**ORIGINAL PAPER**



# **Micro‑habitat features determine oviposition site selection in High Brown and Dark Green Fritillaries**

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#### **Abstract**

The survival of butterfy populations depends on successful oviposition strategies. The limited mobility of early life stages requires females to select sites that refect larval requirements. However, as land use and climate changes are altering habitat conditions and micro-climate, some species may adapt ovipositing strategies and fourish while others, with narrow niche requirements, may be unable to respond. Oviposition site selection and micro-habitat niche is examined for two closely related butterfy species—the specialist High Brown Fritillary (*Fabriciana adippe*) and relative generalist Dark Green Fritillary (*Speyeria aglaja*) through feld observations of egg-laying females and analysis of micro-habitat characteristics. A total of 104 oviposition behaviour observations across both species were recorded in 69 1  $m<sup>2</sup>$  quadrats, with the habitat characteristics compared to randomly selected quadrats in the same area. Results show that higher host plant density was a positively signifcant factor for oviposition site selection only for the High Brown Fritillary. Moreover, the cover of live Bracken (*Pteridium aquilinum*) and grass were important for site selection in both species, with High Brown Fritillaries tolerating less live Bracken and grass cover than Dark Green Fritillaries. This confrms the more specifc requirements and narrower micro-habitat niche of the High Brown Fritillary, which appears to be more sensitive to micro-habitat cooling. **Implications for insect conservation** The management of Bracken mosaic habitats for these two species should aim to supress grass growth and maintain Bracken density within limits, by opening the Bracken canopy on a rotation through grazing or manual cutting, ensuring a continuous supply of suitable micro-habitat.

**Keywords** Lepidoptera · *Fabriciana adippe* · Oviposition preference · Micro-habitat · Niche separation · Bracken

# **Introduction**

Within the extreme global declines in invertebrates (Wagner et al. [2021](#page-12-0)), Lepidoptera are one of the most afected taxa (Sánchez-Bayo and Wyckhuys [2019\)](#page-12-1) with declines reported across Europe (Maes and Van Dyck [2001;](#page-11-0) van

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Swaay et al. [2010](#page-12-2); Stefanescu et al. [2011;](#page-12-3) Warren et al. [2021](#page-12-4)), North America (Forister et al. [2011;](#page-11-1) Wepprich et al. [2019](#page-12-5)) and Asia (Nakamura [2011](#page-11-2); Choi and Kim [2012\)](#page-11-3). Butterfy declines are attributed to agricultural intensifcation, abandonment and fragmentation at the landscape level (van Swaay et al. [2010](#page-12-2)) along with direct habitat loss, changes in management practices and habitat quality at the local level. Within this order, it is the habitat specialists that have disproportionately declined (van Swaay et al. [2006](#page-12-6); Stefanescu et al. [2011\)](#page-12-3) and are more at risk from extinction on a local level (Ellis et al. 2012). An understanding of the autecology of all life-stages is required for successful conservation of lepidoptera, particularly since most species typically spend much of their life cycle in the juvenile stages. Indeed, the requirements of eggs and larvae can be more specifc than that of adults (Smee et al. [2011](#page-12-7)), highlighting the greater need to defne habitat quality for these immature stages prior to habitat management (Thomas et al. [2011\)](#page-12-8).

The limited mobility of early life-stages implies that successful oviposition strategies are critical for the survival of butterfy populations (García-Barros and Fartmann [2009](#page-11-4)). These strategies involve complex processes to select sites with optimal conditions that promote the development of the ofspring. This site selection occurs at the landscape, patch, and micro-habitat scale and can be infuenced by human induced environmental change (García-Barros and Fartmann [2009](#page-11-4)).

A butterfly will detect a suitable site for oviposition through chemical, visual and thermal cues, assessing the habitat, micro-habitat and prospective host plant (Singer [2004;](#page-12-9) García-Barros and Fartmann [2009;](#page-11-4) Eilers et al. [2013](#page-11-5)). The most common strategy is to lay eggs directly on the larval food plant, enabling emerging larvae to feed without depleting their energy reserves (Wiklund [1984\)](#page-12-10). However, species which overwinter in the egg stage and whose larvae feed on herbaceous plants which die out in winter deviate from this approach. Such species avoid laying on the host plant, choosing instead to lay on a sturdy substrate nearby (Wiklund [1984;](#page-12-10) Barnett and Warren [1995;](#page-10-0) Kopper et al. [2000](#page-11-6)).

Such a life-history strategy, with overwintering eggs laid on nearby substrate, is rare with few species recorded in the literature. The only UK butterfy to adopt this strategy is the High Brown Fritillary (*Fabriciana adippe*, Dennis & Schiffermüller, 1775). It is a habitat specialist occupying sheltered south-facing Bracken [*Pteridium aquilinum* (L.) Kuhn] dominated slopes interspersed with grassy areas (Barnett and Warren [1995\)](#page-10-0). Egg laying occurs in gaps in the Bracken canopy, where Bracken litter is dominant and grass is sparse (Warren [1995a](#page-12-11)). In NW England it also utilises early successional habitat in woodland clearings and rides, where egg laying occurs in short vegetation near rock outcrops, with a good cover of moss and sparse grass. These micro-habitats are warmer than ambient temperature and surrounding vegetation on sunny days (Warren [1995a\)](#page-12-11).

The larvae of the High Brown Fritillary hatch in March to feed on the primary larval food plant, Common Dog violet (*Viola riviniana* Rchb.), however Hairy violet (*Viola hirta* L.) is also used in NW England (Warren et al. [1995](#page-12-12)). To aid development, larvae bask on moss or Bracken litter which can be  $15-20$  °C warmer than the surrounding short grassy vegetation (Asher et al. [2001](#page-10-1)). Previous studies have observed larvae in similar habitats to those selected for oviposition (Warren [1995a](#page-12-11)). The visually similar Dark Green Fritillary (*Speyeria aglaja*, Linnaeus, 1758) inhabits a broadly similar habitat although is a relative generalist occupying a wider range of habitats (Polic et al. [2021](#page-11-7)). The Dark Green Fritillary overwinters as larvae and tolerates a cooler vegetation for breeding (Asher et al. [2001](#page-10-1); Ellis et al. n.d.). It also utilises *Viola* spp. as the primary larval host plant in Bracken habitats, although is also known to feed on Bistort [*Persicaria bistorta* (L.) Samp.] in humid grasslands (Fric et al. [2005;](#page-11-8) Zimmermann et al. [2009\)](#page-12-13).

The Dark Green Fritillary is not of conservation concern in Europe due to its stable range but is Near Threatened in the UK due to reductions in distribution with signifcant range contractions over the last 10 years (Fox et al. [2022](#page-11-9)). In contrast, the High Brown Fritillary has sufered declines across Europe, most markedly in the UK, with a severe range contraction  $[-82\% \text{ since } 1985 \text{ (Fox et al. } 2023)]$  $[-82\% \text{ since } 1985 \text{ (Fox et al. } 2023)]$  $[-82\% \text{ since } 1985 \text{ (Fox et al. } 2023)]$  and decrease in abundance [− 65% since 1978 (Fox et al. [2023](#page-11-10))]. As a result, it is the UK's fastest declining butterfy (Ellis et al. [2015\)](#page-11-11), is recognised as Endangered on the Red List of British butterfies (Fox et al. [2022](#page-11-9)) and is protected in UK legislation [Natural Environment and Rural Communities Act (2006); Environment (Wales) Act (2016); Wildlife & Countryside Act (1981)].

Research into these two species typically focuses on dispersal and colonisation (Six [2000;](#page-12-14) Cowley et al. [2001a,](#page-11-12) [2001b;](#page-11-13) Zimmermann et al. [2009;](#page-12-13) Nesbitt [2010](#page-11-14); Polic et al.  $2021$ ). The difficulties of direct observation of oviposition for these species has resulted in only a few published accounts (Thomas [1991](#page-12-15); Zimmermann et al. [2009\)](#page-12-13) and limited research into micro-habitat selection (Warren [1995a,](#page-12-11) [1995b;](#page-12-16) observations  $n=22$ ,  $n=27$  respectively). The females of both species are strong fiers and can rapidly drop into dense Bracken stands making them difficult to observe. Female High Brown Fritillaries will alight on host plants and non-host plants alike to detect suitable oviposition sites (Wiklund [1984\)](#page-12-10). Egg laying occurs on sturdy substrate most often after crawling over the leaf of the nearby host plant, although occasionally eggs are laid soon after landing, before encountering any *Viola* spp. (Barnett and Warren [1995](#page-10-0); Warren [1995a\)](#page-12-11).

Studying the habitat quality at the site of oviposition in two similar species within the same habitat will provide a new understanding of micro-habitat requirements and niche separation. We predict that oviposition sites of both species will have higher density of host plants than the surrounding habitat. Based on the acknowledged diferences in larval thermal preferences, it is expected that High Brown Fritillary females will choose oviposition micro-habitats with greater bracken litter cover and lower cover of grass that will aid micro-climatic warming. In contrast, the relatively generalist Dark Green Fritillaries will be less specifc in choosing oviposition sites using a broader range of micro-habitats.

# **Methodology**

#### **Study area**

Data was collected within the Morecambe Bay area (54°12′07″N, 002°42′07″W), a lowland landscape in NW England characterised by farmland, limestone grasslands, broadleaved woodland and exposed limestone pavements (Natural England [2022](#page-11-15)). This area accounts for most of the UK High Brown Fritillary colonies (Jones et al. [2015](#page-11-16)) and the Dark Green Fritillary is also present.

The colony inhabiting the adjacent management units of Holme Stinted Pastures and the south-western corner of Holme Park Fell was chosen for this study due to its medium population size and recently stable numbers of High Brown Fritillary (Ellis and Wainwright [2008](#page-11-17)). The area is approximately 20 ha, with an elevation of 130–170 m a.s.l and is predominately south-facing. The principal habitat is Bracken and limestone grassland mosaic with scrub (Online Resource, Figure S1). Prior to 2000, Holme Park Fell was heavily grazed by sheep and now has reduced stocking densities of cattle and sheep, whereas Holme Stinted Pastures had historically low stocking densities, with a scrub management programme introduced since 2008 (Ellis and Wainwright [2008\)](#page-11-17). Both management units are now in agrienvironment schemes, are grazed by cattle, and a programme of scrub removal and light Bracken control aimed at increasing breeding habitat has been implemented in recent years.

# **Oviposition observation**

Due to habitat variability across the site, the study area was divided into thirteen sections of potentially suitable habitat. Each section was surveyed for male and female adults by walking a set route on days with dry weather above 13 °C from the beginning of the short fight period of both species. The entire route through all sections was walked 13 times over 10 days between 11th June & 14th August in 2020, and 22 times over 14 days between 12th June & 4th August in 2021, after which sections with the most sightings of adults were targeted to maximise the chance of observing oviposition behaviour. Species ID was carried out in the feld using binoculars & photography to determine species and sex.

After locating a female of either species, they were followed from a distance of at least 3 m for up to 30 min, until they settled or were lost from sight. Upon settling they were approached to 1 m (Kopper et al. [2000\)](#page-11-6) and observed for oviposition behaviour. Oviposition behaviour was determined as an observed and defnite downward turn of the abdomen accompanied by probing of the substrate. Oviposition behaviour sites were waymarked using a handheld GPS device and a coloured peg was placed as close to the location as possible without causing disturbance (Online Resource, Figure S2). Eggs were searched for and only found occasionally. Due to this lack of verifcation of oviposition the marked sites represented oviposition behaviour. If an individual showed no oviposition behaviour within the 30 min further females would be searched for in the same site section (Henry and Schultz [2013](#page-11-18)). If both species were observed at the same

time, preference was given to following High Brown Fritillaries due to the conservation status of this species.

To further verify oviposition success, larval feeding damage to the host plant was recorded the following year on 11th June 2021 & 7th June 2022 in each of the observed oviposition behaviour sites. Larvae were considered present if there were any *Viola* spp. plants with heavy damage (>30% of a leaf) or more than 4 plants with medium damage (11–30% of a leaf) within the quadrat. Due to the presence of several *Viola*-feeding fritillary species on site, the species causing the damage could not be determined. Larval searches were carried out, but observations were low  $(n=1)$ , therefore, larval performance could not be accurately assessed. No significant difference  $(P > 0.05)$  was found in the habitat between oviposition behaviour sites with confrmed eggs or larval damage and those without (Online Resource, Table S3 & 4). This suggests that in the absence of oviposition verifcation, the observation of oviposition behaviour is an appropriate method for determining site selection of ovipositing females for these species.

## **Micro‑habitat data collection**

Within two weeks of the observation a  $1 \text{ m}^2$  quadrat was placed centrally around each point of oviposition behaviour, where the edges were perpendicular to cardinal directions. Oviposition behaviour by the same individual that fell within 30 cm of each other were recorded within the same quadrat to minimise overlap of quadrats.

To determine diferences in micro-habitat preferences between both species, a range of variables which afect the micro-habitat temperatures through shading and insulation, or the ability of the host plant to develop through competition and resource limitation, were measured in each quadrat (Online Resource, Table S5). These were leaf litter depth and percentage cover of Bracken, Bracken litter, grass, moss, bare ground and rock, and Bramble. To determine the infuence of host plant density, the number of *Viola* spp. per m<sup>2</sup> and the distance from the point of oviposition behaviour to the nearest *Viola* spp. plant (mm) were measured. In quadrats with more than one oviposition behaviour observation, measurements taken at the site of oviposition behaviour were averaged to gain one reading.

To determine micro-habitat preference within each species, randomly generated locations of the wider available habitat in each section of the site were surveyed as controls. Randomly generated locations were rejected for survey if they were in areas of dense trees or scrub over 2 m as this did not represent the habitat utilised by either species. To avoid surveyor bias and for ease of resurveying, the  $1m<sup>2</sup>$  quadrat was placed to the north-east of each randomly generated control location with the edges perpendicular to cardinal directions. Control locations were surveyed for the same

variables within four weeks of the frst oviposition behaviour in each section. This led to a total of 97 control locations, refecting the number of oviposition behaviour observations in each section as well as 28 locations in an additional section where fritillaries had been observed regularly but no oviposition behaviour recorded.

#### **Data analysis**

Analyses were carried out with R software, version 4.2.1 (R Core Team [2022](#page-12-17)).

To determine if the butterfies select distinct micro-habitats for oviposition, Principal Component Analyses was used to compare the microhabitat variables at oviposition behaviour sites with those at control locations. One PCA was carried out for each species using the RDA function in the 'vegan' package (Oksanen et al. [2022\)](#page-11-19) and the variables were standardised by mean zero and unit variance.

To test if oviposition behaviour is determined by host plant density and other micro-habitat variables, a binary generalized linear model (GLM) with model averaging was carried out for each species. Prior to modelling, colinearity between variables was checked using the variance infation factor (VIF) function within the 'car' package (Fox and Weisberg [2019](#page-11-20)). Variables with the highest VIF were sequentially dropped until all remaining variables had a VIF value<3 (Zuur et al. [2010](#page-12-18)) (Online Resource, Tables S6 & S7). The rcorr function within the 'Hmisc' package (Harrell Jr [2022](#page-11-21)) was also used to calculate the correlation coefficient (rho) between response variables for each species (Online Resource, Figures S8 & S9). Any remaining variables with a high correlation (rho $> \pm 0.7$ ) were assessed for inclusion in the models and the least important variable in terms of biological rationale was removed. The fnal variables selected for modelling in both species were *Viola* spp. density, average litter depth, cover of Bracken, grass, moss, bare ground and rock, and Bramble (*Rubus fruticosus* agg.) (Online Resource, Table S5). Quadratic terms were included for cover of Bracken and grass for both species and Bramble for Dark Green Fritillary which displayed non-linear relationships. A binomial error distribution was applied to each model with binary response variables  $(0=control, 1=ovi$ position behaviour observation in each species respectively). Three global models were produced for each species to enable the selection of the most appropriate link function with the lowest residual deviance. 'Probit' link function was used for the models of both species.

Multi-model inference with the 'MuMin' package (Barton [2022\)](#page-10-2) was used to assess all possible models for each species as the large combination of variables produced too many models to assess manually. Models were standardised for by mean zero and unit variance. The model with the best ft was determined by the lowest AICc. All models within AICc $\leq$  2 of the best model for each species were checked prior to model averaging with the Hosmer–Lemeshow goodness of ft test using the Hoslem.test function in the 'ResourceSelection' package (Lele et al. [2019](#page-11-22)). All models showed no signifcant diference between the model and the observed data for each species  $(P>0.05)$  and thus represented a good ft. The fnal 'best' model for each species was validated by replacing correlated variables to ensure that the removed variables did not improve the AICc. All variables were assessed for relative variable importance (RVI) based on the sum of the Akaike weights  $(w_i)$  from all possible models for each species (Burnham and Anderson [2002](#page-11-23)). The inclusion of quadratic terms resulted in each variable appearing in an unequal number of models, therefore RVI was divided by the number of models each variable appeared in to gain an adjusted RVI (Kittle et al. [2008\)](#page-11-24).

To explore niche separation, PCA was used to compare the microhabitat variables at the oviposition behaviour sites of each species, as previously described. To determine if the species select diferent microhabitats for oviposition, each habitat variable was tested for signifcant diference using either a Two sample t-test or a non-parametric Wilcoxon rank sum test.

## **Results**

#### **Oviposition observation**

Sixty-fve High Brown Fritillary and thirty-nine Dark Green Fritillary oviposition behaviours were recorded in 69 1  $m<sup>2</sup>$ quadrats across 9 sections of the site (Table [1;](#page-4-0) see Online Resource for further details). Following searches at each location no eggs were found in 2020 while five Dark Green and two High Brown Fritillary eggs were found in 2021. Two of the Dark Green Fritillary eggs were found within the same quadrat on two separate *Viola* spp. plants. All quadrats with eggs had heavy larval damage the following spring and a further twelve Dark Green and seventeen High Brown Fritillary oviposition quadrats also had larval damage. 33% of Dark Green Fritillary and 58% of High Brown Fritillary quadrats did not have signifcant feeding damage or eggs. It could not be confrmed which species or which individual caused the feeding damage and it is possible that unobserved oviposition events by other individuals occurred in the same locations.

#### **High Brown Fritillary oviposition site selection**

A PCA of all habitat variables at High Brown Fritillary oviposition behaviour sites and across the control locations accounted for 49% of the variation in this dataset in PC1 (35%) & PC2 (14%) (Fig. [1;](#page-4-1) Online Resource, Table S10).

<span id="page-4-0"></span>**Table 1** Oviposition behaviour, egg, and larval damage observations for High Brown Fritillary (HBF) and Dark Green Fritillary (DGF) distributed by quadrat and site section during 2020 & 2021

Site section	No. oviposition behaviour observa- tions	No. ovi- position quadrats	No. oviposition quadrats with eggs or larval damage	No. control quadrats	Total No. quadrats surveyed		Site section No. oviposition behaviour obser- vations	No. oviposi- tion quadrats
	HBF	<b>DGF</b>	<b>HBF</b>	$\overline{DGF}$	HBF	DGF		
HSP <sub>0</sub>						$\Omega$		h
HSP <sub>1</sub>	25		20		9	4	25	50
HSP <sub>2</sub>	8						9	18
HSP <sub>3</sub>	25		14			3	19	38
HSP <sub>3b</sub>	$\overline{c}$							8
HSP <sub>4</sub>	0			6	0	6	h	12
HSP <sub>6</sub>	0				0	$\Omega$		
HPF <sub>2</sub>	0					$\Omega$		
HPF <sub>3</sub>	2				0	$\Omega$		
HSP 7 <sup>a</sup>	0					$\Omega$	28	28
Total	65	39	45	24	19	16	97	166

The number of control quadrats for each section are also shown

*HSP* Holme Stinted Pastures*, HPF* Holme Park Fell

<sup>a</sup>No oviposition was observed in this section but fritillaries were regularly present

<span id="page-4-1"></span>**Fig. 1** Principal Component Analysis of High Brown Fritillary oviposition behaviour sites and control locations, using the full set of variables.  $PC1 = 35\%$ ,  $PC2 = 14%$ 



Control locations were distributed across the entire spread of the oviposition behaviour sites in ordination space and thus are a true representation of the available habitat. High Brown Fritillary oviposition behaviour showed a negative association with PC2, which is negatively associated with cover of bare ground and rock and *Viola* spp. density and positively associated with cover of Bramble and leaf litter depth. Within the observed oviposition behaviour sites there was no separation of quadrats with confrmed eggs or larval damage and those without.

Binary GLMs revealed that the best model for predicting oviposition site selection in High Brown Fritillaries included cover of Bracken and cover of grass (both quadratic and linear terms) and *Viola* spp. density. In this model the quadratic term for cover of Bracken and linear term for cover of grass were significant at the  $P < 0.001$  level, whereas *Viola* spp. density was significant at the  $P < 0.05$ level (Table [2](#page-5-0)). The adjusted RVI revealed cover of Bracken and grass to be the most important. Multi-model inference revealed three other valid models within  $AICc \leq 2$  of the

	Best model (AICc 87.58)			Model average $(3 \text{ models} \leq 2AICc)$				RVI Number of models	Adjusted RVI <sup>a</sup>
	Estimate $(\pm SE)$		z value $Pr(>\vert z \vert)$	Estimate $(\pm SE)$		z value $Pr(>\vert z \vert)$		(N)	
(Intercept)	1.011 $(\pm 0.352)$		$2.87$ 0.004**	$1.040 \ (\pm 0.360)$	2.87	$0.004**$			
Cover of Bracken ^2	$-4.556 \ (\pm 1.118)$		$-4.08$ 0.00005***	$-4.570 \ (\pm 1.118) \quad 4.05$		$0.00005***$	1.00 96		1.04
Cover of grass $\sqrt{2}$	$-4.376$ $(\pm 1.459)$		$-3.00$ 0.003**	$-4.514 \left( \pm 1.504 \right)$ 4.62		$0.000004***$	0.99 96		1.03
<i>Viola</i> spp. density	1.172 $(\pm 0.540)$		2.17 0.030*	4.204 $(\pm 1.078)$	2.98	$0.003**$	0.86 144		0.60
Cover of Bracken	$-2.125$ $(\pm 0.499)$			$-4.26$ 0.00002*** $-2.179$ ( $\pm$ 0.518) 4.17		$0.00003***$	1.00 192		0.52
Cover of grass	$-1.902$ $(\pm 0.508)$		$-3.74$ 0.0002 ***	$-1.867 \left( \pm 0.516 \right)$ 3.59		$0.0003***$		1.00 192	0.52
Cover of Bramble	$\overline{\phantom{0}}$			$-0.080 \ (\pm 0.233) \ \ 0.34$		0.732	0.34 144		0.24
Cover of moss				$-0.048 \ (\pm 0.203) \quad 0.24$		0.813	0.28 144		0.19
Cover of bare ground $-$ & rock							0.25 144		0.17
Leaf litter depth							0.25 144		0.17

<span id="page-5-0"></span>**Table 2** Binary linear regression and model averaging of High Brown Fritillary oviposition behaviour sites versus available habitat

Relative variable importance is given for all variables used in searching for the best model

\*P<0.05, \*\*P<0.01, \*\*\*P<0.001

a RVI/N\*100

best model. Model averaging of these four models showed that cover of Bracken, cover of grass (P < 0.001) and *Viola* spp. density  $(P < 0.01)$  were significant in the selection of oviposition sites by the High Brown Fritillary, with relative variable importance indicating *Viola* spp. density to be the least important variable of the three. Cover of Bramble and cover of moss were included in the averaged model but were not found to be signifcant (Table [2](#page-5-0)).

Visualisation of the best model reveals the non-linear relationship between the explanatory variables of Bracken and grass cover and the probability of oviposition behaviour in the High Brown Fritillary (Fig. [2\)](#page-6-0). The probability of oviposition behaviour increases rapidly with the cover of Bracken, reaching more than 0.8 between 30 and 65% cover of Bracken. Above 65% Bracken cover probability rapidly decreases. The probability of oviposition behaviour reaches more than 0.8 when grass cover is between 15 and 35%. Above 45% grass cover probability rapidly decreases. The presence of *Viola* spp. gives a nearly 0.8 probability of oviposition behaviour by High Brown Fritillary. As the number of *Viola* spp. increases to 100 per m<sup>2</sup> probability of oviposition behaviour rises to 0.9.

# **Dark Green Fritillary oviposition site selection**

A PCA of all habitat variables collected at Dark Green Fritillary oviposition sites and across the control locations accounted for 54% of the variation in this dataset in PC1 (39%) & PC2 (15%) (Fig. [3](#page-7-0); Online Resource, Table S11).

Control locations were distributed across the entire spread of the oviposition behaviour sites in ordination space and thus are a true representation of the available habitat. Dark Green Fritillary oviposition behaviour was negatively associated with PC1, which is negatively associated with cover of grass, *Viola* spp. density and cover of moss, and positively associated with cover of Bracken, cover of Bracken litter and distance to nearest *Viola* spp.. Within the observed oviposition behaviour sites there was no separation between quadrats with confrmed eggs or larval damage and those without.

The best model for predicting oviposition site selection in Dark Green Fritillaries included cover of Bracken and cover of grass (both quadratic and linear terms) (Table [3](#page-7-1)). The quadratic & linear terms for cover of Bracken were significant at the  $P < 0.001$  level, and the linear term for cover of grass was significant at the  $P < 0.05$  level. The adjusted RVI revealed cover of Bracken and grass to be the most important. Multi-model inference revealed six other valid models within  $AICc \leq 2$  of the best model. Model averaging also showed that quadratic and linear terms cover of Bracken  $(P<0.05)$  and the quadratic term for grass  $(P<0.01)$  were signifcant in the selection of oviposition sites for the Dark Green Fritillary. Other variables were included in the averaged model but were not signifcant (Table [3\)](#page-7-1).

Visualisation of the best model reveals the non-linear relationship between the significant explanatory variables and the probability of oviposition behaviour in the Dark Green Fritillary (Fig. [4\)](#page-8-0). The probability of oviposition behaviour reaches more than 0.7 between 25 and



<span id="page-6-0"></span>**Fig. 2** Conditional plots of the relationship between probability of High Brown Fritillary oviposition behaviour and each signifcant explanatory variable including; **a** % cover of Bracken  $(P < 0.001)$ , **b** % cover of grass (P<0.001), and **c** *Viola* spp. density (P<0.05), as estimated by the best model  $(AICc=87.58)$ 

55% cover of Bracken, above which probability rapidly decreases. The probability of oviposition behaviour is greatest when grass cover is between 30 and 60%, above which probability slowly decreases.

#### **Micro‑habitat niche‑separation**

A PCA of all habitat variables collected at High Brown Fritillary and Dark Green Fritillary oviposition behaviour sites accounted for 40% of the variation in this dataset in PC1  $(25\%)$  & PC2  $(15\%)$  (Fig. [5\)](#page-8-1). Within the observed oviposition behaviour sites there was no separation of sites with confrmed eggs or larval damage for either species, although there was separation of oviposition behaviour sites between the species. The separation was predominantly associated with PC1, most explained by cover of Bracken litter, cover of Bracken, and cover of grass. Dark Green Fritillary oviposition was also positively associated with PC2, most explained by *Viola* spp. density, cover of grass and cover of Bramble (Online Resource, Table S12).

Assessment of individual habitat variables between oviposition behaviour sites of the two species showed that there was a signifcant diference in the cover of Bracken  $(t=- 2.2318; df=67; P=<0.05)$ , with High Brown Fritillaries choosing sites with more Bracken than Dark Green Fritillaries (Fig. [6a](#page-9-0)). Cover of grass differed significantly between the two species' oviposition behaviour sites  $(W=912.5; N=69; P = <0.001)$ , with High Brown Fritillaries choosing sites with less grass cover than Dark Green Fritillaries (Fig. [6b](#page-9-0)). There was a significant difference in the cover of Bracken litter between the two species' oviposition behaviour sites (W = 264; N = 69; P =  $<$  0.005), with High Brown Fritillaries choosing sites with more Bracken litter than Dark Green Fritillaries (Fig. [6](#page-9-0)c). Cover of Bramble difered signifcantly between the two species oviposition behaviour sites (W = 697; N = 69; P =  $<$  0.05), with High Brown Fritillaries choosing sites with less Bramble cover than Dark Green Fritillaries (Fig. [6](#page-9-0)d). *Viola* spp. density did not significantly differ  $(P>0.05)$  between the oviposition behaviour sites of the two species (Fig. [6](#page-9-0)e). Leaf litter depth, cover of moss, and cover of bare ground and rock were also not signifcantly diferent (all P-values>0.05, see Online Resource, Figure S13).

# **Discussion**

#### **Oviposition site selection**

Oviposition site selection by High Brown Fritillary was signifcantly afected by the cover of live Bracken, the cover of grass and the density of *Viola* spp., whilst oviposition site selection by the Dark Green Fritillary was also signifcantly infuenced by the cover of live Bracken and the cover of grass, but not by density of the host plant. These variables presumably affect the development of larvae through changes in micro-habitat temperature and resource availability and the infuence of host plant density for the <span id="page-7-0"></span>**Fig. 3** Principal Component Analysis of Dark Green Fritillary oviposition behaviour sites and control locations using the full set of variables.  $PC1 = 39\%$ ,  $PC2 = 15%$ 



<span id="page-7-1"></span>**Table 3** Binary linear regression and model average of Dark Green Fritillary oviposition behaviour sites versus available habitat



Relative variable importance is given for all variables used in searching for the best model

a RVI/N\*100

 $*P<0.05$ ,  $*P<0.01$ ,  $**P<0.001$ 

High Brown Fritillary suggests this species has additional requirements.

Warm micro-habitats with larval host plants promote faster larval development through increased (Rytteri et al.  $2021$ ) and more efficient feeding (Porter [1982](#page-12-20)), and in turn can lower the risk of predation and parasitism (Benrey and Denno [1997\)](#page-10-3). Due to micro-climatic cooling by tall, green vegetation (WallisDeVries [2006;](#page-12-21) WallisDeVries and van Swaay [2006](#page-12-22)), it was expected that oviposition behaviour would be signifcantly negatively infuenced by an increase in cover of grass. The results of this study show that, whilst both species tolerate some grass, the chance of oviposition behaviour decreases rapidly beyond a threshold of approximately 45% cover for the High Brown Fritillary and slowly



<span id="page-8-0"></span>**Fig. 4** Conditional plots of the relationship between probability of Dark Green Fritillary oviposition behaviour and each signifcant explanatory variable including  $a \%$  cover of Bracken (P<0.001) and **b** % cover of Grass ( $P < 0.01$ ), as estimated by the best model  $(AICc=83.88)$ 

<span id="page-8-1"></span>**Fig. 5** Principal Component Analysis of High Brown Fritillary & Dark Green Fritillary oviposition sites, using the full set of variables.  $PC1 = 25\%,$ 

 $PC2 = 15%$ 

beyond 60% for the Dark Green Fritillary. Similarly, both species selected sites with an intermediate cover of live Bracken (High Brown Fritillary: 35–65%, Dark Green Fritillary: 25–55%), suggesting intermediate coverage is optimal. These fndings for the High Brown Fritillary align with Warren [\(1995a\)](#page-12-11) where, in Bracken habitats in NW England, oviposition was observed in locations with a Bracken canopy between 30 and 70% and grass cover of 12–40%.

Micro-habitat preferences at the time of oviposition may refect the larval preference for specifc micro-climatic conditions. A dense canopy of Bracken is not present in the early spring when larvae are developing thus not likely to infuence the performance of larvae, although females using thermal cues to fnd optimal conditions for their ofspring could still be deterred by such cool environments in the summer.

Tolerating some microclimatic cooling during oviposition may be a trade-off as the amount of shade produced by standing Bracken provides a canopy for *Viola* spp., a species which thrives in woodlands (Warren and Oates [1995](#page-12-23); Ellis [2005](#page-11-25)). Too little Bracken may be associated with a limited production of Bracken litter and increased presence of grass, which will further cool the micro-habitat at oviposition sites. Due to the absence of a Bracken canopy in the spring, when habitat assessments for the two species are normally carried out, the amount of standing live Bracken has not been previously identifed as a signifcant factor in micro-habitat quality for either species.

The ability of Bracken litter to reach and maintain the optimum temperature for larvae (32 °C) on mild days (Warren [1994\)](#page-12-24) and the numerous accounts of larvae found





<span id="page-9-0"></span>Fig. 6 Boxplots of five habitat variables; a cover of Bracken<sup>a</sup>; b cover of grass<sup>b</sup>; **c** cover of Bracken litter<sup>b</sup>; **d** cover of Bramble<sup>a</sup>; and **e** *Viola* spp. density. Control locations were not included in analysis.  $\text{DGF}=\text{Dark}$  Green Fritillary oviposition behaviour sites (n=24),

HBF=High Brown fritillary oviposition behaviour sites  $(n=45)$ , Control=control locations (n=97). <sup>a</sup>Significant difference (P<0.05) between species. <sup>b</sup>Significant difference (P<0.001) between species

basking in these locations would indicate that it would be infuential in oviposition site selection. As such, it was expected that Bracken litter could positively infuence site selection but due to the co-variation with standing live Bracken, this was not displayed by the models. The preference for standing live Bracken over Bracken litter may refect a diference in visual apparency of these two variables to ovipositing females.

Survival for the relatively immobile early instars is reliant on females selecting oviposition sites within reach of a food resource. Host plant density is shown to positively infuence occurrence, oviposition and emigration in a range of lepidoptera (Menéndez et al. [2002;](#page-11-26) Betzholtz et al. [2007;](#page-11-27) Salz and Fartmann [2009;](#page-12-25) Smee et al. [2011](#page-12-7); Ewing et al. [2020\)](#page-11-28). *Viola* spp. density was an important factor in site selection only for the High Brown Fritillary, however there was no diference in the density of host plants between the sites selected by the two species. The indication that *Viola* spp. density is not an important factor in site selection for the Dark Green Fritillary species is unexpected due to the lack of alternative host plants within Bracken habitats.

*Viola* spp. are not uncommon plants, nor are they restricted to just one habitat. Therefore, the presence of the plants alone is not likely a driving factor in the occurrence of each species. The relative importance of the signifcant variables indicates that High Brown Fritillary females may prioritise habitat structure and composition as an oviposition cue whilst only secondarily assess host plant density. Such prioritisation has been evidenced in other Lepidopteran species (Friberg et al. [2008b](#page-11-29)), although this may not be applicable if host plant density is too low.

If faced with an abundance of host plants, other factors of host plant quality may become infuential in oviposition site selection, this may include more apparent features including host plant size (Anthes et al. [2003\)](#page-10-4) or colour (Myers 1985; Stefanescu et al. 2006). Less apparent physiological measures, such as levels of carbon, nitrogen and phosphorous (Myers 1985; Salz and Fartmann [2009\)](#page-12-25) and defensive compounds (García-Barros and Fartmann [2009](#page-11-4)) could also indirectly infuence oviposition site selection. Consequently, host plant selectiveness may change over time with agricultural intensifcation and anthropogenic nitrogen deposition.

# **Micro‑habitat niche‑separation**

Micro-habitat is of importance for adults of both species when selecting oviposition sites, but our results indicate that the High Brown Fritillary requirements are more specialised (Barnett and Warren [1995a,](#page-12-11) [b](#page-12-16)). Dark Green Fritillaries chose oviposition sites with a signifcantly higher cover of grass and a lower cover of Bracken litter and standing live Bracken than High Brown Fritillaries, affirming the higher tolerance of Dark Green Fritillaries to micro-climatic cooling. Whilst a denser Bracken canopy will cool the micro-climate in summer, the amount of litter that it produces in autumn, together with its ability to supress grass growth, is also of importance to the High Brown Fritillary (Warren and Oates [1995\)](#page-12-23).

The wider variety of suitable habitat types available to the Dark Green Fritillary within its range (Zimmermann et al. [2009](#page-12-13); Polic et al. [2021](#page-11-7)) suggests a wider habitat niche than the specialist High Brown Fritillary. The results of this study also confrm signifcant diferences in micro-habitat niche between the two species within their overlapping range. The wider niche of the Dark Green Fritillary could be advantageous for persistence at the landscape scale as the need for long distance dispersal will be reduced when faced with changes in micro-habitat (Six [2000;](#page-12-14) Polic et al. [2021\)](#page-11-7). However, it is important to take into account that niche separation between two closely related species has been shown to vary with local habitat variation across geographic ranges (Friberg et al. [2008a\)](#page-11-30), and that oviposition preference in High Brown Fritillaries may difer between recently cleared scrub and Bracken habitats (Warren [1995a](#page-12-11)). As such, oviposition preference could difer in a more ubiquitous habitat, where Bracken is more extensive and recently removed scrub is not a key part of the available habitat.

#### **Conservation implications**

Management of Bracken mosaic habitats for the High Brown and Dark Green Fritillary should aim to restore and maintain micro-habitat with a combination of 35- 65% cover of Bracken, less than 45% cover of Grass and a high density of *Viola* spp. (> 100/m<sup>2</sup>). These limits should provide suitable breeding habitat for both species to co-exist in the same habitat area, although the extent of suitable micro-habitat may need to be considered at the site and landscape scale to support viable metapopulations.

The seemingly contradictory habitat requirement of less than 65% cover of live Bracken and a good cover of Bracken litter demonstrates the fne line in management of Bracken for the more specialised High Brown Fritillary. The areas on site where this occurred were largely where cattle had created paths through Bracken in the summer months, keeping the Bracken sward open and breaking down the previous year's litter. Where grazing is not possible, cutting paths through Bracken early in the fight season would open dense swards enabling warming of Bracken litter below. This would also enable regrowth later in the season to supress grass growth and maintain a supply of Bracken litter for the following breeding season. To enable a continuous supply of this habitat, it is likely that the network of paths would need to change year on year as paths remaining open for too long can quickly be invaded by grass. Stocking densities should be low enough for this to happen and cutting will need to be on rotation.

Due to the conservation status and specialised habitat requirements of the High Brown Fritillary, management should be directed towards this species, however the Dark Green Fritillary and other *Viola*-feeding fritillaries with wider niches (e.g. Pearl-bordered and Small Pearl-bordered Fritillaries) are also likely to beneft (Ellis et al. [2019\)](#page-11-31).

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**Ethical approval** The authors confrm that they follow the rules of good scientifc practice and all ethical standards requested by the journal. Ethical approval was received from Edge Hill University research ethics committee, prior to data collection. Collection of habitat data was carried out with licence from appropriate governmental bodies and in an approved and appropriate manner.

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