

Evidence of reproductive cost in the triphasic life history of the red alga *Gracilaria chilensis* (Gracilariales, Rhodophyta)

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Abstract The extent of changes in basic physiological and demographic traits associated with reproduction was investigated in the highly cultivated haploid–diploid red alga, *Gracilaria chilensis*. Sixty individuals bearing vegetative and reproductive fronds collected in the natural population of Niebla (39°52'S, 73°23'W), in Chile, were cultivated under controlled culture conditions. Our results demonstrated that vegetative fronds have a higher survival rate and a better growth rate than reproductive ones irrespective of the type of individual analyzed (male gametophyte, female gametophyte, and tetrasporophyte). Moreover, the reproductive fronds clearly showed a decrease in photosynthetic activity compared to non-reproductive ones. In males and tetrasporophytes, the photosynthetic reduction in reproductive individuals could be explained by a physical effect of reproductive structure development as well as spores release, disrupting the continuity of the photosynthetic cortical tissues. Translocation of photo-assimilates from nearby vegetative tissue or the previous accumulation of photosynthetic products seems to be a prerequisite

for reproductive structure development in this species. Altogether, these results document for the first time in *G. chilensis* that reproduction has a strong physiological effect on male, female, and tetrasporophyte fronds. This trade-off between reproduction, growth, and survival suggest the existence of reproductive costs in the life history of *G. chilensis*.

Keywords Short-term reproductive cost · Dioecy · Gametophyte · Sporophyte · Carposporophyte · Haplo-diploidy · Photosynthesis

Introduction

The existence of reproductive cost has been demonstrated in various animals (Reznick et al. 2000) and plants (Obeso 2002). The concept of reproductive cost, developed by Fisher (1930) and Harper (1967), provides the conceptual framework of the theory of resource allocation (for review, see Bazzaz et al. 2000). This theory predicts the existence of a resource trade-off between reproduction, growth, and survival, assuming that reproductive effort imposes a cost on an organism in terms of reduced growth or lower survival (Fisher 1930; Bell 1984; Stearns 1992). Consequently, the study of resource investment in reproduction constitutes a crucial parameter in the understanding of life history strategies of any given species (Harper 1967). According to Bell (1984), reproductive cost is a necessary condition for the optimization of life histories. Indeed, the evolution of life history is constrained by the presence of trade-offs among some of the traits that contribute to fitness (Stearns 1992).

In terrestrial plants, such trade-offs have been demonstrated experimentally in limited resources conditions either between sexual reproduction and vegetative growth (Obeso 2002) or between reproduction and survival rate (Rameau and Gouyon 1991). In dioecious plants, studies have shown

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clear differences in responses among sexes, with females investing more carbon, nitrogen, and other resources in reproduction than males (Obeso 2002). This difference in reproductive effort has been mainly related to the production of seeds, fruits, and associated structures (Ashman 1994) and has often been linked with differences in photosynthetic performance (Wheelwright and Logan 2004).

In seaweed, cost of reproduction remains poorly documented (for review, see De Wree and Klinger 1990) and has been mainly addressed in Phaeophyceae species in which reproductive organs are clearly differentiated (e.g., receptacles and sporophylls) (Vernet and Harper 1980; McCourt 1985; Ang 1992; Mathieson and Guo 1992; Åberg 1996; Zou et al. 2011). While a reproductive cost, like growth reduction or decrease of survival rate, has been observed in various fucoids (McCourt 1985; Ang 1992; Mathieson and Guo 1992; Åberg 1996; Chu et al. 2011), the existence of resource trade-off in algae has been discussed because reproductive organs are normally pigmented and potentially able to photosynthesis before the release of spores or gametes (De Wree and Klinger 1990). However, the reproductive period coincides generally with the end of growth and can be followed by the senescence and the death of the parental thallus (Zou et al. 2006).

In Rhodophyta species, resource trade-off between reproduction and vegetative growth is more difficult to assess because reproductive organs are generally embedded in the cortical tissues of the photosynthetic thallus except for female individuals showing a parasitic-like development of the cystocarp on the female thallus after fertilization (Hommersand and Fredericq 1995). In *Gracilaria chilensis* and *Gracilaria domingensis*, it has been shown that vegetative growth of female gametophytes decreases after fertilization during the development of cystocarps (Santelices and Varela 1995; Guimaraes et al. 1999), suggesting that resources were limited. Similarly, the results obtained in the three Ceramiacean species, *Antithamnion nipponicum*, *Ceramium boydenii*, and *Ceramium japonicum*, demonstrated that the number and size of spores released depend on the length of the vegetative branch on which cystocarps were produced (Kamiya and Kawai 2002). All these results support the existence of a high photosynthetic investment in reproductive structures of female gametophyte in Rhodophyta. In male and tetrasporophyte individuals, the influence of reproductive structure on survival and growth has been rarely assessed (but see Santelices and Varela 1995). In these two types of individual, the reproductive structure derives from cortical tissues, and the dual role of the thallus (i.e., photosynthesis and reproduction) raises questions regarding the potential trade-offs involved in the maintenance of these two functions. Thornber (2006) hypothesized that haploid (male and female gametophytes) and diploid (tetrasporophyte) individuals could differ in vegetative growth, survival, and respective investment in reproductive structures.

The aim of this study was to analyze the ecophysiological outcomes of the development of reproductive structures in the red seaweed *G. chilensis* for the three types of individuals (males, females, and tetrasporophytes). This species is the most commercially important agarophyte in Chile (Buschmann et al. 2001). We studied the influence of the reproductive status on survival and growth rates as well as basic physiological parameters of interest (i.e., respiration rate, net primary production, and pigment content) by comparing reproductively and vegetative fronds of male gametophyte, female gametophyte, and tetrasporophyte individuals.

Materials and methods

Gracilaria chilensis exhibits a typical Polysiphonia-type life cycle with two free-living isomorphic generations. Meiosis occurs in the reproductive diploid individuals (tetrasporophytes) to produce haploid spores (tetraspores). When liberated, tetraspores attach to the substratum, develop perennial holdfasts, and grow into haploid dioecious gametophytic males and females. Male gametes are liberated in the water column, whereas female gametes are retained on the female thallus. After fertilization, the carposporophyte (cystocarp) develops on the female haploid plant and the zygote undergoes successive mitoses to produce a gonimoblast from which many thousands of identical diploid spores are formed. When liberated, carpospores attached to the substratum develop into perennial holdfasts, subsequently growing into a tetrasporophyte, completing the complex life cycle. In Chile, natural populations formed by individuals growing from a perennial holdfast and attached to the rocky substratum are encountered between Coquimbo and Raul Marin Balmaceda (30° S and 45° S; Bird et al. 1986). As vegetative fronds share the same morphology, phase and sex of individuals can be recognized only during the reproductive period.

Laboratory experimental design

Individuals of *G. chilensis* were collected at low tide in the natural population of Niebla (39°52' S, 73°23' W, Region XIV, Chile) in March 2010. The type of individual was identified according to the reproductive structures (tetrasporangia for tetrasporophytes, spermatangia for males, and cystocarps for fertilized females), using a binocular microscope (Guillemin et al. 2008). Even if reproductive individuals of *G. chilensis* could be encountered all year round (Meneses 1996), a 3-year survey of the Niebla population has shown that the highest percentage of reproductive individuals was encountered during summer (December to April) with at least 85 % of reproductive individuals (ML Guillemin, unpublished data). Twenty tetrasporophytes, 20 females and 20 males, were sampled and

cleaned of epiphytes. For each individual, 2 g of fresh weight of vegetative fronds and 2 g of fresh weight of reproductive fronds were selected to obtain six groups of fronds (vegetative females, males, and tetrasporophytes, and reproductive females, males, and tetrasporophytes). Following the criteria suggested by Guzmán-Urióstegui et al. (2002), we selected for experimental analyses only the healthy fronds with no damage (i.e., alteration of the cortical tissue) or discoloration. Only fronds with early stages of reproductive structures were selected to form the three groups of reproductive fronds. The fronds were incubated for 2 weeks in 1-L Erlenmeyer flask of culture medium (modified SFC culture medium; Correa and McLachlan 1991) aerated with ambient air under controlled photoperiod (12:12 h L/D), photon flux density ($60 \mu\text{mol photons m}^{-2} \text{s}^{-1}$), temperature ($15 \pm 1 \text{ }^\circ\text{C}$), and salinity (35 PSU). An important spore release was observed during the first 48 h for both tetrasporophytes and females. However, a continuous and low spore shedding was observed until the end of the experiment for these two types of individuals (spores not counted during our study).

Basic physiological parameters and survival rate and growth rates of vegetative fronds were measured on fronds without visible reproductive structures under binocular microscope observation. For reproductive fronds, measures were performed on male and tetrasporophyte fronds showing reproductive structures and, on female fronds, with at least two cystocarps per centimeter. Male and tetrasporophyte reproductive thalli selected presented a continuous and homogenous cover of reproductive structures (100 % covering).

Pigments content

Chlorophyll *a* (Chl *a*) was extracted from 10–15 mg fresh weight samples in *N,N*-dimethylformamide following the protocols of Gómez et al. (2005). Absorbance values of the supernatant were measured using a spectrophotometer. Chl *a* concentration were estimated by the equation of Inskeep and Bloom (1985). The phycobilins were extracted from ~50 mg fresh weight samples in 0.1 M phosphate buffer (pH=6.5). After centrifugation (30 min at ca. $4,500 \times g$), the concentrations of phycoerythrin (PE) and phycocyanin (PC) were determined spectrophotometrically using the equations of Beer and Eshel (1985). For each pigment concentration measurement performed, eight samples of the six frond groups were selected randomly.

Metabolic rates and primary productivity

Net primary productivity (NPP) and respiration (*R*) were measured using the light and dark bottle methodology described in Howarth and Michaels (2000). Fresh fronds of

$0.1 \pm 0.02 \text{ g}$ were placed in 50-mL Falcon tubes filled with filtered seawater sterilized with UV light (QL-40; Rainbow Lifegard, USA) and kept in a temperature- and light-controlled incubation chamber ($60 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ and $15 \pm 1 \text{ }^\circ\text{C}$) for 12 h. Initial and final dissolved O_2 concentrations were estimated using an YSI 550A oxymeter (YSI Inc., USA). No blank correction was applied since oxygen values in light and dark control bottles remained constant. All oxygen values ($\text{mg O}_2 \text{L}^{-1} \text{h}^{-1}$) were converted into carbon values ($\text{mg C L}^{-1} \text{h}^{-1}$) by using a factor of 0.375/PQ (i.e., photosynthetic quotient). The PQ value was taken as 1 and expressed as a fresh weight basis. Eight replicate measurements were taken for each of the six groups of fronds. After incubation, the algae from the respective bottles were dried at $60 \text{ }^\circ\text{C}$ until a constant weight was reached.

Survival and growth

Fifty frond segments (without apex), 1 cm in length, were excised from each group of fronds (vegetative and reproductive male, female, and tetrasporophytes). Separately for each group of fronds, ten segments were randomly selected and placed into a 50-mL culture flask with filtered seawater without aeration and under low light intensity of $20 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ (five replicates). Filtered seawater was changed weekly and segments were cleaned using a soft brush. After 50 days, the survival rate was estimated by counting the number of healthy segments (i.e., highly pigmented) and was expressed as a percentage of the original number of segments at the beginning of the experiment. After 60 days, all the bleached fronds fragments had died. The growth rate was estimated after 50 days by measuring the number of new tips formed along the 1 cm length frond segment.

Data analysis

Physiological parameters of interest, survival, and growth estimated under laboratory conditions were compared using a two-way analysis of variance (ANOVA) and Tukey's Honestly Significant Difference (HSD) tests (STATISTICA 7.0, StatSoft, USA). The type of individual (female, male, and tetrasporophyte) and the reproductive status of the fronds (vegetative or reproductive) were used as fixed factors. ANOVA assumptions (homogeneity of variance and normal distribution) were examined using Cochran and Kolmogorov–Smirnov tests, respectively. Transformations preceded the analyses when needed ($\log_{10}(x+1)$ for all three pigment concentrations and $\arcsin \sqrt{(x/100)}$ for the survival rate).

Results

There was a significant interaction between the type of individual (female gametophyte, male gametophyte, and tetrasporophyte) and the reproductive status of the frond (vegetative or reproductive) with respect to the concentration of PE, net productivity, respiration, and growth rates (Table 1). Reproductive females (showing cystocarps) presented the lowest values of pigment content, net productivity, and respiration rate of all the fronds measured (Fig. 1).

The reproductive status of the frond in *G. chilensis* strongly affects all the seven ecophysiological measurements performed during our study ($p < 0.001$, ANOVA; Table 1, Fig. 1). Regardless of sex or phase, vegetative fronds were characterized by a higher pigment content, net productivity, survival, and growth rates than the reproductive fronds (Fig. 1). Concentration of Chl *a* and PE were 1.2 to 4.5 times greater in vegetative fronds than in reproductive ones (Fig. 1a, b). Net productivity for vegetative fronds was almost twice that of reproductive fronds for all three individual types (Tukey's HSD $p < 0.05$, Fig. 1d). On the other hand, the respiration rates were roughly similar for all groups of fronds, with the exception of reproductive females, which presented a reduction in the respiration rate (Fig. 1e). Striking differences between vegetative and reproductive fronds were observed for survival and growth rates. Indeed, after 50 days, whereas male and female reproductive gametophytes segments had not generated new apex, vegetative segments of 1 cm length (whatever the sex or phase) generated approximately two new apices (Fig. 1f, g). The most conspicuous differences between reproductive and vegetative fronds were observed in female gametophytes, and the differences were significant for all the seven measures (Tukey's HSD $p < 0.05$, Fig. 1).

There were no consistent differences between the three types of individual except for the phycobilin concentrations (PC), the net productivity, and respiration rate (ANOVA;

Table 1). PE and PC were slightly higher in tetrasporophytes than in gametophytes, and the lowest net productivity and respiration rate were seen in female gametophytes (Fig. 1).

Discussion

The results of this study indicate that (1) vegetative fronds of *G. chilensis* from the natural population of Niebla (Chile) have a higher survival rate and a better growth rate than reproductive ones and (2) the reproductive fronds clearly show a clear decrease in photosynthetic activity compared to non-reproductive fronds.

Previous studies in Rhodophyta species demonstrated the effect of reproduction on the photosynthetic activity, such as *Gelidiella acerosa* (Ganzon-Fortes 1999), *Schottera nicaensis* (Perrone and Felicini 1988), and *Porphyra yezoensis* (Yang et al. 2012). These authors suggested that the development of new reproductive structures depends on translocation of photoassimilates from nearby vegetative tissue or on the previous accumulation of photosynthetic products. Yang et al. (2012) demonstrated that complete maturation of the spermatia in *P. yezoensis* was determined by photosynthetic activity at the beginning of the spermatangial development (i.e., the four-cell stage), while in the Chlorophyta *Ulva prolifera*, the formation of the mature sporangia depended directly on the thallus accumulation of photosynthetic products (Gao et al. 2010). In the red alga *Gracilaria cornea*, Guzmán-Urióstegui et al. (2002) pointed out the importance of nutrient supply in the development of cystocarps, and Kamiya and Kawai (2002) demonstrated in Ceramiales species that female reproductive effort depends on the length of the vegetative branch where cystocarps are produced. The dependence of reproductive structure upon nearby vegetative tissues in algae are generally associated with (1) a positive correlation between plant size and fecundity (De Wreede and Klinger 1990; Åberg 1992, 1996;

Table 1 Results of ANOVA analyses on pigment contents, net productivity, respiration, survival, and growth of reproductive and vegetative fronds for female gametophytes, male gametophytes, and tetrasporophytes of *G. chilensis*

| Response variable | <i>n</i> | Type of individual <i>df</i> (2) | | | Reproductive status <i>df</i> (1) | | | Interaction <i>df</i> (2) | | |
|--|----------|----------------------------------|---------------|------------------|-----------------------------------|----------------|------------------|---------------------------|---------------|------------------|
| | | MS | <i>F</i> | <i>p</i> | MS | <i>F</i> | <i>p</i> | MS | <i>F</i> | <i>p</i> |
| Chlorophyll <i>a</i> (mg Chl <i>a</i> FW ⁻¹) | 8 | 0.001 | 0.283 | 0.755 | 0.029 | 14.755 | <0.001 | 0.001 | 0.698 | 0.504 |
| Phycocyanin (mg Chl <i>a</i> FW ⁻¹) | 8 | 0.000 | 5.765 | 0.006 | 0.001 | 20.613 | <0.001 | 0.000 | 1.879 | 0.165 |
| Phycocerytrin (mg Chl <i>a</i> FW ⁻¹) | 8 | 0.001 | 14.310 | <0.001 | 0.005 | 116.086 | <0.001 | 0.000 | 8.622 | <0.001 |
| Net productivity (mg C L ⁻¹ h ⁻¹) | 8 | 0.047 | 24.726 | <0.001 | 0.582 | 305.930 | <0.001 | 0.006 | 3.317 | 0.046 |
| Respiration rate (mg C L ⁻¹ h ⁻¹) | 8 | 0.004 | 52.360 | <0.001 | 0.001 | 12.690 | 0.001 | 0.002 | 20.910 | <0.001 |
| Survival (%) | 5 | 0.023 | 0.766 | 0.4757 | 2.135 | 72.039 | <0.001 | 0.044 | 1.485 | 0.247 |
| Growth (%) | 5 | 0.098 | 0.861 | 0.435 | 24.95 | 219.807 | <0.001 | 0.429 | 3.778 | 0.037 |

Bold characters represent significant values

df degrees of freedom, *n* number of replicates, *p* probability level

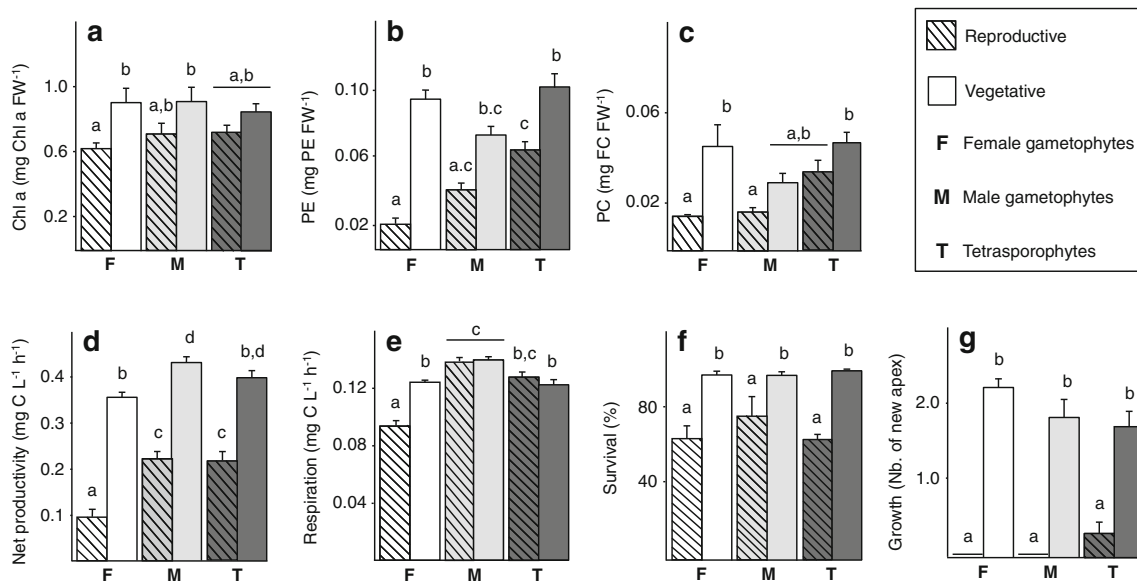


Fig. 1 Comparison between reproductive and vegetative fronds of female gametophytes, male gametophytes, and tetrasporophytes for Chl a concentration (a), PE concentration (b), PC concentration (c), net productivity (d), respiration (e), survival of 1 cm length segments after 50 days (f), and

growth, represented as the number of new apices generated by the 1 cm length segments after 50 days (g). Error bars indicate standard error of the mean, and significant differences are represented by different letters (HSD tests, $p < 0.05$)

Mathieson and Guo 1992; Pfister 1992; Zou et al. 2006) and (2) the existence of a threshold size necessary to achieve reproductive maturity (Gómez and Westermeier 1991; Dyck and De Wreede 2006).

Interestingly, in our study, cystocarp, spermatangia, and sporangia development were concomitant with a reduction of photosynthetic rate. This was usually associated with a slower growth rate and a higher mortality rate of reproductive fronds in comparison to vegetative fronds. We hypothesize that the lower net productivity observed in reproductive fronds of *G. chilensis* is directly linked to lower pigment concentration as suggested by Yokoya et al. (2007). In *Gracilaria*, development of reproductive organs as well as the release of tetraspores borne in cortical tissue could affect directly the photosynthetic activity, decreasing the pigment concentration. The negative effect of carposporophyte (cystocarp) on female growth rate has been reported previously in different *Gracilaria* species (*Gracilaria sjoestedtii*: Zhang and van der Meer 1988; *G. chilensis*: Santelices and Varela 1995; *G. domingensis*: Guimarães et al. 1999). These results suggest that the development of carposporophyte depends nutritionally on the female gametophyte (Hommersand and Fredericq 1995). Similar relationships showing a negative trade-off between gametophyte growth rate and sporophyte production were also observed in some bryophytes (Bisang and Ehrlen 2002; Rydgren and Økland 2003). Our results corroborate previous studies on *G. chilensis* showing that the reproductive stages have lower growth rate compared to the vegetative stages (Santelices and Varela 1995; Halling et al. 2005) and suggest that reduced growth rate is due to extra energy costs for producing gametes and spores (Kain and Destombe 1995).

Generally, the existence of long-term demographic effects of current reproduction on subsequent mortality, growth, or reproduction has remained somewhat unclear in algae, as incongruent results have been reported (De Wreede and Klinger 1990; Ang 1992; Dyck and De Wreede 2006). Low reproductive costs were suggested since reproductive structures are potentially capable of photosynthesis (Santelices 1990). Moreover, it has been hypothesized that the comparatively low cost of reproductive structures in relation to the overall energy budget of the plant might minimize the cost of reproduction in algae. In fucoid species, Vernet and Harper (1980) estimated that a very low proportion of body weight was invested in gamete production (<0.4 % for female gametes and <0.02 % for male gametes). Pfister (1992) suggested that plastic or compensatory responses to the environment may minimize reproduction cost on kelps, and Ang (1992) argued that the failure to detect cost of reproduction with respect to mortality and longevity in *Fucus distichus* may be due to the modular nature of the plants, where cost occurs at the level of the modules (branches or fronds) rather than at the level of the whole plant (individual). In fact, in *G. chilensis*, Santelices and Varela (1995) have proposed that the negative impact of reproductive structures in female (i.e., cystocarps) plants could be highly localized. Even though the long-term cost of reproduction at the individual level was not measured in our study, the lower growth and survival of reproductive fronds may have a strong impact on individual fitness. In short-lived seaweeds (or semelparous), release of gametes or spores usually initiates the degradation of the photosynthetic tissues and bouts of sexual reproduction have a clear negative impact on local algal abundance (Clifton and Clifton 1999;

Sánchez et al. 2003). In perennial species (or iteroparous), reproduction is generally delayed and depends on the individual size (Åberg 1992). While fronds constitute the primary photosynthesis organ and only seasonally bear reproductive structure, spores production and individual volume are inextricably linked in *Gracilaria gracilis* (Engel et al. 2001). In *G. chilensis*, reproductive cost could have a higher demographic impact in estuaries, where populations reproduce mostly by vegetative fragmentation of fronds than in rocky shore, where individuals are fixed by a perennial holdfast and reproduce solely by spores (Guillemin et al. 2008).

In conclusion, the physiological and demographic parameters analyzed in this study allowed the exploration of reproductive cost in red haploid–diploid seaweeds. The results revealed the existence of a short-term reproductive cost in *G. chilensis* whatever the type of individual studied (male gametophyte, female gametophyte, and tetrasporophyte), providing valuable insight into demographic processes at a local scale. In fact, our results demonstrated that vegetative fronds have a higher survival rate and a better growth rate than reproductive ones. These findings could have useful implications in the management and farming practices of *G. chilensis*. Nevertheless, there is a need to carry out new experimental approach at the population level to estimate the impact of reproduction on individual survival in different local ecological environments, such as muddy or rocky shore. Similarly, since ecological differences have been reported in this species (Santelices and Ugarte 1990), it is important to understand to what extent environmental variation can influence changes in the energy allocation and trade-offs between reproduction, growth, and survival across the geographical range of distribution of *G. chilensis* and future studies should be performed in natural populations located in other regions in order to generalized our results.

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References

- Åberg P (1992) A demographic study of two populations of the seaweed *Ascophyllum nodosum* in stochastic environment. *Ecology* 73:1473–1487
- Åberg P (1996) Patterns of reproductive effort in the brown alga *Ascophyllum nodosum*. *Mar Ecol Prog Ser* 138:199–207
- Ang PO (1992) Cost of reproduction in *Fucus distichus*. *Mar Ecol Prog Ser* 28:25–35
- Ashman T (1994) A dynamic perspective on the physiological cost of reproduction in plants. *Am Nat* 144:300–316
- Bazzaz FA, Ackerly DD, Reekie EG (2000) Reproductive allocation in plants. In: Fenner M (ed) *Seeds: the ecology of regeneration in plant communities*. CABI Publishing, Wallingford, pp 1–30
- Beer S, Eshel A (1985) Determining phycoerythrin and phycocyanin concentrations in aqueous crude extracts of red algae. *Aust J Mar Freshw Res* 36:785–792
- Bell G (1984) Measuring the cost of reproduction. 1. The correlation structure of the life table of a plankton rotifer. *Evolution* 38:300–313
- Bird CJ, McLachlan J (1986) The effect of salinity on distribution of species of *Gracilaria* Grev. (Rhodophyta, Gigartinales): an experimental assessment. *Bot Mar* 29:231–238
- Bisang I, Ehrlén J (2002) Reproductive effort and cost of sexual reproduction in female *Dicranum polysetum*. *Bryologist* 105:384–397
- Buschmann AH, Correa JA, Westermeier R, Hernandez-Gonzalez MDC, Norambuena R (2001) Red algal farming in Chile: a review. *Aquaculture* 194:203–20
- Chu S, Zhang Q, Liu S, Zhang S, Tang Y, Lu Z, Yu Y (2011) Trade-off between vegetative regeneration and sexual reproduction of *Sargassum thunbergii*. *Hydrobiologia* 678:127–135
- Clifton KE, Clifton LM (1999) The phenology of sexual reproduction by green algae (Bryopsidales) on Caribbean coral reefs. *J Phycol* 35:24–34
- Correa JA, McLachlan JL (1991) Endophytic algae of *Chondrus crispus* (Rhodophyta). III. Host specificity. *J Phycol* 27:448–459
- De Wreede R, Klinger T (1990) Reproductive strategies in algae. In: Lovett-Doust J, Lovett-Doust L (eds) *Plant reproductive ecology: patterns and strategies*. Oxford University Press, New York, pp 267–284
- Dyck LJ, DeWreede RE (2006) Reproduction and survival in *Mazzaella splendens* (Gigartinales, Rhodophyta). *Phycologia* 45:302–310
- Engel C, Åberg P, Gaggiotti O, Destombe C, Valero M (2001) Population dynamics and stage structure in a haploid–diploid red seaweed, *Gracilaria gracilis*. *J Ecol* 89:436–450
- Fisher RA (1930) *The genetical theory of natural selection*. Clarendon Press, Oxford
- Ganzon-Fortes ET (1999) Photosynthetic and respiratory responses of the agarophyte *Gelidiella acerosa* collected from tidepool, intertidal and subtidal habitats. *Hydrobiologia* 398/399:321–328
- Gao S, Chen XY, Yi QQ, Wang GC, Pan GH, Lin AP, Peng G (2010) A strategy for the proliferation of *Ulva prolifera*, main causative species of green tides, with formation of sporangia by fragmentation. *PLoS One* 5:e8571
- Gómez IM, Westermeier RC (1991) Frond regrowth from basal disc in *Iridaria laminarioides* (Rhodophyta, Gigartinales) at Mehuín, southern Chile. *Mar Ecol Prog Ser* 73:83–91
- Gómez I, Figueroa F, Huovin P, Ulloa N, Morales V (2005) Photosynthesis of the red alga *Gracilaria chilensis* under natural solar radiation in an estuary in southern Chile. *Aquaculture* 244:369–382
- Guillemin ML, Faugeton S, Destombe C, Viard F, Correa JA, Valero M (2008) Genetic variation in wild and cultivated populations of the haploid–diploid red alga *Gracilaria chilensis*: how farming practices favour asexual reproduction and heterozygosity. *Evolution* 62:1500–1519
- Guimarães M, Plastino E, Oliveira E (1999) Life history, reproduction and growth of *Gracilaria domingensis* (Gracilariales, Rhodophyta) from Brazil. *Bot Mar* 42:481–486
- Guzmán-Urióstegui A, García-Jiménez P, Marián FD, Robledo D, Robaina RR (2002) Polyamines influence maturation in reproductive structures of *Gracilaria cornea* (Gracilariales, Rhodophyta). *J Phycol* 38:1169–75
- Halling C, Aroca G, Cifuentes M, Buschmann A, Troell M (2005) Comparison of spore inoculated and vegetative propagated cultivation methods of *Gracilaria chilensis* in an integrated seaweed and fish cage culture. *Aquacult Int* 13:409–422
- Harper JL (1967) A Darwinian approach to plant ecology. *J Ecol* 55:247–270

- Hommersand MH, Fredericq S (1995) Sexual reproduction and cystocarp development. In: Cole KM, Sheath RG (eds) *Biology of the red algae*. Cambridge University Press, New York, pp 305–345
- Howarth R, Michaels A (2000) The measurement of primary production in aquatic ecosystems. In: Sala OE, Jackson RB, Mooney HA, Howarth RW (eds) *Methods in ecosystem science*. Springer-Verlag, New York, pp 74–85
- Inskip WP, Bloom PR (1985) Extinction coefficients of chlorophyll *a* and *b* in *N,N*-dimethylformamide and 80 % acetone. *Plant Physiol* 77:483–485
- Kain JM, Destombe C (1995) A review of the life history, reproduction and phenology of *Gracilaria*. *J Appl Phycol* 7:269–281
- Kamiya M, Kawai H (2002) Dependence of the carposporophyte on the maternal gametophyte in three ceramiacean algae (Rhodophyta), with respect to carposporophyte development, spore production and germination success. *Phycologia* 41:107–115
- Mathieson AC, Guo Z (1992) Patterns of fucoid reproductive biomass allocation. *Br Phycol J* 27:37–41
- McCourt RM (1985) Reproductive biomass allocation in three *Sargassum* species. *Oecologia* 67:113–117
- Meneses I (1996) Sources of morphological variation in populations of *Gracilaria chilensis* Bird, McLachlan & Oliviera of Chile. *Rev Chil Hist Nat* 69:35–44
- Obeso J (2002) The costs of reproduction in plants. *New Phytol* 155:321–348
- Perrone C, Felicini GP (1988) Physiological ecology of *Schottera nicaeensis* (Phylloporaceae, Rhodophyta): functional significance of heterotrichy. *Phycologia* 27:347–354
- Pfister C (1992) Costs of reproduction in an intertidal kelp: patterns of allocation and life history consequences. *Ecology* 73:1586–1596
- Rameau C, Gouyon PH (1991) Resource allocation to growth, reproduction and survival in *Gladiolus*: the cost of male function. *J Evol Biol* 4:291–307
- Reznick D, Nunney L, Tessier A (2000) Big houses, big cars, superfleas and the cost of reproduction. *Trends Ecol Evol* 15:421–425
- Rydgren K, Økland RH (2003) Short-term costs of sexual reproduction in the clonal moss *Hylocomium splendens*. *Bryologist* 106:212–220
- Sánchez I, Fernández C, Rico JM (2003) Distribution, abundance and phenology of two species of *Liagora* (Nemaliales, Rhodophyta) in northern Spain. *Phycologia* 42:7–17
- Santelices B (1990) Patterns of reproduction, dispersal and recruitment in seaweeds. *Oceanogr Mar Biol Annu Rev* 28:177–276
- Santelices B, Ugarte R (1990) Ecological differences among Chilean populations of commercial *Gracilaria*. *J Appl Phycol* 2:17–26
- Santelices B, Varela D (1995) Regenerative capacity of *Gracilaria* fragments: effects of size, reproductive state and position along the axis. *J Appl Phycol* 7:501–506
- Stearns SC (1992) *The evolution of life histories*. Oxford University Press, New York
- Thornber CS (2006) Functional properties of the isomorphic biphasic algal life cycle. *Integr Comp Biol* 46:605–614
- Vernet P, Harper JL (1980) The costs of sex in seaweeds. *Biol J Linn Soc* 13:129–138
- Wheelwright N, Logan BA (2004) Previous-year reproduction reduces photosynthetic capacity and slows lifetime growth in females of a neotropical tree. *Proc Natl Acad Sci USA* 101:8051–8055
- Yang RL, Zhou W, Shen SD, Wang GC, He LW, Pan GH (2012) Morphological and photosynthetic variations in the process of spermatia formation from vegetative cells in *Porphyra yezoensis* Ueda (Bangiales, Rhodophyta) and their responses to desiccation. *Planta* 235:885–893
- Yokoya YS, Necchi O Jr, Martins AP, Gonzalez SF, Plastino EM (2007) Growth responses and photosynthetic characteristics of wild and phycoerythrin-deficient strains of *Hypnea musciformis* (Rhodophyta). *J Appl Phycol* 19:197–205
- Zhang X, van de Meer JP (1988) A genetic study on *Gracilaria sjoestedtii*. *Can J Bot* 66:2022–2026
- Zou DH, Gao KS, Ruan ZX (2006) Seasonal pattern of reproduction of *Hizikia fusiformis* (Sargassaceae, Phaeophyta) from Nanao Island, Shantou, China. *J Appl Phycol* 18:195–201
- Zou D, Gao K, Chen W (2011) Photosynthetic carbon acquisition in *Sargassum henslowianum* (Fucales, Phaeophyta), with special reference to the comparison between the vegetative and reproductive tissues. *Photosynth Res* 107:159–168