

# Effects of nitrate on intracellular nitrite and growth of *Microcystis aeruginosa*

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**Abstract** Although nitrate is a macronutrient and can serve as good nitrogen source for many species of phytoplankton, high nitrate concentrations do not benefit the growth of phytoplankton. We hypothesise that algae cultured under high nitrate concentrations can accumulate intracellular nitrite, which is produced by nitrate reductase (NR) and can inhibit the growth of algae. To assess the validity of this hypothesis, *Microcystis aeruginosa* was grown under different nitrate concentrations from 3.57 to 21.43 mM in low CO<sub>2</sub> and high CO<sub>2</sub> conditions for 15 days. We observed that, with increasing nitrate concentrations, the intracellular nitrite concentrations of the alga increased and the growth rates and photosynthesis declined. When grown under high CO<sub>2</sub> conditions, *M. aeruginosa* showed lower intracellular nitrite concentrations and higher growth rates and  $P_m^{chl a}$ ,  $R_d^{chl a}$ ,  $\alpha^{chl a}$  than under low CO<sub>2</sub> conditions. These results suggest that the accumulation of intracellular nitrite could be the cause of inhibition of algal growth under high nitrate concentrations.

**Keywords** Growth characteristics · Photosynthesis · Intracellular nitrite · Nitrate reductase · Nitrite reductase · *Microcystis aeruginosa* · Cyanobacteria

## Introduction

Under certain conditions, a combination of high nutrient load and warm, stable conditions, cyanobacteria can grow excessively and form blooms which cause a variety of

water quality problems, including toxin production, odors, scums and unsafe drinking water (Paerl 1988; Benoufella et al. 1994). Nutrient concentrations play an important role in the growth of phytoplankton, and the nitrogen sources considered most important for the growth of phytoplankton are nitrate and ammonium (Viaroli et al. 1995; Herndon and Cochlan 2007). Nitrate occurs naturally from mineral sources and animal wastes, and anthropogenically as a by-product of agriculture and human wastes (Madison and Burnett 1985). In many parts of the world, nitrate may build up and, as a result, aquatic nitrate concentrations range from normal background level of below 0.71 mM to over 7.14 mM (Rouse et al. 1999), while in some marine aquaculture systems, nitrate concentrations can approach values of 35.71 mM (De Graaf 1964; Pierce et al. 1993).

Although many studies have reported that nitrate could affect photosynthesis, growth and cellular toxicity of phytoplankton and metal toxicity to phytoplankton (Bates et al. 1993; Rijstenbil et al. 1998; Menéndez 2005), there are few reports on the accumulation of intracellular nitrite under high nitrate concentrations. We know that, once inside the alga, nitrate is converted to ammonium (reduced nitrogen) in two successive steps catalysed by nitrate reductase (NR) and nitrite reductase (NiR) in the cytosol and chloroplast, respectively, before being incorporated into organic compounds. Generally, NR catalysing the reduction of nitrate to nitrite is the rate-limiting step for the conversion of nitrate to ammonium (Maldonado et al. 1996). However, nitrate can regulate NR activity which increases with increasing nitrate concentrations (Crawford 1995; Sivasankar and Oaks 1996), and when nitrite formed by NR is more than the nitrite reduced by NiR, the accumulation of intracellular nitrite will occur.

Nitrite ion has a negative influence upon phytoplankton (Abe et al. 2002; Yang et al. 2004). Sijbesma et al. (1996)

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has reported that nitrite ion acted as a protonophore, an uncoupler that increases the proton permeability of membranes by a shunting mechanism, inhibiting ATP synthesis and stimulating ATP hydrolysis. Almeida et al. (1995) proposed that nitrite could induce a lower efficiency of respiratory-chain-linked energy conservation, due to an increase of proton permeability by the nitrite ion, counteracting the proton pumping effect of ATP-ase.

In order to gain some insight on accumulation of intracellular nitrite in algae under high nitrate concentrations and the effects of increasing intracellular nitrite, this study examined the effects of different levels nitrate on the specific growth rate, NR activities, NiR activities, intracellular nitrite concentrations and photosynthesis of *Microcystis aeruginosa* under both low and high CO<sub>2</sub> conditions. Suzuki et al. (1995) and Hu and Zhang (2007) have reported that high CO<sub>2</sub> concentrations enhanced NiR activities, but had no effect on NR activities, which could decrease intracellular nitrite concentration. *Microcystis aeruginosa* is recognised as one of the most common bloom-forming cyanobacteria found in fresh waters (Colman 1989).

## Materials and methods

*Microcystis aeruginosa* was obtained from the Institute of Hydrobiology, Chinese Academy of Sciences (Wuhan, China). It was grown in 1,000-mL flasks containing 500 mL sterilised BG-11 medium in a plant growth chamber at 25°C, 40 μmol photons m<sup>-2</sup> s<sup>-1</sup> and a 12 h light:12 h dark cycle. NaNO<sub>3</sub> was added to the BG-11 medium (Saito et al. 2002) (N omitted) to gain three treatments (3.57, 10.71, 21.43 mM nitrate). The experiment was divided into a low CO<sub>2</sub> group and a high CO<sub>2</sub> group; in the low CO<sub>2</sub> group, three treatments were aerated with air at 100 mL min<sup>-1</sup>, and in the high CO<sub>2</sub> group were aerated with 5% CO<sub>2</sub> air at 100 mL min<sup>-1</sup>. Every treatment had three replicates. The initial cell density was about 5 × 10<sup>4</sup> cells mL<sup>-1</sup>.

**Cell density** Cell density was monitored daily with a phytoplankton-counting chamber (0.1 mL). Specific growth rates (μ) were calculated using the equation  $\mu = \ln(X_t/X_0)/t$ , where X<sub>0</sub> is the initial cell density, X<sub>t</sub> is the cell density after t days.

**Intracellular nitrite concentration** The culture was centrifuged at 3,000 g for 10 min on day 10, and rinsed twice with a solution of 0.18 mM NaHCO<sub>3</sub>. After centrifugation, the algal pellet was mixed with 10 mL 5% (w/v) trichloroacetic acid, disintegrated ultrasonically for 10 min in an ice bath, and placed in a 45°C water bath for 30 min. The mixture was centrifuged and the nitrite concentrations of supernatant were determined by the N-1-naphthylethylene-

diamine method (Wei and Qi 2002). The intracellular nitrite concentrations of alga were calculated from the biomass of the alga and the nitrite contents of supernatant.

**NR activities and NiR activities** The culture was centrifuged at 3,000 g for 10 min on day 10, and proteins were extracted with a buffer containing 50 mM Hepes (pH 7.6), 10 mM MgCl<sub>2</sub>, 5 μM flavin adenine dinucleotide (FAD), 1 μM leupeptine, 10 mM 2-mercaptoethanol and polyclar 50 g L<sup>-1</sup>. NR activities were measured as described in Pigaglio et al. (1999). NiR activities were assayed as described by Lillo (1984). The NR activities were expressed as mg NO<sub>2</sub><sup>-</sup>N formed per cell and NiR activities were expressed as mg NO<sub>2</sub><sup>-</sup>N reduced per cell.

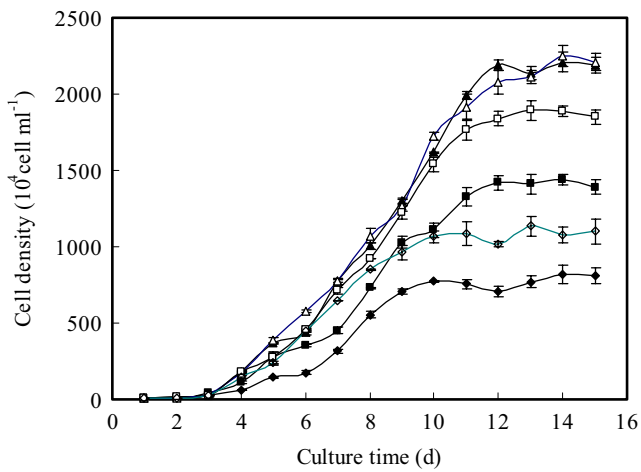
**Photosynthetic activities** The culture was centrifuged at 3,000 g for 10 min on day 10, and resuspended in BG-11 medium. Their photosynthetic activities were assayed by measuring the rate of O<sub>2</sub> evolution under different irradiances using a Clark-type O<sub>2</sub> electrode (Hansatech Instruments, UK). The temperature was kept at 25°C by a circulating water bath. Data were treated by non-linear fitting technique using model  $P = P_m \times \tanh(\alpha \times I/P_m) + R_d$  (Henley 1993), where I = irradiance, and P = photosynthetic rate at certain irradiance, P<sub>m</sub> = light-saturated photosynthesis, α = the initial slope at limiting irradiance, was calculated to assess the photosynthetic efficiency and R<sub>d</sub> = dark respiration rate.

**Statistical analysis** All treatments were analysed using an analysis of variance (ANOVA). Results of all tests were considered significant at 95% confidence if  $p < 0.05$  for a given *F* statistic test value.

## Results

### Growth of *Microcystis aeruginosa*

The effect of different nitrate concentrations on growth are shown in Fig. 1 and Table 1. The growth of *Microcystis aeruginosa* tended to be inhibited with nitrate concentrations increasing in both the high CO<sub>2</sub> group ( $p = 0.011$ , two-way ANOVA) and the low CO<sub>2</sub> group ( $p = 0.009$ , two-way ANOVA). When aerated with 5% CO<sub>2</sub> air, the specific growth rates and maximum cell densities of alga at 10.71 and 21.43 mM nitrate showed a significant increase in comparison with the low CO<sub>2</sub> group ( $p = 0.005$ , two-way ANOVA). Maximum cell densities at 10.71 and 21.43 mM nitrate in the high CO<sub>2</sub> group were, respectively, approx. 1.32- and 1.34-fold that in low CO<sub>2</sub> group, and specific growth rates were approx. 1.05- and 1.06-fold, but these



**Fig. 1** The growth curves of *Microcystis aeruginosa* at different nitrate concentrations. 3.57 mM nitrate (▲), 10.71 mM nitrate (■), 21.43 mM nitrate (◆) in low CO<sub>2</sub> group; 3.57 mM nitrate (△), 10.71 mM nitrate (□), 21.43 mM nitrate (◇) in high CO<sub>2</sub> group. Error bars denote the standard deviation of triplicate incubations

values did not markedly change when nitrate concentrations were 3.57 mM ( $p=0.92$ , one-way ANOVA)

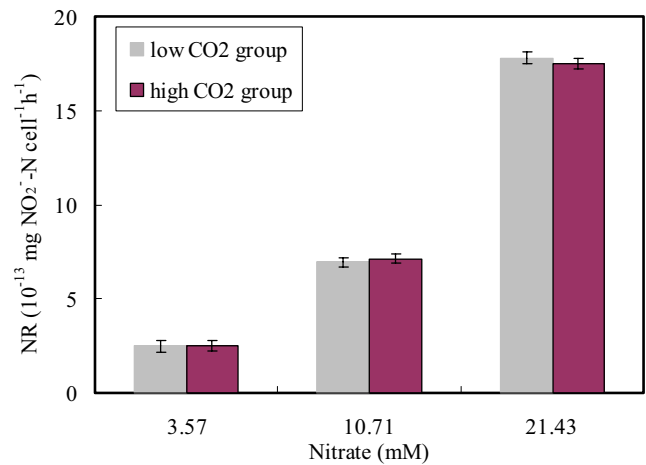
**NR activities and NiR activities**

After 10 days cultivation, NR activities and NiR activities showed a significant increase with increasing nitrate concentrations in both the high CO<sub>2</sub> group ( $p=0.002$ ,  $p=0.004$ , one-way ANOVA) and the low CO<sub>2</sub> group ( $p=0.008$ ,  $p=0.002$ , one-way ANOVA) (Figs. 2 and 3). In the low CO<sub>2</sub> group, the NR activities at 10.71 and 21.43 mM nitrate were, respectively, 2.79- and 7.15-fold higher than at 3.57 mM nitrate, and the NiR activities at 10.71 and 21.43 mM nitrate were, respectively, 1.12- and 1.53-fold higher than at 3.57 mM nitrate. 5% CO<sub>2</sub> air significantly enhanced NiR activities ( $p=0.017$ , two-way ANOVA), compared to the low CO<sub>2</sub> group, the NiR activities at 3.57, 10.71 and 21.43 mM nitrate were increased, respectively by 115, 179 and 175%. However, high CO<sub>2</sub>

**Table 1** Maximum cell densities and specific growth rates of *Microcystis aeruginosa* grown under different nitrate concentrations

Nitrate (mM)	Specific growth rates (day <sup>-1</sup> )	Maximum cell densities (×10 <sup>4</sup> cells mL <sup>-1</sup> )
Aerated with air	3.57	0.43
	10.71	0.40
	21.43	0.36
Aerated with 5% CO <sub>2</sub> air	3.57	0.43
	10.71	0.42
	21.43	0.38
		2,210±110
		1,440±63
		820±56
		2,250±127
		1,895±70
		1,100±68

Data are means ± SD (n=3)

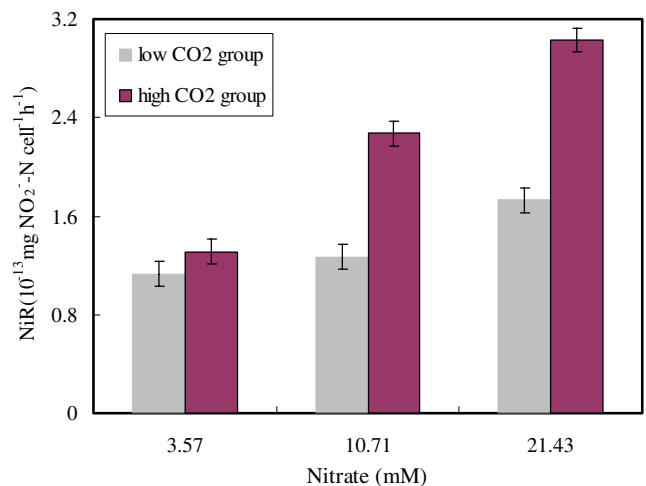


**Fig. 2** The NR activities of *Microcystis aeruginosa* at different nitrate concentrations. Error bars denote the standard deviation of triplicate incubations

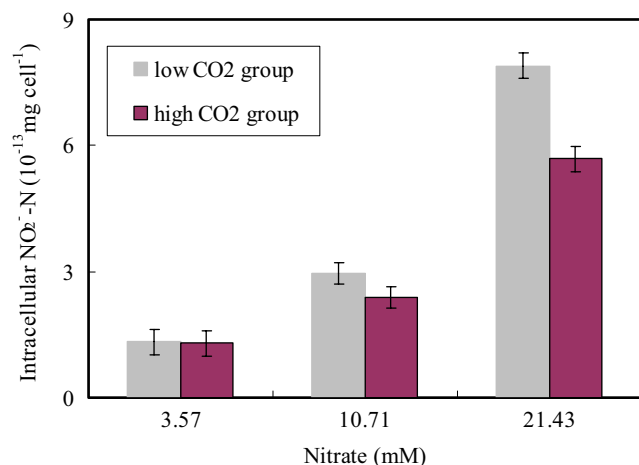
conditions (high CO<sub>2</sub> group) did not significantly influence NR activities compared to low CO<sub>2</sub> conditions (low CO<sub>2</sub> group) ( $p=0.717$ , two-way ANOVA).

**Intracellular nitrite concentrations**

On day 10, intracellular nitrite concentrations of *M. aeruginosa* increased with nitrate increasing concentrations in both the high CO<sub>2</sub> group ( $p=0.003$ , one-way ANOVA) and the low CO<sub>2</sub> group ( $p=0.004$ , one-way ANOVA) (Fig. 4). In the low CO<sub>2</sub> group, the intracellular nitrite concentrations at 10.71 and 21.43 mM nitrate were, respectively, 2.12- and 13.38-fold higher than at 3.57 mM nitrate. When aerated with 5% CO<sub>2</sub> air, the intracellular nitrite concentrations at 10.71 and 21.43 mM nitrate were, respectively, 0.81- and 0.72-fold that of the low CO<sub>2</sub> group, but no significant difference was



**Fig. 3** The NiR activities of *Microcystis aeruginosa* at different nitrate concentrations. Error bars denote the standard deviation of triplicate incubations



**Fig. 4** The intracellular nitrite concentrations of *Microcystis aeruginosa* at different nitrate concentrations. Error bars denote the standard deviation of triplicate incubations

observed for alga at 3.57 mM nitrate ( $p=0.68$ , one-way ANOVA).

#### Photosynthesis

Increasing nitrate concentrations inhibited the chlorophyll *a*-specific light-saturated photosynthetic rate ( $P_m^{chl a}$ ), dark respiration rate ( $R_d^{chl a}$ ) and photosynthetic efficiency ( $\alpha^{chl a}$ ) in both high the CO<sub>2</sub> group ( $p=0.009$ ,  $p=0.012$ ,  $p=0.032$ , one-way ANOVA) and the low CO<sub>2</sub> group ( $p=0.002$ ,  $p=0.007$ ,  $p=0.041$ , one-way ANOVA) (Table 2).  $P_m^{chl a}$ ,  $R_d^{chl a}$  and  $\alpha^{chl a}$  at 10.71 and 21.43 mM nitrate in the high CO<sub>2</sub> group showed a significant increase, compared to the low CO<sub>2</sub> group ( $p=0.011$ ,  $p=0.007$ ,  $p=0.032$ , two-way ANOVA). However, at 3.57 mM nitrate there was no significant difference in  $P_m^{chl a}$ ,  $R_d^{chl a}$  and  $\alpha^{chl a}$  between the low and high CO<sub>2</sub> groups ( $p=0.95$ ,  $p=0.79$ ,  $p=0.34$ , one-way ANOVA).

#### Discussion

Nitrate is a macronutrient and can serve as a good nitrogen source for phytoplankton (Dortch, 1990; Syrett 1981). Menéndez (2005) reported that moderate levels of fertilizing nitrate could increase the maximum rate of net photosynthesis of the green alga *Chaetomorpha linum*. Touzet et al. (2007) also found that the dinoflagellate *Alexandrium minutum* showed higher maximum cell densities with increasing nitrate concentrations. Similarly, Leong et al. (2004) showed a positive correlation between the growth rates of *A. tamarense* and nitrate concentrations. However, Shi et al. (2005) observed that there was a negative effect of excess nitrate on the growth of *A. tamarense*, and Hwang and Lu (2000) also reported *A.*

*minutum* exposed to excess nitrate had a low growth rate. These findings suggested that, although nitrate was a good nitrogen source, excess nitrate supply was harmful to phytoplankton and could depress growth, which is in agreement with the results of the present study.

As mentioned in the Introduction, nitrate is converted to ammonium in two successive steps catalysed by NR and NiR, and when nitrite formed by NR was more than nitrite reduced by NiR, the accumulation of intracellular nitrite of alga will appear. In the present study, *M. aeruginosa* grown under high nitrate concentrations showed a significant increase in NR activities, which is consistent with the results of Sivasankar and Oaks (1996) and Crawford (1995), where the activities of NR increased with nitrate increase. Although the NiR activities also increased with increasing nitrate concentrations, the extent of the NiR activity increases was less than that of NR activities, which caused an increase in intracellular nitrite concentrations under high nitrate concentrations. Compared to the low CO<sub>2</sub> group, at the high CO<sub>2</sub> condition (high CO<sub>2</sub> group), NiR activities were higher and NR activities remained constant, due to the high CO<sub>2</sub> condition enhancing NiR activities, but not NR activities (Suzuki et al. 1995; Hu and Zhang 2007). This resulted in a decrease in intracellular nitrite concentrations at the high CO<sub>2</sub> condition.

In general, excessive nitrate can be stored in cells without detrimental effect (Tylova-Munzarova et al. 2005), but nitrite is an inorganic monovalent anion which affects the process of photosynthesis quite significantly. It is known to inhibit photosynthetic electron transport (Spiller and Boger 1977; Loranger and Carpentier 1994), change intracellular pH and damage cell membranes of

**Table 2** Photosynthetic parameters of *Microcystis aeruginosa* grown under different nitrate concentrations

Photosynthetic parameters	Nitrate (mM)			
	3.57	10.71	21.43	
Aerated with air	$P_m^{chl a}$ $\mu\text{mol O}_2$ (mg chl <i>a</i> h) <sup>-1</sup>	863±11.3	575±10.5	349±10.8
	$\alpha^{chl a}$ $\mu\text{mol O}_2$ (mg chl <i>a</i> h) <sup>-1</sup> (μmol photons m <sup>-2</sup> s <sup>-1</sup> ) <sup>-1</sup>	1.3±0.2	0.9±0.1	0.4±0.1
	$R_d^{chl a}$ $\mu\text{mol O}_2$ (mg chl <i>a</i> h) <sup>-1</sup>	62.8±4.1	36.7±5.1	15.4±3.2
Aerated with 5% CO <sub>2</sub> air	$P_m^{chl a}$ $\mu\text{mol O}_2$ (mg chl <i>a</i> h) <sup>-1</sup>	872±13.2	729±9.3	517±11.2
	$\alpha^{chl a}$ $\mu\text{mol O}_2$ (mg chl <i>a</i> h) <sup>-1</sup> (μmol photons m <sup>-2</sup> s <sup>-1</sup> ) <sup>-1</sup>	1.3±0.1	1.1±0.1	0.6±0.1
	$R_d^{chl a}$ $\mu\text{mol O}_2$ (mg chl <i>a</i> h) <sup>-1</sup>	64.5±3.9	52.8±4.2	29.7±5.2

These parameters were determined by fitting a three-parameter model,  $P = P_m \times \tanh(\alpha \times I/P_m) + R_d$  (see "Materials and Methods") Data are means ± SD ( $n=3$ )



algae (Almeida et al. 1995; Sijbesma et al. 1996; Yang et al. 2004). In the present study, a significant increase was observed in intracellular nitrite concentrations under high nitrate concentrations. Therefore, it is possible that the increase of intracellular nitrite resulted in the decrease of growth and  $P_m^{chl a}$ ,  $R_d^{chl a}$ ,  $\alpha^{chl a}$  observed at high nitrate concentrations. Our data clearly support this conclusion. At 3.57 mM nitrate, the intracellular nitrite concentrations did not change significantly in the high and low CO<sub>2</sub> groups and, as a result,  $P_m^{chl a}$ ,  $R_d^{chl a}$ ,  $\alpha^{chl a}$  remained constant. However, at 10.71 and 21.43 mM nitrate, the intracellular nitrite concentrations were lower in the high CO<sub>2</sub> group than in the low CO<sub>2</sub> group, which resulted in an increase in  $P_m^{chl a}$ ,  $R_d^{chl a}$ ,  $\alpha^{chl a}$  in the high CO<sub>2</sub> group compared to the low CO<sub>2</sub> group.

The levels of nitrate concentrations used in the present study ranged from 3.57 to 21.43 mM. Much higher nitrate concentrations (from 7.14 to 35.71 mM) have been found in aquatic ecosystems which were strongly contaminated by agricultural and urban activities (Rouse et al. 1999; De Graaf 1964; Pierce et al. 1993). According to the present study, these high nitrate concentrations could become an inhibitor of growth of phytoplankton. Therefore, further research is required to investigate the actual effect of high nitrate concentrations on phytoplankton community composition and the occurrence of blooms in the field.

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## References

- Abe K, Imamaki A, Hirano M (2002) Removal of nitrate, nitrite, ammonium and phosphate ions from water by the aerial microalga *Trentepohlia aurea*. J Appl Phycol 14:129–134. doi:10.1023/A:1019599216554
- Almeida JS, Júlio SM, Reis MAM, Carrondo MJT (1995) Nitrite inhibition of denitrification by *Pseudomonas fluorescens*. Biotechnol Bioeng 46:194–201. doi:10.1002/bit.260460303
- Bates SS, Worms J, Smith JC (1993) Effects of ammonium and nitrate on growth and domoic acid production by *Nitzschia pungens* in batch culture. Can J Fish Aquat Sci 50:1248–1254. doi:10.1139/f93-141
- Benoufella F, Laplanche A, Boisdon V, Bourbigot MM (1994) Elimination of *Microcystis* cyanobacteria (blue-green algae) by an ozoflotation process: a pilot plant study. Water Sci Technol 30:245–257
- Colman B (1989) Photosynthetic carbon assimilation and the suppression of photorespiration in the cyanobacteria. Aquat Bot 34:211–231. doi:10.1016/0304-3770(89)90057-0
- Crawford NM (1995) Nitrate: nutrient and signal for plant growth. Plant Cell 7:859–868
- De Graaf F (1964) Maintenance problems in large public aquaria. Arch Neerland Zool 16:142–143
- Dortch Q (1990) Review: the interaction between ammonium and nitrate uptake in phytoplankton. Mar Ecol Prog Ser 61:183–201. doi:10.3354/meps061183
- Henley WJ (1993) Measurement and interpretation of photosynthetic light-response curves in alga in the context of photoinhibition and diel changes. J Phycol 29:729–739. doi:10.1111/j.0022-3646.1993.00729.x
- Herndon J, Cochlan WP (2007) Nitrogen utilization by the raphidophyte *Heterosigma akashiwo*: growth and uptake kinetics in laboratory cultures. Harmful Algae 6:260–270. doi:10.1016/j.hal.2006.08.006
- Hu HH, Zhang X (2007) Nitrite utilization by *Chaetoceros muelleri* under elevated CO<sub>2</sub> concentration. World J Microbiol Biotechnol 24:891–894. doi:10.1007/S 11274-007-9553-X
- Hwang DF, Lu YH (2000) Influence of environmental and nutritional factors on growth toxicity and toxin profile of dinoflagellate *Alexandrium minutum*. Toxicon 38:1491–1503. doi:10.1016/S0041-0101(00)00080-5
- Leong SCY, Murata A, Nagashima Y, Taguchi S (2004) Variability in toxicity of the dinoflagellate *Alexandrium tamarense* in response to different nitrogen sources and concentrations. Toxicon 43:407–415. doi:10.1016/j.toxicon.2004.01.015
- Lillo C (1984) Diurnal variations of nitrite reductase, glutamine synthetase, glutamate synthase, alanine aminotransferase and aspartate aminotransferase in barley leaves. Plant Physiol 61:214–218. doi:10.1111/j.1399-3054.1984.tb05899.x
- Loranger C, Carpentier R (1994) A fast bioassay for phytotoxicity measurements using immobilized photosynthetic membranes. Biotechnol Bioeng 44:178–183. doi:10.1002/bit.260440206
- Madison RJ, Burnett JO (1985) Overview of the occurrence of nitrate in groundwater of the United States. US Geol Surv Water Supply Pap 2275:93–105
- Maldonado JM, Barro F, Gonzales-Fontes A (1996) Nitrate assimilation and protein accumulation in tritordeum. Plant Physiol Biochem 34:721–726
- Menéndez M (2005) Effect of nutrient pulses on photosynthesis of *Chaetomorpha linum* from a shallow Mediterranean coastal lagoon. Aquat Bot 82:181–192. doi:10.1016/j.aquabot.2005.04.004
- Paerl HW (1988) Nuisance phytoplankton blooms in coastal estuarine inland waters. Limnol Oceanogr 33:823–847
- Pierce RH, Weeks JM, Prappas JM (1993) Nitrate toxicity to five species of marine fish. J World Aquacult Soc 24:105–107. doi:10.1111/j.1749-7345.1993.tb00156.x
- Pigaglio E, Durand N, Meyer C (1999) A conserved acidic motif in the N-terminal domain of nitrate reductase is necessary for the inactivation of the enzyme in the dark by phosphorylation and 14-3-3 binding. Plant Physiol 119:219–230. doi:10.1104/pp.119.1.219
- Rijstenbil JW, Dehairs F, Ehrlich R, Wijnholds JA (1998) Effect of the nitrogen status on copper accumulation and pools of metal-binding peptides in the planktonic diatom *Thalassiosira pseudonana*. Aquatic Toxicol 42:187–209. doi:10.1016/S0166-445X(97)00091-X
- Rouse JD, Bishop CA, Struger J (1999) Nitrogen pollution: an assessment of its threat to amphibian survival. Environ Health Perspect 107:799–803. doi:10.2307/3454576
- Saito K, Ishii H, Nishida F (2002) Purification of microcystins by DEAE and C<sub>18</sub> cartridge chromatography. Toxicon 40:97–101. doi:10.1016/S0041-0101(01)00201-X
- Shi Y, Hu H, Cong W (2005) Positive effects of continuous low nitrate levels on growth and photosynthesis of *Alexandrium tamarense* (Gonyaulacales, Dinophyceae). Phycol Res 53:43–48. doi:10.1111/j.1440-1835.2005.tb00356.x
- Sijbesma WFH, Almeida JS, Reis MAM, Santos H (1996) Uncoupling effect of nitrite during denitrification by *Pseudomonas fluorescens*: an in vivo P-NMR study. Biotechnol Bioeng 52:176–192. doi:10.1002/(SICI)1097-0290(19961005)52:1<176::AID-BIT18>3.0.CO;2-M

- Sivasankar S, Oaks A (1996) Nitrate assimilation in higher plants: the effect of metabolites and light. *Plant Physiol Biochem* 34:609–620
- Spiller H, Boger P (1977) Photosynthetic nitrite reduction by dithioerythritol and the effect of nitrite on electron transport in isolated chloroplasts. *Photochem Photobiol* 26:397–402. doi:10.1111/j.1751-1097.1977.tb07504.x
- Suzuki I, Sugiyama T, Omata T (1995) Regulation of nitrite reductase activity under CO<sub>2</sub> limitation in the cyanobacterium *Synechococcus* sp. PCC7942. *Plant Physiol* 107:791–796. doi:10.1104/pp.107.1.299
- Syrett PJ (1981) Nitrogen metabolism of microalgae. In: Platt T (ed) *Physiological Bases of phytoplankton Ecology*. Can Bull Fish. Aquatic Sciences 201:182–210.
- Touzet N, Franco JM, Raine R (2007) Influence of inorganic nutrition on growth and PSP toxin production of *Alexandrium minutum* (Dinophyceae) from Cork Harbour, Ireland. *Toxicon* 50:106–119. doi:10.1016/j.toxicon.2007.03.001
- Tylova-Munzarova E, Lorenzen B, Brix H, Votrubova O (2005) The effects of NH<sub>4</sub><sup>+</sup> and NO<sub>3</sub><sup>-</sup> on growth, resource allocation and nitrogen uptake kinetics of *Phragmites australis* and *Glyceria maxima*. *Aquat Bot* 81:326–342. doi:10.1016/j.aquabot.2005.01.006
- Viaroli P, Bartoli M, Bondavalli C, Naldi M (1995) Oxygen fluxes and dystrophy in a coastal lagoon colonized by *Ulva rigida* (Sacca di Goro, Po River Delta, northern Italy). *Fresenius Environ Bull* 4:381–386
- Wei FS, Qi WQ (2002) *Inspection measurement of water and waste*. China Environmental Science Press, pp 260–263
- Yang S, Wang J, Cong W, Cai ZL, Fan OY (2004) Utilization of nitrite as nitrogen source by *Botryococcus braunii*. *Biotechnol Lett* 26:239–243. doi:10.1023/B:BILE.0000013722.45527.18