-IR spectra of (i) UCNP@nSiO₂@mSiO₂, (ii) UCNP@nSiO₂@mSiO₂-NH₂, (iii) UCNP@nSiO₂@mSiO₂-CHO.

Fig. 7A. Figure 7B displays the in vivo fluorescence images of UCNPs@nSiO2@mSiO2-CHO after intravenous injection into the mice through a tail vein. Under 980-nm excitation, clear fluorescence signals in the mice were detected after 2 h of injection. Over time, the fluorescence of mice tail gradually weakened, and fluorescence signals appeared in the left kidney, indicating that the particles gradually entered the kidney with blood circulation. High fluorescence intensity was detected in the urine and urethra, as shown in Fig. 5B-d, which means UCNPs@nSiO2@mSiO2-CHO nanoparticles exhibit good biocompatibility and can be excreted in urine. The in vivo results confirm the high luminous efficiency and low systematic toxicity of the self-gated nanoparticles.

Drug loading and release

To determine the gating effect of DOX, we further delivery explore the drug capacity of UCNP@nSiO₂@mSiO₂-CHO. The amount of DOX in the nanocarrier solution can be determined by the characteristic DOX absorption peak at 500 nm. Figure 8a shows the loading profile of composite nanoparticles under different DOX initial concentrations. With the concentration increasing, the drug adsorption capacity of the carrier increases gradually. When the initial concentration increases above $60 \mu g/ml$, the adsorption capacity of the carrier reaches saturation, up to 20 μ g/mg or more, which is higher than UCNP@nSiO2@mSiO2 without any functional treatment (Fig. S2). This phenomenon

Figure 7 A Fluorescence spectra of UCNP@ $nSiO_2@mSiO_2$ -CHO, B in vivo fluorescence images of injection after (a) 2 h, (b) 24 h, (c) 72 h, (d) 86 h.

attributes to the capillary action and van der Waals force of porous structure of the composite nanoparticles. The capillary action of pore and van der Waals force make the drug to be easily loaded, and with the increase in loading, the action force gradually decreases, thus reaching the saturated adsorption of the drug. Then, the covalent benzoic–imine linkage achieves the capping effect in alkaline condition.

Figure 8b displays the DOX release profiles in PBS buffer solution with different pH values. Obviously, the DOX release behavior exhibits a significantly pHdependent property. In alkaline and neutral buffer (pH = 7.4, 6.8), the release rate of drug in the carrier is extremely low, and the cumulative release of DOX is only about 17%, which displays the excellent DOX self-gating effect under physiological conditions. In contrast, under acidic environment (PH = 5), large amount of DOX (~ 45.5%) release is observed over the same time period. However, there is no pH response for drug release process of single UCNP@nSiO2@mSiO₂, and the release rate is very high (Fig. S3). Clearly, in this work, the pH-controlled drug release system has been successfully constructed. The gatekeeper pH-responsive Schiff base reaction plays the vital role of controlled release of DOX. Under alkaline condition, the covalent benzoicimine bond between DOX and -CHO groups seals the pore entrances of nanoparticles, causing the DOX molecules encapsulated in the blocked pore channels. A few release of DOX may be due to the adsorption of DOX on the external surface of UCNP@nSiO₂@mSiO₂-CHO. Under acidic environment, inverse

reaction takes place and the covalent benzoic-imine bond is broken, resulting that the pore entrance is opened. These results demonstrate that without any auxiliary capping agents, the DOX-gated method in this study is a feasible strategy that meets the requirements of drug loading and achieves pH-responsive drug release, exhibiting its potential applicability in combination therapy.

Cytotoxicity

Cytotoxicity of the synthesized nanocomposite is evaluated by 3-[4,5-dimethylthiazol-2-yl]-2,5diphenyltetrazolium bromide (MTT) assay on HeLa cells. From Fig. 9, after incubation with UCNP and

Figure 9 In vitro cell viability assay of UCNP, UCNP@nSiO₂@mSiO₂-CHO, UCNP@nSiO₂@mSiO₂-CHO-DOX and free DOX.

Figure 8 a DOX loading capacity of as-prepared UCNP@nSiO2@mSiO2-CHO; b DOX release at different pH conditions.

Figure 10 In vitro cytotoxicity against HeLa cells of UCNP@nSiO2@mSiO2-CHO-DOX to cells.

UCNP@nSiO₂@mSiO₂ nanoparticles with various concentrations of 0–80 μ g/mL for 24 h, the cellular survival rate descended slightly with the increased concentration, especially cells with UCNP@nSiO₂@mSiO₂, and the cellular viability was still higher than 90.72%, which implied that the samples exhibit a negligible cytotoxicity. The cells incubated with UCNP@nSiO₂@mSiO₂ exhibited a relative high survival rate, which indicated that SiO₂ coating modification can improve the biocompatibility of the nanocomposite. The prepared composite fluorescent nanoparticles display wonderful biocompatibility and can be used as drug carrier in the biomedical fields.

Furthermore, the cytotoxic effect of drug-loaded nanoparticles on HeLa cells was also investigated. Upon drug loading, the self-gated group UCNP@nSiO2@mSiO2-CHO-DOX could remarkably inhibit HeLa cell growth at a low concentration range. The cellular survival rate descended seriously with the increased concentration, displaying a similar concentration-dependent toxicity effect to the pure DOX. It mainly depends on the positive surface potential for cell internalization and subsequently the sustained DOX release from the pore channels of mesoporous nanocomposites. The cell viability assay for different times is shown in Fig. S4, a time-dependent toxicity can be observed for UCNP@nSiO₂@ mSiO₂-CHO-DOX nanoparticles. Figure 10 shows the effect of UCNP@nSiO2@mSiO2-CHO-DOX on the

apoptosis rate under different concentrations after 24 h. With the concentrations increasing, cell viability decreased rapidly. In the images of live cells (green) and dead cells (red), it is apparent that the tumor cells are almost totally killed when cultured with UCNP@nSiO₂@mSiO₂-CHO-DOX at 80 μ g/mL after 24 h compared with the cells incubated with pure culture. Taken together, these results suggest the drug loading nanoparticles own the potential to selectively kill more cancerous cells than normal tissue/cells at weakly acidic tumor microenvironments.

Conclusion

We designed a multifunctional antitumor nanocarrier, which consists of facile drug self-gated, pHsensitive and near-infrared fluorescence properties. Such a new pH-responsive system, gated by antitumor drug DOX itself, could tactfully bypass the use of complex capping agents. The pH-sensitive benzoicimine covalent bond can effectively control the release of drugs. The new system, which exhibits desirable drug release at pH = 5, can effectively inhibit HeLa cells growth. More inspiringly, nanocomposites display distinctive autofluorescence characteristics because of rare-earth upconversion particles in the core. Excellent in vivo imaging data show that particles can be excreted with urine, reducing unpredictable medical risks. The autofluorescence characteristics also show general potential

for locating tumor tissues and real-time tracing drug delivery. The developed functional system provides promising applications in future cancer therapy by integrating cancer diagnosis and collaborative therapy.

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