

Biosorption capacity and kinetics of cadmium(II) on live and dead *Chlorella vulgaris*

Jinfeng Cheng¹ · Wenke Yin¹ · Zhaoyang Chang¹ · Nina Lundholm² · Zaimin Jiang¹

Received: 29 December 2015 / Revised and accepted: 10 July 2016 / Published online: 27 July 2016 © Springer Science+Business Media Dordrecht 2016

Abstract Pollution of aquatic environments with heavy metals from natural water is a serious problem because of the toxicity of heavy metals to humans, fish, and other live organisms. Cheap and environmentally friendly methods for removing heavy metals from water are therefore needed. Algae have emerged as a promising biosorbent to bioextract heavy metal ions by adsorption, and our objective was to evaluate the biosorption capacity and kinetics of cadmium ions by live and dead cells of the microalga Chlorella vulgaris. The biosorption of cadmium was assessed by varying the sorption parameters: use of dead or live material of C. vulgaris, contact time, initial metal ion concentration, and algal dosage. Cadmium ion removal was rapid with more than 95 % of total adsorption taking place in 5 min, and with equilibrium attained in 105 min. Chlorella vulgaris had high adsorption capacity for cadmium, with 96.8 and 95.2 % of the total amount of cadmium being removed by the dead algal and the live algal biomass, respectively. The biosorption capacity increased with increasing cadmium concentration, and the maximum adsorption capacity for cadmium at equilibrium was found to be 16.34 mg Cd(II) g^{-1} biomass using live C. vulgaris cells and 16.65 mg Cd(II) g^{-1} biomass using dead C. vulgaris cells. A positive correlation was found between the adsorption efficiency and (1) the concentration of Cd(II) until adsorption equilibrium of the live and dead C. vulgaris, and (2) with the adsorbent dosage of the live and dead

Jinfeng Cheng chengjinfeng@nwsuaf.edu.cn *C. vulgaris*. The adsorption efficiency was consistently above 60 % in natural water. The kinetic data showed that a pseudo-first-order model described the sorption kinetics of Cd(II) ions by live algae better than a pseudo-second-order or an Elovich model, and use of dead algal cells was best modeled by a pseudo-second-order model. The results using both live and dead *C. vulgaris* fitted well to the Sips isotherm compared with other two-parameter (Langmuir, Freundlich) and three-parameter (Khan) isotherm models.

Keywords Chlorella vulgaris \cdot Cadmium \cdot Adsorption capacity \cdot Heavy metal

Introduction

Heavy metal pollution of air, soil, and water is a growing global problem (Matlock et al. 2002). Heavy metal contamination may have devastating effects on the environment and the diversity of aquatic organisms (Farombi et al. 2007), and heavy metals may also affect humans causing diseases such as the Minamata disease caused by mercury (Baby et al. 2010) or itai-itai caused by cadmium (WHO 2011). Serious pollution of aquatic environments due to heavy metals in natural water has recently become an increasing problem, and governmental environmental regulations have therefore been established to protect surface and ground water from heavy metals like Cd, Cu, Pb, Hg, Cr, and Fe (Matlock et al. 2002, Rai 2012, Naser 2013).

Cadmium is one of the most toxic heavy metals. Cd(II) can induce oxidative stress by generating reactive oxygen species (ROS), which may damage proteins, amino acids, nucleic acids, and membrane lipids. A decrease in biomass production, as well as photosynthesis, is a typical symptom of Cd(II) contamination in the environment (Perales-Vela et al.

¹ College of Life Sciences, Northwest A &F University, Yangling 712100, Shaanxi, China

² Natural History Museum of Denmark, University of Copenhagen, Sølvgade 83S, DK-1307 Copenhagen K, Denmark

2007; Bačkor and Fahselt 2008). Cadmium is thus extremely toxic, and because of the low permissible exposure limit, overexposures may occur even at low concentrations. WHO (2010) therefore announced that "National, regional and global actions are needed to decrease global environmental cadmium releases and reduce occupational and environmental exposure."

Removal of excess heavy metals like cadmium from natural water is thus very important. Traditional methods such as chemical precipitation, ion-exchange, adsorption, and reverse osmosis are commonly used processes for heavy metal removal, but these methods are of limited use because of technical and/or economic constraints (Ahmet et al. 2006). It is therefore crucial to explore the use of potential materials with a strong tolerance and adsorption capacity for heavy metals. Different types of materials have been used as biosorbents for removal of heavy metal, and a summary of studies on different biosorbents for Cd(II) is provided in Table 1.

Algae are photosynthetic organisms and responsible for the majority of the primary production in aquatic environments, thus playing an important role in the food webs of aquatic ecosystems. Heavy metal pollutants have been shown to depress photosynthesis, disrupt electron transport in photosystem II, reduce pigment concentrations, and affect the permeability of the plasma membrane in higher plants and microalgae (Xia et al. 2004). Heavy metals have also been found to adsorb on algal cells (Xia et al. 2004). When compared to other microbial organisms such as yeast and different fungi, heavy metal adsorption capacity of algae proved to be the highest (Gupta et al. 2015a, 2015b), because of the algal cell wall being composed of a fiber-like structure and an amorphous embedding matrix of various polysaccharides (Bayramoğlu and Arica 2009). Several functional chemical groups on the algal cell surface can attract and sequester heavy metal ions (Lagoa and Rodrigues 2007), and the process of adsorption is often relatively inexpensive, non-hazardous, and may permit recovery of the metals from the adsorbing biomass (Bayramoğlu and Arica 2008).

Adsorption is the process by which a pollutant is removed by an adsorbent, and thermodynamics and kinetics of the process are important for comparing details of the adsorption performance and mechanisms. From the kinetic analyses, the adsorption capacity, solute uptake rate, and the equilibrium time may be established. Kinetic models have been widely employed to describe the kinetic process of adsorption (Chen et al. 2008; Cheung et al. 2001; Aksu 2001; Qiu et al. 2009; Edris et al. 2014). An adsorption isotherm describes the equilibrium of the sorption of a material at a surface at constant temperature. From the absorption isotherm estimate, biosorption performance may be predicted and compared. Two-parameter isotherm models are available for modeling adsorption data. Two two-parameter isotherm models, namely Langmuir and Freundlich models, have often been used to describe the sorption data generated from cadmium biosorption using dead algae (Edris et al. 2014). A few three-parameter models such as Sips and Khan models also have been used to describe the biosorption isotherm (Vijayaraghavan et al. 2006).

Species of red, green, and brown algae as well as several microalgal and cyanobacterial species have been found to be able to remove heavy metals, but *Chlorella vulgaris* appears as the most promising (Edris et al. 2014; Khattar et al. 2015). *Chlorella vulgaris* is a unicellular freshwater green algae with a cell diameter of $3-8 \mu m$ and is a highly efficient photosynthetic organism. Previous studies have reported using the use of dead algae to remove heavy metals from aqueous solutions (Edris et al. 2014; Mirghaffari et al. 2015). However, little attention has been devoted to remove heavy metals from natural water and to compare the biosorption capacity and kinetic of cadmium(II) of live and dead *C. vulgaris*.

This study aimed to compare the ability of the dead and live cells of *C. vulgaris* to eliminate cadmium(II) ions from artificial and natural water. The objective was achieved by examining biosorption of cadmium(II) using *C. vulgaris* and by varying sorption parameters such as contact time, initial metal ion concentration, and algal concentration. The sorption kinetic analysis of metal ion biosorption was carried using pseudo-first-order, pseudo-second-order, and Elovich kinetic models. The equilibrium uptake of cadmium biosorption by live and dead *C. vulgaris* was compared using three two-parameter isotherm models namely Langmuri, Freundlich, and Temkin isotherm models and two three-parameter isotherm models namely Sips and Khan isotherm models by a non-linear method.

Materials and methods

Chlorella vulgaris growth conditions

A clonal culture of *Chlorella vulgaris* was established by micropipette isolation of a single cell from a polluted water sample collected in Yangling, Shaanxi province, China. The *C. vulgaris* culture was grown under sterile conditions in glass triangular flasks using BG11 medium (Stanier et al. 1971) and maintained at 25 °C in a 12:12 light/dark (L/D) cycle at a light intensity of 75 µmol photons m⁻² s⁻¹.

Batch adsorption experiments

Cd(II) solutions were prepared from 1 g L^{-1} stock solutions of $3CdSO_4 \cdot 8H_2O$ in distilled water. In the experiments, we compared adsorption by dead and live algae under different conditions, using (1) different contact time, (2) different initial cadmium concentration, and (3) different algal dosage.

Table 1 Biosorption capacity of adsorbents for removal of Cd(II) from waste water (modified from Gupta et al. 2015a, 2015b)

Adsorbents	Adsorption capacity $(mg g^{-1})$	Reference
Various biomass of plant		
Natural rice husk	73.96	Akhtar et al. 2010
Wheat straw	39.22	Farooq et al. 2011
Coconut waste	285.70	Pino et al. 2006
Orange peel (chem mod)	136.05	Sha et al. 2009
Banana peel	35.52	Memon et al. 2008
Pomelo peel	21.83	Saikaew et al. 2009
Coffee waste, raw	15.65	Azouaou et al. 2010
Tea waste	11.29	Cay et al. 2004
Pinus roxburghii bark	3.01	Padmini and Sridhar 2007
Coir pith	93.40	Kadirvelu and Namasivayam 2003
Mango peel	68.92	Iqbal et al. 2009
Bacteria		
Aeromonas caviae	155.3	Loukidou et al. 2004
Enterobacter sp.	46.2	Lu et al. 2006
Pseudomonas sp.	278.0	Ziagova et al. 2007
Staphylococcus xylosus	250.0	Ziagova et al. 2007
Streptomyces rimosus	64.9	Selatnia et al. 2004
Pseudomonas fluorescens	66.25	Yu et al. 2011
Fungi		
Saccharomyces cerevisiae (waste brewer's yeast)	15.4	Chen and Wang 2007
Baker's yeast (lab cultured)	11.63	Yu et al. 2007
Phomopsis sp. (lab cultured)	29	Saiano et al. 2005
Algae		
Ulva onoi	61.9	Suzuki et al. 2005
Gelidium sesquipedale	18.0	Vilar et al. 2006
Parthenium hysterophorous	27	Ajmal et al. 2006
Spirodela polyrhiza	36	Meitei and Prasad 2013
Spirulina	357	Solisio et al. 2008
Fucus ceranoides	90	Herrero et al. 2006
Oedogonium	88.90	Gupta and Rastogi 2008, 2009
Nostoc muscorum	666.7	Dixit and Singh 2014
Scenedesmus-24	50	Jena et al. 2015
Scenedesmus quadricauda	135.1	Mirghaffari, et al. 2015
Ectocarpus siliculosus	41	Winter, et al. 1994
Cladophora fracta	0.240	Ji et al. 2012
Inorganic		
Red mud	68.00	Vaclavikova et al. 2005
Red mud + H_2O_2	13.00	Gupta and Sharma 2002
Bagasse fly ash	2.00	Gupta et al. 2003
BF (slag)	18.72	Gupta et al. 1997

For acquiring dead algal material, live algal cells were harvested in exponential growth phase, centrifuged for 10 min at 2415×g, and dried at 110 °C for 24 h stored in the refrigerator for further study.

-

For comparing the adsorption of dead and live algae, the solutions of dead and live algae were added to 500 mL Erlenmeyer flasks containing 200 mL cadmium solution with a metal concentration of 100 mg L^{-1} , and kept shaken on an

orbital shaker at 150 rpm at 25 °C. At 5, 10, 15 min, and thereafter every 15 min until equilibrium was attained, 10 mL solution was removed and centrifuged to obtain the supernatant, and the residual Cd(II) concentration in the solution was measured with an atomic absorption spectrophotometer (Hitachi Z 2000).

For exploring the effect of increasing concentrations of adsorbate (cadmium) and adsorbent (algal biomass) on the sorption quantity and efficiency, removal of Cd(II) was studied using 0.18,1.8, and 18 mg of dry algal biomass and 5, 10, 50, 100, and 150 mg L^{-1} Cd(II) concentration. Otherwise, the conditions were as in the initial experiment. At the time of equilibrium, the residual Cd(II) concentration was measured as above. The experiment was performed with live and dead algae as above at 25 °C.

The adsorption percentage was obtained by using the following expression:

$$R(\%) = \frac{(C_0 - C_t)}{C_0} \times 100$$

where *R* is the percentage of metal adsorbed by algae, C_0 is the initial Cd(II) concentrations (mg L⁻¹), and C_t is the remaining concentrations of the metal ions at *t* time (mg L⁻¹).

Adsorption capacity calculations:

At equilibrium adsorption, Cd(II) uptake by algae biomass was determined according to the following:

$$q = \frac{v(c_0 - c_e)}{w}$$

where q is the metal uptake in mg (metal) g^{-1} biomass, v is the volume of metal-containing solution in contact with the biosorbent in L, C_0 is the initial concentration of metals in the solution in mg L⁻¹, C_e is the equilibrium concentration of metals in mg L⁻¹, and w is the weight of added algae dosage in gram.

Natural water adsorption experiments

Natural water was sampled at three different local water resource localities (34° 16′ 56.24″ N, 108° 4′ 27.95″ E), in Weihui canal, Hou river, and Gaogan Canaqu, and from discharged water from a factory (fruit plant). The first three localities represent the main water resources for irrigation in Yangling, Shaanxi, China. The Cd(II) concentrations of water from the localities were 0.097, 0.129, 0.260, and 0.497 mg L⁻¹ for the fruit plant, Gaogan Canaqu, Hou river, and Weihui canal, respectively. For each of the four localities, 200 mL natural water in 500 mL Erlenmeyer flasks was used to study the adsorption efficiency of 1.8 mg dead and a corresponding amount of live algal biomass; otherwise, the conditions were as in the previous experiments. The Cd(II) concentration was measured before the experiment and after 105 min (at equilibrium).

Kinetic model analyses

Kinetic models were used to provide information on the mechanism and process of the adsorption onto adsorbents. In batch adsorption systems, the kinetics is described by models based on adsorption equilibrium parameters such as the Lagergren pseudo-first-order, the Ho Y S pseudo-second-order, and the Elovich kinetic models. The kinetic models can be presented in the following forms:

$$\ln(q_e - q_t) = \ln q_e - k_1 t \tag{1}$$

$$\frac{t}{q_t} = \frac{1}{k_2 q_e^2} + \frac{1}{q_e} t$$
(2)

$$q_t = \frac{1}{\beta} \ln(\alpha\beta) + \frac{1}{\beta} \ln(t) \tag{3}$$

where $q_t (\text{mg g}^{-1})$ is the adsorption capacity at time t (min), q_e (mg g⁻¹) is the adsorption capacity at equilibrium, k_I (min⁻¹) and k_2 (g mg⁻¹ min⁻¹) are the pseudo-first-order and pseudo-second-order reaction rate constants for the kinetic model, and α and β are the initial adsorption rate and the desorption rate constant, respectively.

The data for the adsorption kinetic of cadmium adsorption by *C. vulgaris* were explored using an initial cadmium concentration of $C_0 = 100 \text{ mg L}^{-1}$ and the obtained data were analyzed by pseudo-first-order, pseudo-second-order, and Elovich kinetic models. The model parameters (q_e , k_1 , k_2 , α , and β) were determined, and statistical analyses were performed using the non-linear regression software Polymath (Zimmerman et al. 2004).

Biosorption isotherm models

For optimizing the model of the adsorption process for the removal of heavy metals, acquired experimental data were fitted to different sorption isotherm models in order to examine the relationship between sorption and metal concentration at equilibrium. The relationship between cadmium biosorption capacity and cadmium concentration at equilibrium was tested using two two-parameter isotherm models (Langmuir and Freundlich model) and two three-parameter isotherm models (Sips and Khan model). The isotherm model constants were calculated by nonlinear regression using the Polymath software.

Two-parameter models

The Langmuir isotherm may be presented as follows:

$$q_e = q_m \frac{k_1 c_e}{1 + k_1 c_e}$$

where q_m is the maximum metal uptake (mg g⁻¹) and k_I (L mg⁻¹) is the Langmuir constant. In Langmuir formulation, it is assumed that all sites possess equal affinity for the sorbate, and binding to the surface is primarily by physical forces and implicit in its derivation (Davis et al. 2003). This model estimates the maximum metal uptake values where they could not be reached in the experiments (Vijayaraghavan et al. 2006).

The Freundlich isotherm model has generally expressed as follows:

$$q_e = k_f c_e^{-1} / n$$

where k_f is the Freundlich constant (L g⁻¹), and *n* is the Freundlich exponent. The model assumes that the stronger binding sites are occupied first and that the binding strength decreases with the increasing degree of site occupation (Vijayaraghavan et al. 2006). Values of *n* greater than 1 show favorable nature of adsorption (Günay et al. 2007).

Three-parameter models

Sips isotherm is expressed in the following form:

$$q_e = q_{\max} \frac{\left(k_s c_e\right)^{\gamma}}{1 + \left(k_s c_e\right)^{\gamma}}.$$

Sips isotherm (Sips, 1948) is a combined form of Langmuir and Freundlich expressions deduced for predicting the heterogeneous adsorption systems and circumventing the limitation of the rising adsorbate concentration (Foo and Hameed 2010).

Khan isotherm has the following form:

$$q_e = \frac{q_m b_k c_e}{\left(1 + b_k c_e\right)^{a_k}}$$

where b_k is the Khan model constant, and a_k is the Khan model exponent.

Statistical analyses

All experiments were carried out in triplicate. One-factor analysis of variance (ANOVA) was used to determine significant differences between each variable using the software IBM SPSS Statistics 19 version. The Polymath software (Zimmerman et al. 2004) was used to analyze the kinetic and isotherm models by non-linear progression.

Results

Effects of algal material and contact time

The adsorption process of cadmium by live and dead *C. vulgaris* was studied using an initial concentration of 100 mg L⁻¹ cadmium. The adsorption rate was very rapid at first, with 96.8 and 95.2 % of the total amount of cadmium being removed by the dead algal and the live algal biomass, respectively, within the first 5 min (Fig. 1). Subsequently, the adsorption increased slowly for both live and dead algal material until it reached an equilibrium state after 105 min.

Effects of the initial concentration of cadmium

The effect of different initial concentrations of cadmium on adsorption on *C. vulgaris* was studied using five initial concentrations of cadmium (5, 10, 50, 100, and 150 mg L⁻¹). The adsorption efficiency of dead algae was found to be slightly higher (90 %) but not significantly different than that of live algae (86 %) using an initial cadmium concentrations, the efficiency was above 96 % for both types of algal material. No difference was found between dead algae and live algae using initial cadmium concentrations are using initial cadmium concentrations. The metal adsorption capacity by both live and dead algae increased significantly with an increasing initial metal concentration (Fig. 2b), with no difference between live and dead algae. The initial cadmium concentration thus had a significant effect on the adsorption.

Effects of the algal dosage

The adsorbent dosage played a significant role in the biosorption of metals (p < 0.05, Fig. 3). The influence of algal dose on adsorption efficiency of Cd(II) by *C. vulgaris* was investigated by adding dead algae and live algae (dry algae weights were 0.18, 1.8, and 18 mg of dead algae and a corresponding amount of live algae) to 200 mL of 100 mg L⁻¹ Cd(II) solution while keeping other parameters (contact time, agitation speed, temperature) constant. The results showed that the removal efficiency of *C. vulgaris* increased from 89 to 95 % as the adsorbent dosage was increased from 0.18 to 18 mg (Fig. 3a), and the adsorption capacity decreased with an increase in algal dose (Fig. 3b). No significant difference was found between dead or live algae.



Fig. 1 Effect of contact time on the adsorption of Cd(II) by *C. vulgaris*. **a** The adsorption efficiency by live and dead *C. vulgaris*. **b** The adsorption capacity by live algae and dead *C. vulgaris*

The adsorption efficiency of C. vulgaris in natural water

After biosorption, the adsorption efficiency of Cd(II) by *C. vulgaris* in natural water increased with the increasing Cd(II) concentration, similar to the results under batch adsorption experiment. Biosorption efficiency value was 85.1 and 61.6 % for dead and live algae, respectively, when Cd(II) concentration was 0.097 mg L⁻¹. But the efficiency increased to 96.4 and 90.8 % for dead and live algae with Cd(II) concentration increasing to 0.497 mg L⁻¹ (Fig. 4a). The adsorption quantity was low (Fig. 4b), probably because the cadmium concentration was low and because other metal ions were present in the natural water.

Kinetic model analyses

The data for the adsorption kinetic of cadmium adsorption by *C. vulgaris* were explored for an initial cadmium concentration of $C_0 = 100 \text{ mg L}^{-1}$ and the obtained data were analyzed by pseudo-first-order, pseudo-second-order, and Elovich kinetic models. According to the R^2 listed in Table 2, the pseudo-first-order kinetic equation ($R^2 = 0.996$) fits the adsorption of cadmium by live algae cells better than the Elovich model ($R^2 = 0.510$). The pseudo-second-order kinetic



Fig. 2 Effect of initial Cd(II) concentration on adsorption by *C. vulgaris.* **a** The adsorption efficiency by live and dead *C. vulgaris.* **b** The adsorption capacity by live and dead *C. vulgaris.* Data are means \pm SD (triplicates). *Different letters* indicate significant differences according to Tukey's test (p < 0.05)

equation ($R^2 = 0.999$) fits the adsorption of cadmium by dead algae cells better than the Elovich model ($R^2 = 0.512$). The predicted q_e (the adsorption amounts per unit biomass of algae) was 10.1 mg g⁻¹ from the pseudo-first-order kinetic model for live algal cells and was very close to the experimental q_e (10.60 mg g⁻¹). The calculated q_e of the pseudo-secondorder kinetic model for dead algal cells was10.85 mg g⁻¹ and was very close to the experimental q_e (10.89 mg g⁻¹). The results indicate that dead algal cells have higher adsorption abilities for cadmium than the live algal cells and suggest that the biosorption system of dead algae was a pseudo secondorder reaction, whereas live algae cells were best modeled by a pseudo first-order reaction.

Biosorption isotherm models

The relationship between cadmium biosorption capacity and cadmium concentration at equilibrium was tested using two two-parameter isotherm models and two three-parameter isotherm models listed in Tables 3 and 4.



Fig. 3 The influence of algae dosage on the adsorption efficiency and the adsorption quantity. a The adsorption efficiency by live and dead C. vulgaris. b The adsorption quantity by live and dead C. vulgaris. Data are means ± SD (triplicates). Different letters indicate significant differences according to Tukey's test (p < 0.05)

Two-parameter models The Langmuir isotherm model estimated the maximum adsorption capacity to be 35.31 mg g^{-1} for dead algae which was higher than the maximum adsorption amount of live algae (31.05 mg g⁻¹). The k_1 was calculated to be 0.19 and 0.16 L mg⁻¹ for live and dead algae, respectively. Higher values of k_1 indicate a steep initial slope of a sorption isotherm and high affinity. In general, high q_m and a steep initial isotherm slope are desirable (Davis et al. 2003). The correlation coefficients (0.902 and 0.938) were lower, which showed that the Langmuir models are not well



Biosorption efficiency (%)

Natural water collection site (Cd(II) concentration(mg L⁻¹))

Fig. 4 The adsorption of C. vulgaris in natural water. a The adsorption efficiency of C. vulgaris. b The adsorption capacity of C. vulgaris. Data are means ± SD (triplicates). Different letters indicate significant differences according to Tukey's test (p < 0.05)

suitable for describing the biosorption equilibrium of cadmium by the algal cells in the studied concentration range.

Similarly, the Freundlich isotherm model in general had low correlation coefficients (Table 3).

Three-parameter models The Sips isotherm model had high correlation coefficients (0.972 and 0.996 for live algae and dead algae, respectively, and low mean squared error (MSE) values (Table 4), indicating that the Sips isotherm model explains the expressed biosorption of cadmium onto C. vulgaris.

Table 2 Kinetic parameters of Cd(II) adsorption by C. vulgaris

Live algae	Pseudo-first-order kinetic model			Elovich					
U	$k_1 (\min^{-1})$	$q_{e(\text{cal.})} (\text{mg g}^{-1})$	$q_{e(\exp,1)} (\text{mg g}^{-1})$	R^2	MSE	α	β	R^2	MSE
	5.63 ± 0.40	10.41 ± 0.02	10.60 ± 0.03	0.996	0.05	45.06 ± 13.61	0.72 ± 0.01	0.510	0.55
Dead algae	Pseudo-second-order	kinetic model				Elovich			
	$K_2 (g mg^{-1} min^{-1})$	$q_{e(\text{cal.})} (\text{mg g}^{-1})$	$q_{e(\exp.)} (\mathrm{mg \ g}^{-1})$	R^2	MSE	α	β	R^2	MSE
	1.87 ± 0.69	10.85 ± 0.01	10.89 ± 0.02	0.999	0.01	45.39 ± 6.94	0.70 ± 0.01	0.512	0.60

Data are means ± standard error

Two-parameter models	Live algae	Dead algae	
Langmuir			
$q_m (\mathrm{mg \ g}^{-1})$	31.05 ± 15.04	35.31 ± 17.04	
$k (L mg^{-1})$	0.19 ± 0.16	0.16 ± 0.13	
R^2	0.902	0.938	
MSE	0.83	0.66	
Freundlich			
kf	4.76 ± 1.50	4.68 ± 1.26	
п	1.40 ± 1.4	1.31 ± 0.32	
R^2	0.870	0.912	
MSE	0.96	0.79	

 Table 3
 Isotherm constants of two-parameter models for Cd(II)

 biosorption on C. vulgaris

Data are means ± standard error

The maximum adsorption quantity values q_m were not well predicted by the Khan isotherm model, as illustrated by the relatively low correlation coefficients (Table 4).

Discussion

The adsorption efficiency of dead and live algae was found to be up to 96.83 and 95.17 %, respectively, after the first 5 min. The adsorption efficiency increased with time until it reached equilibrium at 105 min. The adsorption efficiency of dead algae was higher than that of live algae, but with no significant difference between the live and dead algae. For the live algae, the biosorption mechanism includes processes like active transport, intracellular and extracellular metal complexing, protein synthesis, metabolism, secretion caused by extracellular deposition, and biological adsorption. For the dead algae,

Table 4 Isotherm constants of three-parameter models for Cd(II)
 biosorption on *C. vulgaris* Cd(II)

Three-parameter models	Live algae	Dead algae
Sips isotherm		
q_m	16.34 ± 1.97	16.65 ± 0.85
ks	0.68 ± 0.12	0.62 ± 0.04
γ	2.63 ± 0.88	2.81 ± 0.49
R^2	0.972	0.996
MSE	0.45	0.18
Khan isotherm		
q_m	102.00 ± 12.05	101.99 ± 23.01
а	2.58 ± 0.10	2.40 ± 0.20
b	0.06 ± 0.01	0.06 ± 0.01
R^2	0.908	0.942
MSE	0.80	0.64

Data are means ± standard error

the biosorption is mainly an independent process which has nothing to do with metabolic processes (Liu et al. 2002). Compared to live algae, the dead algal material has internal functional groups exposed because the cell walls are damaged. In the adsorption process, parameters like the amount of biosorbent, initial concentration of metal ions, and existence of other ions (which may compete with the ions of interest for the active adsorption sites) in the adsorption medium may affect the adsorption rate (Bayramoglu and Arica 2008).

The adsorption capacities increased with increasing initial concentration of cadmium in the adsorption medium (Fig. 2b). The maximum adsorption efficiency can be as high as 98.02 % (Fig. 2a). To a large extent, the removal efficiency of algae is depending on the concentration of heavy metals. With biomass concentration being constant, the number of binding sites was the same, but the number of cadmium ions increases with the concomitant increase in cadmium concentration (Bhat et al. 2008). In the adsorption process, the adsorption efficiency can continue to increase with an increase in metal ion concentration, but only when the metal ion concentration is low. This adsorption phenomenon is caused by a larger driving force to overcome all mass transfer resistance of cadmium between the solid and the aqueous phase. Thus, a higher initial metal concentration results in higher metal ion adsorption (Edris et al. 2014). Ghimire et al. (2008) found that the adsorption increases with an increase in metal ion concentration until equilibrium is reached at a high metal concentration. The maximum metal ion adsorption of Scenedesmus quadricauda and the adsorption of metal ions increased as the initial concentration of the metal ions increased in the medium (Bayramoglu and YArica 2011). The initial cadmium concentration thus had a significant effect on the adsorption capacity. Similar results have been also found by Chen et al. (2008), Riaz et al. (2009), Aksu (2001), and Edris et al. (2014).

The adsorption efficiency also was affected by algal dosage (Fig. 3a). In the present study, the adsorption efficiency increased, from 89 to 95 % as the algal dosage increased. This can be due to an increase in the number of adsorption sites, and the decrease in unit adsorption is basically due to the biosorption sites that remain unsaturated during adsorption reaction (Vinod et al. 2009). Similar trends were shown by Ozsoy et al. (2008) and Edris et al. (2014). The adsorption capacity per unit algal material decreased when algal dose increased from 0.18 to 1.8 mg (Fig. 3b). This can simply be attributed to a lack of cadmium left for the sorption process. Moreover, the increase in adsorption efficiency of metals by increasing the biomass dosage is due to an increase in the number of active sites and available surface area (Edris et al. 2014).

The Cd(II) concentrations in the natural water localities were so high that the use of the water resources for irrigation without removal of cadmium could cause ecological problems and affect humans. Ouality standard value for Cd(II) in irrigation water of the basic control project is 0.01 mg L^{-1} . For the adsorption of cadmium in the natural water, we found that the adsorption efficiency of both live algae and dead algae increased with increasing cadmium ion concentration (Fig. 3a). Under laboratory conditions, live and dead algal adsorption efficiency reached 95.97 and 98.12 %, respectively, at equilibrium (Figs. 1a and 2a). Whereas in natural water, the adsorption efficiency of live and dead algae at adsorption equilibrium was lower (Fig. 3a). The smallest live and dead algae adsorption efficiency can reach 61.57 and 85.13 %. The reason for this reduction adsorption efficiency might be the presence of other meal ions besides cadmium in the natural water. These metal ions may also be absorbed by C. vulgaris and thereby creating a competitive adsorption. If the number of binding sites of the biomass was constant, this would result in a reduction in the adsorption efficiency of cadmium.

The experimental data for adsorption kinetics of cadmium by live algal cells obeyed a pseudo-first-order model (Table 2). In recent years, the pseudo-first-order model has been widely used for describing adsorption of pollutants from natural water in different fields (Hameed and EI-Khaiary MI, 2008a; 2008b; Tan et al. 2008). The sorption system of dead algal cells is best described by a second-order model, based on the assumption that it predicts the behavior over the whole range of adsorption and the rate-limiting step may be the biosorption.

At low adsorbate concentration, the slope of the initial portion of the isotherm was small, and with increasing adsorbate concentration, the adsorbing capacity raised linearly (Fig. 2b). This indicates that the distribution ratio of the adsorbate in the liquid phase and the surface of the adsorbent is constant. The adsorption isotherm is in accordance with the constant partition type of adsorption isotherms concerning the adsorption on solid surfaces from solution (Zhu and Zhao 1996). The Sips constants showed easy uptake of Cd(II) with high adsorptive capacity of the fresh and dried *C. vulgaris* cells (Tables 3 and 4).

In conclusion, the adsorption of cadmium by live and dead *C. vulgaris* was highly influenced by parameters such as contact time, adsorbent dosage, and initial concentration of cadmium. The adsorption capacity was not significantly different between dead algae and live algae, so by using live algae to remove heavy metal contamination instead of dry algal material, one may save time as well as costs involved in the drying procedure. Using both live and dead algal materials, the adsorption capacity increased with an increase in the initial cadmium concentration and decreased with an increase in adsorbent dosage.

The study showed that *C. vulgaris* makes up a promising candidate algal species for the removal of Cd(II) from natural water. The high cadmium concentrations measured in natural waters in combination with the identification of a suitable

candidate for removal of cadmium suggest that regulatory measures should be taken by the authorities to limit or remove the metal pollutants in the water environment. The obtained kinetic parameters, equilibrium time, rate constants, and initial biomass dosage can be used for optimizing the adsorption design.

Acknowledgments This study was supported by the National Natural Science Foundation of China (31000099) and the Fundamental Research Funds of the Northwest A&F University (2014YB038).

References

- Ahmet C, Tamer A, Sibel T, Ozge T (2006) Biosorption characteristics of Bacillus sp. ATS-2 immobilized in silica gel for removal of Pb(II). J Hazard Mater 136:317–323
- Ajmal M, Rao RAK, Ahmad R, Khan MA (2006) Adsorption studies on Parthenium hysterophorous weed: removal and recovery of Cd(II) from wastewater. J Hazard Mater 135:242–248
- Akhtar M, Iqbal S, Kausar A, Bhanger MI, Shaheen MA (2010) An economically viable method for the removal of selected divalent metal ions from aqueous solutions using activated rice husk. Colloids Surf B 75:149–155
- Aksu Z (2001) Equilibrium and kinetic modeling of cadmium (II) biosorption by *C. vulgaris* in a batch system: effect of temperature. Sep Purif Technol 21:285–294
- Azouaou N, Sadaoui Z, Djaafri A, Mokaddem H (2010) Adsorption of cadmium from aqueous solution onto untreated coffee grounds: equilibrium, kinetics and thermodynamics. J Hazard Mater 184:126–134
- Baby J, Raj JS, Biby ET, Sankarganesh P, Jeevitha MV, Ajisha SU, Rajan SS (2010) Toxic effect of heavy metals on aquatic environment. Int J Biol Chen Sci 4:939–952
- Bačkor M, Fahselt D (2008) Lichen photobionts and metal toxicity. Symbiosis 46:1–10
- Bayramoglu G, Arica MY (2008) Enzymatic removal of phenol and p-chlorophenol in enzyme reactor: horseradish peroxidase immobilized on magnetic beads. J Hazard Mater 156:148
- Bayramoğlu G, Arica MY (2008) Removal of heavy mercury (II), cadmium (II) and zinc (II) metal ions by live and heat inactivated *Lentinus edodes* pellets. Chem Eng J 143:133–140
- Bayramoğlu G, Arica MY (2009) Construction a hybrid biosorbent using Scenedesmus quadricauda and Ca-alginate for biosorption of Cu(II), Zn(II) and Ni(II): kinetics and equilibrium studies. Bioresour Technol 100:186–193
- Bayramoglu G, YArica MY (2011) Preparation of a composite biosorbent using *Scenedesmus quadricauda* biomass and alginate/polyvinyl alcohol for removal of Cu(II) and Cd(II) ions: isotherms, kinetics, and thermodynamic studies. Water Air Soil Pollut 221:391–403
- Bhat SV, Melo JS, Chaugule BB, D'Souza SF (2008) Adsorption characteristics of uranium (VI) from aqueous medium onto *Catenella repens*, a red alga. J Hazard Mater 158:628–635
- Cay S, Uyanık A, Özas A (2004) Single and binary component adsorption of copper (II) and cadmium(II) from aqueous solutions using tea-industry waste. Sep Purif Technol 38:273–280
- Chen C, Wang J (2007) Influence of metal ionic characteristics on their biosorption capacity by Saccharomyces cerevisiae. Appl Microbiol Biotechnol 74:911–917
- Chen Z, Ma W, Han M (2008) Biosorption of nickel and copper onto treated alga (*Undaria pinnatifida*): application of isotherm and kinetic models. J Hazard Mater 155:327–222

- Cheung CW, Porter JF, McKay G (2001) Sorption kinetic analysis for the removal of cadmium ions from effluents using bone char. Water Res 35:605–621
- Davis TA, Volesky B, Mucci A (2003) A review of the biochemistry of heavy metal biosorption by brown algae. Water Res 37:4311–4330
- Dixit S, Singh DP (2014) An evaluation of phycoremediation potential of cyanobacterium *Nostoc muscorum*: characterization of heavy metal removal efficiency. J Appl Phycol 26:1331–1342
- Edris G, Alhamed Y, Alzahrani A (2014) Biosorption of cadmium and lead from aqueous solutions by *Chloralla vulgaris* biomass: equilibrium and kinetic study. Arab J Sci Eng 39:87–93
- Farombi EO, Adelowo OA, Ajimoko YR (2007) Biomarkers of oxidative stress and heavy metal levels as indicators of environmental pollution in African cat fish (*Clarias gariepinius*) from Nigeria Ogun River. Int J Environ Res Public Health 4:158–165
- Farooq U, Khan MA, Athar M, Kozinski JA (2011) Effect of modification of environmentally friendly biosorbent wheat (*Triticum aestivum*) on the biosorptive removal of cadmium (II) ions from aqueous solution. Chem Eng J 171:400–410
- Foo KY, Hameed BH (2010) Insights into the modeling of adsorption isotherm systems. Chem Eng J 156:2–10
- Ghimire KN, Inoue K, Ohto K, Hayashida T (2008) Adsorption study of metal ions onto crosslinked seaweed *Laminaria japonica*. Bioresour Technol 99:32–37
- Günay A, Arslankaya E, Tosun I (2007) Lead removal from aqueous solution by natural and pretreated clinoptilolite: adsorption equilibrium and kinetics. J Hazard Mater 146:362–371
- Gupta VK, Rastogi A (2008) Equilibrium and kinetic modeling of cadmium (II) biosorption by nonliving algal biomass *Oedogonium* sp. from aqueous phase. J Hazard Mater 153:759–766
- Gupta VK, Rastogi A (2009) Biosorption of hexavalent chromium by raw and acid-treated green alga *Oedogonium hatei* from aqueous solutions. J Hazard Mater 163:396–402
- Gupta VK, Sharma S (2002) Removal of cadmium and zinc from aqueous solutions using red mud. Environ Sci Technol 36:3612–3617
- Gupta VK, Rastogi A, Dwivedi MK, Mohan D (1997) Process development for the removal of zinc and cadmium from wastewater using slag—a blast furnace waste material. Sep Sci Technol 32:2883–2912
- Gupta VK, Jain CK, Ali I, Sharma M, Saini VK (2003) Removal of cadmium and nickel from wastewater using baggasse fly ash—a sugar industry waste. Water Res 37:4038–4044
- Gupta VK, Nayak A, Agarwal S (2015a) Bioadsorbents for remediation of heavy metals: current status and their future prospects. Environ Eng Res 20:1–18
- Gupta VK, Nayak A, Bhushan B, Agarwal S (2015b) A critical analysis on the efficiency of activated carbons from low-cost precursors for heavy metals remediation. Environ Sci Technol 45:613–668
- Hameed BH, EI-Khaiary M (2008b) Sorption kinetics and isotherm studies of a cationic dye using agriculture waste: broad bean peels. J Hazard Mater 154:639–648
- Hameed BH, EI-Khaiary MI (2008a) Batch removal of malachite green from aqueous solutions by adsorption on oil palm trunk fibre: equilibrium isotherms and kinetic studies. J Hazard Mater 154:237–244
- Herrero R, Cordero B, Lodeiro P, Rey-Castro C, Sastre de Vicente M (2006) Interaction of cadmium (II) and protons with dead biomass of marine algae *Fucus* sp. Mar Chem 99:106–116
- Iqbal M, Saeed A, Zafar SI (2009) FTIR spectrophotometry, kinetics and adsorption isotherms modeling, ion exchange, and EDX analysis for understanding the mechanism of Cd²⁺ and Pb²⁺ removal by mango peel waste. J Hazard Mater 164:161–171
- Jena J, Pradhan N, Aishvarya V, Nayak RR, Dash BP, Sukla LB, Panda PK, Mishra BK (2015) Biological sequestration and retention of cadmium as CdS nanoparticles by the microalga *Scenedesmus*-24. J Appl Phycol 27:2251–2260

- Ji L, Xie SL, Feng J, Li YH, Chen L (2012) Heavy metal uptake capacities by the common freshwater green alga *Cladophora fracta*. J Appl Phycol 24:979–983
- Kadirvelu K, Namasivayam C (2003) Activated carbon from coconut coirpith as metal adsorbent: adsorption of Cd(II) from aqueous solution. Adv Envrion Res 7:471–478
- Khattar JIS, Parveen S, Singh Y, Singh DP, Gulati A (2015) Intracellular uptake and reduction of hexavalent chromium by the cyanobacterium *Synechocystis* sp. PUPCCC 62. J Appl Phycol 27:827–837
- Lagoa R, Rodrigues JR (2007) Evaluation of dry protonated calcium alginate beads for biosorption applications and studies of lead uptake. Appl Biochem Biotechnol 143:115–128
- Liu RX, Tang HX, Lao WX (2002) Advances in biosorption mechanism and equilibrium modeling for heavy metals on biomaterials. Prog Chem 14:87–92
- Loukidou MX, Karapantsios TD, Zouboulis AI, Matis KA (2004) Diffusion kinetic study of cadmium(II) biosorption by *Aeromonas caviae*. J Chem Technol Biotechnol 79:711–719
- Lu WB, Shi JJ, Wang CH, Chang JS (2006) Biosorption of lead, copper and cadmium by an indigenous isolate *Enterobacter* sp. J1 possessing high heavy-metal resistance. J Hazard Mater 134:80–86
- Matlock MM, Henke KR, Atwood DA (2002) Effectiveness of commercial reagents for heavy metal removal from water with new insights for future chelate designs. J Hazard Mater 92:129–42
- Meitei MD, Prasad MNV (2013) Lead (II) and cadmium (II) biosorption on *Spirodela polyrhiza* (L.) Schleiden biomass. J Environ Chem Eng 1:200–207
- Memon JR, Memon SQ, Bhanger MI, Zuhra Memon G, El-Turki A, Allen GC (2008) Characterization of banana peel by scanning electron microscopy and FT-IR spectroscopy and its use for cadmium removal. Colloids Surf B 66:260–265
- Mirghaffari N, Moeini E, Farhadian O (2015) Biosorption of Cd and Pb ions from aqueous solutions by biomass of the green microalga, *Scenedesmus quadricauda*. J Appl Phycol 27:311–320
- Naser HA (2013) Assessment and management of heavy metal pollution in the marine environment of the Arabian Gulf: a review. Mar Poll Bull 72:6–13
- Ozsoy HD, Kumbur H, Saha B, Leeuwen JHV (2008) Use of *Rhizopus* oligosporus produced from food processing natural water as a biosorbent for Cu (II) ions removal from the aqueous solutions. Bioresour Technol 99:4943–4948
- Padmini E, Sridhar S (2007) Effect of pH and contact time on the uptake of heavy metals from industrial effluents by *Pongamia pinnata* Bark. Asian J Microbiol Biotechnol Environ Sci 9:187–190
- Perales-Vela HV, Gonzáles-Moreno S, Montes-Horcasitas C, Canizares-Villanueva RO (2007) Growth, photosynthetic and respiratory responses to sub-lethal copper concentrations in *Scenedesmus incrassatulus* (Chlorophyceae). Chemosphere 67:2274–2281
- Pino GH, Mesquita LMS, Torem ML, Pinto GASP (2006) Biosorption of cadmium by green coconut shell powder. Miner Eng 19:380–387
- Qiu H, Lu LV, Pan BC, Zhang QJ, Zhang WM, Zhang QX (2009) Critical review in adsorption kinetic models. J Zhejiang Univ Science A 10(5):716–724
- Rai PK (2012) An eco-sustainable green approach for heavy metals management: two case studies of developing industrial region. Environ Monit Assess 184:421–48
- Riaz M, Nadeem R, Hanif MA, Ansari TM, Rehman KU (2009) Pb(II) biosorption from hazardous aqueous streams using *Gossypium hirsutum* (cotton) waste biomass. J Hazard Mater 161:88–94
- Saiano F, Ciofalo M, Cacciola SO, Ramirez S (2005) Metal ion adsorption by *Phomopsis* sp. biomaterial in laboratory experiments and real wastewater treatment. Water Res 39:2273–2280
- Saikaew W, Kaewsam P, Saikaew W (2009) Pomelo peel: agricultural waste for biosorption of cadmium ions from aqueous solutions. World Acad Sci Eng Technol 56:287–291

- Selatnia A, Bakhti MZ, Madani A, Kertous L, Mansouri Y (2004) Biosorption of Cd²⁺ from aqueous solution by a NaOH-treated bacterial dead *Streptomyces rimosus* biomass. Hydrometallurgy 75:11–24
- Sha L, Xueyi G, Ningchuan F, Qinghua T (2009) Adsorption of Cu²⁺ and Cd²⁺ from aqueous solution by mercapto-acetic acid modified orange peel. Colloids Surf B 73:10–14
- Sips R (1948) Combined form of Langmuir and Freundlich equations. J Chem Phys 16:490–495
- Solisio C, Lodi A, Soletto D, Converti A (2008) Cadmium biosorption on Spirulina platensis biomass. Bioresour Technol 99:5933–5937
- Stanier RY, Kunisawa R, Mandel M, Cohen-Bazire G (1971) Purification and properties of unicellular blue-green algae (Order Chroococcales). Bact Rev 35:171–205
- Suzuki Y, Kametani T, Maruyama T (2005) Removal of heavy metals from aqueous solution by nonliving *Ulva* seaweed as biosorbent. Water Res 39:1803–1808
- Tan IAW, Ahmad AL, Hameed BH (2008) Adsorption of basic dye on high-surface-area activated carbon prepared from coconut husk: equilibrium, kinetic and thermodynamic studies. J Hazard Mater 154:337–346
- Vaclavikova M, Misaelides P, Gallios G, Jakabsky S, Hredzak S (2005) Removal of cadmium, zinc, copper and lead by red mud, an iron oxides containing hydrometallurgical waste. Stud Surf Sci Catal 155:517–525
- Vijayaraghavan K, Padmesh TVN, Palanivelu K, Velan M (2006) Biosorption of nickel (II) ions onto *Sargassum wightii*: application of two-parameter and three-parameter isotherm models. J Hazard Mater B 133:304–308
- Vilar VJP, Botelho CMS, Boaventura RAR (2006) Equilibrium and kinetic modeling of Cd(II) biosorption by algae *Gelidium* and agar extraction algal waste. Water Res 40:291–302
- Vinod VTP, Sashidhar RB, Sreedharc B (2009) Interaction of Pb²⁺and Cd²⁺ with gum kondagogu (*Cochlospermum gossypium*): a natural

carbohydrate polymer with biosorbent properties. Carbohydr Polym 78:894-901

- WHO (2010) Exposure to cadmium: a major public health concern http://www.who.int/ipcs/features/cadmium.pdf?ua=1. Accessed 20 June 2015
- WHO (2011) Cadmium in drinking-water background document for development of WHO guidelines for drinking-water quality, World Health Organization (WHO/SDE/WSH/03.04/ 11/Rev/1). http://www.who.int/water_sanitation_ health/dwq/chemicals/cadmium.pdf. Accessed 20 June 2015
- Winter C, Winter M, Pohl P (1994) Cadmium adsorption by non-living biomass of the semi-macroscopic brown alga, *Ectocarpus* siliculosus, grown in axenic mass culture and localisation of the adsorbed Cd by transmission electron microscopy. J Appl Phycol 6:479–487
- Xia J, Li Y, Lu J (2004) Effects of copper and cadmium on growth, photosynthesis, and pigment content in *Gracilaria lemaneiformis*. Bull Environ Contam Toxicol 73:979–986
- Yu J, Tong M, Sun X, Li B (2007) Cystine-modified biomass for Cd(II) and Pb(II) biosorption. J Hazard Mater 143:277–284
- Yu CL, Lu ZP, Ge FZ, Zhao EL (2011) Biosorption of cadmium onto *Pseudomonas fluorescens*: application of isotherm and kinetic models. Adv Mat Res 171–172:49–52
- Zhu BY, Zhao ZG (1996) The foundation of interface chemistry. Chemical Industry Press, Beijing
- Ziagova M, Dimitriadis G, Aslanidou D, Papaioannou X, Tzannetaki EL, Liakopoulou-Kyriakides M (2007) Comparative study of Cd(II) and Cr(VI) biosorption on *Staphylococcus xylosus* and *Pseudomonas* sp. in single and binary mixtures. Bioresour Technol 98:2859–2965
- Zimmerman AR, Coyne KW, Chorover J (2004) Problem solving in chemical and biochemical engineering with POLYMATH, Excel, and MATLAB. Org Geochem 35:355–375