



Ecological interactions between invasive and native fouling species in the reservoir of a hydroelectric plant

Ana Paula da Silva Bertão · Raissa Vitória Vieira Leite · Aline Horodesky · Marcio Roberto Pie · Thiago Luis Zanin · Otto Samuel Mader Netto · Antonio Ostrensky

Received: 28 September 2020 / Revised: 17 September 2021 / Accepted: 18 September 2021 / Published online: 7 October 2021
© The Author(s), under exclusive licence to Springer Nature Switzerland AG 2021

Abstract In this study, we investigate the main ecological interactions between fouling aquatic organisms (both invasive and native) present in the reservoir of the Governador José Richa hydroelectric plant, located in southern Brazil, and to identify the most suitable period for the interruption of machinery operation for cleaning and maintenance of the hydraulic systems of this plant. A total of 32 experimental plates were fixed to a metallic structure positioned close to the plant's water intake. Three species of invasive fouling were identified in our samples (*Limnoperna fortunei* [Mollusca], *Cordy-*

lophora sp., and *Hydra* sp. [Cnidaria]) and six native taxa belonging to the phyla Protozoa, Ciliophora, Amoebozoa, and Arthropoda. Spring and summer were the seasons with the highest fouling rates, as well as densities of fouling organisms. The highest levels of diversity were recorded during the colder seasons. Several interactions between the organisms were identified, such as mutualism, commensalism, competition, epibiosis, cannibalism, and predation. The data obtained suggest that, from the biological point of view, the most suitable period for machine shutdown destined for the removal of biological fouling in the

Handling editor: Eric R. Larson

A. P. da Silva Bertão (✉) · R. V. V. Leite · A. Ostrensky
Universidade Federal do Paraná - Graduate Program in
Animal Science, Curitiba, Brazil
e-mail: anabertaopaula@gmail.com

R. V. V. Leite
e-mail: Raisasleite0@hotmail.com

A. Ostrensky
e-mail: ostrensky@ufpr.br

A. P. da Silva Bertão · R. V. V. Leite ·
A. Horodesky · A. Ostrensky
Universidade Federal do Paraná - Integrated Group of
Aquaculture and Environment – GIA, R. dos
Funcionários, 1540, Juvevê, Curitiba,
PR 80035-050, Brazil
e-mail: aline.horo@yahoo.com.br

A. Horodesky · M. R. Pie · A. Ostrensky
Universidade Federal do Paraná - Graduate Program in
Zoology, Curitiba, Brazil
e-mail: marcio.pie@ufpr.br

T. L. Zanin
COPEL Geração e Transmissão S/A, GeT/SGE/DPEG/
VECM, Rua José Izidoro Biazetto, 158 - Mossunguê,
Curitiba, PR 82305-240, Brazil
e-mail: thiago.zanin@copel.com

O. S. M. Netto
Aliança Prestadora de Serviços Ltda, Av. Sete de
Setembro, 5402, Cj. 41 Batel, Curitiba,
PR 80240-000, Brazil
e-mail: ottomader@gmail.com

hydraulic systems of the studied plant is between the end of spring and the beginning of summer.

Keywords Ecological succession · Facilitation · Freshwater environment · Biofouling · Seasonality

Introduction

Fouling invasive aquatic organisms have caused severe economic impacts around the world (Morton, 1977; Callow, 1993; Simberloff & Von Holle, 1999; Pimentel et al., 2001; Ricciardi & MacIsaac, 2008; Nakano & Strayer, 2014; Simberloff & Vitule, 2014). There have been records of clogging of pipes in water collection and treatment stations (Melo & Bott, 1997; Pu et al., 2009; Rajagopal & van der Velde, 2012). Industries such as pulp and paper mills are also frequently impacted by fouling organisms, as these sectors depend directly on the capture of water from natural bodies to carry out their production processes (Coetser & Cloete, 2005; Rajagopal & van der Velde, 2012). In these cases, the damage caused by fouling organisms is associated with both increased corrosion rates and clogging of hydraulic structures (Characklis, 1981; Melo & Bott, 1997; Pu et al., 2009; Martin et al., 2016; Farhat et al., 2019; Singh et al., 2020). However, the most widely affected sector has been hydroelectric power generation (Armour et al., 1993; MacIsaac, 1996; Cataldo, 2001; Boltovskoy et al., 2006; Portella et al., 2009; Boltovskoy & Correa, 2015; Mansur et al., 2016; Boltovskoy, 2017; Jernelöv, 2017).

In hydroelectric projects affected by fouling, the frequency of interventions aimed at cleaning and maintaining hydraulic structures increases due to the accumulation of biological fouling (e.g., molluscs, cnidarians, protozoans), with a consequent increase in operating costs (Flemming & Grohmann, 2008b; Cloete 2010; Flemming, 2011; Booy et al., 2017; Nelson, 2019). Each machine stoppage requires skilled labor to carry out cleaning activities, in addition to the interruption of energy production during the entire cleaning period. Frota et al. (2014), based on the value of the energy priced in the Brazilian spot market, concluded that the lost profits for the energy not generated in a single day of maintenance of a 44 MW Francis turbine would be US\$ 180,000. The associated economic impact is even greater when

considering that machine shutdown for maintenance and cleaning are always greater than 24 h (Flemming, 2002; Grohmann, 2008a; Pucherelli et al., 2018). Therefore, identifying the most suitable periods for machine shutdowns for maintenance and cleaning would allow the optimization of efforts and costs in the mid- and long-term, given that it would allow for maintenance efforts to be more effective and concentrated in time, which in turn would minimize control efforts in the intervening months (Grohmann, 2008c; Venkatesan & Murthy, 2008; Oreska & Aldridge, 2011; Frota et al., 2014; Boltovskoy et al., 2015b; Latombe et al., 2017).

In South America, three fouling invasive aquatic species have been reported to cause significant operational and economic losses to companies in the hydroelectric sector: the Asian clam, *Corbicula fluminea* (OF Müller, 1774), the cnidarian *Cordylophora caspia* (Pallas, 1771), and the golden mussel *Limnoperna fortunei* (Dunker, 1857). *Corbicula fluminea* obstructs heat exchangers of plants due to the accumulation of their shells (Karatayev et al., 2005; Ludwig et al., 2014; Mansur et al., 2016). *Cordylophora caspia* affects hydraulic pipes, filters, and protective grids (Darrigran & De Drago, 2000; Portella et al., 2009; Sylvester et al., 2011; Nakano & Strayer, 2014). Finally, *L. fortunei* causes damage to the protection grids (Darrigran, 2002), filters, generators, floodgates (Boltovskoy et al., 2006), pipes, and heat exchangers (Cataldo et al., 2003; Darrigran & Damborenea, 2011; Uliano-Silva et al., 2013).

In addition to the economic impacts, fouling aquatic organisms have generated several negative impacts on the ecosystem in freshwater environments, interfering on the food webs of native species (David et al., 2017; Emery-Butcher et al., 2020). There are relatively well-documented cases of competition between these organisms and incumbent species for food and/or space, as well as shifts of nutrient cycling, among other environmental disturbances (Ricciardi & MacIsaac, 2011; David et al., 2017). Invasive species often interact with each other and often act as “facilitators” (a term that refers to those species that have direct positive effects on other species) (Hacker & Gaines, 1997). This interaction can favor the increase in density or biomass of at least one of the involved species, which can facilitate the bioinvasion process (Ricciardi & MacIsaac, 2000; Rodriguez, 2006; Velde & Rajagopal, 2006; Silknetter et al.,

2019). Thus, this type of interaction would contribute to the increase in the number of establishment or spread of species in aquatic environments into aquatic environments (Simberloff & Von Holle, 1999). The impacts of facilitating species are more frequent when they favor or provide a limiting resource, increasing the complexity of the habitat, functionally replacing a native species or competing for resources and ecological niche (Ricciardi et al., 1996; Simberloff & Von Holle, 1999; Ricciardi, 2001; Simberloff, 2006; Kéfi et al., 2016). The present study aimed to investigate the main ecological interactions between the invasive fouling aquatic species and the native species present in the reservoir of a hydroelectric plant located in the southern region of Brazil and to evaluate the most suitable period for machine shutdown for cleaning and maintenance of the hydraulic systems of this plant.

Methods

Study design

The experiment was carried out between September 2018 and December 2019, at the HPP Governador José Richa reservoir (25° 32' 34.99" S, 53° 29' 45.82" W), located in the Lower Iguazu River section, in the border between the municipalities of Capitão Leônidas Marques and Nova Prata do Iguazu, state of Paraná, Brazil. Samples were obtained using polystyrene experimental settlement plates (15 × 15 cm), fixed with nylon clamps to a metallic structure of 0.58 m². This structure was positioned one meter below the surface and was fixed by ropes to the log boom located near the dam of the HPP. For five seasons (Spring/2018: October, November, and December; Summer/2019: January, February, and March; Autumn/2019: April, May, and June; Winter/2019: July, August, and September; Spring /2019: October, November, and December), a total of 32 experimental plates were sequentially installed, six plates (two replicas/month) in each season, and two more test plates that were installed in the first month (September 2018) and removed only in the last month (December 2019). Each month, two experimental plates were removed, so that at the end of the season, all six seasonal plates had been removed (two plates/month). At the beginning of the next season, the procedure was repeated (Fig. 1).

The experimental settlement plates removed from the floating structures were packed in plastic bags. The biological material was fixed in 6% formaldehyde and preserved in 70% ethanol. Automated hourly measurement of the reservoir water temperature during the 15 months of the study was carried out and the data made available by Companhia Paranaense de Energia.

Analysis of experimental plates

In the laboratory, all experimental settlement plates were photographed, both with a digital camera (Nikon, Coolpix B500 Brindes, Japan) and in a stereoscopic microscope (Zeiss, Stereo Discovery.V8 Crisp, Germany), in order to organize an image database for visual analysis of the fouling process. The images were analyzed using the ImageJ software (version 1.52a) (Schneider et al., 2012) to identify the recruitment pattern and the degree of occupation of the experimental plates. The calculation of the coverage area in each experimental plate was performed using the Watershed plugin, a tool that calculates based on the intensity or gray level of the pixels present (Papadopoulos et al., 2007). The values were then transformed into the percentage of the fouled area in relation to the total area of the plate. After measuring the area occupied by fouling organisms, macroscopic individuals (> 1 cm) were removed from the experimental plates, with the aid of surgical instruments, fixed in 4% formaldehyde buffer, preserved in 70% alcohol, identified to the lowest possible taxonomic level, and quantified. The mollusks were also weighed to obtain the wet total weight (tissue and shell), on a scale (Bell Engineering, S2202h—2200 g, Italy), making it possible to calculate the relative biomass (g/cm²).

For the identification and quantification of microscopic organisms (< 1 cm), two methodological procedures were established. When the incrustated area was greater than 30% of the total area of the experimental settlement plate (as observed in Fig. 2a), subsampling was performed to quantify the organisms. For this purpose, a standardized grid was used, containing 225 squares (1 × 1 cm), of which 20 (8.8% of the total area) were analyzed. The selection of the squares was carried out randomly over the entire plate, including non-fouled surfaces (Fig. 2b), following the methodology proposed by Borges (2013). After this procedure, the biological material embedded in these

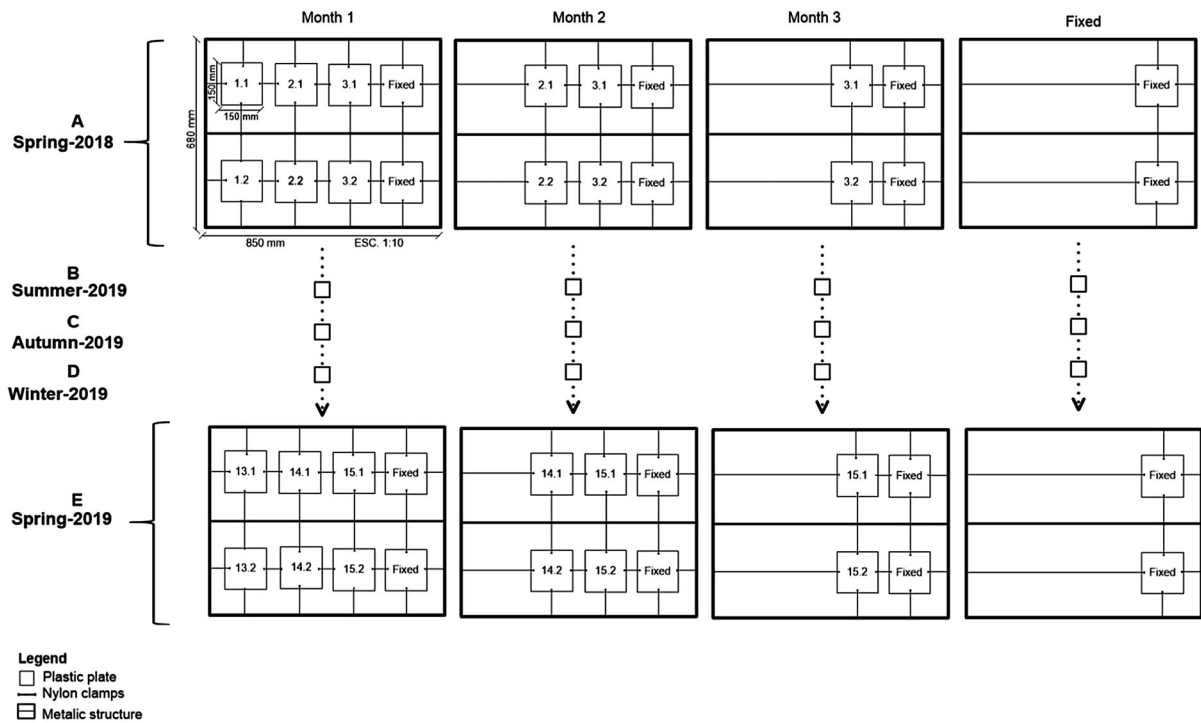


Fig. 1 Schematic representation of the supporting structure of the experimental settlement plates and the methodological procedures adopted: at the beginning of each season, the structure had eight experimental plates; at the end of each

month, two of them were removed for analysis of incrustations and two remaining plates (“fixed”) were removed after 15 months of monitoring; **A** Spring-2018, **B** Summer-2019, **C** Autumn-2019, **D** Winter-2019, and **E** Spring-2019

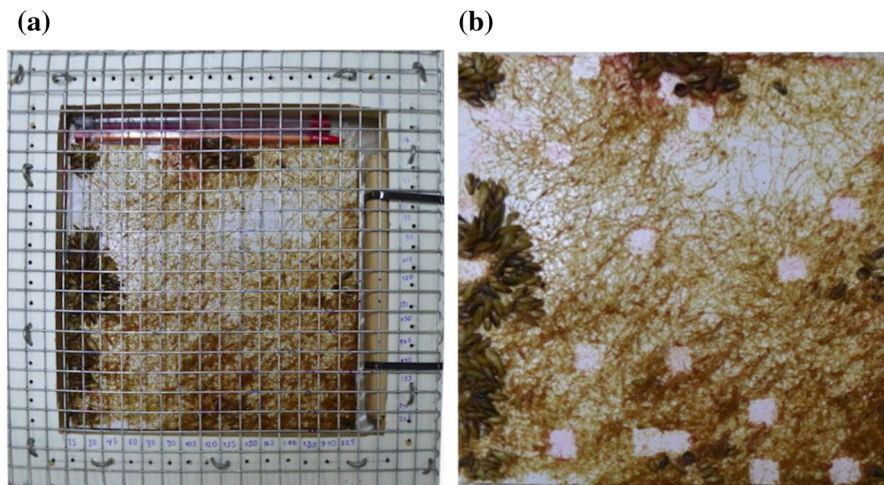


Fig. 2 Auxiliary structure for analyzing recruitment patterns of fouling and native invasive aquatic groups. **a** Grid with 225 squares (1 × 1 cm) showing the pattern of visualization of the

incrustation on the experimental settlement plate, below the grid; **b** experimental settlement plate after taking samples in the 20 random squares

selected cells was removed with the aid of a scalpel, stained with cane rose, and identified using a stereoscopic microscope (Pinto-Coelho, 2004).

Alternatively, if the fouling area covered 30% or less of the experimental settlement plates, they were scrapped, the material stained with a cane rose and

analyzed in a stereomicroscope. The relative density of each taxon on each plate was directly calculated for Amoebidae, Daphniidae, Calocalanidae, Chironomidae, Vaginicolidae, Centropxyidae, and Mytilidae. However, obtaining reliable counts for Cnidaria is made difficult by the frequent presence of fragmented individuals in the samples, mainly in the case of Cordylophoridae. Therefore, the analysis of Cordylophoridae and Hydridae was done qualitatively, considering the number of hydrorrhizae (filaments) or hydrae present in each sample. The recruitment patterns, as well as interactions between the groups found, were based on direct observations under a stereoscopic microscope. The observational data were supplemented with bibliographic information about the reproductive biology, behavior, and ecological relationships between individuals invading limnic environments, as well as between invasive and native species.

Data analysis

We used the Shannon–Wiener diversity index to characterize the diversity of organisms by sampling

period (Pielou, 1966). Normality and homogeneity of the variances of density, biomass, diversity, and temperature data were analyzed by the Levene and Cochran tests; the homoscedasticity was analyzed using the residuals of variance. The Kruskal–Wallis analysis was used to determine the significant differences between the variables tested, always at the 95% confidence level. Spearman's correlation coefficient was applied to assess the degree of association between dependent variables (coverage area, density, biomass, diversity) and water temperature (independent variable). The analyses were performed using the software Statistica 10.0 (StatSoft®).

Results

Environmental conditions

The highest average water temperatures in the reservoir were recorded in January 2019 and the lowest in August 2019. When temperatures are compared across seasons, they were similar ($P > 0.05$) in spring (2018

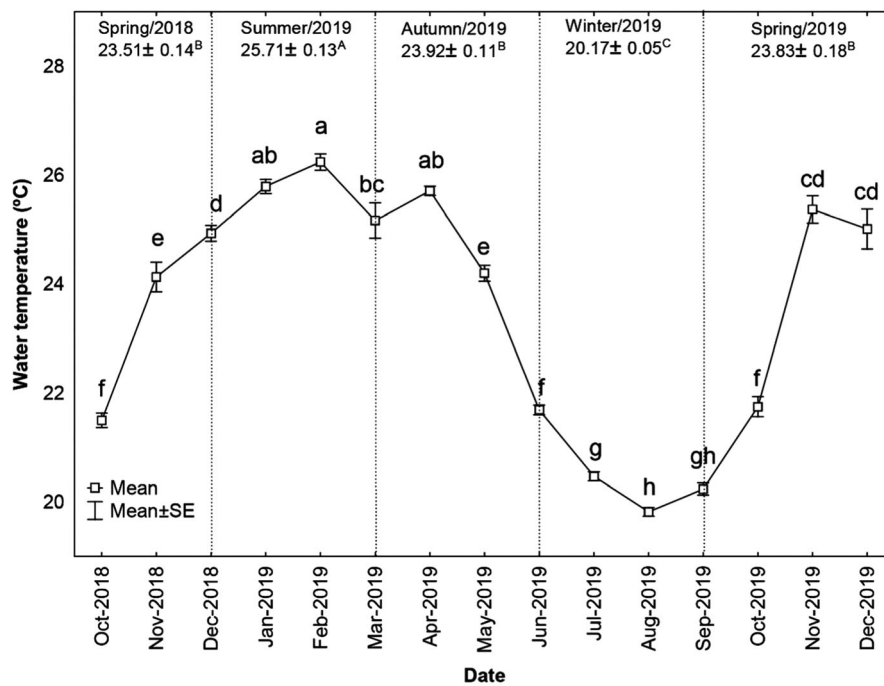


Fig. 3 Temporal variation (average values and standard error) of the water temperature in the reservoir of the HPP José Richa, from October 2018 to December 2019. Similar capital letters

indicate statistical significance between the seasons and the lower letters between the months of the year

and 2019) and in autumn (2019), but higher in summer and lower in winter (Fig. 3).

Identification of the recorded taxa

Macro and microinvertebrate organisms present in the analyzed samples were identified, and three of them (*Limnoperna fortunei*, *Cordylophora* sp. and *Hydra* sp.) are considered fouling invasive species belonging to the phyla Mollusca and Cnidaria (Table 1).

Recruitment pattern of identified taxa

The plates kept submerged for 15 consecutive months had, on average, 79.3% of their area occupied by colonizing organisms. The highest colonization rate was recorded in the spring of 2018 (56.0%) of the occupied area by colonizing organisms. In relation to the seasons, the highest occupations involved plates obtained during the spring of 2018, which showed 56.0% of their occupied area at the end of three months. The values quantified in these periods were significantly higher than those observed in other seasons (Fig. 4a). The average density of organisms present in the test experimental settlement plates was higher in the spring 2018 and the summer 2019 (Fig. 4b). The highest rates of seasonal diversity were recorded in the winter and autumn of 2019, with lower diversities observed on the test experimental settlement plates kept submerged for 15 months and in the spring of 2019 (Fig. 4c). The period considered most critical regarding the presence of fouling invasive organisms took place between late spring and early

summer. In the following seasons, the relative density of these organisms was significantly reduced. Particularly in the case of specimens (plates) kept submerged for 15 months, the density was close to the values found in early summer, with the difference that in this case, there were practically only one encrusted species (*L. fortunei*) (Fig. 4d).

The average density of organisms was 10.4 individuals/cm² during the spring 2018, 6.3 in the summer 2019, 0.3 in the spring 2019, 0.2 in the autumn 2019, and 0.1 in the winter 2019. *Limnoperna fortunei* was the predominant invasive organism throughout the analyzed period. The average density of *L. fortunei* was higher in the spring 2018 and the summer 2019. This species also completely dominated the experimental settlement plates kept submerged for 15 consecutive months (Fig. 5). The largest biomass of *L. fortunei* was recorded in the experimental settlement plates kept submerged for 15 months in the reservoir. The highest seasonal biomasses were recorded in winter (Table 2). There was a significant ($P < 0.05$) but low correlation ($r = -0.39$) between the biomass of *L. fortunei* and water temperature.

Ecological interactions between the families of the identified organisms

The ecological interactions between the main groups identified in the experimental settlement plates were classified as positive and direct (mutualism), positive and indirect (commensalism), negative and direct (competition), negative and indirect (epibiosis, cannibalism, and predation) (Fig. 6). Mutualism was

Table 1 Taxonomic classification of families identified in the experimental settlement plates in the reservoir of the HPP Gov. José Richa

Phylum	Class	Order	Family	Species	Category
Protozoa	Lobosa	Amoebida	Amoebidae	NI*	Native
Arthropoda	Branchiopoda	Cladocera	Daphniidae	NI	Native
Arthropoda	Maxillopoda	Calanoida	Calocalanidae	NI	Native
Arthropoda	Insecta	Diptera	Chironomidae	NI	Native
Ciliophora	Oligohymenophorea	Sessilida	Vaginicolidae	NI	Native
Amoebozoa	Tubulinea	Tubulunida	Centropyxidae	NI	Native
Mollusca	Bivalvia	Mytilida	Mytilidae	<i>Limnoperna fortunei</i>	Invasive
Cnidaria	Hydrozoa	Anthoathecata	Cordylophoridae	<i>Cordylophora</i> sp.	Invasive
Cnidaria	Hydrozoa	Anthoathecata	Hydridae	<i>Hydra</i> sp.	Invasive

*NI - Taxa that could not be identified at the species level

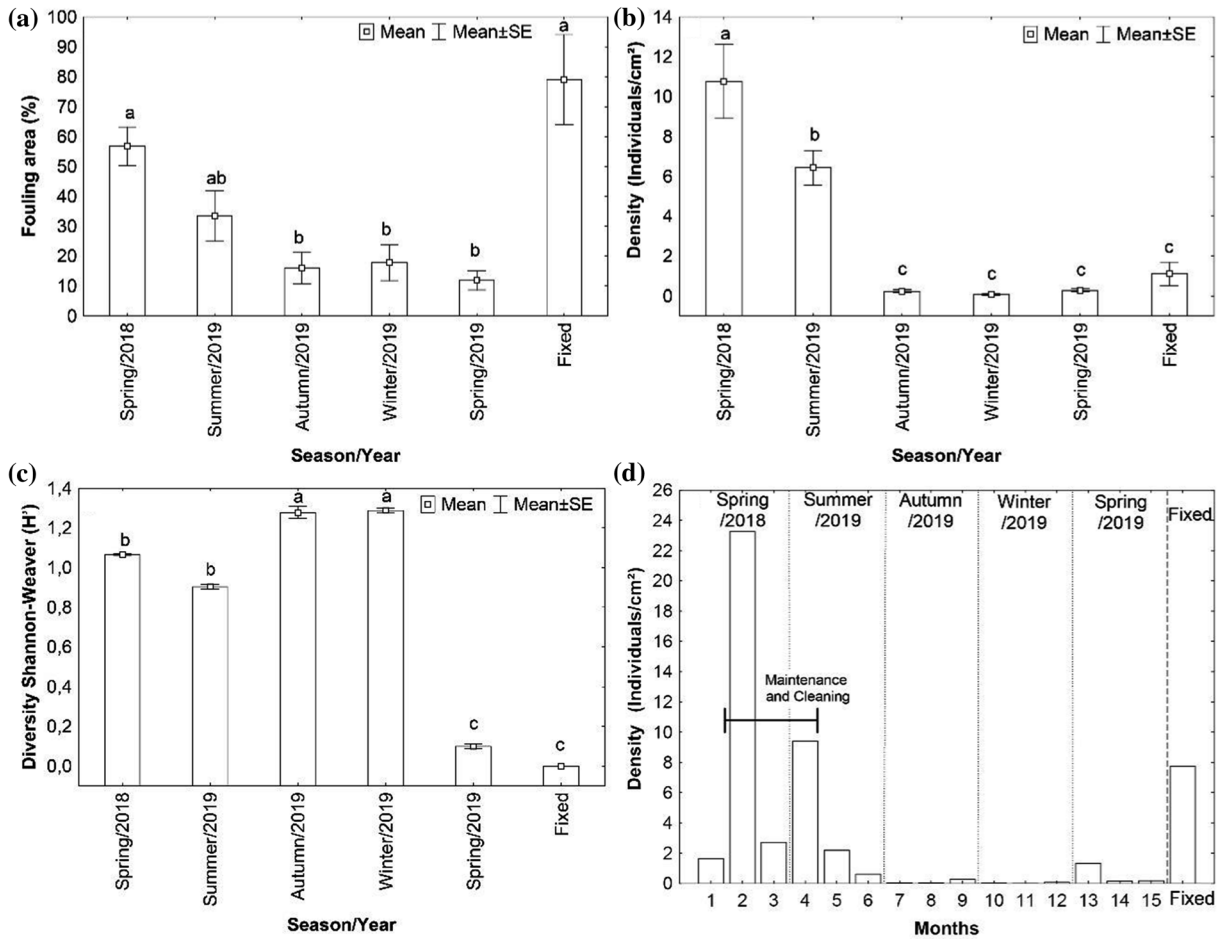


Fig. 4 **a** The area occupied by invasive and native organisms. **b** Seasonal variation in the total density of identified organisms. **c** Seasonal variation of the Shannon–Wiener diversity index. **d** Monthly density and the recommended period for the

maintenance and cleaning of the hydraulic installations (Month: 2- November; 3-December; and 4- January) of the Gobernador José Richa hydroelectric power plant

observed between adult and juvenile individuals of the family Mytilidae, as well as between individuals of the families Cordylophoridae and Mytilidae. Commensalism occurred between Mytilidae and insect larvae of the Chironomidae family. Competition for food and space was observed among fouling invaders (Cordylophoridae, Mytilidae, and Hydridae), especially when the space available for fouling on the substrate was more limited. Competition was observed between invasive and native organisms (Daphniidae, Calocalanidae, Chironomidae, Vaginicolidae, and Centropyxidae). Cannibalism was identified only in adult individuals of Mytilidae, which can feed on larvae of the same species. Epibiosis was characterized by the use of organisms from certain groups, such as

basibionts. For example, individuals from Hydridae were used as substrate by Cordylophoridae, Mytilidae, and Centropyxidae. Cordylophoridae, on the other hand, were recorded hosting Hydridae, Centropyxidae, and Mytilidae. Among the organisms involved in epibiosis, the relationship between Cordylophoridae and Mytilidae was the most evident and recorded on several occasions. Predation was observed, with Hydridae preying on Chironomidae, Daphniidae, and Calocalanidae. Cordylophoridae can feed on larvae of Mytilidae, as observed in the study by Molina et al. (2015), as well as individuals from the family Daphniidae and Calocalanidae in the juvenile and adult phases. Mytilidae can prey on protozoa including Vaginicolidae and Centropyxidae, and

microcrustaceans in the juvenile stage like Daphniidae and Calocalanidae.

Discussion

In subtropical aquatic environments, higher temperatures tend to have positive effects on the rates of fouling and colonization of aquatic communities (Rodrigues et al., 2015; Lansac-Tôha et al., 2019), especially on the density of micro- and macro-invertebrates (Belz et al., 2010; Borges, 2014; Borges et al., 2017). Temperature interferes with reproductive cycles, the supply of recruits, and the trophic conditions of aquatic environments (McPherson et al., 1984; Patil & Anil, 2005; Cifuentes et al., 2010; Chen et al., 2019; Mieczan & Rudyk-Leuska, 2019). In the present study, water temperature did not positively influence the composition or abundance of most identified communities. Temperature correlated only with the biomass of *L. fortunei*, with a weak, negative correlation ($r = -0.39$). On the other hand, the highest rates of fouling and density of organisms in the experimental settlement plates were recorded during spring and summer. The density of organisms present on the plates installed during spring 2018 and spring 2019 did not follow the same pattern of fouling or maximum achieved densities. This seasonal and interannual variation underscores the high degree of complexity of fouling processes by invasive aquatic organisms, which has also been reported in other studies (Underwood & Anderson, 1994; Berntsson & Jonsson, 2003; Dehmordi et al., 2011; Fortunato et al., 2017). It is possible, therefore, to postulate that other environmental factors may have contributed to the results, even those that were not individually monitored here, but were observed in other studies. Among these factors, we can highlight the availability of nutrients and food, precipitation patterns, incidence of sunlight (Melo & Bott, 1997; Pamplin et al., 2006; Fernandez & Navarrete, 2015; Masi et al., 2016; Navarrete et al., 2019), or related to the physical characteristics of the reservoir, such as hydrodynamics or its depth (Albano & Obenat, 2019), which might have contributed to the results obtained.

Both the density and the area occupied in the experimental plates by invading organisms were to those related to native organisms. It is known that the behavior of invasive species is associated with their

Fig. 5 Schematic representation of the ecological succession observed in the experimental settlement plates in the reservoir of the Governador José Richa HPP, indicating the groups of dominant organisms throughout the seasons: Spring 2018; Summer 2019; Autumn 2019; Winter 2019; Spring 2019. The experimental settlement plates kept submerged for 15 months (“Fixed” treatments are those that will remain for the entire 15 months). The values presented graphically represent the average density of the organisms. **a** Cordylophoridae, **b** Hydridae, **c** Mytilidae, **d** Vaginicolidae, **e** Centropyxidae, **f** Chironomidae, **g** Calocalanidae, and **h** Daphniidae

biological characteristics (such as high growth rates, the existence of successful reproduction strategies, and ease of dispersal) and, mainly, reproductive (early sexual maturity and high fertility), which facilitate the bioinvasion process. Such characteristics are present in *Cordylophota* sp. (Folino-Rorem, 2000; Folino-Rorem et al., 2006; Nakano & Strayer, 2014), *Hydra* sp. (Hobmayer et al., 2012; Rodrigues et al., 2016; Deserti et al., 2017), and *L. fortunei* (Darrigran, 2002; Boltovskoy, 2015a, b; Boltovskoy & Correa, 2015; Boltovskoy et al., 2015a).

The settlement periods of the studied groups were well defined, mainly for *L. fortunei*, in all monitored seasons. This observation coincides with Damborenea & Penchaszadeh (2006), which concluded that *L. fortunei* could produce sperm and oocytes continuously throughout the year. Boltovskoy & Correa (2015) showed that in about 80% of the time series studied in South American reservoirs, researchers had identified a density of at least 10 larvae/m³ of golden mussel in the analyzed samples. In the present study, the highest occurrence of *L. fortunei* settlement occurred in spring 2018 and summer 2019 coinciding with the reproduction peaks already reported in several other studies (Magara et al., 2001; Canzi et al., 2005; Darrigran & Damborenea, 2006; Boltovskoy et al., 2009).

The biomass of *L. fortunei* was higher in those experimental settlement plates maintained throughout the study period. On these plates, the individuals present were predominantly adults. In adulthood, *L. fortunei* generally shows greater adaptation to ecological ranges and to stressful environmental factors (Liu et al., 2020). The smallest golden mussel biomass was observed in autumn 2019. This period was characterized by intermediate temperatures, which tend to limit the growth rates of individuals and interfere with the quality and availability of food (Boltovskoy et al.,

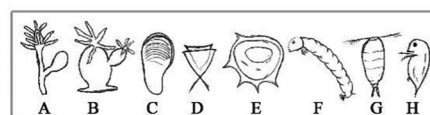
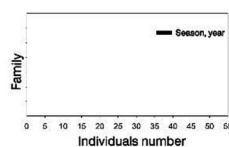
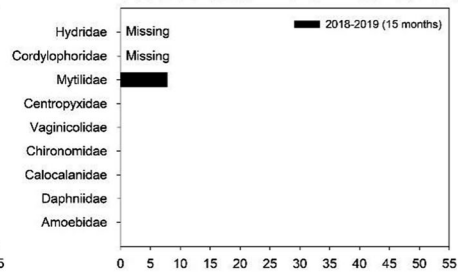
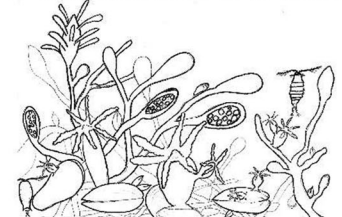
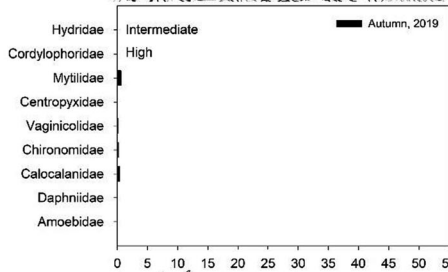
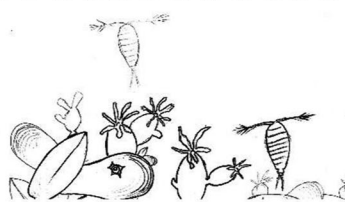
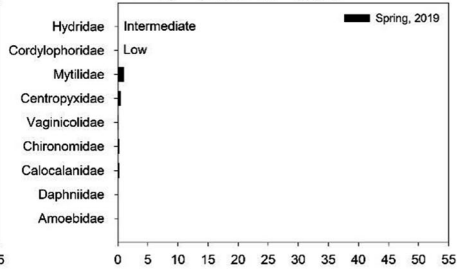
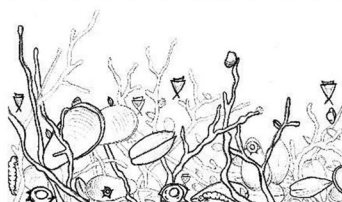
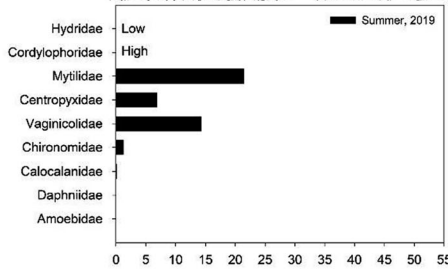
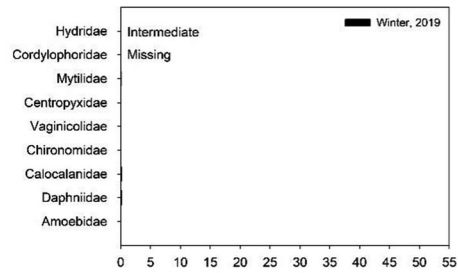
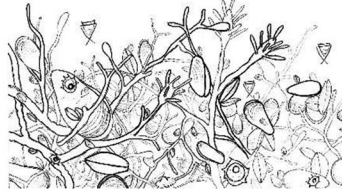
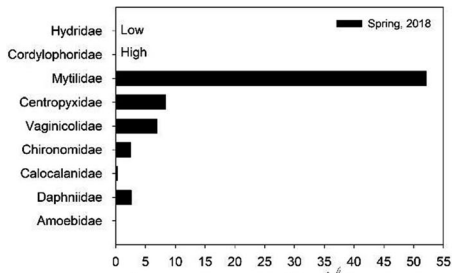


Table 2 Variation of the mean biomass of *Limnoperna fortunei* in the experimental settlement plates kept in the reservoir of Governador José Richa HPP. Different letters indicate statistical significance ($p < 0.05$) among seasons

Season/Year	The average number of specimens	Biomass (mg/cm ²) of <i>Limnoperna fortunei</i> Mean	SE
Spring, 2018	30	0.9 ^{abc}	0.13
Summer, 2019	78	0.5 ^c	0.04
Autumn, 2019	46	0.5 ^c	0.07
Winter, 2019	40	1.1 ^a	0.12
Spring, 2019	40	0.9 ^{abc}	0.14
2018–2019 (15 months)	175	293.8 ^a	87.50

2009; Nakano et al., 2011; Boltovskoy & Correa, 2015; Oliveira et al., 2015).

Over the course of ecological succession, the supporting organisms are the first to establish themselves (microorganisms, filamentous algae, diatoms, protozoa, and others) (Martín-Rodríguez et al., 2015; Silknetter et al., 2019), followed by macroinvertebrates (mollusks, insects, and cnidarians) (Abarzua & Jakubowski, 1995). *Cordylophora* sp. and *L. fortunei* were predominant in this process. The cnidarian has hydrorrhizae, a filamentous structure responsible for adhesion and incrustation on different substrates. These filaments end up serving as the preferred substrate for the settlement of *L. fortunei* larvae, which use them to adhere, protect themselves from local hydrodynamics, and develop during the first stages of life. Similar behavior has also been reported between *Cordylophora caspia* and *Dreissena polymorpha* (Pallas, 1771) in North America (Ackerman et al., 1994; Folino-Rorem et al., 2006; Pucherelli et al., 2016).

The trophic position and the capacity that each group has to occupy the substrate changes in the structuring of these communities over time. Some individuals end up dominating the substrates and making them more homogeneous (Ricciardi & MacIsaac, 2011; Simberloff & Ricciardi, 2020). In the present case, the highest levels of diversity occurred in a period of lower rates of incrustation of the experimental plates. As *L. fortunei* was the dominant species in almost all seasons, when it was less abundant, there was a greater opportunity for the colonization of the experimental plates by other organisms, which increased the diversity indexes. Spaccesi & Rodrigues (2012) also recorded lower

rates of diversity in substrates dominated by *L. fortunei* in the La Plata River. The authors also report that diversity was also lower in periods of low temperatures (autumn and winter).

Experimental studies carried out in South American reservoirs that assess ecological interactions, including fouling and native invasive groups, are still relatively limited (Nakano et al., 2011; Boltovskoy & Correa, 2015). In the present case, it was possible to identify interactions with positive effects (mutualism and commensalism) and indirect negative effects (competition, cannibalism, epibiosis, and predation). The substrate used for the settlement can be a limiting resource in certain environments and, when mutualistic interactions occurred, in the present case, they involved the sharing of substrates by *L. fortunei* and *Cordylophora* sp. Portella et al. (2009) evaluated biological fouling in the same reservoir of HPP José Richa and reported the presence of these two invaders in coexistence, that contribute to operational and economic impacts to the plant. According to Simberloff & Von Holle (1999) and Gallardo & Aldridge (2018), mutualistic relationships between freshwater invaders are poorly studied in the literature. However, understanding the role of each group involved in mutualism is of great importance, given that the effects of both species individually, in addition to their ecological interactions, often lead to severe negative economic impacts (Green et al., 2011; Rolla et al., 2019; Wegner et al., 2019). This mutualistic relationship can also contribute to establishing and magnify the spread of secondary invaders at different levels of the ecosystem (Ricciardi & Reisinger, 1994; Ricciardi, 2001, 2015; Green et al., 2011).

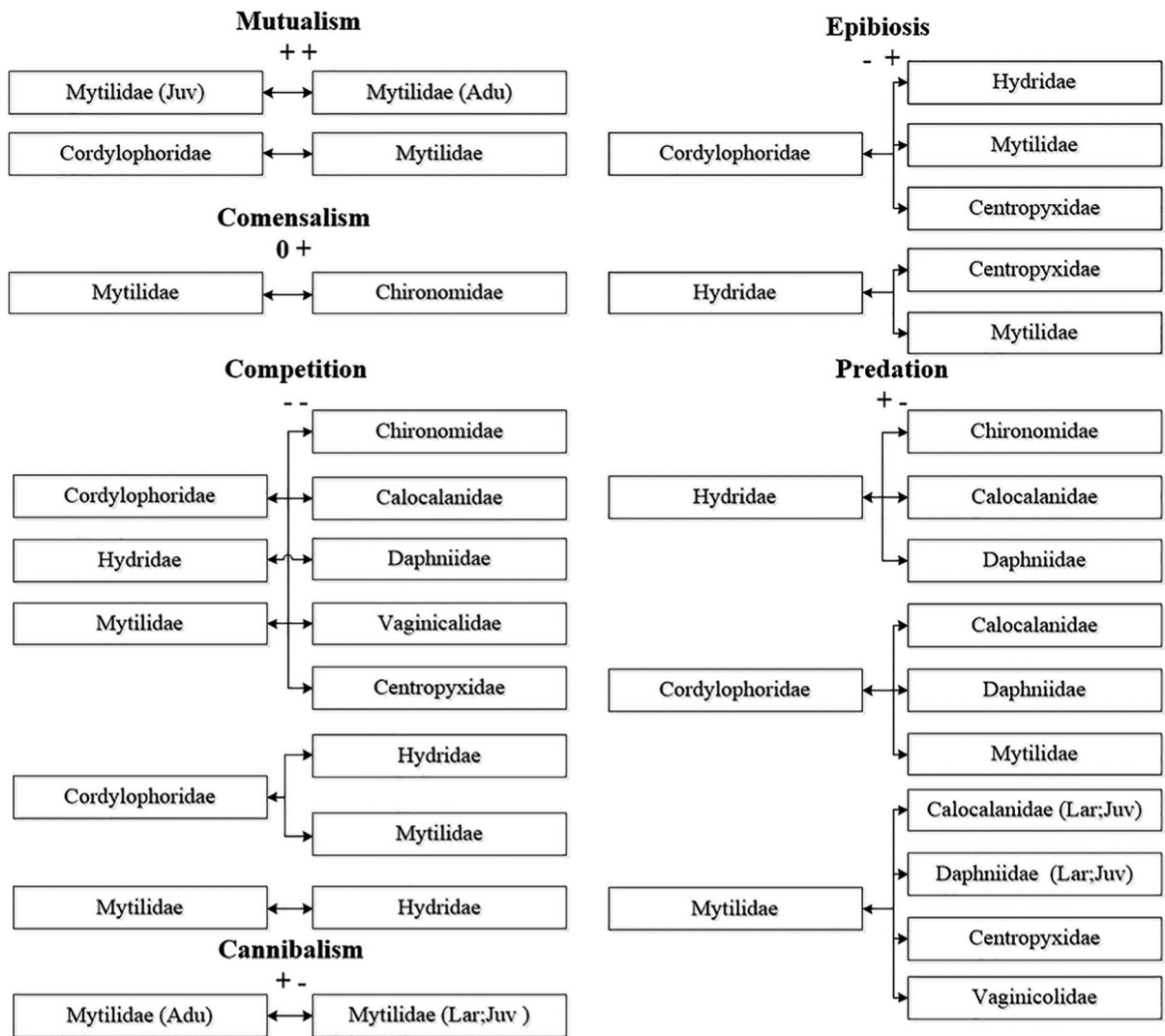


Fig. 6 Schematic representation of the ecological interactions recorded between the families of the fouling and native invader groups present in the experimental settlement plates positioned in the reservoir of the Governador José Richa HPP

Invaders also modify the habitats of native species and contribute to making these environments more homogeneous (Ricciardi & MacIsaac, 2011; Simberloff & Ricciardi, 2020). For example, *L. fortunei* may favor an increase in the number of macroinvertebrates in the Chironomidae family. This happens due to the accumulation of shells of the golden mussel clusters, which end up increasing the physical complexity of the substrate (Gutiérrez et al., 2003; Burlakova et al., 2012), as well as enriching these environments with the increase of stool bio-deposits and pseudofeces (Darrigran et al., 1998; Sardiña et al.,

2008). This enrichment causes changes in the density of benthic fauna, as well as leading to the biological imbalance of these environments (Karatayev et al., 2007; Darrigran & Damborenea, 2011). Burlakova et al. (2012) evaluated the composition of communities in locations with the presence and absence of two ecosystem engineers: *L. fortunei* and *D. polymorpha*. Where the species were present, the authors observed an increase in the richness and density of native macroinvertebrates (Burlakova et al., 2012). However, over time, native communities tend to become more homogeneous (Stewart et al., 1998; Ricciardi, 2001;

Ricciardi & MacIsaac, 2011; Simberloff & Ricciardi, 2020).

Among the potential interactions, competition was the most prevalent, as in other (Jackson; Kuebbing & Nuñez, 2015). This relationship can happen for several reasons, including competition for territories, substrates, and food (Jones et al., 2012). *Limnoperna fortunei* and *Cordylophora* sp. compete for substrate, especially when they are scarce. These organisms are fixed in common structures (both species can be found in the same type of substrate), which makes them close at different times or stages of life during the bioinvasion process. Competition for substrates is also reported between *C. caspia* and *D. polymorpha* (Folino-Rorem et al., 2006; Pucherelli et al., 2016).

Cannibalism was recorded among adults and larvae of *L. fortunei*. Adult mollusks can feed on particles ranging in size from 4 to 1000 µm in length, which includes their own larvae (Molina et al., 2015). In studies with *D. polymorpha*, cannibalism was responsible for mortality rates of up to 70% (MacIsaac et al., 1991). In epibiosis, *Cordylophora* sp. and *Hydra* sp. can serve as a substrate for other groups, such as *L. fortunei* and individuals of the family Centropxyidae. The relationship between *Cordylophora* sp. and *L. fortunei* is the most evident and involves several stages during the fouling process. However, it is more noticeable when *L. fortunei* is in its settlement phase, when they prefer to settle in places protected from turbulent currents (Cataldo & Boltovskoy, 1999; Sylvester et al., 2007).

Simultaneously with epibiosis, the facilitation relationship can occur between these same invaders (*L. fortunei* and *Cordylophora* sp.). The colonies of *Cordylophora* sp. have a three-dimensional structure, which initially favors the settlement of *L. fortunei*. After settling on *Cordylophora* sp. colonies, the mussels develop a preference for harder substrates and to continue their development they start to colonize these hard substrates originally occupied by *Cordylophora* sp., compromising the survival of the organisms of this species. Epibiosis between zebra mussels and other different groups of fouling invaders is well studied, especially the sponges *Ephydatia fluviatilis* (Linnaeus, 1759) (Ricciardi et al., 1996; Gaino, 2005; Ricciardi, 2005), *Eunapius fragilis* (Leidy, 1851) (Molloy et al., 1997; Ricciardi, 2005); the bryozoans *Pectinatella magnifica* (Leidy, 1851), *Plumatella fungosa* (Pallas, 1768) (Ricciardi &

Reiswig, 1994), *Lophopodella carteri* Hyatt, 1866 (Lauer et al., 1999; Cummings & Graf, 2010); and also with the cnidarian *Cordylophora caspia* (Pucherelli et al., 2016).

Predation has also been described for several groups in this study, for example, between *Cordylophora* sp. and larvae of *L. fortunei*. This cnidarian can also use juvenile and adult golden mussel shells as substrate (Olenin & Leppäkoski, 1999; Folino-Rorem et al., 2006). *Limnoperna fortunei* can prey on juvenile micro crustaceans, such as Daphniidae and Calocalanidae, as well as protozoa Vaginicolidae and Centropxyidae. Micro-crustaceans are among the favorite foods of the golden mussel, as they have a larger size and even greater biomass compared to phytoplankton (Molina et al., 2015).

It is recommended that maintenance and cleaning of hydraulic structures to remove biofouling should be carried out preferentially in late spring or early summer. This recommendation is based on abundance values, recruitment patterns, ecological succession, and the interactions observed between dominant groups in the José Richa HPP. This is the most critical period in terms of recruitment of young forms of fouling organisms and, in particular, *L. fortunei*, which is the species that most causes problems with the clogging of filters, grids, and pipes. In autumn and winter, biological fouling rates naturally fall by up to 80%, reducing the operational and economic risks of the machine shutdowns and cleaning. This conclusion is also supported by other studies, which reported decreased fouling levels and speed during colder seasons, which would be related to the heterothermia of invertebrates and their consequent dependence on temperature for regulating metabolic processes (Pörtner, 2002; Poloczanska et al., 2010); (Underwood & Anderson, 1994; Berntsson & Jonsson, 2003; Dehmordi et al., 2011; Fortunato et al., 2017). Therefore, by promoting the shutdown of machines and cleaning of hydraulic systems in the most critical period of this process, there would be less time for the establishment of colonies during the peak of spring and clean structures during the beginning of peak summer colonization, ensuring operating conditions.

Acknowledgements We thank the National Council for Scientific and Technological Development (CNPq) for granting funding to AO (Grant 381091/2014-7).

Author contributions Conceptualization: AB, RL, and AO; methodology and field collection: AB, AO, RL, and AH, and ON; software: AO and AB; writing—preparing the manuscript: AB, AO, RL, and AH; writing—review and editing: AB, AO, AH, RL, and MP; supervision: AO, MP, and TZ.

Funding This paper presents part of the results of the P&D project, code PD-06491-0383/2015, executed by the Federal University of Paraná and Aliança Prestadora de Serviços Ltda. and funded by COPEL Geração e Transmissão SA, under the Research and Technological Development Program of Electricity Sector, regulated by the National Electric Energy Agency (Aneel).

Data availability Raw data used in our analyses are available from the authors upon request.

Declarations

Conflict of interest The authors declare that they have no conflict of interest.

References

- Abarzua, S. & S. Jakubowski, 1995. Biotechnological investigation for the prevention of biofouling. I. Biological and biochemical principles for the prevention of biofouling. *Marine Ecology Progress Series* 123:301-312.
- Ackerman, J. D., B. Sim, S. J. Nichols & R. Claudi, 1994. A review of the early life history of zebra mussels (*Dreissena polymorpha*): comparisons with marine bivalves. *Canadian Journal of Zoology* 72(7):1169-1179.
- Albano, M. J. & S. M. Obenat, 2019. Fouling assemblages of native, non-indigenous and cryptogenic species on artificial structures, depths and temporal variation. *Journal of Sea Research* 144:1-15.
- Aneel, A. N. E. E., 2016. Plano de Dados Abertos, Informações Gerenciais – Tópico: Geração.
- Armour, A., J. Tsou & P., 1993. Wiancko, Zebra mussels: the industrial impact. In: Proceedings of the 3rd international zebra mussel conference, Toronto.
- Belz, C. E., G. Darrigran, N. Bonel & O. S. Mäder Netto, 2010. Density, Recruitment, and Shell Growth of *Limnoperna fortunei* (Mytilidae), an Invasive Mussel in Tropical South America. *Journal of Freshwater Ecology* 25(2):227-233.
- Berntsson, K. M. & P. R. Jonsson, 2003. Temporal and spatial patterns in recruitment and succession of a temperate marine fouling assemblage: a comparison of static panels and boat hulls during the boating season. *Biofouling* 19(3):187-195.
- Boltovskoy, D., 2015. *Limnoperna fortunei*: the ecology, distribution and control of a swiftly spreading invasive fouling mussel, vol 10. Springer.
- Boltovskoy, D., 2017. Traits and impacts of invasive species: Myths and evidences from the perspective of introduced freshwater mussels. *Aquatic Ecosystem Health & Management* 20(4):334-343.
- Boltovskoy, D. & N. Correa, 2015. Ecosystem impacts of the invasive bivalve *Limnoperna fortunei* (golden mussel) in South America. *Hydrobiologia* 746(1):81-95.
- Boltovskoy, D., N. Correa, D. Cataldo & F. Sylvester, 2006. Dispersion and Ecological Impact of the Invasive Freshwater Bivalve *Limnoperna fortunei* in the Río de la Plata Watershed and Beyond. Springer.
- Boltovskoy, D., F. Sylvester, A. Otaegui, V. Leites & D. H. Cataldo, 2009. Environmental modulation of reproductive activity of the invasive mussel *Limnoperna fortunei*: implications for antifouling strategies. *Austral Ecology* 34(7):719-730.
- Boltovskoy, D., B. Morton, N. Correa, D. Cataldo, C. Damborenea, P. E. Penchaszadeh & F. Sylvester, 2015a. Reproductive output and seasonality of *Limnoperna fortunei*. In *Limnoperna Fortunei*. Springer: 77–103.
- Boltovskoy, D., M. Xu & D. Nakano, 2015b. Impacts of *Limnoperna fortunei* on man-made structures and control strategies: general overview. In *Limnoperna fortunei* :375–393.
- Booy, O., L. Cornwell, D. Parrott, M. Sutton-Croft & F. Williams, 2017. Impact of biological invasions on infrastructure. In *Impact of Biological Invasions on Ecosystem Services*. Springer: 235–247.
- Borges, P. D., 2013. Aspectos do ciclo de vida da espécie invasora *Cordylophora caspia* (Cnidaria) no reservatório da Usina Hidrelétrica Governador José Richa, Rio Iguaçu, Paraná.
- Borges, P. D., 2014. *Limnoperna fortunei* (Bivalvia: Mytilidae) e o setor elétrico brasileiro: distribuição, impactos, estudo de caso da dispersão no Rio Iguaçu e teste de protocolo de uso de larvas na caracterização do perfil genético de populações.
- Borges, P. D., S. Ludwig & W. A. Boeger, 2017. Testing hypotheses on the origin and dispersion of *Limnoperna fortunei* (Bivalvia, Mytilidae) in the Iguassu River (Paraná, Brazil): molecular markers in larvae and adults. *Limnology* 18(1):31-39.
- Burlakova, L. E., A. Y. Karatayev & V. A. Karatayev, 2012. Invasive mussels induce community changes by increasing habitat complexity. *Hydrobiologia* 685(1):121-134.
- Callow, M. E., 1993. A review of fouling in freshwaters. *Biofouling* 7(4):313-327.
- Canzi, C., L. Bortoluzzi & D. Rodriguez Fernandez, 2005. Ocorrência e situação atual do mexilhão dourado (*Limnoperna fortunei*) no reservatório da central hidrelétrica de Itaipu. I Simpósio Brasileiro sobre Espécies Exóticas Invasoras, Brasília (Brazil).
- Cataldo, R., 2001. 'Musseling' in on the Ninth District economy. *Fedgazette* 13(1):15-15.
- Cataldo, D. & D. Boltovskoy, 1999. Population dynamics of *Corbicula fluminea* (Bivalvia) in the Paraná River Delta. *Hydrobiologia* 153–163.
- Cataldo, D., D. Boltovskoy & M. Pose, 2003. Toxicity of chlorine and three nonoxidizing molluscicides to the pest mussel *Limnoperna fortunei*. *Journal-American Water Works Association* 95(1):66-78.
- Characklis, W. G., 1981. Bioengineering report: fouling biofilm development: a process analysis. *Biotechnology and Bioengineering* 23(9):1923-1960.

- Chen, X., X. Xiong, X. Jiang, H. Shi & C. Wu, 2019. Sinking of floating plastic debris caused by biofilm development in a freshwater lake. *Chemosphere* 222:856–864.
- Cifuentes, M., I. Krueger, C. P. Dumont, M. Lenz & M. Thiel, 2010. Does primary colonization or community structure determine the succession of fouling communities? *Journal of Experimental Marine Biology and Ecology* 395(1-2):10–20.
- Coetser, S. & T. E. Cloete, 2005. Biofouling and biocorrosion in industrial water systems. *Critical reviews in microbiology* 31(4):213–232.
- Cummings, K. S. & D. L. Graf, 2010. Mollusca: bivalvia. In *Ecology and Classification of North American Freshwater Invertebrates*. Elsevier: 309–384.
- Damborenea, C. & P. Penchaszadeh, 2006. Biología reproductiva de *Limnoperna fortunei*. Bio-invasión del mejillón dorado en el continente americano:71–84.
- Darrigran, G. & I. E. De Drago, 2000. Invasion of the exotic freshwater mussel *Limnoperna fortunei* (Dunker, 1857)(Bivalvia: Mytilidae) in South America. *The Nautilus* 114:69–73.
- Darrigran, G., 2002. Potential impact of filter-feeding invaders on temperate inland freshwater environments. *Biological Invasions* 4(1-2):145–156.
- Darrigran, G. & C. Damborenea, 2011. Ecosystem engineering impact of *Limnoperna fortunei* in South America. *Zoological science* 28(1):1–8.
- Darrigran, G., S. M. Martin, B. Gullo & L. Armendariz, 1998. Macroinvertebrates associated with *Limnoperna fortunei* (Dunker, 1857)(Bivalvia, Mytilidae) in Rio de la Plata, Argentina. *Hydrobiologia* 367(1-3):223–230.
- Darrigran, G. & C. Damborenea, 2006. Bio-invasión del mejillón dorado en el continente americano. Editorial de la Universidad Nacional de La Plata (EDULP).
- David, P., E. Thebault, O. Anneville, P.-F. Duyck, E. Chapuis & N. Loeuille, 2017. Impacts of invasive species on food webs: a review of empirical data. In *Advances in Ecological Research*, Vol 56. Elsevier: 1–60.
- Dehmordi, L. M., L. Karami, N. Safarpour & B. Alesadi, 2011. Taxonomic identification and distribution of biofouling organisms in Deilam port in Iran. *Journal of Ecology and the Natural Environment* 3(14):441–445.
- Deserti, M. I., K. S. Esquius, A. H. Escalante & F. H. Acuña, 2017. Trophic ecology and diet of *Hydra vulgaris* (Cnidaria; Hydrozoa). *Animal Biology* 67(3-4):287–300.
- Emery-Butcher, H. E., S. J. Beatty & B. J. Robson, 2020. The impacts of invasive ecosystem engineers in freshwaters: A review. *Freshwater Biology*.
- Farhat, N., L. Javier, M. Van Loosdrecht, J. Kruithof & J. S. Vrouwenvelder, 2019. Role of feed water biodegradable substrate concentration on biofouling: Biofilm characteristics, membrane performance and cleanability. *Water research* 150:1–11.
- Fernandez, M. O. & S. A. Navarrete, 2015. A comparison of temporal turnover of species from benthic cnidarian assemblages in tropical and subtropical harbours. *Taylor & Francis*.
- Flemming, H.-C., 2002. Biofouling in water systems—cases, causes and countermeasures. *Applied microbiology and biotechnology* 59(6):629–640.
- Flemming, H. C., 2011. *Microbial biofouling: unsolved problems, insufficient approaches, and possible solutions*. Springer.
- Flemming, H.-C. & E. Cloete, 2010. Environmental impact of controlling biofouling and biocorrosion in cooling water systems. In *Operational and Environmental Consequences of Large Industrial Cooling Water Systems* :365–380.
- Folino-Rorem, N. C., 2000. The freshwater expansion and classification of the colonial hydroid *Cordylophora* (Phylum Cnidaria, Class Hydrozoa). In Pederson, J. (ed.) *Marine Bioinvasions: Proceedings of the First National Conference*. Massachusetts Institute of Technology Sea Grant College Program, Cambridge MA.
- Folino-Rorem, N. C., J. Stoeckel, E. Thorn & L. Page, 2006. Effects of artificial filamentous substrate on zebra mussel (*Dreissena polymorpha*) settlement. *Biological Invasions* 8(1):89–96.
- Fortunato, L., S. Jeong & T. Leiknes, 2017. Time-resolved monitoring of biofouling development on a flat sheet membrane using optical coherence tomography. *Scientific reports* 7(1):1–9.
- Frota, M. N., E. M. Ticona, A. V. Neves, R. P. Marques, S. L. Braga & G. Valente, 2014. On-line cleaning technique for mitigation of biofouling in heat exchangers: A case study of a hydroelectric power plant in Brazil. *Experimental thermal and fluid science* 53:197–206.
- Gaino, T. L. E., 2005. Competition between the freshwater sponge *Ephydatia fluviatilis* and the zebra mussel *Dreissena polymorpha* in Lake Trasimeno (central Italy). *Italian Journal of Zoology* 72(1):27–32.
- Gallardo, B. & D. C. Aldridge, 2018. Inter-basin water transfers and the expansion of aquatic invasive species. Elsevier.
- Green, P. T., D. J. O'Dowd, K. L. Abbott, M. Jeffery, K. Retallick & R. Mac Nally, 2011. Invasional meltdown: invader–invader mutualism facilitates a secondary invasion. *Ecology* 92(9):1758–1768.
- Grohmann, P. A., 2008a. Bioencrustation in the turbine cooling system at the funil hydroelectric power plant, Itatiaia, Rio de Janeiro, Brazil. *Naturalia*.
- Grohmann, P. A., 2008b. Bioencrustation caused by a hydroid species in the turbine cooling system at the Funil hydroelectric power plant. *Naturalia* 31:16–21.
- Grohmann, P. A., 2008c. Bioencrustation caused by a hydroid species in the turbine cooling system at the funil hydroelectric power plant, Itatiaia Naturalia, Rio Claro 31:16–21.
- Gutiérrez, J. L., C. G. Jones, D. L. Strayer & O. O. Iribarne, 2003. Mollusks as ecosystem engineers: the role of shell production in aquatic habitats. *Oikos* 101(1):79–90.
- Hacker, S. a. D. & S. D. Gaines, 1997. Some implications of direct positive interactions for community species diversity. *Wiley Online Library*.
- Hobmayer, B., M. Jenewein, D. Eder, M.-K. Eder, S. Glasauer, S. Gufler, M. Hartl & W. Salvenmoser, 2012. Stemness in *Hydra*—a current perspective. *International Journal of Developmental Biology* 56(6-7-8):509–517.
- Jackson, M. C., 2015. Interactions among multiple invasive animals. *Ecology* 96(8):2035–2041.
- Jernelöv, A., 2017. The long-term fate of invasive species. Aliens forever or integrated immigrants with time.

- Jones, E. I., J. L. Bronstein & R. Ferrière, 2012. The fundamental role of competition in the ecology and evolution of mutualisms. Wiley Online Library.
- Karatayev, A. Y., L. E. Burlakova & D. K. Padilla, 2005. Contrasting distribution and impacts of two freshwater exotic suspension feeders, *Dreissena polymorpha* and *Corbicula fluminea*. In The comparative roles of suspension-feeders in ecosystems. Springer: 239–262.
- Karatayev, A. Y., D. K. Padilla, D. Minchin, D. Boltovskoy & L. E. Burlakova, 2007. Changes in global economies and trade: the potential spread of exotic freshwater bivalves. Springer.
- Kéfi, S., M. Holmgren & M. Scheffer, 2016. When can positive interactions cause alternative stable states in ecosystems? *Functional Ecology* 30(1):88–97.
- Kuebbing, S. E. & M. A. Nuñez, 2015. Negative, neutral, and positive interactions among nonnative plants: patterns, processes, and management implications. *Global Change Biology* 21(2):926–934.
- Lansac-Tôha, F. M., J. Heino, B. A. Quirino, G. A. Moresco, O. Peláez, B. R. Meira, L. C. Rodrigues, S. Jati, F. A. Lansac-Tôha & L. F. M. Velho, 2019. Differently dispersing organism groups show contrasting beta diversity patterns in a dammed subtropical river basin. *Science of The Total Environment* 691:1271–1281.
- Latombe, G., P. Pyšek, J. M. Jeschke, T. M. Blackburn, S. Bacher, C. Capinha, M. J. Costello, M. Fernández, R. D. Gregory & D. Hobern, 2017. A vision for global monitoring of biological invasions. *Biological Conservation* 213:295–308.
- Lauer, T. E., D. K. Barnes, A. Ricciardi & A. Spacie, 1999. Evidence of recruitment inhibition of zebra mussels (*Dreissena polymorpha*) by a freshwater bryozoan (*Lophopodella carteri*). *Journal of the North American Benthological Society* 18(3):406–413.
- Liu, W., M. Xu, J. Zhang & T. Zhang, 2020. Survival and attachment of biofouling freshwater mussel (*Limnoperna fortunei*) to environmental conditions: potential implications in its invasion, infection and biofouling control. *Limnology*:1–11.
- Ludwig, S., M. K. Tschá, R. Patella, A. J. Oliveira & W. A. Boeger, 2014. Looking for a needle in a haystack: molecular detection of larvae of invasive *Corbicula* clams. *Management of biological Invasions* 5(2):143.
- MacIsaac, H. J., 1996. Potential abiotic and biotic impacts of zebra mussels on the inland waters of North America. *American zoologist* 36(3):287–299.
- MacIsaac, H. J., W. G. Sprules & J. Leach, 1991. Ingestion of small-bodied zooplankton by zebra mussels (*Dreissena polymorpha*): can cannibalism on larvae influence population dynamics? *Canadian journal of fisheries and aquatic sciences* 48(11):2051–2060.
- Magara, Y., Y. Matsui, Y. Goto & A. Yuasa, 2001. Invasion of the non-indigenous nuisance mussel, *Limnoperna fortunei*, into water supply facilities in Japan. *Journal of Water Supply: Research and Technology – AQUA* 50(3):113–124.
- Mansur, M. C. D., C. P. Santos, D. Pereira, P. E. A. Bergonci & C. T. Callil, 2016. Moluscos límnicos - bivalves.
- Martin, R. A. M., D. V. Obrecht, G. A. Stoner, R. A. O’Hearn, S. W. Lanning & W. R. Mabee, 2016. First record of occurrence of the invasive hydroid *Cordylophora caspia* (Cnidaria: Hydrozoa) in Missouri. *The Southwestern Naturalist* 61(3):260–264.
- Martín-Rodríguez, A. J., J. M. Babarro, F. Lahoz, M. Sansón, V. S. Martín, M. Norte & J. J. Fernández, 2015. From broad-spectrum biocides to quorum sensing disruptors and mussel repellents: Antifouling profile of alkyl triphenylphosphonium salts. *PLoS One* 10(4).
- Masi, B. P., I. Zalmon & R. Coutinho, 2016. Effects of structural factors on upwelling fouling community, Southeast Brazil. *Brazilian Journal of Oceanography* 64(4):387–400.
- McPherson, B. F., W. H. Sonntag & M. Sabanskas, 1984. Fouling community of the Loxahatchee River estuary, Florida, 1980–81. *Estuaries* 7(2):149–157.
- Melo, L. & T. Bott, 1997. Biofouling in water systems. *Experimental thermal and fluid science* 14(4):375–381.
- Mieczan, T. & N. Rudyk-Leuska, 2019. Seasonal dynamics of the epibiont food web on *Unio tumidus* (Philippson, 1788) in a eutrophic reservoir. *European journal of protistology* 69:138–150.
- Molina, F. R., S. B. J. de Paggi & J. C. Paggi, 2015. Impacts of *Limnoperna fortunei* on Zooplankton. *Limnoperna Fortunei*. Springer: 177–190.
- Molloy, D. P., A. Y. Karatayev, L. E. Burlakova, D. P. Kurandina & F. Laruelle, 1997. Natural enemies of zebra mussels: predators, parasites, and ecological competitors. Taylor & Francis.
- Morton, B., 1977. Freshwater fouling bivalves. In: Symposium, held at Texas Christian University, Fort Worth, Texas, Texas Christian University.
- Nakano, D. & D. L. Strayer, 2014. Biofouling animals in fresh water: biology, impacts, and ecosystem engineering. *Frontiers in Ecology and the Environment* 12(3):167–175.
- Nakano, D., T. Kobayashi, N. Endo & I. Sakaguchi, 2011. Growth rate and settlement of *Limnoperna fortunei* in a temperate reservoir. *Journal of Molluscan Studies* 77(2):142–148.
- Navarrete, S. A., M. Parragué, N. Osiadacz, F. Rojas, J. Bonicelli, M. Fernández, C. Arboleda-Baena, A. Perez-Matus & R. Finke, 2019. Abundance, composition and succession of sessile subtidal assemblages in high wave-energy environments of Central Chile: Temporal and depth variation. *Journal of experimental marine biology and ecology* 512:51–62.
- Nelson, N. M., 2019. Enumeration of potential economic costs of dreissenid mussel infestation in montana.
- Olenin, S. & E. Leppäkoski, 1999. Non-native animals in the Baltic Sea: alteration of benthic habitats in coastal inlets and lagoons. *Hydrobiologia* 393:233–243.
- Oliveira, M. D., M. C. S. Campos, E. M. Paolucci, M. C. D. Mansur & S. K. Hamilton, 2015. Colonization and spread of *Limnoperna fortunei* in South America. In *Limnoperna Fortunei*. Springer: 333–355.
- Oreska, M. P. & D. C. Aldridge, 2011. Estimating the financial costs of freshwater invasive species in Great Britain: a standardized approach to invasive species costing. *Biological Invasions* 13(2):305–319.
- Pamplin, P., T. Almeida & O. Rocha, 2006. Composition and distribution of benthic macroinvertebrates in Americana Reservoir(SP, Brazil). *Acta Limnologica Brasiliensia* 18(2):121–132.

- Papadopoulos, F., M. Spinelli, S. Valente, L. Foroni, C. Orrico, F. Alviano & G. Pasquini, 2007. Common tasks in microscopic and ultrastructural image analysis using ImageJ. *Ultrastructural pathology* 31(6):401-407.
- Patil, J. S. & A. C. Anil, 2005. Biofilm diatom community structure: influence of temporal and substratum variability. *Biofouling* 21(3-4):189-206.
- Pielou, E. C., 1966. Shannon's formula as a measure of specific diversity: its use and misuse. *The American Naturalist* 100(914):463-465.
- Pimentel, D., S. McNair, J. Janecka, J. Wightman, C. Simmonds, C. O'Connell, E. Wong, L. Russel, J. Zern & T. Aquino, 2001. Economic and environmental threats of alien plant, animal, and microbe invasions. *Agriculture, Ecosystems & Environment* 84(1):1-20.
- Pinto-Coelho, R. M., 2004. Métodos de coleta, preservação, contagem e determinação de biomassa em zooplâncton de águas continentais. In Bicudo, C. E. (ed.) *Amostragem em limnologia*: 149–167.
- Poloczanska, E., A. B.-. Biofouling, U. 2010 & A. Butler, 2010. Biofouling and climate change. books.google.com.
- Portella, K. F., A. Joukoski, A. S. d. Silva, N. M. Brassac & C. E. Belz, 2009. Biofouling and chemical biodeterioration in hydroelectric power plant portland cement mortar. *Quimica Nova* 32(4):1047-1051.
- Pörtner, H.-O., 2002. Climate variations and the physiological basis of temperature dependent biogeography: systemic to molecular hierarchy of thermal tolerance in animals. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology* 132(4):739-761.
- Pu, H., G.-l. Ding, X.-k. Ma, H.-t. Hu & Y.-f. Gao, 2009. Effects of biofouling on air-side heat transfer and pressure drop for finned tube heat exchangers. *International journal of refrigeration* 32(5):1032-1040.
- Pucherelli, S. F., J. Keele, Y. J. Passamaneck, J. R. Beaver & T. R. Renicker, 2016. Range expansion of the invasive hydroid, *Cordylophora caspia* (Pallas, 1771), in Colorado River reservoirs. *BioInvasions Records* 5(3):133-137.
- Pucherelli, S. F., R. Claudi & T. Prescott, 2018. Control of biofouling in hydropower cooling systems using HOD ultraviolet light. *Management of Biological Invasions* 9(4):451.
- Rajagopal, S. & G. van der Velde, 2012. Invasive species: implications for industrial cooling water systems. In *Operational and Environmental Consequences of Large Industrial Cooling Water Systems*. Springer, Boston: 127–162.
- Ricciardi, A., 2005. Facilitation and Synergistic Interactions Between Introduced Aquatic Species. *Scientific Committee on Problems of the Environment International Council of Scientific Unions*, Vol. 63: 162.
- Ricciardi, A., 2001. Facilitative interactions among aquatic invaders: is an "invasional meltdown" occurring in the Great Lakes? *Canadian journal of fisheries and aquatic sciences* 58(12):2513-2525.
- Ricciardi, A., 2015. Ecology of Invasive Alien Invertebrates Thorp and Covich's Freshwater Invertebrates (Fourth Edition). Elsevier: 83–91.
- Ricciardi, A. & H. J. MacIsaac, 2000. Recent mass invasion of the North American Great Lakes by Ponto-Caspian species. *Trends in Ecology & Evolution* 15(2):62-65.
- Ricciardi, A. & H. J. MacIsaac, 2008. The book that began invasion ecology. *Nature* 452(7183):34-34.
- Ricciardi, A. & H. J. MacIsaac, 2011. Impacts of biological invasions on freshwater ecosystems. Fifty years of invasion ecology: the legacy of Charles Elton 1:211-224.
- Ricciardi, A. & H. M. Reisinger, 1994. Taxonomy, distribution, and ecology of the freshwater bryozoans (Ectoprocta) of eastern Canada. *Canadian Journal of Zoology* 72(2):339-359.
- Ricciardi, A., F. G. Whoriskey & J. Rasmussen, 1996. Impact of the invasion on native unionid bivalves in the upper St. Lawrence River. *Canadian Journal of Fisheries and Aquatic Sciences* 53:1434-1444. <https://doi.org/10.1139/cjfas-53-6-1434>.
- Rodrigues, L. C., N. R. Simões, V. M. Bovo-Scomparin, S. Jati, N. F. Santana, M. C. Roberto & S. Train, 2015. Phytoplankton alpha diversity as an indicator of environmental changes in a neotropical floodplain. *Ecological Indicators* 48:334-341.
- Rodrigues, M., T. Ostermann, L. Kremeser, H. Lindner, C. Beisel, E. Berezikov, B. Hobmayer & P. Ladurner, 2016. Profiling of adhesive-related genes in the freshwater cnidarian *Hydra magnipapillata* by transcriptomics and proteomics. *Biofouling* 32(9):1115-1129.
- Rodriguez, L. F., 2006. Can invasive species facilitate native species? Evidence of how, when, and why these impacts occur. Springer.
- Rolla, M., S. Consuegra, E. Carrington, D. Hall & C. G. de Leaniz, 2019. Experimental evidence of invasion facilitation in the zebra mussel-killer shrimp system. [bioRxiv:626432](https://doi.org/10.1101/2019.06.26.266432).
- Sardiña, P., D. Cataldo & D. Boltovskoy, 2008. The effects of the invasive mussel, *Limnoperna fortunei*, on associated fauna in South American freshwaters: importance of physical structure and food supply. [ingentaconnect.com](https://doi.org/10.1007/s10841-008-9264-3).
- Schneider, C. A., W. S. Rasband & K. W. Eliceiri, 2012. NIH Image to ImageJ: 25 years of image analysis. *Nature methods* 9(7):671.
- Silknetter, S., R. P. Creed, B. L. Brown, E. A. Frimpong, J. Skelton & B. K. Peoples, 2019. Positive biotic interactions in freshwaters: A review and research directive. *Freshwater Biology* doi:<https://doi.org/10.1111/fwb.13476>.
- Simberloff, D., 2006. Invasional meltdown 6 years later: important phenomenon, unfortunate metaphor, or both? *Ecology Letters* 9(8):912-919.
- Simberloff, D. & J. R. Vitale, 2014. A call for an end to calls for the end of invasion biology. *Oikos* 123(4):408-413.
- Simberloff, D. & B. Von Holle, 1999. Positive interactions of nonindigenous species: invasional meltdown? *Biological Invasions* 1(1):21-32.
- Simberloff, D. & A. Ricciardi, 2020. In Chap. 3, Elton invoked the notion of a homogenized world owing. In *The Ecology of Invasions by Animals and Plants*: 53.
- Singh, M. P., P. Singh, H.-B. Li, Q.-Q. Song & R. K. Singh, 2020. Microbial biofilms: Development, structure, and their social assemblage for beneficial applications. In *New and Future Developments in Microbial Biotechnology and Bioengineering: Microbial Biofilms*. Elsevier: 125–138.
- Spaccesi, F. & C. A. Rodrigues, 2012. Benthic communities on hard substrates covered by *Limnoperna fortunei* Dunker

- (Bivalvia, Mytilidae) at an estuarine beach (Río de la Plata, Argentina). *Journal of Limnology* 71.
- Stewart, T. W., J. G. Miner & R. L. Lowe, 1998. Quantifying mechanisms for zebra mussel effects on benthic macroinvertebrates: organic matter production and shell-generated habitat. *Journal of the North American Benthological Society* 17(1):81-94.
- Sylvester, F., D. Boltovskoy & D. Cataldo, 2007. The invasive bivalve *Limnoperna fortunei* enhances benthic invertebrate densities in South American floodplain rivers. Springer.
- Sylvester, F., O. Kalaci, B. Leung, A. Lacoursière-Roussel, C. C. Murray, F. M. Choi, M. A. Bravo, T. W. Therriault & H. J. MacIsaac, 2011. Hull fouling as an invasion vector: can simple models explain a complex problem? *Journal of Applied Ecology* 48(2):415-423.
- Uliano-Silva, M., F. F. C. F. Fernandes, I. B. B. Holanda & M. F. Rebelo, 2013. Invasive species as a threat to biodiversity: the golden mussel *Limnoperna fortunei* approaching the Amazon River basin. *Exploring Themes on Aquatic Toxicology Kerala* P135–148.
- Underwood, A. & M. Anderson, 1994. Seasonal and temporal aspects of recruitment and succession in an intertidal estuarine fouling assemblage. *Journal of the Marine Biological Association of the United Kingdom* 74(3):563-584.
- Velde, G. v. d. & S. Rajagopal, 2006. *Biological invasions: concepts to understand and predict a global threat*. Springer.
- Venkatesan, R. & P. S. Murthy, 2008. Macrofouling control in power plants. *Series on Biofilms* doi:https://doi.org/10.1007/7142_2008_14.
- Wegner, B., A. L. Kronsbein, M. Gillefalk, K. Van de Weyer, J. Köhler, E. Funke, M. T. Monaghan & S. Hilt, 2019. Mutual facilitation among invading *Nuttall's* waterweed and quagga mussels. *Frontiers in plant science* 10.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.